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MUNI

Advanced biophysical methods for
characterization of biosamples

Atomic Force Microscopy of biosamples

Jan PŘIBYL

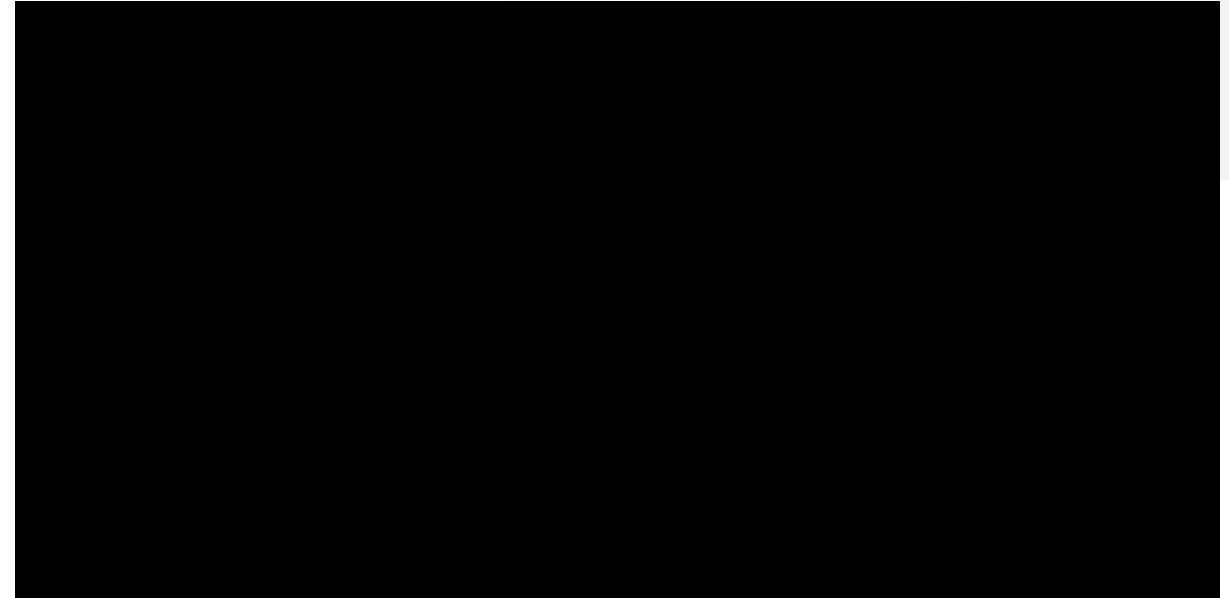
CEITEC MU, Core Facility NanoBio

Masaryk university, Brno, Czech Republic

E-mail: jan.pribyl@ceitec.muni.cz

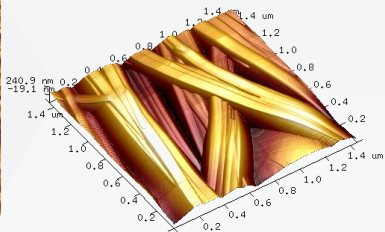
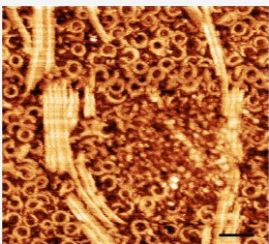
Content

- **AFM, BioAFM technology**
- **AFM operation modes: Imaging, Indentation, ...**
- **Examples**
- **Core Facility NanoBiotechnology**



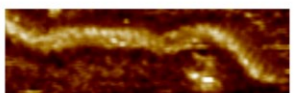
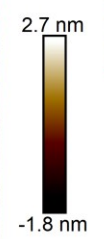
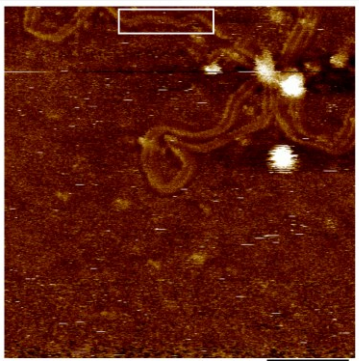
Introduction

Bio Atomic Force Microscopy (BioAFM)

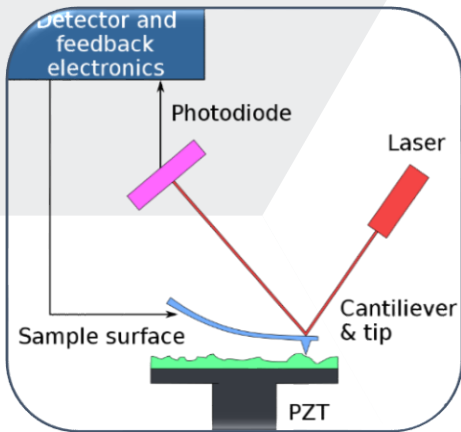
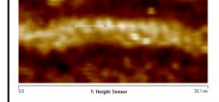
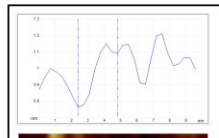


IMAGING

Proteins, DNA, Nanoobjects



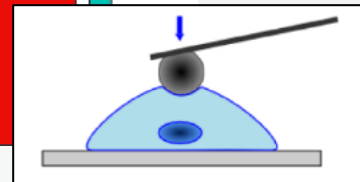
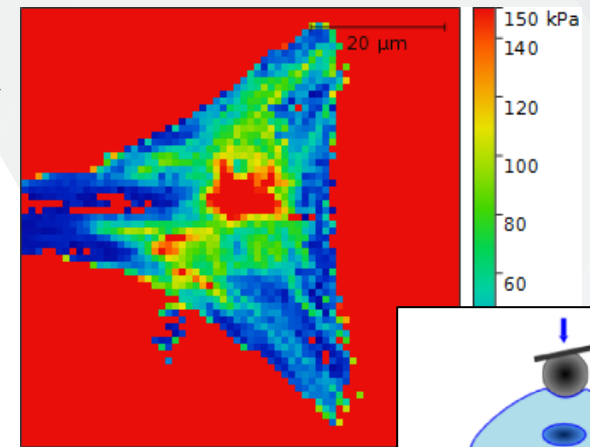
after low-pass filtering



by OverloadQ

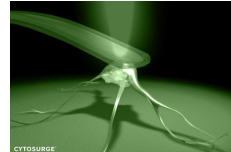
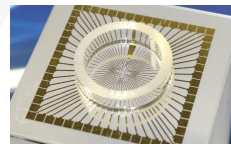
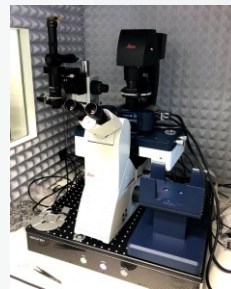
NANOINDENTATION

Stiffness mapping

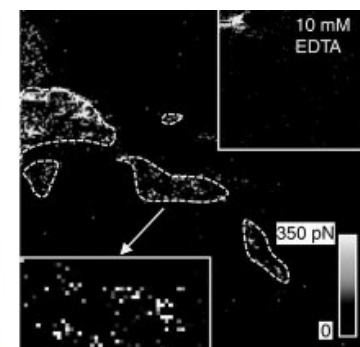
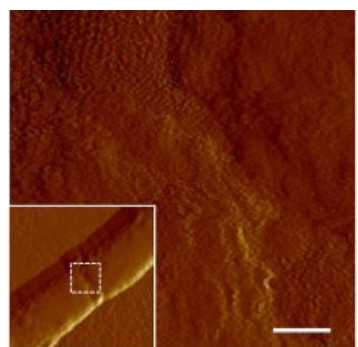
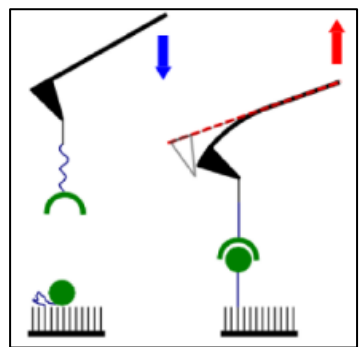


Combination with other techniques

- Optical (fluorescence) microscopy
- Multielectrode Array (MEA)
- FluidFM

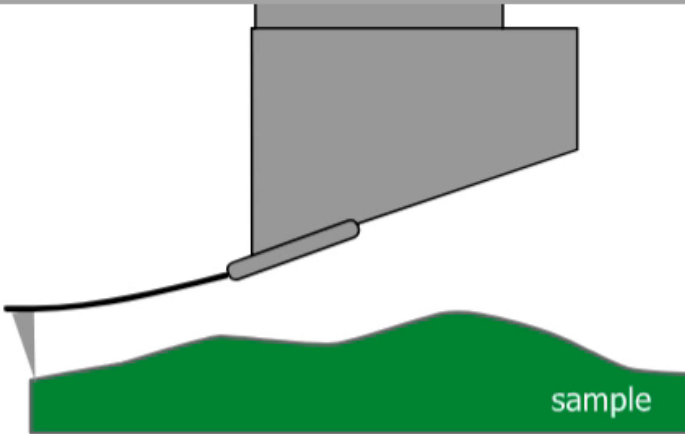


AFFINITY INTERACTION



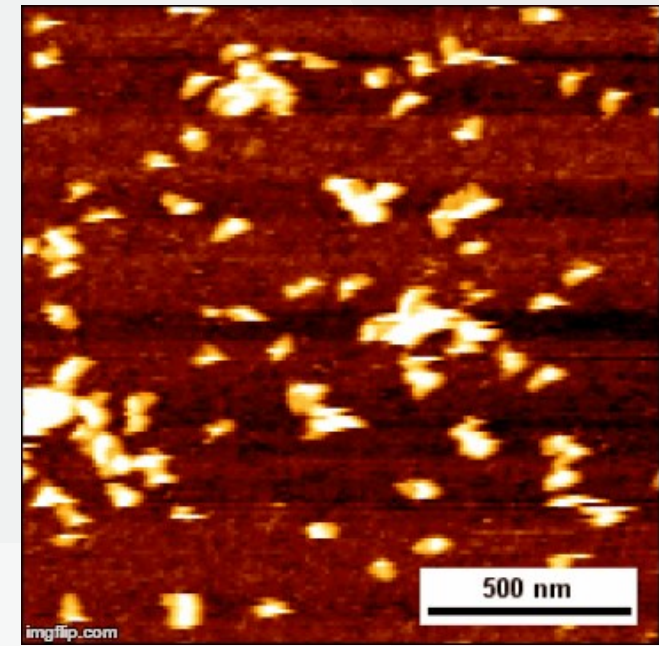
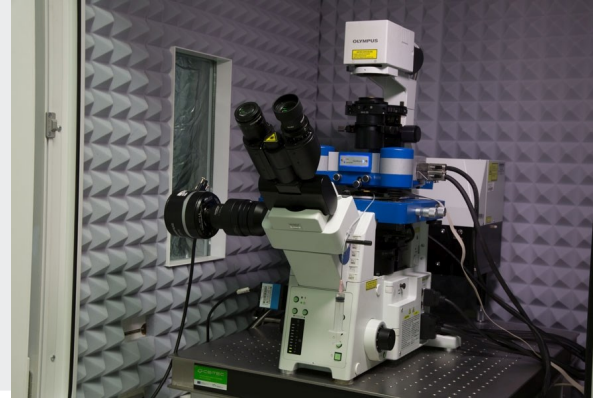
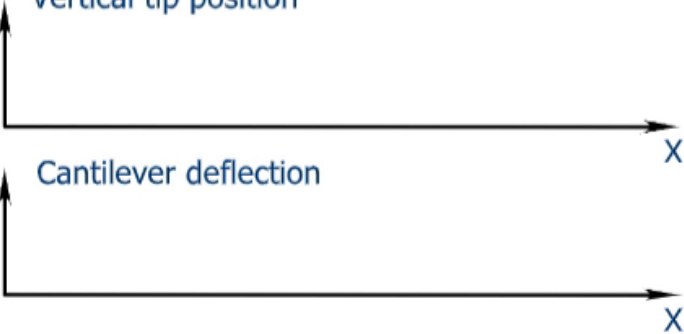
Jak funguje AFM

Constant Force mode



Vertical tip position

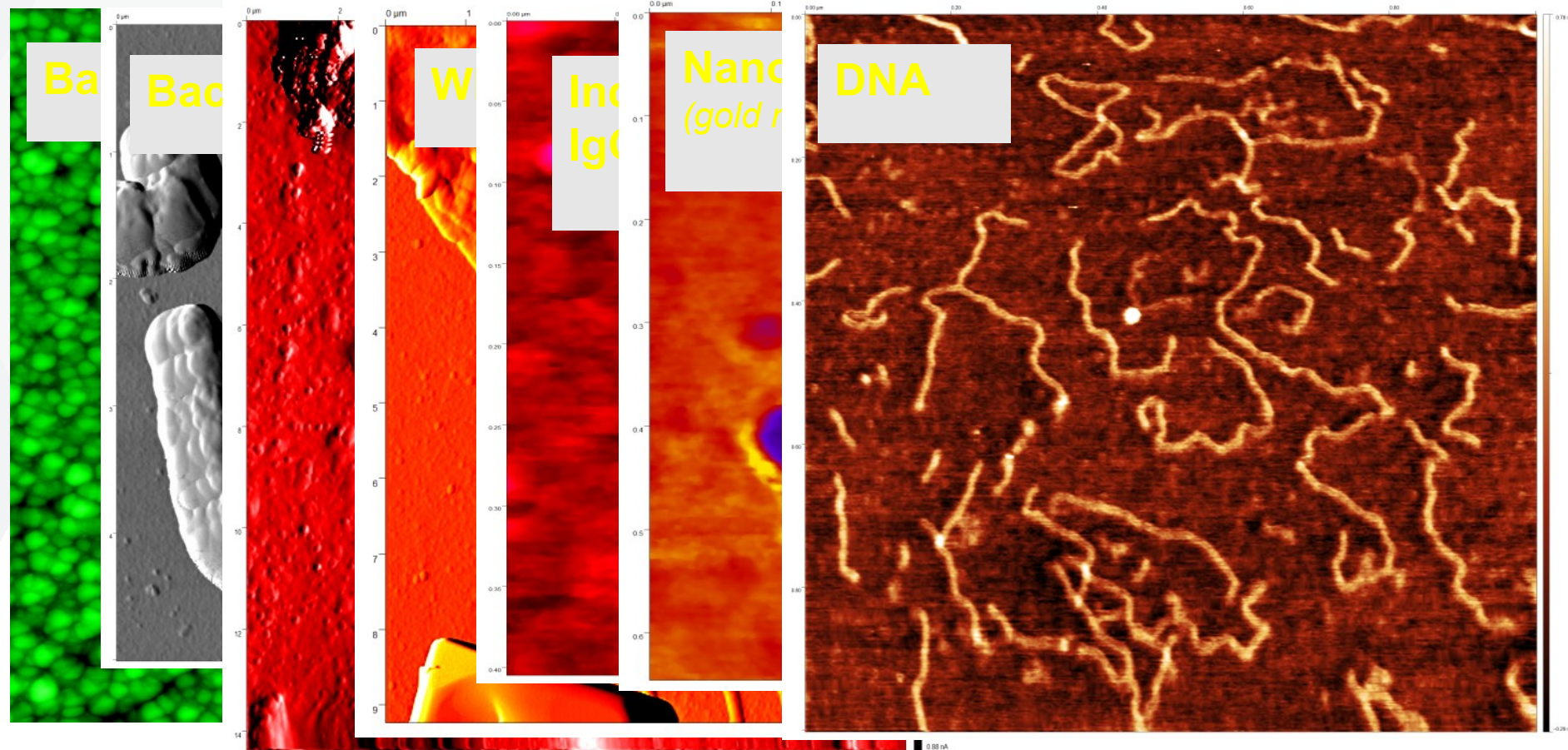
Cantilever deflection



AFM (Atomic Force Microscope)

- Z rodiny tzv. Probe microscopies = skenující próbou
- Posun hrotu nad vzorkem = topografie povrchu
- Další vlastnosti vzorku (tuhost, adhezivita) z analýzy signálu

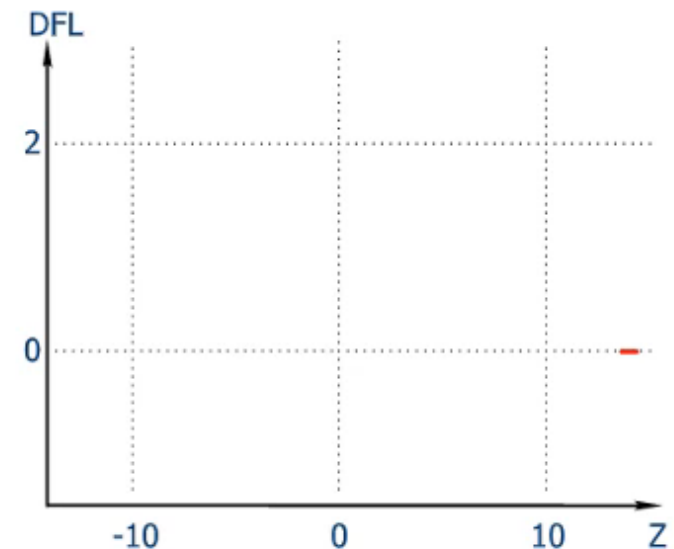
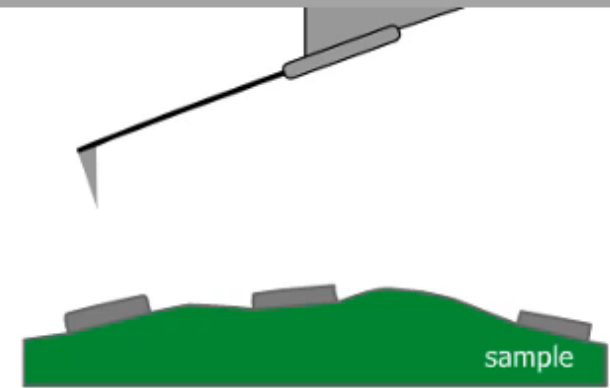
AFM visualization of biomolecules and bioobjects



AFM indentation

- AFM cantilever = nanosensor
- Force-distance curve (FDC) – describes interaction/deformation
- FDC = Young's modulus, adhesivity, rigidity, etc.

FORCE-DISTANCE CURVES

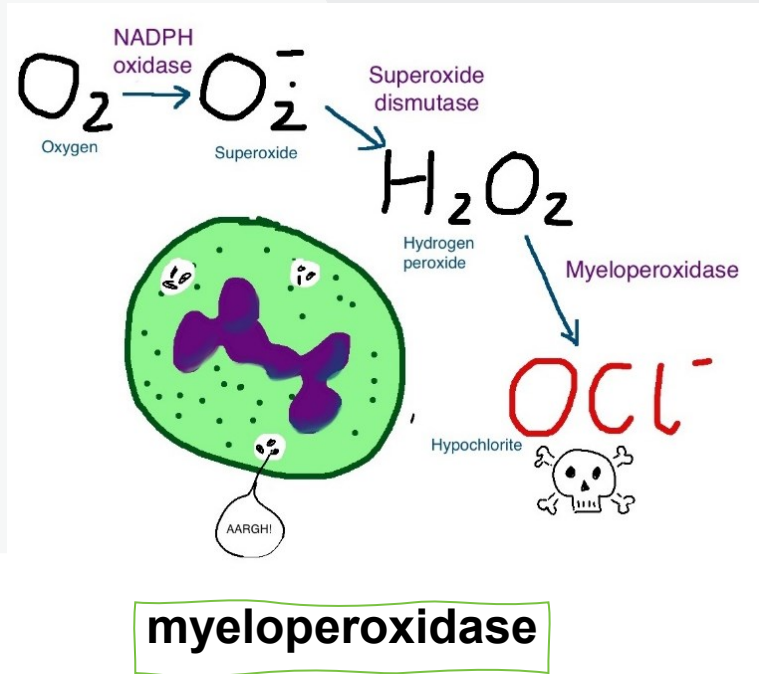


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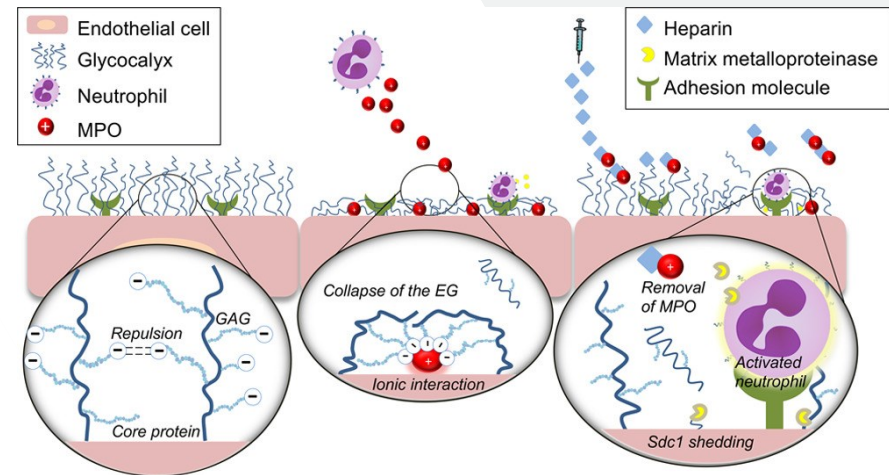
www.ntmdt-si.com

Imaging is not enough...

Hyaluronan-myeloperoxidase complex

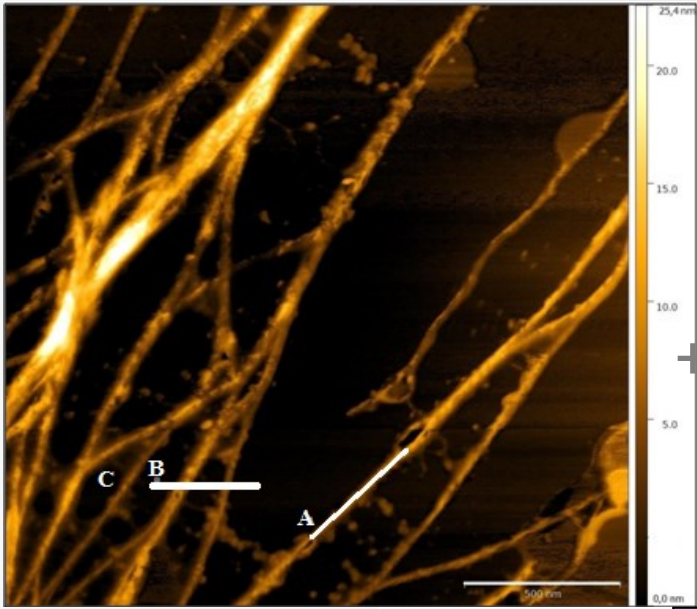


MPO (Myeloperoxidase) Reduces Endothelial Glycocalyx Thickness Dependent on Its Cationic Charge

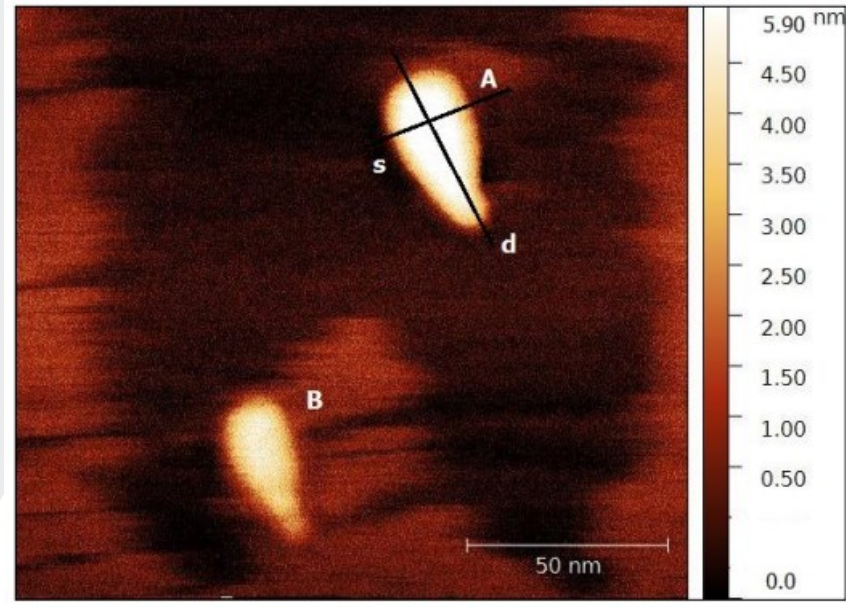


Kashish Manchanda, et al. Arteriosclerosis, Thrombosis, and Vascular Biology. 2018;38:1859–1867

