

Před analýzou

>P12345 Yeast chromosome1

GATTACAGATTACAGATTACAGATTACAGATTACAG
ATTACAGATTACAGATTACAGATTACAGATTACAGA
TTACAGATTACAGATTACAGATTACAGATTACAGAT
TACAGATTAGAGATTACAGATTACAGATTACAGATT
ACAGATTACAGATTACAGATTACAGATTACAGATTA
CAGATTACAGATTACAGATTACAGATTACAGATTAC
AGATTACAGATTACAGATTACAGATTACAGATTACA
GATTACAGATTACAGATTACAGATTACAGATTACAG
ATTACAGATTACAGATTACAGATTACAGATTACAGA
TTACAGATTACAGATTACAGATTACAGATTACAGAT

Po částečné analýze

>P12345 Gene_1 - gen kodující
protein alkoholdehydrogenazy ...

TATA	TATAAA
	CGATTGACGATGACGAT
start	ATG
exon1	TACAGATTACAGATTACAGATTACAGATGT
intron1	CAGATTACAGATTACAGATTACAGATTACAGATTCA
exon2	AGATTACAGATTACAGATTACAGA
stop	TAA

>P12346 Protein_1
MASAQSFYLLDHNQNQNFDDHLAVDIVMILSHERFMN

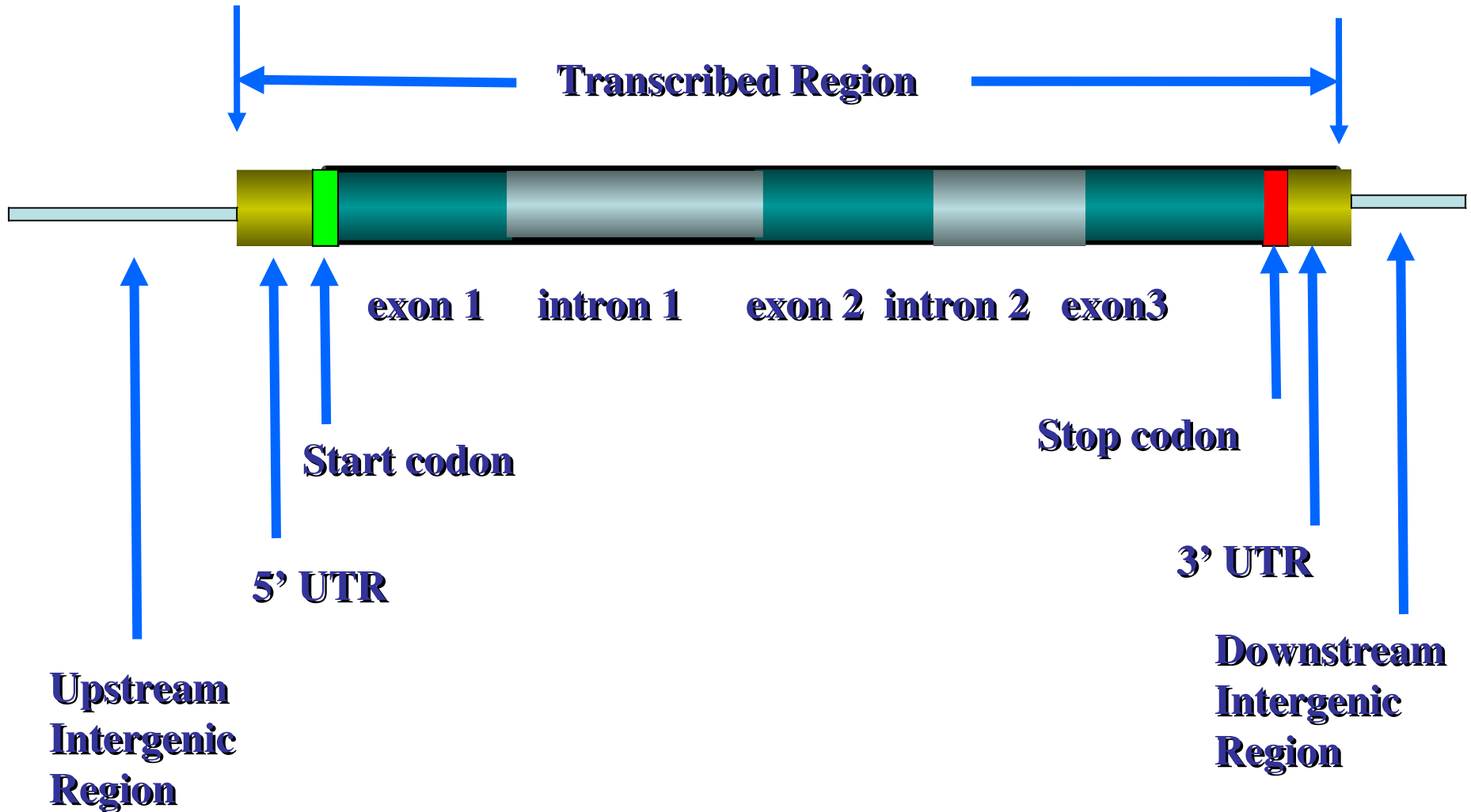
Analýza DNA sekvence

- ☀ = ~ anotace genomu (sekvence)
 - ✳ identifikace signálů a genů
 - ✳ anotace genů (jejich kódujících sekvencí)

Anotace genů \approx anotace proteinů

- ✦ Identifikace a popis fyzikálně-chemických, funkčních a strukturních vlastností daného genu/proteinu
 - ✦ sekvence DNA, AA, pozice v genomu, délka, složení
 - ✦ běžné názvy, odkazy na literaturu
 - ✦ příslušnost do rodiny, evoluce
 - ✦ partneři pro interakci, aktivita, regulační mechanismy
 - ✦ struktura, aktivní místa, role v metabolismu buňky

Eukaryotic Gene Structure



Analýza DNA sekvence

- ✦ Statistika

- ✦ frekvence n-gramů a jiných prvků, repetice, kodony

- ✦ Signální prvky

- ✦ TATA (promotor), ATG (start), STOP, GT (donor), AG (akceptor) a pod

- ✦ Kódující část

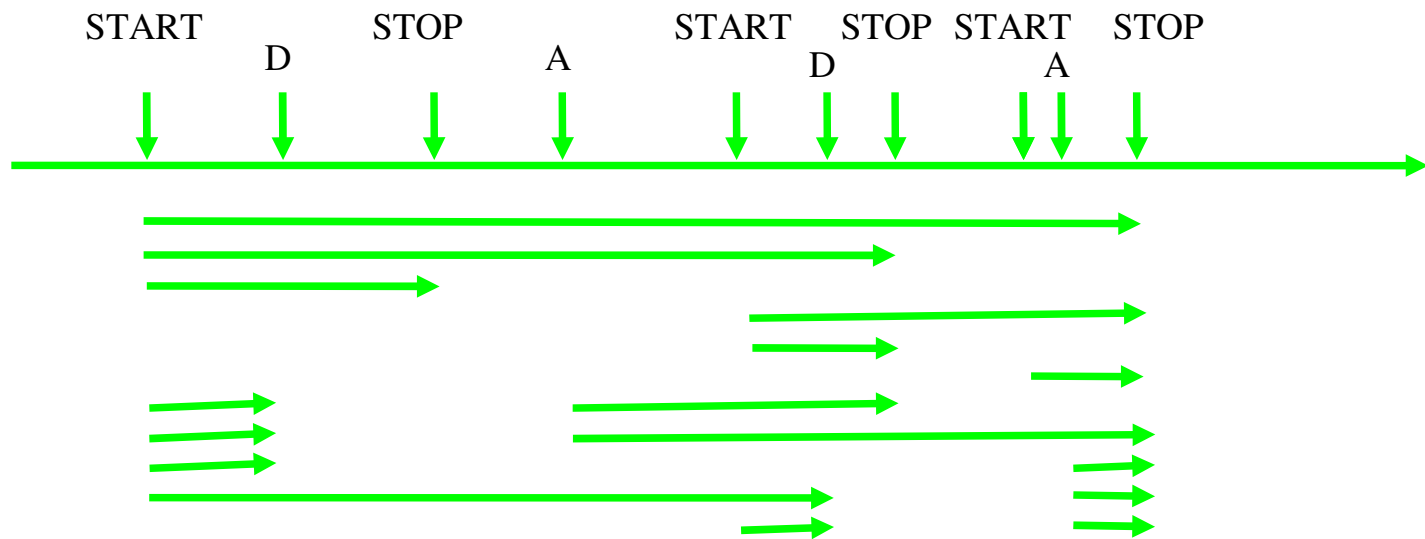
- ✦ podobnost kódované sekvence s jinými proteiny

