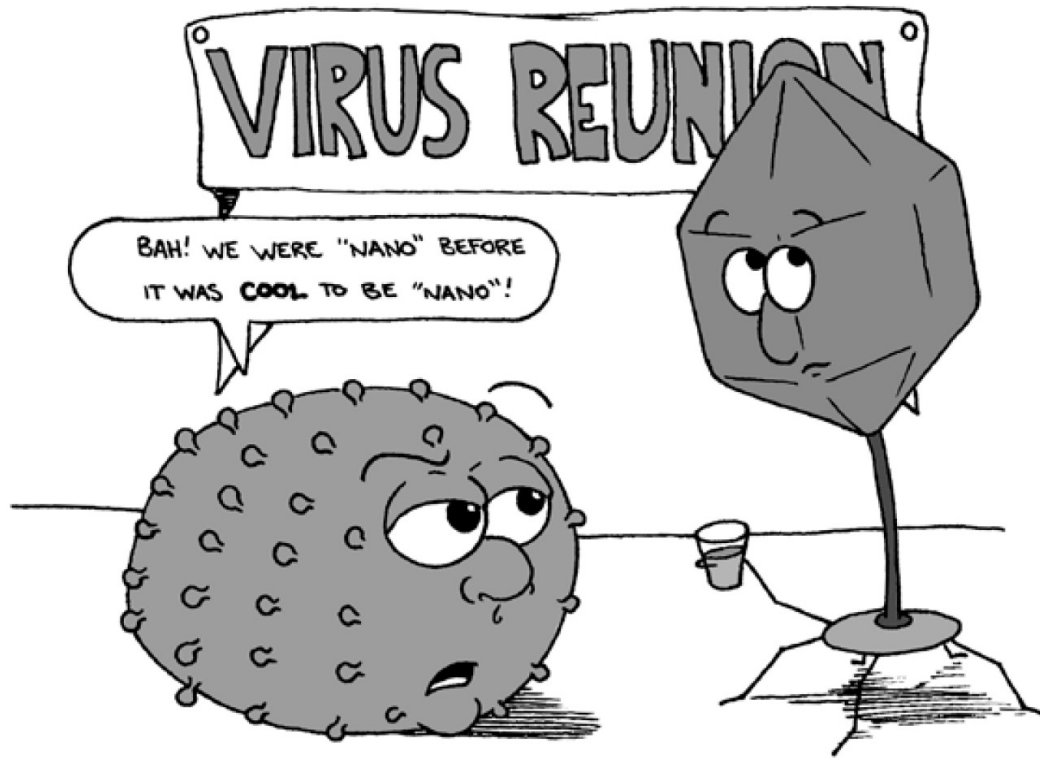


Origins, structure, and impact of viruses on the evolution of cellular life



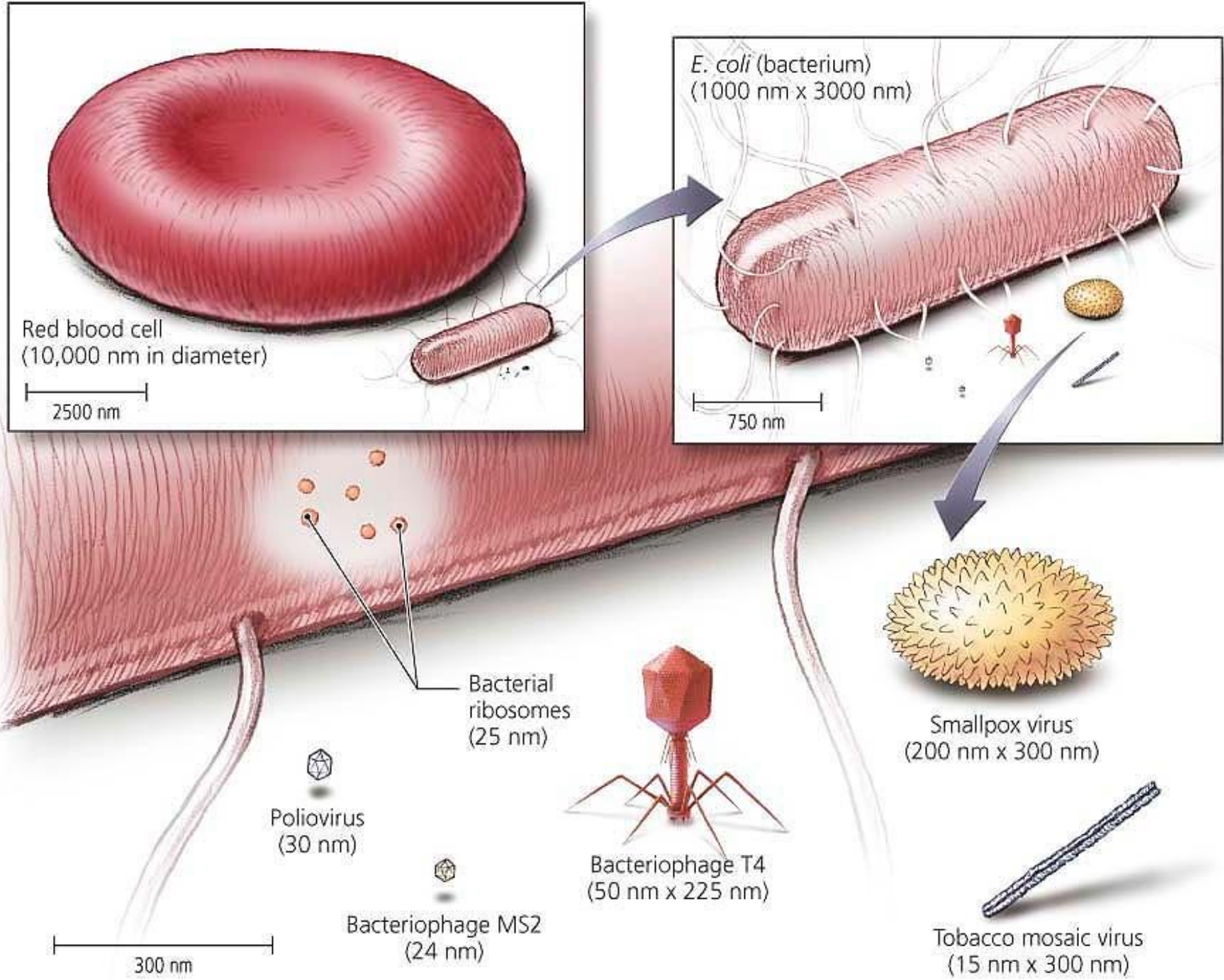
Pavel Plevka

What is a virus?

Origins of viruses

Virus structures and infection processes

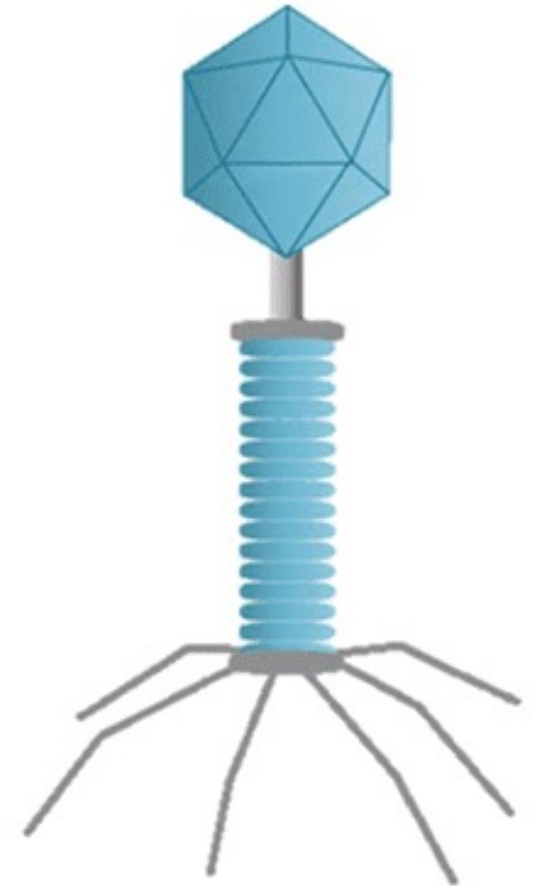
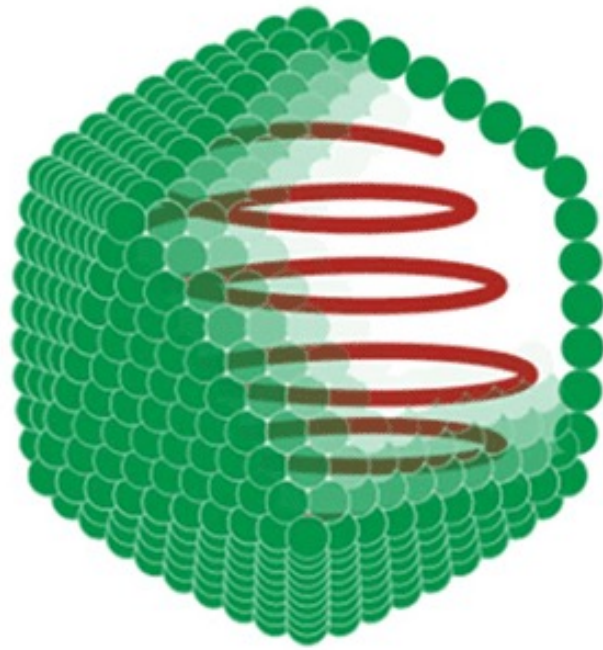
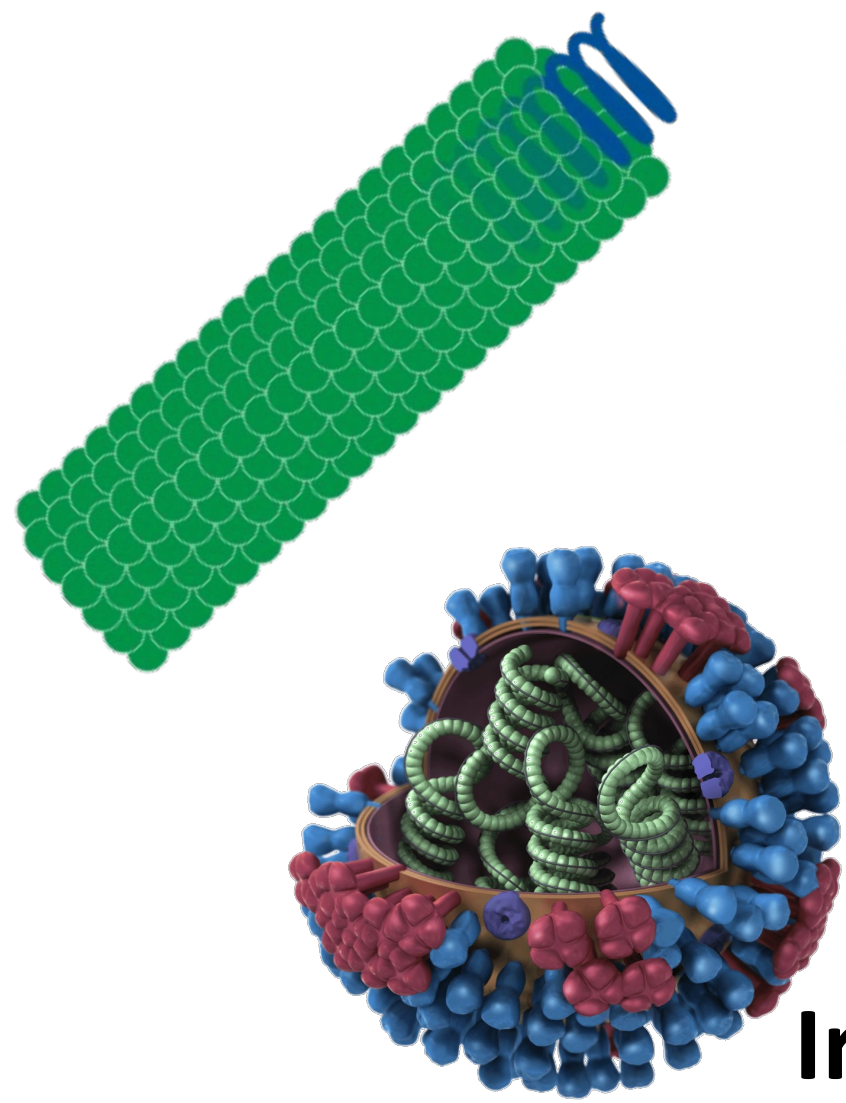
Extra



Helical

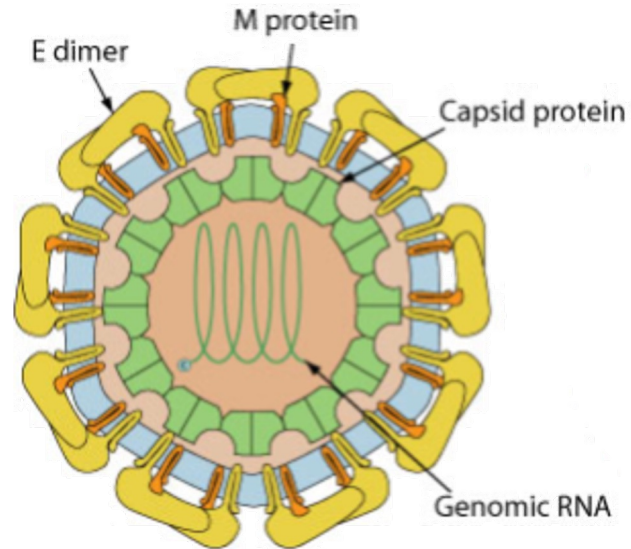
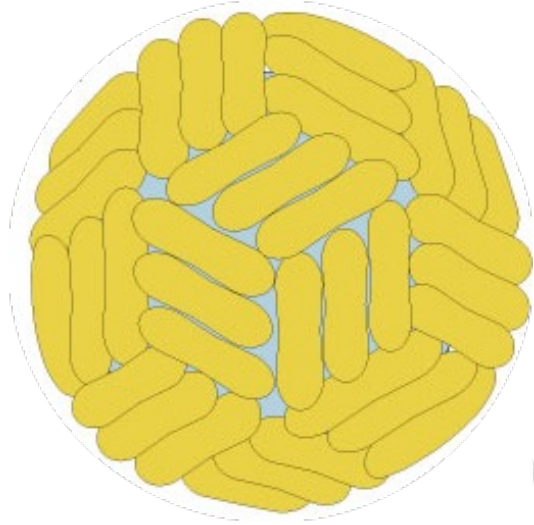
Icosahedral

Complex

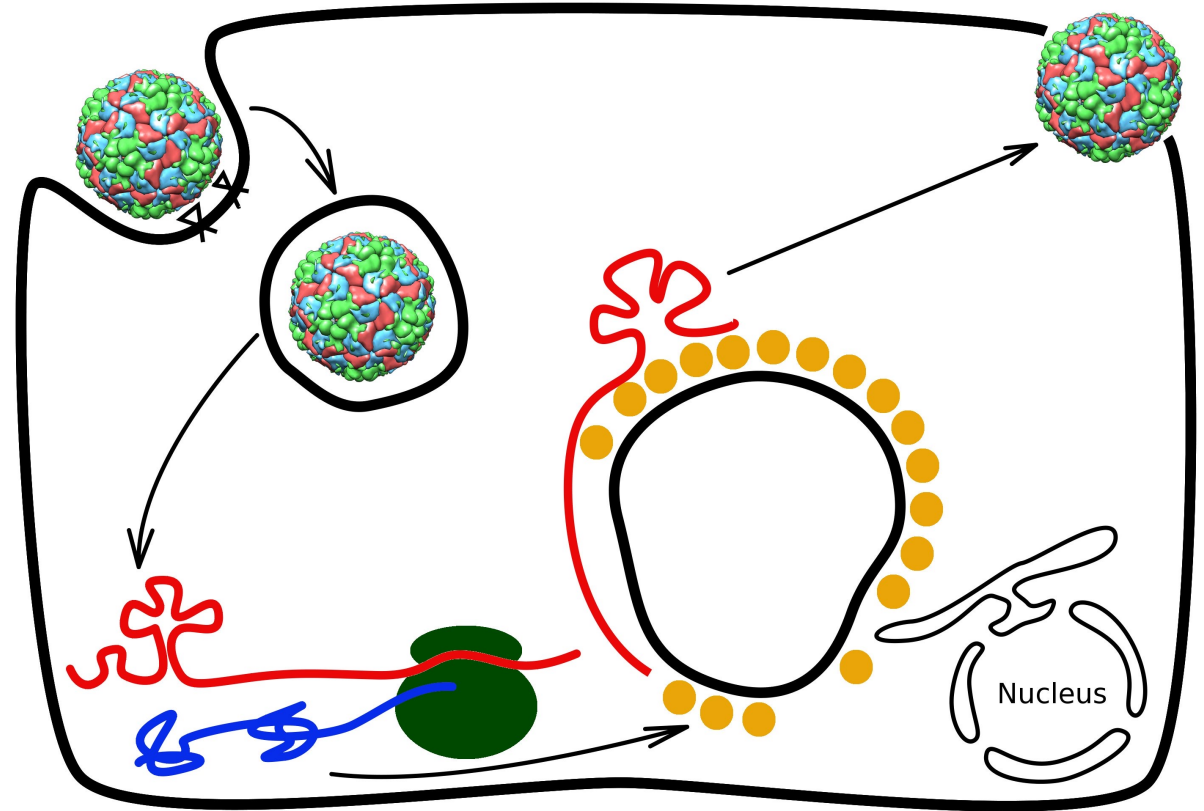


Irregular

Virion



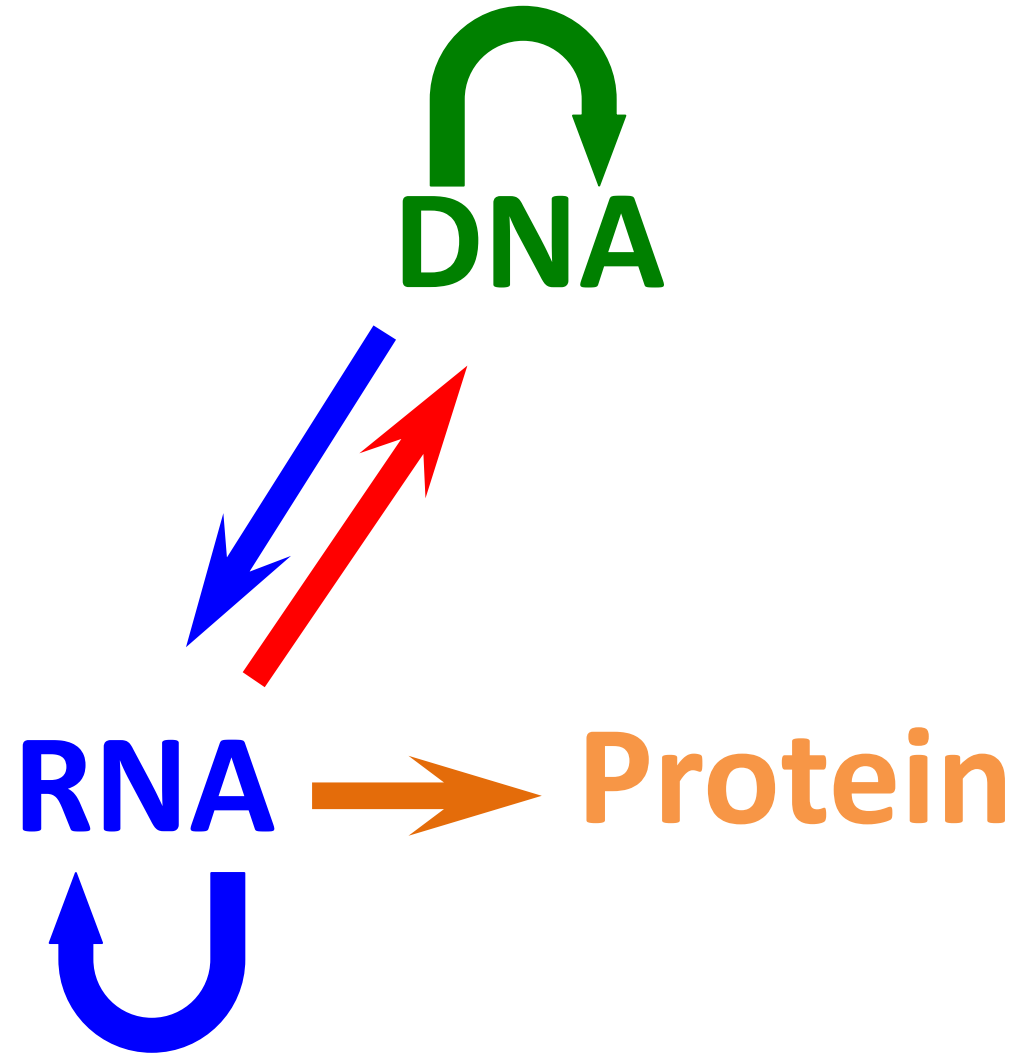
Infected cell

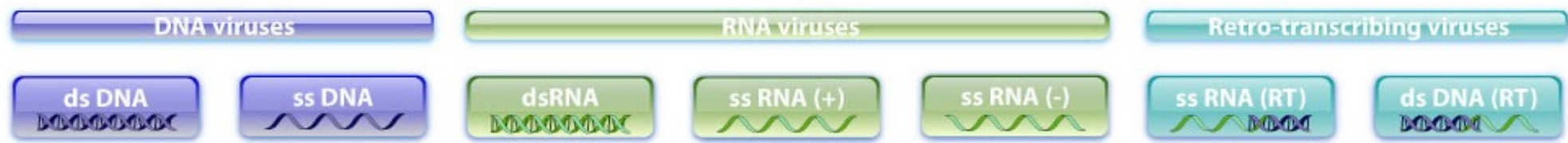


Cellular organisms

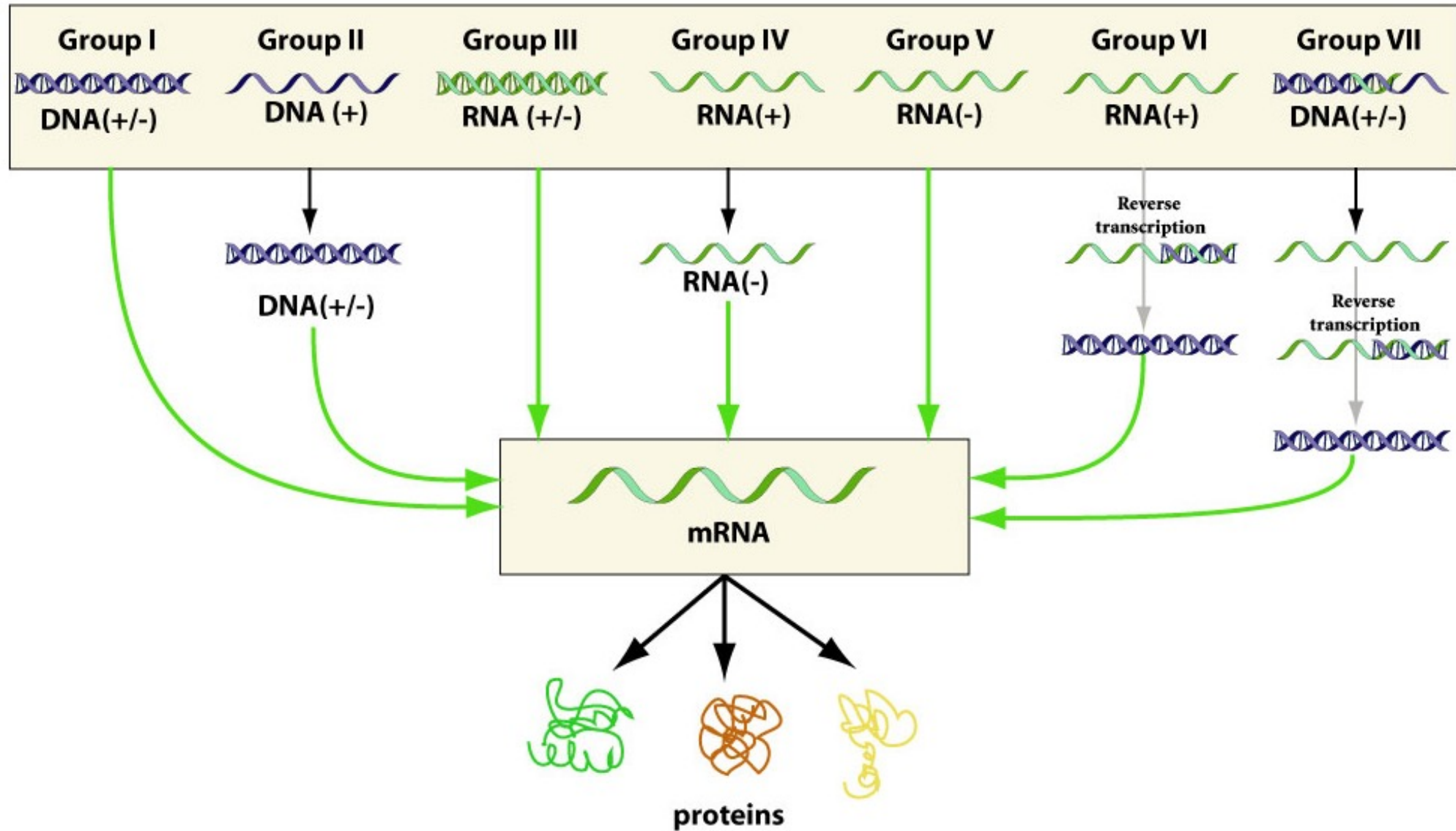


Viruses





Genetic material present in the virion



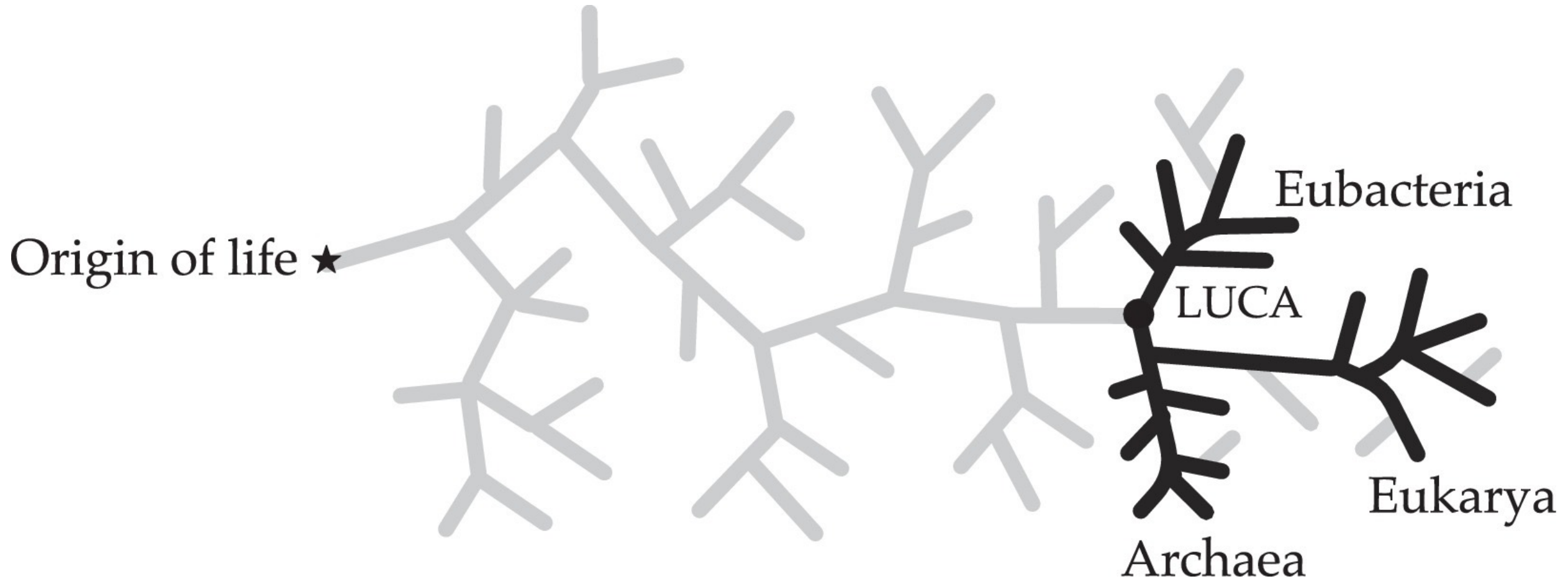
What is a virus?

