

# Před analýzou

>P12345 Yeast chromosome1

GATTACAGATTACAGATTACAGATTACAGATTACAG  
ATTACAGATTACAGATTACAGATTACAGATTACAGA  
TTACAGATTACAGATTACAGATTACAGATTACAGAT  
TACAGATTAGAGATTACAGATTACAGATTACAGATT  
ACAGATTACAGATTACAGATTACAGATTACAGATTA  
CAGATTACAGATTACAGATTACAGATTACAGATTAC  
AGATTACAGATTACAGATTACAGATTACAGATTACA  
GATTACAGATTACAGATTACAGATTACAGATTACAG  
ATTACAGATTACAGATTACAGATTACAGATTACAGA  
TTACAGATTACAGATTACAGATTACAGATTACAGAT

# Po částečné analýze

>P12345 Gene\_1 - gen kodující  
protein alkoholdehydrogenazy ...

TATA	TATAAA
	CGATTGACGATGACGAT
start	ATG
exon1	TACAGATTACAGATTACAGATTACAGATGT
intron1	CAGATTACAGATTACAGATTACAGATTACAGATTCA
exon2	AGATTACAGATTACAGATTACAGA
stop	TAA

>P12346 Protein\_1  
MASAQSFYLLDHNQNNQNFDDHLAVDIVMILSHERFMN

# Analýza DNA sekvence

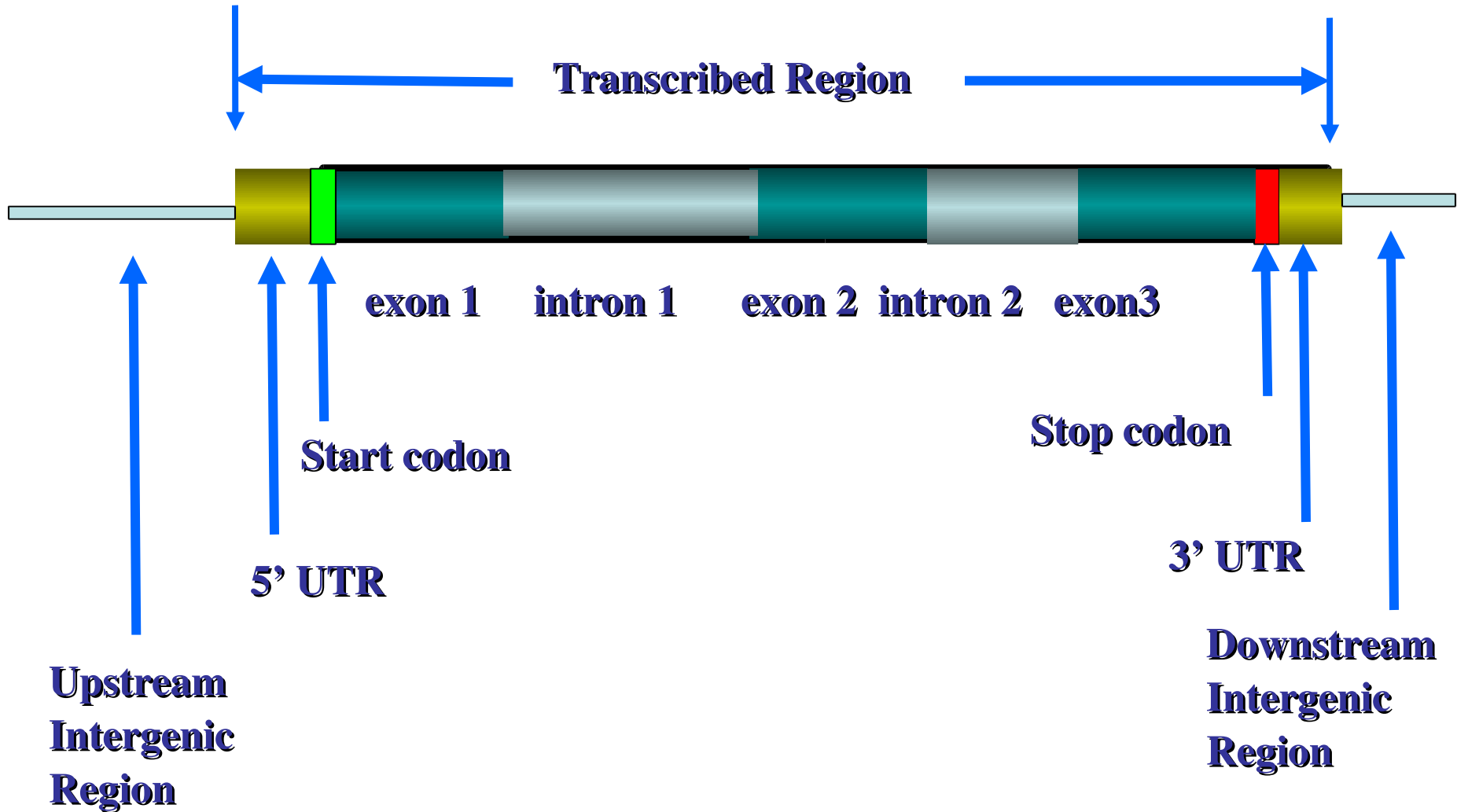
☀️ ≈ anotace genomu (sekvence)

- ☀️ identifikace signálů a genů
- ☀️ anotace genů (jejich kódujících sekvencí)

# Anotace genů $\approx$ anotace proteinů

- ☀ Identifikace a popis fyzikálně-chemických, funkčních a strukturních vlastností daného genu/proteinu
  - ☀ sekvence DNA, AA, pozice v genomu, délka, složení
  - ☀ běžné názvy, odkazy na literaturu
  - ☀ příslušnost do rodiny, evoluce
  - ☀ partneři pro interakci, aktivita, regulační mechanismy
  - ☀ struktura, aktivní místa, role v metabolismu buňky

# Eukaryotic Gene Structure



# Analýza DNA sekvence

- ☀ Statistika

- ☀ frekvence n-gramů a jiných prvků, repetice, kodony

- ☀ Signální prvky

- ☀ TATA (promotor), ATG (start), STOP, GT (donor), AG (akceptor) a pod

- ☀ Kódující část

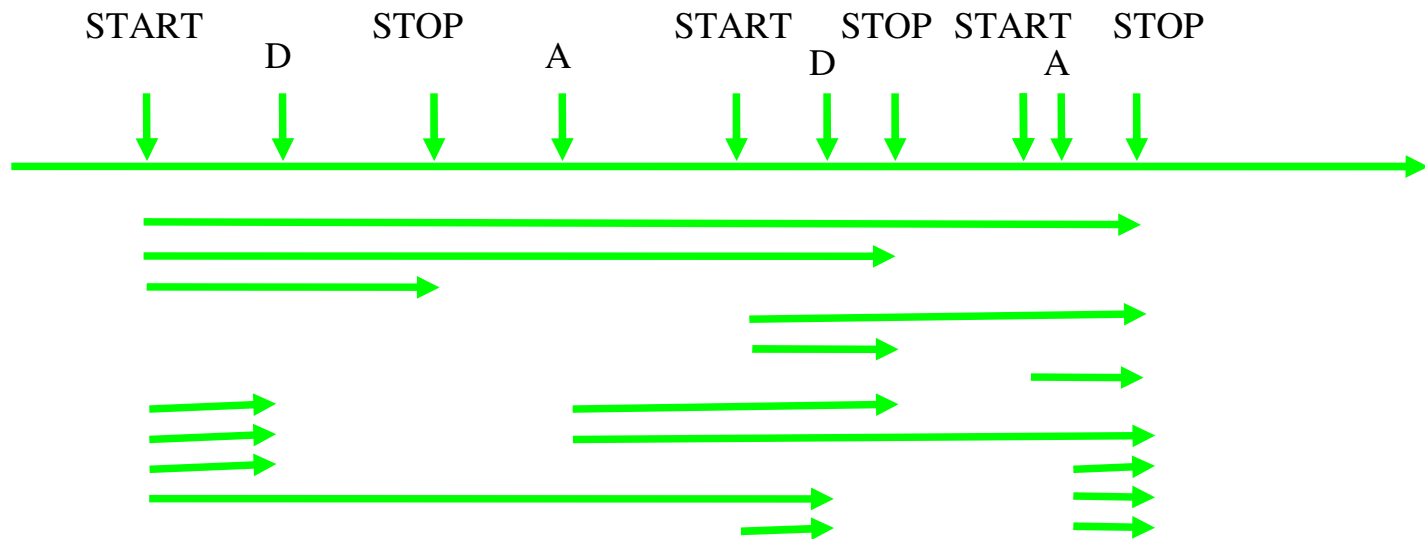
- ☀ podobnost kódované sekvence s jinými proteiny