- ✦ Kombinované přístupy

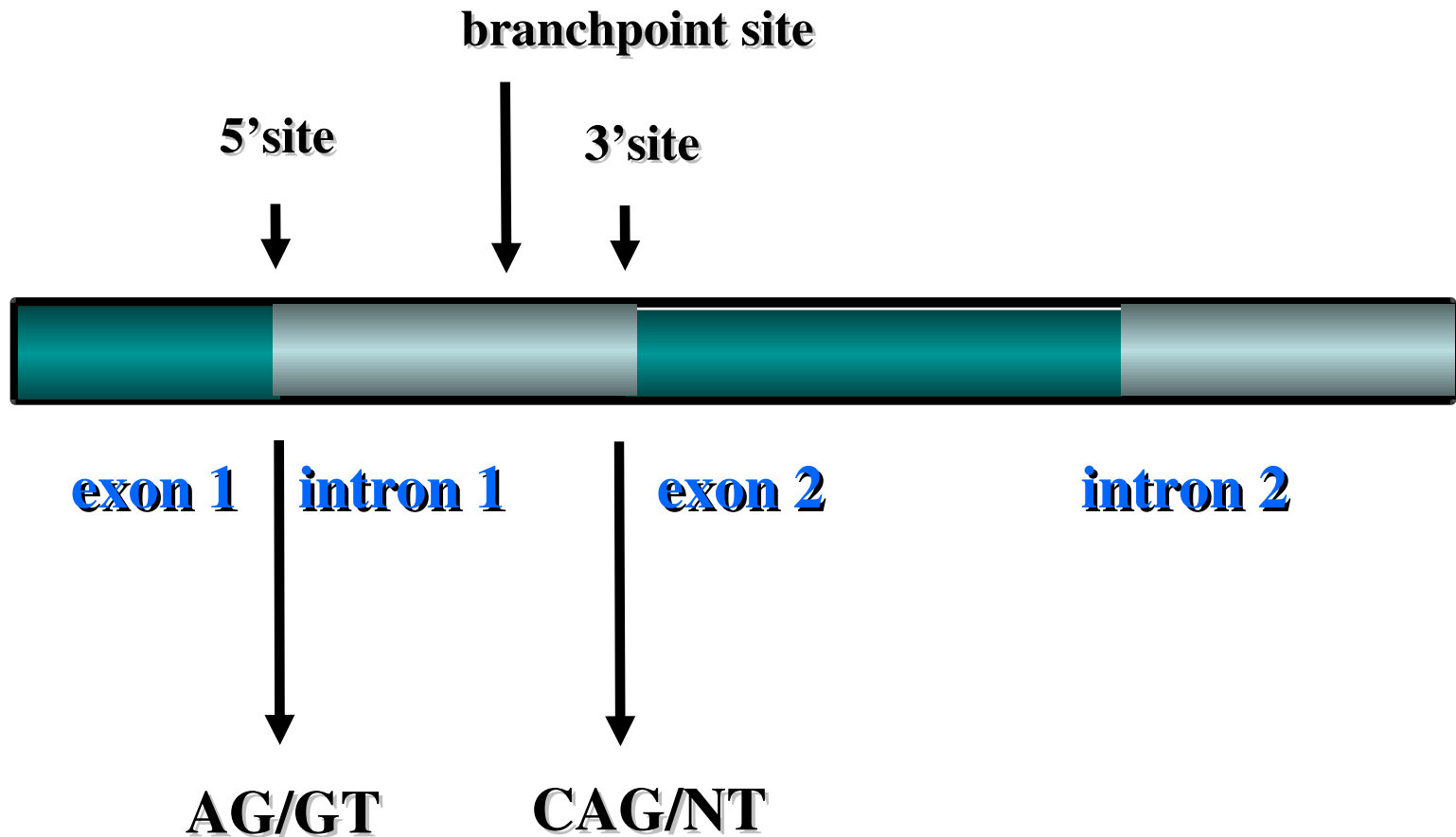
Identifikace genů

- ☀ U prokaryotů 95-100% spolehlivost, u složitějších eukaryotů 90% na úrovni bazí, 70% na úrovni exonů/intronů
 - ✳ existence intronů
 - ✳ větší genomy
 - ✳ nízká hustota genů (<30%; 3% u Homo sapiens)
 - ✳ alternativní splicing (zhruba u poloviny genů)
 - ✳ velké množství repetitivních sekvencí
 - ✳ občasný překryv genů

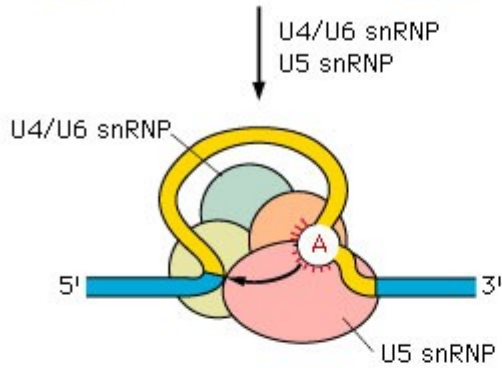
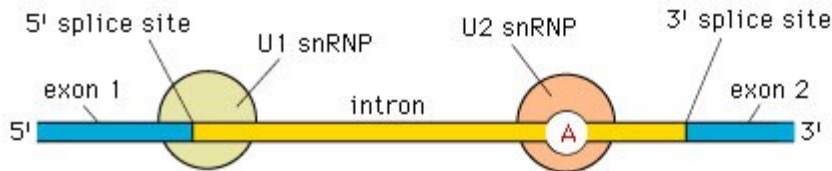
Identifikace genů



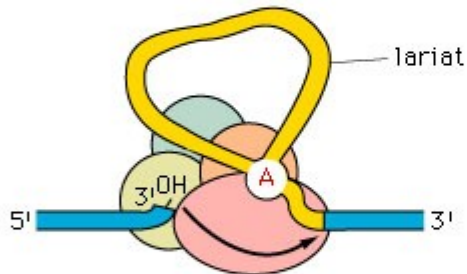
Eukaryotic Gene Structure



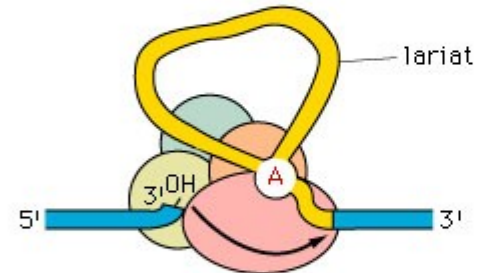
RNA Splicing



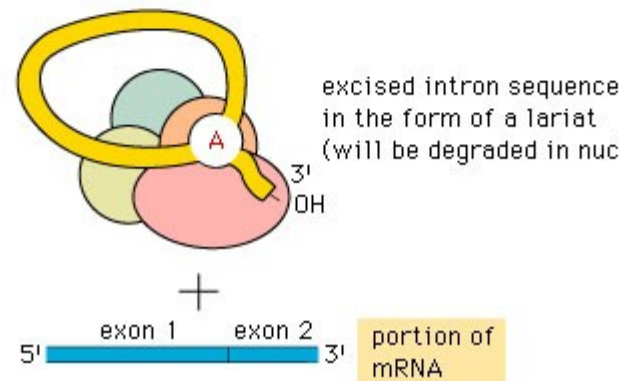
LARIAT FORMATION
AND 5' SPLICE SITE
CLEAVAGE



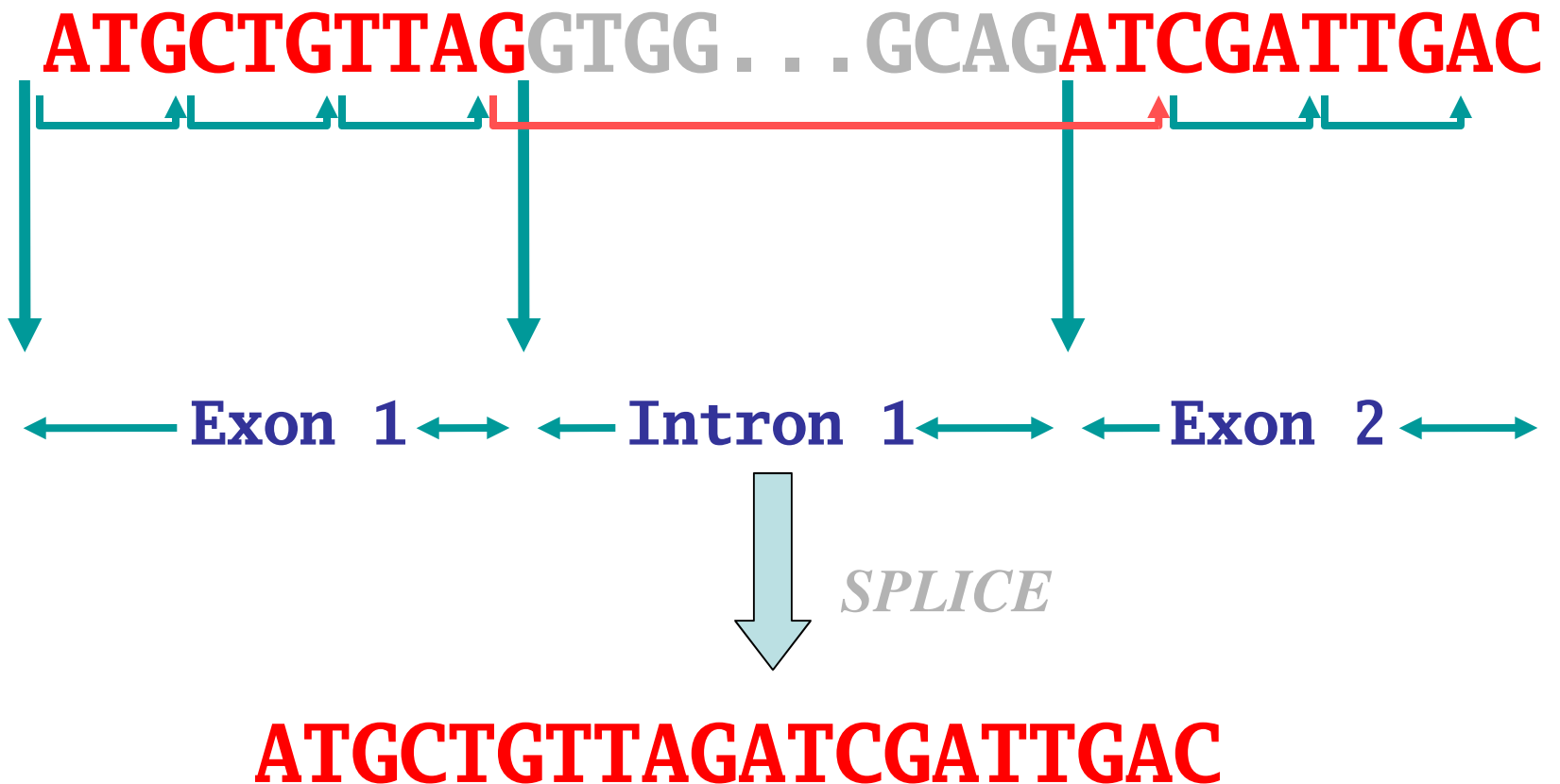
LARIAT FORMATION
AND 5' SPLICE SITE
CLEAVAGE



