

# 3D Structural Alignment and Tunnel Computation

David Sehnal

# Contents

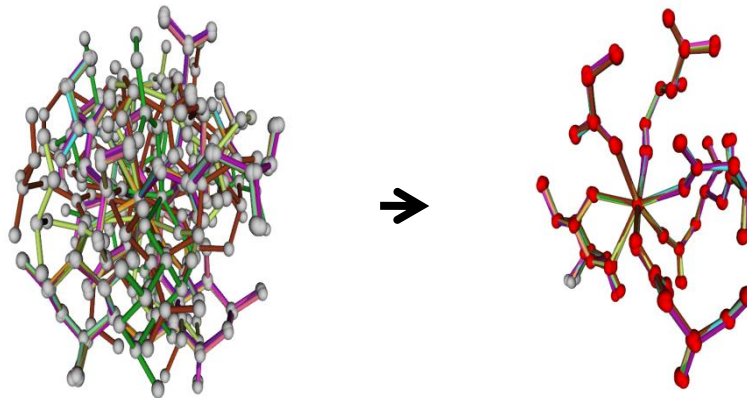
- Two Worlds
  - 3D Structural Alignment (on the small scale)
  - Tunnel Computation
- The Worlds Collide?

# 3D Alignment

- **Large scale** (proteins)
  - Global (classification, evolutionary links)
  - Local (compare smaller sub-structures)
- **Small scale**

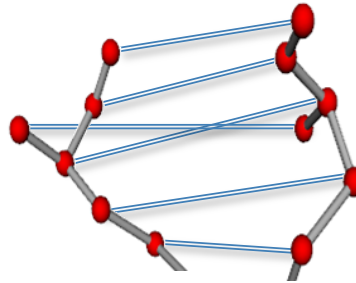
Global FTFTALILLAVAV  
F--TAL-LLA-AV

Local FTFTALILL-AVAV  
--FTAL-LLAAV--



# 3D Alignment

- What we need?
  - Know which atoms/amino acids/... belong together



- Find a transformation that fits the corresponding pairs

$$\text{RMSD}(A, B) = \sqrt{\frac{1}{n} \sum_{i=1}^n \|A_i - B_i\|^2}$$

# The Transformation Part

- Horn K.P.: Closed-form solution of absolute orientation using unit quaternions, Journal of the Optical Society of America, 1986.
- Karney C.F.F.: *Quaternions in molecular modeling*. Journal of Molecular Graphics and Modelling, 2007.

$$\text{RMSD}(T(A), B) = \sqrt{\frac{1}{n} \sum_{i=1}^n \|R(A_i) + t - B_i\|^2}$$

Rotation component
Translation component

Stays the same Maximize this

$$\sum_{i=1}^n \|R(A_i) - B_i\|^2 = \sum_{i=1}^n \|R(A_i)\|^2 + \sum_{i=1}^n \|B_i\|^2 - 2 \sum_{i=1}^n R(A_i) \cdot B_i$$

$$q^T \left( \sum_{i=1}^n \hat{A}_i^T B_i \right) q$$

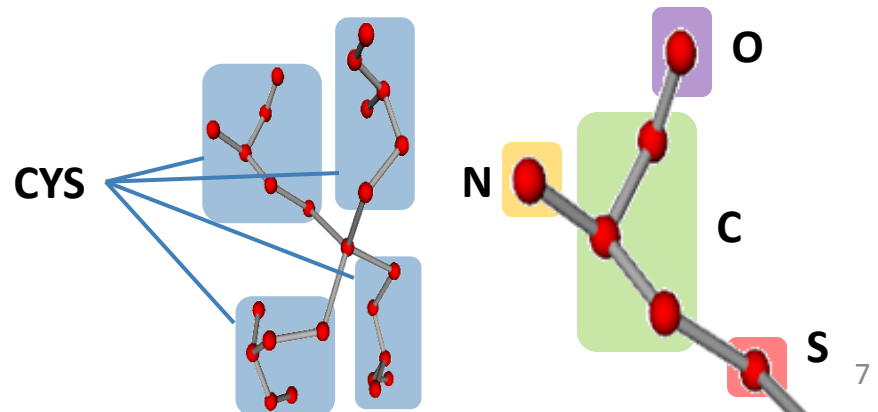
$R(x) = qxq^*$

# Pairing the Elements

- Large Scale
  - Usually work on the secondary structure (amino acids)
  - Combinatorial extensions, subgraph isomorphism, dynamic programming, ...
- **Small Scale**
  - Usually work on 3D structure
  - Exhaustive search, pairing based on distance, subgraph isomorphism, ...