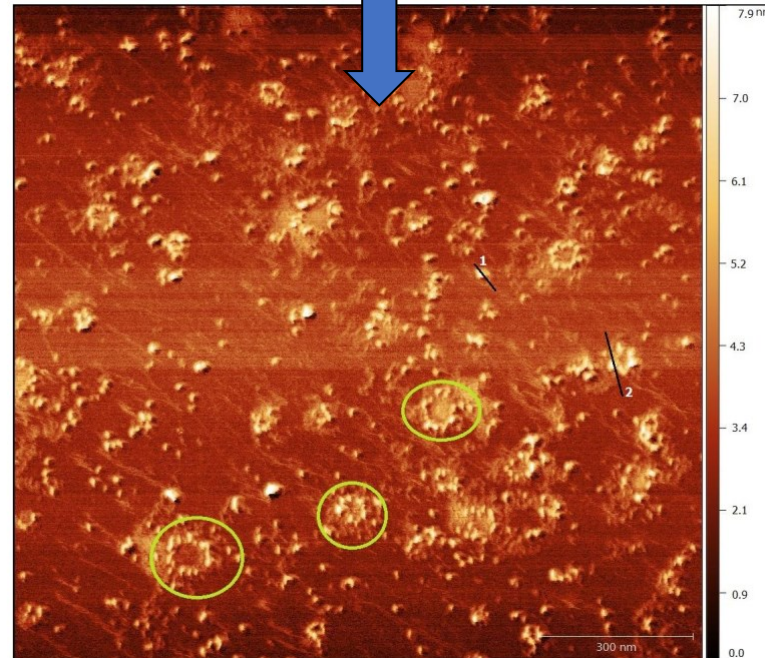
Imaging



hyaluronan



myeloperoxidase

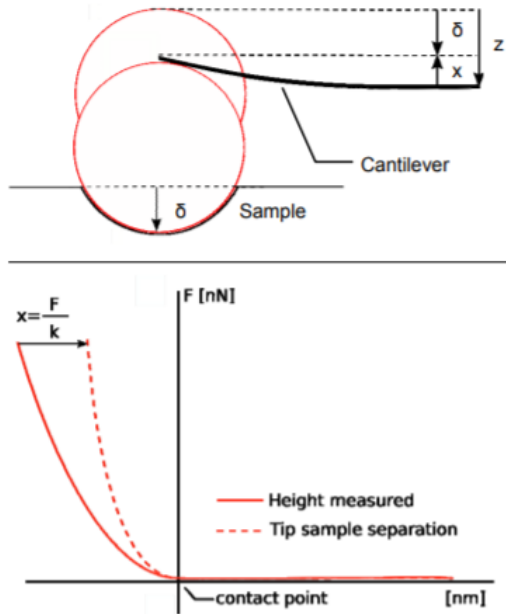


Molecular complex

FDC analysis

Young's modulus

Experiment +
FDC recording



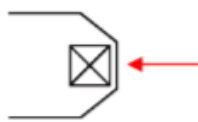
Tip (indenter) definition

Four-sided pyramid

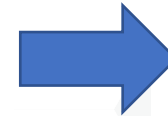
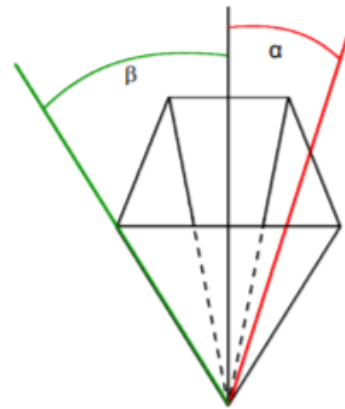
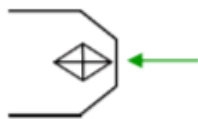
$$F = \frac{E}{1-\nu^2} \frac{\tan \alpha}{\sqrt{2}} \delta^2$$

$$a = \frac{\tan \alpha}{\sqrt{2}} \delta$$

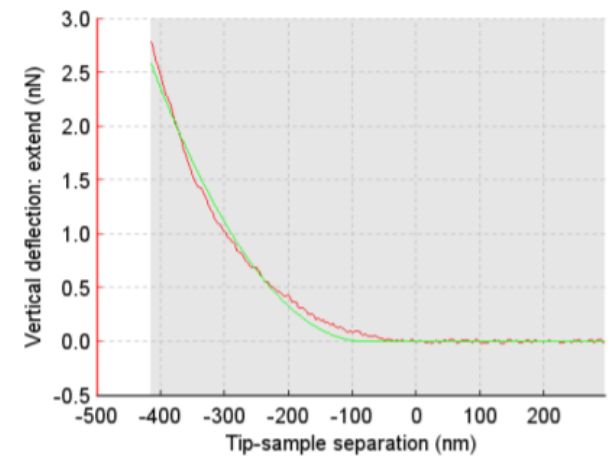
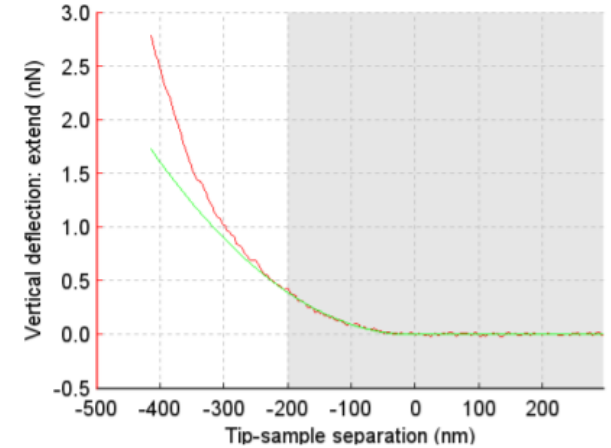
α = face angle, usually given for Si_3N_4 -cantilevers



β = edge angle, usually given for Si-cantilevers



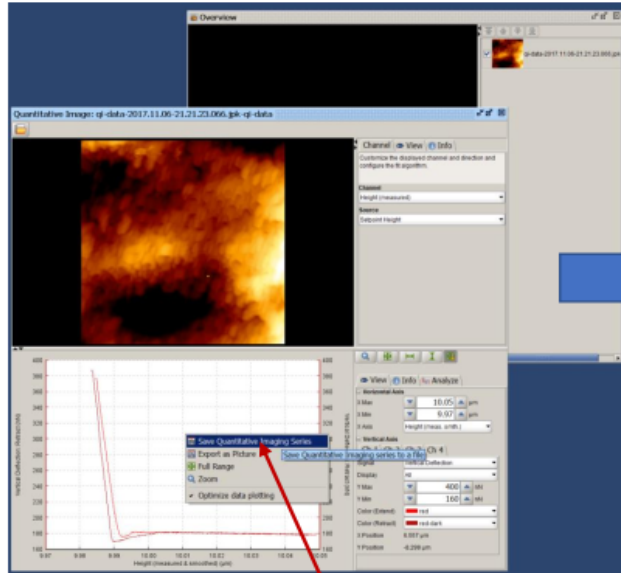
FDC fit



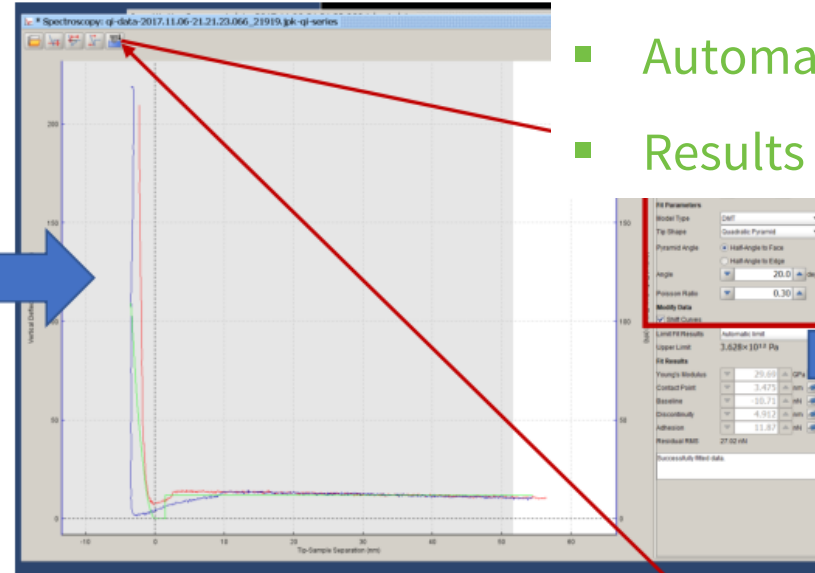
FDC automatic analysis

Young's modulus

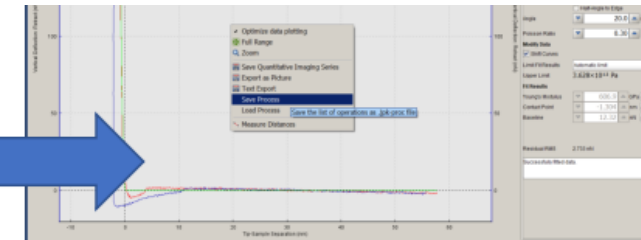
- Force-Distance curves maps
- Typically, 32x32, 64x64 and more
- Automated FDC processing
- Results – stiffness maps, adhesivity maps, ...



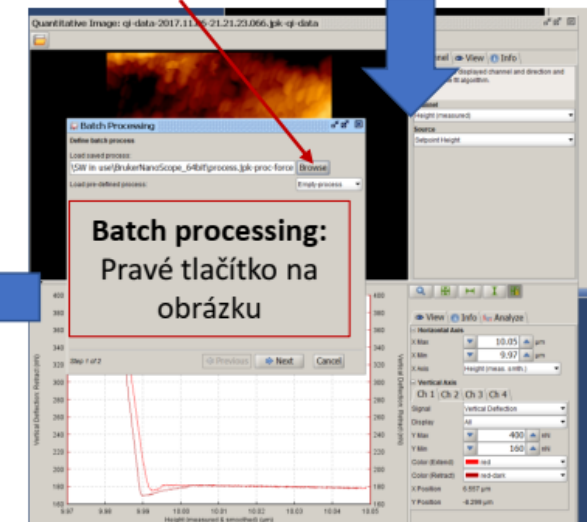
Vybrat pěknou křivku - uložit (pravé tlačítko na křivce)



Křivku otevřít – nadefinovat požadovaný proces



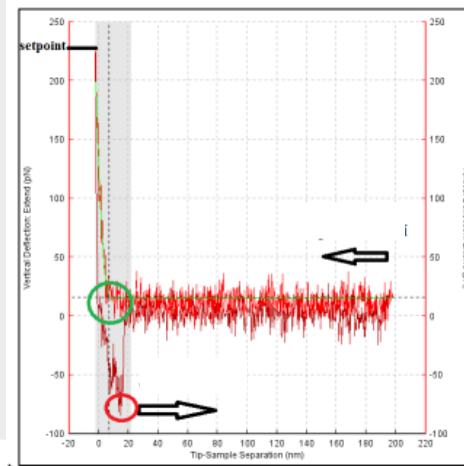
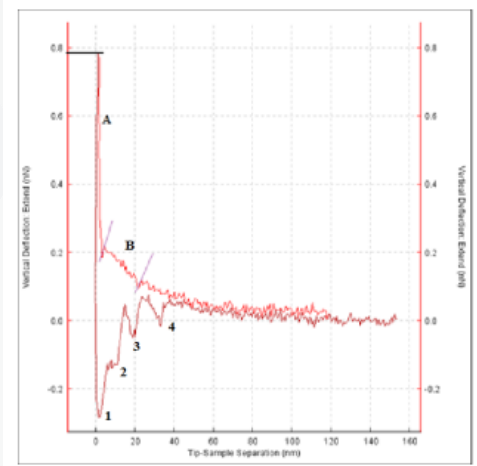
Uložit např. jako:
process.jpk-proc-force



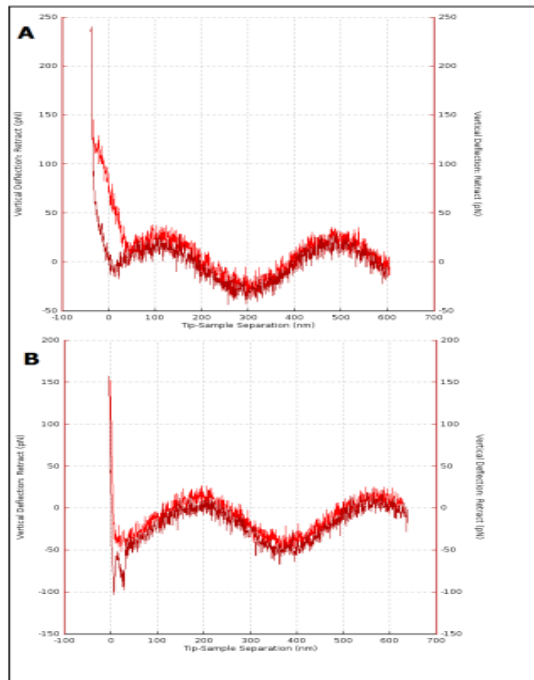
Batch processing:
Pravé tlačítko na
obrazku

... další strana

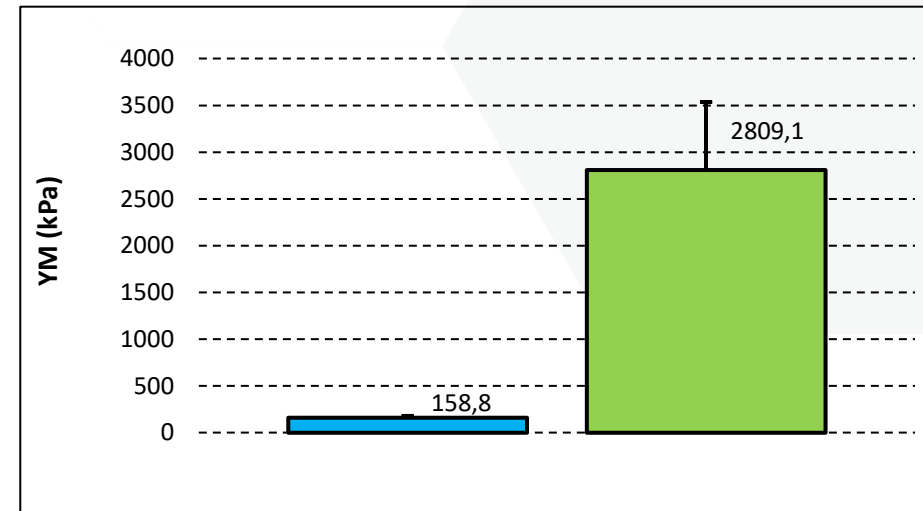
FD curve recording and analysis



Not too strong!

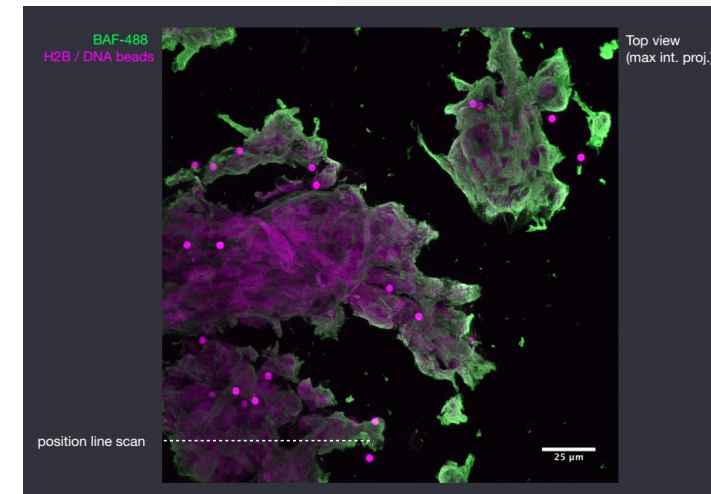
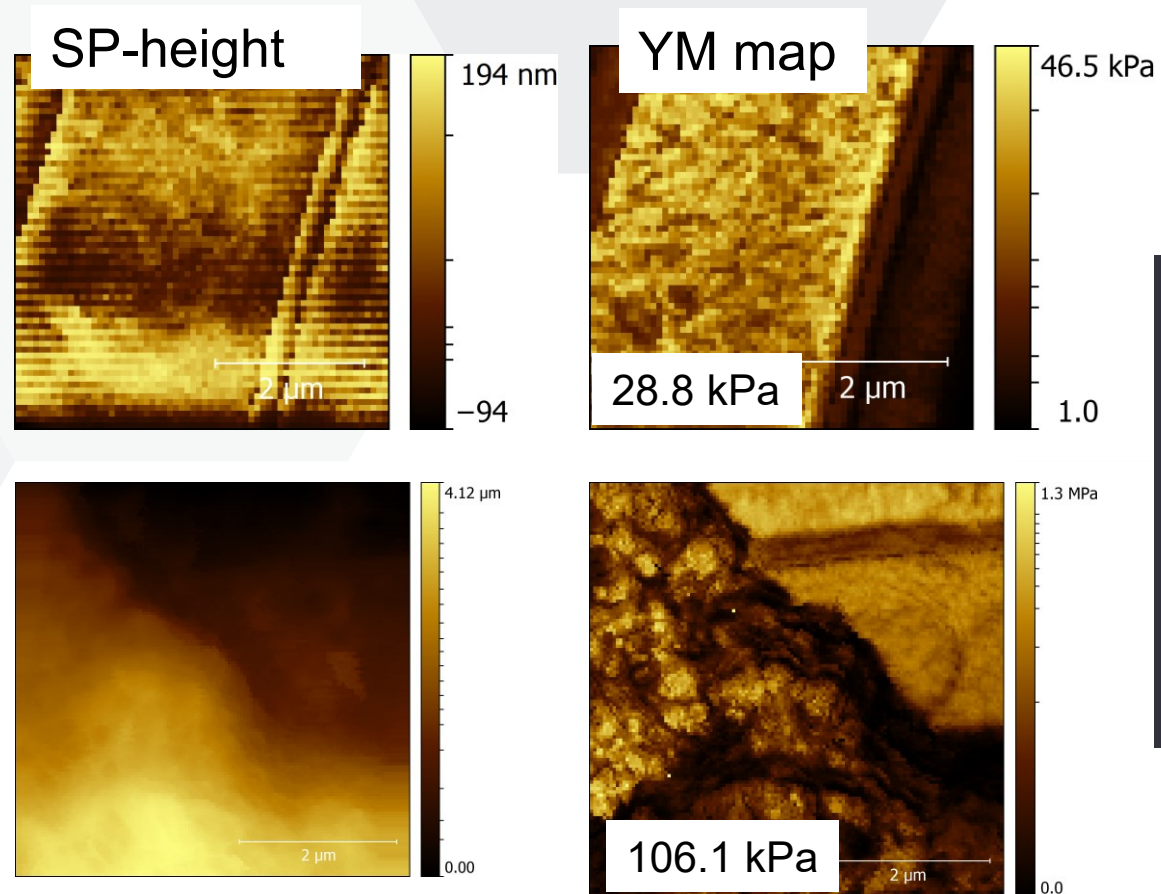


HA-MPO complex is 20x stiffer comparing to HA

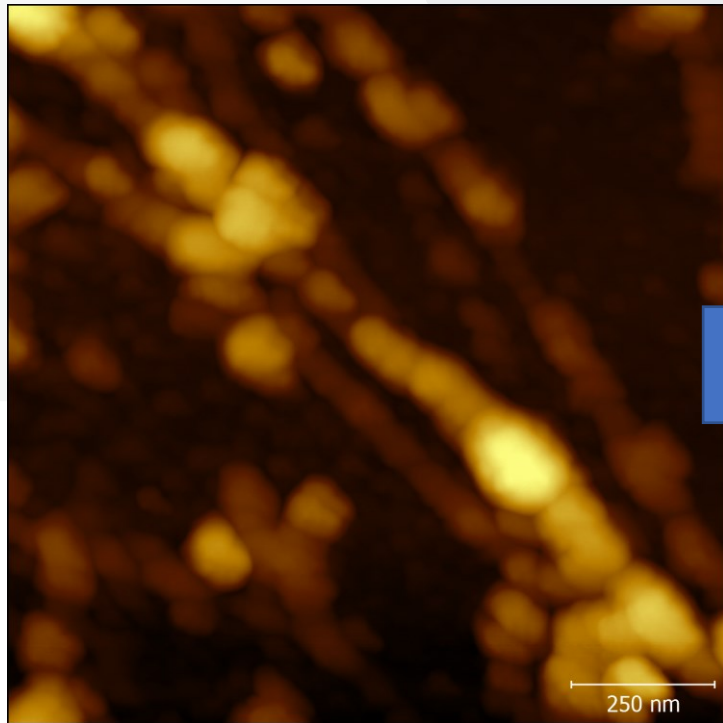


Chromatin-protein interaction

- For tissue engineering
ForceMapping as imaging method
- Collapsible material
 - Soft material (macroscopic view)

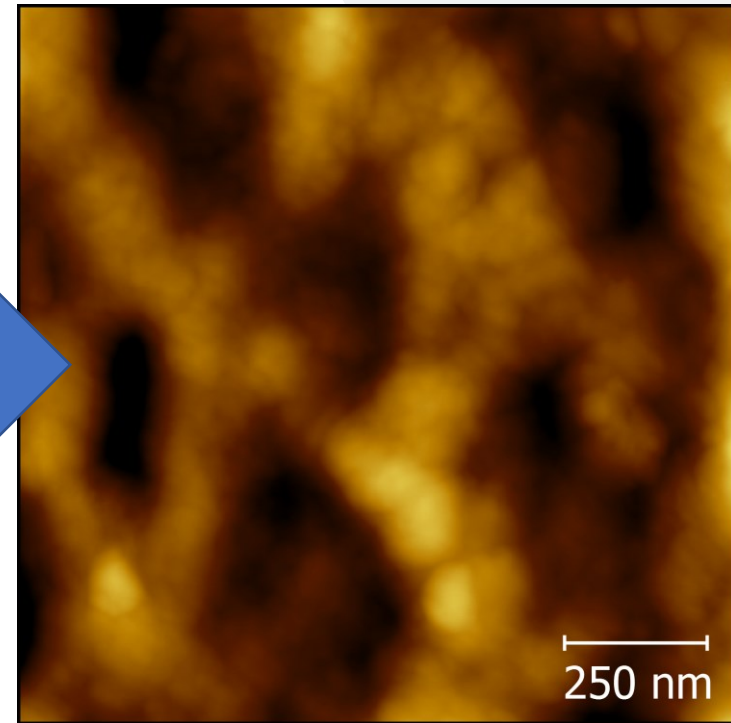
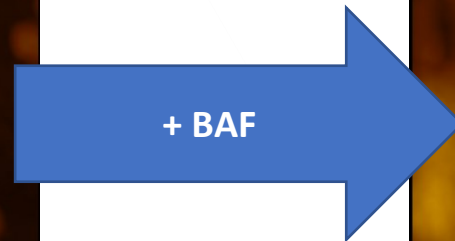


Chromatin only
Chains composed of ellipsoids



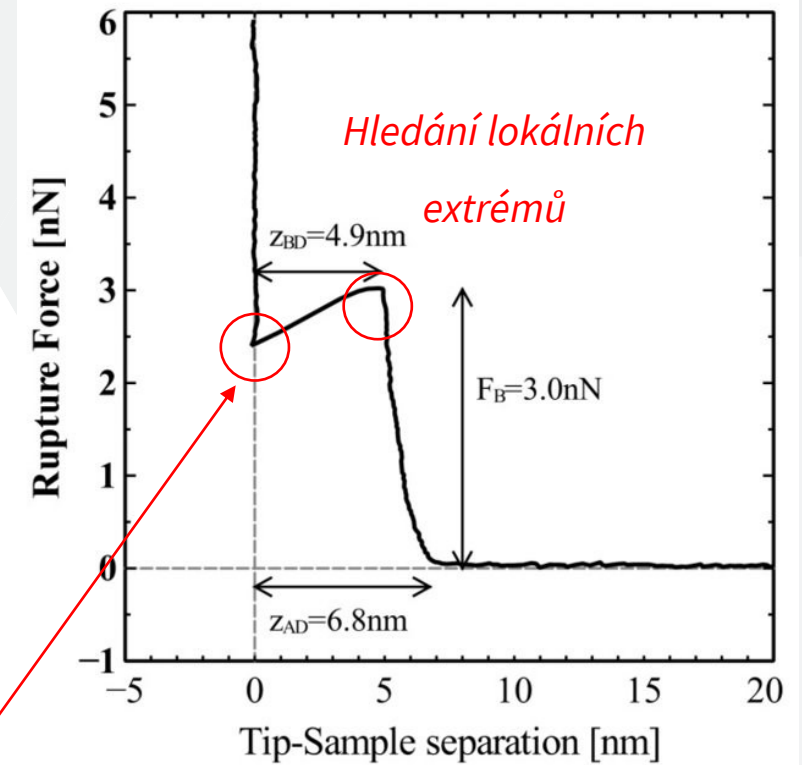
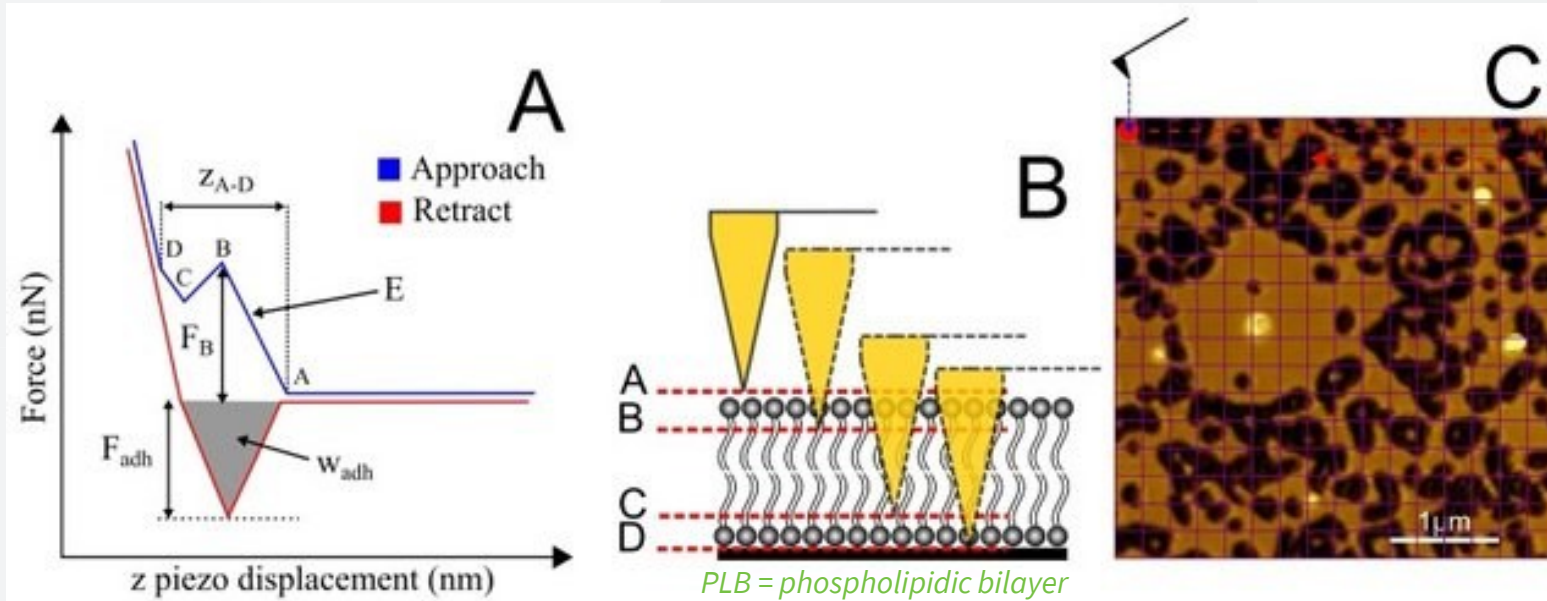
SUMMARY

Chromatin-BAF
Granular structure, grains
composed of fibers (?) (~17nm)