3' SPLICE SITE
CLEAVAGE AND
JOINING OF TWO
EXON SEQUENCES



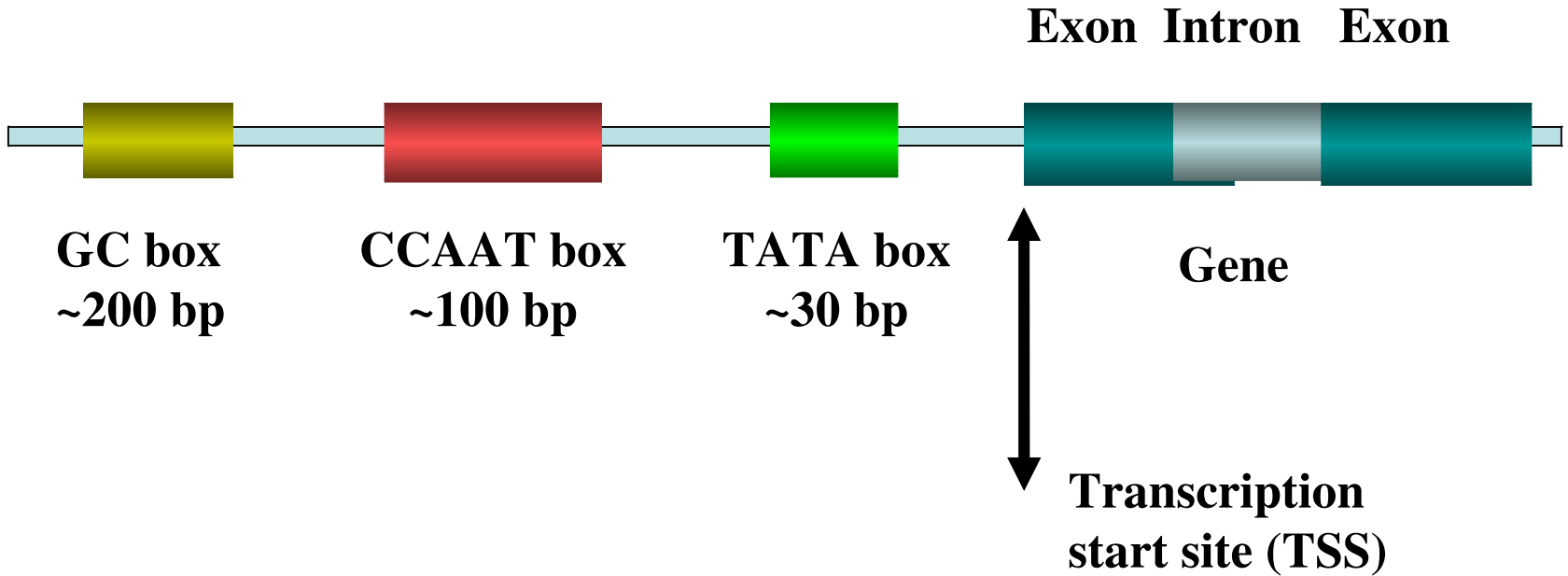
Exon/Intron Structure (Detail)



Typické signály v eukaryotických sekvencích

- ✦ Promotorové elementy
 - ✦ CAP, CCAAT, GC a TATA
- ✦ Kozakova sekvence (rozpoznávána ribozomem = RBS)
- ✦ Splicing (donor, acceptor a lariat)
- ✦ Terminační signál
- ✦ Polyadenylační signál

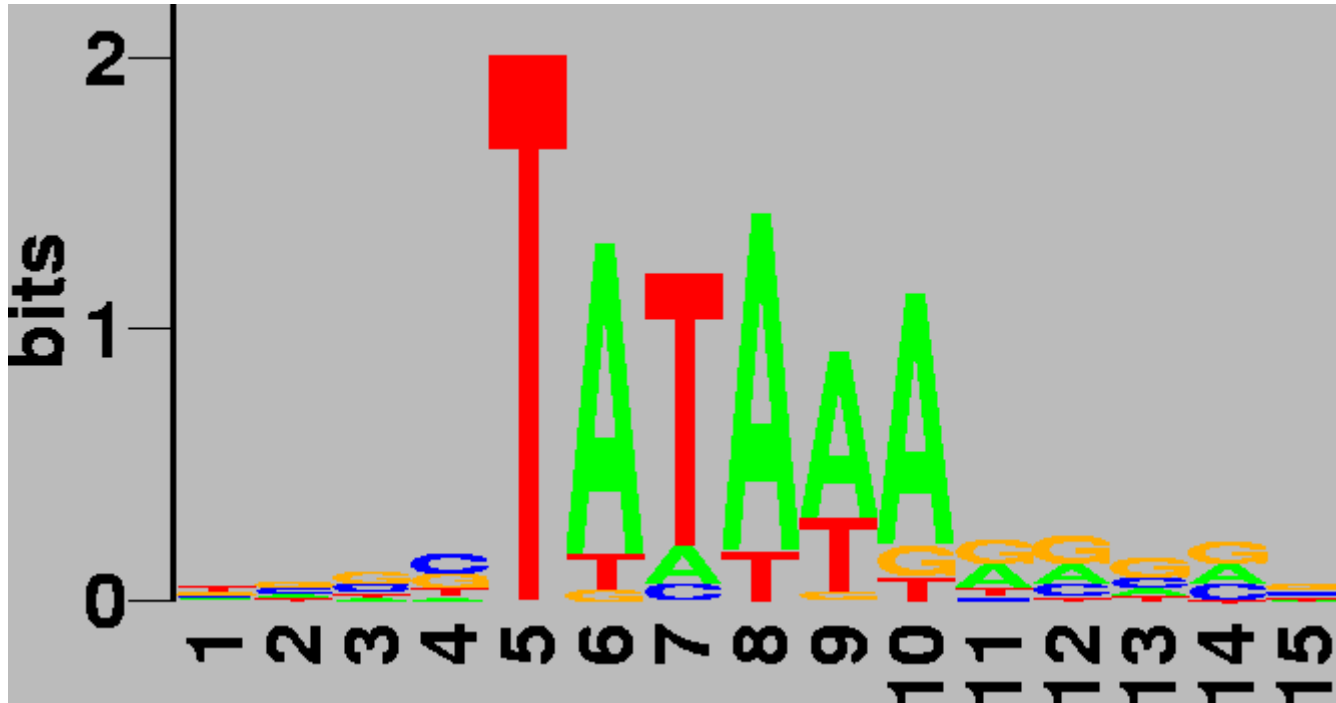
Pol II Promoter Elements



Pol II Promoter Elements

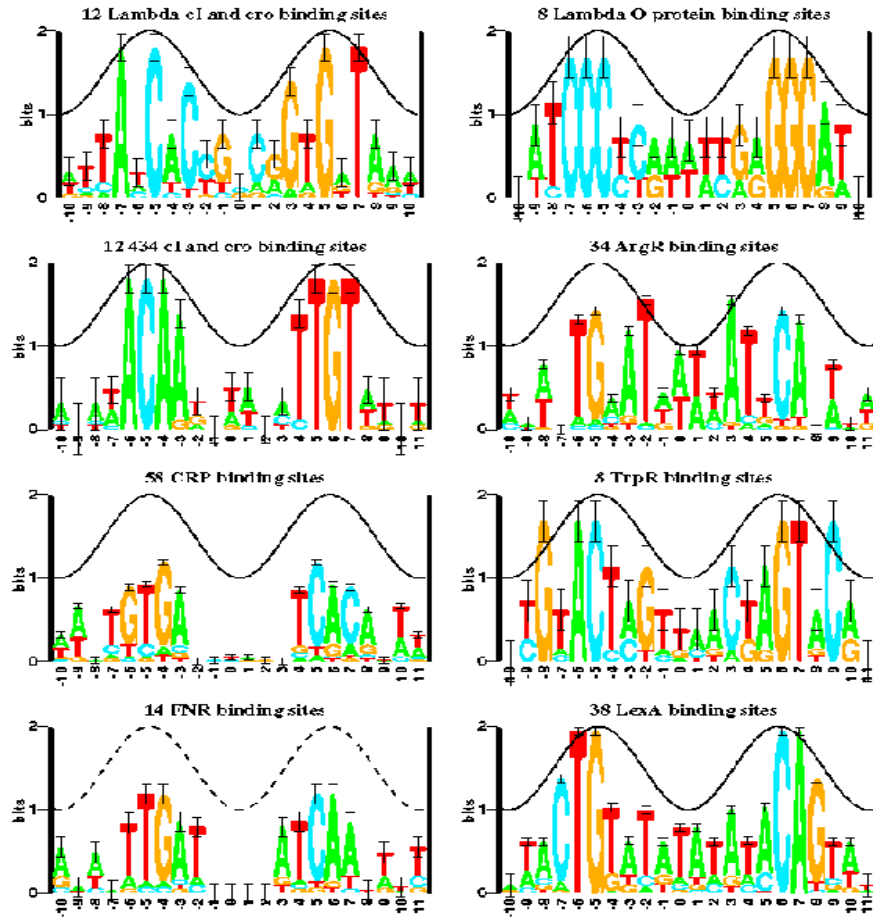
- **Cap Region/Signal**
 - **n C A G T n G**
- **TATA box (~ 25 bp upstream)**
 - **T A T A A n G C C C**
- **CCAAT box (~100 bp upstream)**
 - **T A G C C A A T G**
- **GC box (~200 bp upstream)**
 - **A T A G G C G nGA**