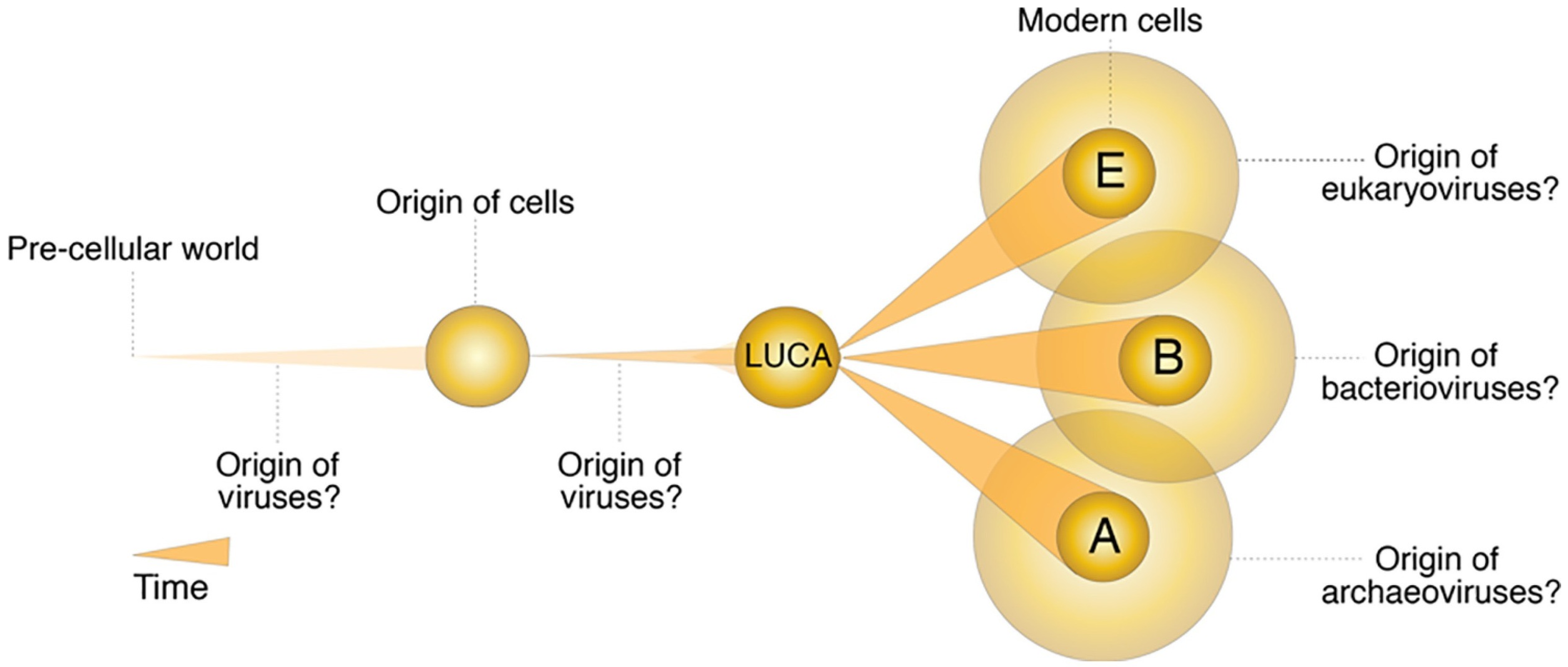
Origins of viruses

Virus structures and infection processes

Extra

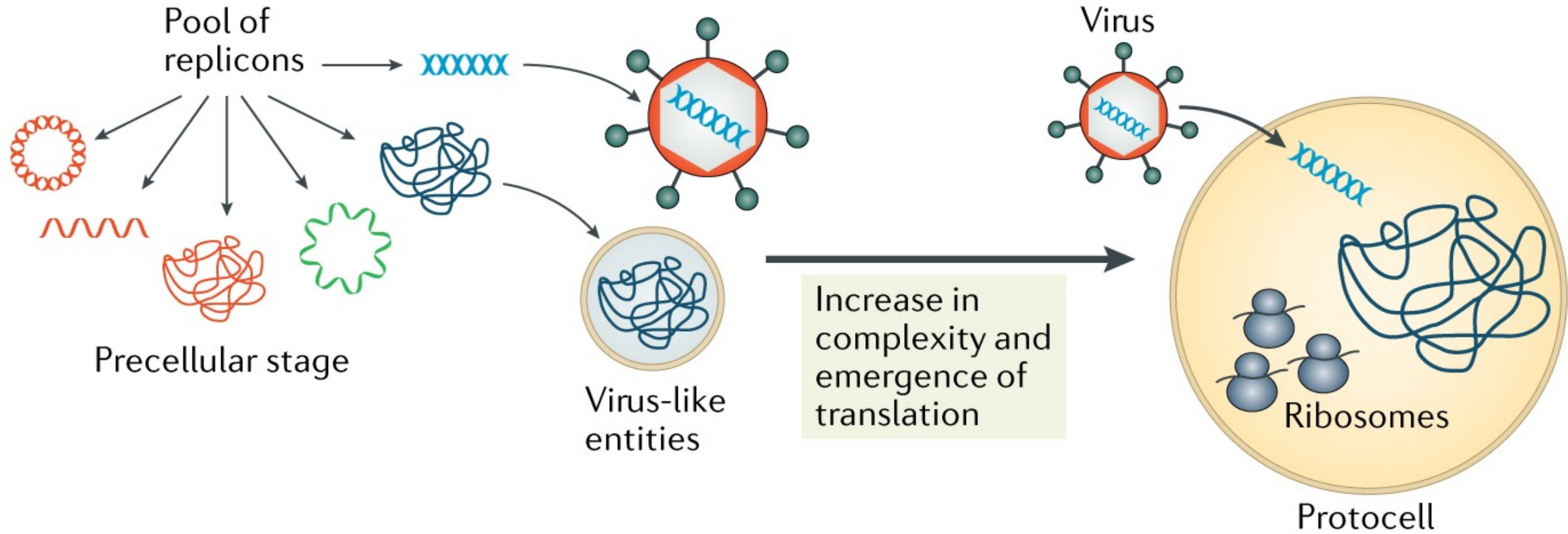
Last Universal Common Ancestor (LUCA)





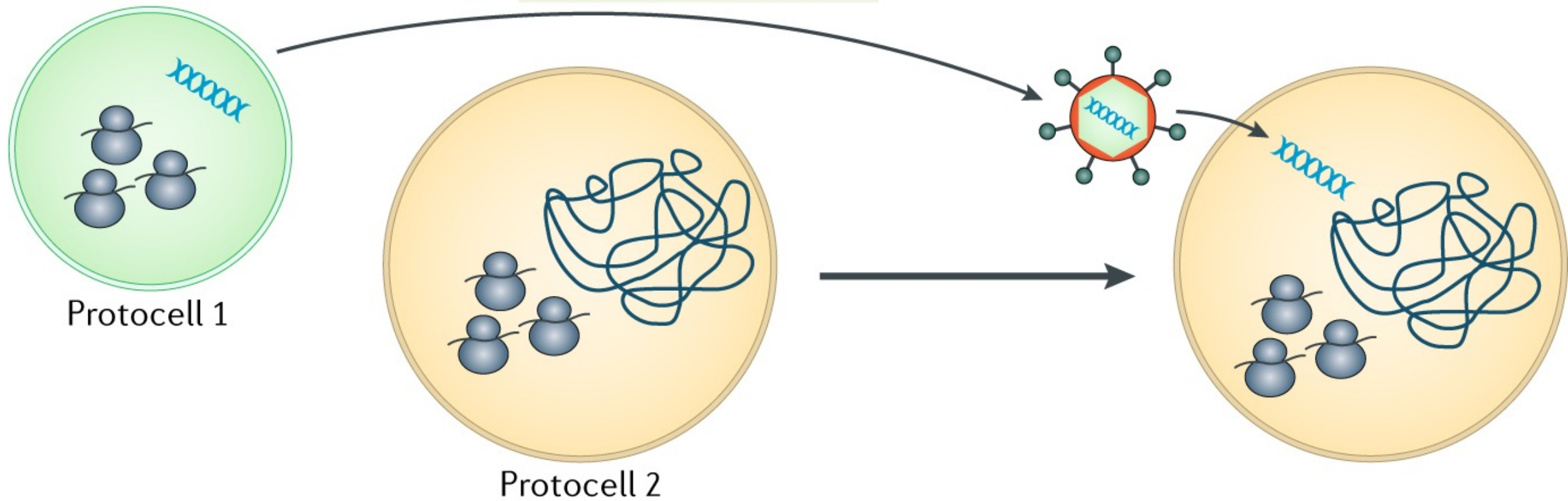
Trends in Microbiology

'Virus early' hypothesis

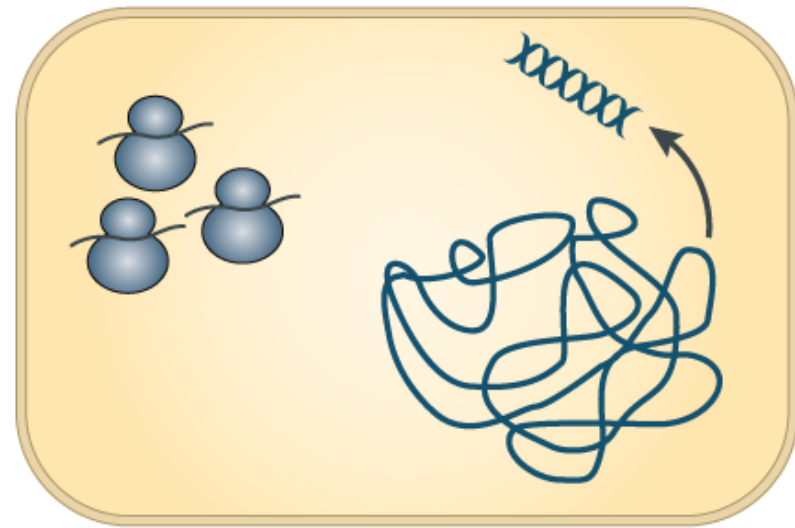


'Regression' hypothesis

Reductive evolution,
loss of translation and
obligate parasitism



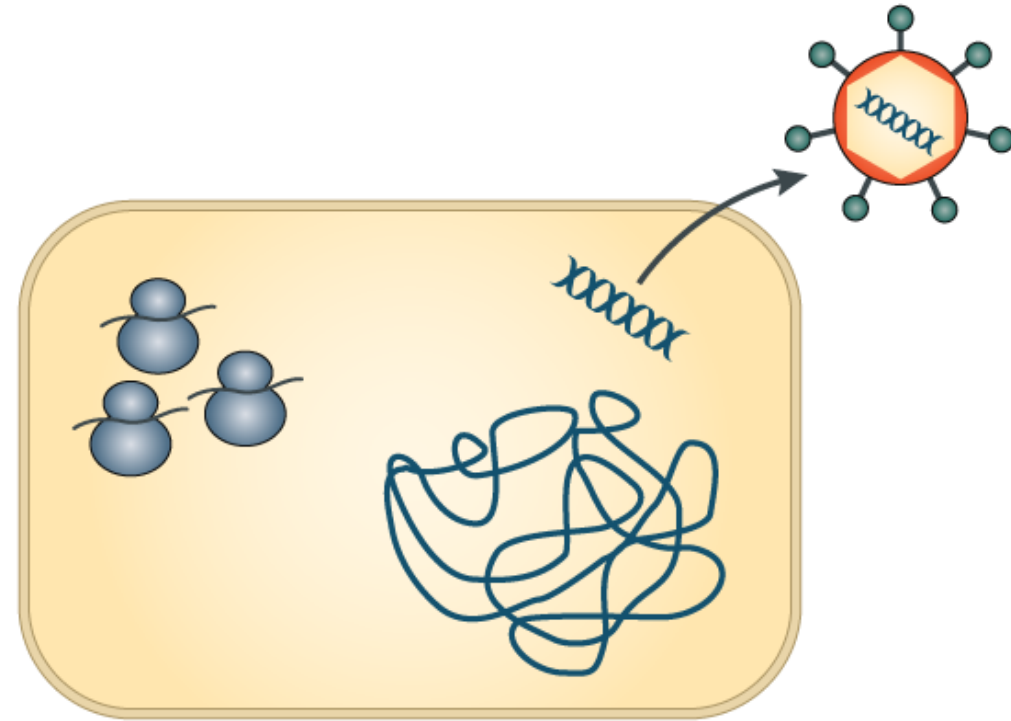
'Escaped genes' hypothesis



Modern cell



Autonomous replication of selfish cellular genes



What is a virus?

Origins of viruses

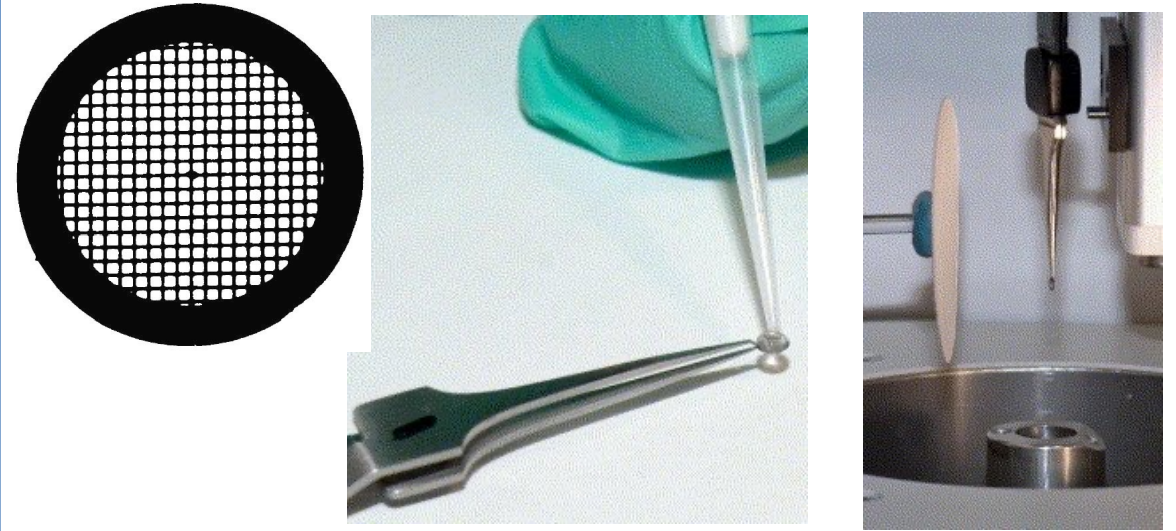
Virus structures and infection processes

Extra

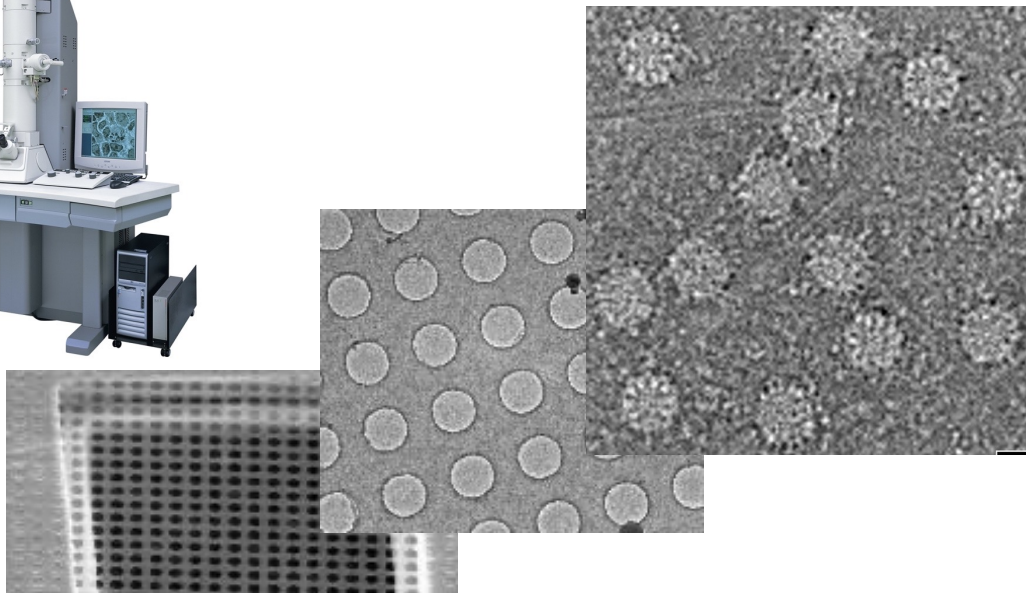
1. Virus purification



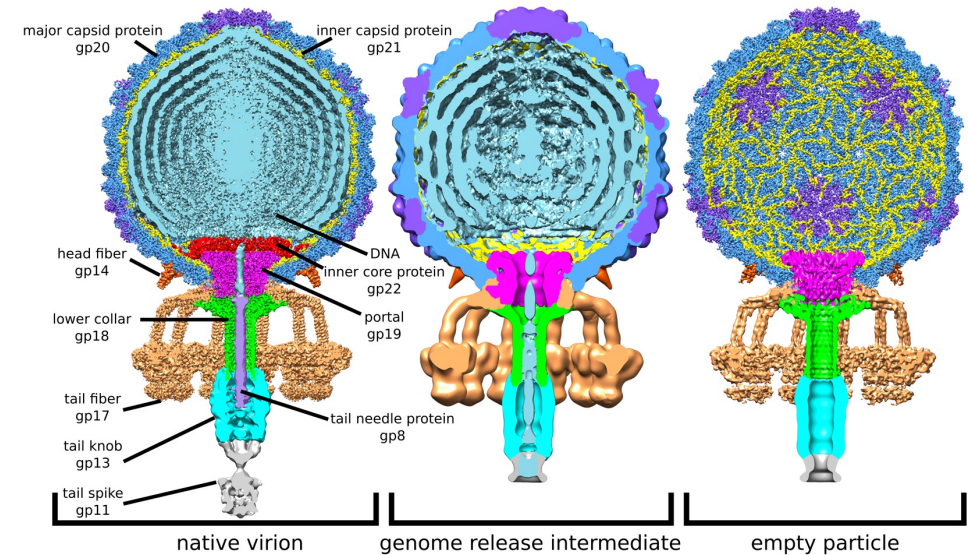
2. Grid preparation

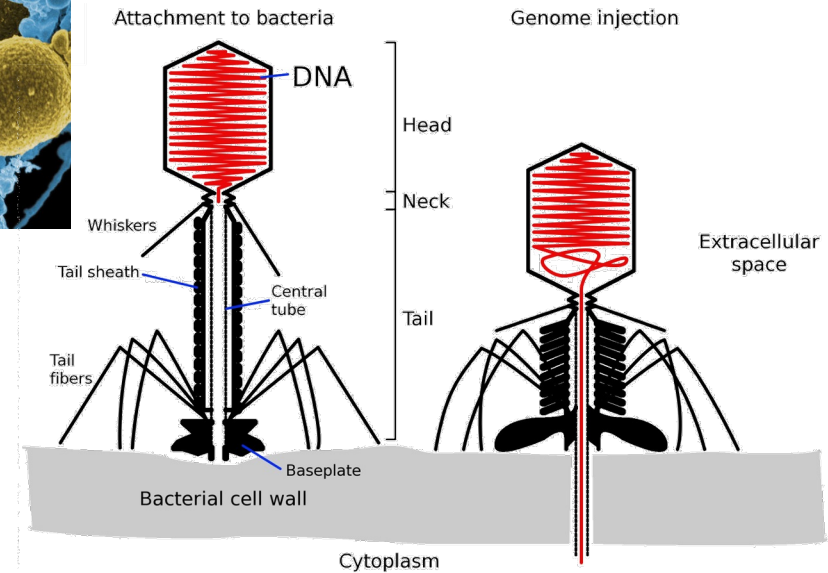
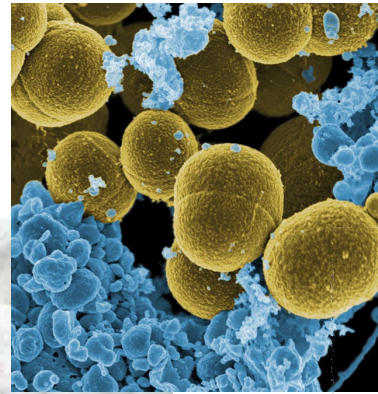
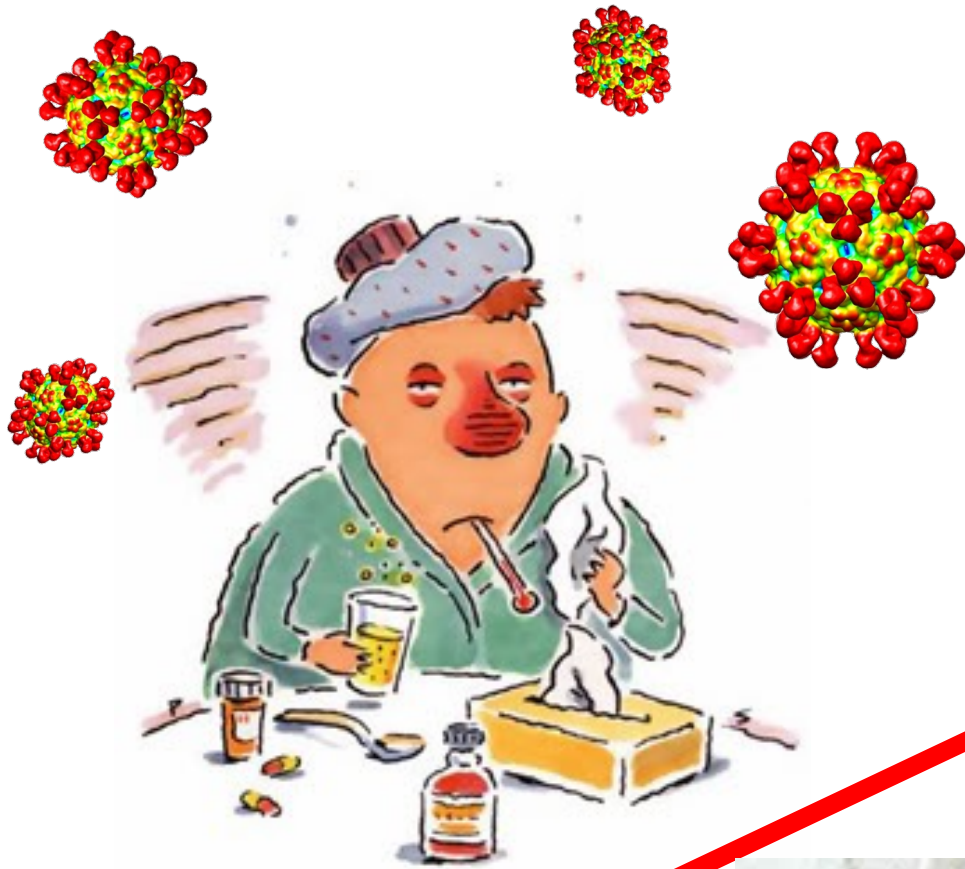


3. cryo-EM

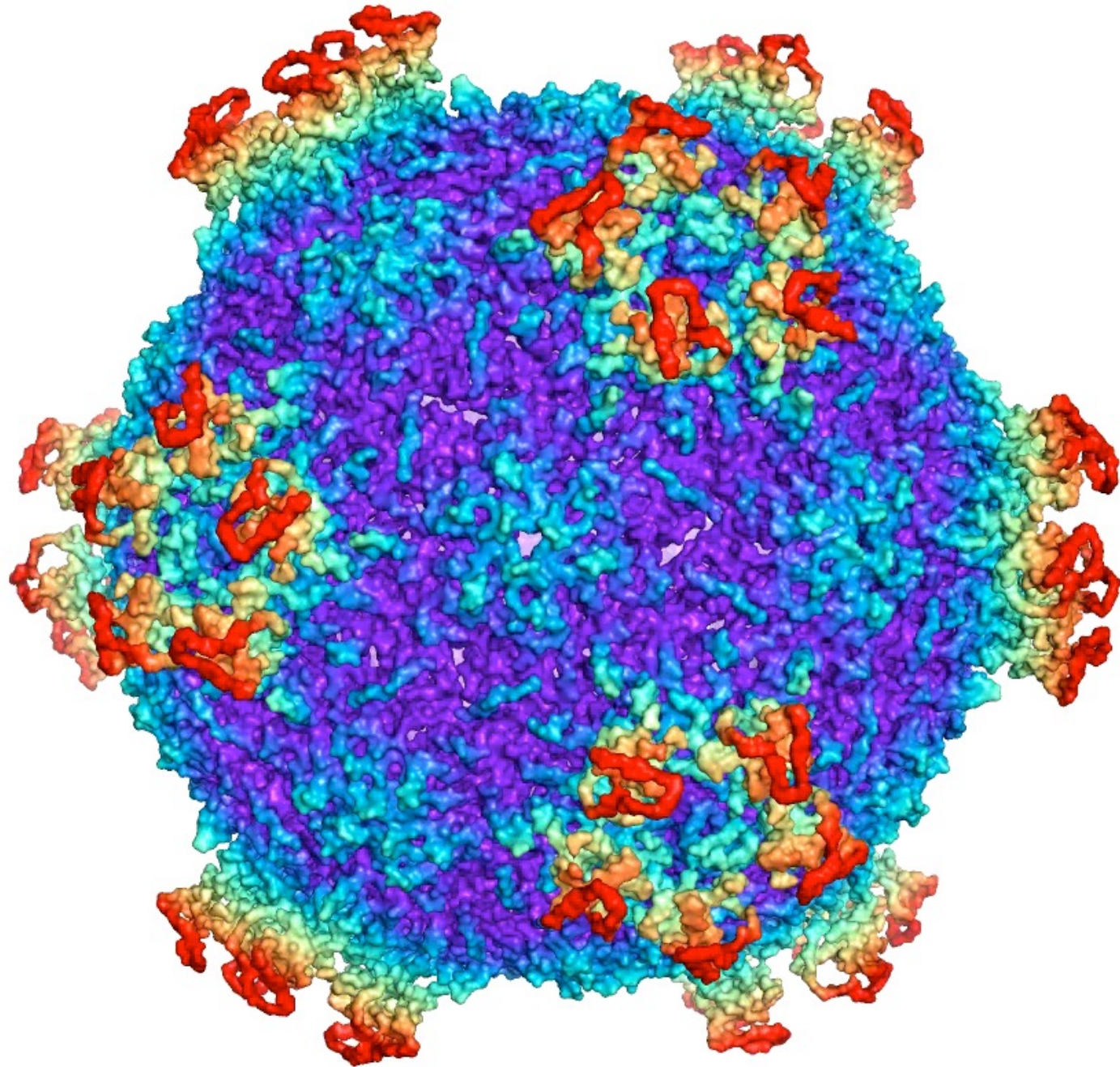


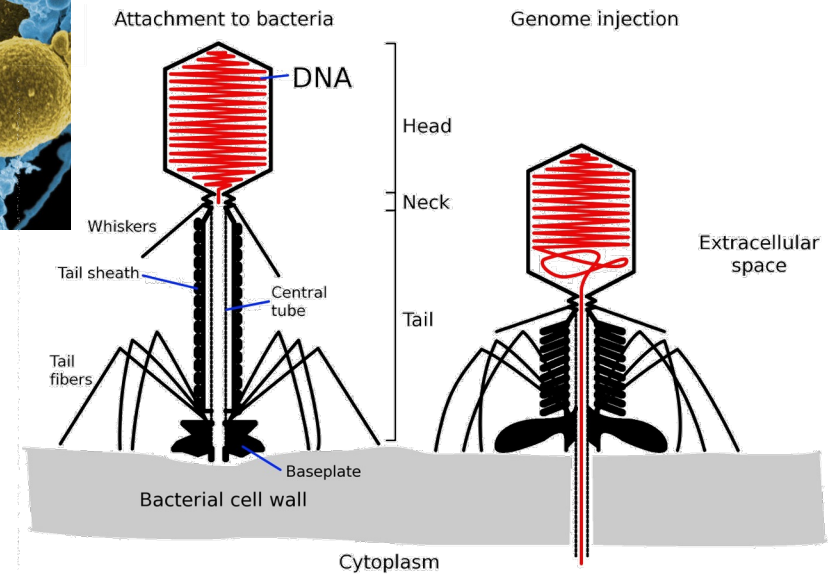
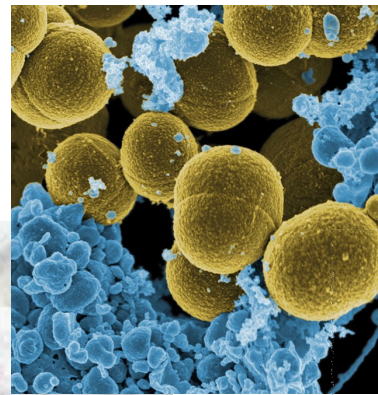
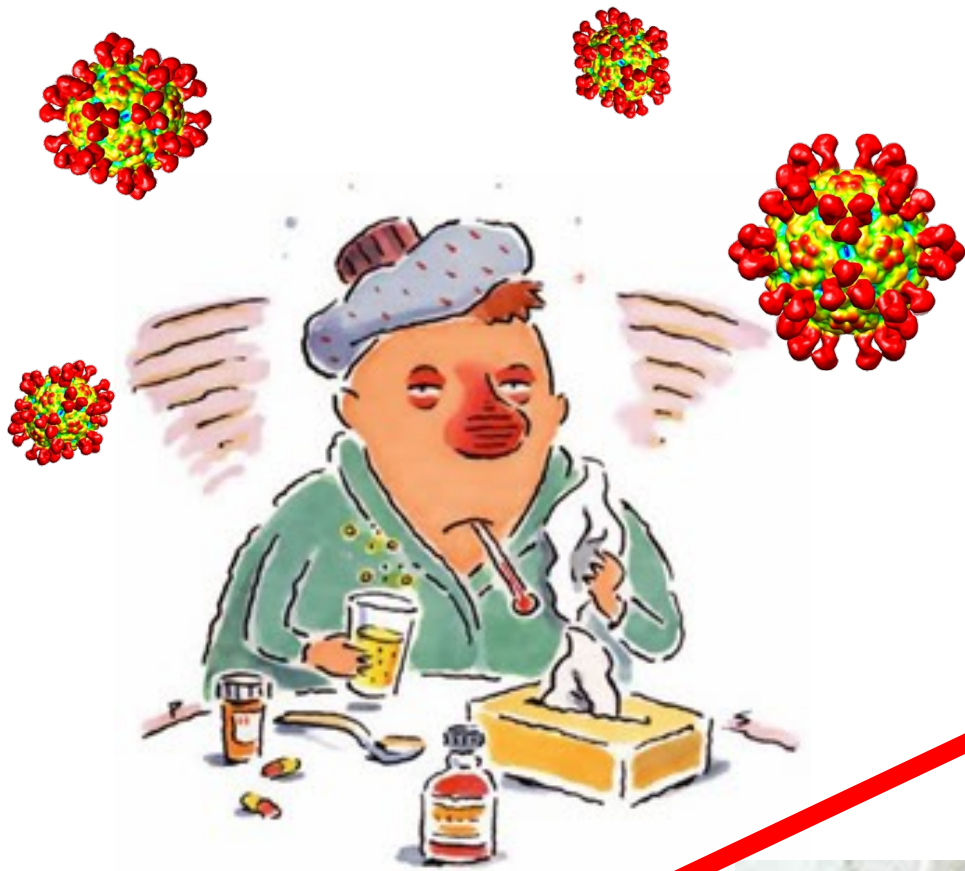
4. Reconstruction