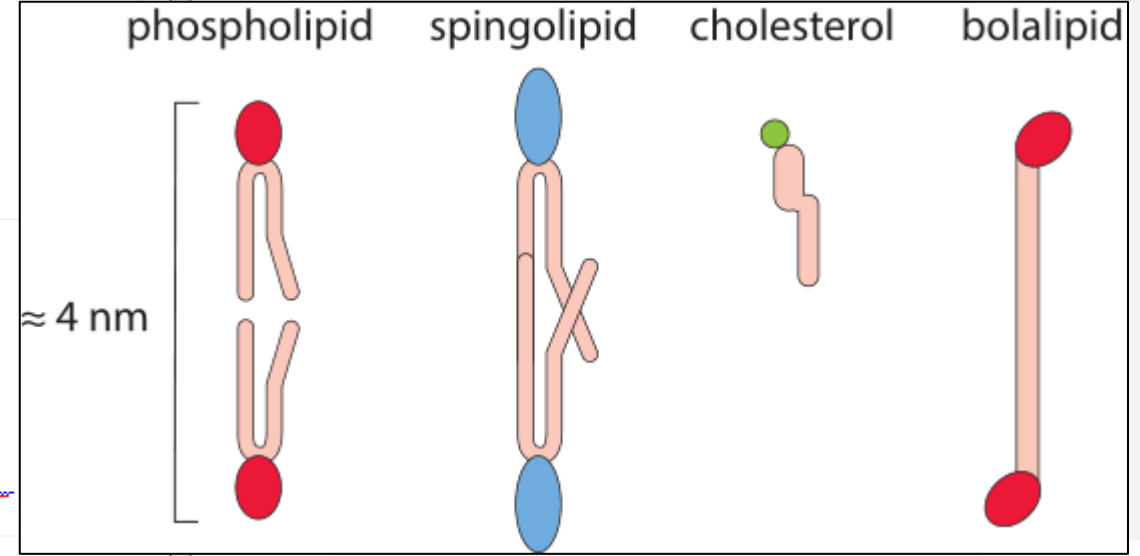
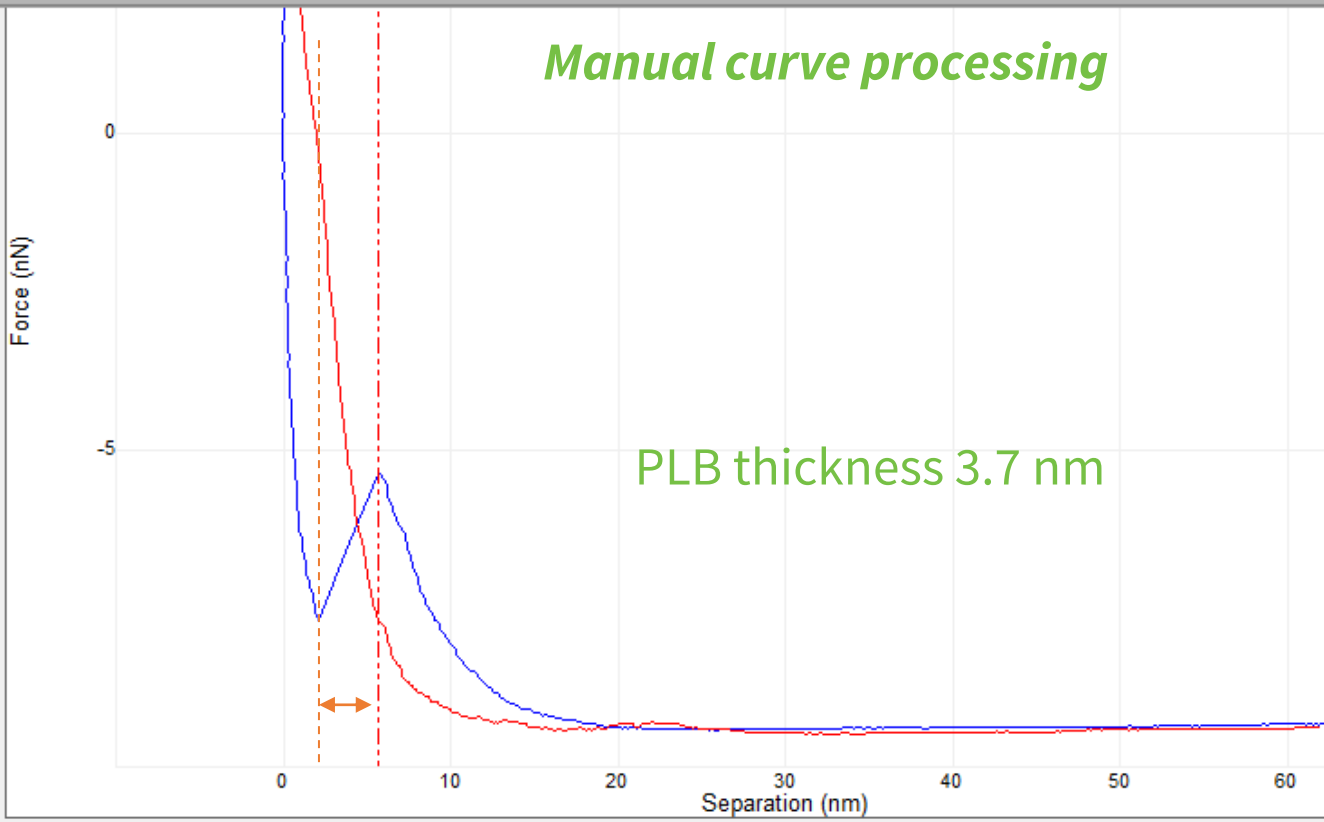
AFM as destructive sensor

FDC, Rupture events



- PLB = PhosphoLipidic Bilayer (= cell membrane *in-vitro*)
- Force application = bilayer rupter (=rupture event)
- Z = PLB thickness
- F = PLB strength
- Difficult automatic analysis

Manual curve processing



Channel data

- Image Data
 - X Data Type
 - Sens. DeflSens
 - Plot Units
 - Spring Constant
 - Display Mode
 - Plot Invert
- Deflection Error
- Z
- 43.00 nm/V
- Force
- 0.7000 N/m
- Deflection Error vs. Sep
- Normal

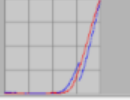


Separation signal

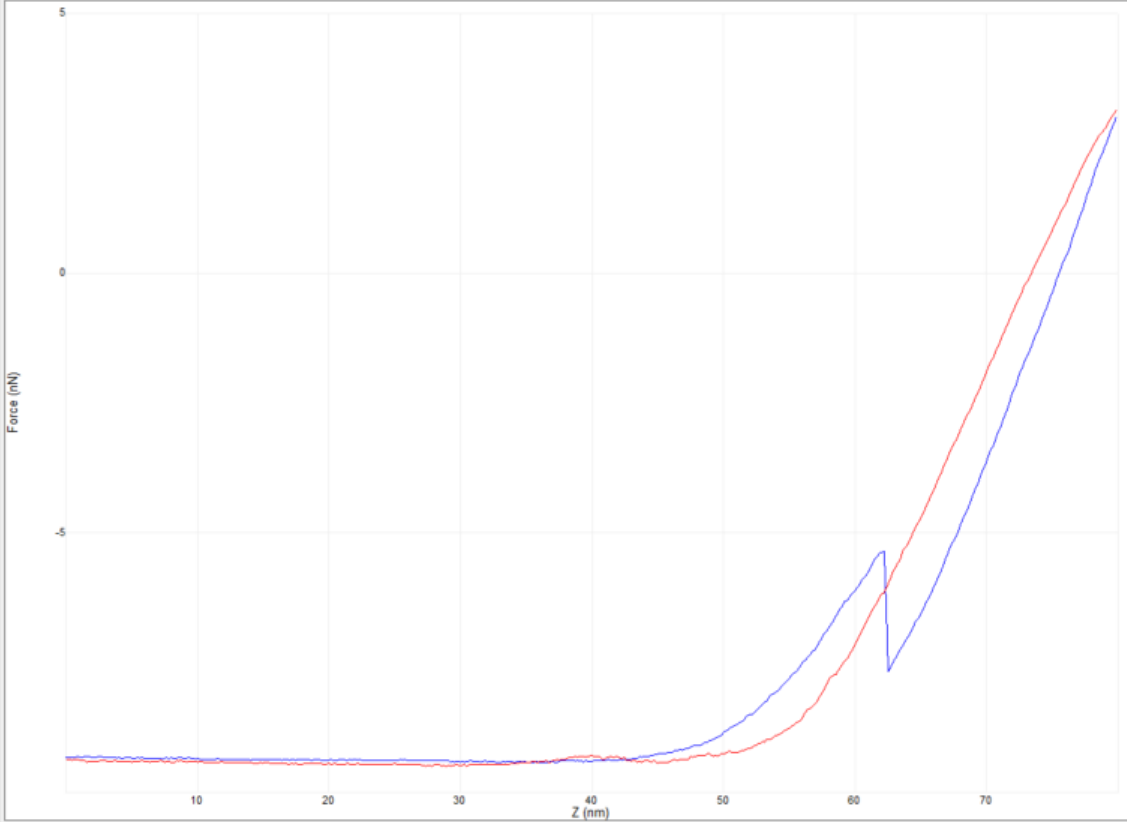
Roi.mca

cr00012.0....spm

Deflection Error



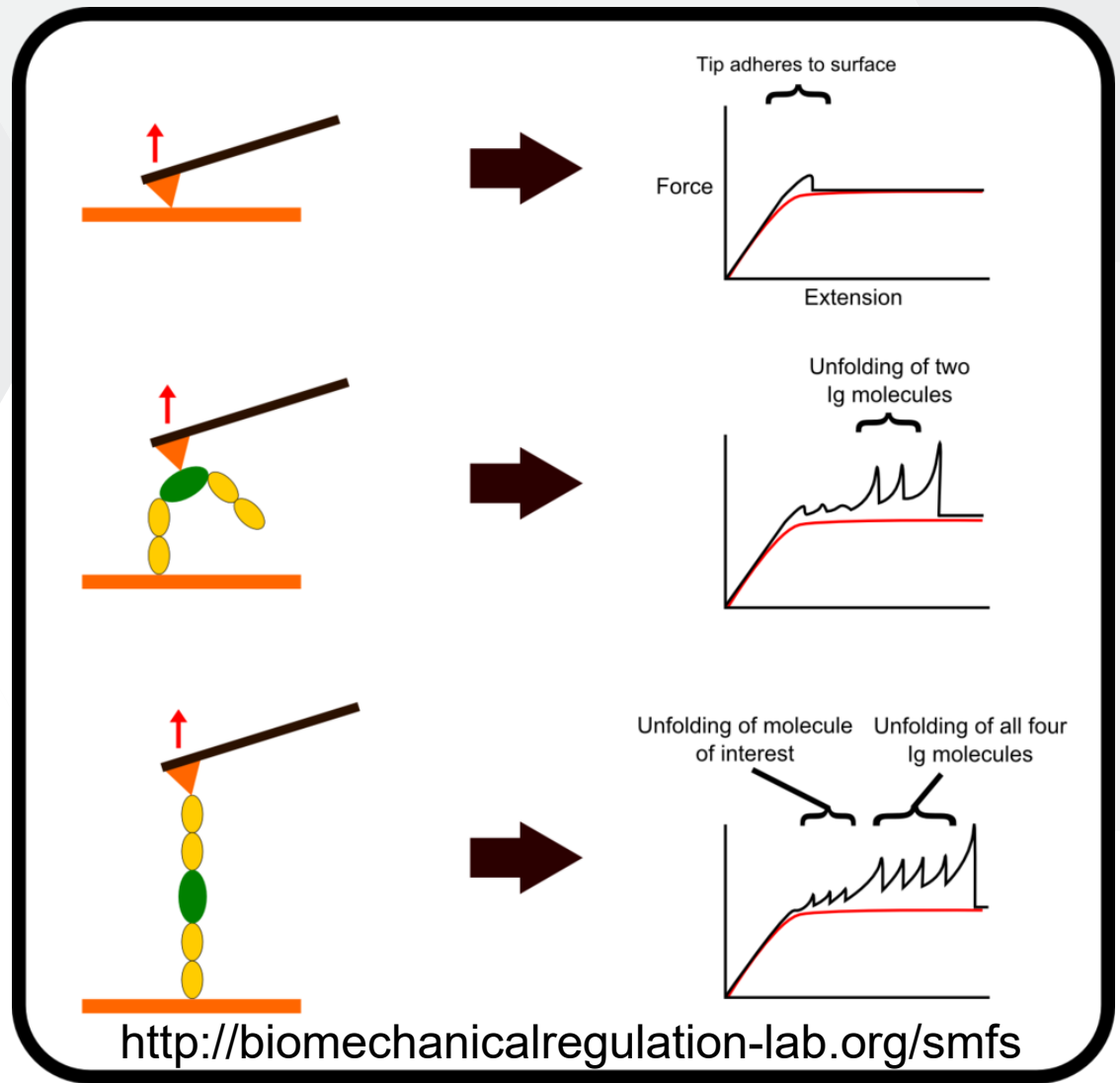
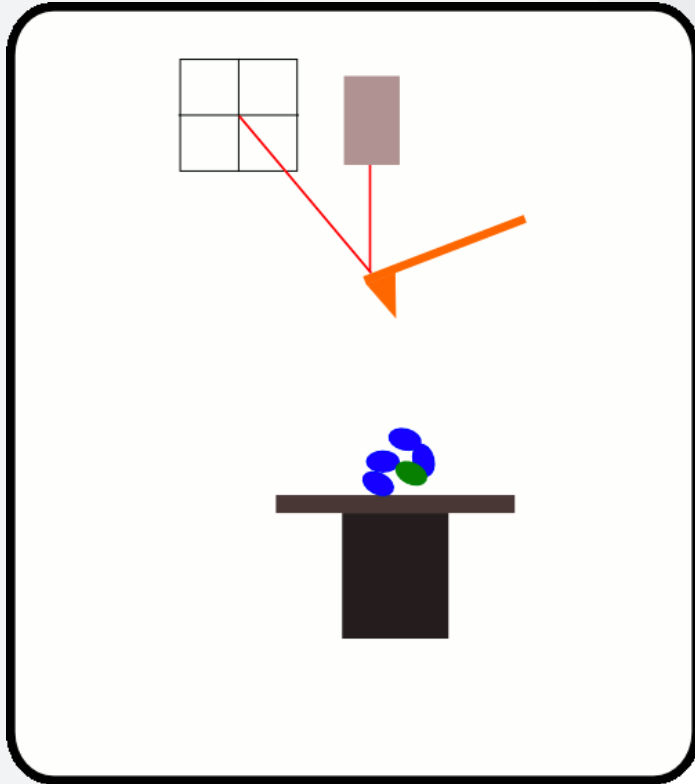
Ramp Data



Marker Pair 1	
- X0	0.000 nm
- X1	0.000 nm
- Y0	0.000 nN
- Y1	0.000 nN
- X1 - X0	0.000 nm
- Y1 - Y0	0.000 nN
- dY/dX	
- dX/dY	
- Least Squares Equation	
- Root Mean Square	0.000 nN
- Average Deviation	0.000 nN
Marker Pair 2	
- X2	0.000 nm
- X3	0.000 nm
- Y2	0.000 nN
- Y3	0.000 nN
- X3 - X2	0.000 nm
- Y3 - Y2	0.000 nN
- dY/dX	
- dX/dY	
- Least Squares Equation	
- Root Mean Square	0.000 nN
- Average Deviation	0.000 nN

Channel data	
- Image Data	
- X Data Type	
- Sens. DeflSens	43.00 nm/V
- Plot Units	Force
- Spring Constant	0.7000 N/m
- Display Mode	Deflection Error vs. Ch2:
- Plot Invert	Normal
- Phase Offset	0.00 ms
- Fit Type	None
- Active Curve	Retract

Single-molecule force spectroscopy (SMFS)



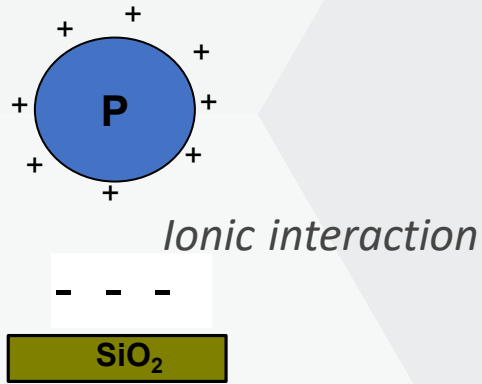
Force dependency of biochemical reactions measured by single-molecule force-clamp spectroscopy

Ionel Popa Pallav Kosuri Jorge Alegre-Cebollada Sergi Garcia-Manyes Julio M. Fernandez

Nature Protocols June, 2013

Protein immobilization

■ Glass

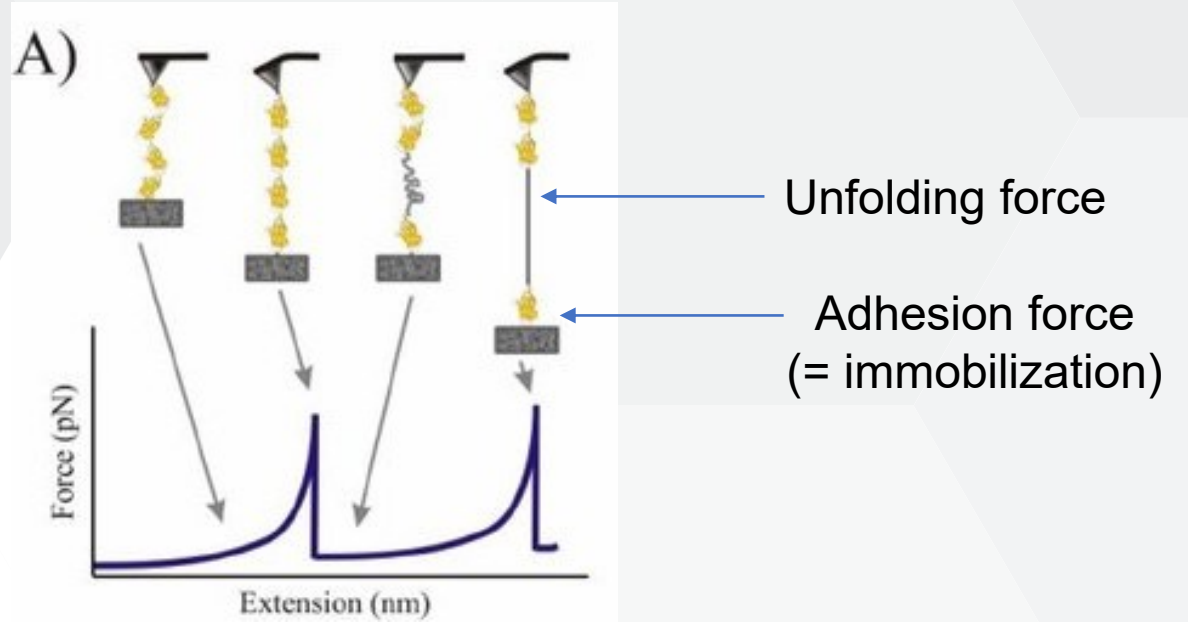


■ Gold:

- Cysteine (SH-) tags
- $F \sim 1 \text{ nN}$ → Ok for most of the unfolding forces

■ Ni-NTA

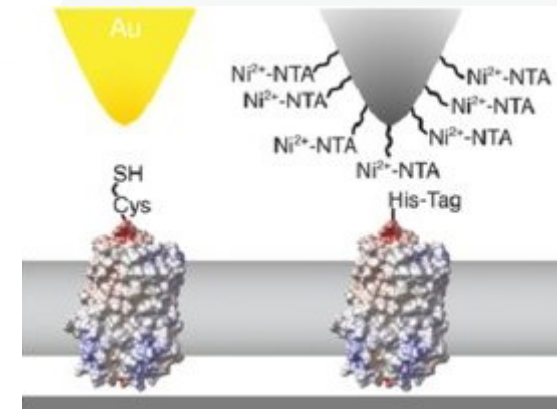
- $F \sim 50 \text{ pN}$
- OK for spectrin, ankyrin or C2 domains
- NOT sufficient e.g., for Ig-domains



Adhesion force > Unfolding force

Probe (cantilever and tip) selection

- **Gold-coated tip** (*few types on the market*)
- **Very soft** ($k < 0.01 \text{ N/m}$)
- **Silicon Nitride material** (*typical for operations in liquid*)
- **Smaller is better** (*lower noise in liquid*)



Probe (cantilever and tip) selection

- **Olympus Biolever** and **Bruker MCLT-Au** – *most of publications*
- **Discontinued** 😞

OLYMPUS Micro Cantilevers

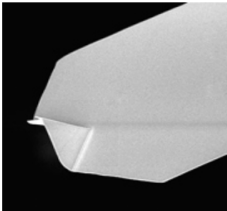
Home | Product features | Measurement example | P

Home > Product Line > BioLever

Silicon cantilever for AC mode

BioLever **Discontinued**

○ Cantilevers for Biological Sample Measurements In Water



Both side Gold coating

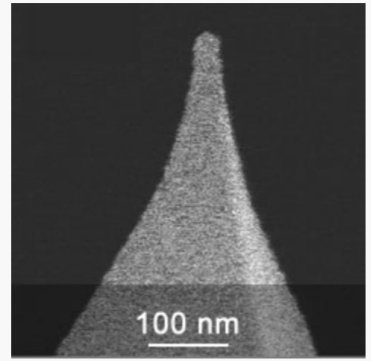
- BL-RC150VB-HW (210 chips/box, Strip Type)
- BL-RC150VB-C1 (24 chips/box, Pre-separated chips)


Material	
Probe apex	Silicon Nitride
Lever	Silicon Nitride
Coating Metal	
Probe side	Gold
Reflex side	Gold

BL-RC150VB-HW, BL-RC150VB-C1

Probe (cantilever and tip) selection

- *New cantilever types*
- *Ordered – to be tested*



HQ:CSC38/Cr-Au 

AFM Probe with 3 Different Gold Coated Contact Mode AFM Cantilevers

Manufacturer: MikroMasch

Coating: Gold Overall
AFM tip shape: Rotated


This probe features 3 cantilevers


F	20 kHz	C	0.09 N/m	L	250 µm
F	10 kHz	C	0.03 N/m	L	350 µm
F	14 kHz	C	0.05 N/m	L	300 µm

*nominal values

Applications


- > Life Science AFM Probes
- > Conductive AFM Probes

How to optimize AFM scan parameters 

NPG-10 

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Overview
Au coated tips, 4 Cantilevers 0.06-0.35N/m

Geometry	Tip Radius (nm)
▼ B Triangular	Nom: 30 Max: 90
Frequency (KHz)	Length (µm)
Nom: 23 Min: 16 Max: 28	Nom: 205 Min: 200 Max: 210
Spring Const (N/m)	Width (µm)
Nom: 0.12 Min: 0.06 Max: 0.24	Nom: 40 Min: 35 Max: 45


Tip Specification
Gold coating on the tip side makes functionalizing the tip easier (e.g., using thiol chemistry).

Cantilever Specification
Cantilever Orientation diagram can be found here. DNP probes have less than 2deg of cantilever. The nominal stress specification for NP probes.

Contact

NANO WORLD INNOVATIVE TECHNOLOGIES

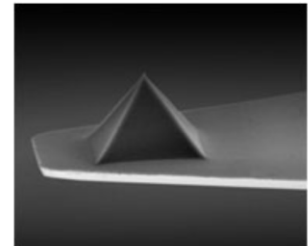
AFM PROBES HOW TO BUY ABOUT US CONTACT BLOG

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Home >

Type: **PNP-TR-Au** **Pyrex-Nitride Probe – TRIangular Cantilevers Au coated**

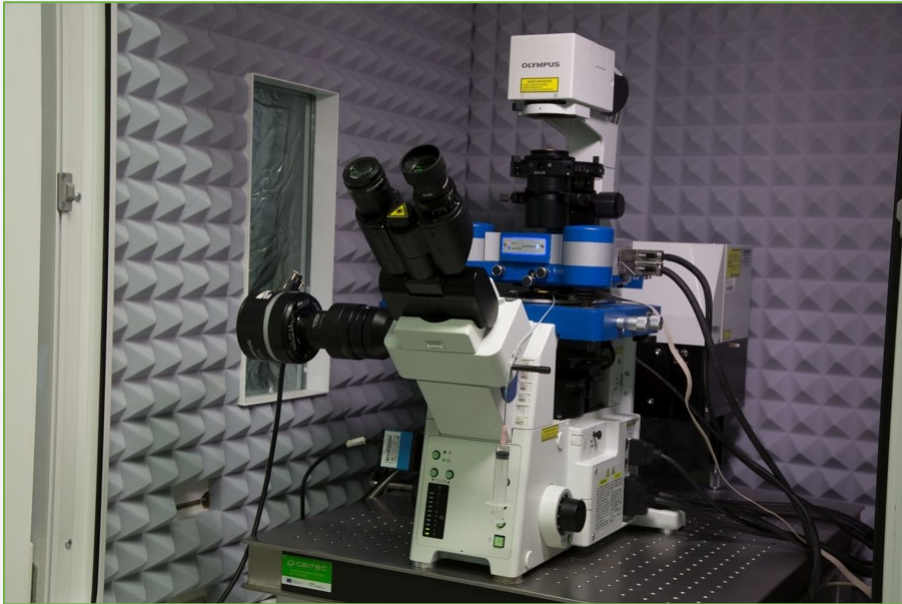
Cantilever Data*	Cant. 1	Cant. 2
Shape	Triangle	
Resonance Frequency	67 kHz	17 kHz
Force Constant	0.32 N/m	0.08 N/m
Length	100 µm	200 µm
Mean Width	13.5 µm	28 µm
Thickness	500 nm	500 nm



Pyrex-Nitride oxide sharpened, pyramidal tip More images

Experimental settings

JPK NanoWizard 3



ATOMIC FORCE MICROSCOPE

ForceRobot® 300

Automated Force Spectroscopy

Label-free, single molecule technique
Measure forces between and within molecules.
Characterize molecular and receptor-ligand interactions and protein (un)folding events. Target individual molecules.



ForceRobot measurement head

Fully automated

200,000 Force curves per day

Powerful software for efficient data collection, processing, and evaluation. Automated laser and detector alignment. Standardized auto-calibration procedures.