Pol II Promoter Elements



TATA box is found in ~70% of promoters

WebLogos



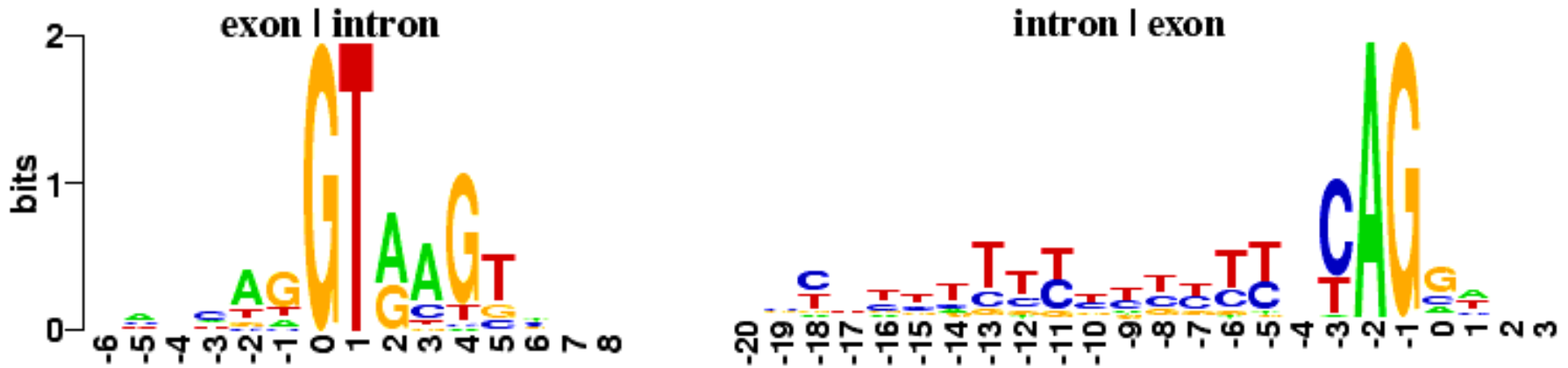
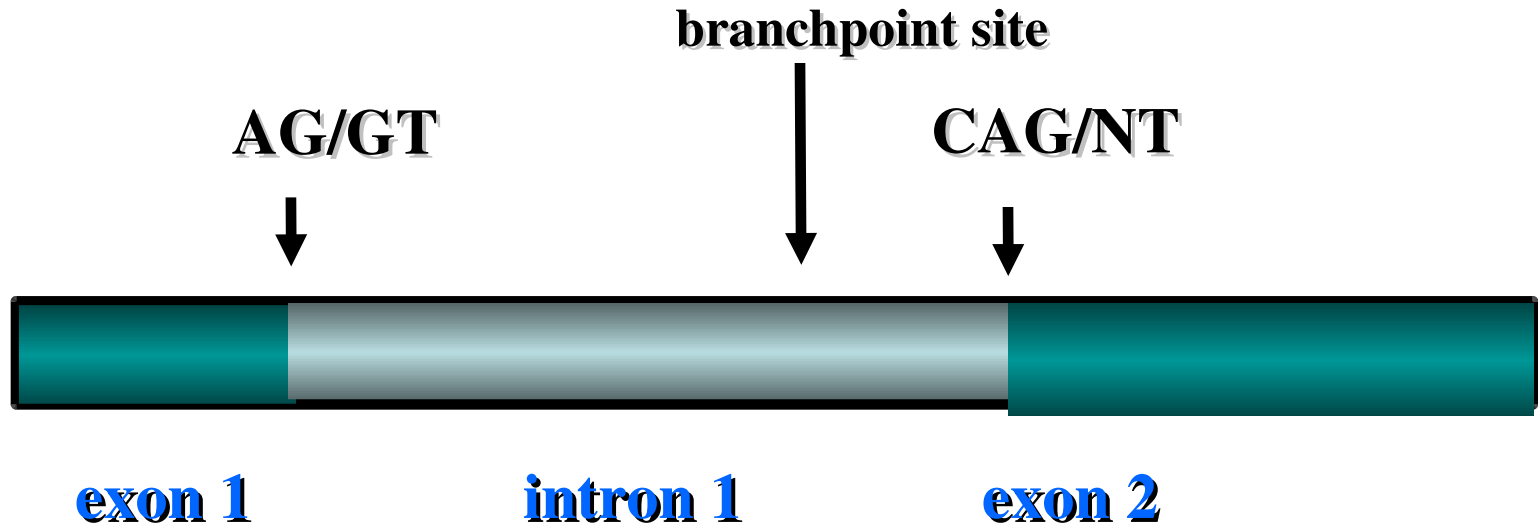
<http://www.bio.cam.ac.uk/cgi-bin/seqlogo/logo.cgi>

Kozak (RBS) Sequence

-7 -6 -5 -4 -3 -2 -1 0 1 2 3
A G C C A C C **A T G** G



Splice Signals

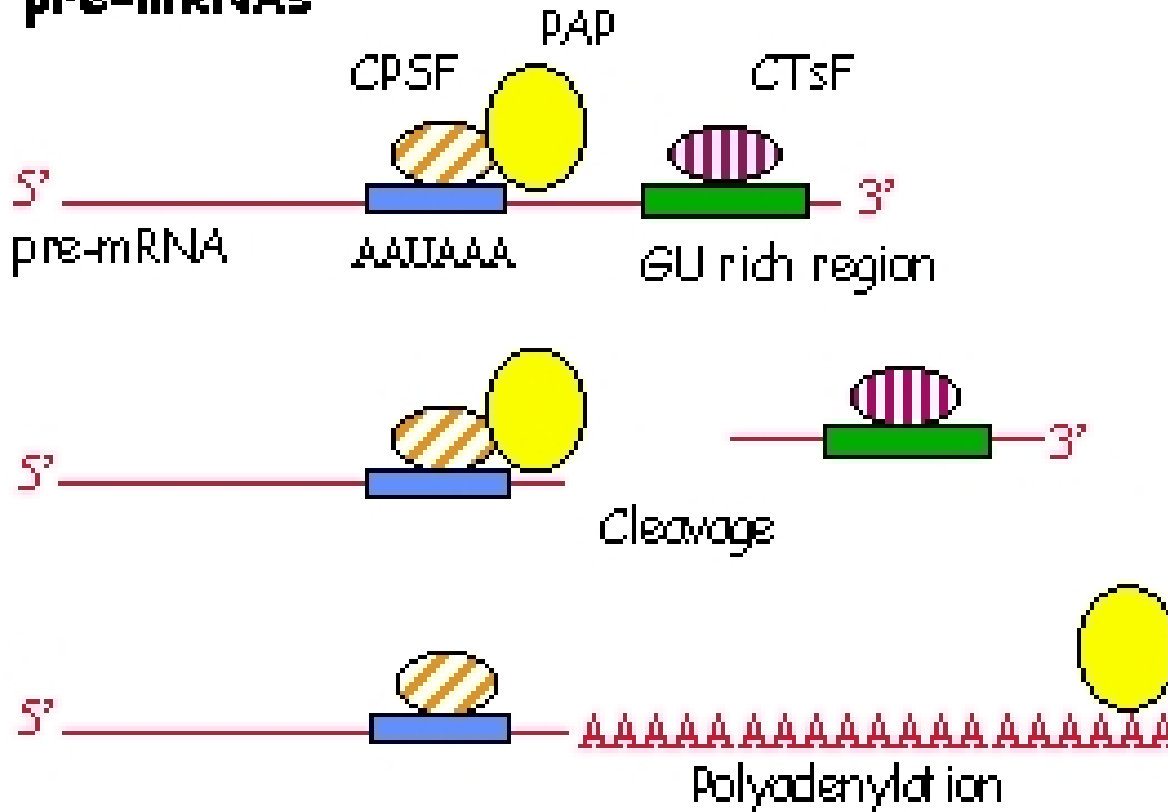


Miscellaneous Signals

- **Polyadenylation signal**
 - **A A T A A A or A T T A A A**
 - Located 20 bp upstream of poly-A cleavage site
- **Termination Signal**
 - **A G T G T T C A**
 - Located ~30 bp downstream of poly-A cleavage site

Polyadenylation

Cleavage and Polyadenylation of Eukaryotic pre-mRNAs



CPSF – Cleavage & Polyadenylation Specificity Factor

PAP – Poly-A Polymerase

CTsF – Cleavage Stimulation Factor

Analýza genomu – kombinované metody

- ☀ Neurónové sítě

- ☀ Grail, GeneParser

- ☀ Lineární diskriminační analýza

- ☀ GeneFinder, GeneID, MZEF

- ☀ Lingvistická

- ☀ GeneLang

- ☀ Markovovy řetězce

- ☀ Genie, GeneMark, GenScan, VEIL

- ☀ Podobnosti

- ☀ Procrustes, AAT

- ☀ Rozhodovací stromy

Neural Network

Training Set

ACGAAG
AGGAAG
AGCAAG
ACGAAA
AGCAAC



Definitions

A = [001]
C = [010]
G = [100]



E = [01]
N = [00]

Sliding Window

ACGAAG



[010100001]