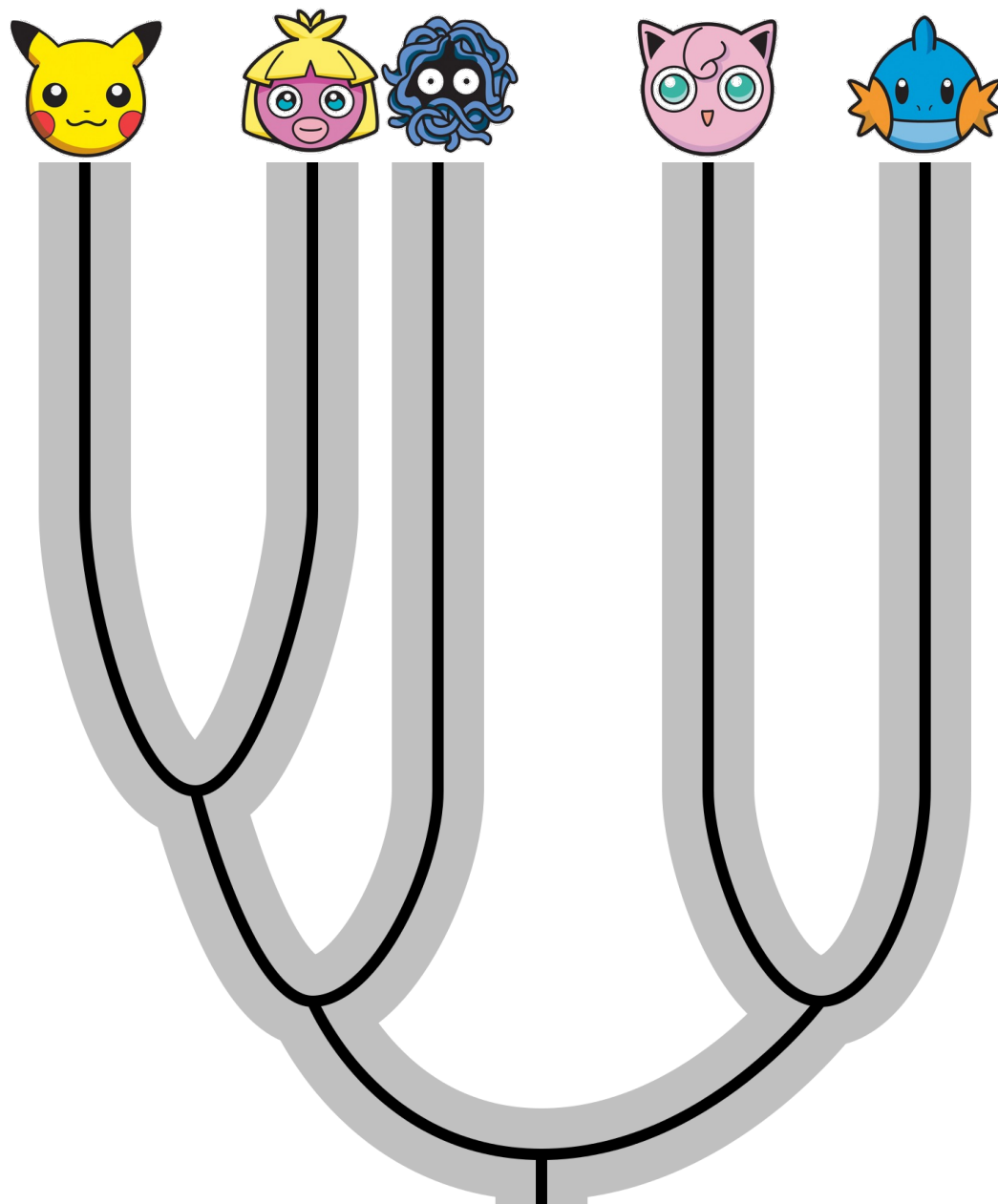


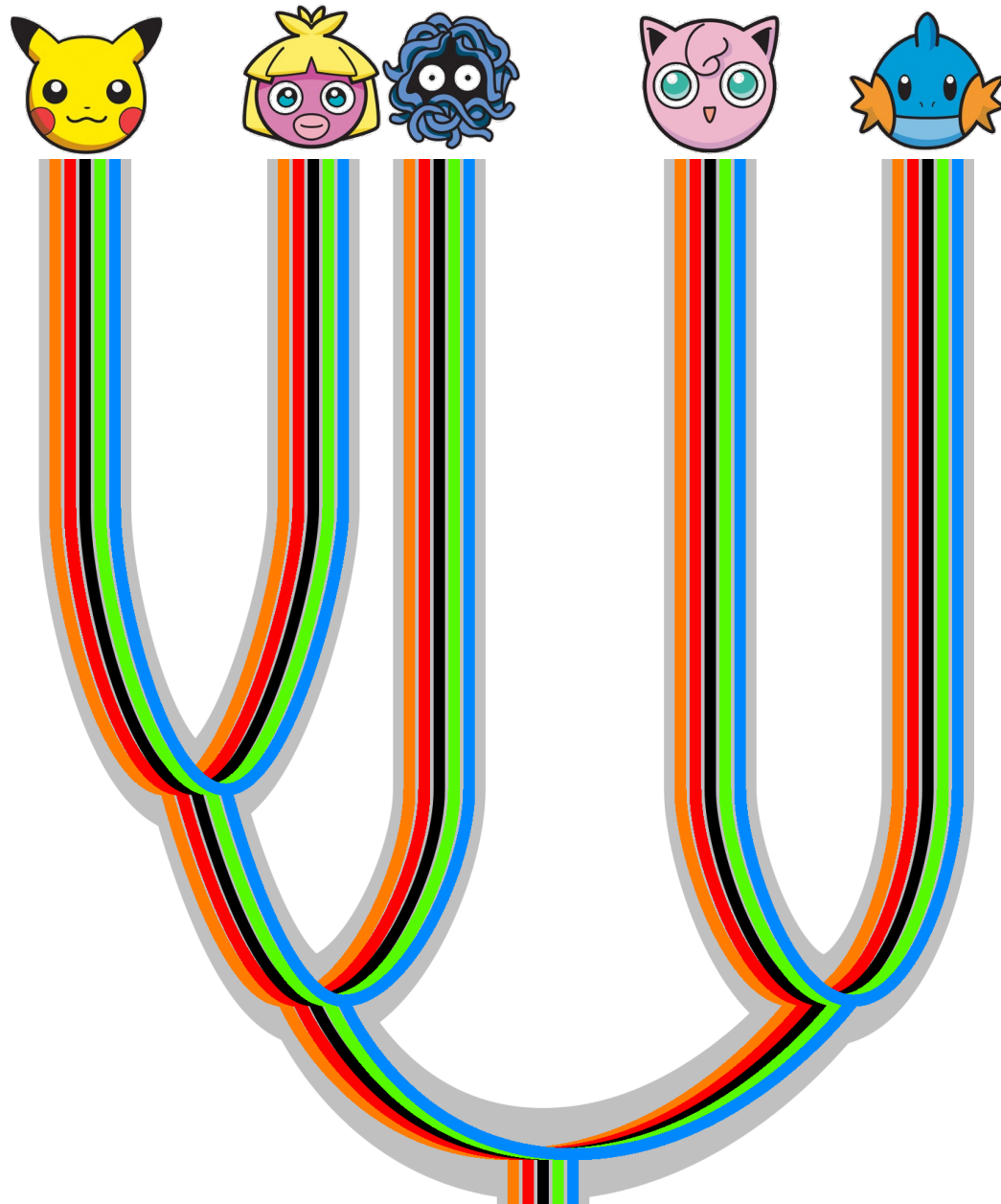


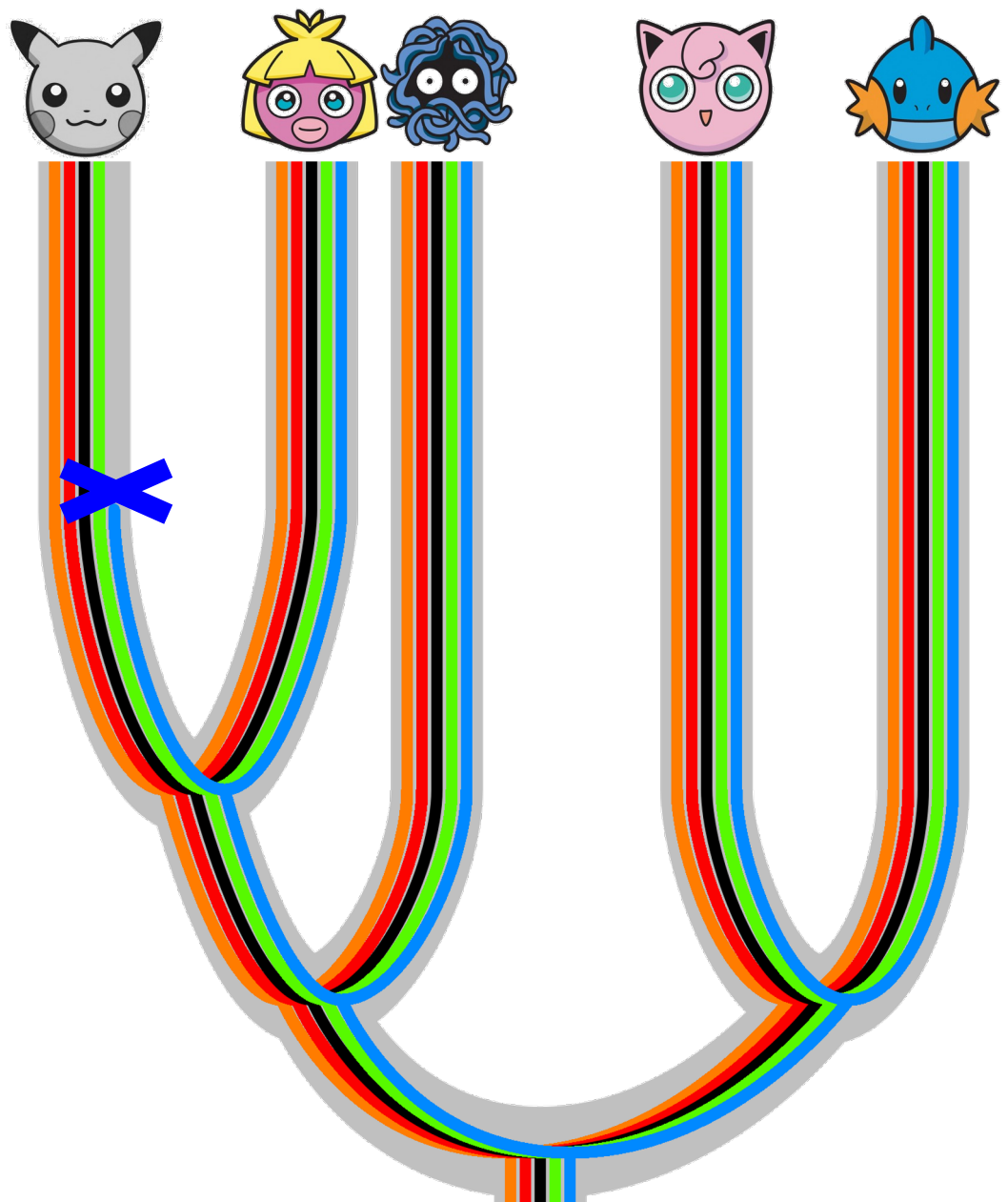
Slow bee
paralysis
virus

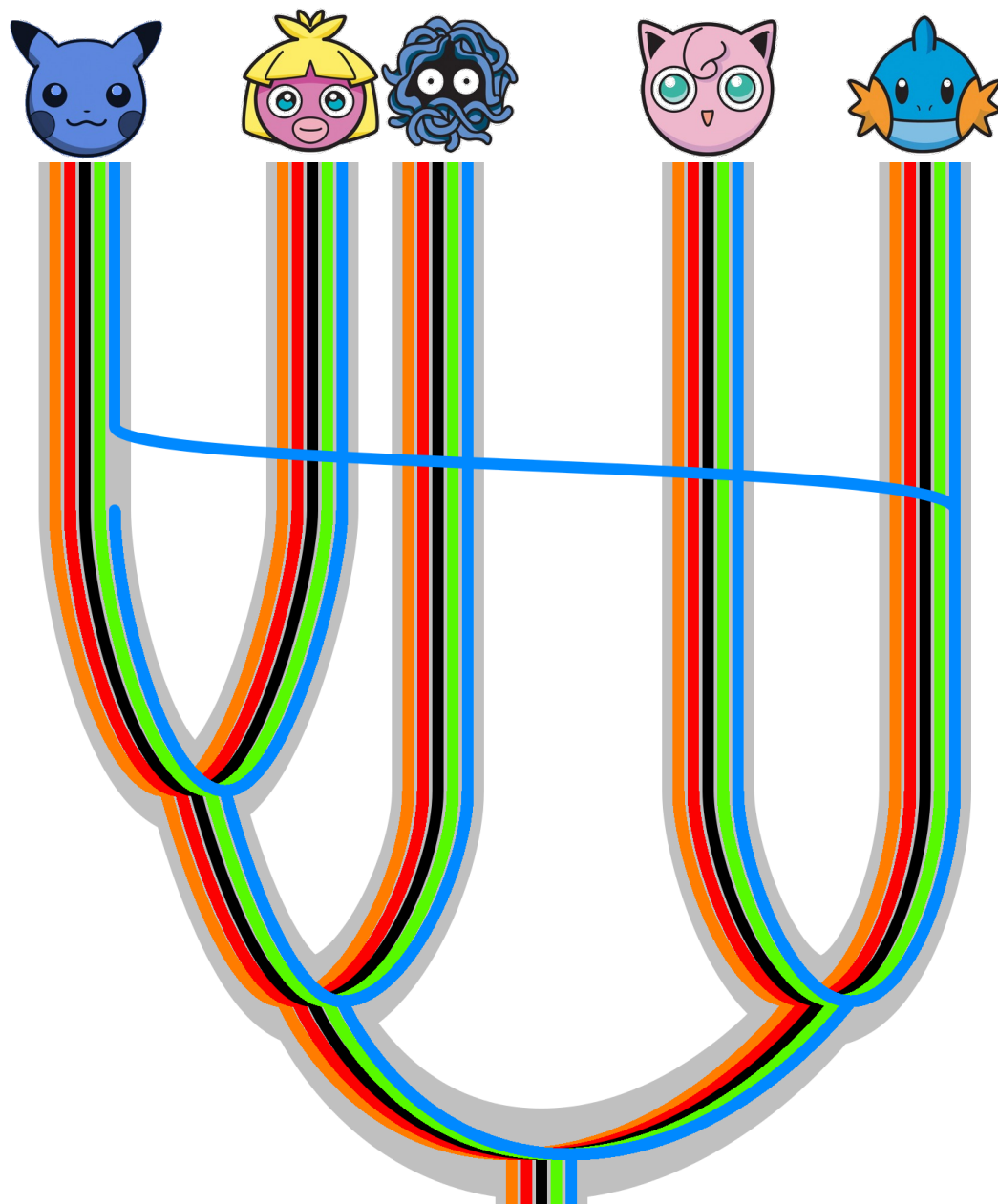


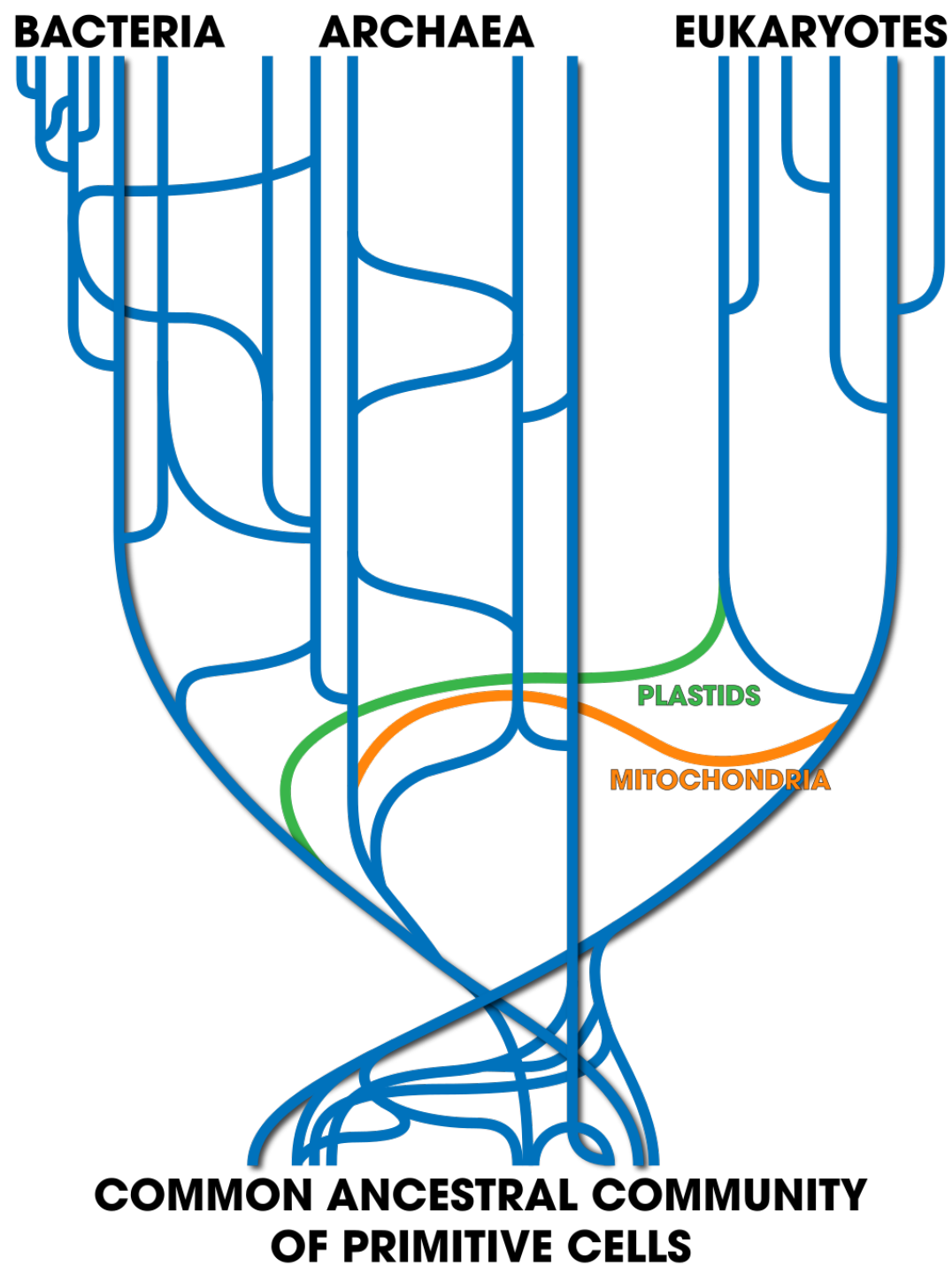




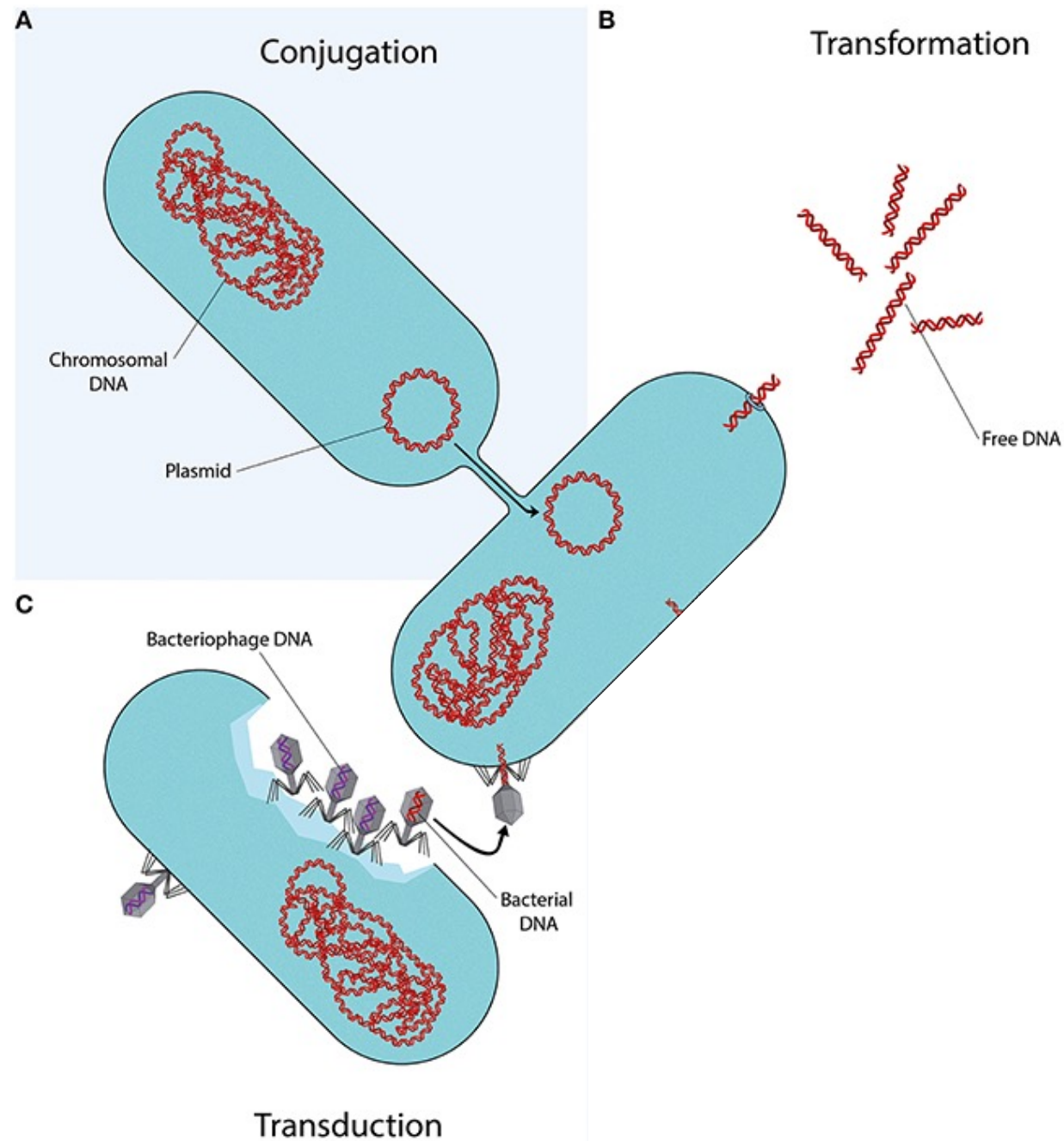




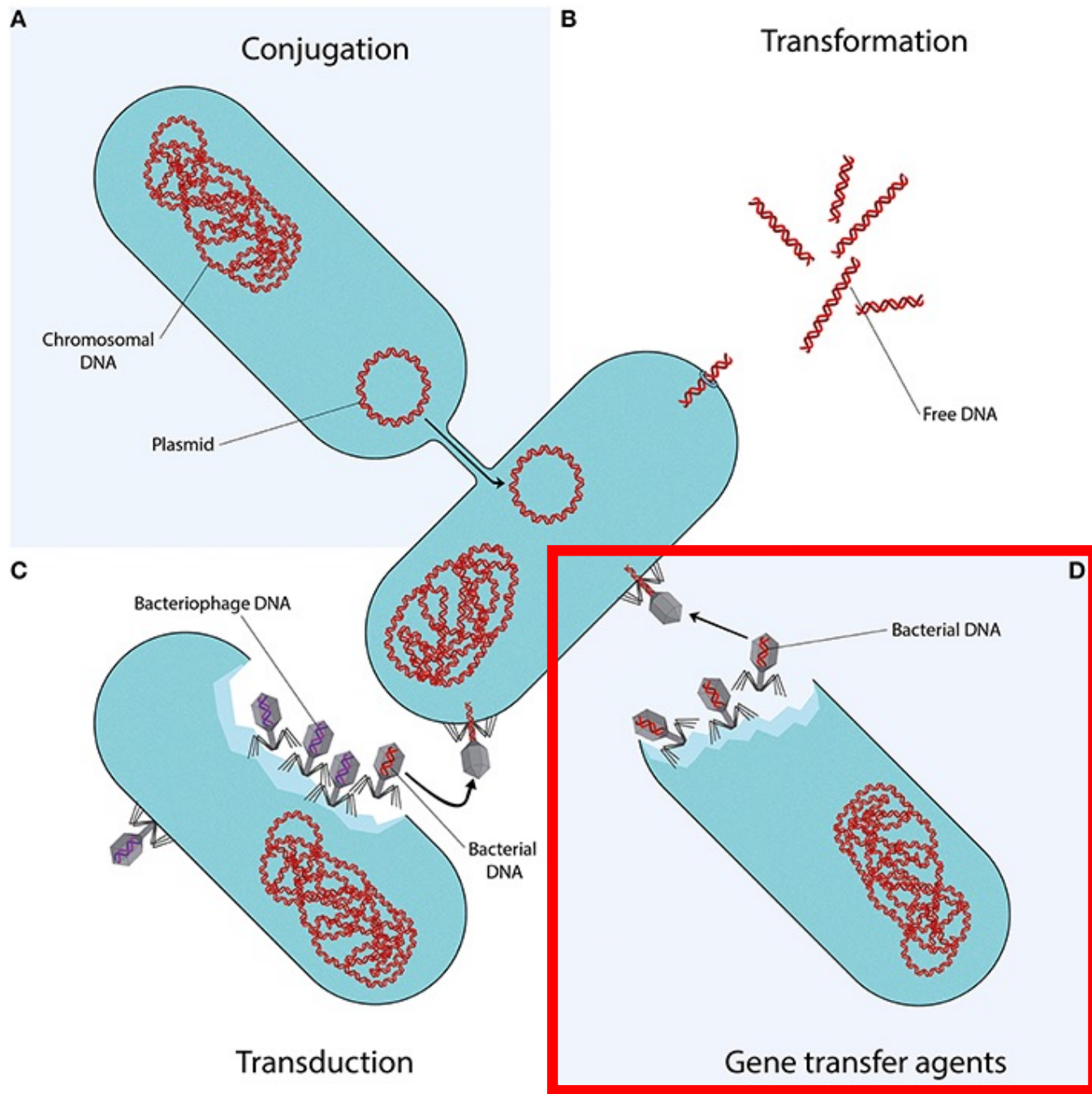




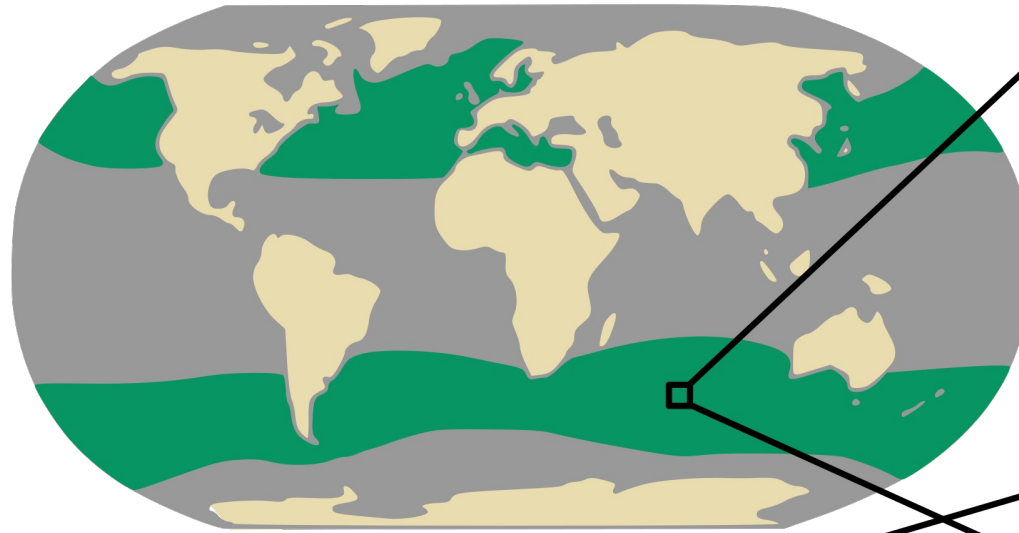
Mechanisms of horizontal gene transfer



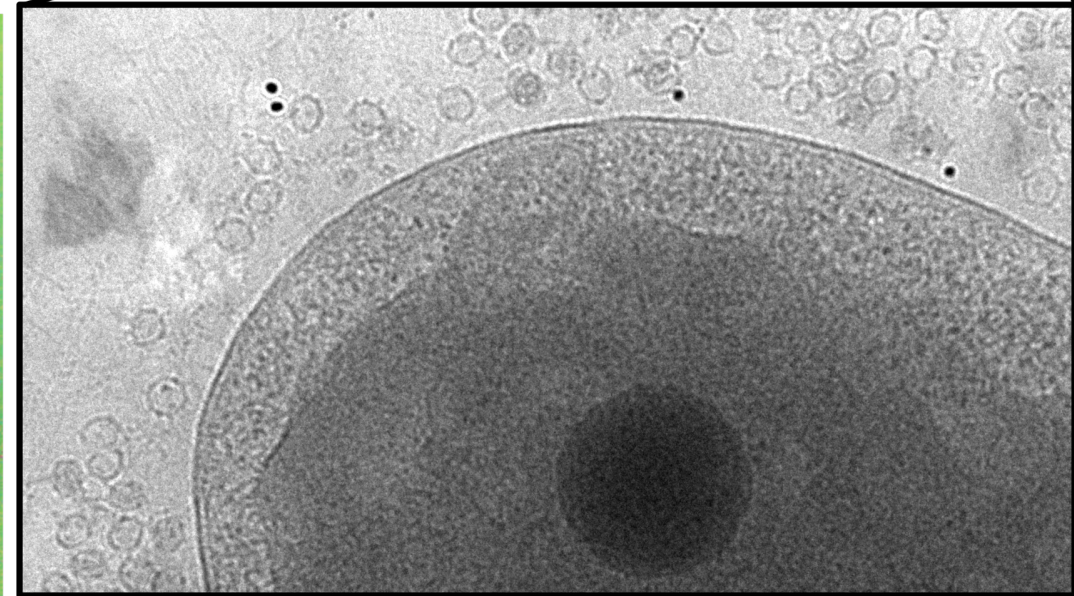
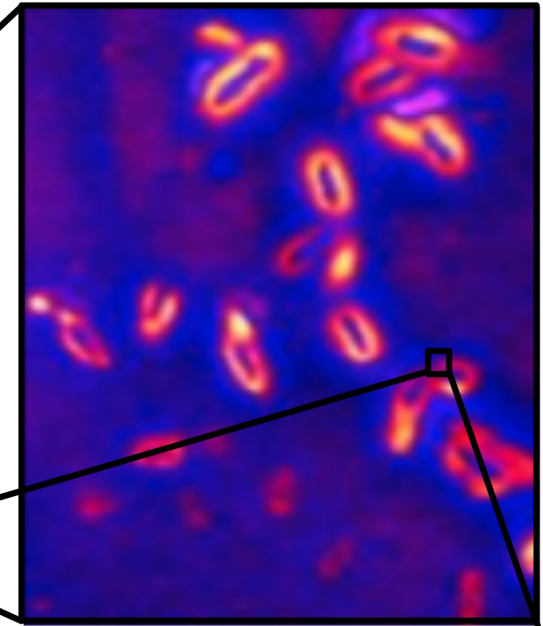
Mechanisms of horizontal gene transfer



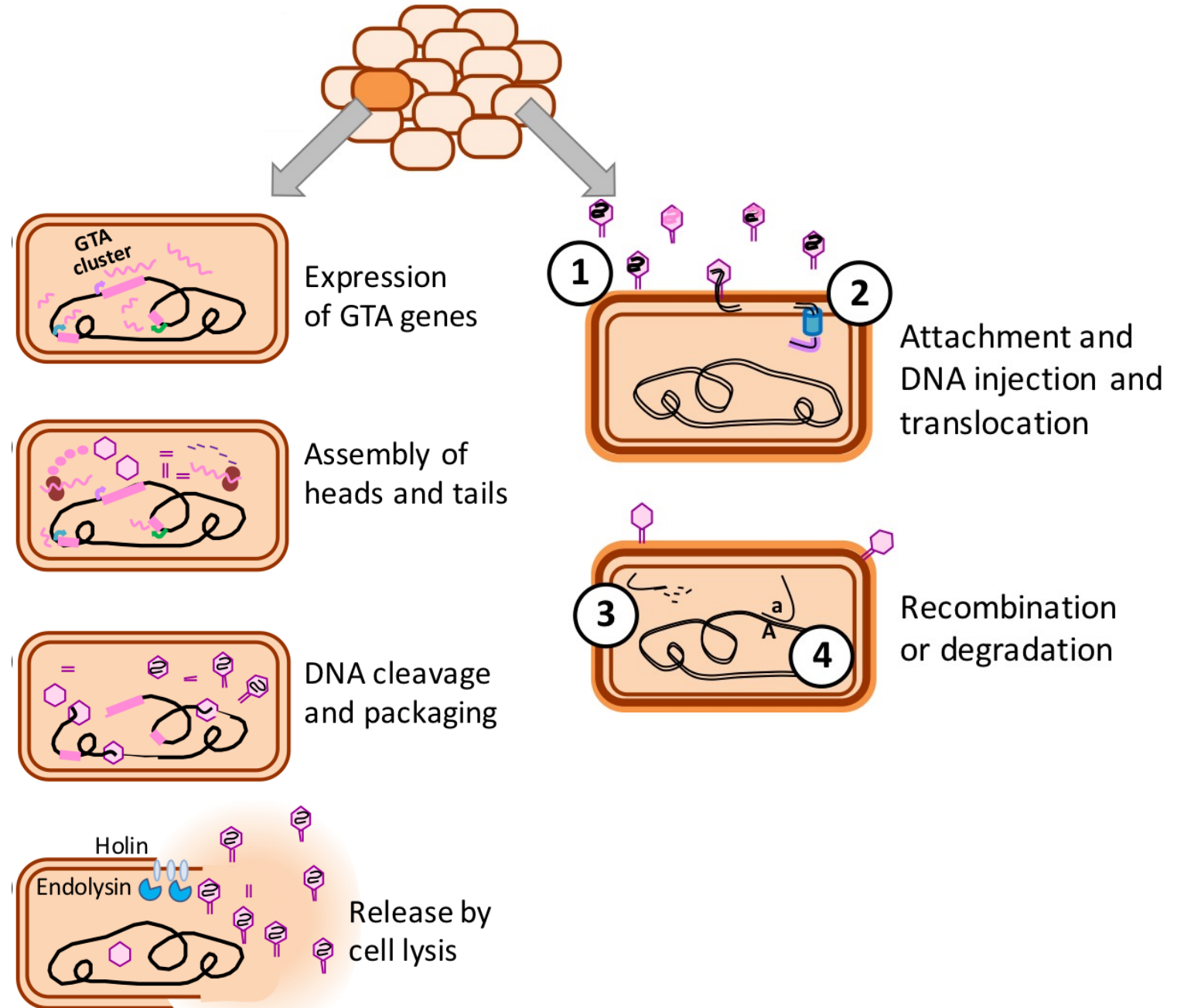
*Rhodobacter
capsulatus*
gene transfer
agent



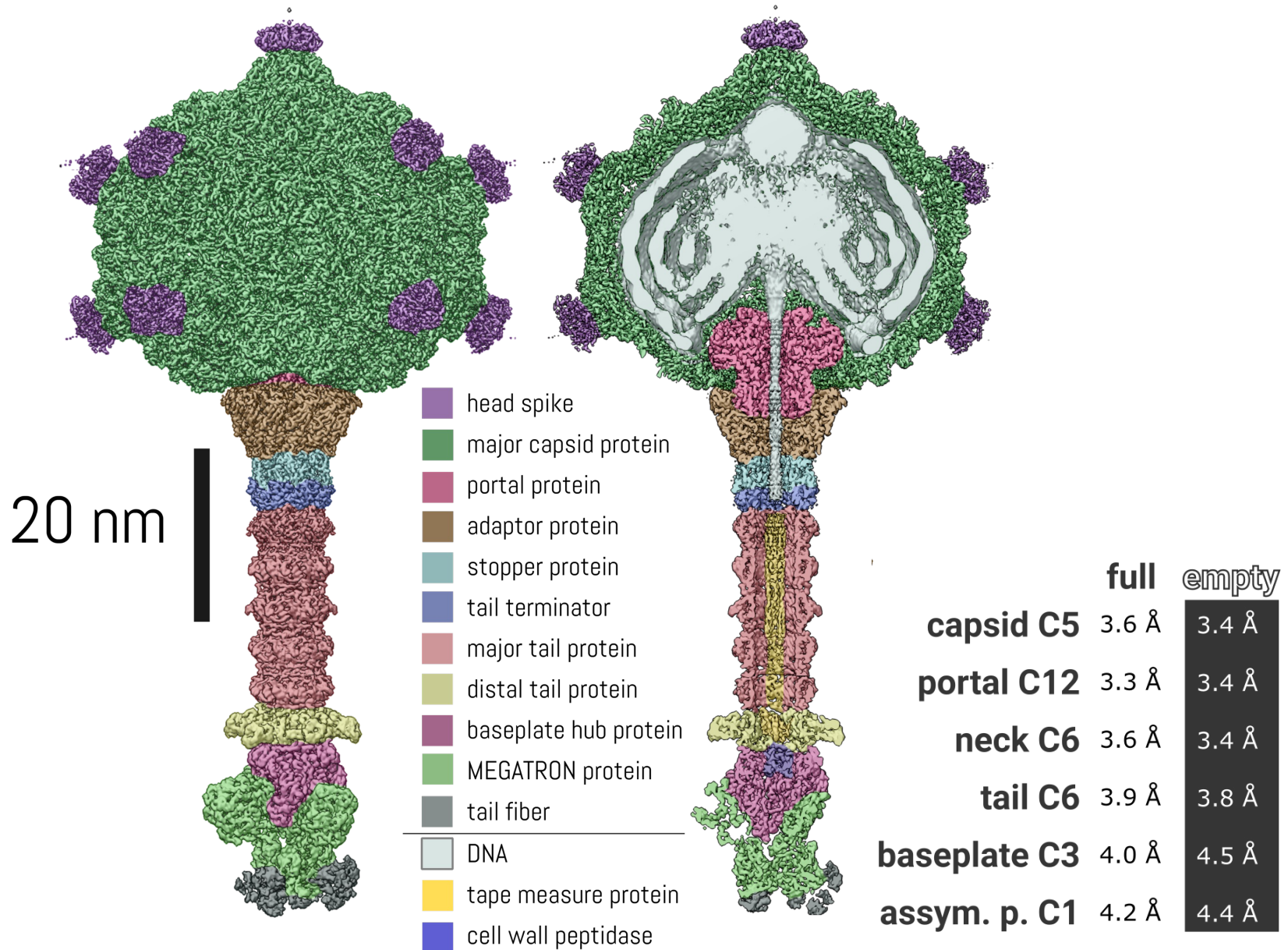
■ Rhodobacterales > 50%