- ☀ Kombinované přístupy

# Identifikace genů

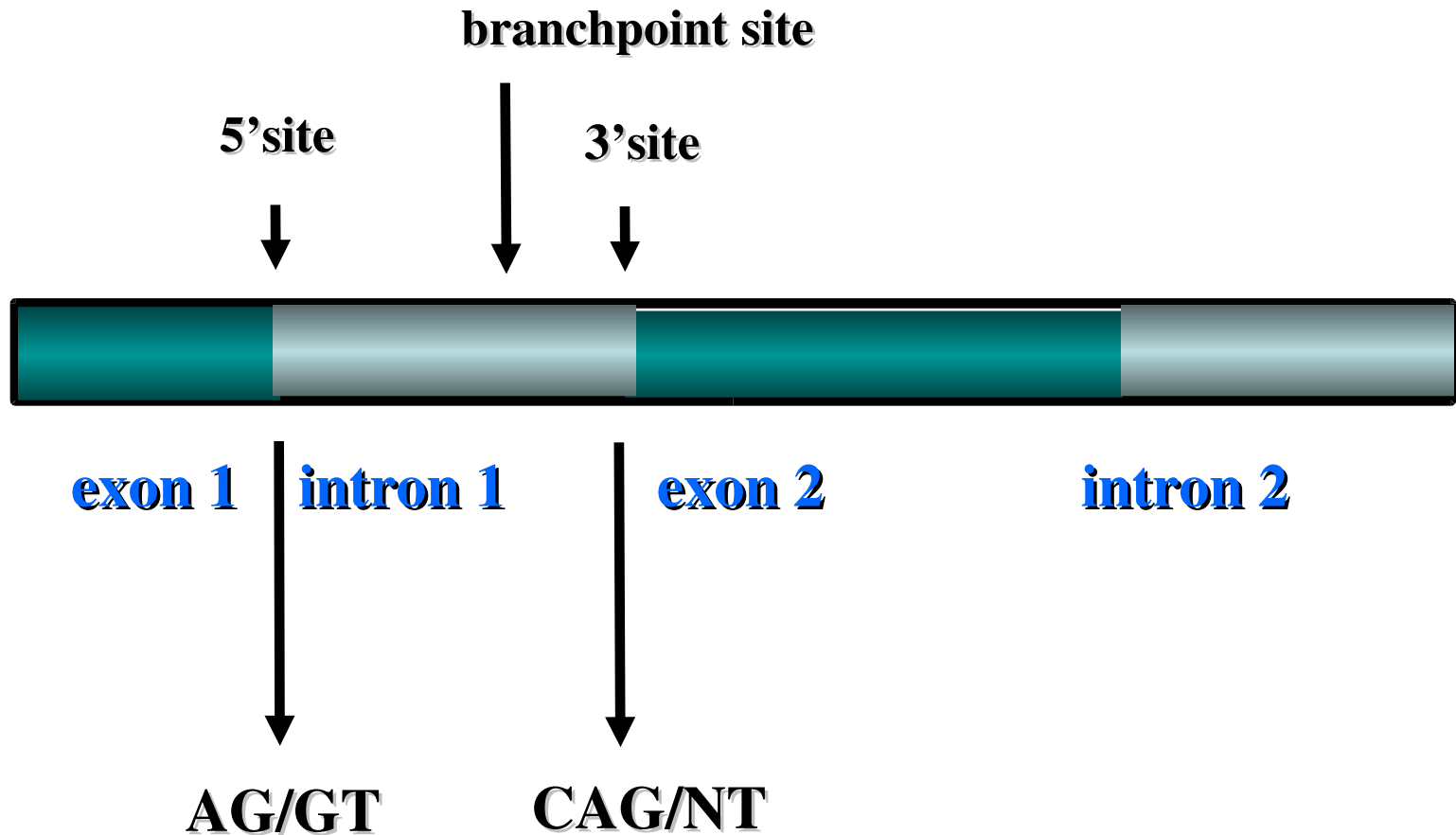
- ☀ U prokaryotů 95-100% spolehlivost, u složitějších eukaryotů 90% na úrovni bazí, 70% na úrovni exonů/intronů
  - ✳ existence intronů
  - ✳ větší genomy
  - ✳ nízká hustota genů (<30%; 3% u Homo sapiens)
  - ✳ alternativní splicing (zhruba u poloviny genů)
  - ✳ velké množství repetitivních sekvencí
  - ✳ občasný překryv genů

# Identifikace genů

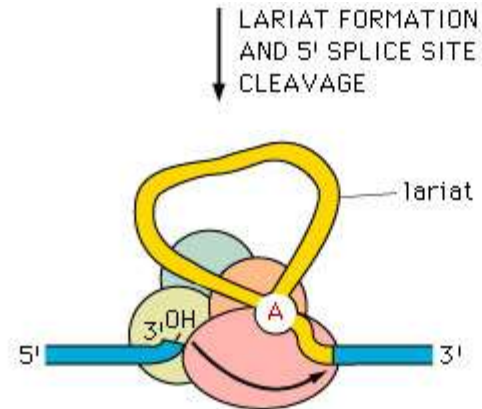
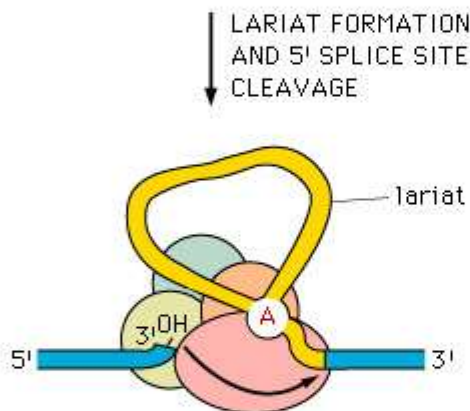
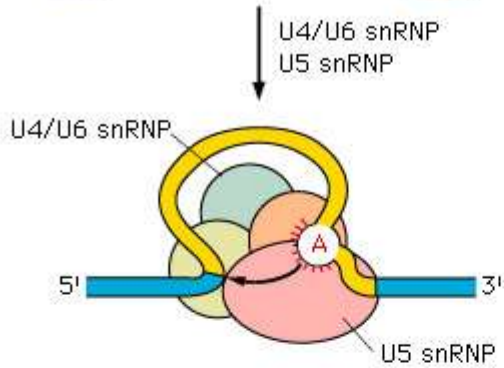
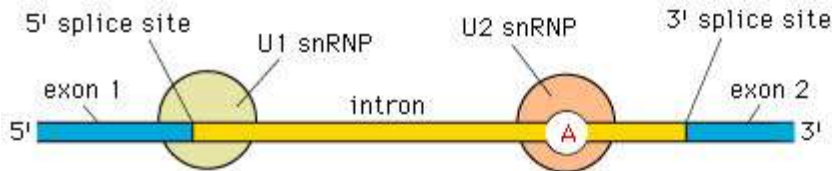