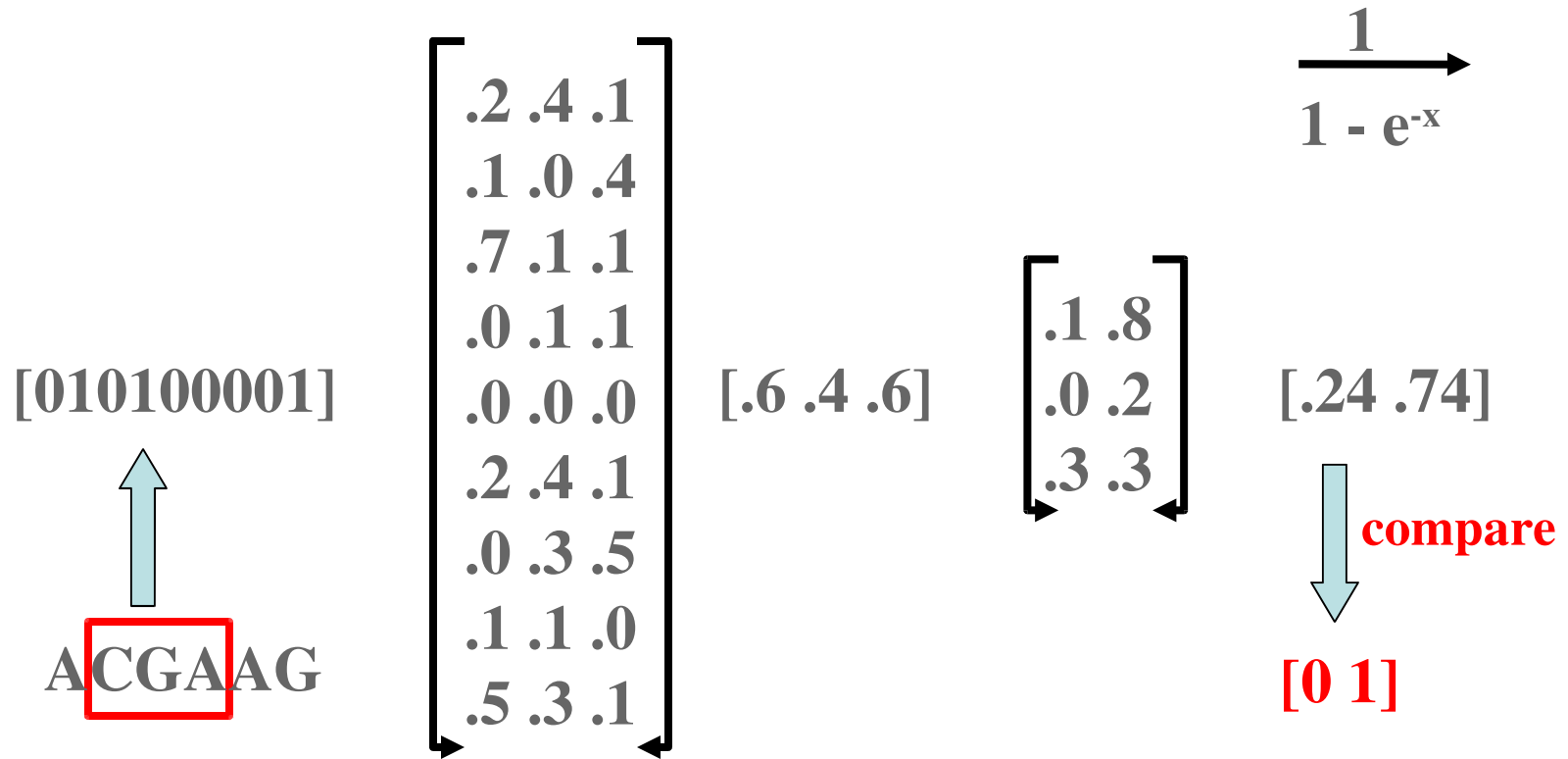
Input Vector

[01]

Output Vector

Desired Output

Neural Network Training



**Input
Vector**

**Weight
Matrix1**

**Hidden
Layer**

**Weight
Matrix2**

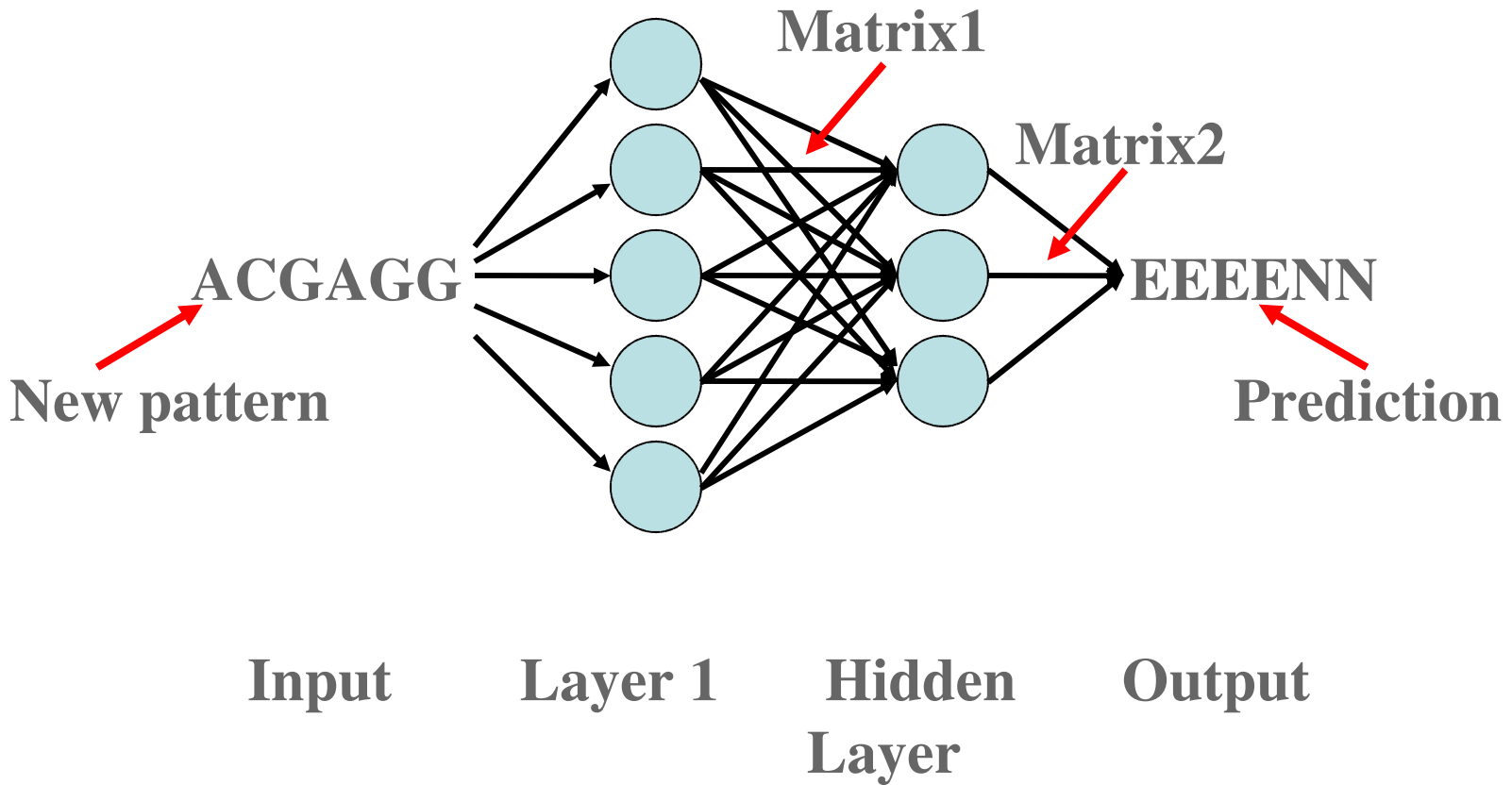
**Output
Vector**

After Many Iterations....

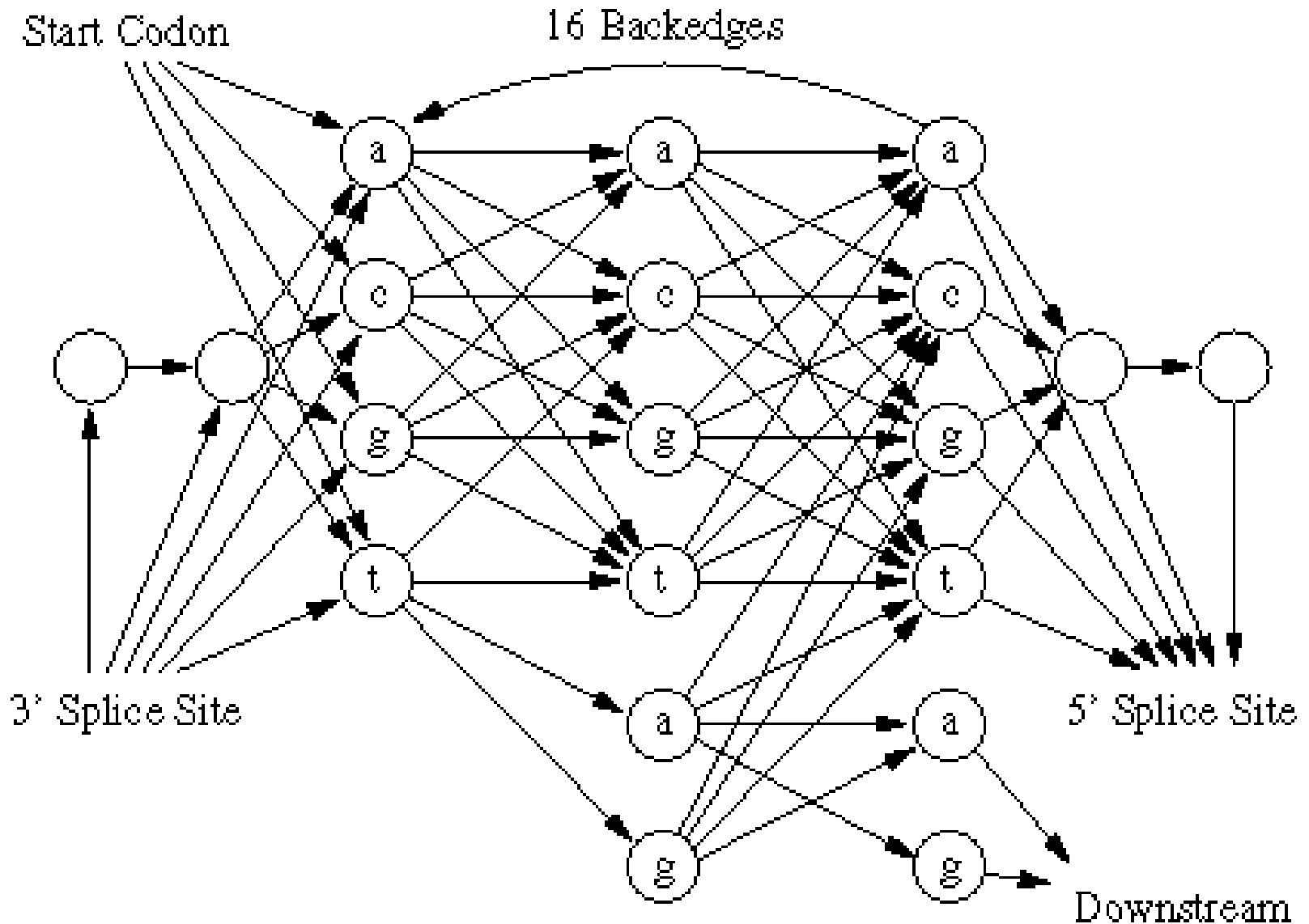
$$\begin{bmatrix} .13 & .08 & .12 \\ .24 & .01 & .45 \\ .76 & .01 & .31 \\ .06 & .32 & .14 \\ .03 & .11 & .23 \\ .21 & .21 & .51 \\ .10 & .33 & .85 \\ .12 & .34 & .09 \\ .51 & .31 & .33 \end{bmatrix} \quad \begin{bmatrix} .03 & .93 \\ .01 & .24 \\ .12 & .23 \end{bmatrix}$$