Experimental settings

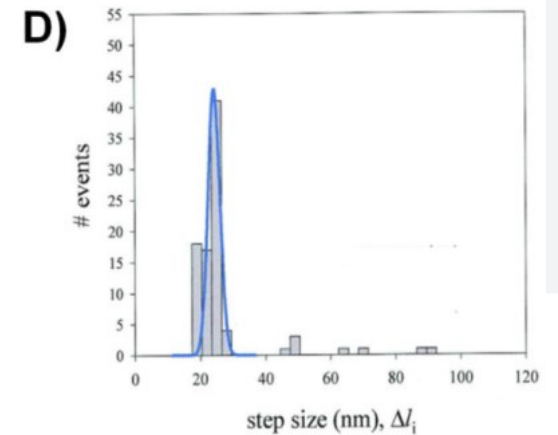
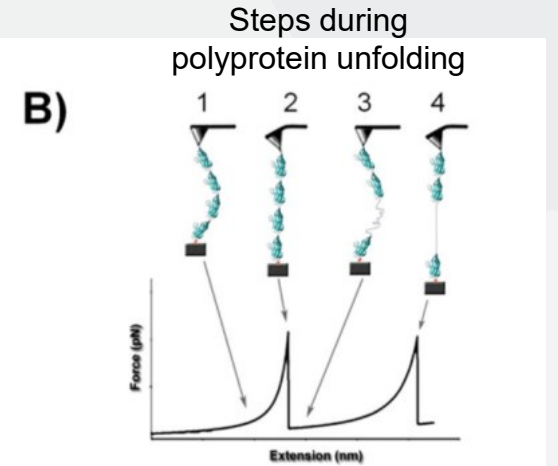
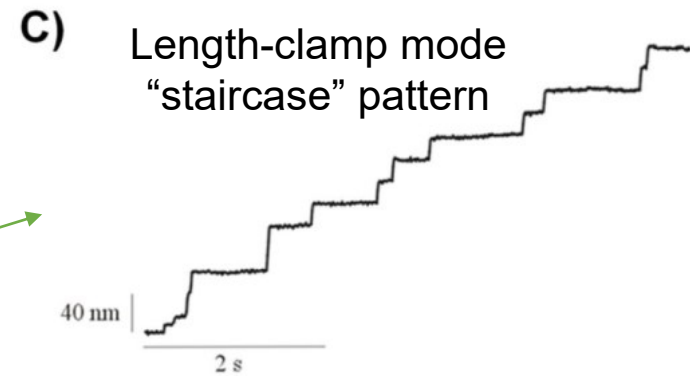
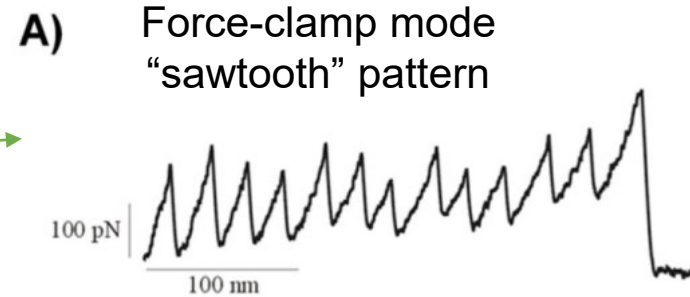
Modes of operation

■ **Force-clamp mode**

- *Protein folding/unfolding; Mechanisms of enzyme catalysis*
- *Force measurement with high speed of feedback gain – 0.1 to 5 milliseconds*
- *Typical settings: $F = 50\text{-}500\text{ pN}$, pulling speed $1\text{ }\mu\text{m/s}$*

■ **Length-clamp mode**

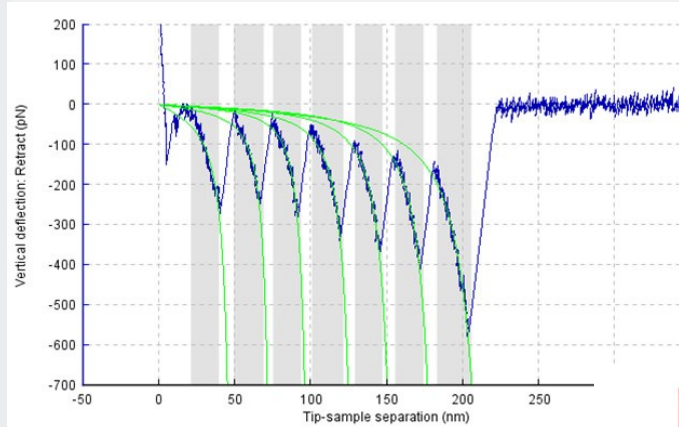
- *Difficult interpretation*
- *For multidomain proteins (native multi-domain proteins (such as titin, tenascin or spectrin))*
- *Typical settings: $F = 50\text{-}500\text{ pN}$, pulling speed $1\text{ }\mu\text{m/s}$, no feedback driving*



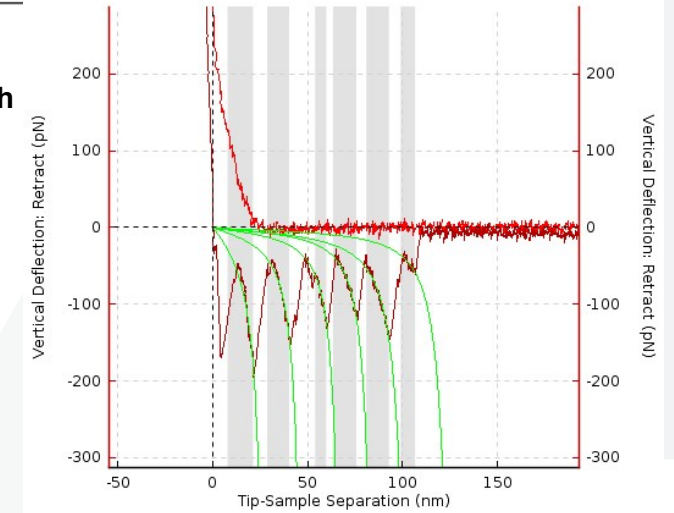
Data processing

JPK Data Processing software

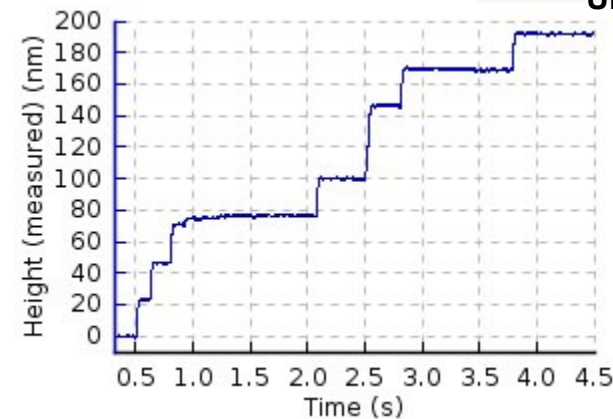
- **Predefined functions**
- **Batch processing of curves**
- **Data sorting**



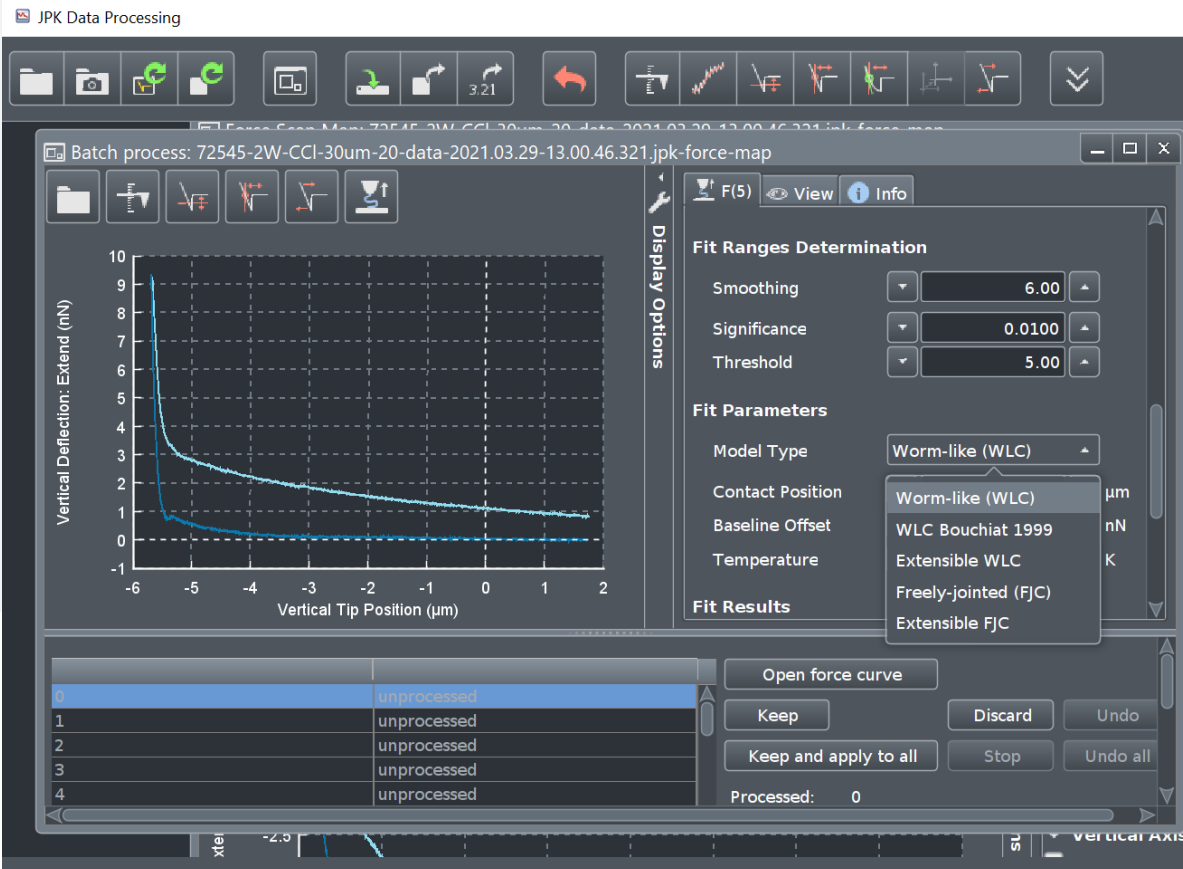
Fitting of force-extension profile with worm-like fit



Unfolding of artificial polyprotein GB1

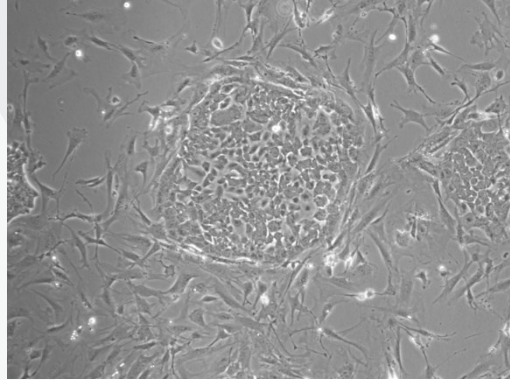


Unfolding of individual titin domain

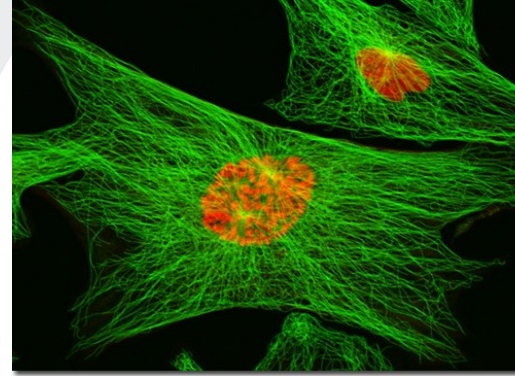


Nanomechanical mapping of living cells

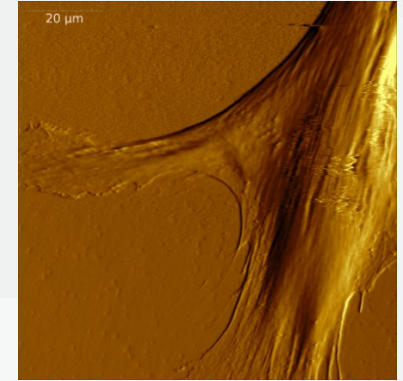
Optical microscopy



Confocal microscopy



AFM



Young's modulus mapping

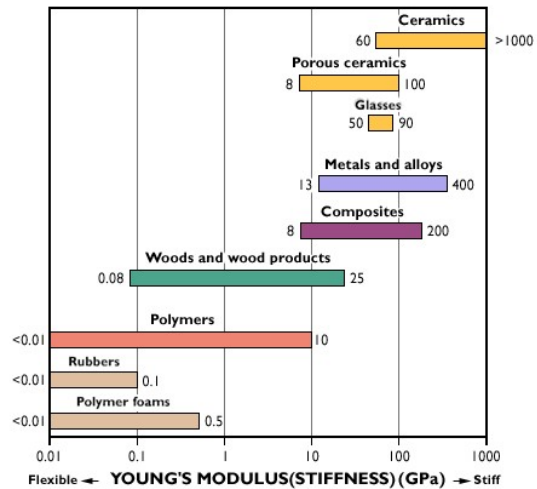


Motivation

Why to quantify elasticity of (living) objects?

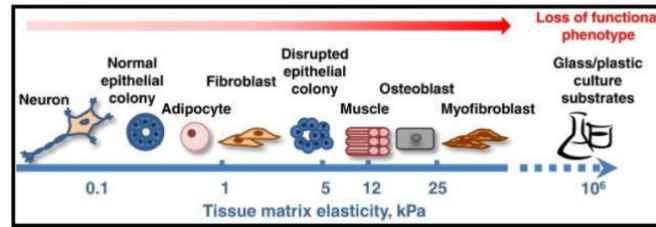
- **Stiffness** (Young's modulus) mapping
→ stiffness = basic parameter of any material
- **Elasticity-phenotype** relation ship
- **Mechanobiological** characterization
- **Driving of instrument** properties (QNM, QI)

Young's modulus (YM) of materials

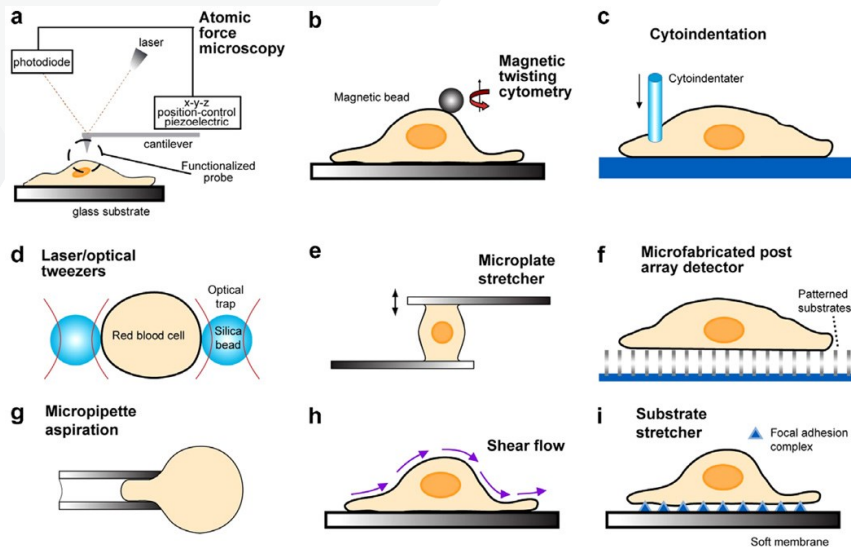


Tissue's Young Modulus

Tissue elastic modulus (E) is given by the resistance offered by the tissues to deformation effects, i.e. the tissue stiffness.



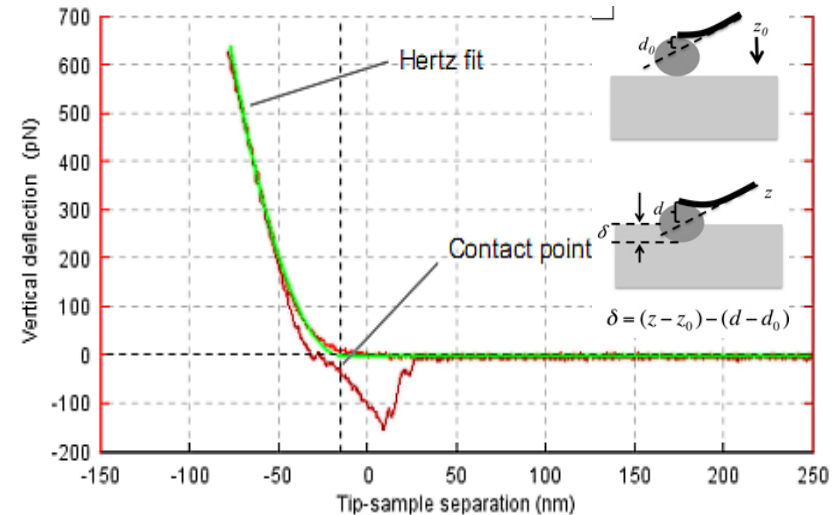
www-materials.eng.cam.ac.uk/



+ Holographic Q-phase microscope

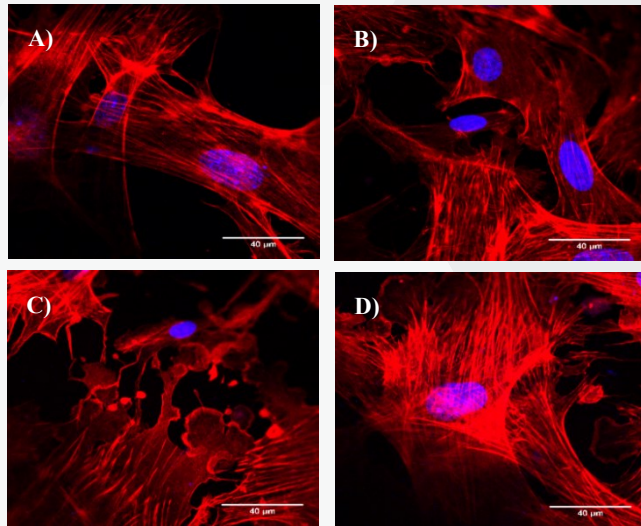
Acta Biomater. 2007 Jul; 3(4): 413–438.

Methods for YM measurement



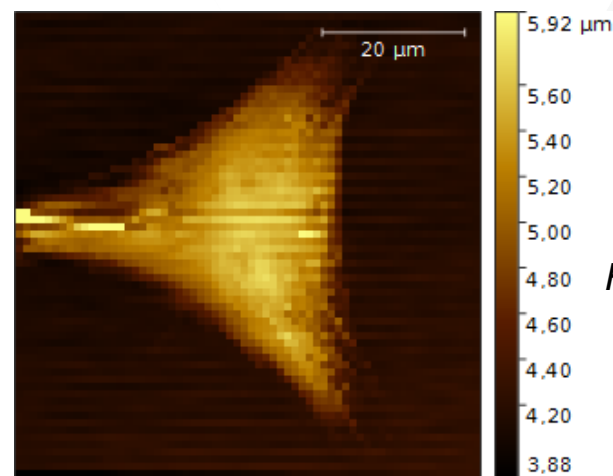
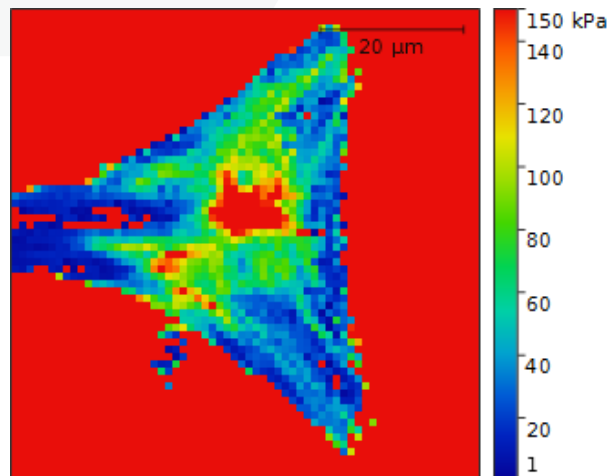
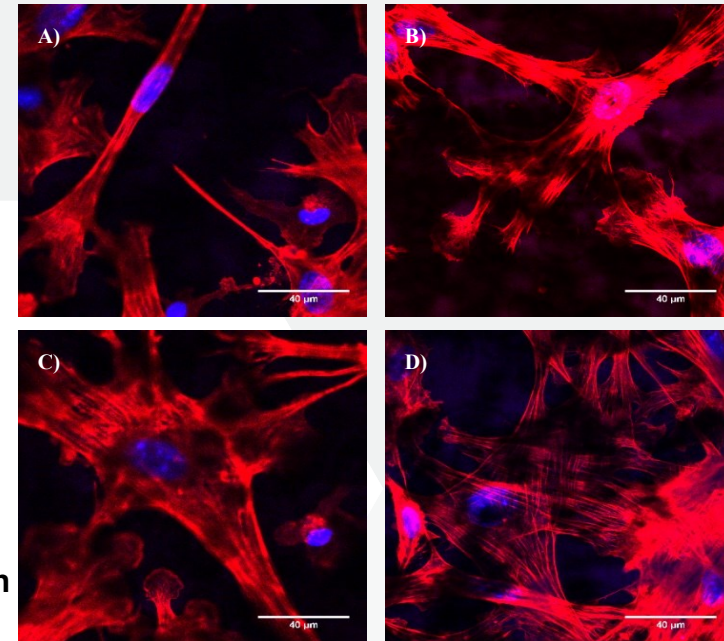
J. Vis. Exp. (76), e50497, doi:10.3791/50497 (2013).

Confocal vs. AFM



Confocal microscopy

- DAPI nucleus staining
- Actin staining by Phalloidin

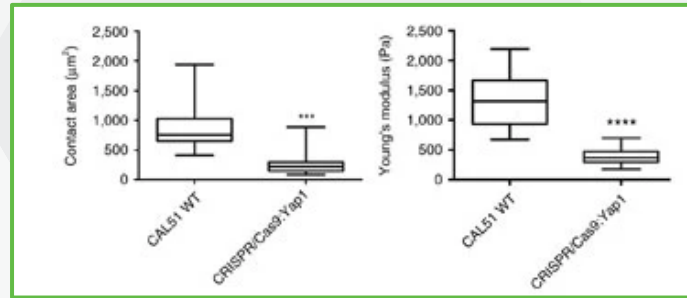
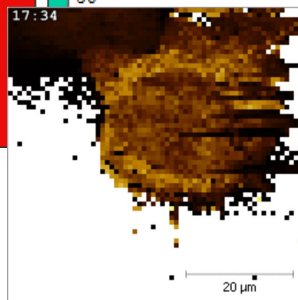
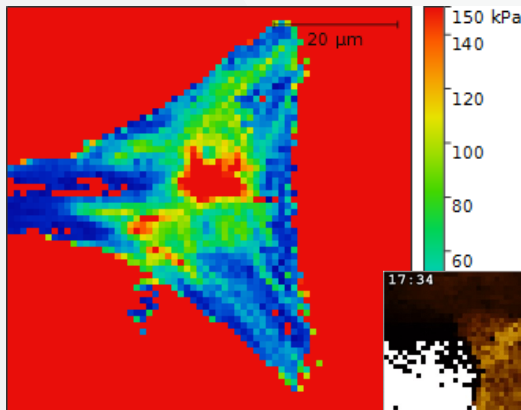
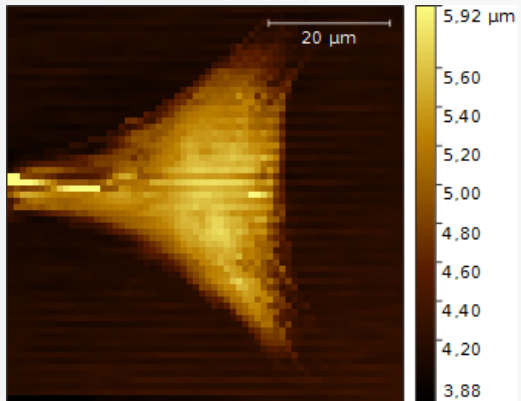


Force Mapping on single cells

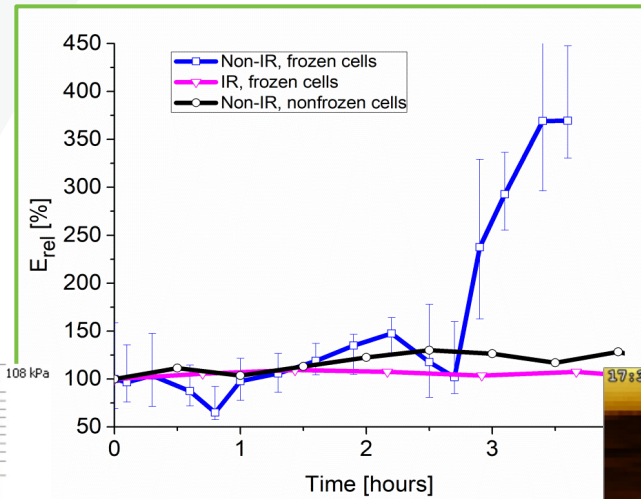
Cellular nanomechanics

By means of AFM

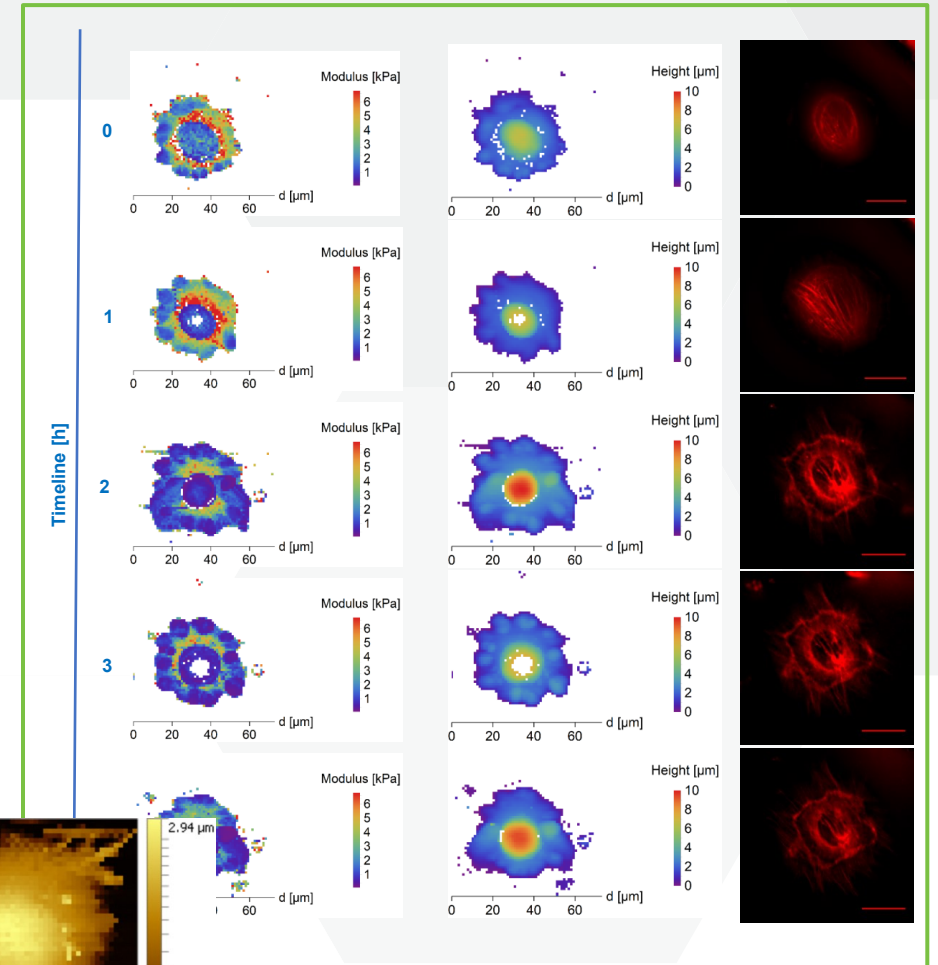
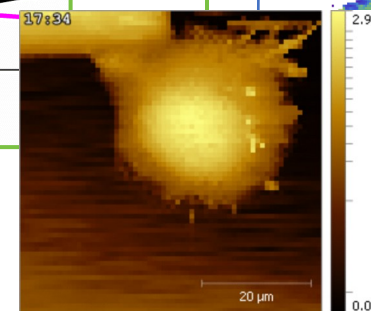
AFM mapping - correlation with fluorescence microscopy