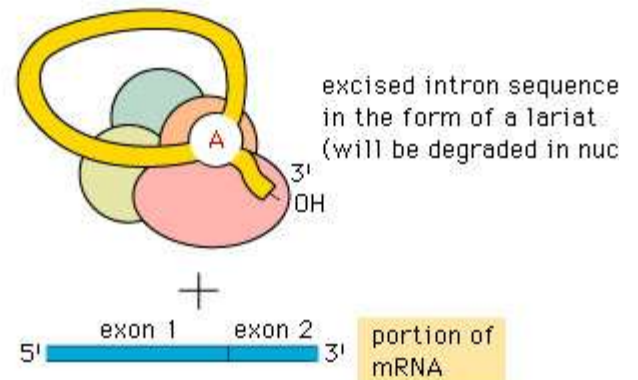
# Eukaryotic Gene Structure



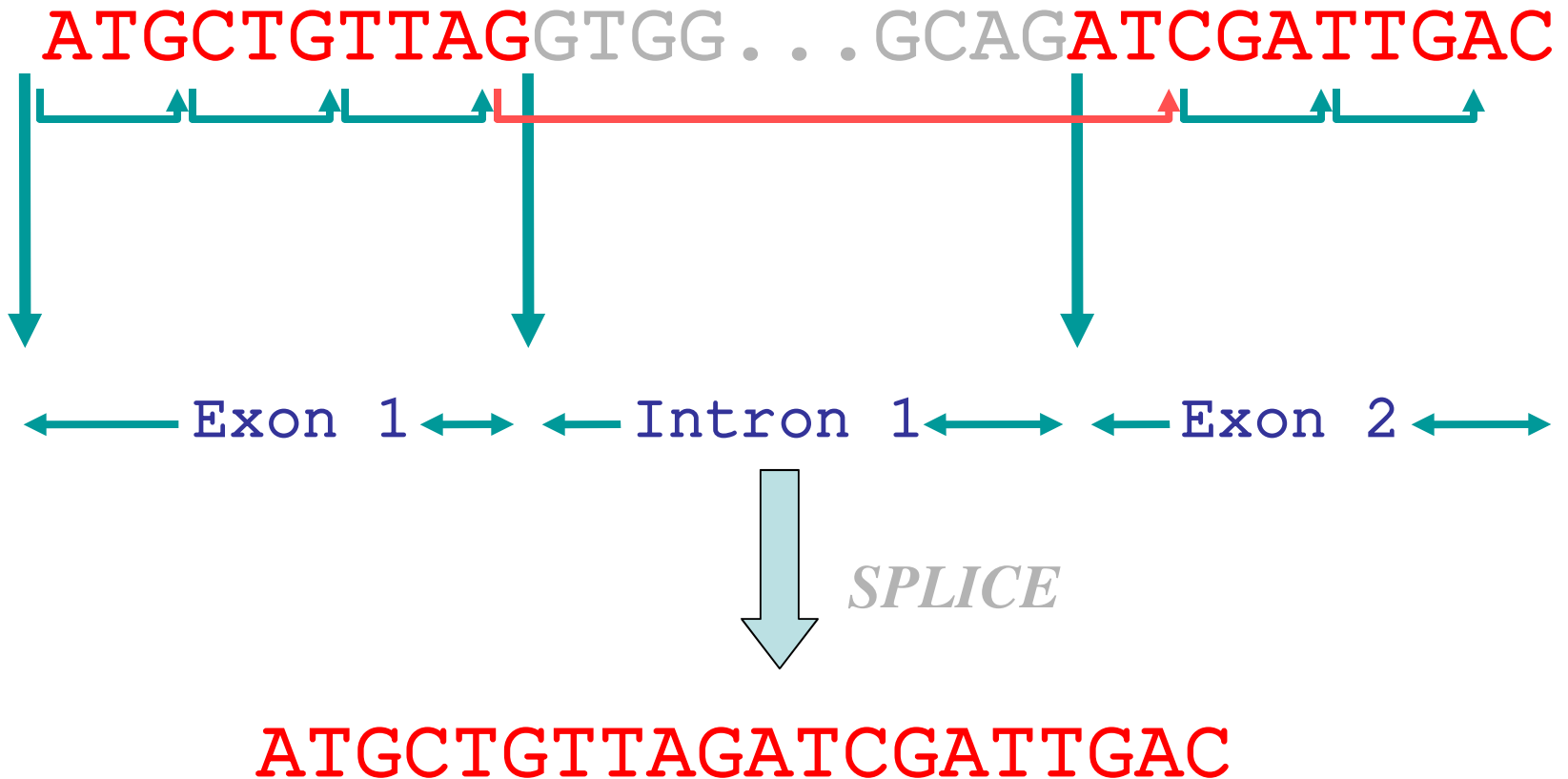
# RNA Splicing



3' SPLICE SITE CLEAVAGE AND JOINING OF TWO EXON SEQUENCES



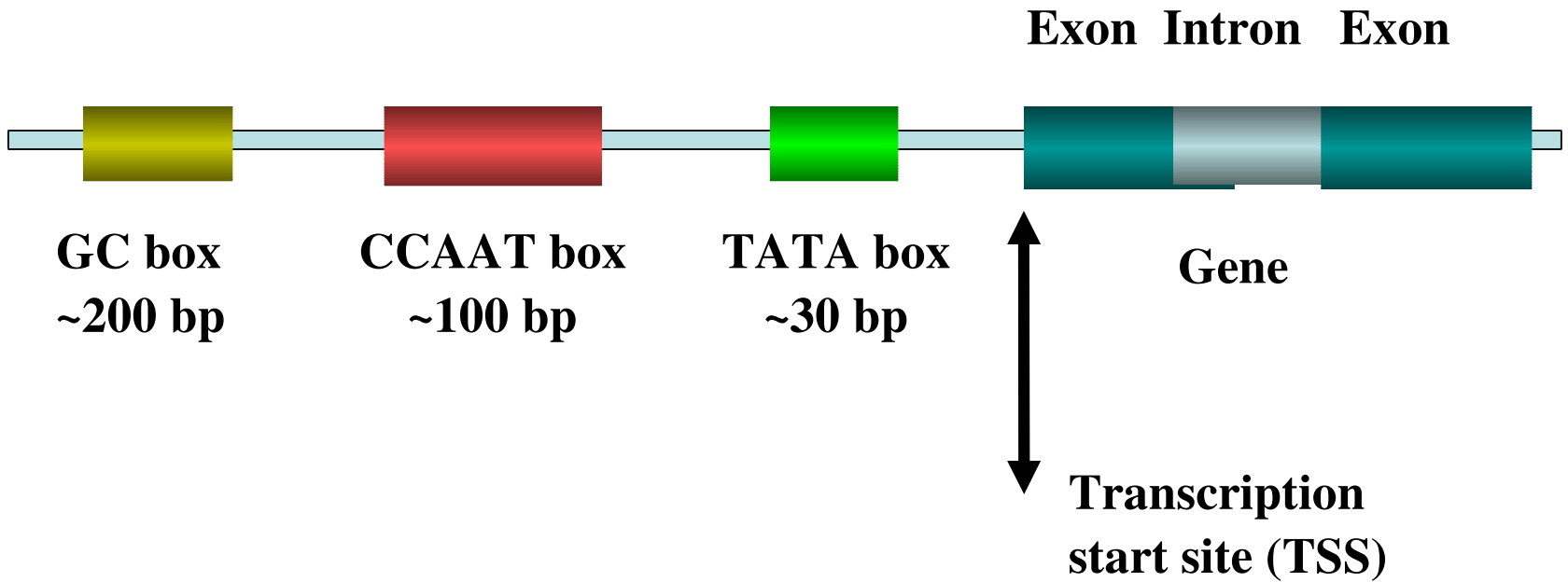
# Exon/Intron Structure (Detail)



# Typické signály v eukaryotických sekvencích

- ✦ Promotorové elementy
  - ✦ CAP, CCAAT, GC a TATA
- ✦ Kozakova sekvence (rozpoznávána ribozomem = RBS)
- ✦ Splicing (donor, acceptor a lariat)
- ✦ Terminační signál
- ✦ Polyadenylační signál

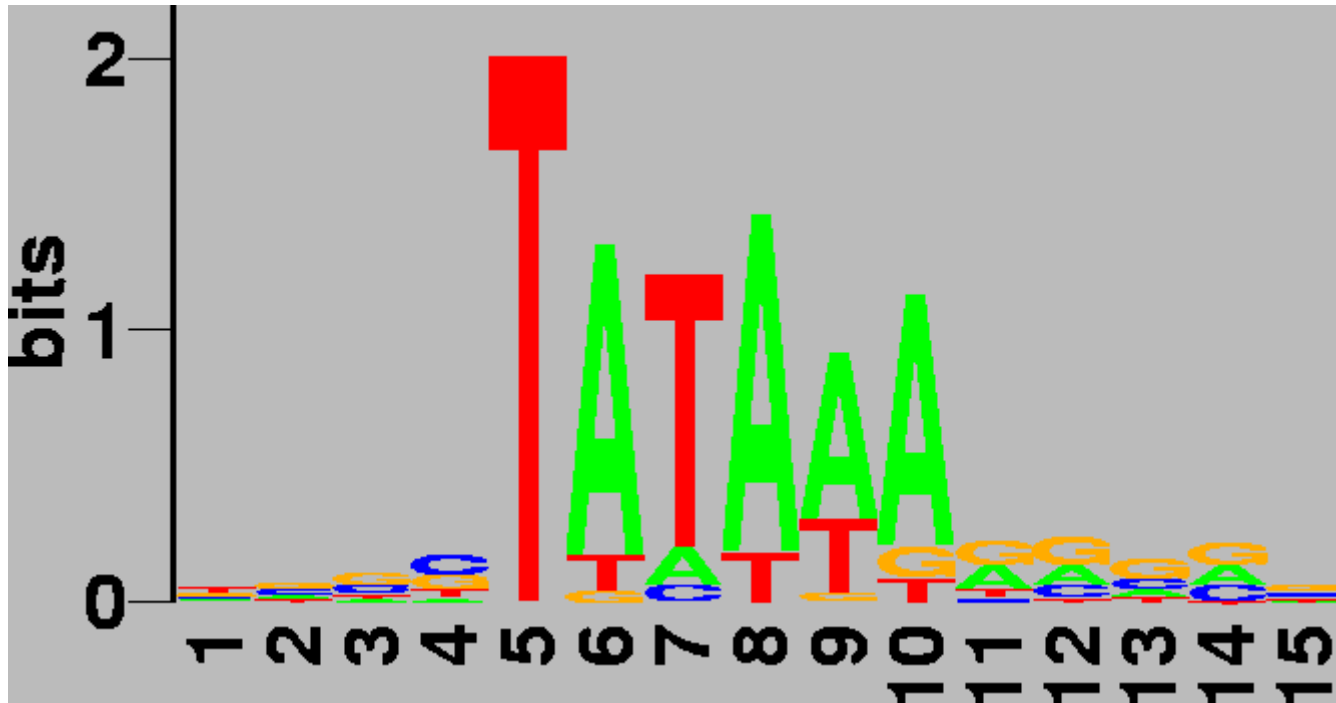
# Pol II Promoter Elements



# Pol II Promoter Elements

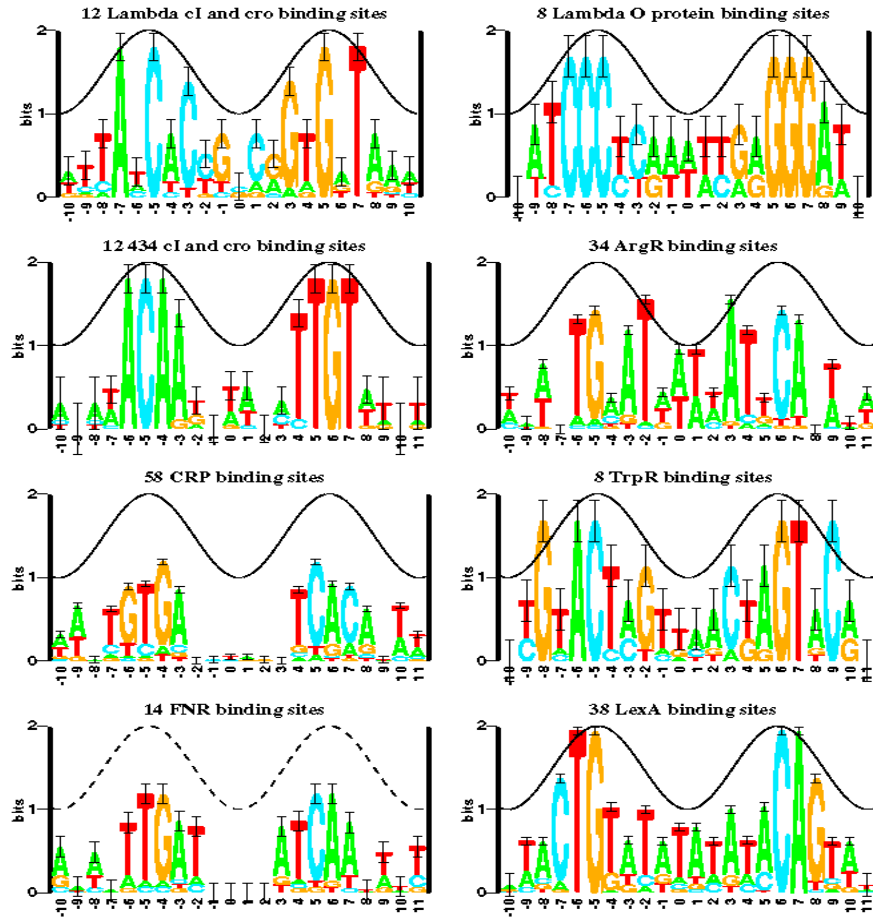
- **Cap Region/Signal**
  - **n C A G T n G**
- **TATA box (~ 25 bp upstream)**
  - **T A T A A n G C C C**
- **CCAAT box (~100 bp upstream)**
  - **T A G C C A A T G**
- **GC box (~200 bp upstream)**
  - **A T A G G C G nGA**