Two “Generalized” Weight Matrices

Neural Networks



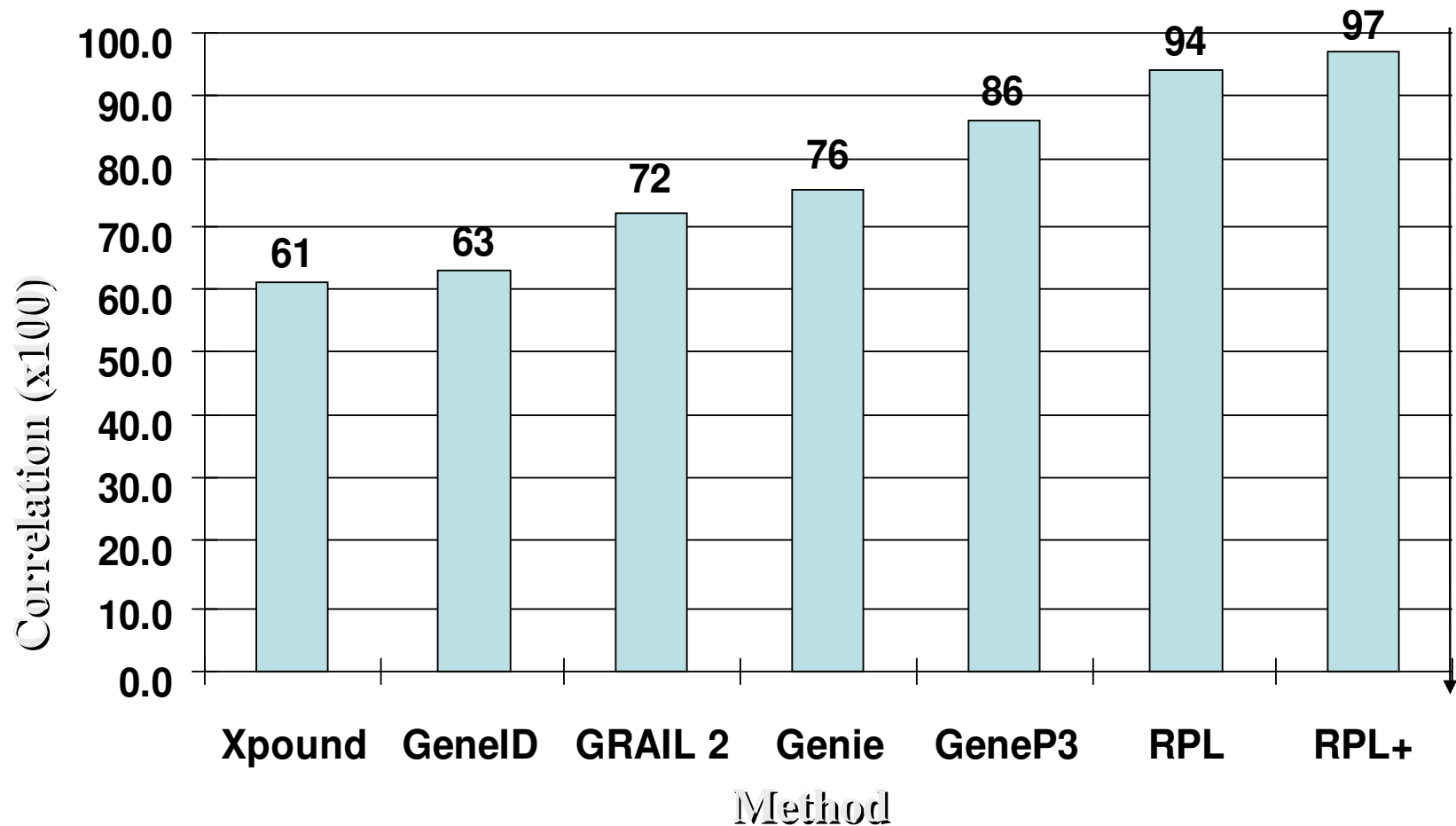
HMM for Gene Finding



Combined Methods

- **Bring 2 or more methods together (usually site detection + composition)**
- **GRAIL (<http://compbio.ornl.gov/Grail-1.3/>)**
- **FGENEH (<http://genomic.sanger.ac.uk/gf/gf.shtml>)**
- **HMMgene (<http://www.cbs.dtu.dk/services/HMMgene/>)**
- **GENSCAN(<http://genes.mit.edu/GENSCAN.html>)**
- **Gene Parser (<http://beagle.colorado.edu/~eesnyder/GeneParser.html>)**
- **GRPL (GeneTool/BioTools)**

How Well Do They Do?



Burset & Guigio test set (1996)

How Well Do They Do?

<i>Programs</i>	<i># of seq</i>	<i>Nucleotide accuracy</i>				<i>Exon accuracy</i>								
		<i>Sn</i>	<i>Sp</i>	<i>AC</i>	<i>CC</i>	<i>ESn</i>	<i>ESp</i>	$(ESn+ESp)/2$	<i>ME</i>	<i>WE</i>	<i>PCa</i>	<i>PCp</i>	<i>OL</i>	
FGENES	195(5)	0.86	0.88	0.84	0.83	0.67	0.67	0.69	0.12	0.09	0.20	0.17	0.02	
GeneMark	195(0)	0.87	0.89	0.84	0.83	0.53	0.54	0.54	0.13	0.11	0.29	0.27	0.09	
Genie	195(15)	0.91	0.90	0.89	0.88	0.71	0.70	0.71	0.19	0.11	0.15	0.15	0.02	
Genscan	195(3)	0.95	0.90	0.91	0.91	0.70	0.70	0.71	0.08	0.09	0.21	0.19	0.02	
HMMgene	195(5)	0.93	0.93	0.91	0.91	0.76	0.77	0.76	0.12	0.07	0.14	0.14	0.02	
Morgan	127(0)	0.75	0.74	0.70	0.69	0.46	0.41	0.43	0.20	0.28	0.28	0.25	0.07	
MZEF	119(8)	0.70	0.73	0.68	0.66	0.58	0.59	0.59	0.32	0.23	0.08	0.16	0.01	

"Evaluation of gene finding programs" S. Rogic, A. K. Mackworth and B. F. F. Ouellette. *Genome Research*, 11: 817-832 (2001).

GenomeScan -

<http://genes.mit.edu/genomescan.html>

Run GenomeScan:

Organism:

Sequence name (optional):

Print options:

Upload your DNA sequence file (one-letter code, upper or lower case, spaces/numbers ignored):

Browse...

Or paste your DNA sequence here (one-letter code, upper or lower case, spaces/numbers ignored):

TwinScan -

<http://genes.cs.wustl.edu/>

The screenshot shows the TwinScan web application interface. On the left is a dark red sidebar with navigation links: Home, Run TWINSCAN, Examples, Resources, and Brent Lab. The main content area has a dark teal background with the word "TWINSCAN" in large, bold, red letters. Below the title, there is a form with an "Organism:" label, a dropdown menu showing "Select Organism", and a "(Required)" note. To the right of the dropdown is a small window titled "mouse annotations of the UCSC browser." with "Human" and "Mous" buttons. Below the organism selection, there is a text input field with a "Browse..." button. Underneath is a large text area for pasting a sequence. At the bottom of the main area are "Run TWINSCAN" and "Clear" buttons. The browser's address bar shows "Document: Done" and the Windows taskbar is visible at the bottom.

Washington University
St. Louis, MO

TWINSCAN

Organism: (Required)

mouse annotations of the UCSC browser.
Human Mous

You can either upload a text file or cut and paste your sequence into the box below.

Browse...

Run TWINSCAN Clear

Document: Done

SLAM -

<http://baboon.math.berkeley.edu/~syntenic/slam.html>

The SLAM server: submit pairs of syntenic sequences for gene annotation and alignment

The server is currently configured for human (first sequence) and mouse (second sequence), but will work on other sequences at similar evolutionary distances. Please make sure that both sequences are in the same orientation.