Function of gene transfer agents

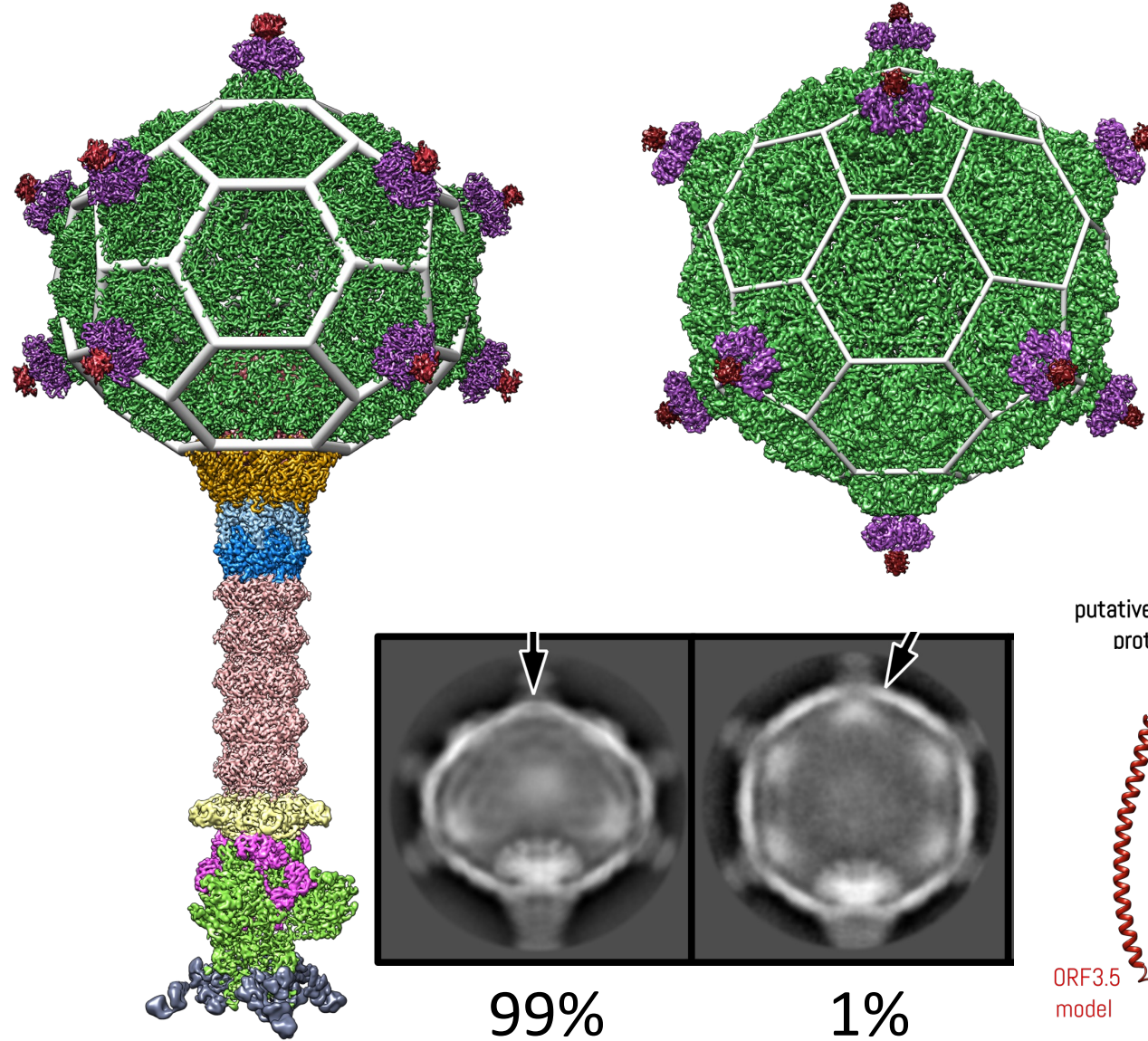
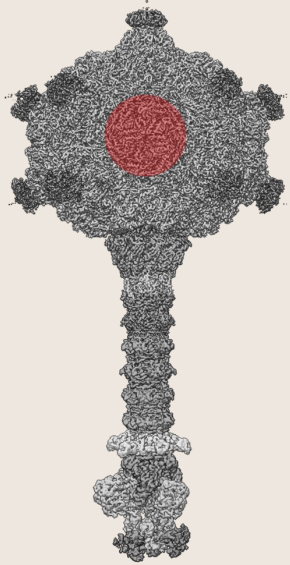


Virion of the GTA particle

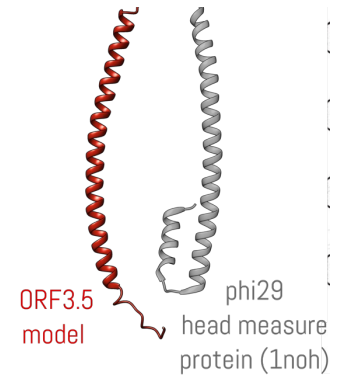


RESULTS

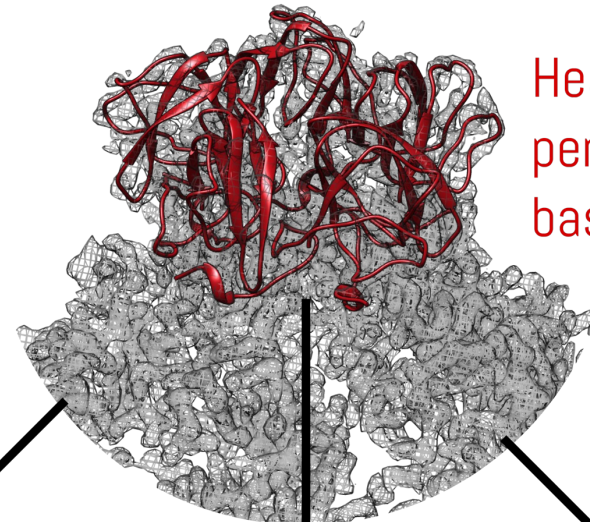
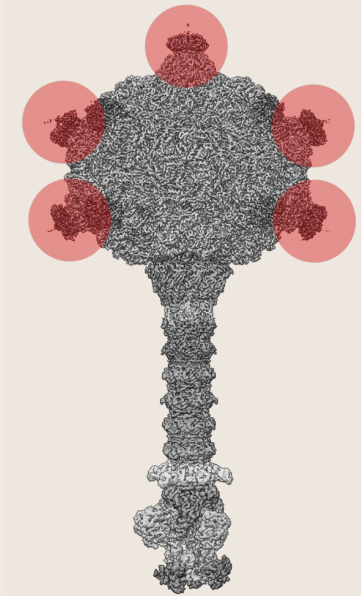
Capsid of the GTA



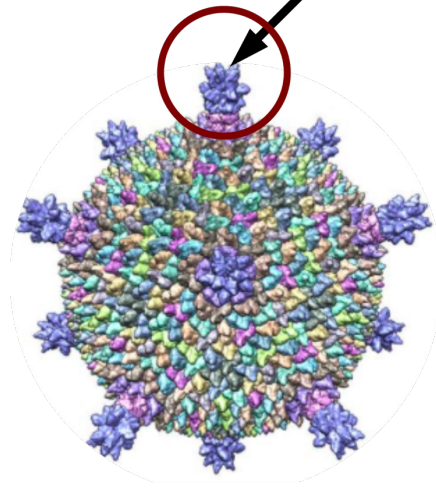
putative head-measure protein ORF3.5



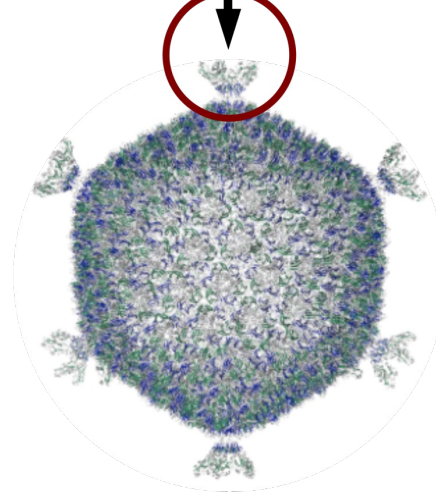
Structural alignment of GTA head-spike



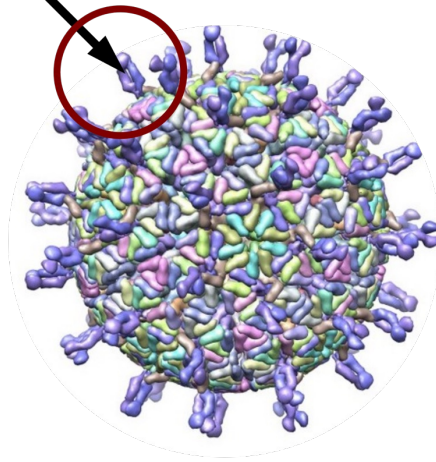
Head-spike
penton
base



Archeal virus
Sulfolobus turreted IV1
(3j31)

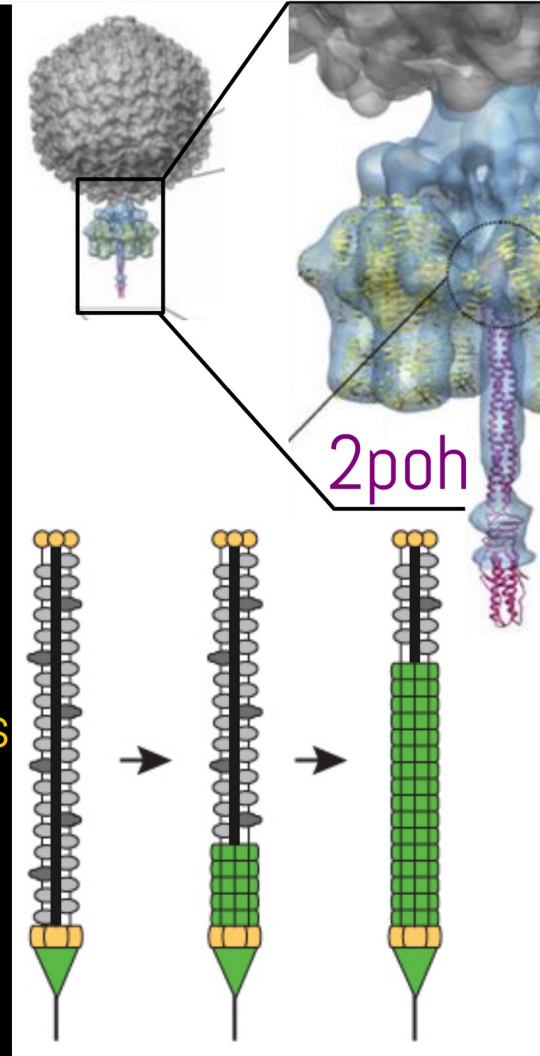
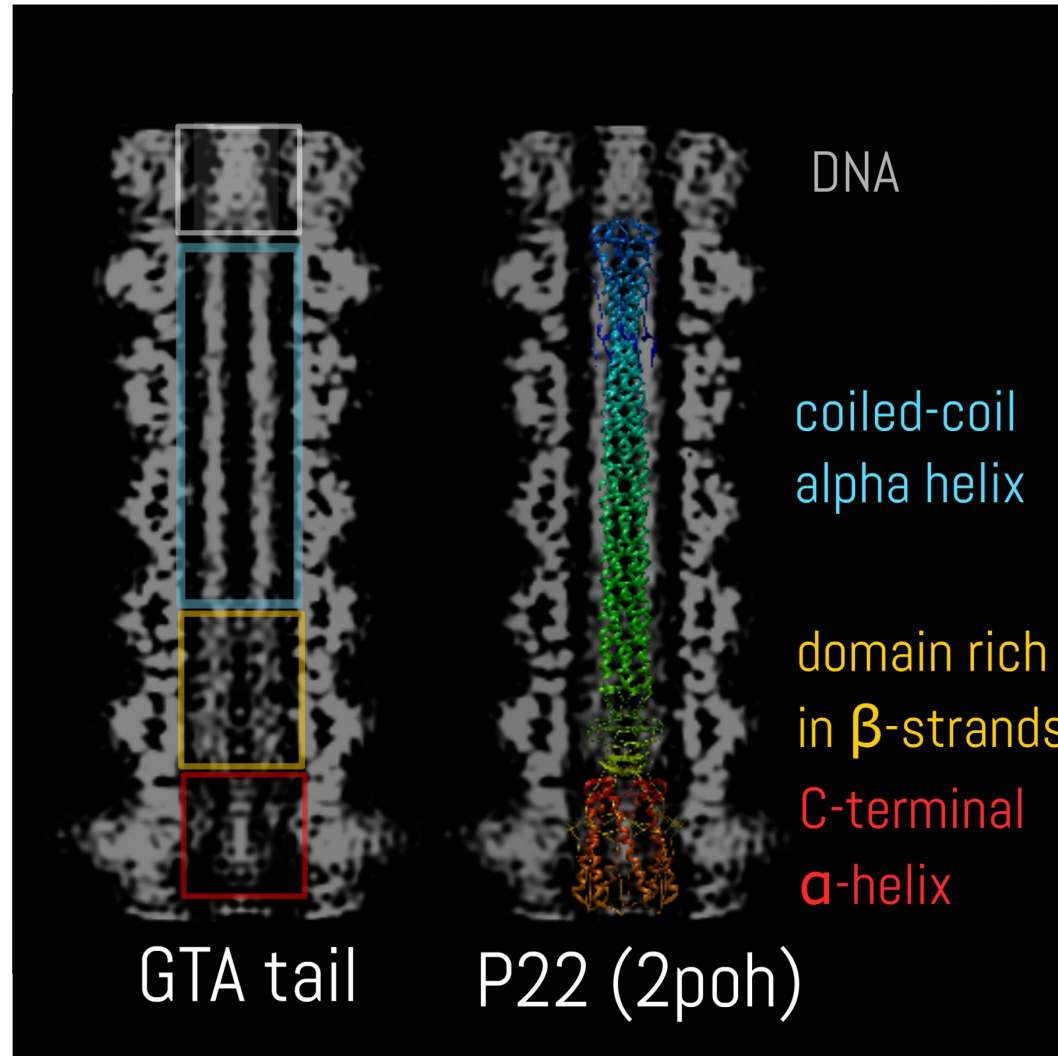
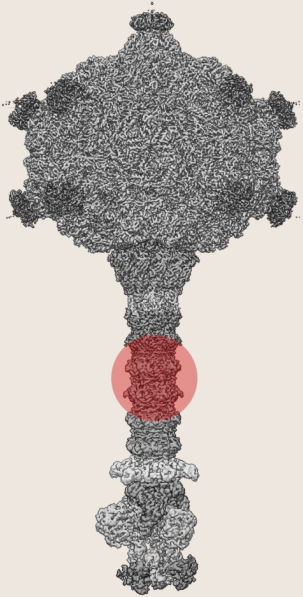