# Pol II Promoter Elements



**TATA box is found in ~70% of promoters**

# WebLogos



<http://www.bio.cam.ac.uk/cgi-bin/seqlogo/logo.cgi>

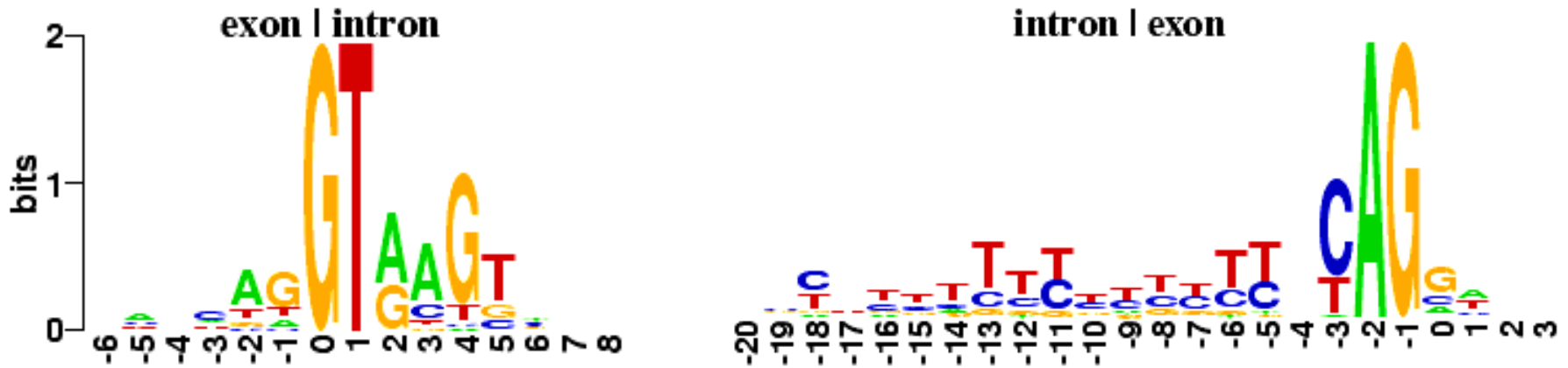
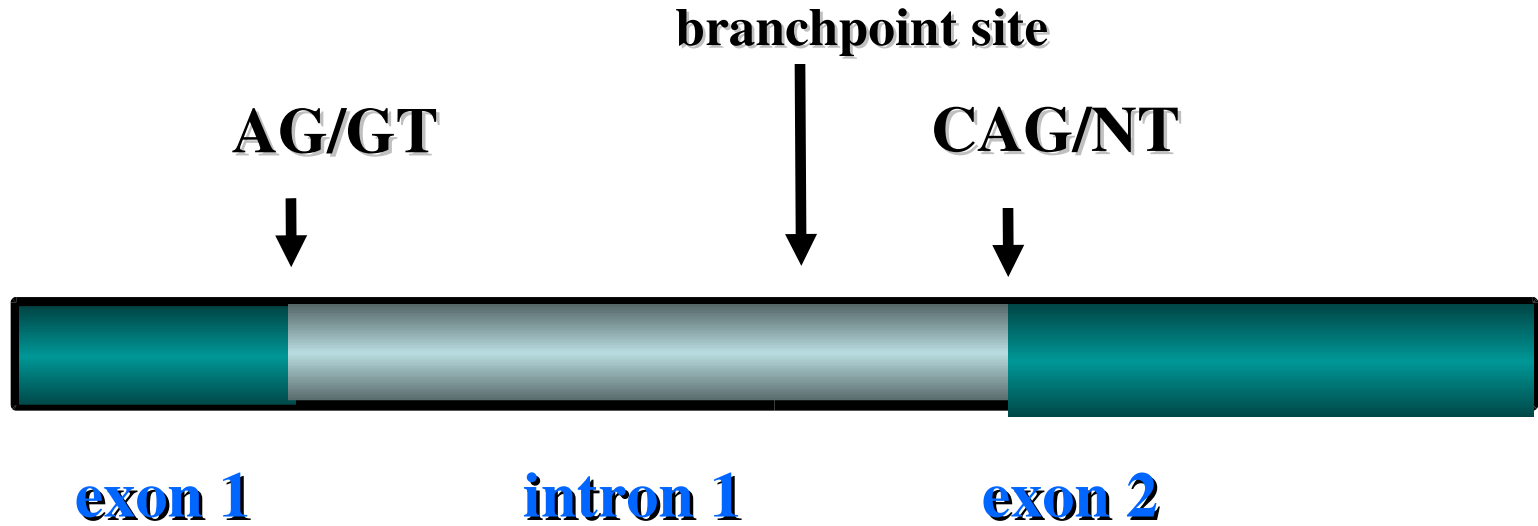


# Kozak (RBS) Sequence

-7 -6 -5 -4 -3 -2 -1 0 1 2 3  
A G C C A C C **A** **T** **G** G



# Splice Signals

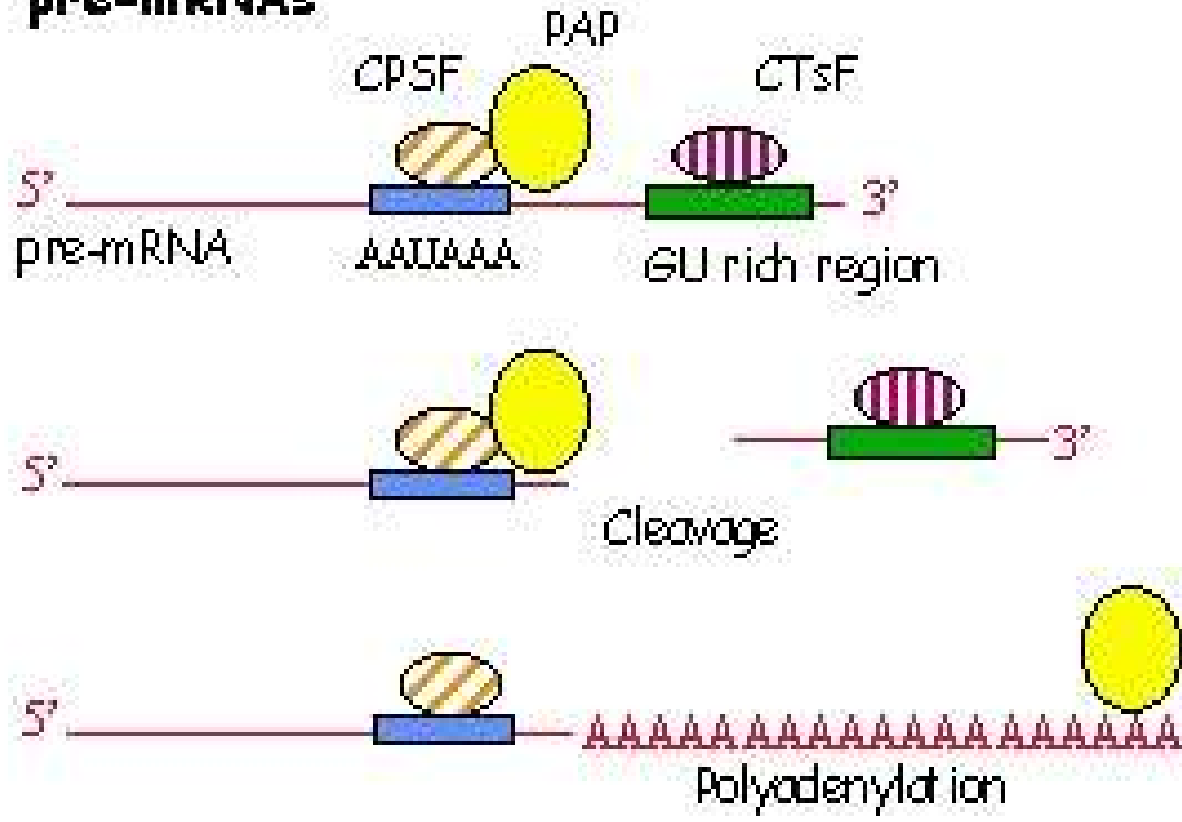


# Miscellaneous Signals

- **Polyadenylation signal**
  - **A A T A A A or A T T A A A**
  - Located 20 bp upstream of poly-A cleavage site
- **Termination Signal**
  - **A G T G T T C A**
  - Located ~30 bp downstream of poly-A cleavage site

# Polyadenylation

## Cleavage and Polyadenylation of Eukaryotic pre-mRNAs



CPSF – Cleavage & Polyadenylation Specificity Factor

PAP – Poly-A Polymerase

CTsF – Cleavage Stimulation Factor

# Analýza genomu – kombinované metody

- ✦ Neurónové sítě
  - ✦ Grail, GeneParser
- ✦ Lineární diskriminační analýza
  - ✦ GeneFinder, GeneID, MZEF
- ✦ Lingvistická
  - ✦ GeneLang
- ✦ Markovovy řetězce
  - ✦ Genie, GeneMark, GenScan, VEIL
- ✦ Podobnosti
  - ✦ Procrustes, AAT
- ✦ Rozhodovací stromy

# Neural Network

## Training Set

ACGAAG  
AGGAAG  
AGCAAG  
ACGAAA  
AGCAAC



## Definitions

A = [001]  
C = [010]  
G = [100]  
  
E = [01]  
N = [00]