Enter your email address (for obtaining results):


The first sequence (in [FASTA](#) format):

The second sequence (in [FASTA](#) format):

Document: Done

GeneComber -

<http://www.bioinformatics.ubc.ca/genecomber/submit.php>

UBiC  **GeneComber**
UBC Bioinformatics Centre *ab initio gene prediction server*

[About](#) | [Documentation](#) | [Submit Sequences](#) | [Retrieve Results](#) | [Display Submissions](#) | [Downloads](#)

[contact](#) | [helpdesk](#) | [report bugs](#)

GeneComber Submission

Genecomber - Submit a Job

GenBank Accession Number:

Upload FastA DNA sequence: **Browse...**

Upload Genscan output: **Browse...**

Genscan Training Set:

Upload HMMGene output: **Browse...**

Processing Method(s): EUI GI EUI_Frame

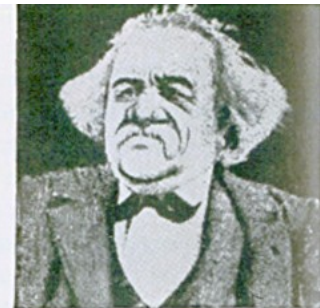
e-mail address (required):

Submit

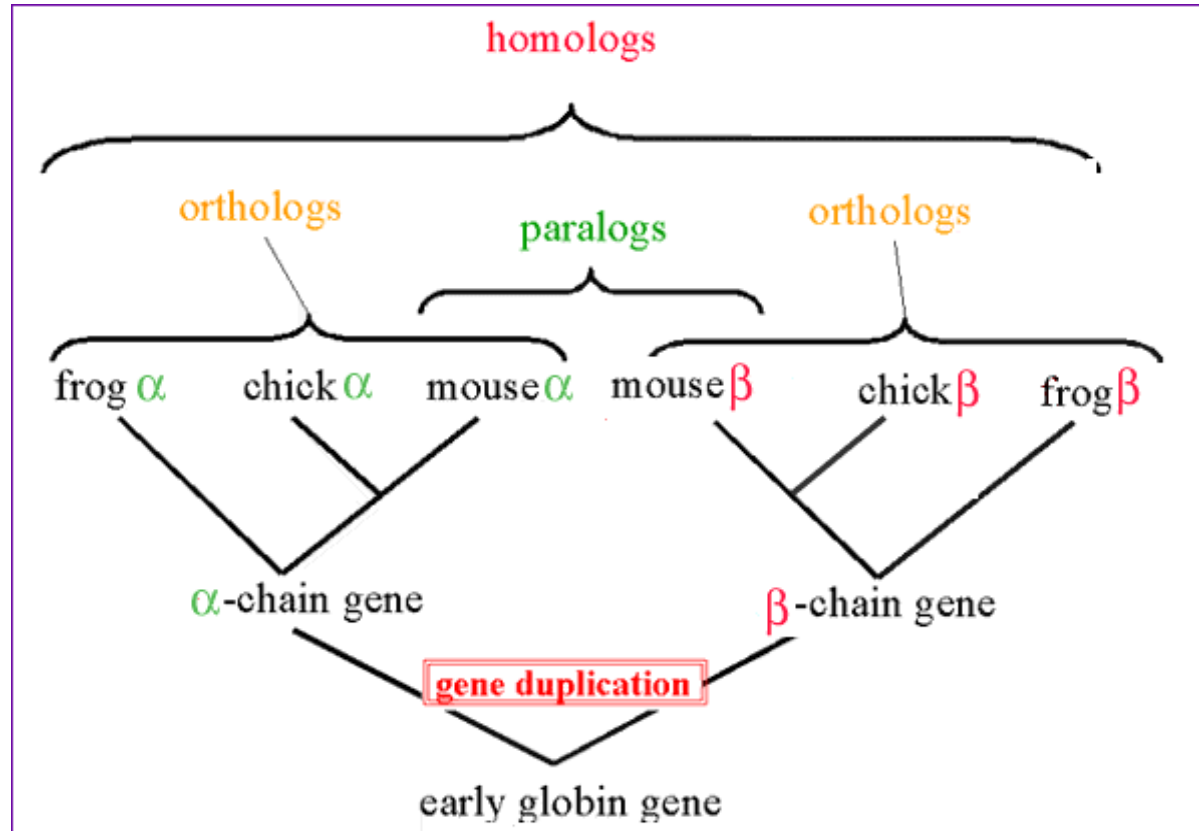
[Home](#) | [About](#) | [Documentation](#) | [Submit Sequences](#) | [Retrieve Results](#) | [Display Submissions](#) | [Downloads](#)

Document: Done

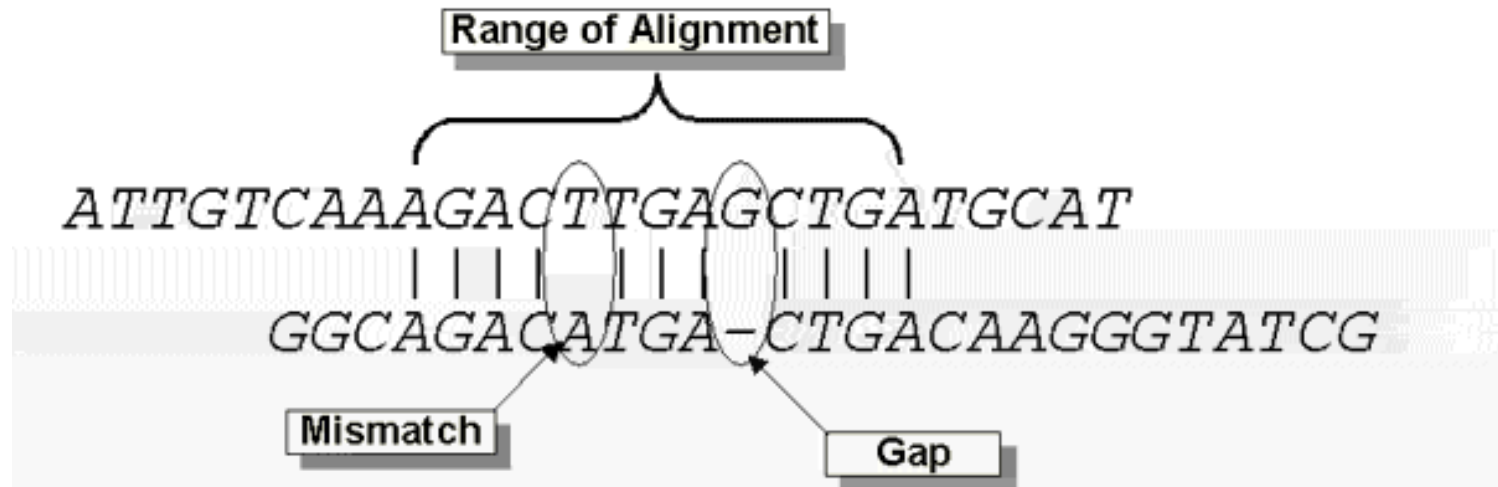
Srovnávání sekvencí



Různé příčiny podobnosti



Hodnocení podobnosti



$$S = \sum(\text{identities, mismatches}) - \sum(\text{gap penalties})$$

$$\text{Score} = \text{Max}(S)$$

Zarovnání sekvencí

ACGTGA -> ACGTGA ->
CGTG -> CGTG -> 4

ACGTGA
TCGTA

ACGTGATGCAG
GGAGAGCACG

ACAGTTGACGAGATGGCAGGATGCGCGATGCAGCA
GACGAGCGTGAGTGCGATCGATGACAGTGTATAT

Zarovnání sekvencí

ACGTGA

: : : :

4

CGTG

ACGTGA

: : : :

4

TCGT-A

ACGTGATGCA-G

: : : : :

7

GGAGA-GCACG

Aligning Two Sequences

ATTGCAGTGATCG

ATTGCGTCGATCG

Solution 1:

Solution 2:

ATTGCAGTGATCG
| | | | | | | | | |
ATTGCGTCGATCG

ATTGCAGT-GATCG
| | | | | | | | | |
ATTGC-GTCGATCG

Which alignment is better?

ATTGCAGTGATCG

ATTGCGTCGATCG

Solution 1:

Solution 2:

ATTGCAGTGATCG
| | | | | | | | | |
ATTGCGTCGATCG

ATTGCAGT-GATCG
| | | | | | | | | |
ATTGC-GTCGATCG

10 matches+ 3 mismatches

12 matches+2 gaps

Scoring Scheme

Match	+1
Mismatch	-1
Indel	-2

Which alignment is better?

ATTGCAGTGATCG

ATTGCGTCGATCG

Solution 1:

Solution 2:

ATTGCAGTGATCG

||||| |||||

ATTGCGTCGATCG

ATTGCAGT-GATCG

||||| || |||||

ATTGC-GTCGATCG

Score=7

Score=8

**Finding the best alignment
for long
sequences is tedious**

**For two sequences of length 300
bases there are 10^{179} different
alignments**



Dynamic programming

Dynamické programování

Needleman-Wunsch (1970)

Smith-Waterman (1981)

- ✦ První krok je triviální a pokrývá částečné řešení
- ✦ Každé další řešení je hodnoceno na základě předcházejících zjištění
- ✦ Zarovnání je tak postupně prodlužováno o další triviální úseky
- ✦ Opakování předchozích kroků vyústí v konečné řešení

Dynamic Programming Algorithm

Seq 1) * A G C
Seq 2) * A A A C

		A	G	C
	0	1	2	3
0				
A 1				
A 2				
A 3				
C 4				

Needelman-Wunsch algorithm (1970)