Lipid phage PM2
(2woc)



eukaryotic
dsRNA Simian rotavirus SA11
(4v7q)

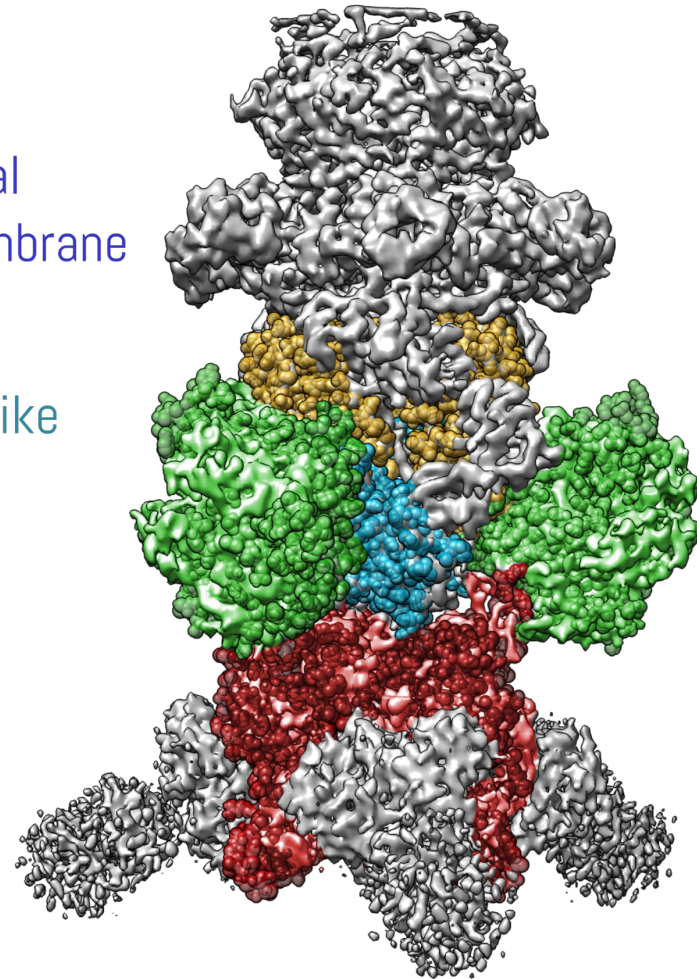
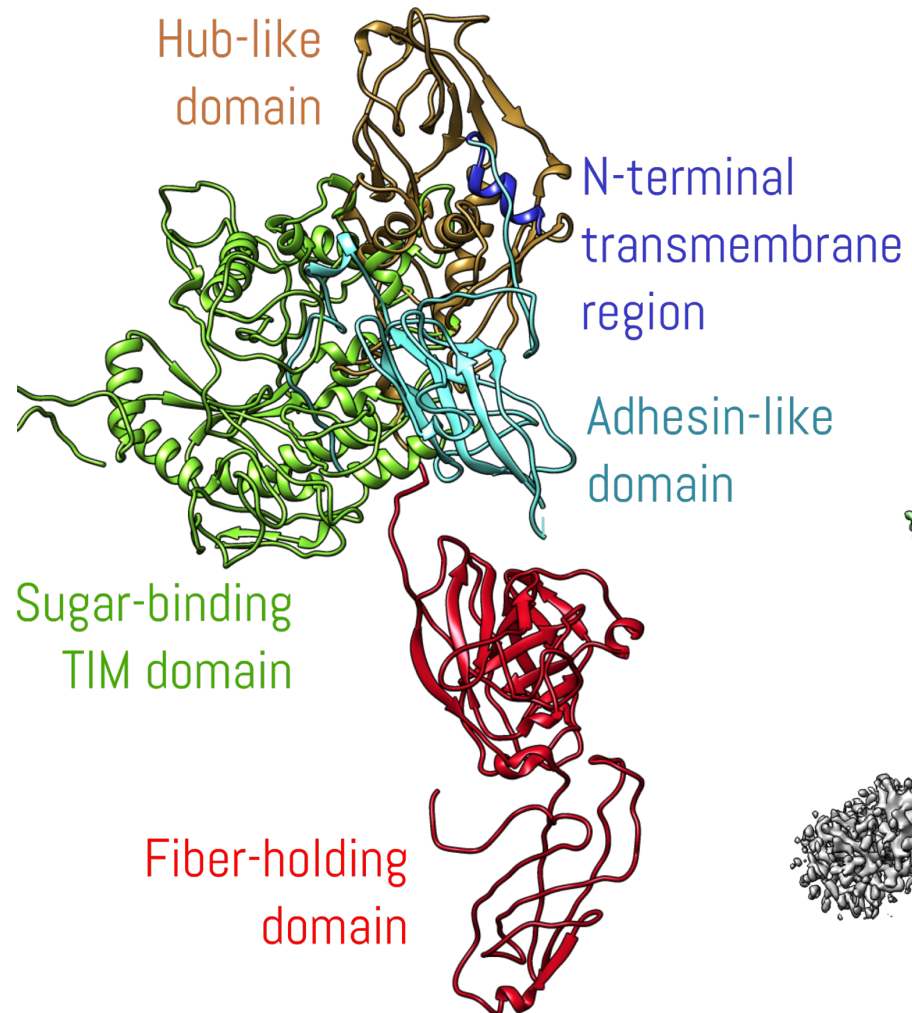
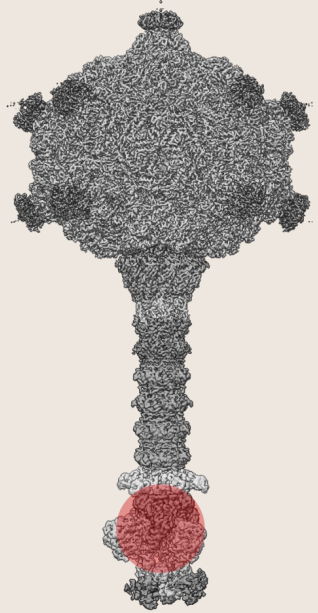
Tape-measure protein of the GTA



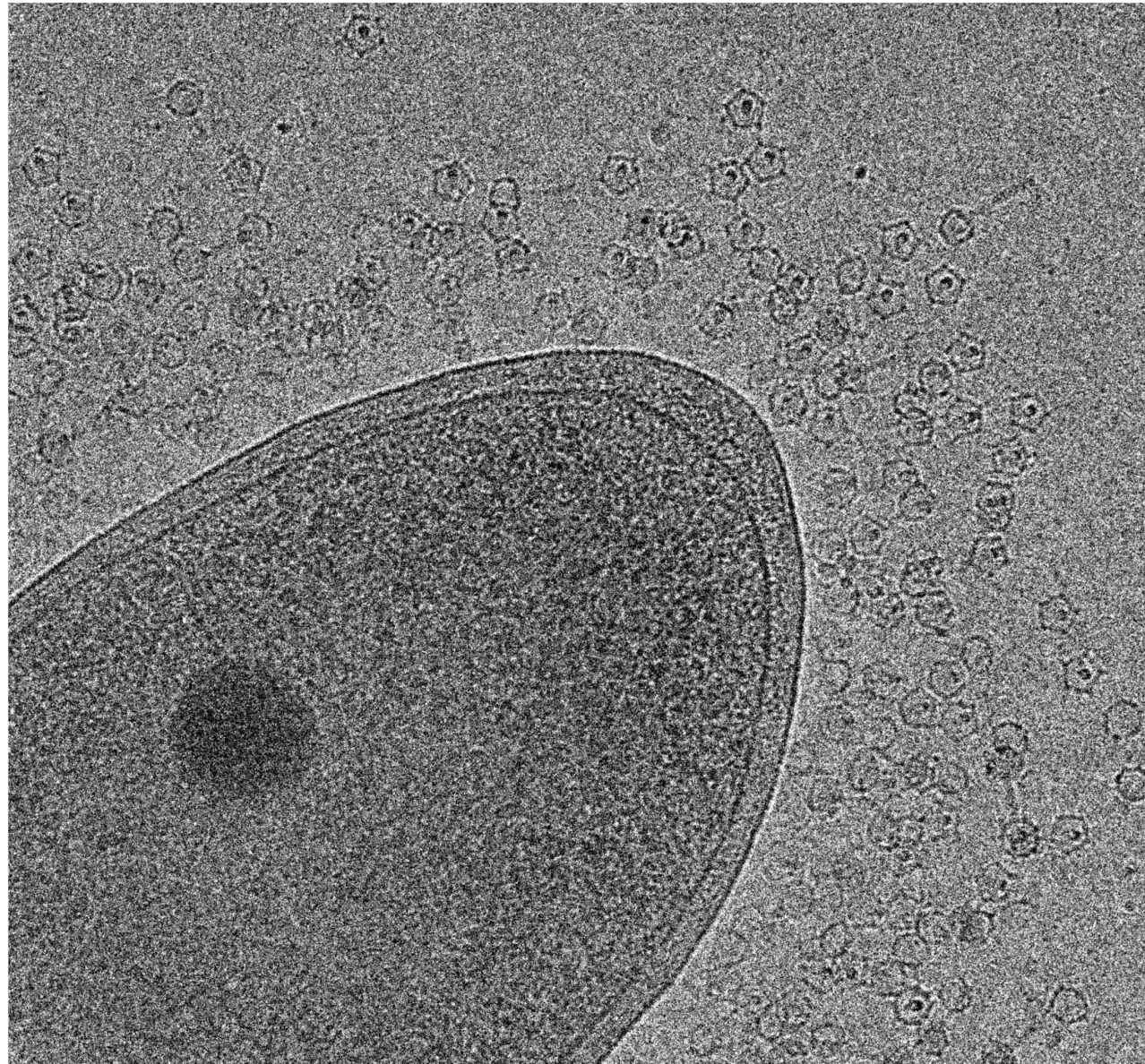
RESULTS

MEGATRON protein

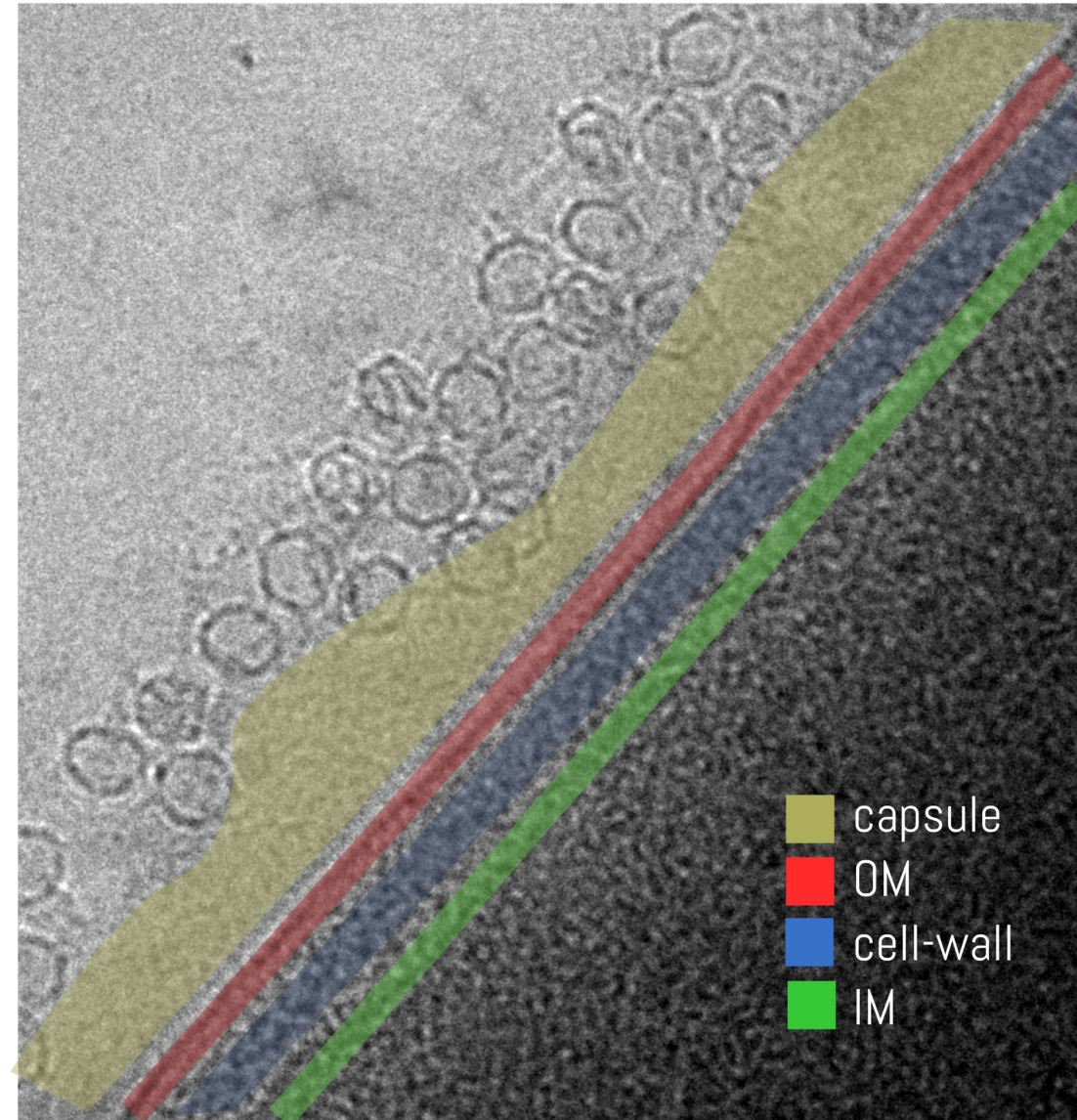
1  1303



DNA-delivery mechanism of the GTA

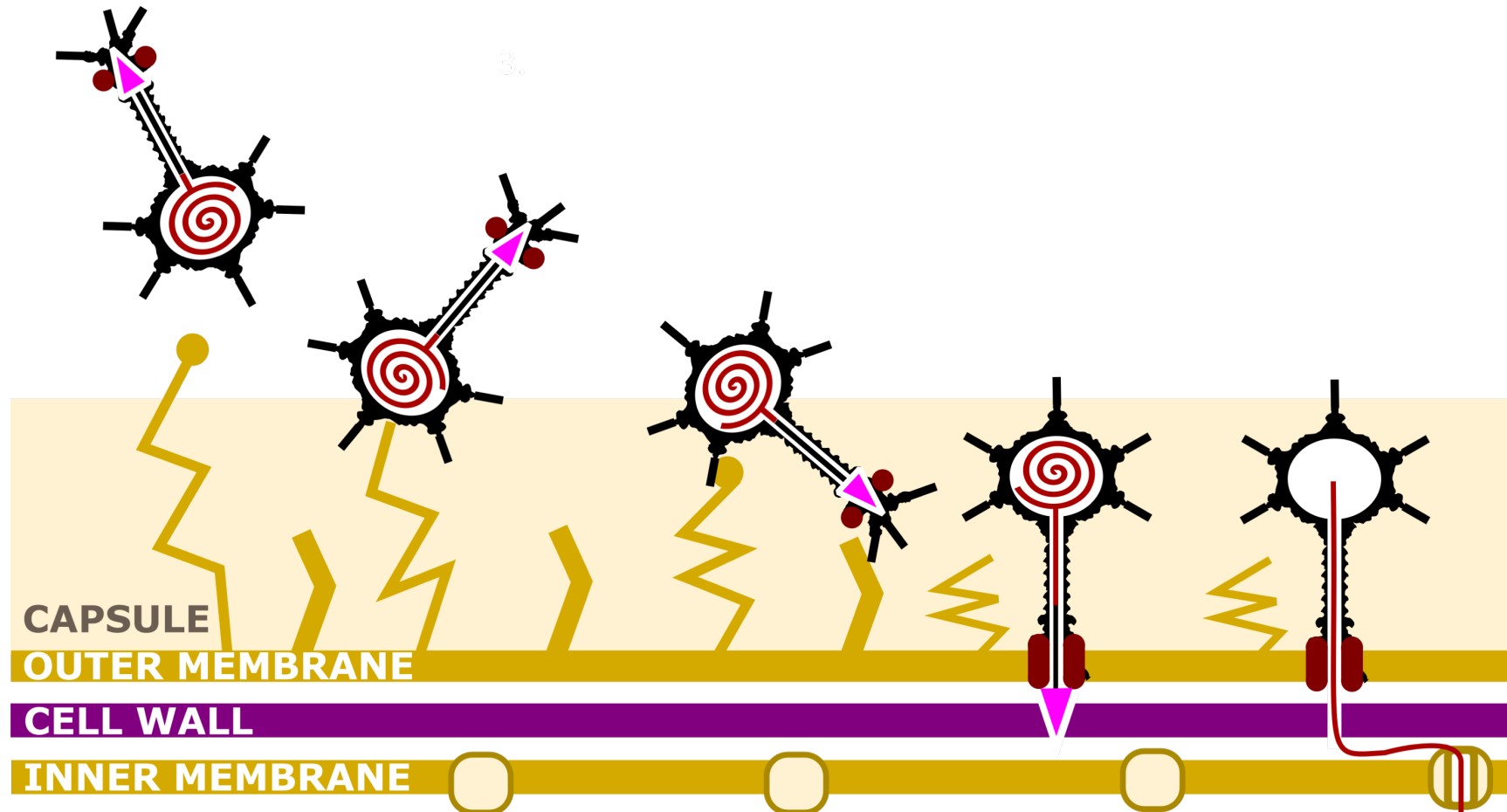


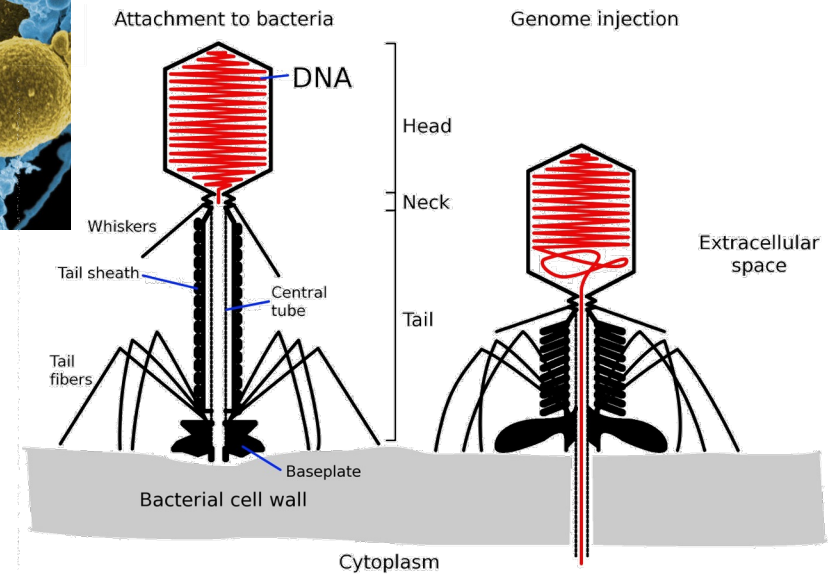
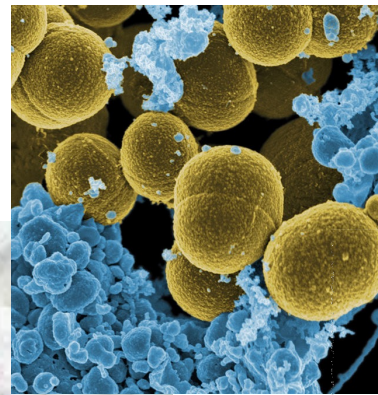
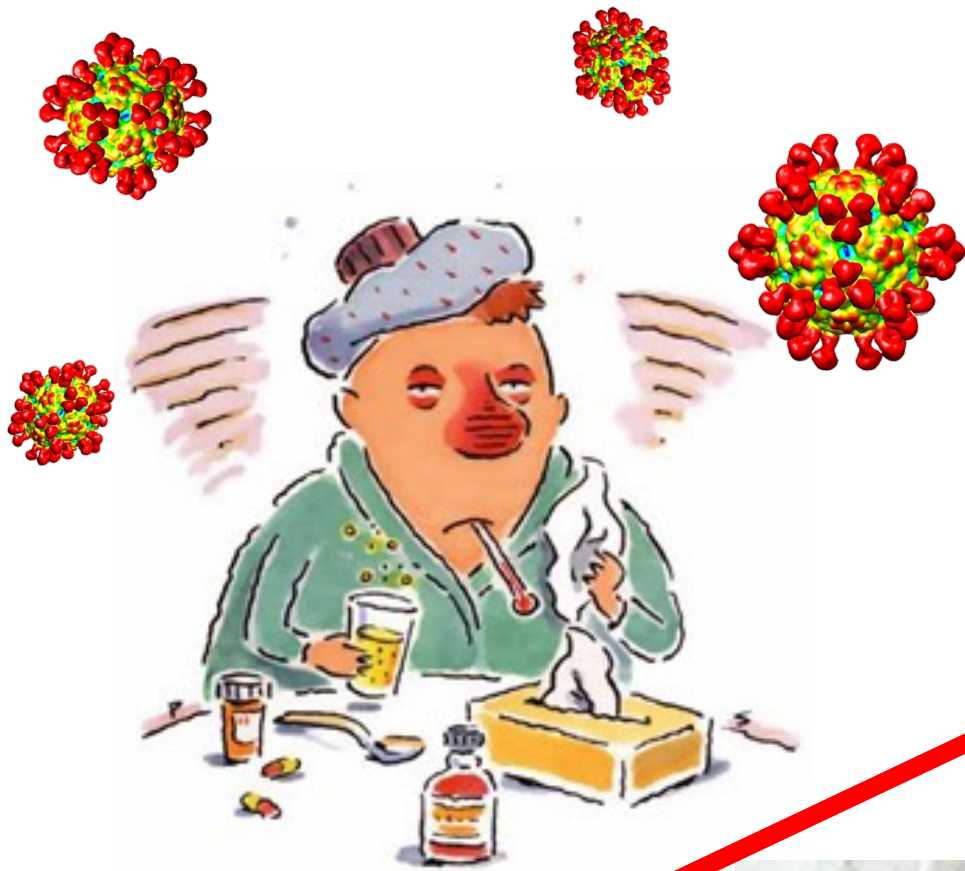
DNA-delivery mechanism of the GTA