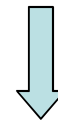


## Desired Output

EEEENN

## Sliding Window

ACGAAG



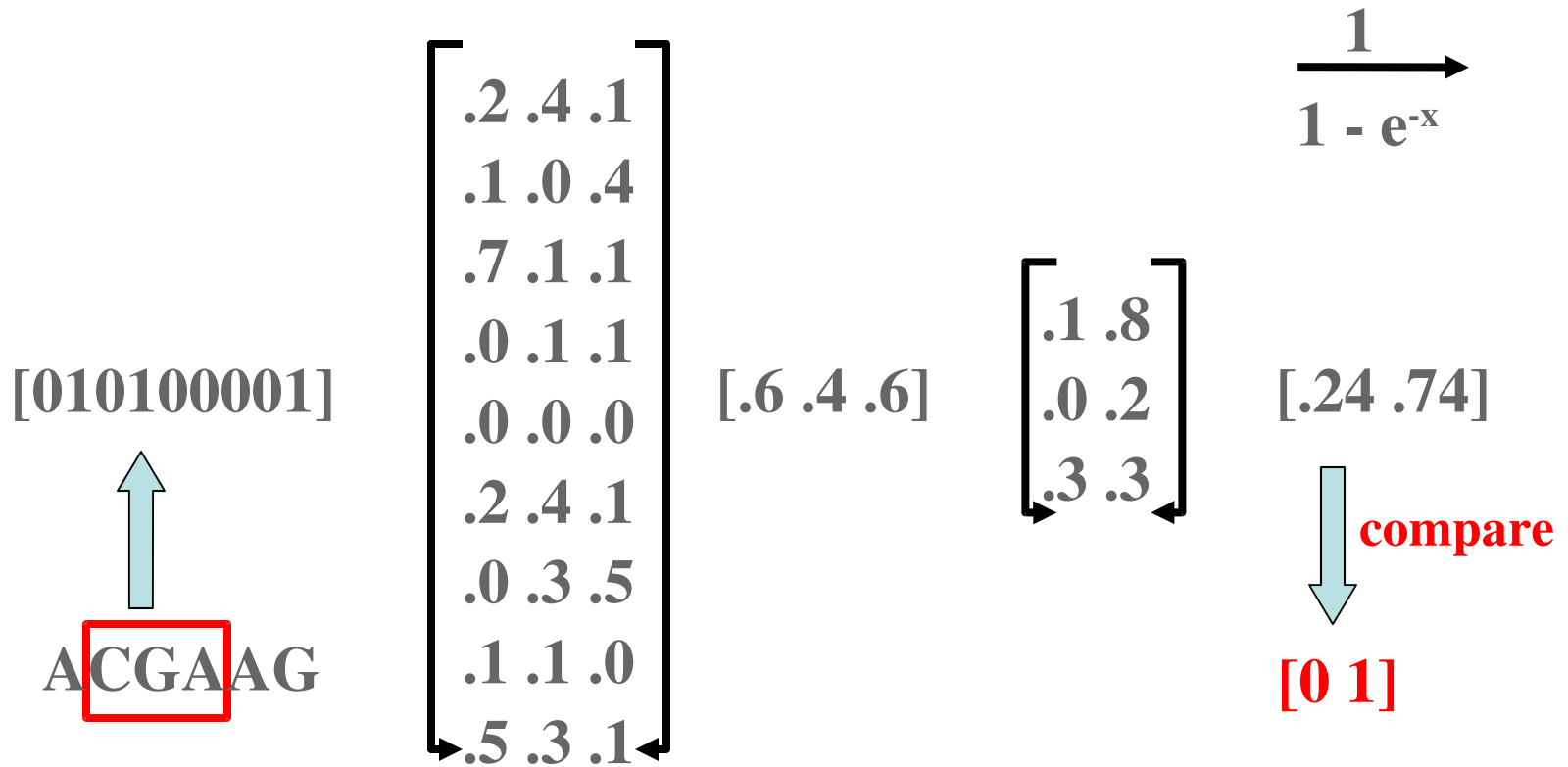
[010100001]

## Input Vector

[01]

## Output Vector

# Neural Network Training



**Input  
Vector**

**Weight  
Matrix1**

**Hidden  
Layer**

**Weight  
Matrix2**

**Output  
Vector**

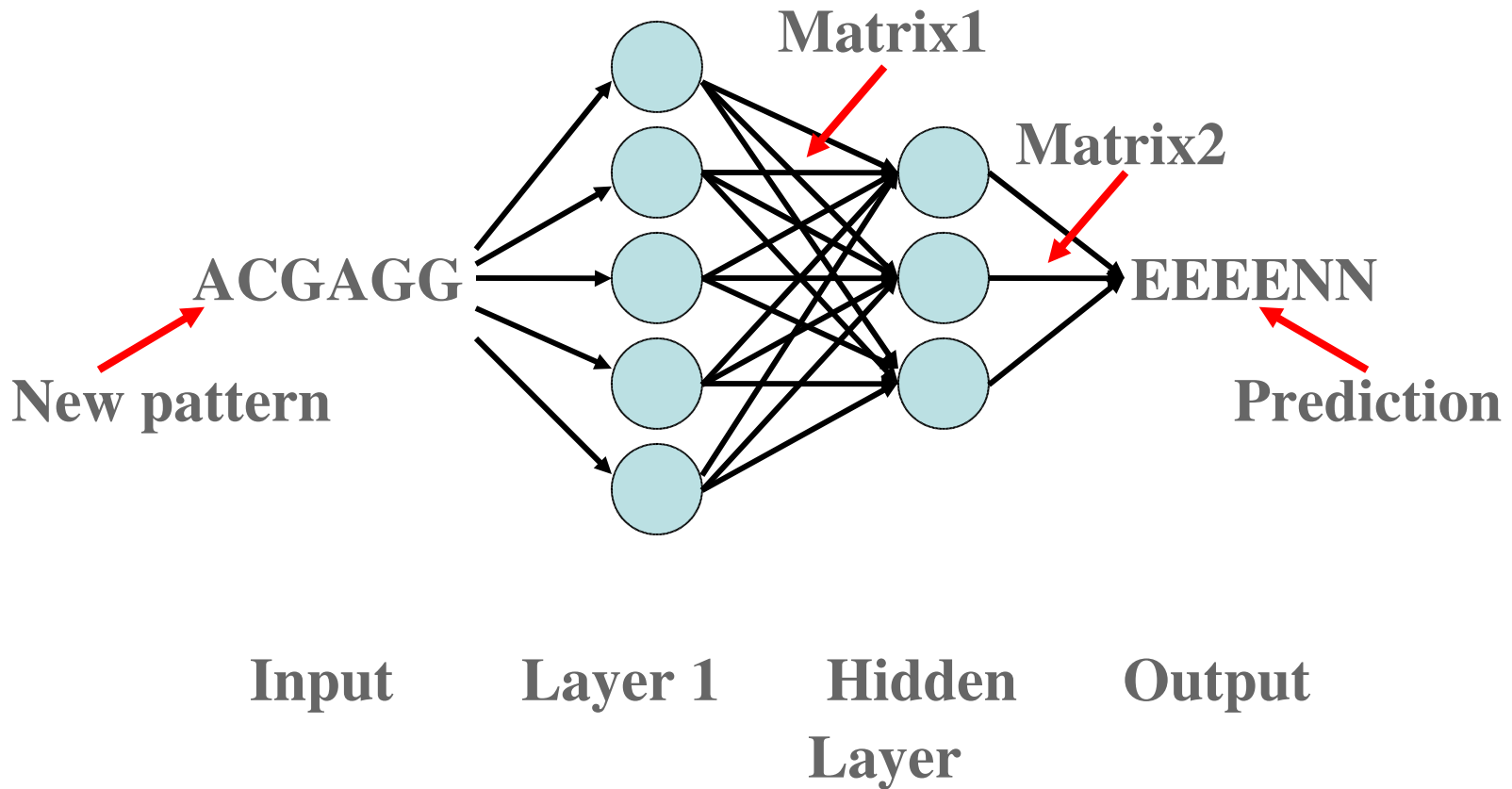
# After Many Iterations....

$$\begin{bmatrix} .13 & .08 & .12 \\ .24 & .01 & .45 \\ .76 & .01 & .31 \\ .06 & .32 & .14 \\ .03 & .11 & .23 \\ .21 & .21 & .51 \\ .10 & .33 & .85 \\ .12 & .34 & .09 \\ .51 & .31 & .33 \end{bmatrix} \quad \begin{bmatrix} .03 & .93 \\ .01 & .24 \\ .12 & .23 \end{bmatrix}$$