Evaluation - statistics

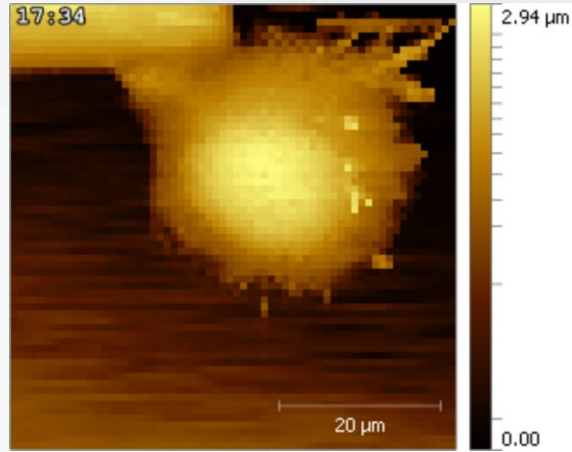


Time-lapsed biomechanics

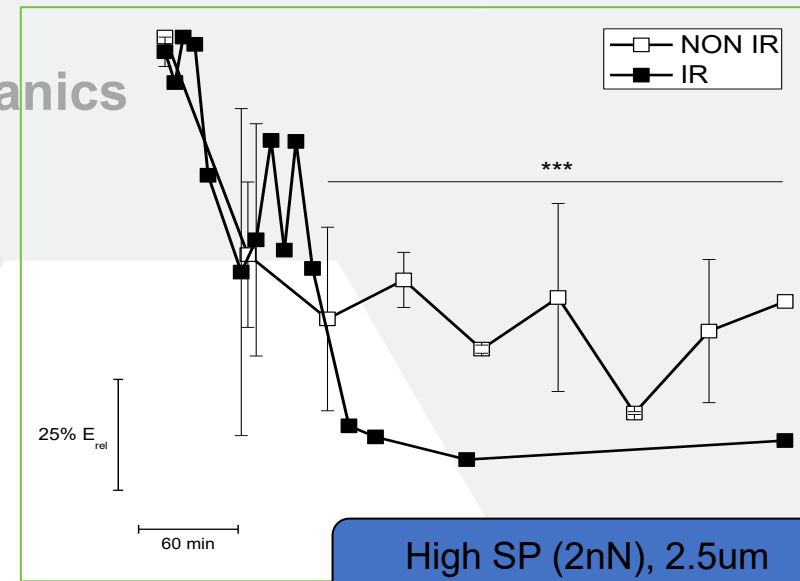


Fibroblasts thawing process

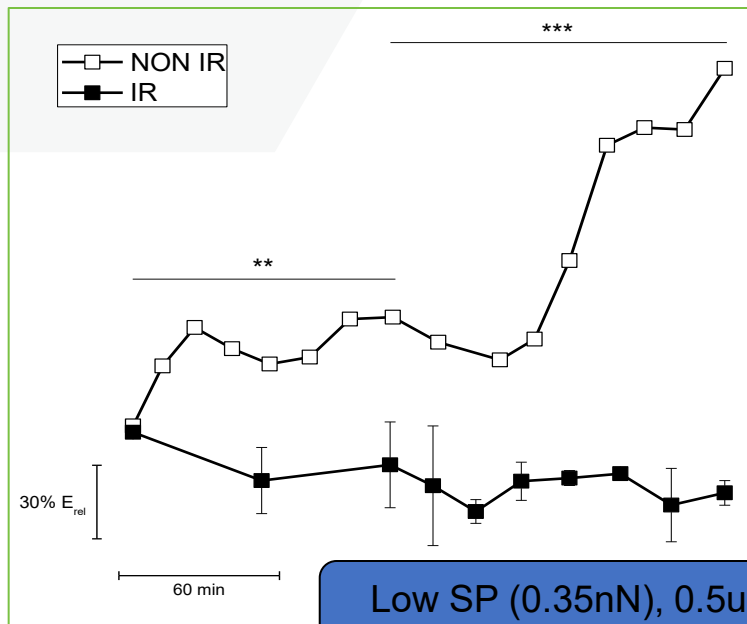
Time-lapsed monitoring of cell biomechanics



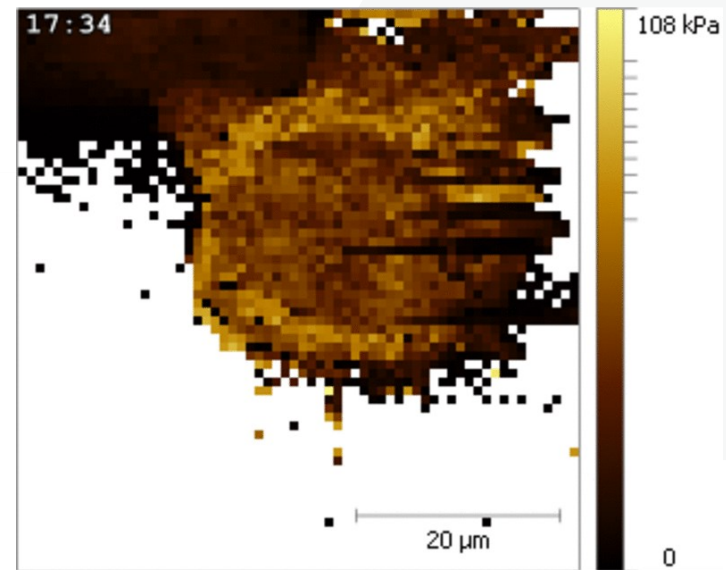
SP height profile (12 hours)



High SP (2nN), 2.5 μm indent. Depth (35%)

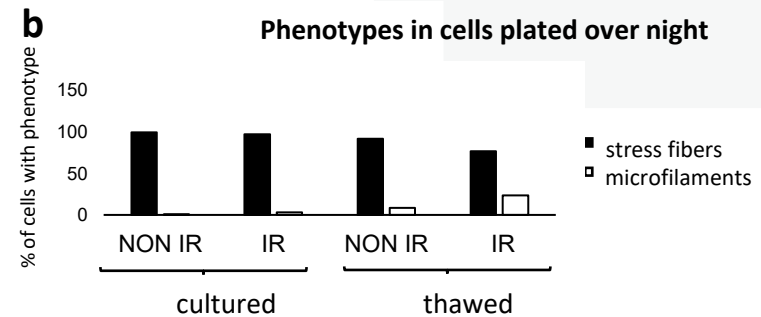
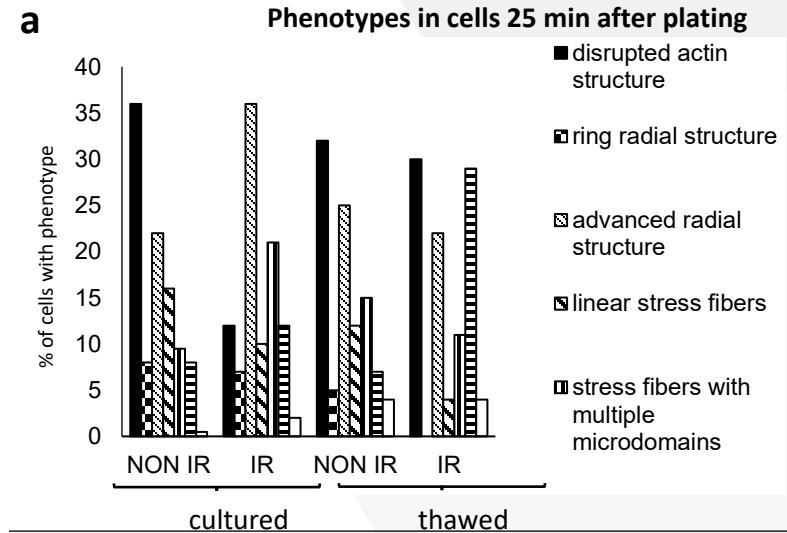
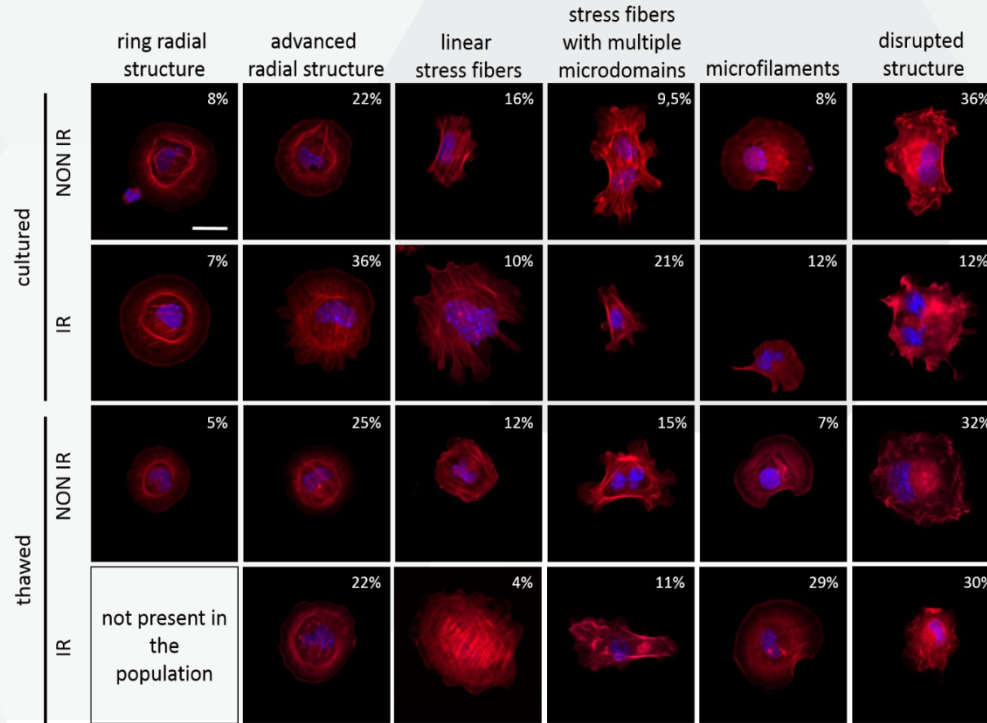


Low SP (0.35nN), 0.5 μm indent. Depth (6%)

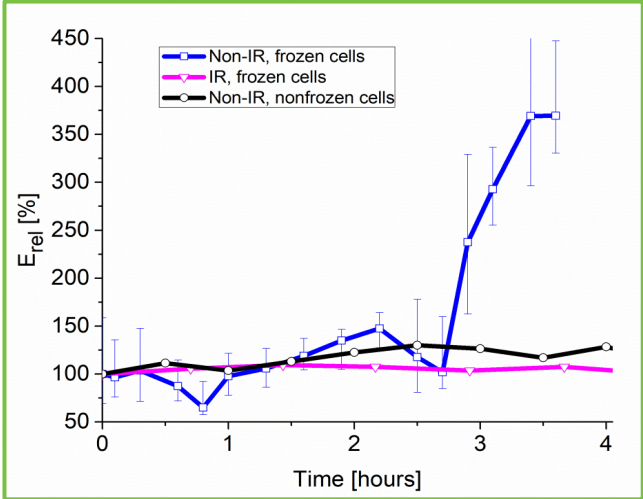
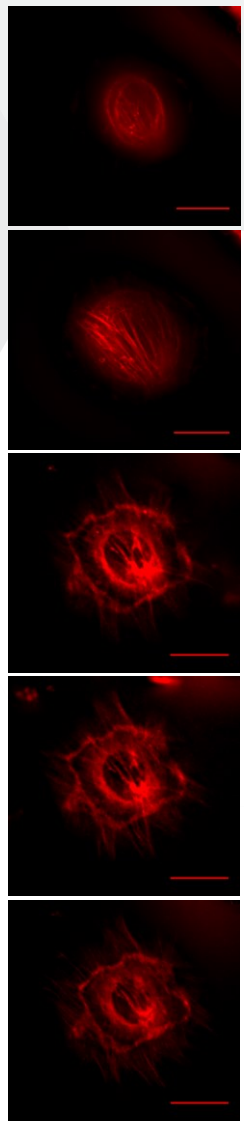
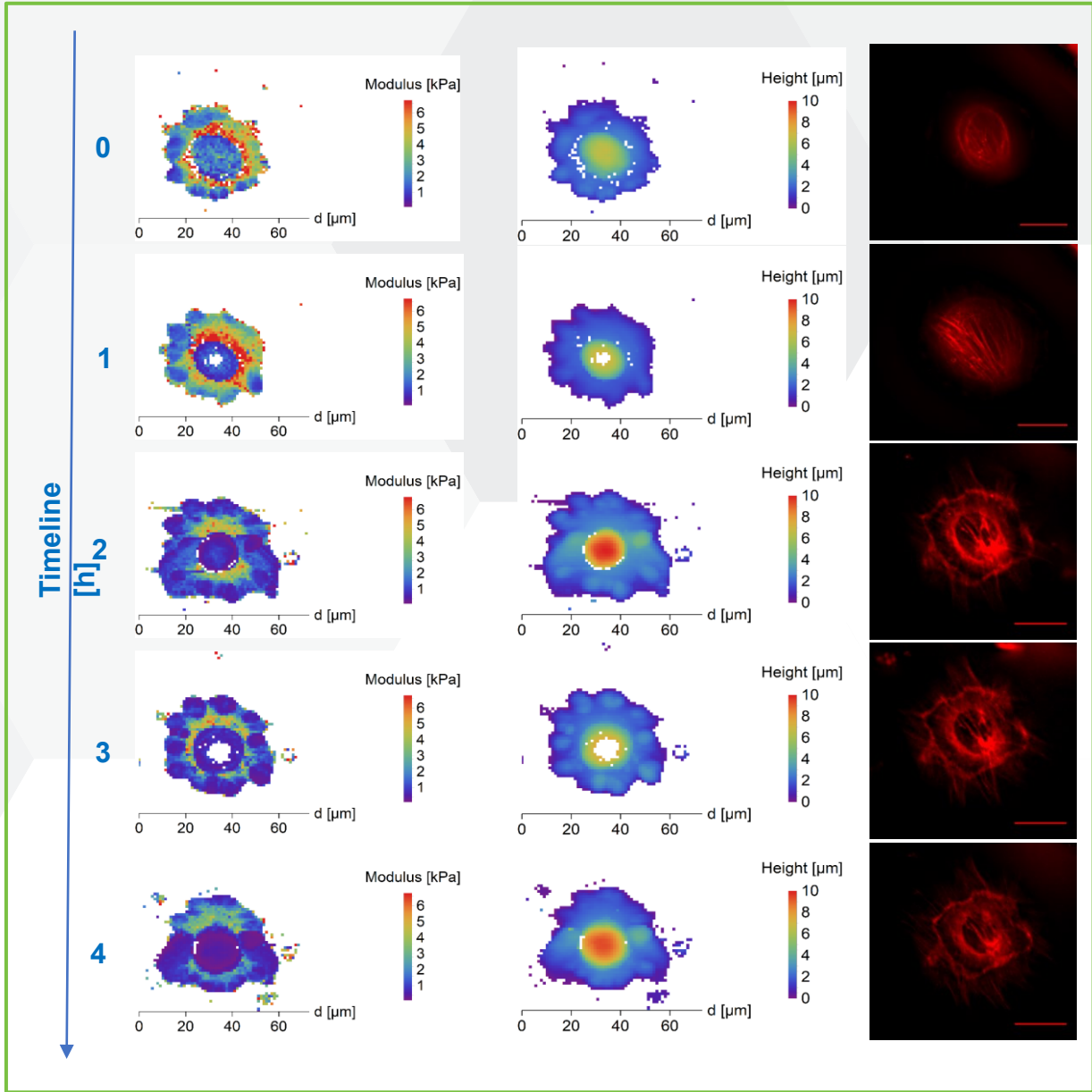


YM maps (12 hours)

Phenotypes in cells 25 min after plating

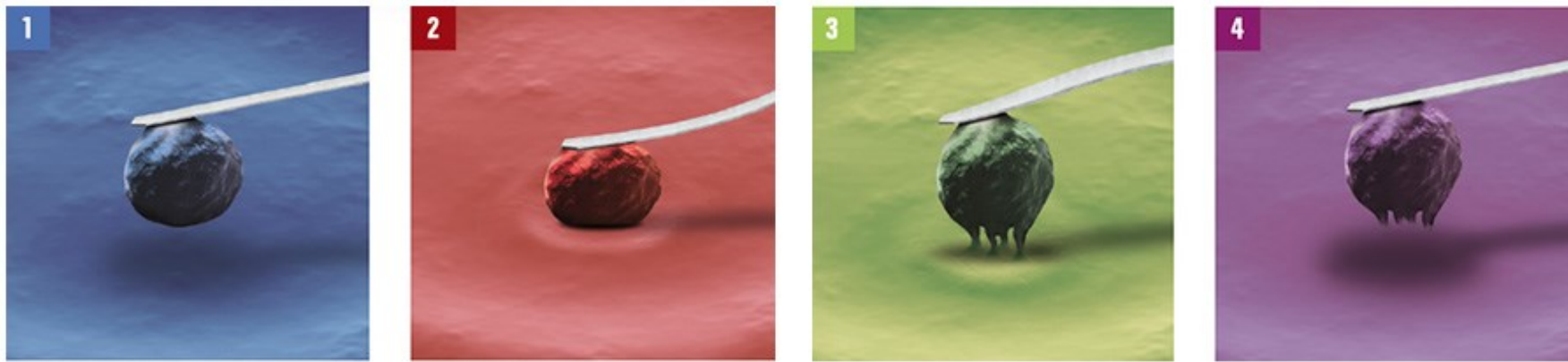


Cytoskeleton staining
(Phalloidin/DAPI)

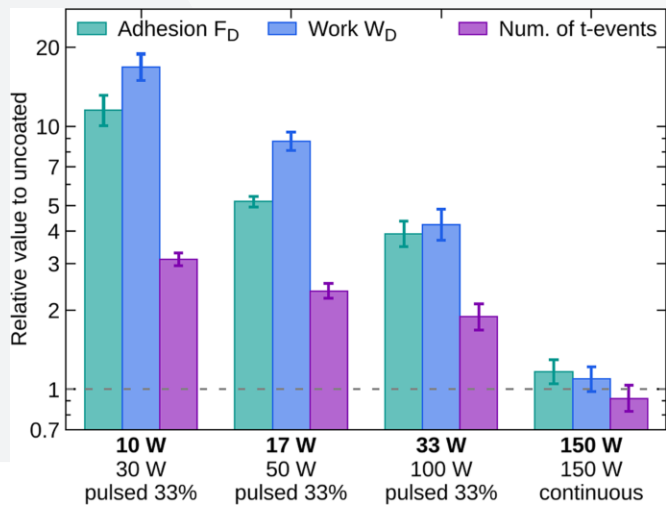


SiR
live actin staining

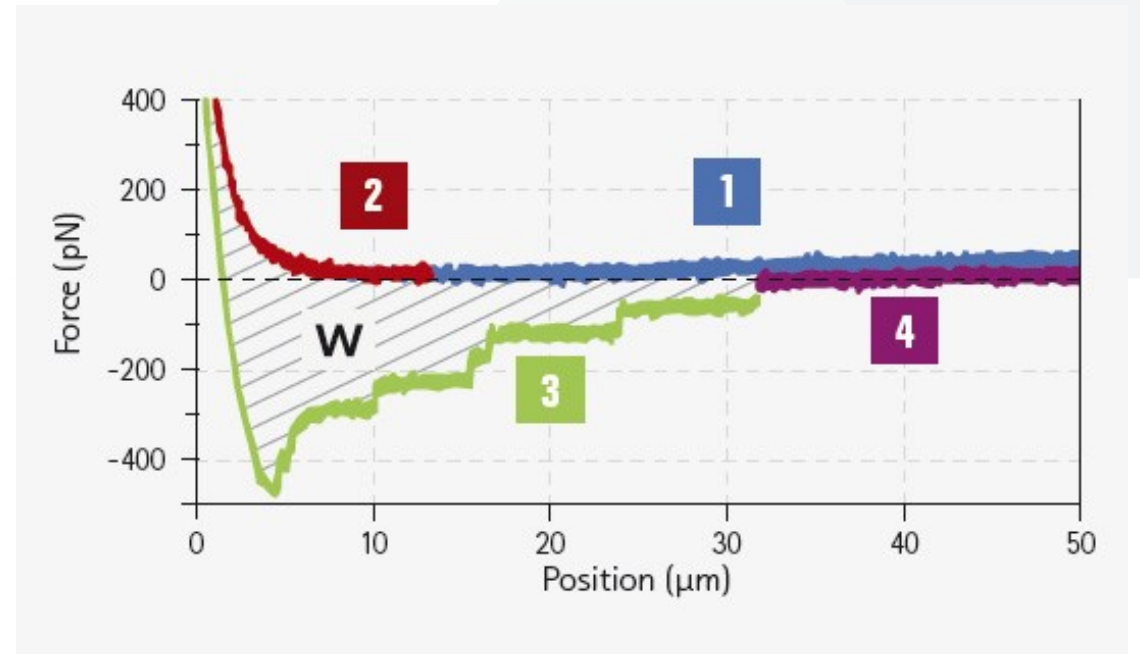
Cell-adhesion experiments



www.jpk.com



Relative adhesion of cells to plasma modified surfaces

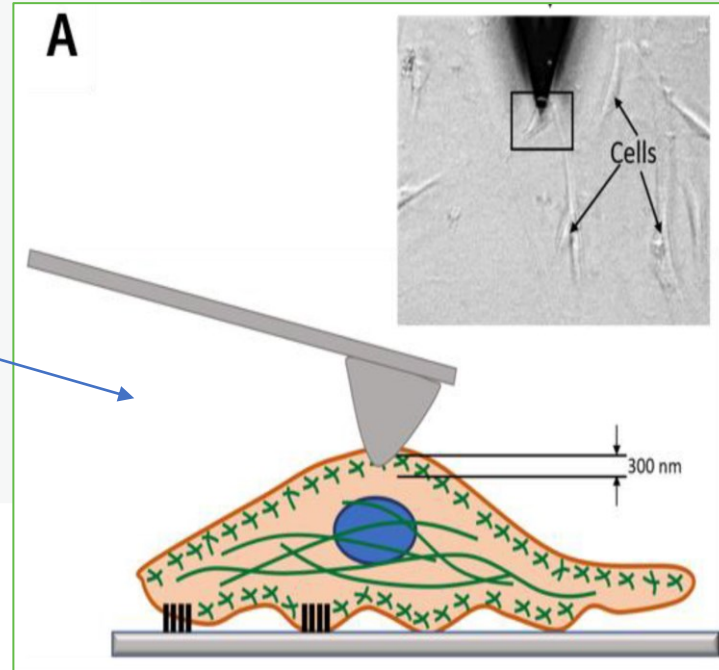


Černochová, P., Blahová, L., Medalová, J. *et al.* Cell type specific adhesion to surfaces functionalised by amine plasma polymers. *Sci Rep* **10**, 9357 (2020). <https://doi.org/10.1038/s41598-020-65889-y>

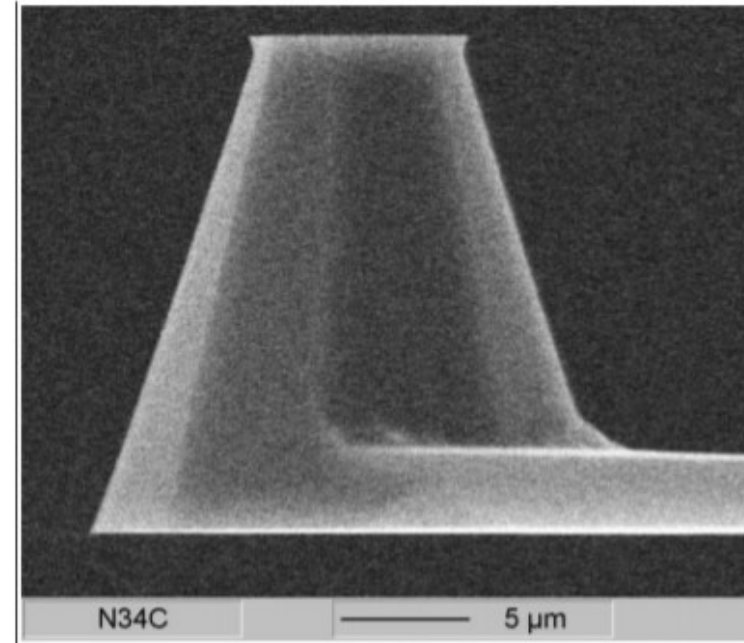
Other applications of AFM-based biomechanics

Cell scratching = cell adhesion

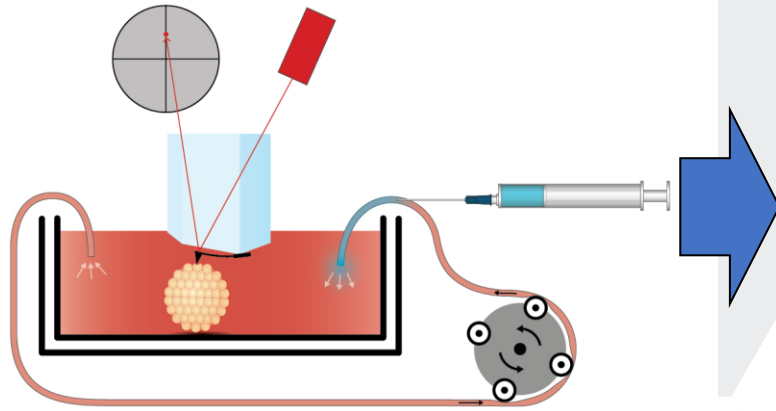
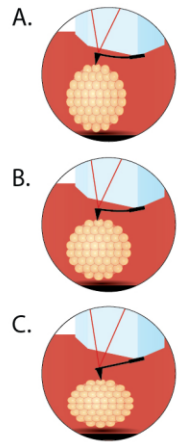
Scratching movement
Force variation