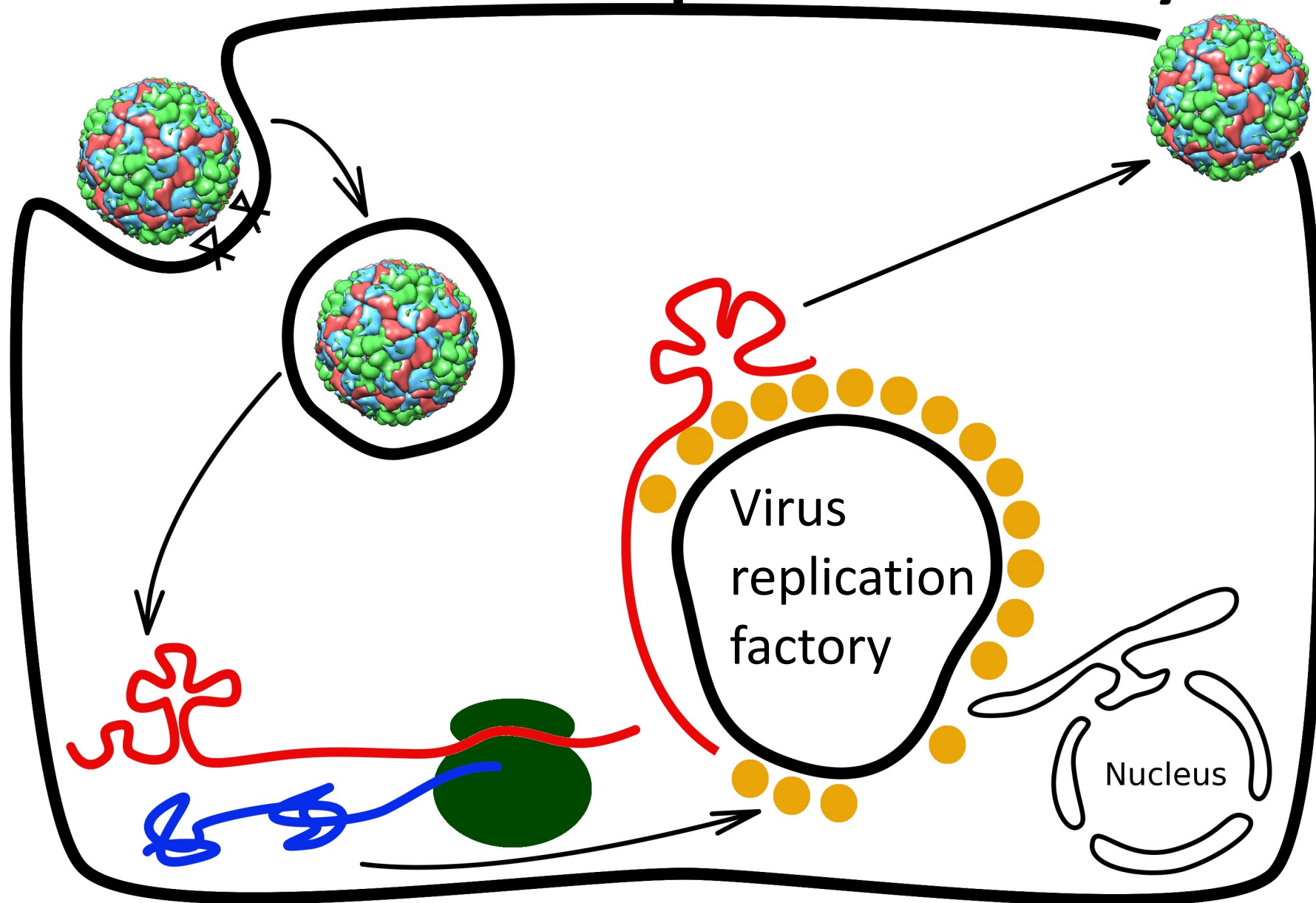
RESULTS

DNA-delivery mechanism of the GTA

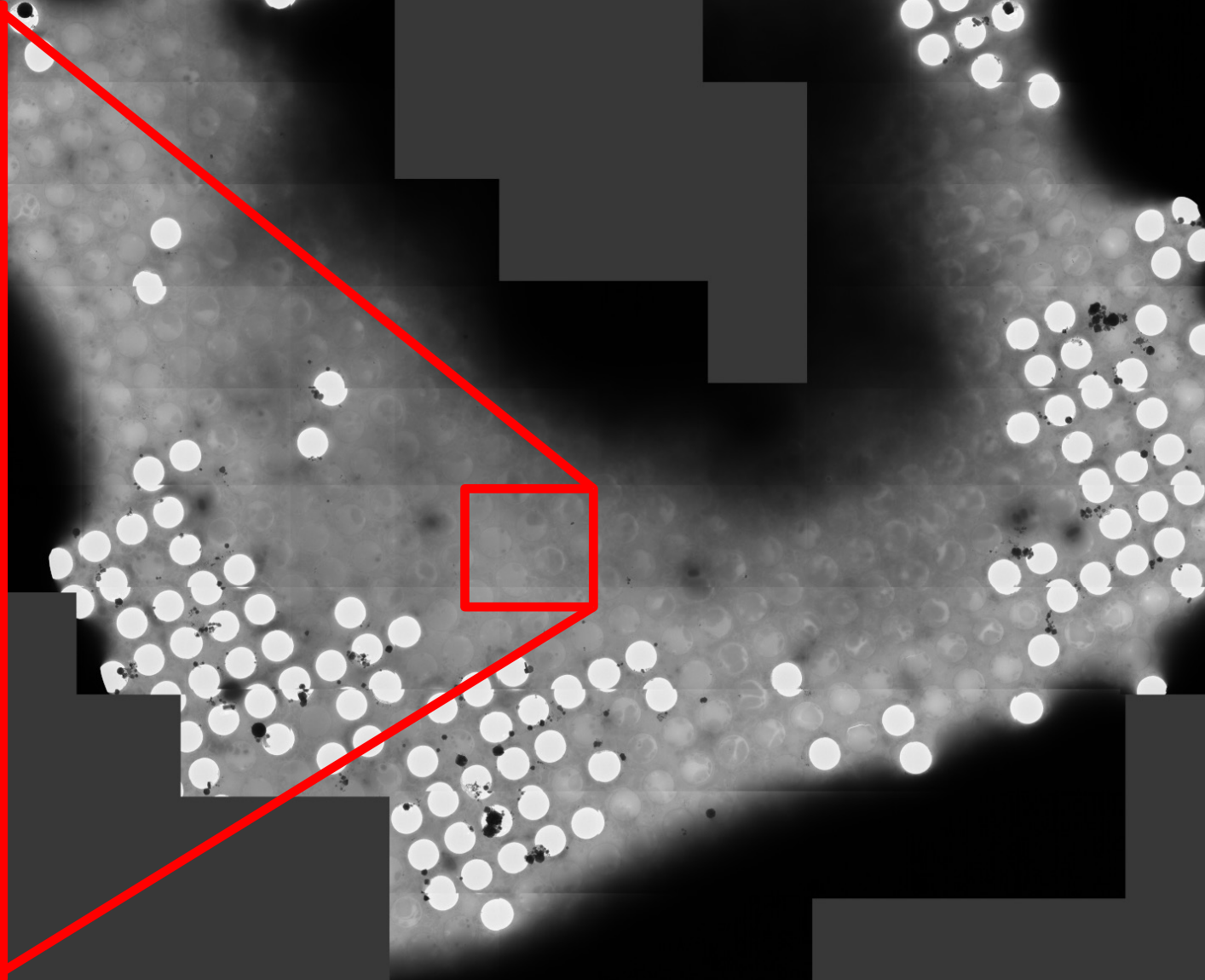
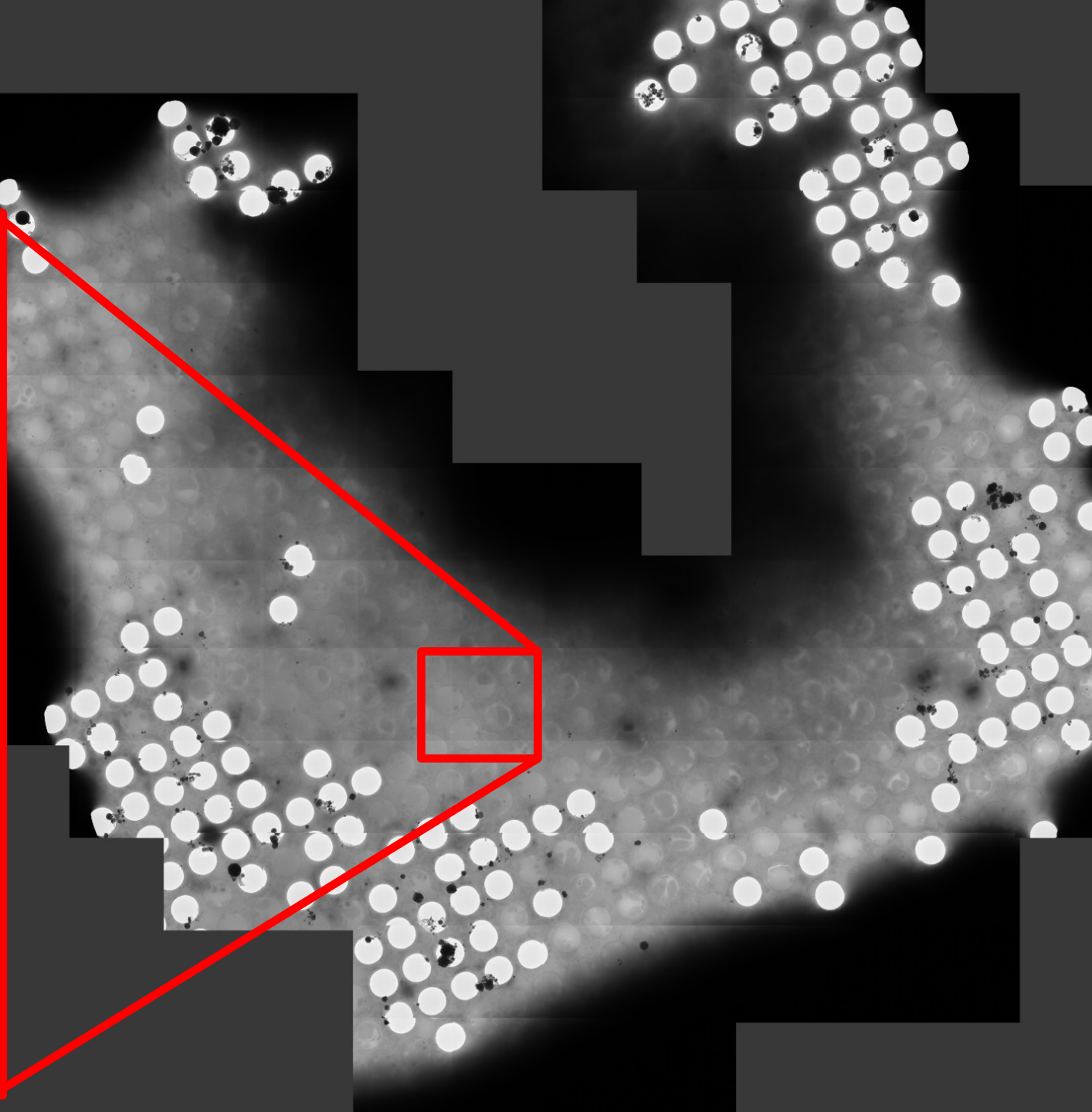




Enterovirus replication cycle

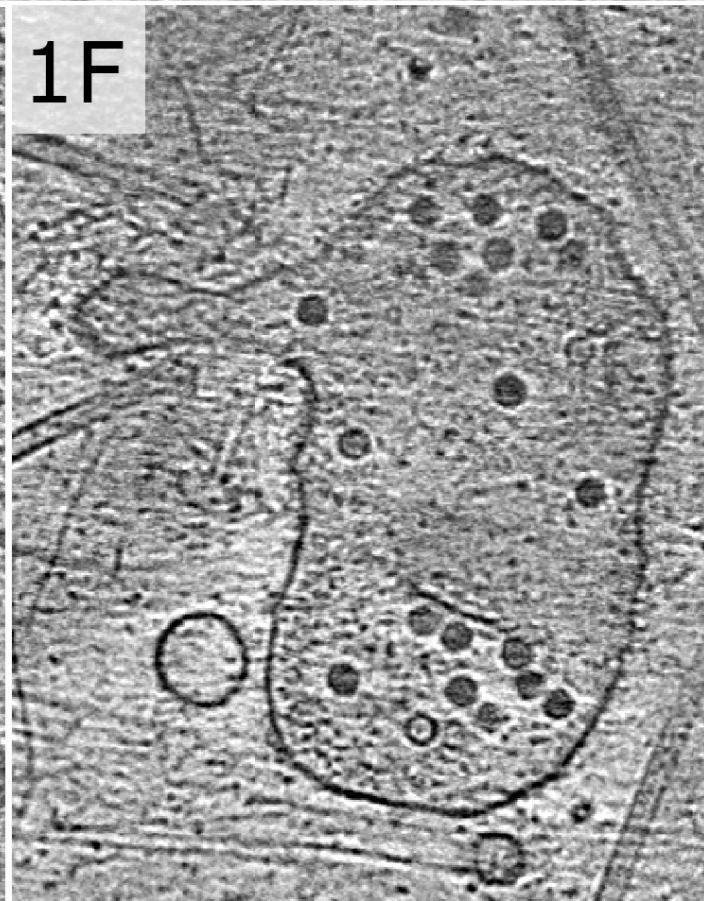
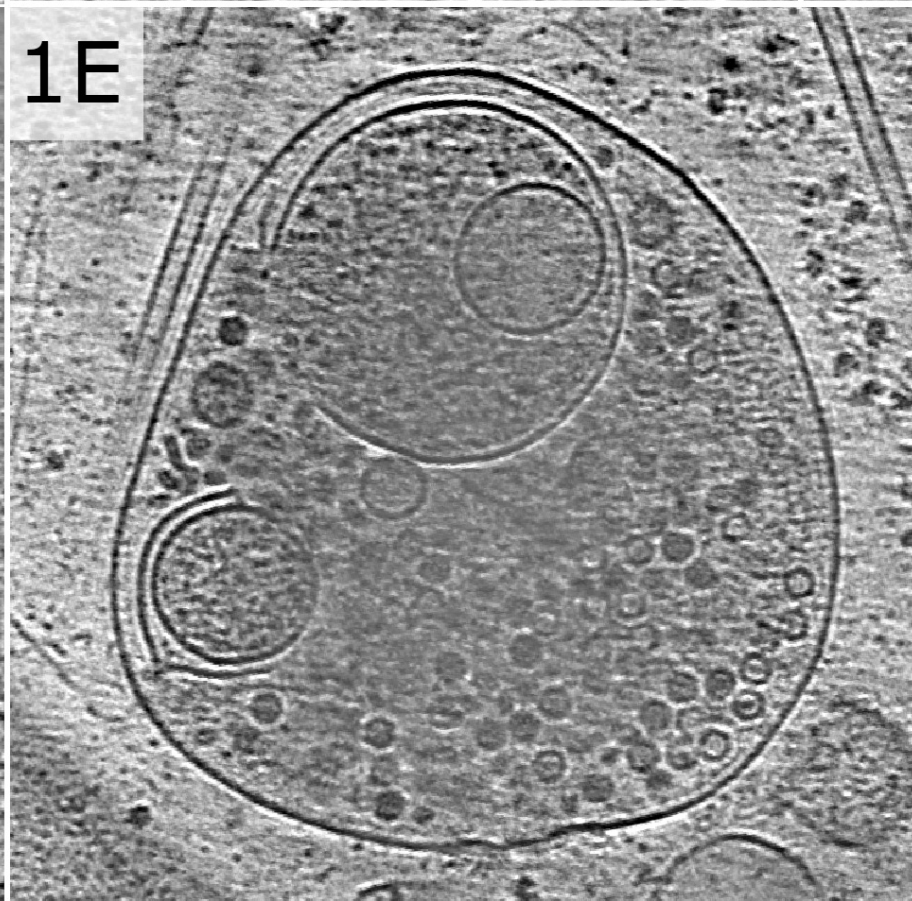
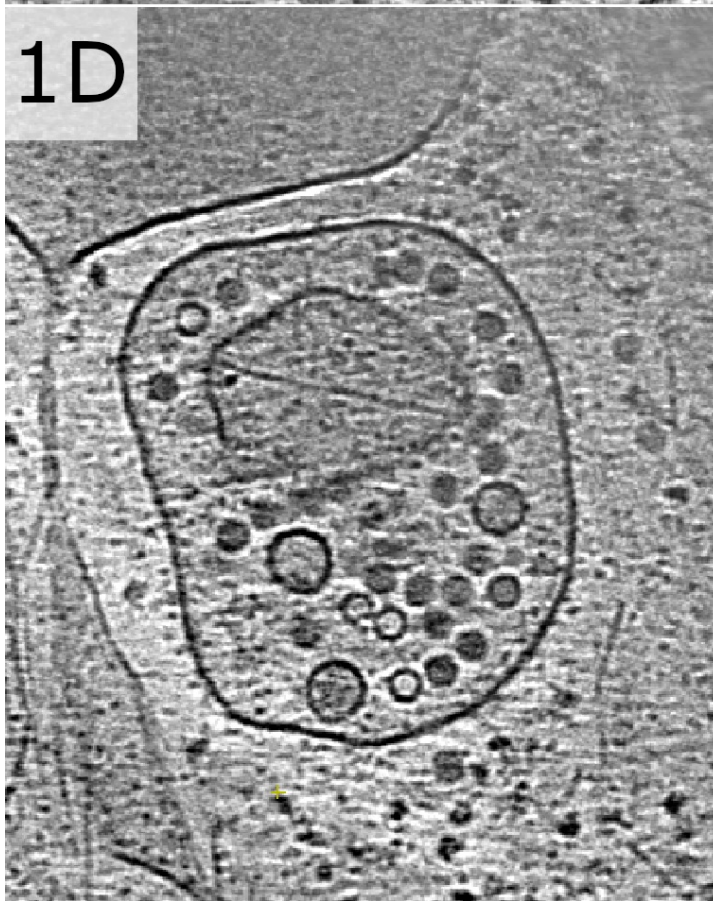
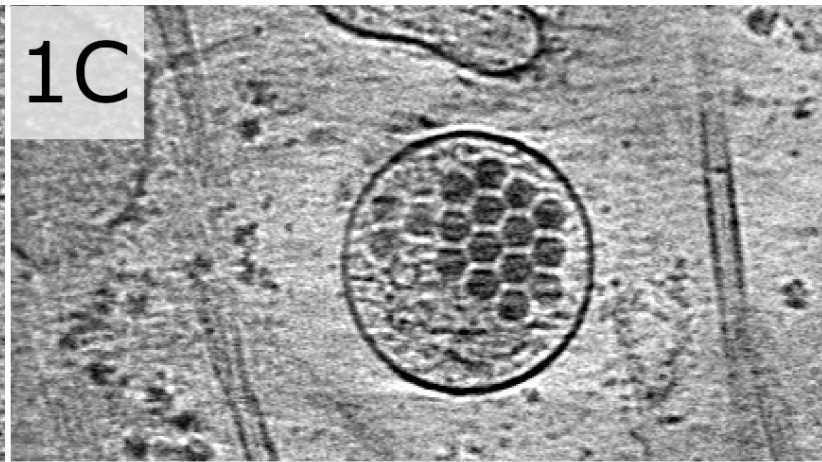
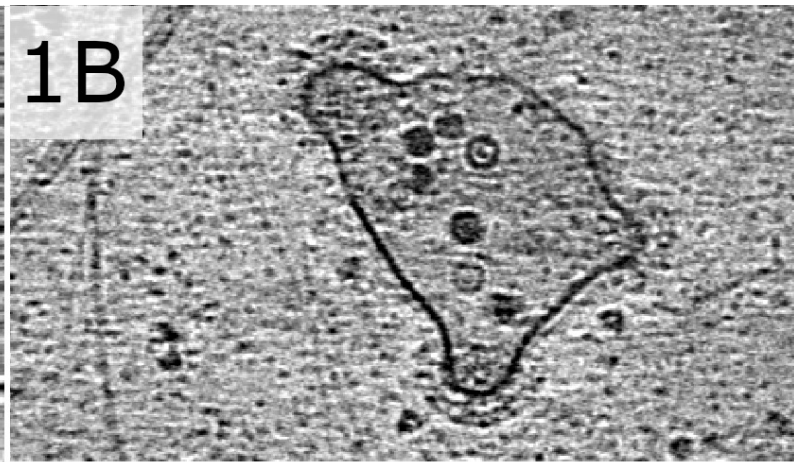
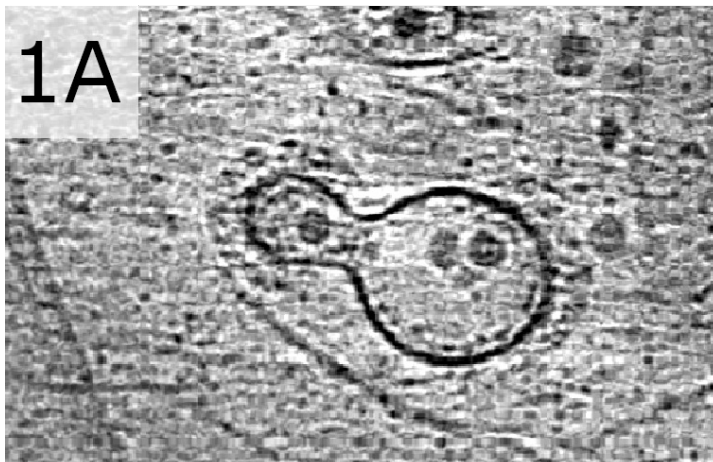


in situ cryo-electron
tomography

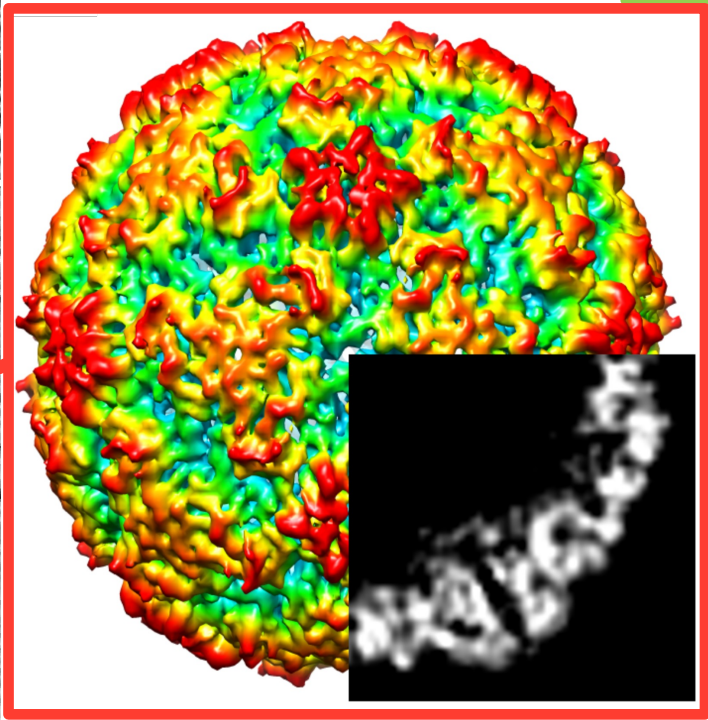
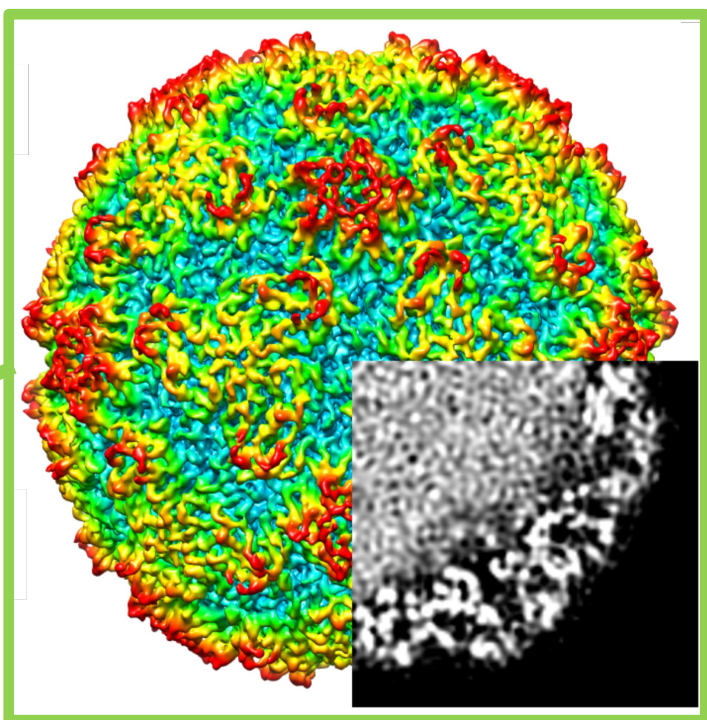
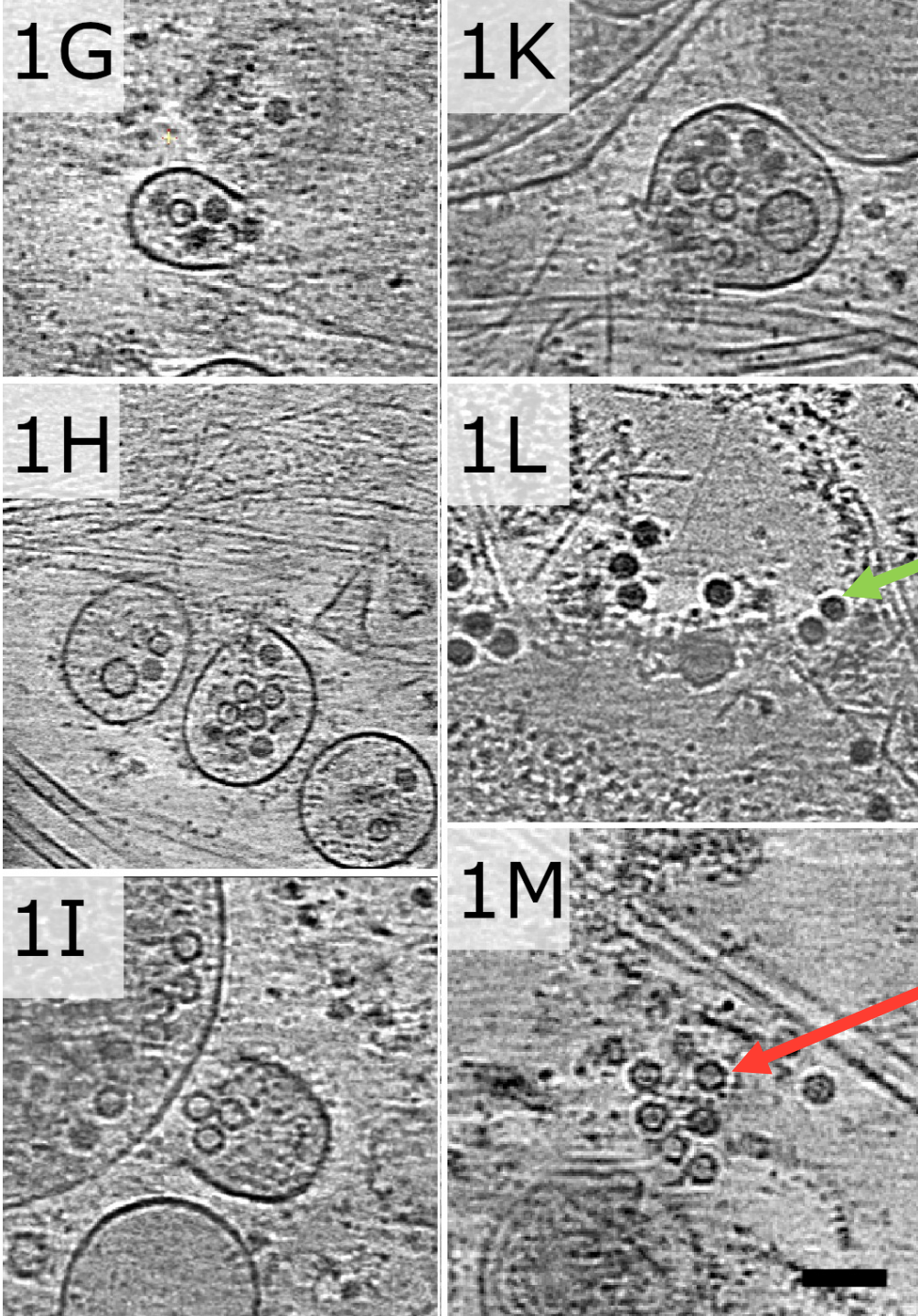




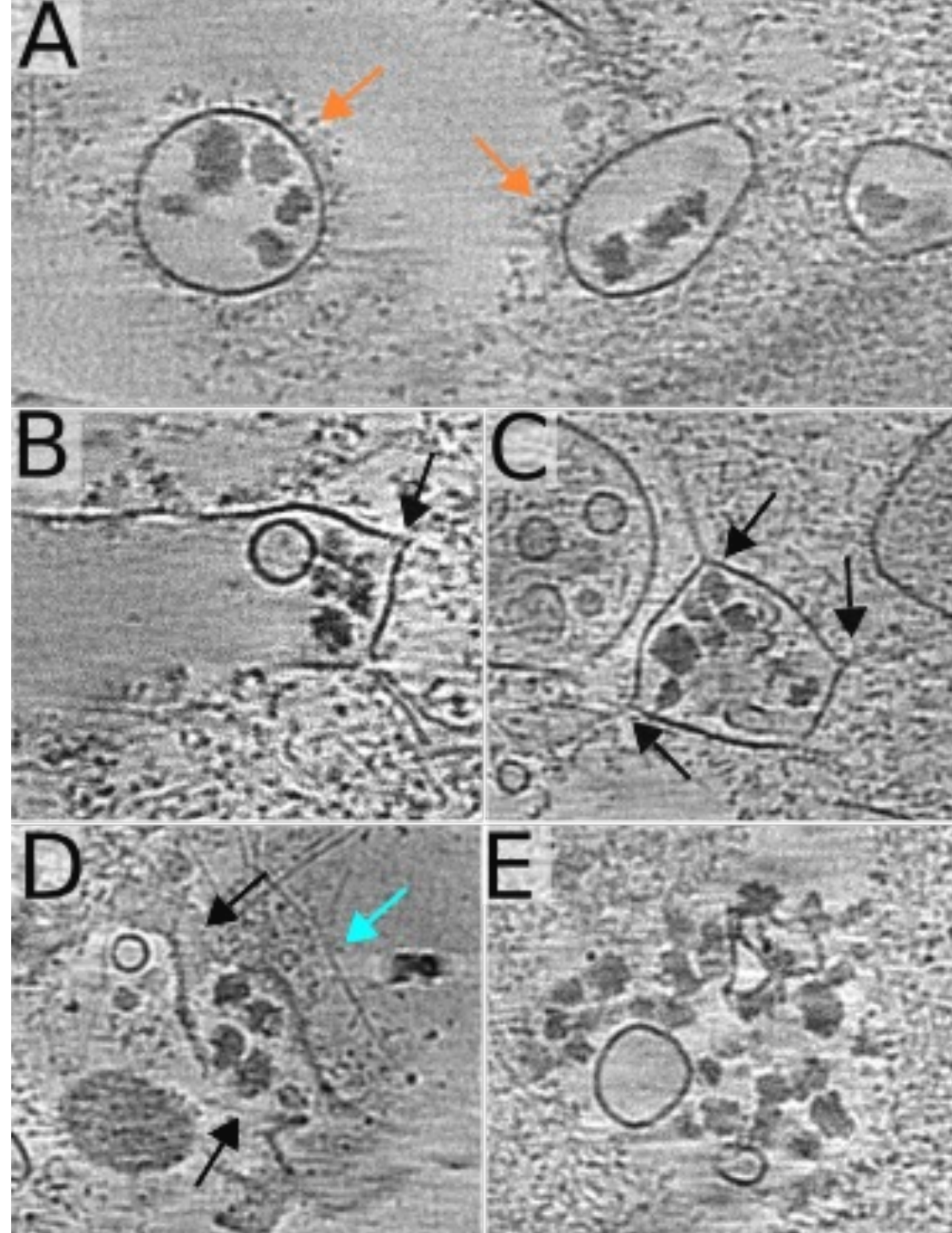
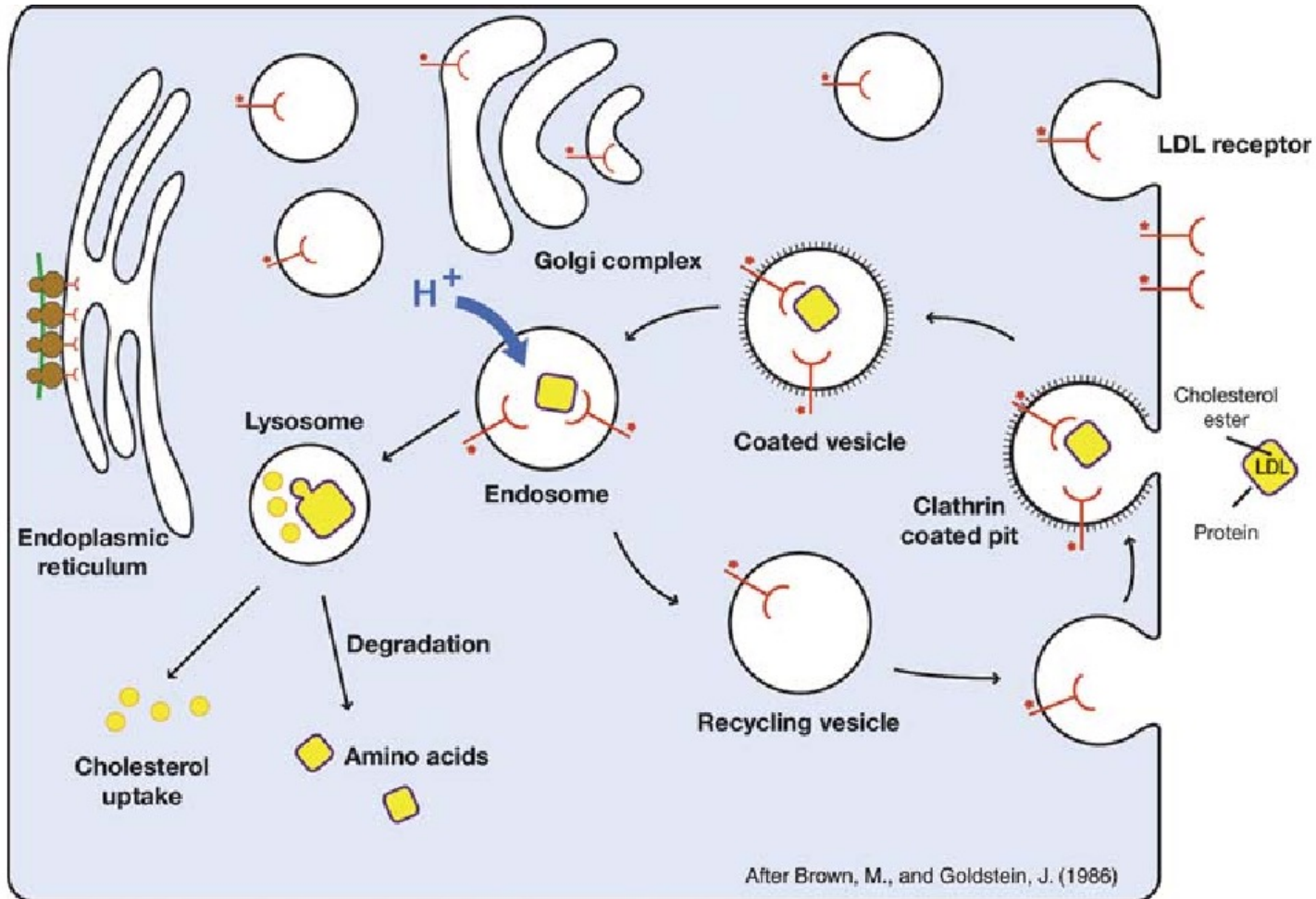
Cell entry of rhinovirus 2