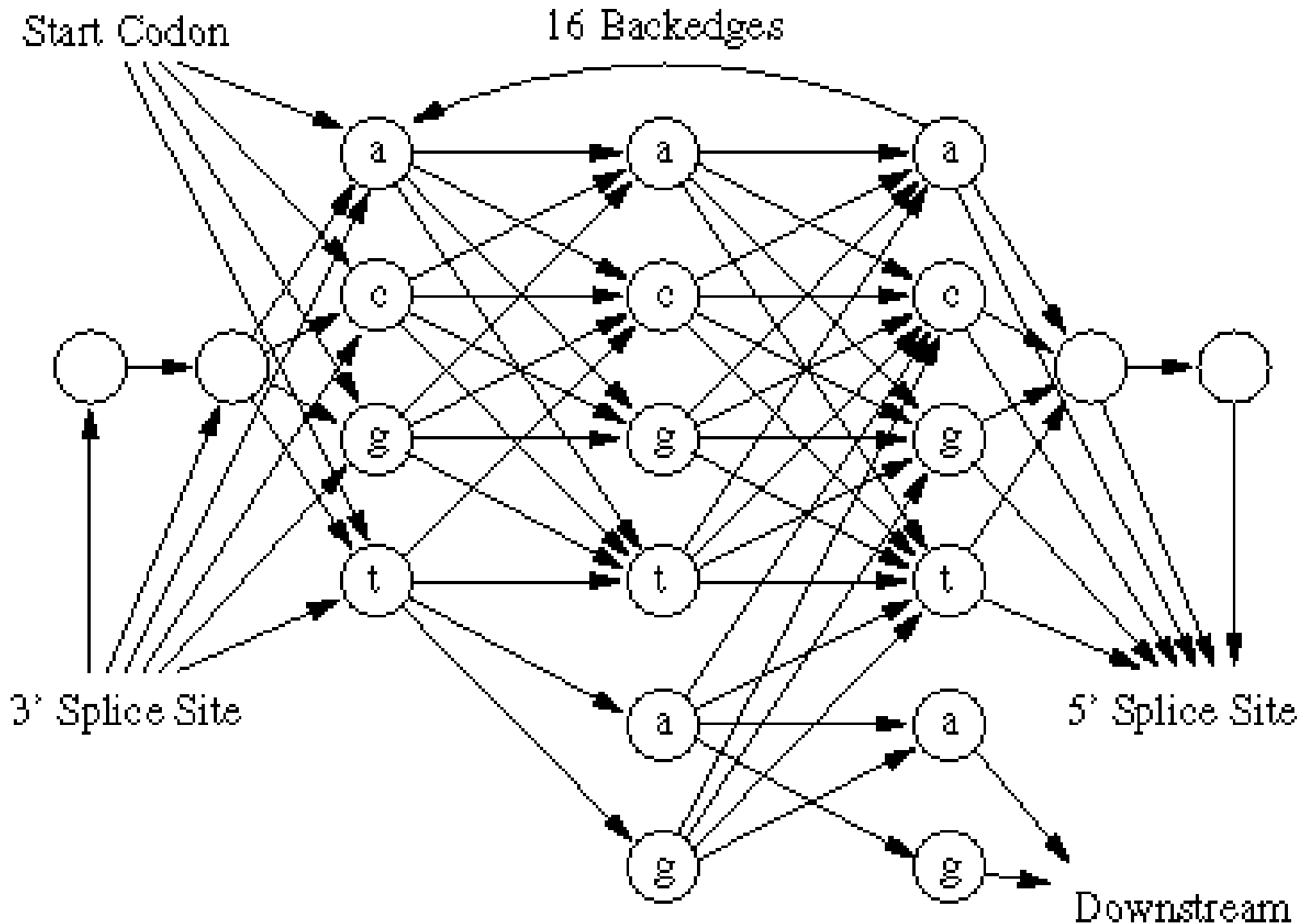
**Two “Generalized” Weight Matrices**



# Neural Networks



# HMM for Gene Finding



# Combined Methods

- **Bring 2 or more methods together (usually site detection + composition)**
- **GRAIL** (<http://compbio.ornl.gov/Grail-1.3/>)
- **FGENEH** (<http://genomic.sanger.ac.uk/gf/gf.shtml>)
- **HMMgene** (<http://www.cbs.dtu.dk/services/HMMgene/>)
- **GENSCAN**(<http://genes.mit.edu/GENSCAN.html>)
- **Gene Parser** (<http://beagle.colorado.edu/~eesnyder/GeneParser.html>)
- **GRPL (GeneTool/BioTools)**

# How Well Do They Do?

<i>Programs</i>	<i># of seq</i>	<i>Nucleotide accuracy</i>				<i>Exon accuracy</i>								
		<i>Sn</i>	<i>Sp</i>	<i>AC</i>	<i>CC</i>	<i>ESn</i>	<i>ESp</i>	$(ESn+ESp)/2$	<i>ME</i>	<i>WE</i>	<i>PCa</i>	<i>PCp</i>	<i>OL</i>	
FGENES	195(5)	0.86	0.88	0.84	0.83	0.67	0.67	0.69	0.12	0.09	0.20	0.17	0.02	
GeneMark	195(0)	0.87	0.89	0.84	0.83	0.53	0.54	0.54	0.13	0.11	0.29	0.27	0.09	
Gene	195(15)	0.91	0.90	0.89	0.88	0.71	0.70	0.71	0.19	0.11	0.15	0.15	0.02	
Genscan	195(3)	0.95	0.90	0.91	0.91	0.70	0.70	0.71	0.08	0.09	0.21	0.19	0.02	
HMMgene	195(5)	0.93	0.93	0.91	0.91	0.76	0.77	0.76	0.12	0.07	0.14	0.14	0.02	
Morgan	127(0)	0.75	0.74	0.70	0.69	0.46	0.41	0.43	0.20	0.28	0.28	0.25	0.07	
MZEF	119(8)	0.70	0.73	0.68	0.66	0.58	0.59	0.59	0.32	0.23	0.08	0.16	0.01	

"Evaluation of gene finding programs" S. Rogic, A. K. Mackworth and B. F. F. Ouellette. Genome Research, 11: 817-832 (2001).

# GenomeScan -

<http://genes.mit.edu/genomescan.html>

## Run GenomeScan:

Organism:

Sequence name (optional):

Print options:

Upload your DNA sequence file (one-letter code, upper or lower case, spaces/numbers ignored):

**Browse...**

Or paste your DNA sequence here (one-letter code, upper or lower case, spaces/numbers ignored):

# TwinScan -

<http://genes.cs.wustl.edu/>

The screenshot shows the TwinScan web application interface. On the left is a dark red sidebar with navigation links: Home, Run TWINSCAN, Examples, Resources, and Brent Lab. The main content area has a dark teal background with the word "TWINSCAN" in large, bold, red letters at the top. Below the title, there is a form for "Organism:" with a dropdown menu currently showing "Select Organism" and a "(Required)" label. To the right of the form is a small window titled "mouse annotations of the UCSC browser." with buttons for "Human" and "Mous". Below the organism selection, a text box contains the instruction: "You can either upload a text file or cut and paste your sequence into the box below." Underneath this text is a file upload field with a "Browse..." button. A large, empty text area for pasting the sequence is located below the file upload field. At the bottom of the main area are two buttons: "Run TWINSCAN" and "Clear". The browser's address bar at the bottom shows "Document: Done".

# SLAM -

<http://baboon.math.berkeley.edu/~syntenic/slam.html>



The screenshot shows a web browser window displaying the SLAM server interface. At the top, there is a banner with the word "slam" in a stylized font. Below the banner are navigation links: "About", "Download links", "FAQ", and "Help". The main heading reads "The SLAM server: submit pairs of syntenic sequences for gene annotation and alignment". Below this, a paragraph explains the server's configuration for human and mouse sequences. A form for entering an email address is present, followed by two input fields for FASTA sequences, each with a "Browse..." button. At the bottom of the form are "Reset" and "Submit sequences" buttons. The browser's address bar shows the URL, and the taskbar at the bottom indicates the document is "Done".

[About](#) [Download links](#) [FAQ](#) [Help](#)

## The SLAM server: submit pairs of syntenic sequences for gene annotation and alignment

The server is currently configured for human (first sequence) and mouse (second sequence), but will work on other sequences at similar evolutionary distances. Please make sure that both sequences are in the same orientation.


Enter your email address (for obtaining results):

The first sequence (in [FASTA](#) format):

The second sequence (in [FASTA](#) format):

# GeneComber -

<http://www.bioinformatics.ubc.ca/genecomber/submit.php>

**UBiC**  **GeneComber**  
UBC Bioinformatics Centre *ab initio gene prediction server*

[About](#) | [Documentation](#) | [Submit Sequences](#) | [Retrieve Results](#) | [Display Submissions](#) | [Downloads](#)

[contact](#) | [helpdesk](#) | [report bugs](#)

## GeneComber Submission

### Genecomber - Submit a Job

GenBank Accession Number:

Upload FastA DNA sequence:  **Browse...**

Upload Genscan output:  **Browse...**

Genscan Training Set:

Upload HMMGene output:  **Browse...**

Processing Method(s):  EUI  GI  EUI\_Frame

e-mail address (required):

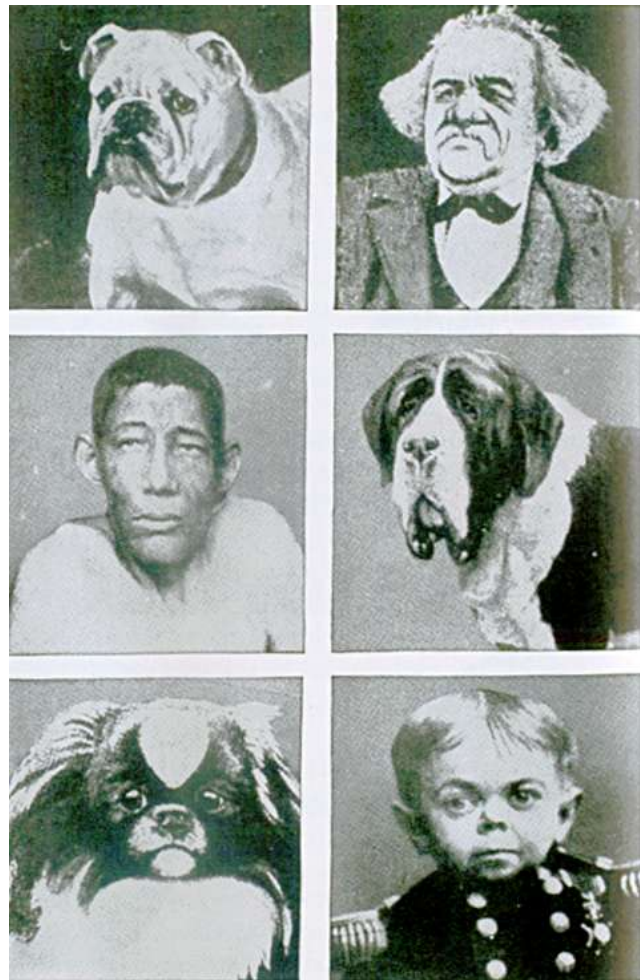
**Submit**

[Home](#) | [About](#) | [Documentation](#) | [Submit Sequences](#) | [Retrieve Results](#) | [Display Submissions](#) | [Downloads](#)