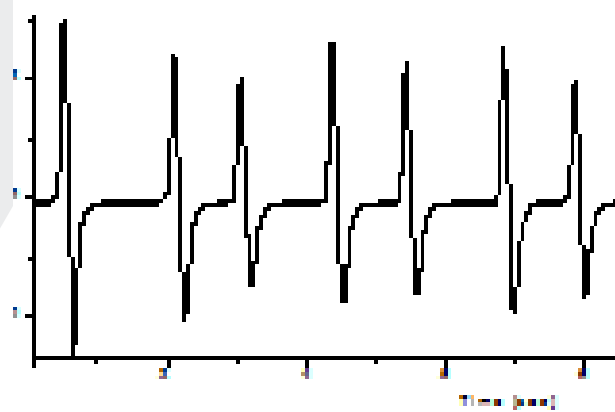
Large Plateau AFM Tips



Other applications of AFM-based biomechanics

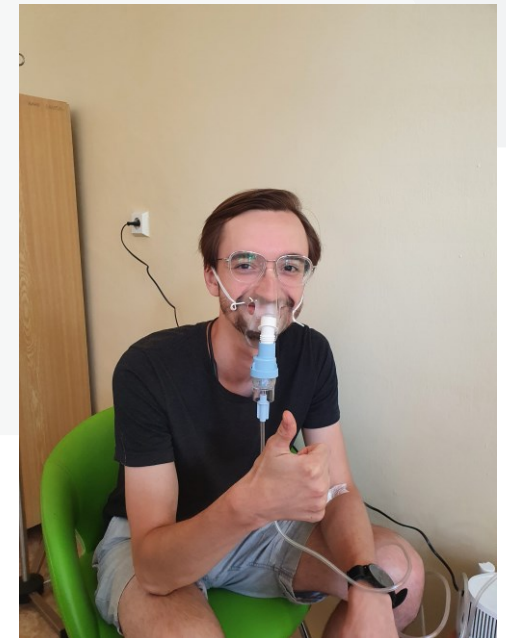
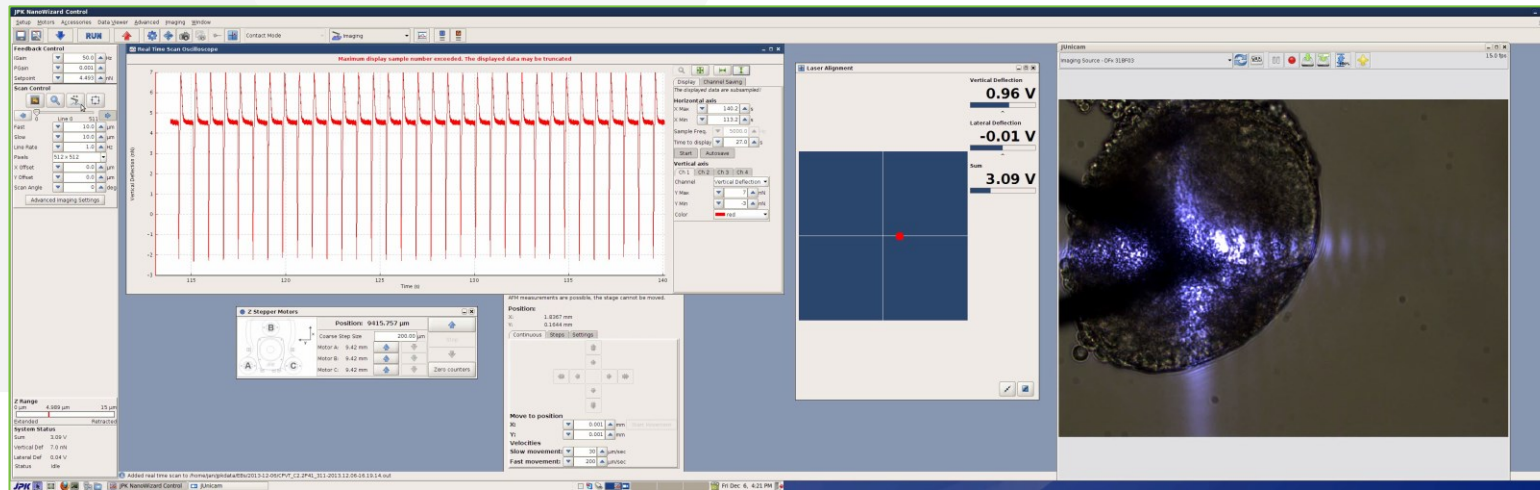


Setup scheme



MCG = mechanocardiogram

Cardiac cells biomechanics

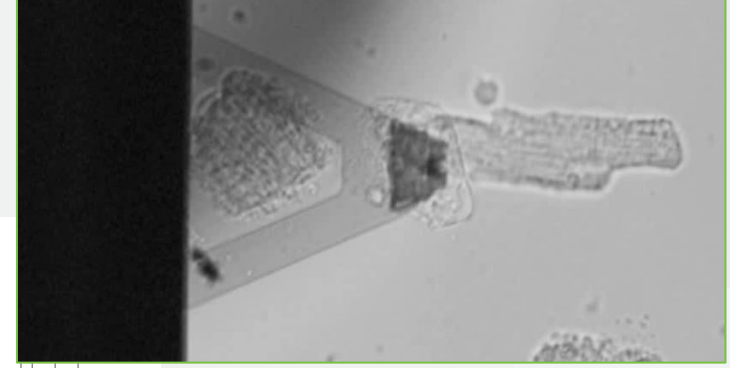


Pesl M, Pribyl J, et al. 2016 *Biosensors and Bioelectronics* **85** 751–7

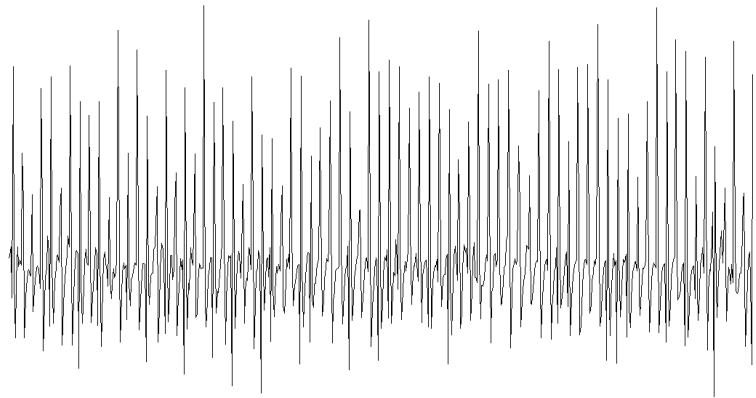
Pesl M, Pribyl J, et al. 2016 *J Mol Recognit* n/a-n/a

Pesl M, Acimovic I, Pribyl J, et al. 2014 *Heart Vessels* 29 834–46

Typical DFL vs. time curves recorded as a result of various drug treatment

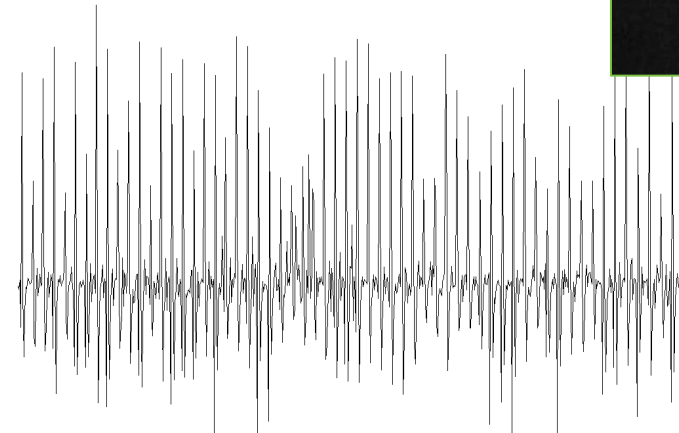


A



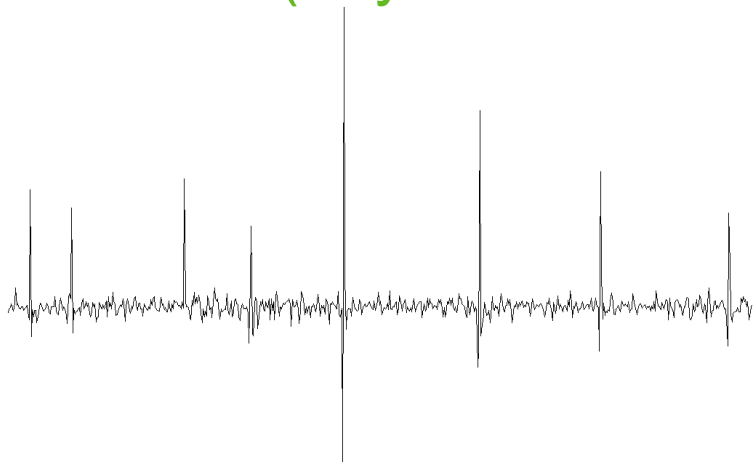
Base line (Thyroid medium)

C



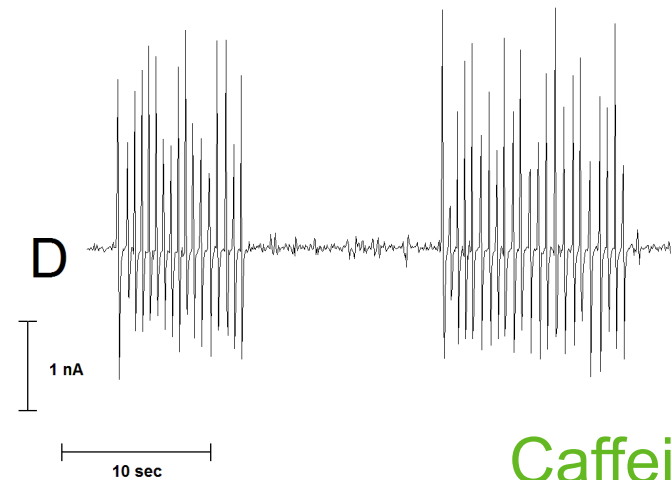
Isoproterenol (activator)

B



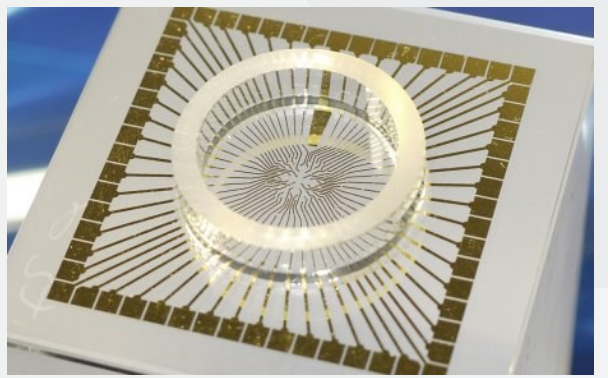
Metoprolol (betablocker)

D

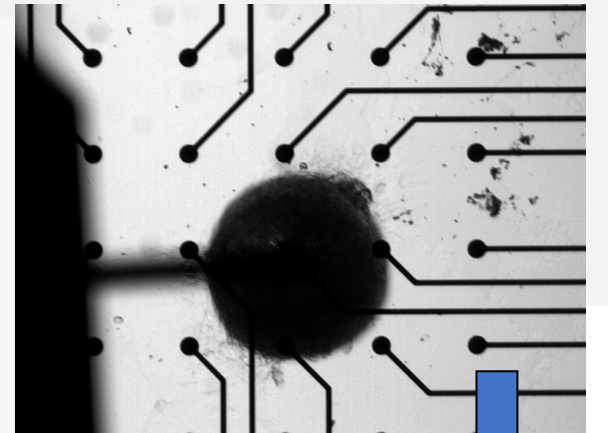


Caffeine

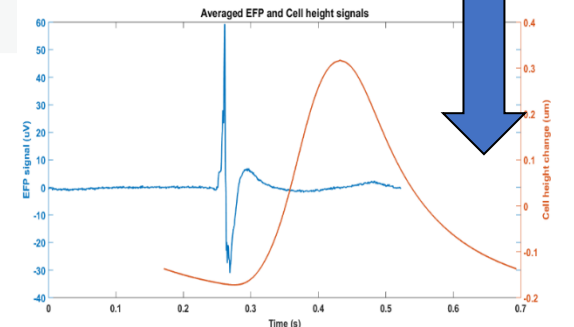
Combination with other techniques



MultiElectrode Array
Extracellular Cell Potential
Cardiac cells and Neurons

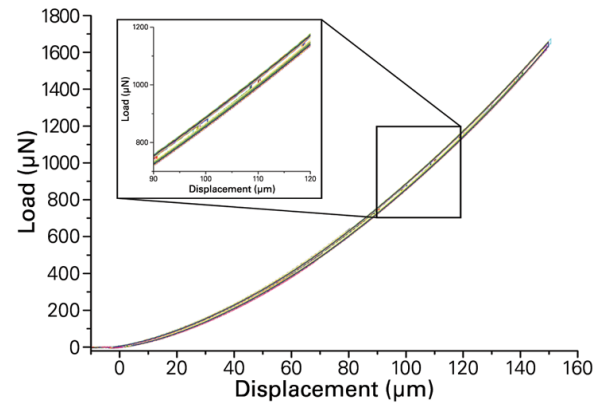
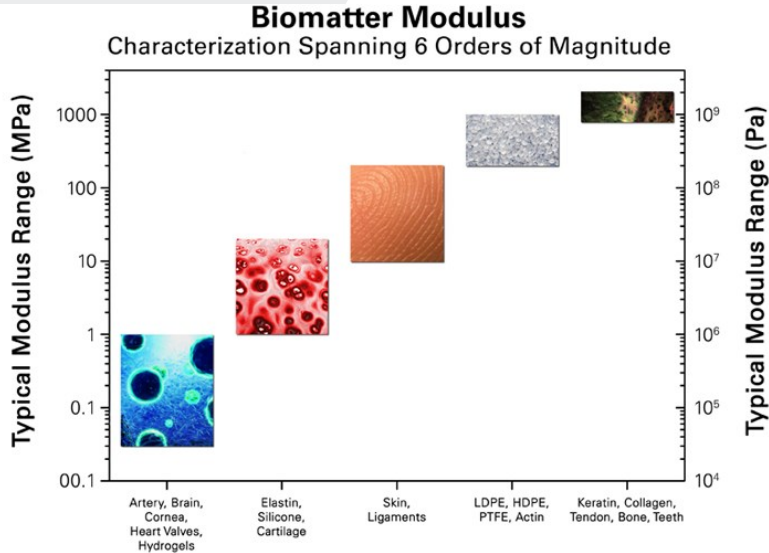


AFM + MEA
= electromechanical (de)coupling

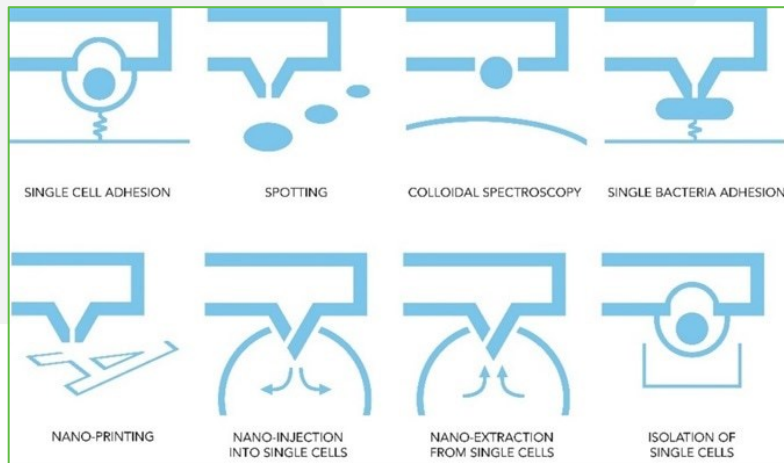
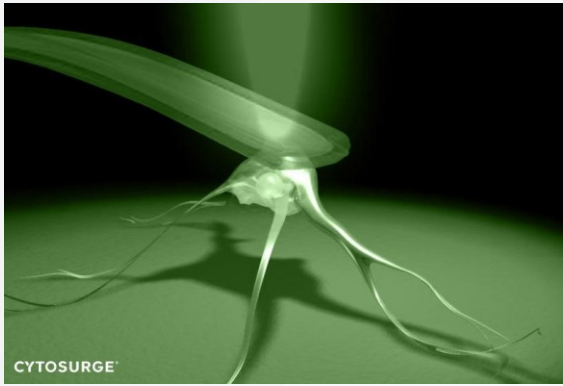


NanoIndentation

Single point indentation curves

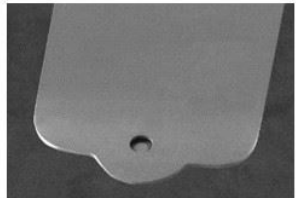
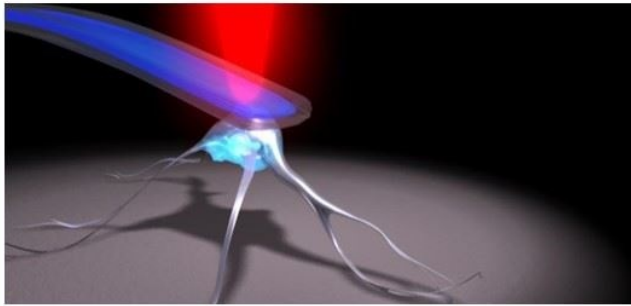


CytoSurge Fluid FM module



FluidFM – microfluidic force microscopy

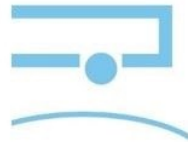
FluidFM micropipette



2, 4, 8 μm aperture

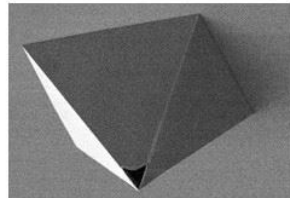
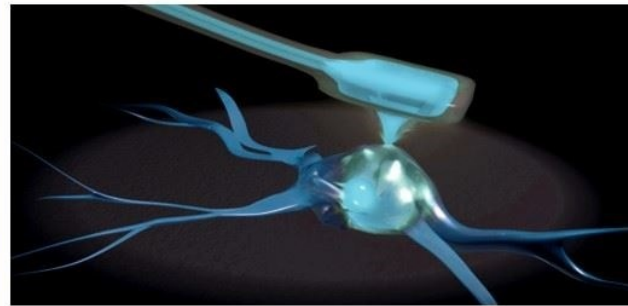


Adhesion of single cells



Colloidal spectroscopy

FluidFM nanosyringe



300 - 800 nm aperture

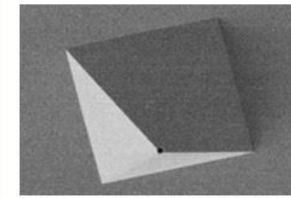
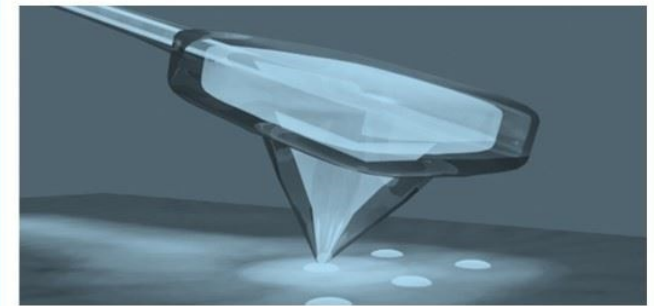


Nano-injection into single cells



Nano-extraction from single cells

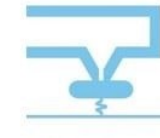
FluidFM nanopipette



300 nm aperture



Spotting



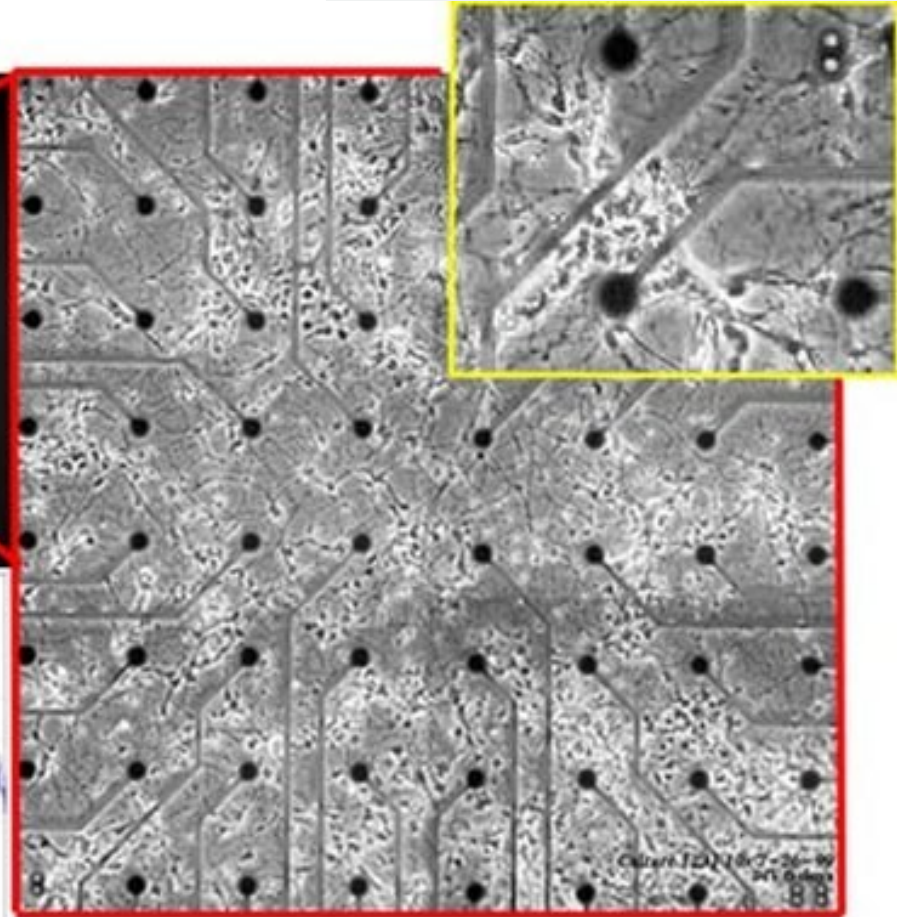
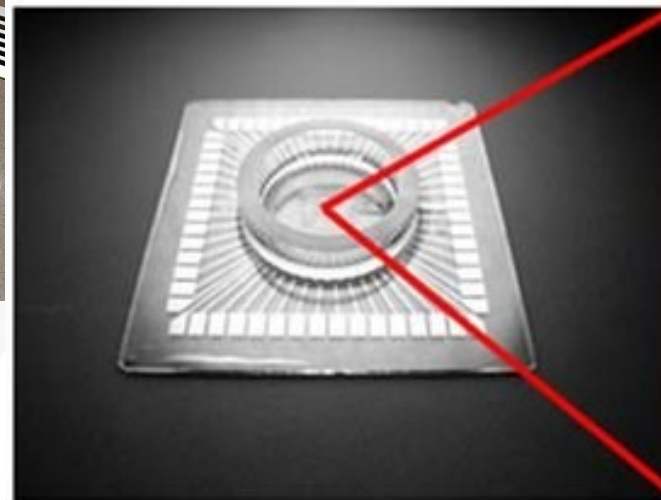
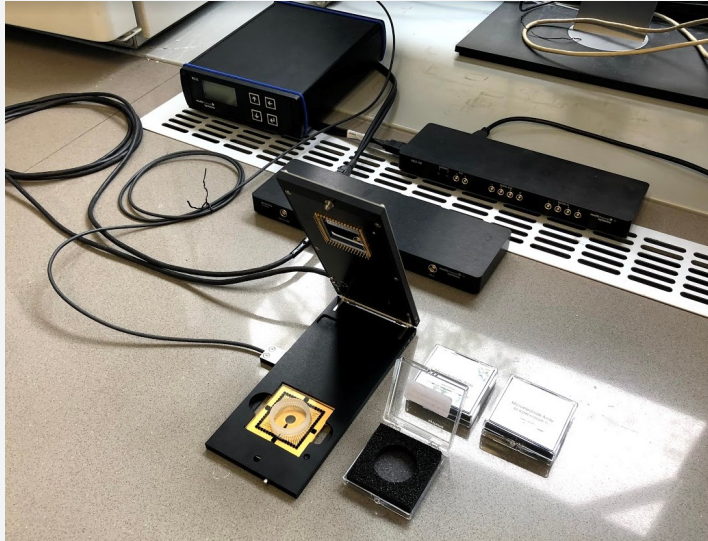
Adhesion of single bacteria



Nano-printing

<https://www.news-medical.net>

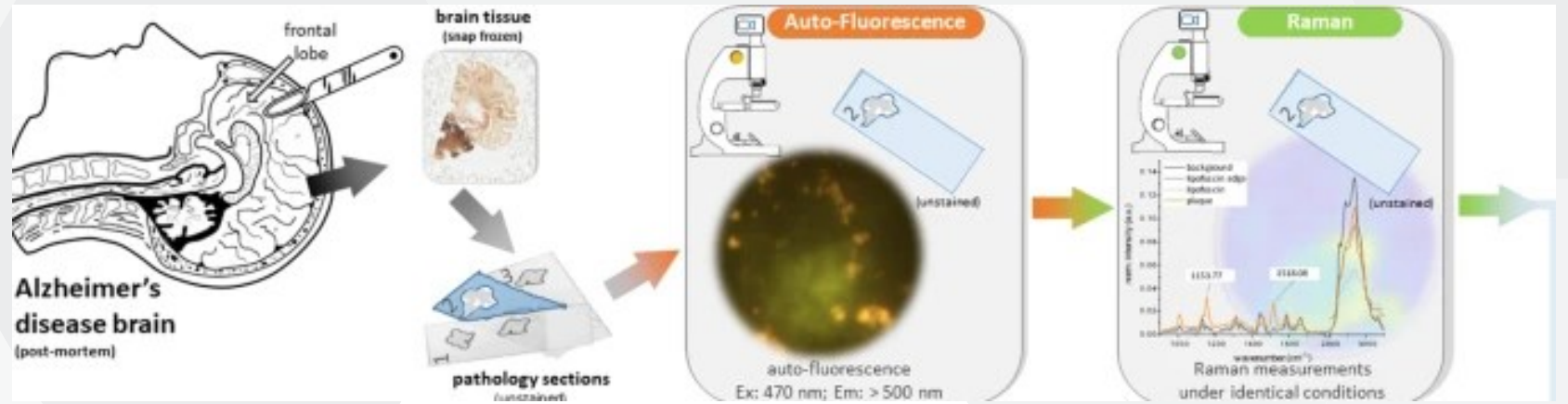
MEA – microelectrode array cellular electrophysiology



Raman microscopy

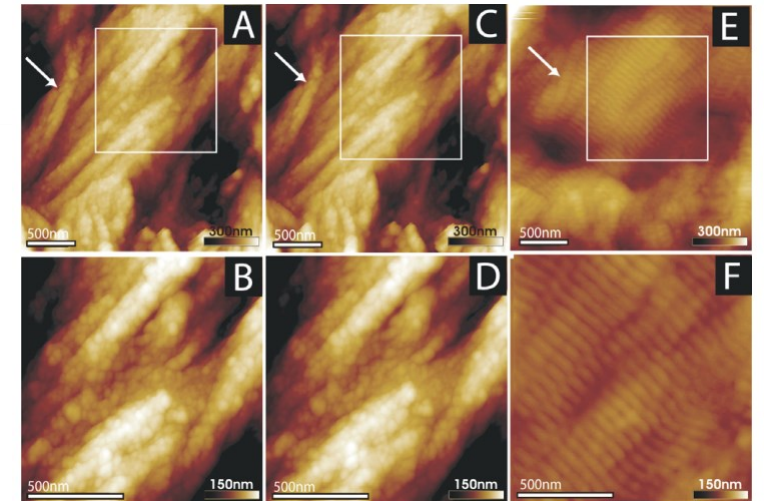
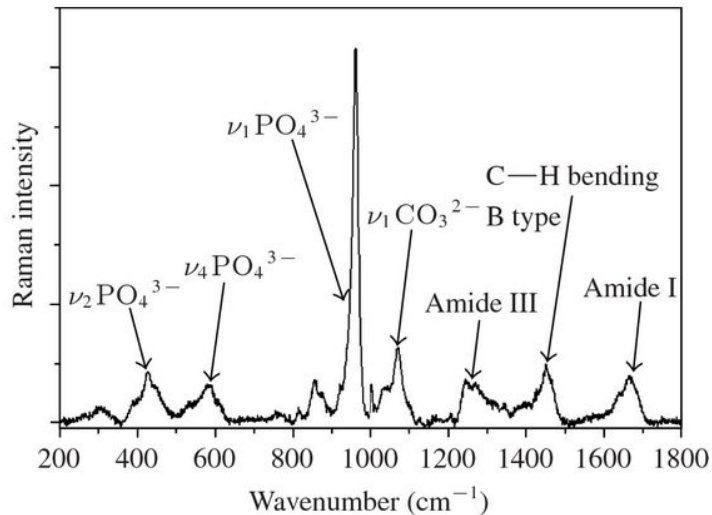
On bio samples

Lochocki, B., Boon, B.D.C., Verheul, S.R. *et al.* Multimodal, label-free fluorescence and Raman imaging of amyloid deposits in snap-frozen Alzheimer's disease human brain tissue. *Commun Biol* **4**, 474 (2021).