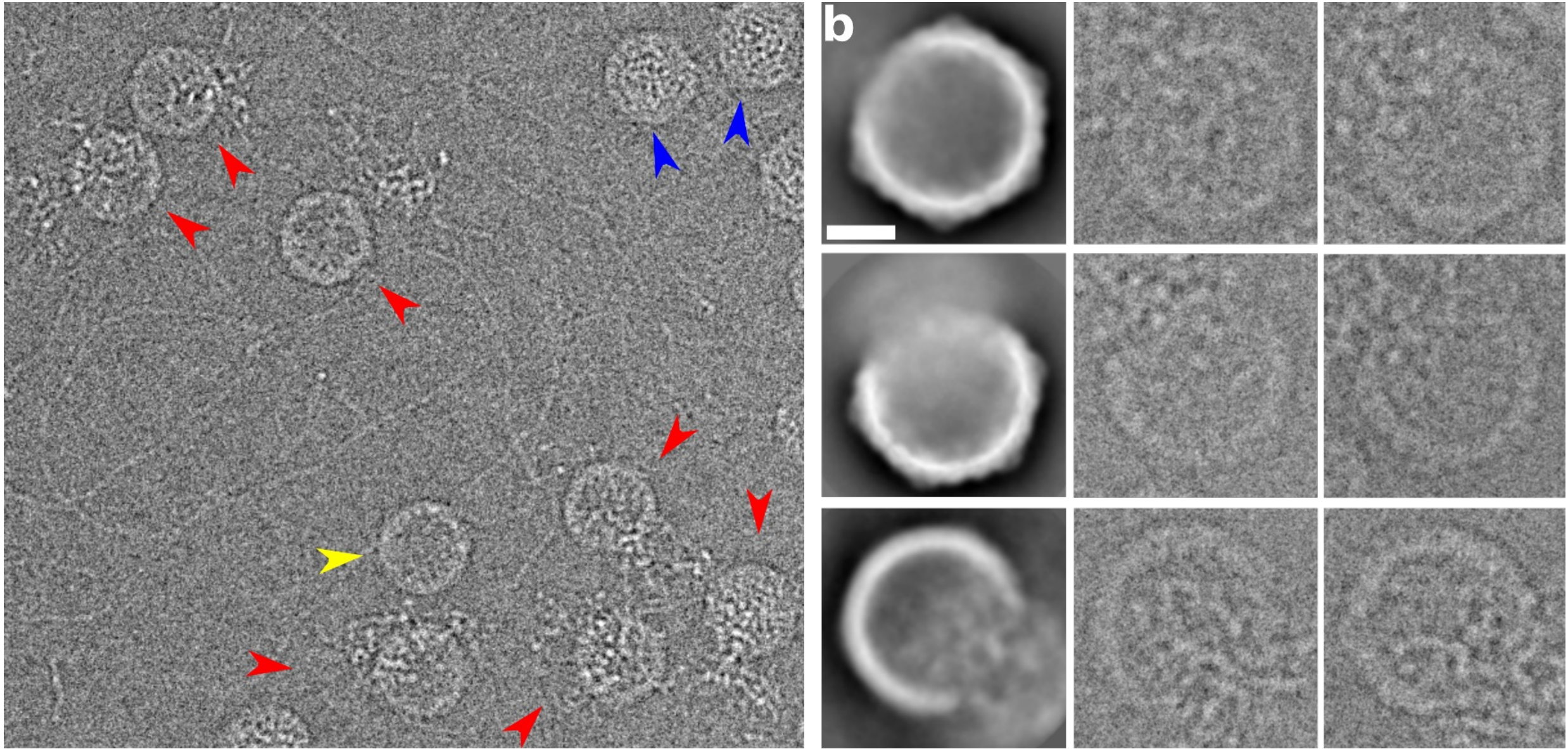
Cell entry of rhinovirus 2



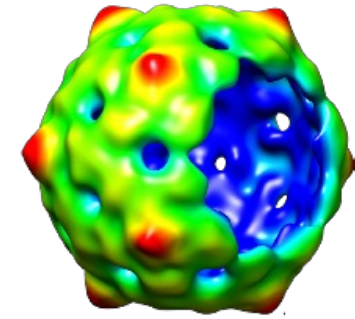
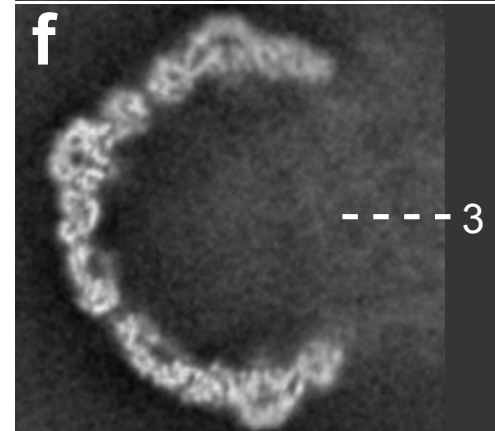
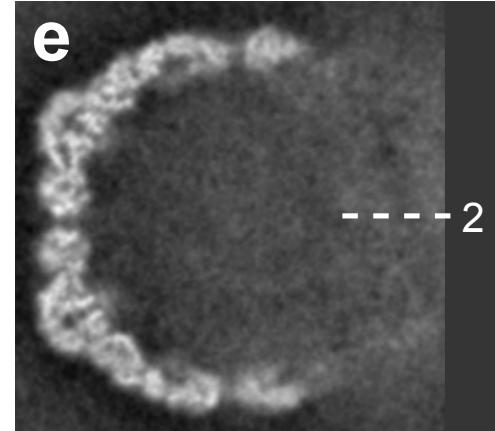
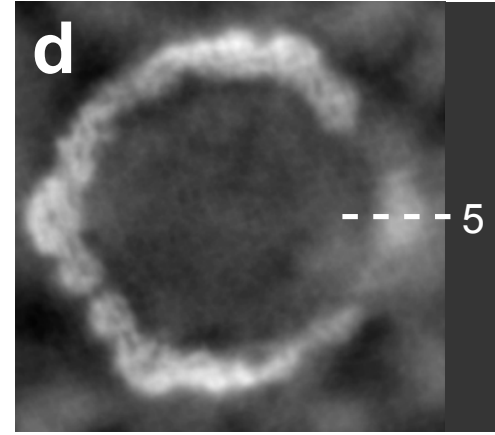
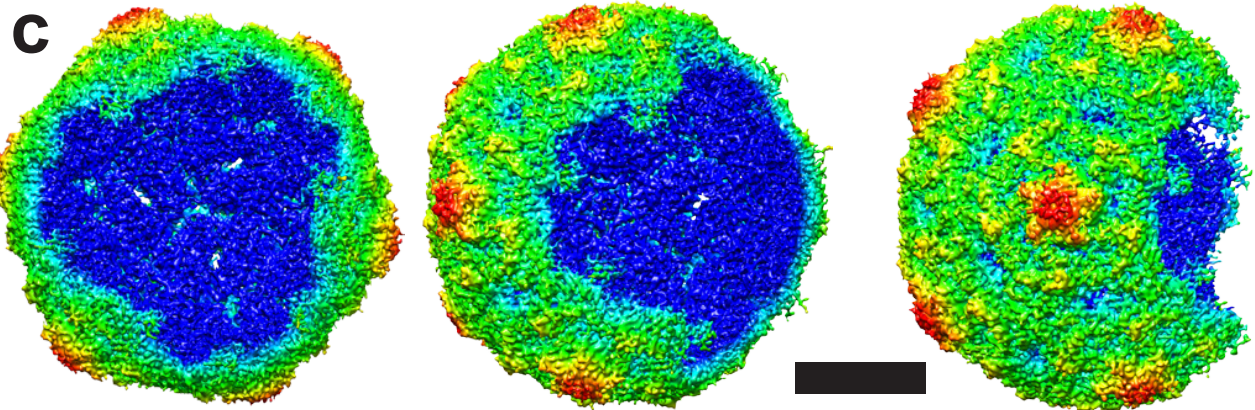
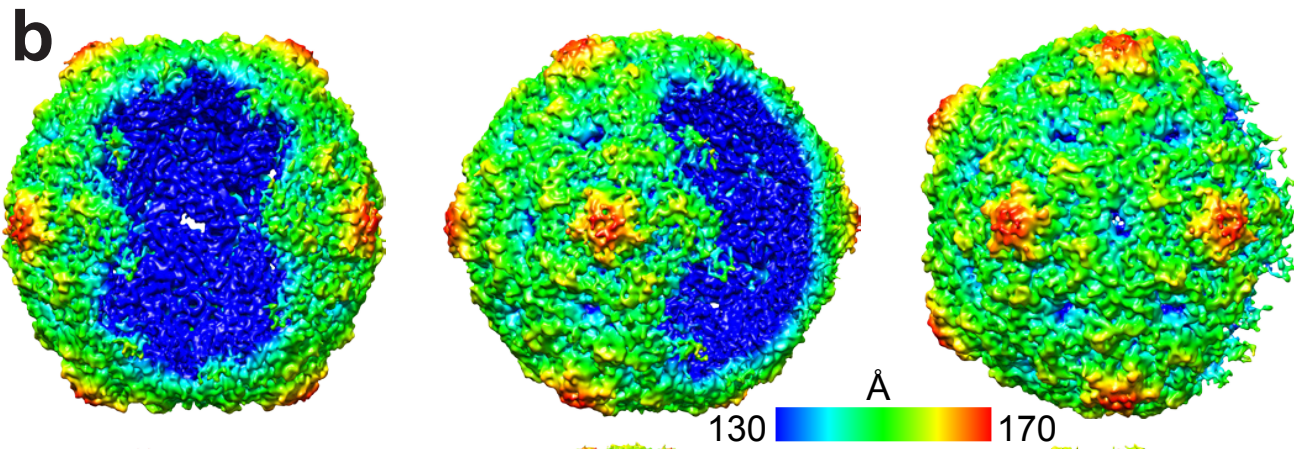
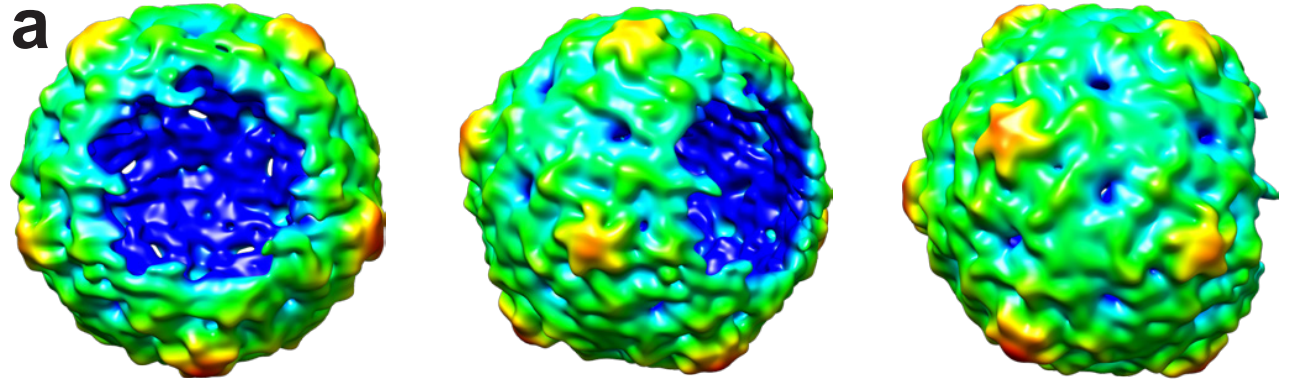
Endocytosis of VLDL - native cargo of VLDLR



Genome release intermediates of echovirus 18



Open particles of echovirus 18

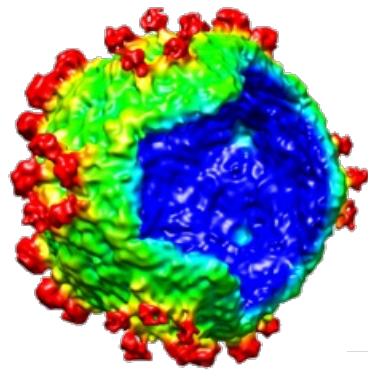
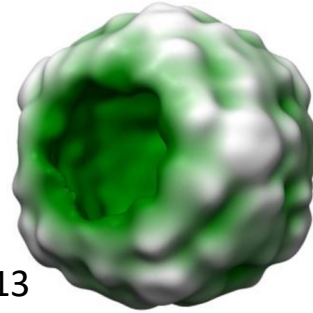


Echovirus 30
Picornaviridae

Buchta et al. 2019

Human
rhinovirus 2
Picornaviridae

Harutyunyan et al. 2013

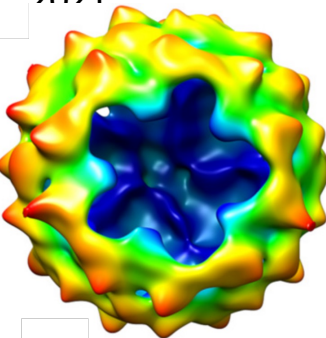


Slow bee
paralysis virus
Iflaviridae

Škubník et al.
2021

Kashmir bee virus
Dicistroviridae

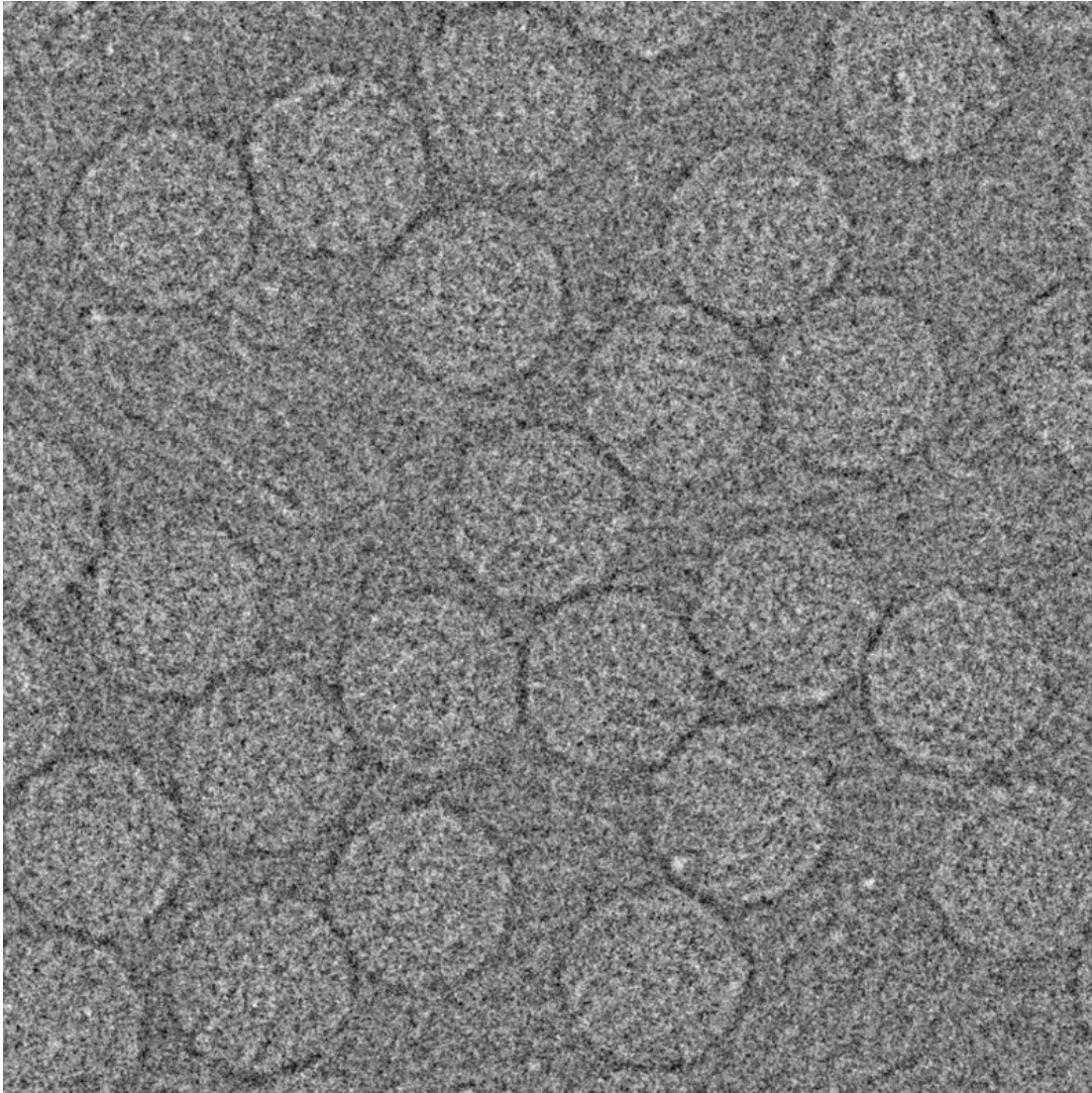
Mukhamedova et al.
2021



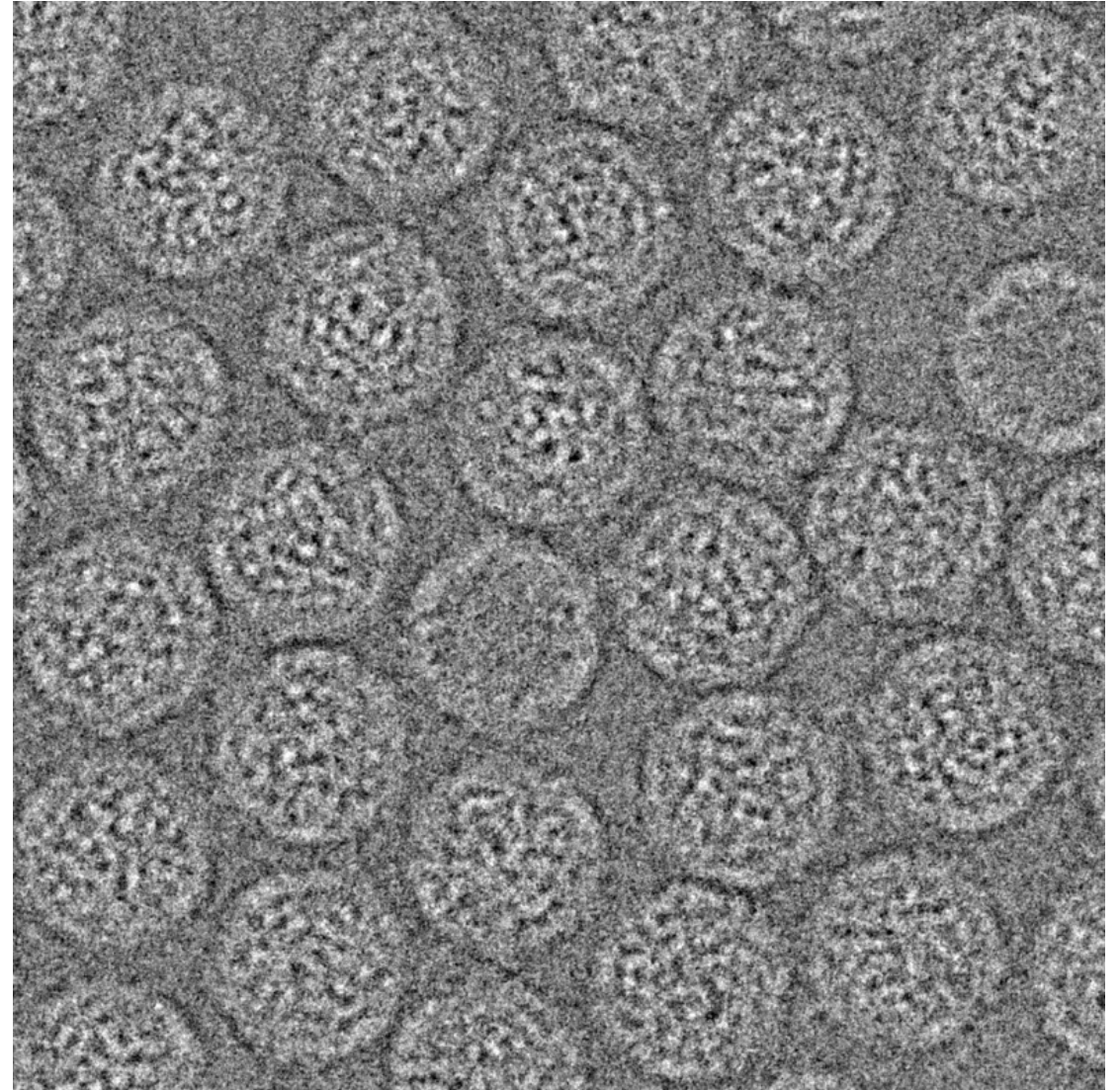
Buchta et al. 2019

Acidic pH induces genome reorganization

Virions at neutral pH



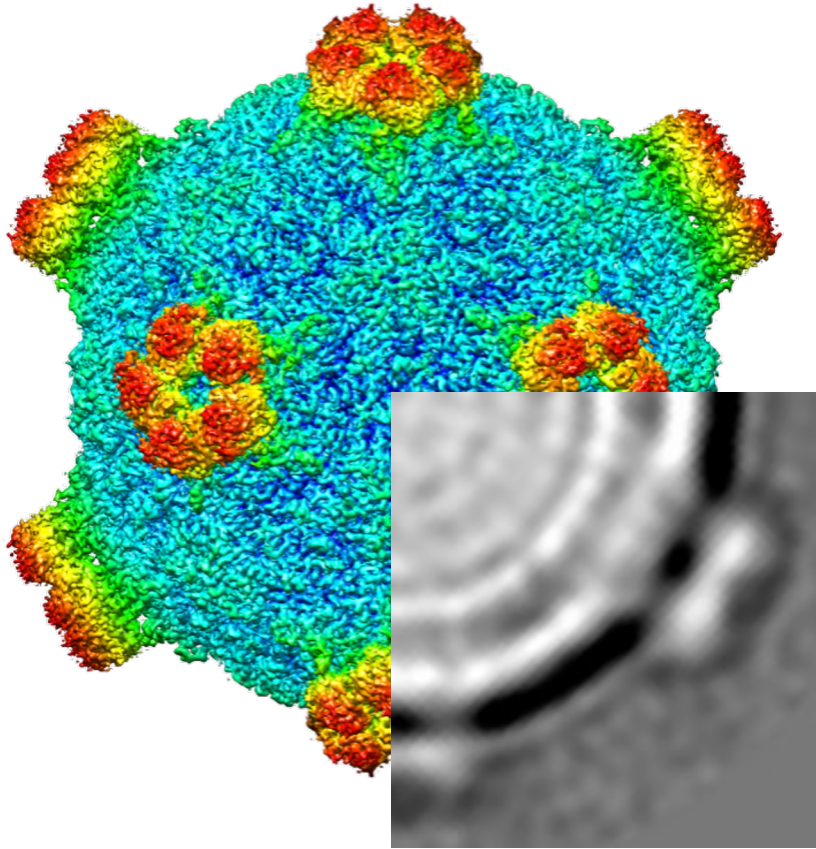
Activated particles at acidic pH



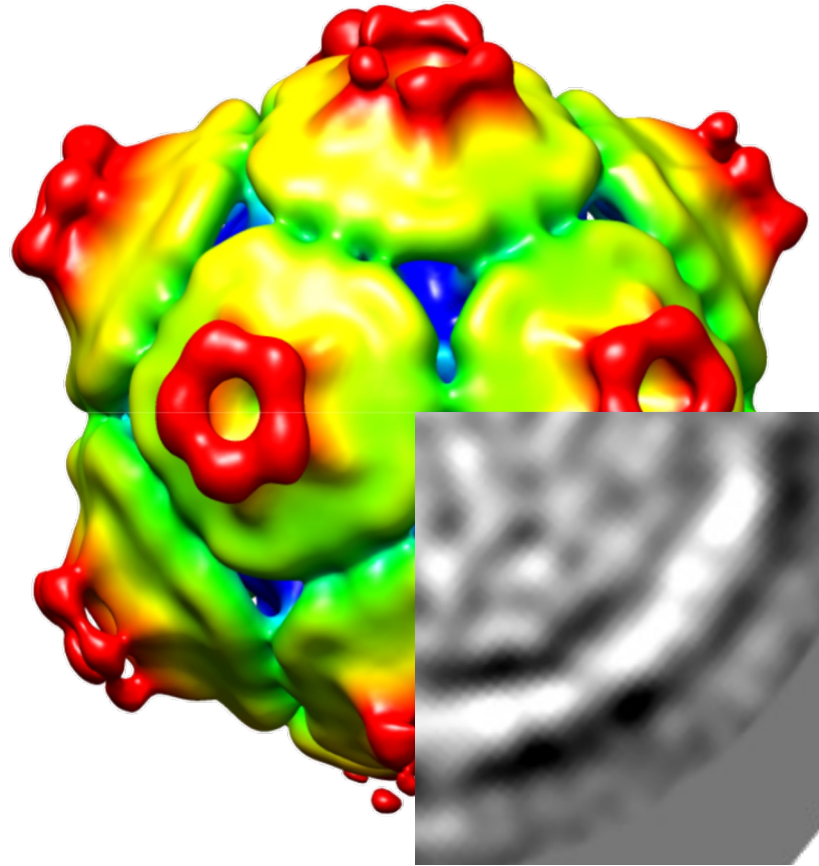
Deformed wing virus (*Iflaviridae*)