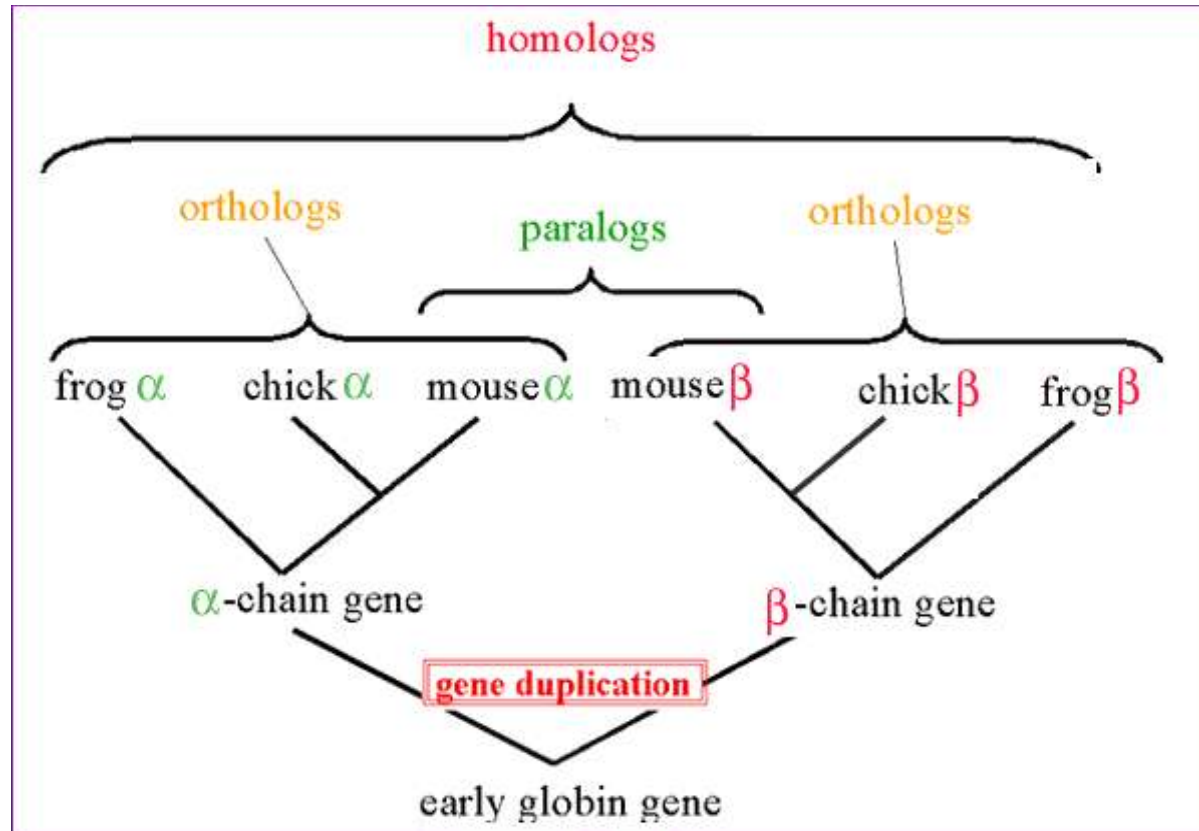
Document: Done



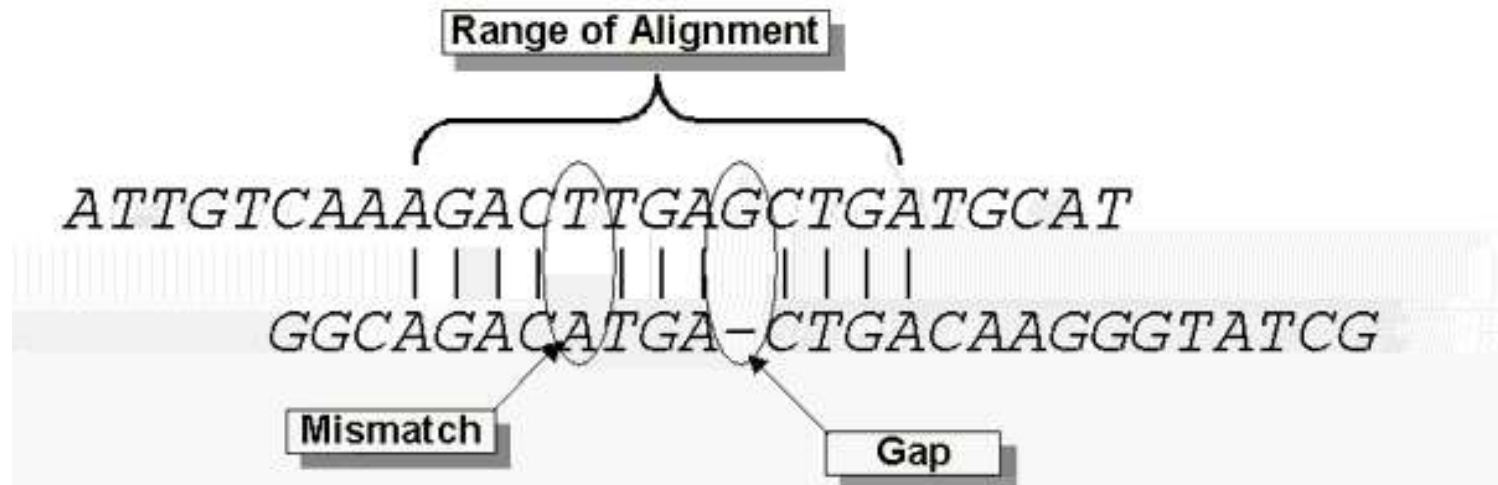
# Srovnávání sekvencí



# Různé kategorie podobnosti



# Hodnocení podobnosti



$$S = \sum(\text{identities, mismatches}) - \sum(\text{gap penalties})$$

$$\text{Score} = \text{Max}(S)$$

# Zarovnání sekvencí

**ACGTGA      ->      ACGTGA      ->**  
**CGTG        ->      CGTG        ->      4**

**ACGTGA**  
**TCGTA**

**ACGTGATGCAG**  
**GGAGAGCACG**

**ACAGTTGACGAGATGGCAGGATGCGCGATGCAGCA**  
**GACGAGCGTGAGTGCGATCGATGACAGTGTATAT**

# Zarovnání sekvencí

**ACGTGA**

**: : : :**

**4**

**CGTG**

**ACGTGA**

**: : : :**

**4**

**TCGT-A**

**ACGTGATGCA-G**

**: : : : :**

**7**

**GGAGA-GCACG**

# Aligning Two Sequences

*ATTGCAGTGATCG*

*ATTGCGTCGATCG*

*Solution 1:*

*Solution 2:*

*ATTGCAGTGATCG*  
| | | | | | | |  
*ATTGCGTCGATCG*

*ATTGCAGT-GATCG*  
| | | | | | | |  
*ATTGC-GTCGATCG*

# Which alignment is better?

*ATTGCAGTGATCG*

*ATTGCGTCGATCG*

*Solution 1:*

*Solution 2:*

*ATTGCAGTGATCG*

*/////       /////*

*ATTGCGTCGATCG*

*ATTGCAGT-GATCG*

*/////   //   /////*

*ATTGC-GTCGATCG*

10 matches+ 3 mismatches

12 matches+2 gaps

# Scoring Scheme

Match	+1
Mismatch	-1
Indel	-2



# Which alignment is better?

*ATTGCAGTGATCG*

*ATTGCGTCGATCG*

*Solution 1:*

*Solution 2:*

*ATTGCAGTGATCG*

*/ / / / / / / / / /*

*ATTGCGTCGATCG*

*ATTGCAGT-GATCG*

*/ / / / / / / / / /*

*ATTGC-GTCGATCG*

Score=7

Score=8

# Finding the best alignment for long sequences is tedious

For two sequences of length 300  
bases there are  $10^{179}$  different  
alignments



Dynamic programming

# Dynamické programování

Needleman-Wunsch (1970)

Smith-Waterman (1981)

- ✦ První krok je triviální a pokrývá částečné řešení
- ✦ Každé další řešení je hodnoceno na základě předcházejících zjištění
- ✦ Zarovnání je tak postupně prodlužováno o další triviální úseky
- ✦ Opakování předchozích kroků vyústí v konečné řešení

# Dynamic Programming Algorithm

Seq 1) \* A G C  
Seq 2) \* A A A C

		A	G	C
	0	1	2	3
0				
A 1				
A 2				
A 3				
C 4				

Needelman-Wunsch algorithm (1970)

# Dynamic Programming Algorithm

```
* - - - - A G C
* A A A C
```

```
match=1
mismatch=-1
indel=-2
```

		<b>A</b>	<b>G</b>	<b>C</b>
	<b>0</b>	<b>1</b>	<b>2</b>	<b>3</b>
<b>0</b>	0	-2	-4	-6
<b>A 1</b>	-2			
<b>A 2</b>	-4			
<b>A 3</b>	-6			
<b>C 4</b>	-8			