Raman imaging of **amyloid** deposits in snap-frozen **Alzheimer's disease** human brain tissue

Calcification level and Collagen Fibers Arrangement in Bone Tissue

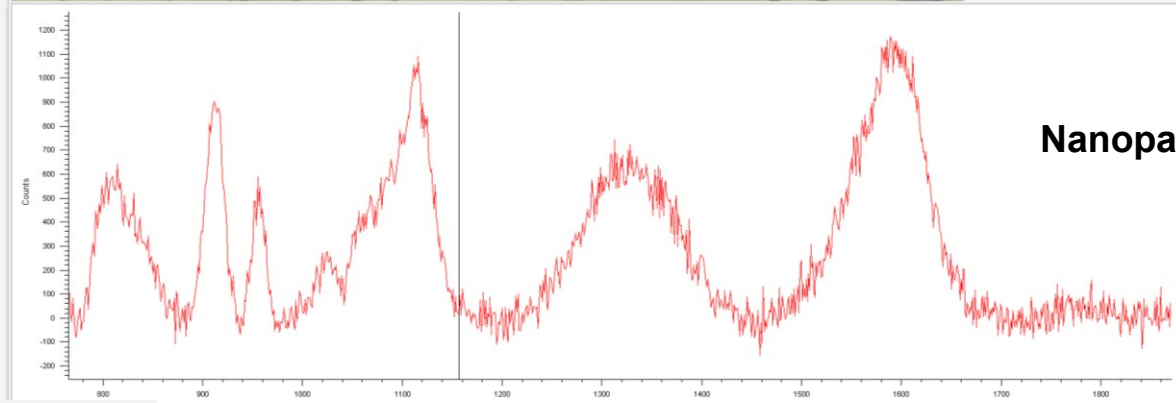
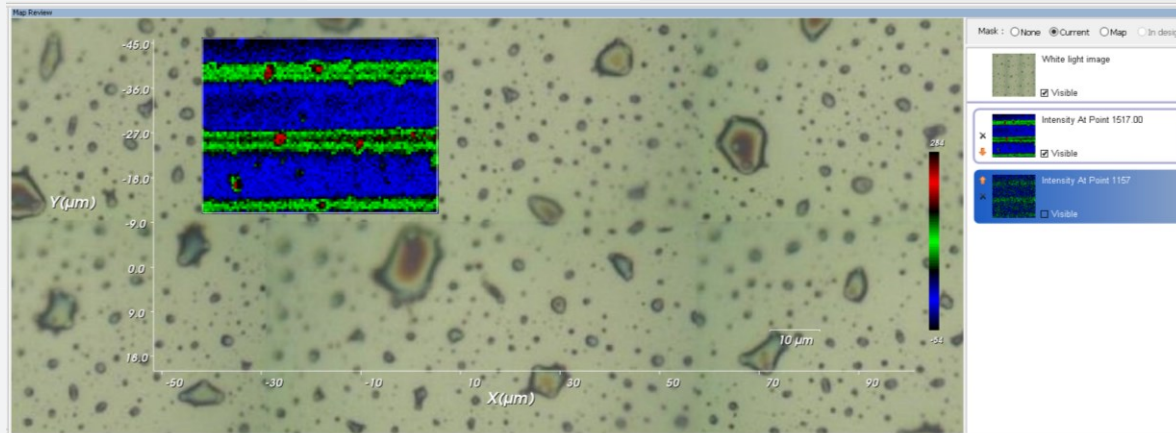


+ combination with AFM topography

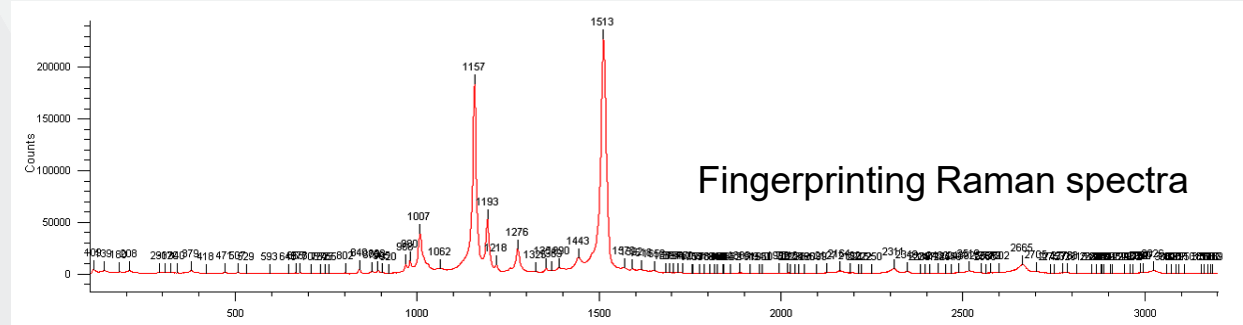


Raman microscopy

Chemical mapping

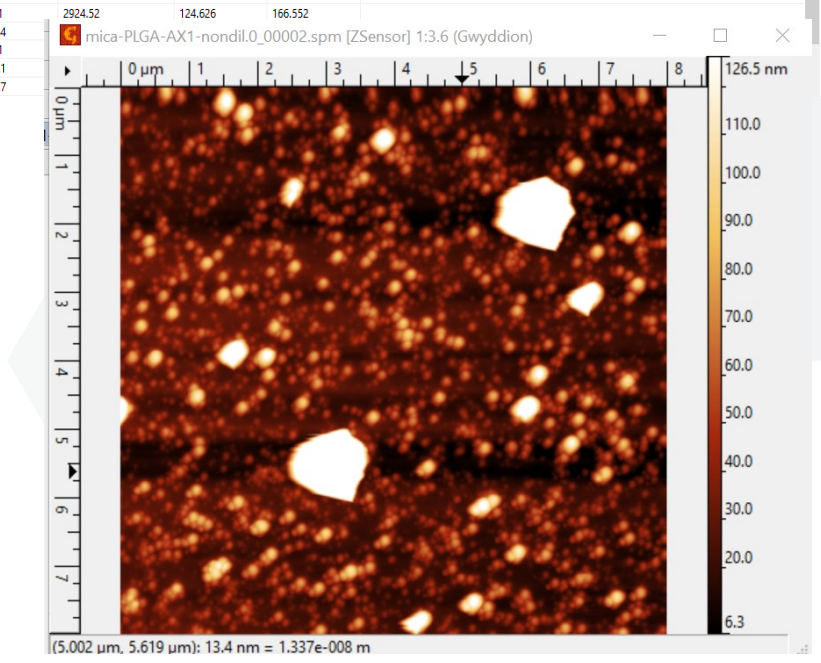


Nanoparticles loading study



Fingerprinting Raman spectra

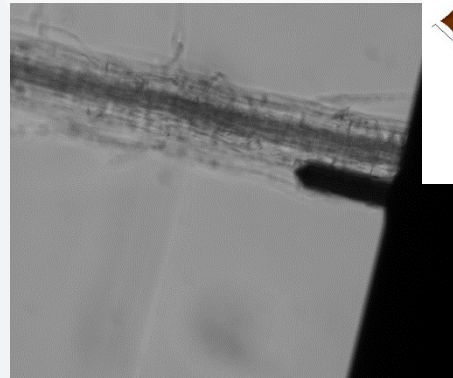
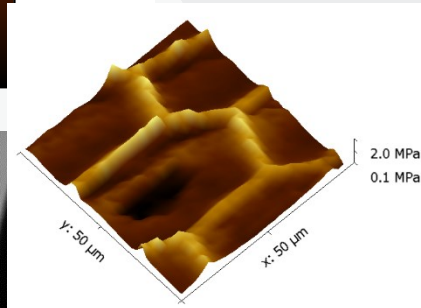
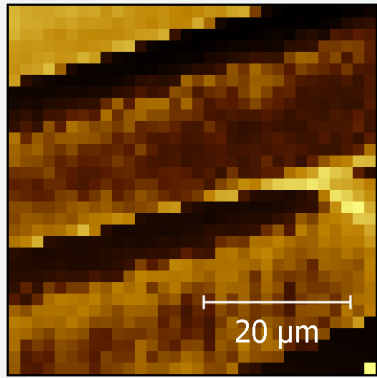
Peak no.	Centre	Height	Width	Area	Absolute intensity	Low edge	High edge
1	108.873	3059.03	9.28978	78556.2	4036.61	102.247	123.312
2	139.016	1668.17	14.5499	122921	2924.52	124.626	166.552
3	180.065	804.539	9.00485	55816.4			
4	208.109	1870.6	14.0121	110061			
5	291.483	692.559	7.83973	51650.1			
6	306.84	476.022	6.30329	31306.7			



+ combination with AFM topography

AFM-based biomechanics

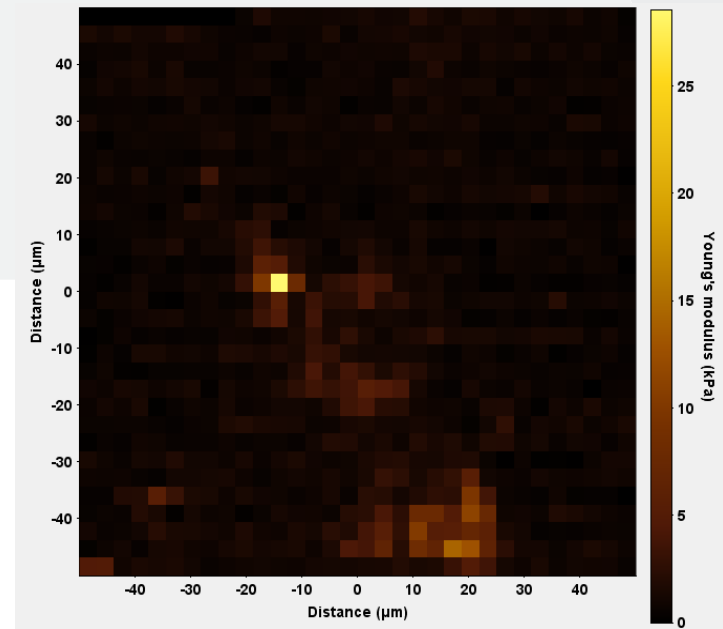
On a tissue level



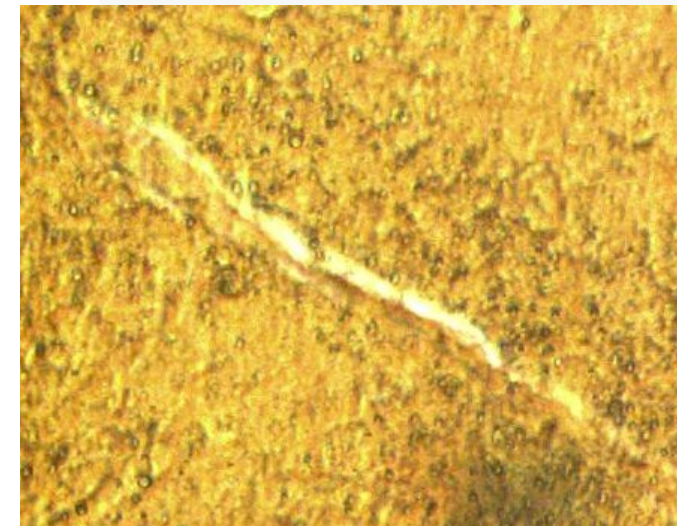
Plant samples
(hypocotyl)



by **Marçal Gallemí**
Eva Benkova Lab
& Jan Hejtko Lab



Liver cirrhosis
Correlation of
Collagen fibers by polarized microscopy
AFM nanoindentation



by **Srikant Ojha**
Martin Gregor Lab

Core Facility NanoBiotechnology

Standard operations of the CF

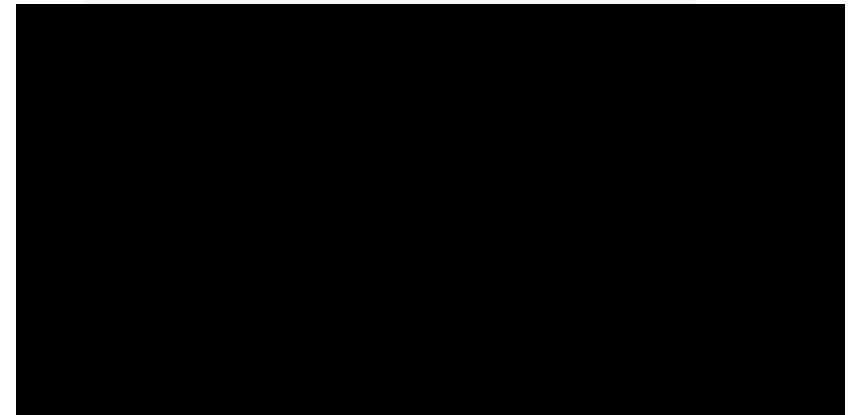
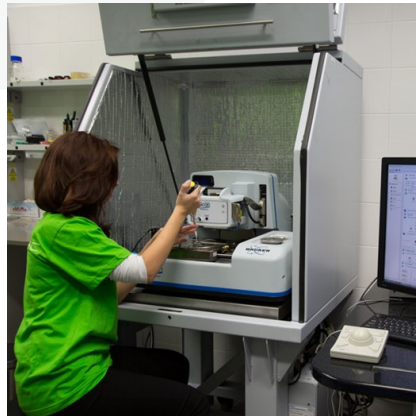
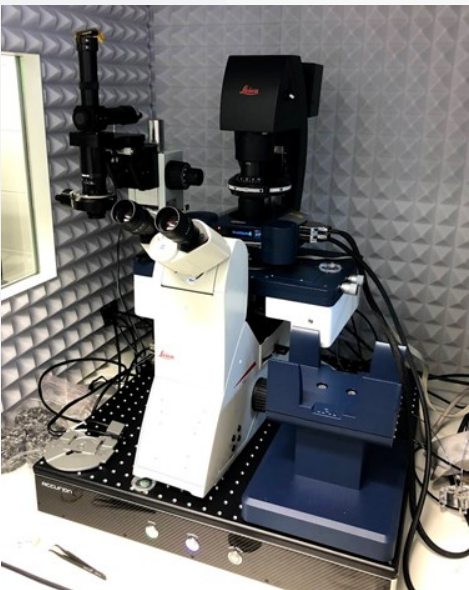
- **BioAFM microscopy** – biomolecules, nano-objects
- **Stiffness mapping** – cells, tissues
- **Combination** of AFM with other techniques (BF/fluorescence microscopy)

2020/21 Innovations

- **FluidFM** – microfluidic force microscopy
- **Raman** chemical mapping
- **MEA** – micro-electrode array cell electrophysiology



Core Facility NanoBio



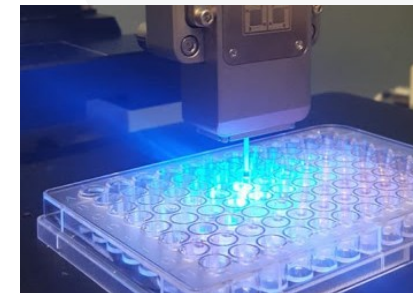
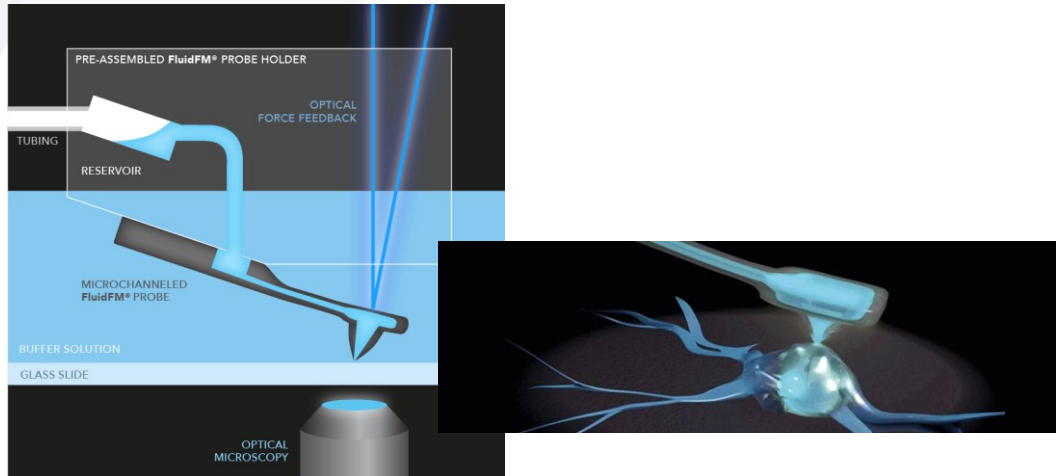
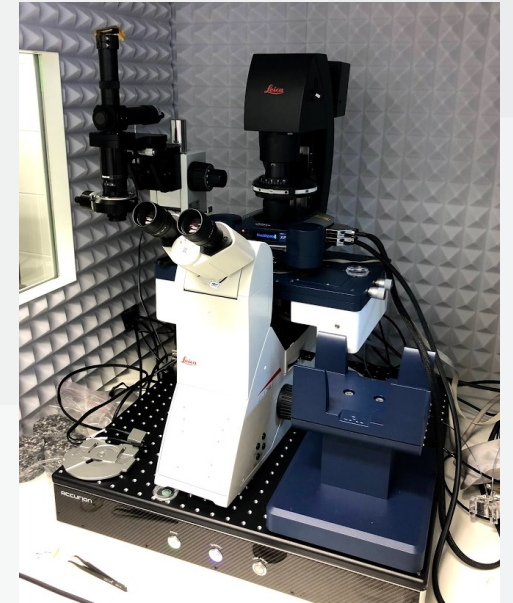
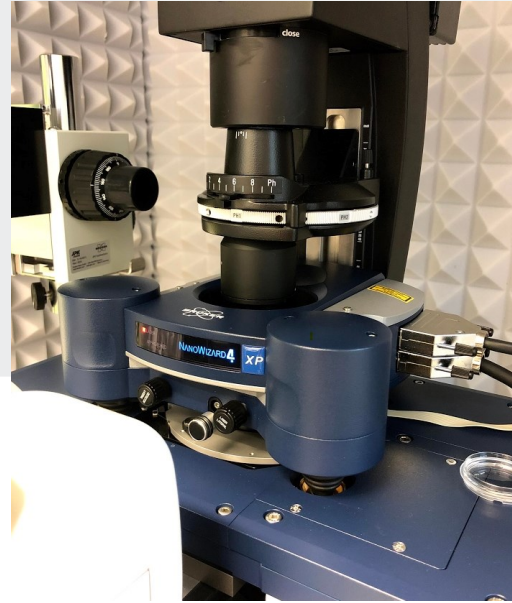
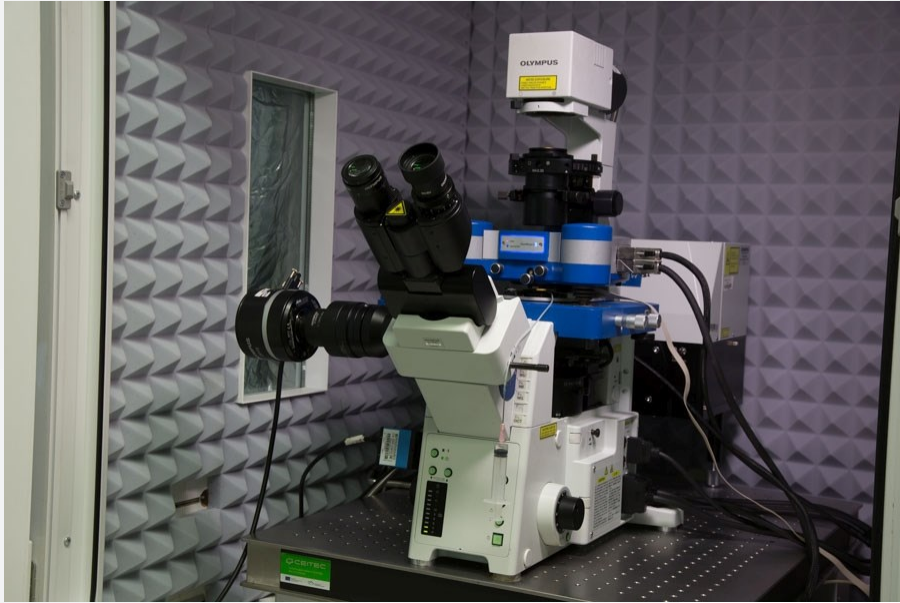


JPK
Instruments

Core Facility
Nanobiotechnology
XCIISB

JPK NanoWizard 3 and 4 with extended scanning range

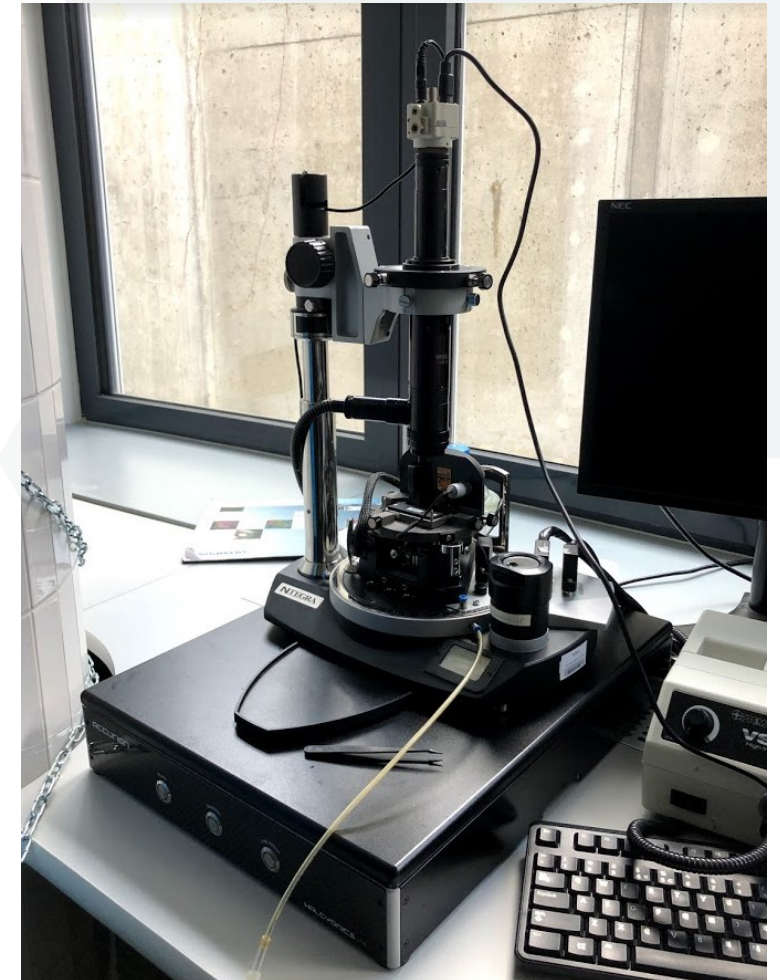
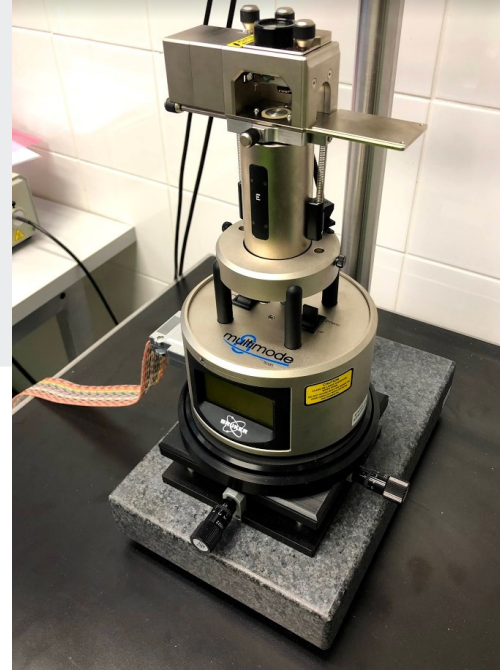
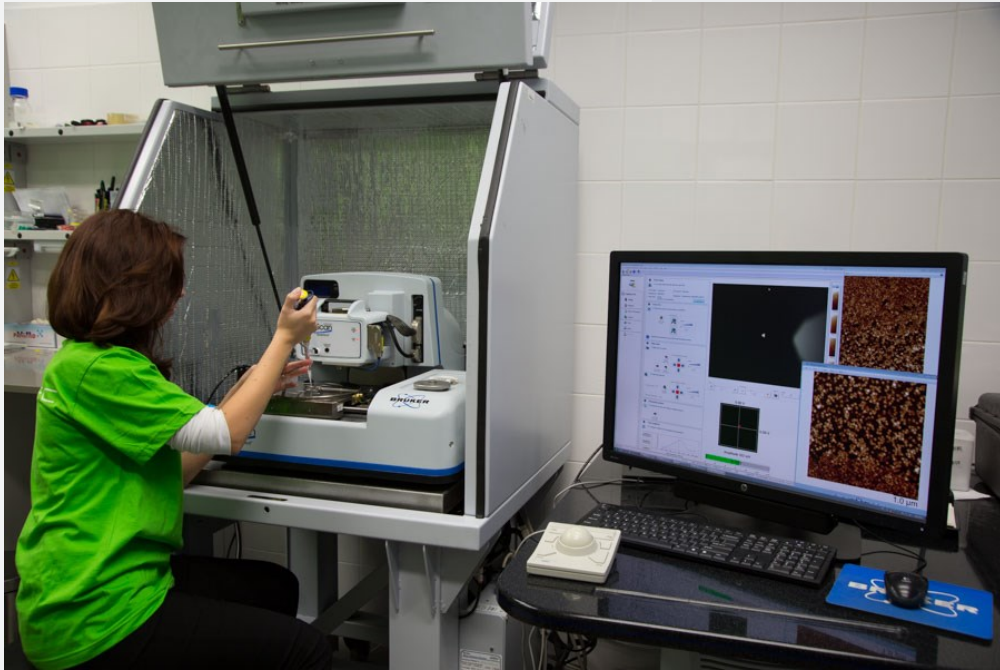
BioAFM – living cells and tissues