Dynamic Programming Algorithm

* - - - - A G C
* A A A C

match=1
mismatch=-1
indel=-2

		A	G	C
0	0	-2	-4	-6
A 1	-2			
A 2	-4			
A 3	-6			
C 4	-8			

Dynamic Programming Algorithm

* A G C

* A - - A A C

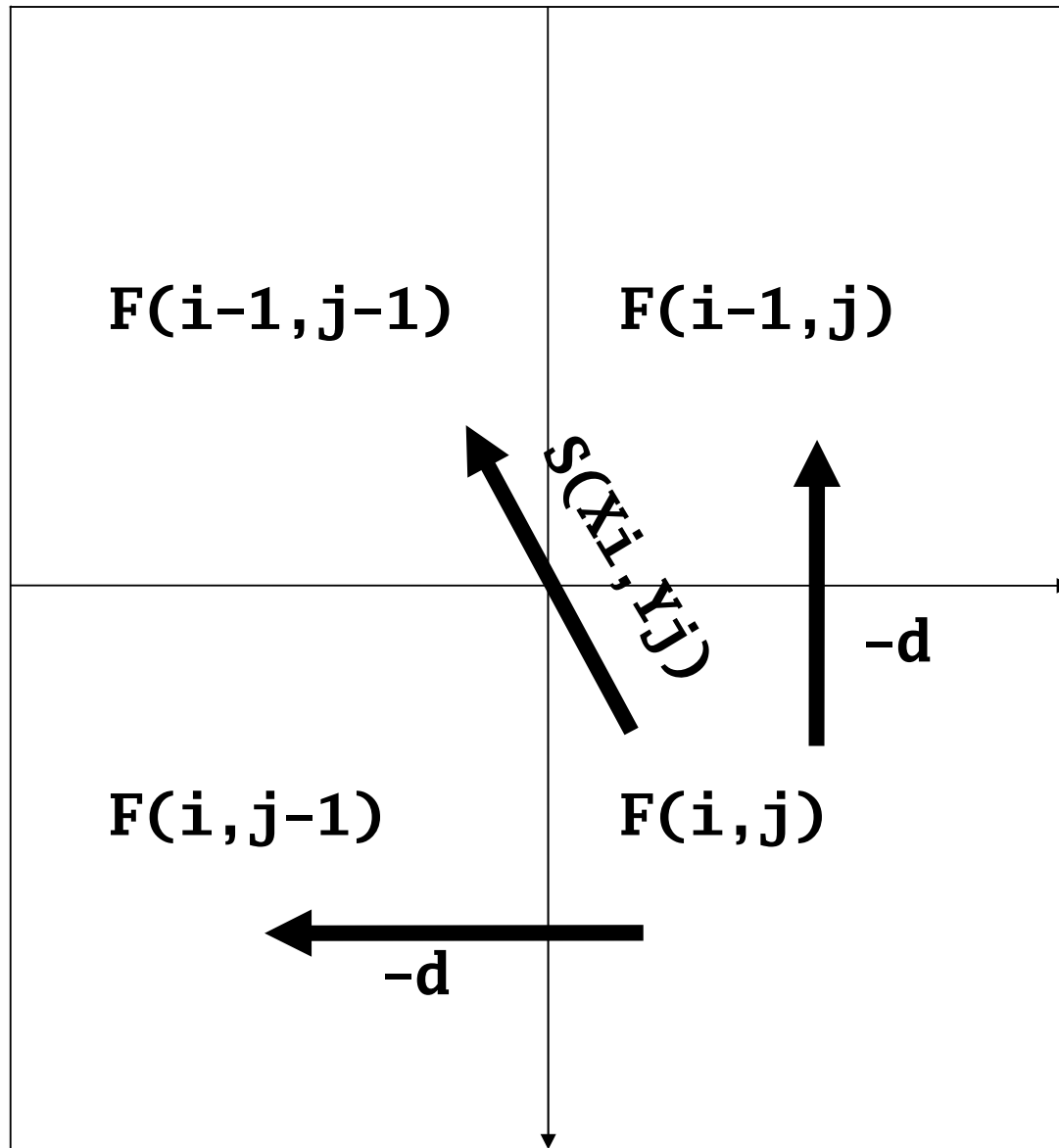
match=1

mismatch=-1

indel=-2

		A	G	C
0	0	-2	-4	-6
A 1	-2	1	-1	-3
A 2	-4			
A 3	-6			
C 4	-8			

Diagram illustrating the dynamic programming algorithm for sequence alignment. The table shows the alignment of sequence A (A G C) against sequence B (A - - A A C). The values in the table represent the score of the alignment at each position. Arrows indicate the path of the alignment: from (0,0) to (1,1), then from (1,1) to (1,2), and finally from (1,2) to (1,3).



Global pairwise alignment

$$F(i,j) = \max \begin{cases} F(i-1, j-1) + s(x_i, y_j) \\ F(i-1, j) - d \\ F(i, j-1) - d \end{cases}$$

Finding the Best Score

		A	G	C
	0	1	2	3
0	0 ← -2 ← -4 ← -6			
A 1	-2 ↑ ↘ 1 ← -1 ← -3			
A 2	-4 ↑ ↘ 1 ↑ ↘ 0 ← -2			
A 3	-6 ↑ ↘ -3 ↑ ↘ -2 ← -1			
C 4	-8 ↑ ↘ -5 ↑ ↘ -4 ↑ ↘ -1			

Tracing the Best Alignment

		A	G	C
0	0 ← -2 ← -4 ← -6			
A 1	-2 ↑ 1 ← -1 ← -3			
A 2	-4 ↑ -1 ↑ 0 ← -2			
A 3	-6 ↑ -3 ↑ -2 -1			
C 4	-8 ↑ -5 ↑ -4 -1			

The table shows a dynamic programming matrix for sequence alignment. Red arrows trace the path from the bottom-right cell (row C 4, column C) to the top-left cell (row 0, column 0). The value -1 in the bottom-right cell is circled in red.

A	G	-	C
A	A	A	C

Tracing the Best Alignment

		A	G	C
	0	1	2	3
0	0 ← -2 ← -4 ← -6			
A 1	-2 ↑ 1 ← -1 ← -3			
A 2	-4 ↑ -1 ↑ 0 ← -2			
A 3	-6 ↑ -3 ↑ -2 -1			
C 4	-8 ↑ -5 ↑ -4 -1			

A	-	G	C
A	A	A	C

Tracing the Best Alignment

		A	G	C
	0	1	2	3
0	0 ← -2 ← -4 ← -6			
A 1	-2 ↑ -1 ← -3	1 ← -1 ← -3		
A 2	-4 ↑ -1 ← -2	-1 ↑ 0 ← -2		
A 3	-6 ↑ -3 ← -1	-3 ↑ -2 ← -1		
C 4	-8 ↑ -5 ← -4 ← -1	-5 ↑ -4 ← -1		

- A G C
A A A C

Local Alignment Example

		A	T	C	T	A	A
	0	1	2	3	4	5	6
ATCTAA	0						
T	1						
A	2						
A	3						
T	4						
A	5						

Smith-Waterman algorithm, 1981

Local Alignment

$$F(i,j) = \max \begin{cases} F(i-1, j-1) + s(x_i, y_j) \\ F(i-1, j) - d \\ F(i, j-1) - d \\ 0 \end{cases}$$

Local Alignment Example

TCATAA
TAATA

		T	A	C	T	A	A
	0	1	2	3	4	5	6
0	0	0	0	0	0	0	0
T 1	0	1	0	0	1	0	0
A 2	0	0	2	0	0	2	1
A 3	0	0	1	1	0	1	3
T 4	0	0	0	0	2	0	1
A 5	0	0	1	0	0	3	1

Local Alignment Example



		T	A	C	T	A	A
	0	1	2	3	4	5	6
0	0	0	0	0	0	0	0
T 1	0	1	0	0	1	0	0
A 2	0	0	2	0	0	2	1
A 3	0	0	1	1	0	1	3
T 4	0	0	0	0	2	0	1
A 5	0	0	1	0	0	3	1

Local Alignment Example

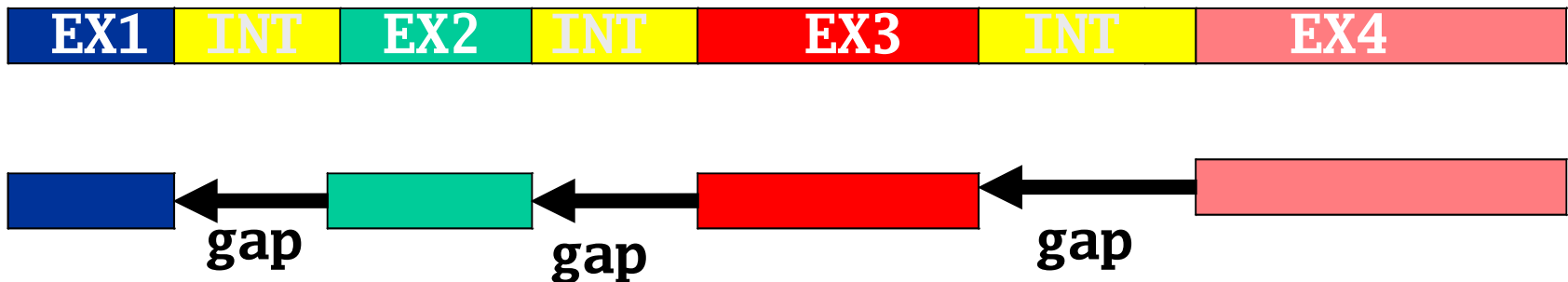
TACTAA
TAATA

		T	A	C	T	A	A
	0	1	2	3	4	5	6
0	0	0	0	0	0	0	0
T 1	0	1	0	0	1	0	0
A 2	0	0	2	0	0	2	1
A 3	0	0	1	1	0	1	3
T 4	0	0	0	0	2	0	1
A 5	0	0	1	0	0	3	1

Examples : Genomic DNA versus mRNA



Alignment



Gap Penalties

AAC-AATTAAG-ACTAC-GTTCATGAC

A-CGA-TTA-GCAC-ACTG-T-A-GA-

AACAATTAAGACTACGTTCATGAC---

AACAATT-----GTTCATGACGCA

Scoring Gaps

I AAC-AATTAAG-ACTAC-GTTCATGAC -6

A-CGA-TTA-GCAC-ACTG-T-A-GA-

II AACCAATTAAGACTACGTTCATGAC--- 12

AACAATT-----GTTCATGACGCA

Scoring parameters

match: +1; Gap_open: -2

Scoring Insertions/Deletions

AAC-AATTAAG-ACTAC-GTTCATGAC -6

I

A-CGA-TTA-GCAC-ACTG-T-A-GA-

AACAATTAAGACTACGTTCATGAC--- -6

II

AACAATT-----GTTCATGACGCA

**Scoring parameters
match:+1;indel:-2**

Considering Gap Opening and Gap Extension

I AAC-AATTAAG-ACTAC-GTTCATGAC -17

A-CGA-TTA-GCAC-ACTG-T-A-GA-

II AACCAATTAAGACTACGTTCATGAC---

AACAATT-----GTTCATGACGCA¹

Scoring parameters

match: +1; Gap_open: -2; Gap_exten: -1