# Dynamic Programming Algorithm

\* A G C

\* A - - A A C

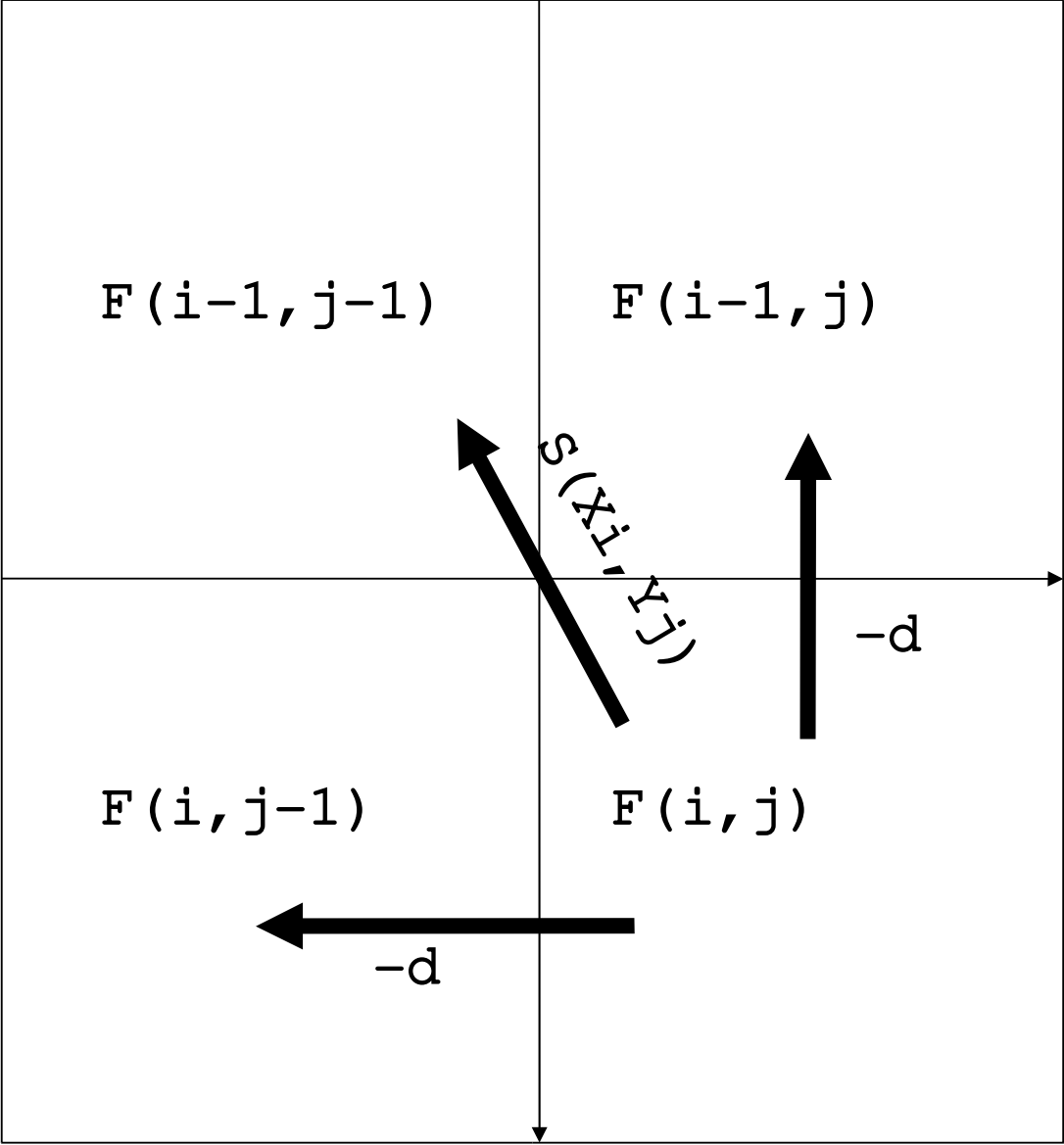
match=1

mismatch=-1

indel=-2

		A	G	C
0	0	-2	-4	-6
A 1	-2	1	-1	-3
A 2	-4			
A 3	-6			
C 4	-8			

Arrows indicate the path from (0,0) to (1,1), (1,1) to (1,2), and (1,2) to (1,3).



# Global pairwise alignment

$$F(i,j) = \max \left\{ \begin{array}{l} F(i-1, j-1) + s(x_i, y_j) \\ F(i-1, j) - d \\ F(i, j-1) - d \end{array} \right.$$



# Finding the Best Score

		A	G	C
	0	1	2	3
0	0 ← -2 ← -4 ← -6			
A 1	-2 ↑ ↘ 1 ← -1 ← -3			
A 2	-4 ↑ ↘ -1 ↑ ↘ 0 ← -2			
A 3	-6 ↑ ↘ -3 ↑ ↘ -2 ↑ ↘ -1			
C 4	-8 ↑ ↘ -5 ↑ ↘ -4 ↑ ↘ -1			

# Tracing the Best Alignment

		A	G	C
	0	1	2	3
0	0	-2	-4	-6
A 1	-2	1	-1	-3
A 2	-4	-1	0	-2
A 3	-6	-3	-2	-1
C 4	-8	-5	-4	-1

Tracing arrows: Red arrows point from (0,0) to (1,1), (1,1) to (2,2), (2,2) to (3,3), and (3,3) to (4,4). Black arrows point from (0,0) to (1,0), (1,0) to (2,1), (2,1) to (3,2), (3,2) to (4,3), (0,0) to (1,1), (1,1) to (2,0), (2,0) to (3,1), (3,1) to (4,2), (0,1) to (1,2), (1,2) to (2,3), (2,3) to (3,4), (0,2) to (1,3), (1,3) to (2,4), (0,3) to (1,4), (1,4) to (2,5), (2,5) to (3,6), (3,6) to (4,7).

A	G	-	C
A	A	A	C

# Tracing the Best Alignment

		A	G	C
	0	1	2	3
0	0 ← -2 ← -4 ← -6			
A 1	↑ -2    1 ← -1 ← -3	↑		
A 2	↑ -4    ↑ -1    0 ← -2		↑	
A 3	↑ -6    ↑ -3    ↑ -2    -1			↑
C 4	↑ -8    ↑ -5    ↑ -4    -1			

A	-	G	C
A	A	A	C

# Tracing the Best Alignment

		A	G	C
	0	1	2	3
0	0	-2	-4	-6
A 1	-2	1	-1	-3
A 2	-4	-1	0	-2
A 3	-6	-3	-2	-1
C 4	-8	-5	-4	-1

Red arrows trace the path from (0,0) to (4,4). The cell (4,4) containing -1 is circled in red.

-	A	G	C
A	A	A	C

# Local Alignment Example

		A	T	C	T	A	A
	0	1	2	3	4	5	6
0							
ATCTAA							
T 1							
A 2							
A 3							
T 4							
A 5							

Smith-Waterman algorithm, 1981

# Local Alignment

$$F(i,j) = \max \left\{ \begin{array}{l} F(i-1, j-1) + s(x_i, y_j) \\ F(i-1, j) - d \\ F(i, j-1) - d \\ 0 \end{array} \right.$$

# Local Alignment Example

TCATAA  
TAATA

		T	A	C	T	A	A
	0	1	2	3	4	5	6
0	0	0	0	0	0	0	0
T 1	0	1	0	0	1	0	0
A 2	0	0	2	0	0	2	1
A 3	0	0	1	1	0	1	3
T 4	0	0	0	0	2	0	1
A 5	0	0	1	0	0	3	1

# Local Alignment Example



		T	A	C	T	A	A
	0	1	2	3	4	5	6
0	0	0	0	0	0	0	0
T 1	0	1	0	0	1	0	0
A 2	0	0	2	0	0	2	1
A 3	0	0	1	1	0	1	3
T 4	0	0	0	0	2	0	1
A 5	0	0	1	0	0	3	1



# Local Alignment Example

TACTAA  
TAATA

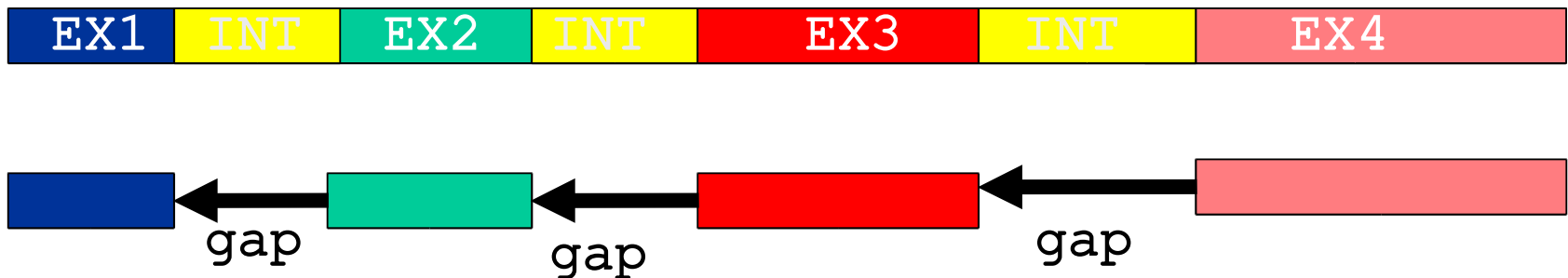
		T	A	C	T	A	A
	0	1	2	3	4	5	6
0	0	0	0	0	0	0	0
T 1	0	1	0	0	1	0	0
A 2	0	0	2	0	0	2	1
A 3	0	0	1	1	0	1	3
T 4	0	0	0	0	2	0	1
A 5	0	0	1	0	0	3	1

# Examples :

## Genomic DNA versus mRNA



Alignment



# Gap Penalties

AAC-AATTAAG-ACTAC-GTTCATGAC

A-CGA-TTA-GCAC-ACTG-T-A-GA-

AACAATTAAGACTACGTTCATGAC---

AACAATT-----GTTCATGACGCA

# Scoring Gaps

I  
AAC-AATTAAG-ACTAC-GTTCATGAC -6  
A-CGA-TTA-GCAC-ACTG-T-A-GA-

II  
AACCAATTAAGACTACGTTCATGAC--- 12  
AACCAATT-----GTTCATGACGCA

Scoring parameters  
match:+1;Gap\_open:-2

# Scoring Insertions/Deletions

AAC-AATTAAG-ACTAC-GTTCATGAC -6

I

A-CGA-TTA-GCAC-ACTG-T-A-GA-

AACAATTAAGACTACGTTCATGAC--- -6

II

AACAATT-----GTTCATGACGCA

Scoring parameters  
match:+1;indel:-2

# Considering Gap Opening and Gap Extension

I AAC-AATTAAG-ACTAC-GTTCATGAC -17  
A-CGA-TTA-GCAC-ACTG-T-A-GA-

II AACCAATTAAGACTACGTTCATGAC--- 1  
AACCAATT-----GTTCATGACGCA

Scoring parameters

match:+1; Gap\_open:-2; Gap\_exten:-1