Neutral pH

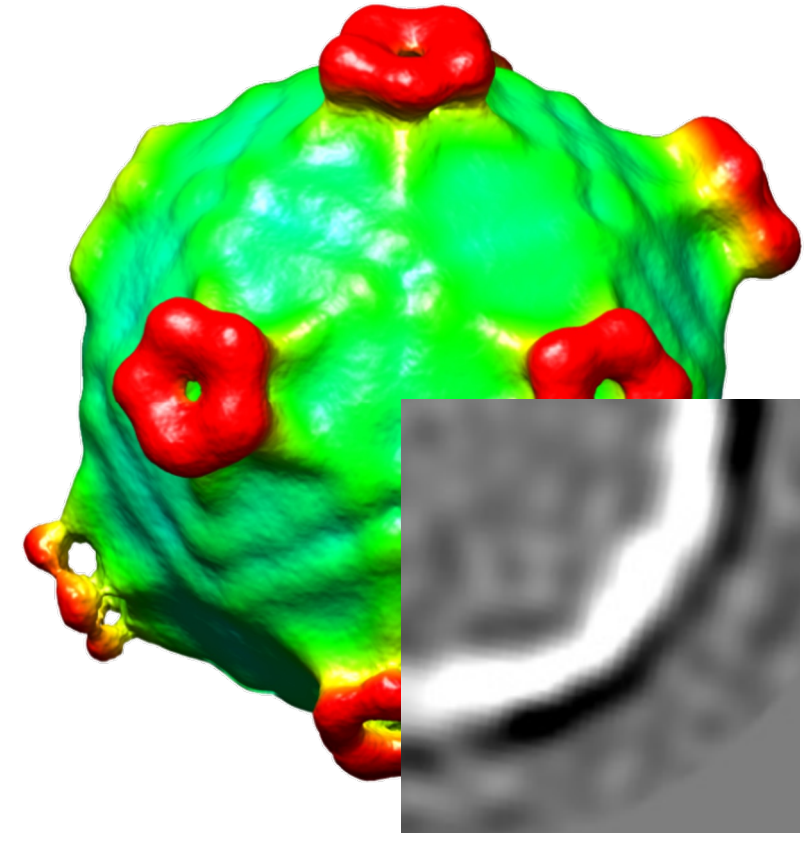
Acidic pH



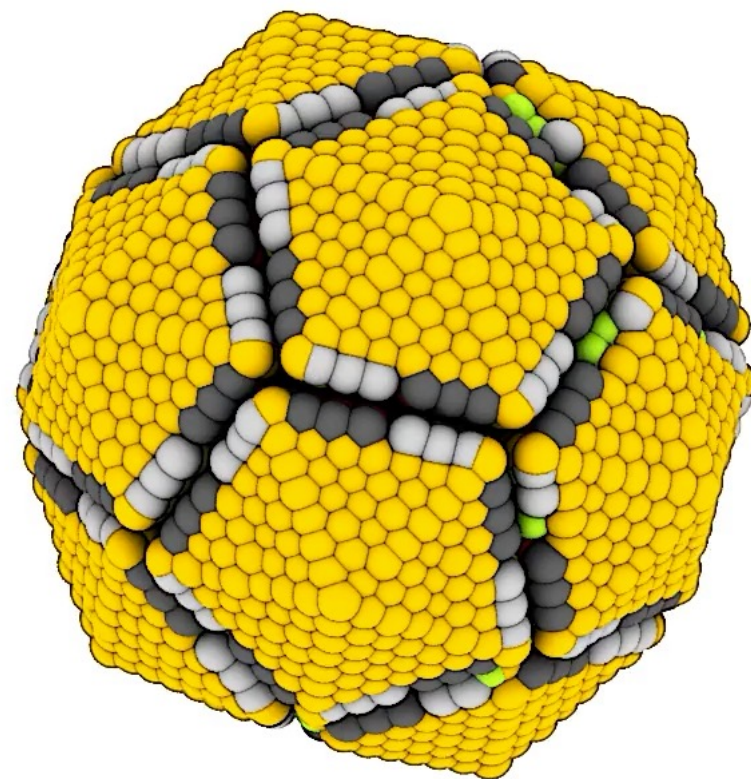
Virion



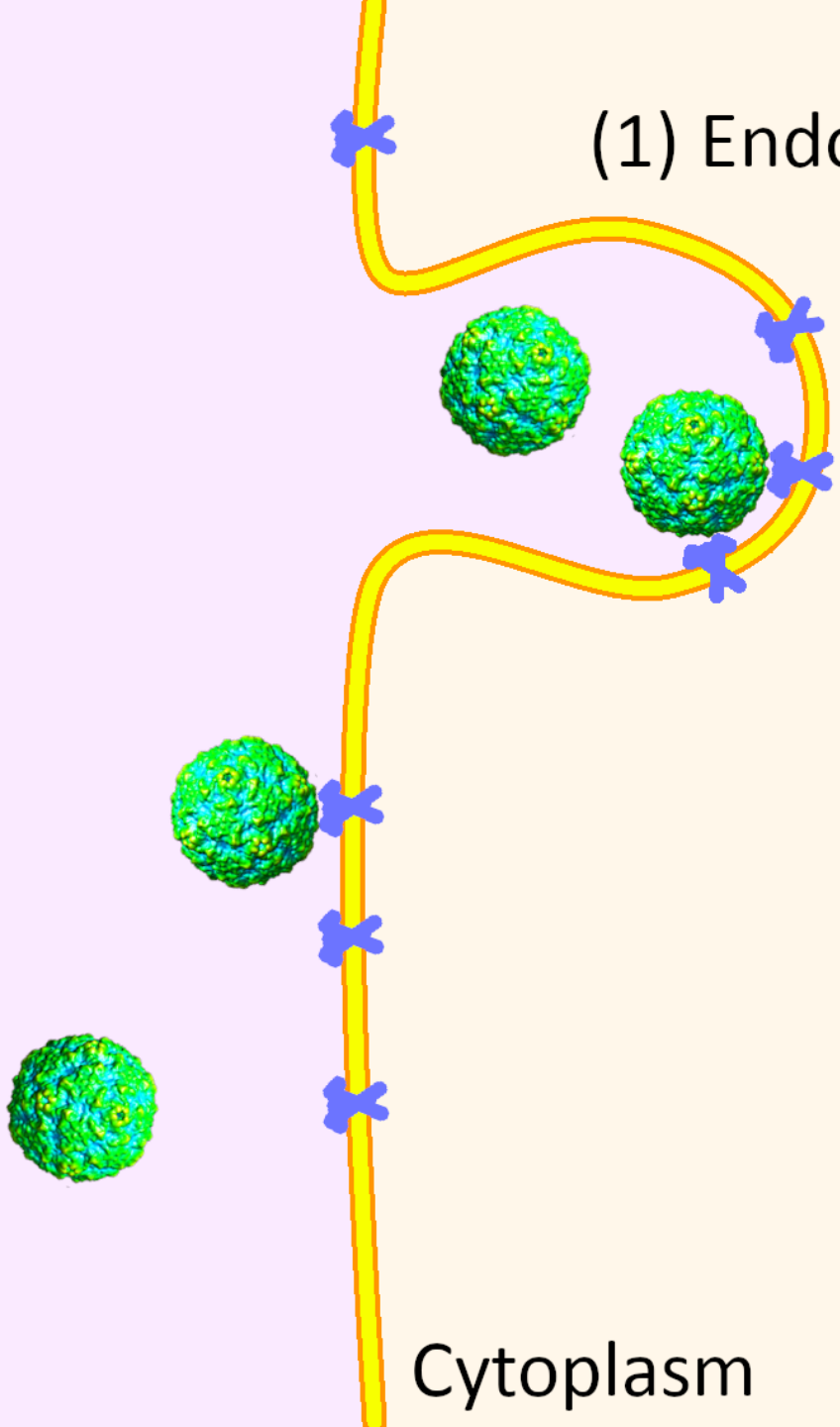
Full particle



Empty particle



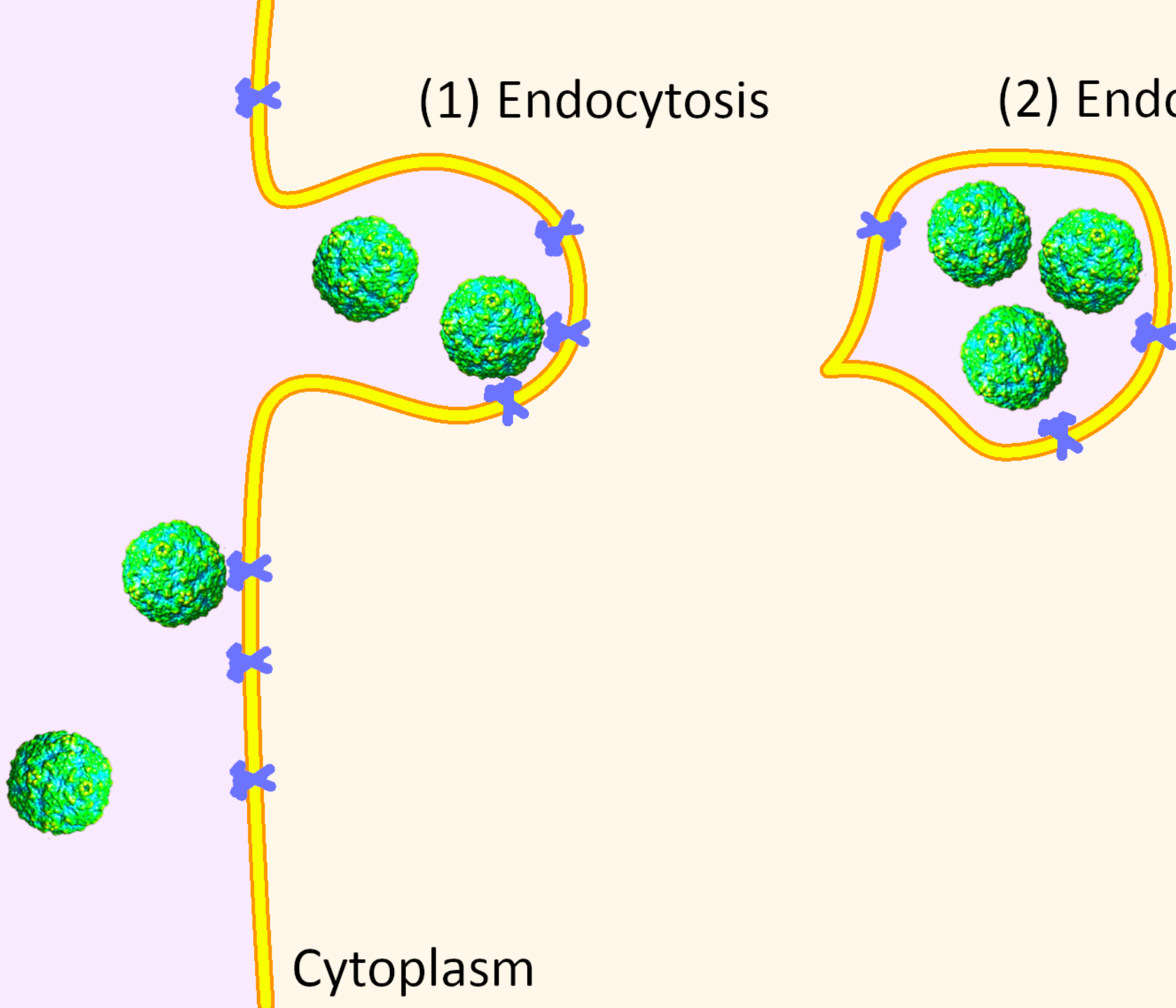
(1) Endocytosis



Cytoplasm

(1) Endocytosis

(2) Endosome

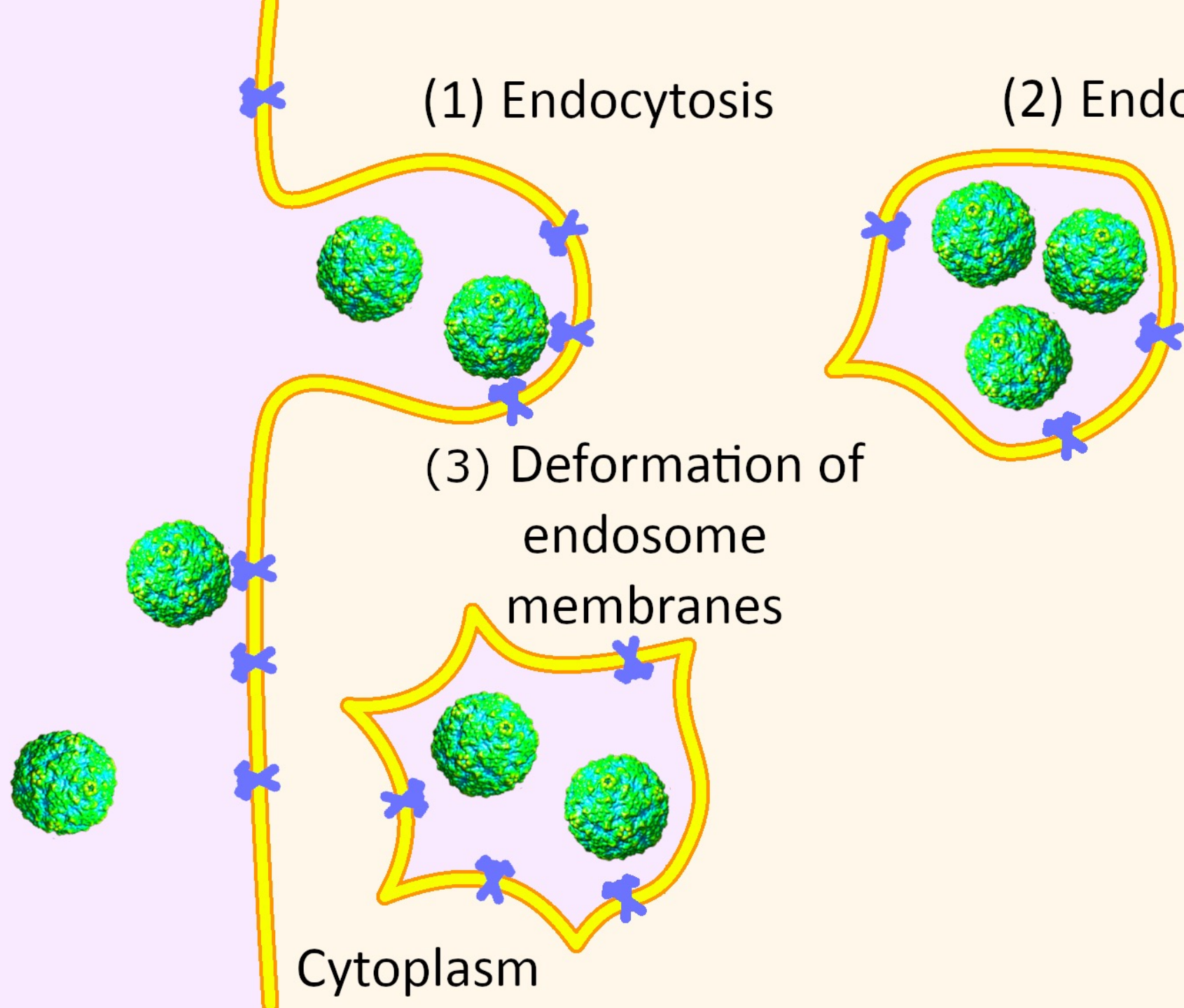


(1) Endocytosis

(2) Endosome

(3) Deformation of endosome membranes

Cytoplasm



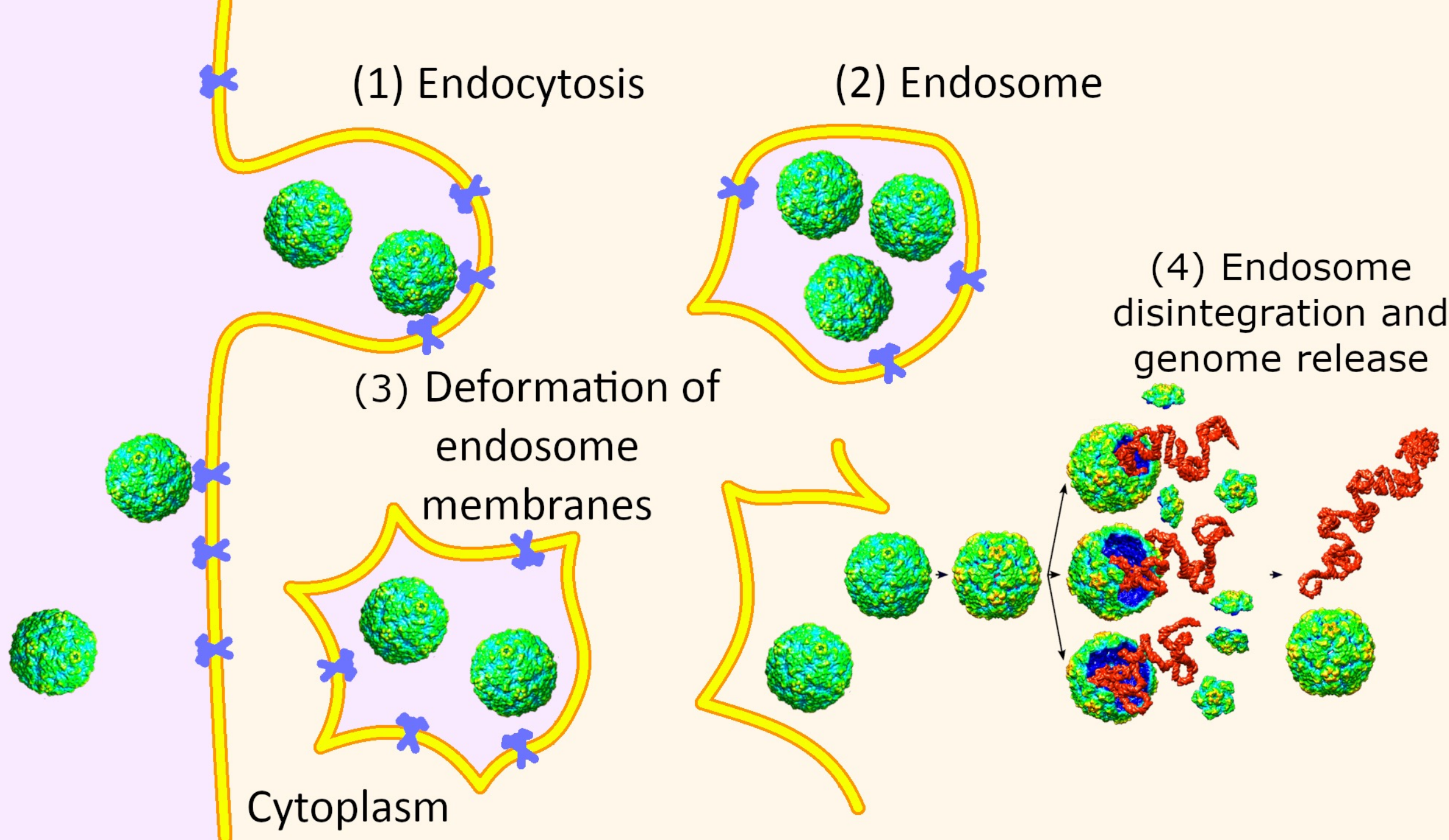
(1) Endocytosis

(2) Endosome

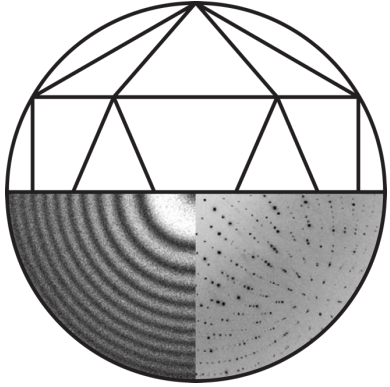
(3) Deformation of endosome membranes

(4) Endosome disintegration and genome release

Cytoplasm



Acknowledgments:



Robert Vácha, Lukáš Sukeník, CEITEC Masaryk University, Czech Republic

Antonín Přidal, Mendel University, Czech Republic

Robert Paxton, MLU Halle-Wittenberg, Germany

Joachim de Miranda, Swedish University of Agricultural Sciences, Sweden

Jiří Nováček, CEITEC Masaryk University, Czech Republic

Roman Pantůček, Martin Benešík, Masaryk University, Czech Republic



What is a virus?

Origins of viruses

Virus structures and infection processes

Extra

I'm in college!

What is important?

Lectures × Bachelor's and Master's theses

- Lectures are the theoretical basis, but you will gain practical skills by solving real problems.
- In a bachelor's/master's thesis, you will encounter questions without a known solution for the first time.

A good supervisor is the basis for your career

The thesis topic and the supervisor are an indivisible whole.

The topic of your bachelor's thesis will define your study specialisation and thus determine your employment.



How to choose a supervisor and topic for your bachelor's and master's thesis

- Think and invest time in the choice.
- Internet and MU website.
- Ask **sensible** colleagues and teachers.

Choosing the right supervisor requires information



Starting to work on BC work from the first semester means less stress, better results, and more experience.

What NOT to choose by

- academic degrees
- I like her/him (how she/he lectures)
- I attended his/her lecture
- I have a friend working for him



Desirable traits of a supervisor

- receives individual research funding from GAČR, TAČR, and EU
 - Read CV
- publishes as corresponding author in international journals
 - Read CV
- students and post-docs are at the first place of their supervisor's publications
- studied or worked abroad

Structure of deformed wing virus, a major honey bee pathogen

Karel Škubník^a, Jiří Nováček^a, Tibor Füzik^a, Antonín Přidal^b, Robert J. Paxton^c, and Pavel Plevka^{a,1}

^aStructural Virology, Central European Institute of Technology, Masaryk University, 62500 Brno, Czech Republic; ^bDepartment of Zoology, Fishery, Hydrobiology, and Apidology, Faculty of Agronomy, Mendel University in Brno, 613 00 Brno, Czech Republic; and ^cInstitute of Biology/Zoology, Martin Luther University Halle-Wittenberg, 06120 Halle, Germany

Edited by Wolfgang Baumeister, Max Planck Institute of Chemistry, Martinsried, Germany, and approved February 10, 2017 (received for review September 20, 2016)

The worldwide population of western honey bees (*Apis mellifera*) is under pressure from habitat loss, environmental stress, and pathogens, particularly viruses that cause lethal epidemics. Deformed wing virus (DWV) from the family *Iflaviridae*, together with its vector, the mite *Varroa destructor*, is likely the major threat to the world's honey bees. However, lack of knowledge of the atomic structures of iflaviruses has hindered the development of effective treatments against them. Here, we present the virion structures of DWV determined to a resolution of 3.1 Å using cryo-electron microscopy and 3.8 Å by X-ray crystallography. The C-terminal extension of capsid protein VP3 folds into a globular protruding (P) domain, exposed on the virion surface. The P domain contains an Asp-His-Ser catalytic triad that is, together with five residues that are spatially close, conserved among iflaviruses. These residues may participate in receptor binding or provide the protease, lipase, or esterase activity required for entry of the virus into a host cell. Furthermore, nucleotides of the DWV RNA genome interact with VP3 subunits. The capsid protein residues involved in the RNA binding are conserved among honey bee iflaviruses, suggesting a putative role of the genome in stabilizing the virion or facilitating capsid assembly. Identifying the RNA-binding and putative catalytic sites within the DWV virion structure enables future analyses of how DWV and other iflaviruses infect insect cells and also opens up possibilities for the development of antiviral treatments.

colony collapse disorder | virus | structure | *Apis mellifera* | honey bee

The western honey bee (*Apis mellifera*) plays a vital role in world agriculture by providing pollination services to diverse commercial crops, a service valued at US\$ 215 billion annually (1). In addition, honey bees pollinate numerous wild flowering plants, thereby supporting biodiversity (2, 3). However, over the past two decades, honey bees have suffered from elevated mortality in North America and Europe (4, 5). Colony losses have been associated with the exotic ectoparasitic mite *Varroa destructor*, which feeds on honey bee hemolymph, thereby vectoring numerous honey bee viral pathogens, in particular the iflavirus deformed wing virus (DWV). In the absence of varroa, DWV levels are low, and the virus causes asymptomatic infections. Varroa-infested colonies show elevated levels of DWV (6, 7). Symptoms associated with acute DWV infections include the death of pupae, as well as deformed wings, shortened abdomen, and cuticle discoloration of adult bees that die soon after pupation, causing colony collapse (6, 8). Indeed, winter colony mortality is strongly correlated with the presence of DWV, irrespective of the levels of varroa infestation (8, 9). DWV-induced loss of honey bees, coupled with a long-term decline in beekeeping, has become a serious threat to adequate provision of pollination services, threatening food security and ecosystem stability (1).

Viruses from the order *Picomavirales*, including the family *Iflaviridae*, have nonenveloped icosahedral virions that are about 30 nm in diameter (10). Iflavirus capsids protect 10,000-nt-long ssRNA genomes, which are translated into polyproteins that are cotranslationally and posttranslationally cleaved by viral proteases

to produce structural (capsid-forming) and nonstructural proteins (11). The major capsid proteins VP1, VP2, and VP3 originating from a single polyprotein form a protomer, the basic building block of the pseudo-T3 icosahedral capsid. The entire capsid consists of 60 such protomers, arranged in 12 pentamer units of 5 protomers each.

Previously, the structure of the iflavirus Chinese sacbrood virus was characterized to a resolution of 25 Å by cryo-electron microscopy. The structure confirmed the pseudo-T3 icosahedral symmetry of its capsid and a smooth outer surface of the virion (12). Recently, we determined the structure of the iflavirus slow bee paralysis virus (SBPV) to a resolution of 2.6 Å by X-ray crystallography (13). Despite its efficient transmission by *V. destructor*, SBPV infection is a rare disease of honey bees (14). The structure revealed that the C-terminal extension of capsid protein VP3 of SBPV forms a globular protruding (P) domain positioned at the virion surface. The P domain is anchored to the core of the VP3 subunit by a 23-residue-long flexible linker that allows the P domain to attach to different areas of the capsid (13). In addition, the P domain contains the putative active site Asp-His-Ser, which is conserved among several iflaviruses (13). Iflaviruses were also proposed to harbor short VP4 subunits consisting of only about 20 residues (11, 14); however, electron density

Significance

Honey bee populations in Europe and North America have been decreasing since the 1950s. Deformed wing virus (DWV), which is undergoing a worldwide epidemic, causes the deaths of individual honey bees and collapse of whole colonies. We determined three-dimensional structures of DWV at different conditions and show that the virus surface is decorated with protruding globular extensions of capsid proteins. The protruding domains contain a putative catalytic site that is probably required for the entry of the virus into the host cell. In addition, parts of the DWV RNA genome interact with the inside of the virus capsid. Identifying the RNA binding and catalytic sites within the DWV virion offers prospects for the development of antiviral treatments.

Author contributions: P.P. designed research; K.Š. and A.P. performed research; R.J.P. contributed new reagents/analytic tools; K.Š., J.N., T.F., and P.P. analyzed data; and K.Š., R.J.P., and P.P. wrote the paper.

The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

Freely available online through the PNAS open access option.

Data deposition: Cryo-EM maps of the DWV virions from different conditions have been deposited in the Electron Microscopy Data Bank (EMDB) (accession nos. EMD-4014, EMD-3574, EMD-3570, and EMD-3575); the corresponding coordinates and structure factors have been deposited in the Protein Data Bank (PDB), www.pdb.org (PDB ID codes 5LBQ, 5MV5, 5MUP, and 5MV6). The crystal structures of the DWV virion and P domain have been deposited under PDB ID codes 5G52 and 5G51.

¹To whom correspondence should be addressed. Email: pavel.plevka@ceitec.muni.cz.

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Structure of deformed wing virus, a major honey bee pathogen

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Abstract
 The role of the genome in stabilizing the virion or facilitating capsid assembly. Identifying the RNA-binding and putative catalytic sites within the DWV virion structure enables future analyses of how DWV and other iflaviruses infect insect cells and also opens up possibilities for the development of antiviral treatments.

colony collapse disorder | virus | structure | *Apis mellifera* | honey bee

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allows the P domain to attach to different areas of the capsid (13). In addition, the P domain contains the putative active site Asp-His-Ser, which is conserved among several iflaviruses (13).

Iflaviruses were also shown to be involved in the development of only about 10% of the honey bee colonies.

Significance

Honey bee population has been decreasing worldwide, which is underpinned by the development of individual honey bee colonies. We determined three-dimensional structures of DWV virions and P domain protruding globular domains. Our findings are likely to be generally applicable to other iflaviruses. In addition, parts of the virion structure are likely to be required for the development of antiviral treatments.

Author contributions: P.P. designed research; K.Š. and A.P. performed research; R.J.P. contributed new reagents/analytic tools; K.Š., R.J.P., and P.P. analyzed data; and K.Š., R.J.P., and P.P. wrote the paper.
 The authors declare no conflict of interest.
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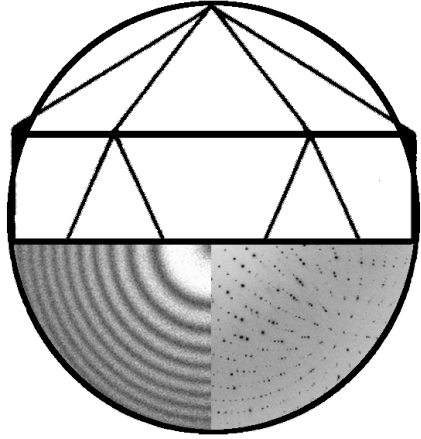
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Plan

- at least 3 personal interviews with potential supervisors
- the supervisors will be happy if you come to talk to them casually about job opportunities
- get to know the members of the lab (students, technicians, post-docs)
- internship or short project (2 months)

Acknowledgments:



Jiří Nováček, CEITEC Masaryk University, Czech Republic
Roman Pantůček, Masaryk University, Czech Republic