+ CytoSurge FluidFM module

+ Biosoft NanoIndenter

BioAFM – molecules, nanoobjects, molecular complexes



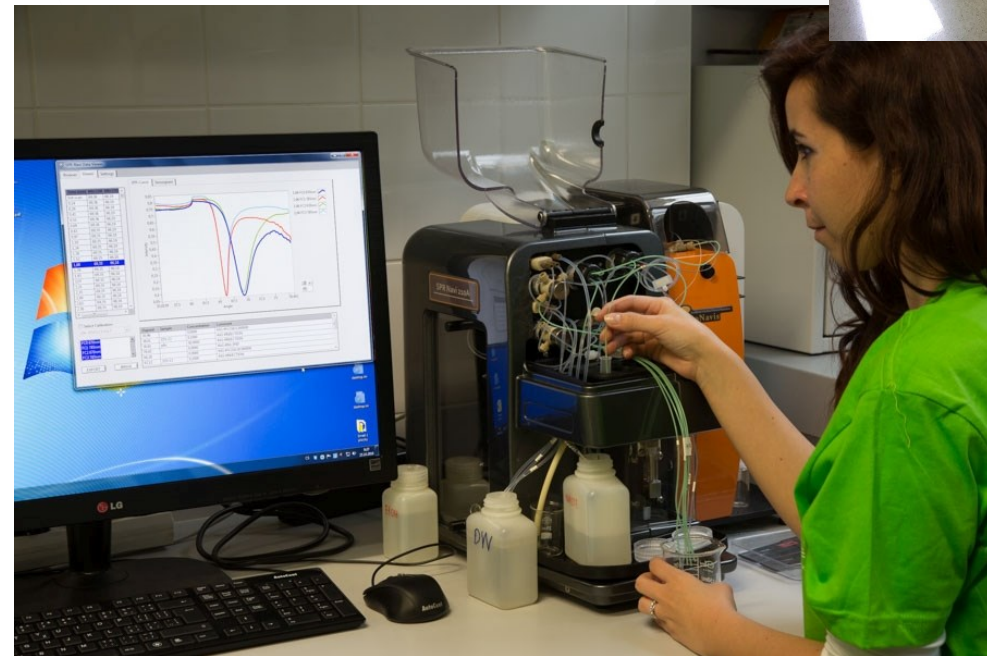
**Bruker Dimension Icon FastScan and MultiMode 8HR
NTMDT Ntgra Vita**

Raman microscopy, SPR affinity biosensor, Upconverting particles UCNP reader

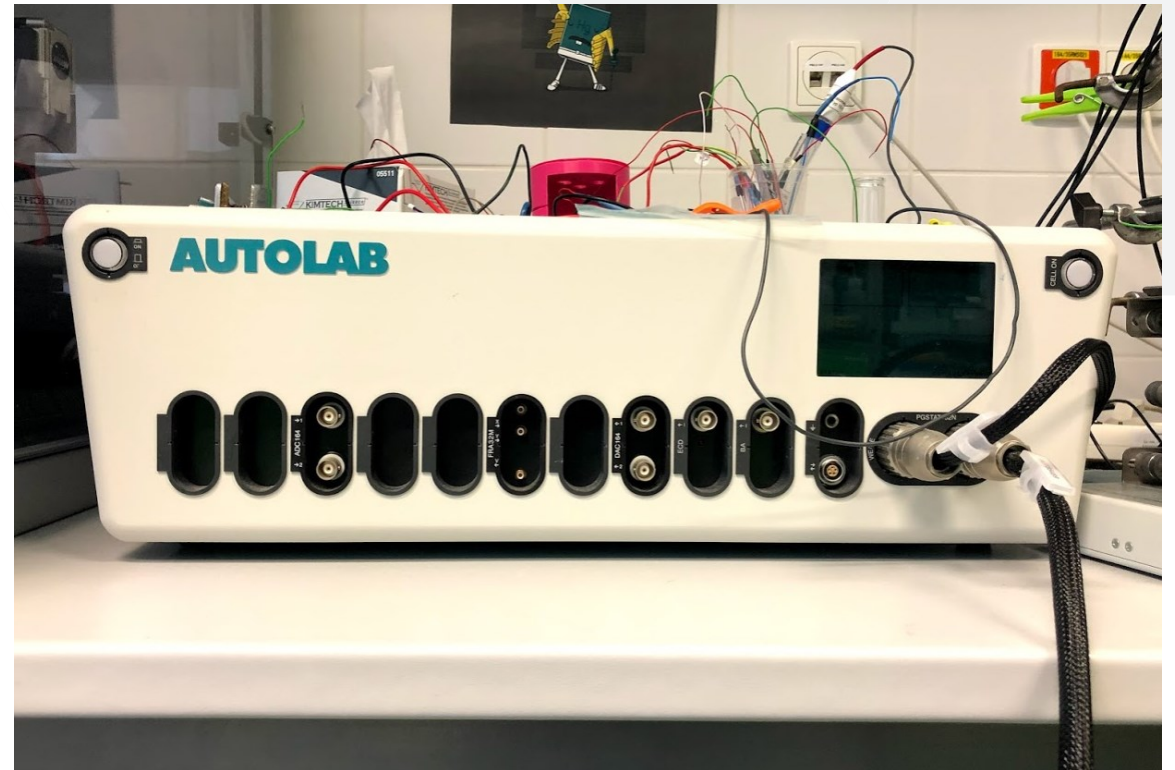
Renishaw InVia Raman microscope

Bionavis SPR biosensor device

Labrox UPCON reader



**Autolab Modular potentiostat
MultiChannel MEA2100Lite**



Technology and Expertise

List of services

1. **Cells – mechanical properties**
2. **Cells - imaging**
3. **Biomolecules - imaging**
4. **Nano-objects imaging**
5. **Raman-AFM combined microscopy**
6. **Raman microscopy**
7. **Electrochemical measurements**
8. **Nanodeposition system**
9. **SPR biosensor**
10. **Scanning of upconversion luminescence**
11. **Multielectrode array recording of cellular potential**

FULL SERVICE / MEASUREMENT only / DATA PROCESSING only

User Training

- 2019 – 2021: **6 workshops**
- **Over 200 participants**
- Workshop content shared **online** – youtube, Data Storage

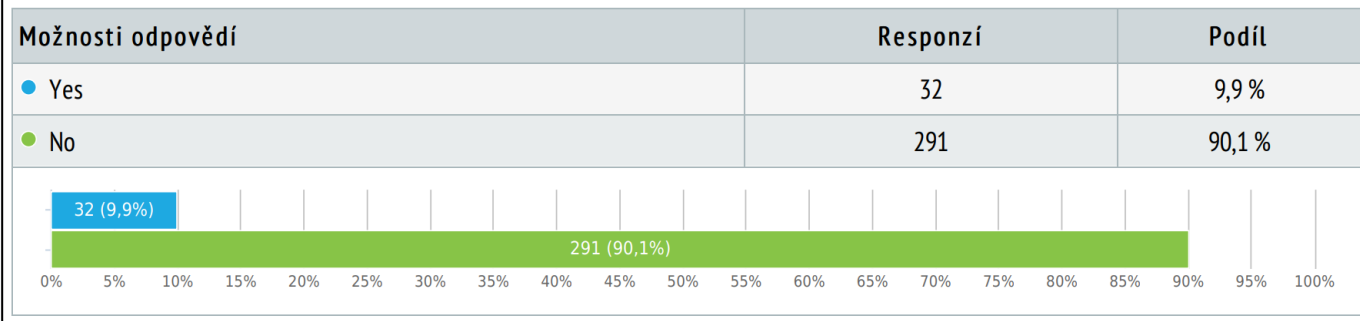
Workshop title	Date	Main objectives	No of participants
Atomic Force Microscopy (AFM) for Bio Applications	April 16-17, 2019	Combined characterization of biosamples by AFM, practical applications, hands-on session	20
Characterization of nanoparticles and proteins by Atomic Force Microscopy	July 30-31, 2019	Characterization of nano-objects and proteins by AFM, practical applications, hands-on session	25
Spring Workshop on BioAFM Microscopy	April 6-8, 2020	Theoretical background and new aspects of bio-AFM microscopy, sample preparation, hands-on session, social event	Canceled
Introduction to Raman microscope Renishaw inVia	June 23 rd , 2020	Introducing the Raman microscope Renishaw inVia, User samples characterization	12
(Bio) Atomic Force Microscopy (bioAFM), Basic Course	October 1 st , 2020	Basics of AFM, Sample preparation techniques, Data processing	80
Introduction to JPK NanoWizard 4 AFM microscope	May 4 th , 2021	Introduction to a new JPK NanoWizard AFM system combined with CytoSurge and NanoIndenter module	10



User Survey

173. Have you used Nanobiotechnology (Nanobio) core facility?

Výběr z možností, zodpovězeno 323x, nezodpovězeno 0x



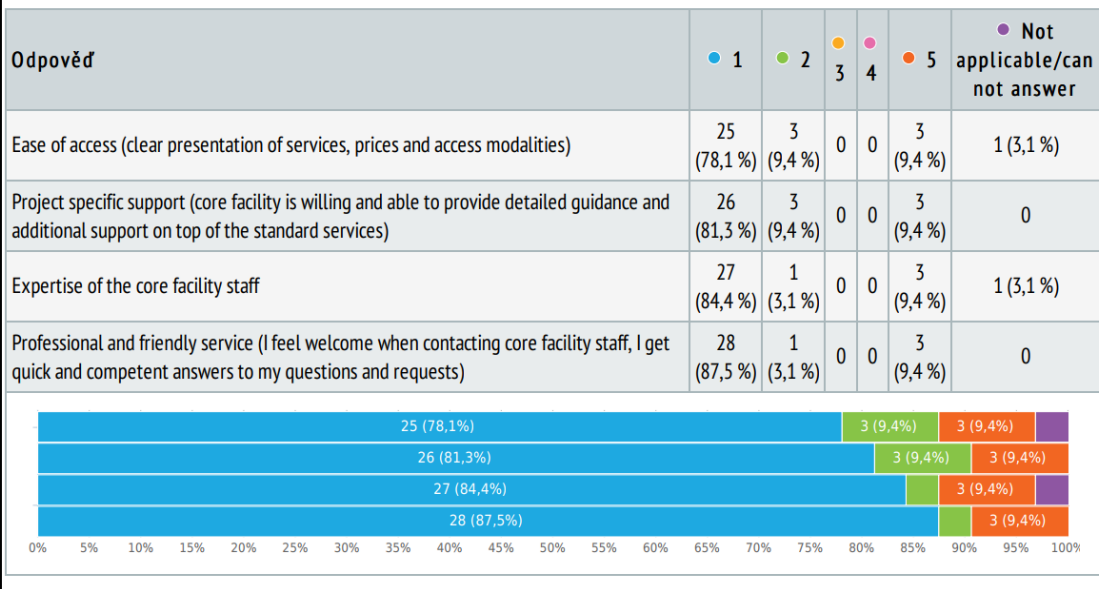
176. Do you have any comments about service quality?

Textová odpověď, zodpovězeno 13x, nezodpovězeno 310x

- CF personel is underestimated
- Everything ok.
- OK
- excellent
- No
- I especially welcome help with adjustment of measurement parameters
- Staff is always willing to help, experts in their fields.
- prompt measurement but until now no results processed
- Dr Pribyl has excellent knowledge not only in AFM area
- Excellent service, facility is willing to help with anything needed. Always available for your questions.
- Jan Pribyl was a great help during our intense measurements on AFM NanoWizard 3.
- (2x) no

175. Service quality: How satisfied are you with service quality at Nanobio?

Matice výběru z možností, zodpovězeno 32x, nezodpovězeno 291x



- N/A
- 20 samples per month
- Possibly some software for volumetric analysis
- pink gloves
- Nanoparticle characterization, stability studies - DLS, zeta-potential. On a regular basis..
- no idea
- Nap room, so that the short stay of AFM measurement would allow more data measured by the visitor, into very late hours and with short periods of sleep
- -
- no

Technology offers for industrial partners

Cooperation with industrial partners (<http://industry.ceitec.cz/>), Daniela Tršová manages this topic.

Bio-AFM microscopy imaging and biomechanical studies

AFM microscopy (structure and mechanical properties) of bio-samples (biomolecules, cells, tissues) under semi-physiological conditions (37 °C, liquid media).

- **Raman microscopy of biosamples**

Raman mapping of biosamples (molecular complexes, cells, tissues) with high resolution (~ 500 nm)

- **Drug testing on cardiac cells**

Biomechanical (bioAFM) and electrical field potential signal as a response to drug exposition. Human stem cells and/or primary animal cells can be used. *! Coordination with Vladimir Rotrekl RG – essential!!!*






- **Tunable hydrogel system**

The new system of stable and Robust biocompatible hydrogel system with tunable mechanical properties.

! Coordination with Vladimir Rotrekl RG – essential!!!

Booking system

Under development for last 2 years..

Plánovací tabule Seznam rezervací Požadavky ▾ Infrastruktura ▾ Původní verze     

Vyberte službu pro požadavek

Other

Biomolecules - imaging
Zobrazování biomolekul (proteiny, DNA, makromolekuly) a jejich komplexů. Standardní podklad – slída (mica), lze použít i jiné – HOPG, křemík, kovové elektrody, atp. Metody: pokleповý režim, PF-QNM, QI, Force Volume. Vyhodnocení a export dat.

Cells - imaging
Buněčné kultury ve standardní Petriho misce (TPP 93040), lze použít i misky pro konfokální mikroskopii (vhodný typ nejprve konzultujte s námi). Fixované (např. PFA) buňky na skle. Metody – kontaktní mód, QI, PF-QNM, Force Volume. Post-processing a export dat. Možná kombinace s optickou mikroskopii (BF, fluorescence, konfokální mikroskopie) – možnost nezávislého nebo overlay snímkování. Místnost je vybavena CO2 inkubátorem a malým laminárním boxem. UV sterilizace prostoru.

Cells - mechanical properties
Buněčné kultury ve standardní Petriho misce (TPP 93040), lze použít i misky pro konfokální mikroskopii (vhodný typ nejprve konzultujte s námi). Metoda Force-Mapping, biomechanická charakterizace kardiomyocytů. Vyhodnocení naměřených dat matematickými modely (Hertz-Sneddon, DMT, JKR, atd.), post-processing. Možná kombinace s optickou mikroskopii (BF, fluorescence, konfokální mikroskopie) – možnost nezávislého nebo overlay snímkování. Místnost je vybavena CO2 inkubátorem a malým laminárním boxem. UV sterilizace prostoru.

Electrochemical measurements
Elektrochemický analyzátor pro voltametrická, amperometrická a impedanční měření (EIS) na různých typech elektrod a sensorů. Možnost dvoukanálových měření, vysoká citlivost, nízký šum. SW Autolab Nova pro analýzu dat.

SPR biosensor
Dvoukanálový průtočný SPR (bio)sensor využívající metody rezonance povrchového plasmonu. Sledování a charakterizace optických vlastností tenkých vrstev a jejich změn v reálném čase – v kapalině i nasucho. Velmi široký úhlový rozsah díky použití goniometru. Využití 2 vlnových délek umožňuje měření indexu lomu a tloušťky vrstev. Dále lze simultánně provádět elektrochemická měření. Možnost sledování a charakterizace interakcí biomolekul bez potřeby jejich značení, jeden vazebný partner musí být imobilizován na povrchu měřicího čipu, druhý je volný v roztoku. Určování kinetických parametrů, vazebných konstant či měření koncentrace různých analytů.

Nano-objects imaging
Zobrazování nano-objektů (nanočástice, nanotrubičky, nanodrátky, atp.) a jejich komplexů Standardní podklad – slída (mica), lze použít i jiné – HOPG, křemík, kovové elektrody, atp. Metody: pokleповý režim, PF-QNM, QI, Force Volume. Vyhodnocení a export dat.

Data Sharing

- Medium Storage of MU – complicated for external users
- OneDrive – limited space to 1 TB
- IT manager missing

711025-Core Facility Nanobiotechnology	ADR	10.03.2021	17:34:28
711025-Core Facility Nanobiotechnology-BIOLOGY	ADR	04.02.2020	14:32:55
711025-Core Facility Nanobiotechnology-CF_Internal	ADR	16.04.2021	14:08:04
711025-Core Facility Nanobiotechnology-Guides	ADR	04.04.2021	21:07:03
711025-Core Facility Nanobiotechnology-Workshops	ADR	18.12.2020	13:10:26
<hr/>			
_MACHINES_backups	ADR	04.12.2020	10:06:32
_Guides	ADR	04.04.2021	21:07:03
_Software	ADR	17.03.2021	13:03:19
_Workshops	ADR	18.12.2020	13:10:26
A-beta	ADR	19.02.2021	18:01:57
Andrej Besse	ADR	23.03.2020	12:45:00
BIOLOGY	ADR	04.02.2020	14:32:55
BOUCHAL	ADR	21.12.2019	17:34:19

Sharing of

- *Data*
- *Workshop content*
- *Software*
- *Guides*

CF involvement in the Correlative Microscopy project

Integration of our CF in the project **correlative microscopy** has been proposed in the **Strategic Plan** of the Central European Institute of Technology at Masaryk University for **2021-2028**, section "At the frontiers of technology: Correlative approaches to connect dynamics and structure of living systems."

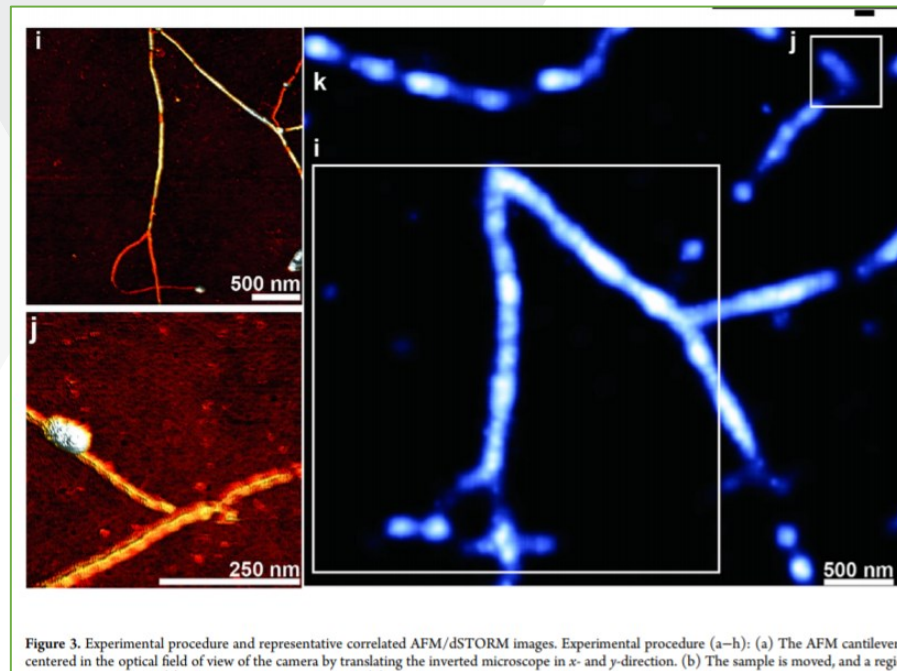
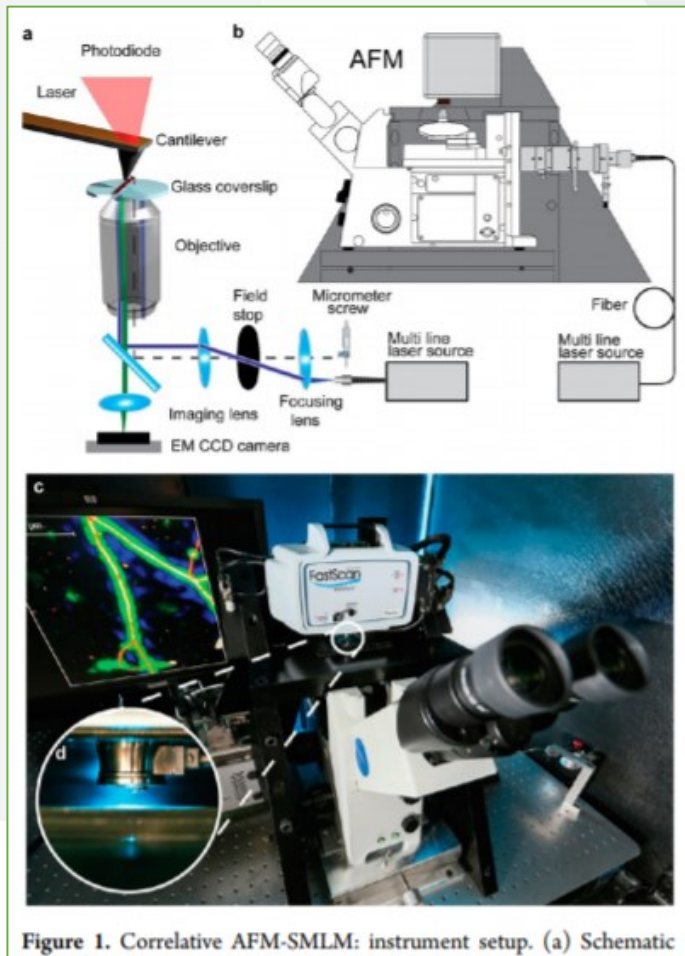
Correlative AFM-STED microscopy highlight a new and essential aspect and generate a warning: Fluorescence techniques cannot characterize all the products derived from the in vitro aggregation of misfolded proteins. Therefore, the combination of microscopic techniques brings a better understanding of the physiological processes.

Mechanical properties (AFM), calcium levels (fluorescent dyes), and cellular potential (MEA) – **mechano-physical-electrical** coupling

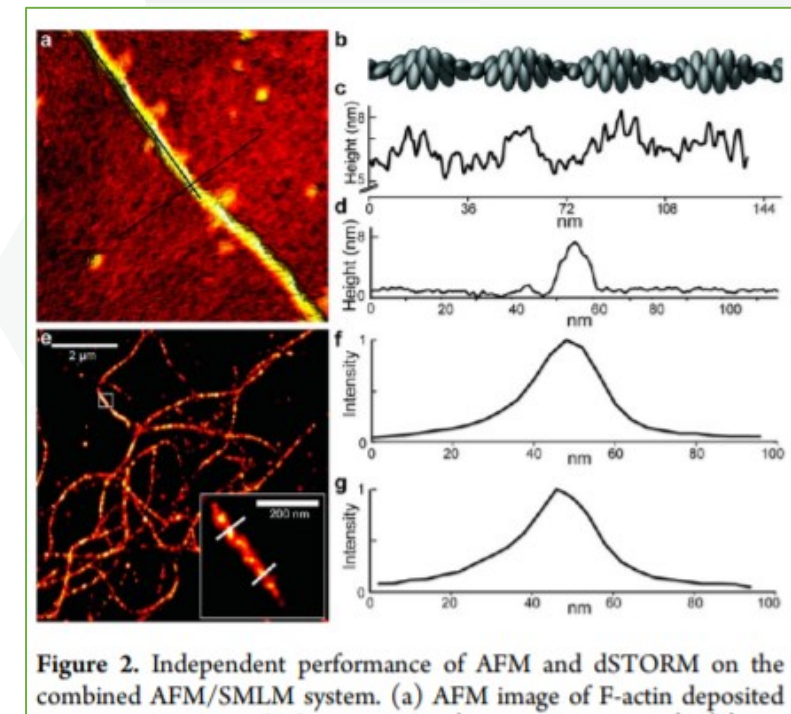
"Mikroskopie" (= Microscopy) educational project and in the LLL remote training and propagation project started in the time of the corona crisis.

High-Resolution Correlative Microscopy: Bridging the Gap between Single Molecule Localization Microscopy and Atomic Force Microscopy

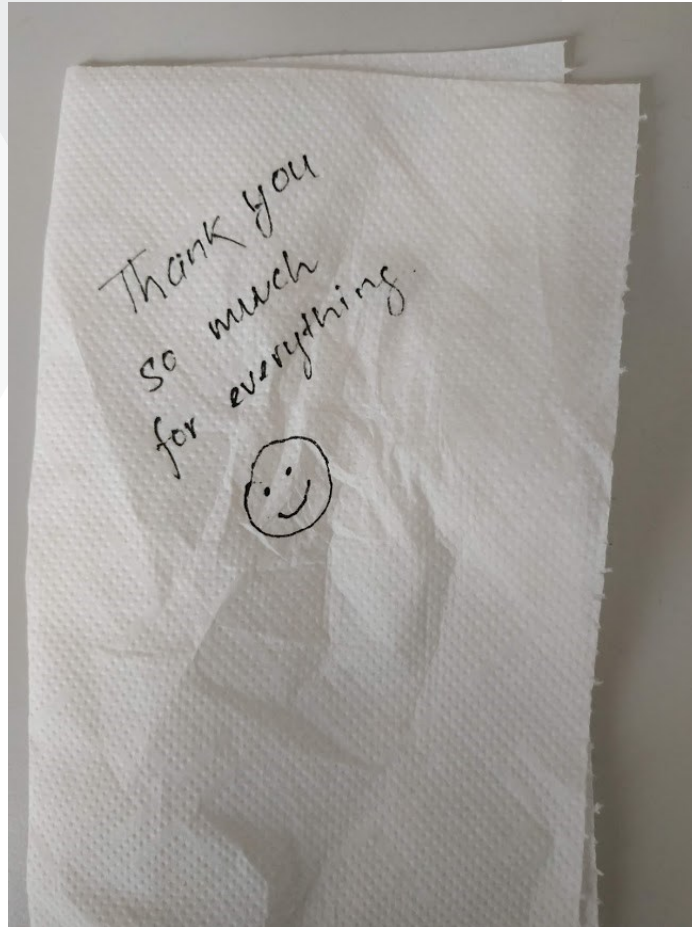
P.D. Odermatt et al., *Nano Lett.* 2015, 15, 8, 4896–4904



F-Actin fibers study



Let's all the measurements end up with this...



Conclusions

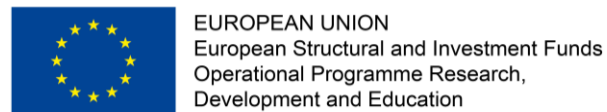
- **BioAFM microscopy - more than just imaging method**
- **Easy combination with other methods**
- **Sample range from molecules to tissue slices**
- **Mechanobiology - information related to the pathophysiology**

Acknowledgment

- Petr Skládal – FS MU
- J. Víteček, L. Kubala – IBP Brno
- M. Samwer, IMBA Wien
- G. Forte, G. Nardone – ICRC Brno
- M. Pesl, V. Rotrekl, S. Jelinkova, ... - MU Brno
- I. Kratochvilova – IF CAS, Prague
- E. Benkova, M. Gallemi – IST Wien

Acknowledgment text - CIISB

- **Preferred version:** *„CIISB, Instruct-CZ Centre of Instruct-ERIC EU consortium, funded by MEYS CR infrastructure project LM2018127, is gratefully acknowledged for the financial support of the measurements at the CF Nanobiotechnology.“*
- **Short version:** *„We acknowledge CF Nanobiotechnology of CIISB, Instruct-CZ Centre, supported by MEYS CR (LM2018127).“*



OP VVV CZ.02.1.01/0.0/0.0/18_046/0015974

Thank you for your attention!