# (My) Simple Algorithm

1. For each bijection  $f: \text{Atoms}(A) \rightarrow \text{Atoms}(B)$ 
    - Align sequences  $f(A)$  and  $B$  using the quaternion method
  2. Return the alignment that yields the lowest RMSD
- Improvement for small protein substructures:
    - Grouping of atoms

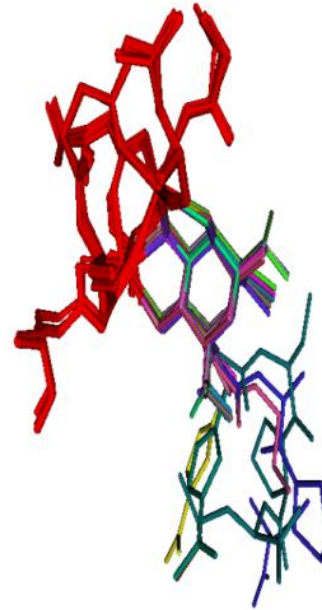


# Applications



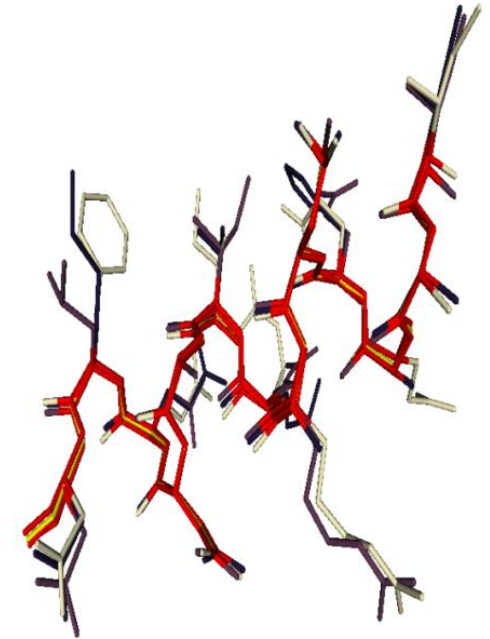
## Reverse Proteins

Classification



## Lectines

66 structures  
56 atom on each



## Apoptosis

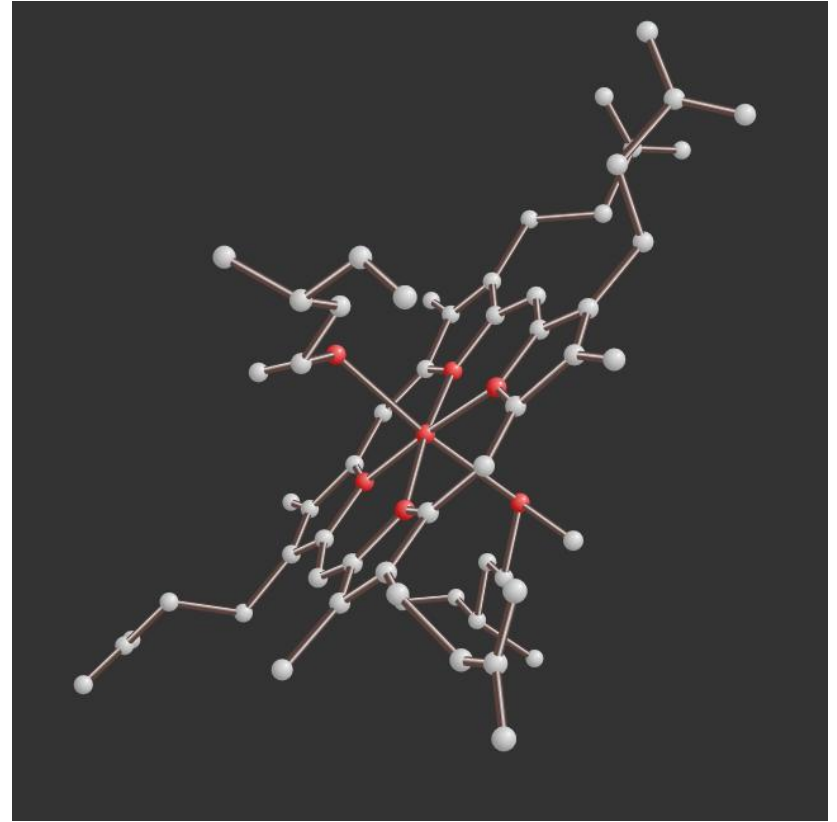
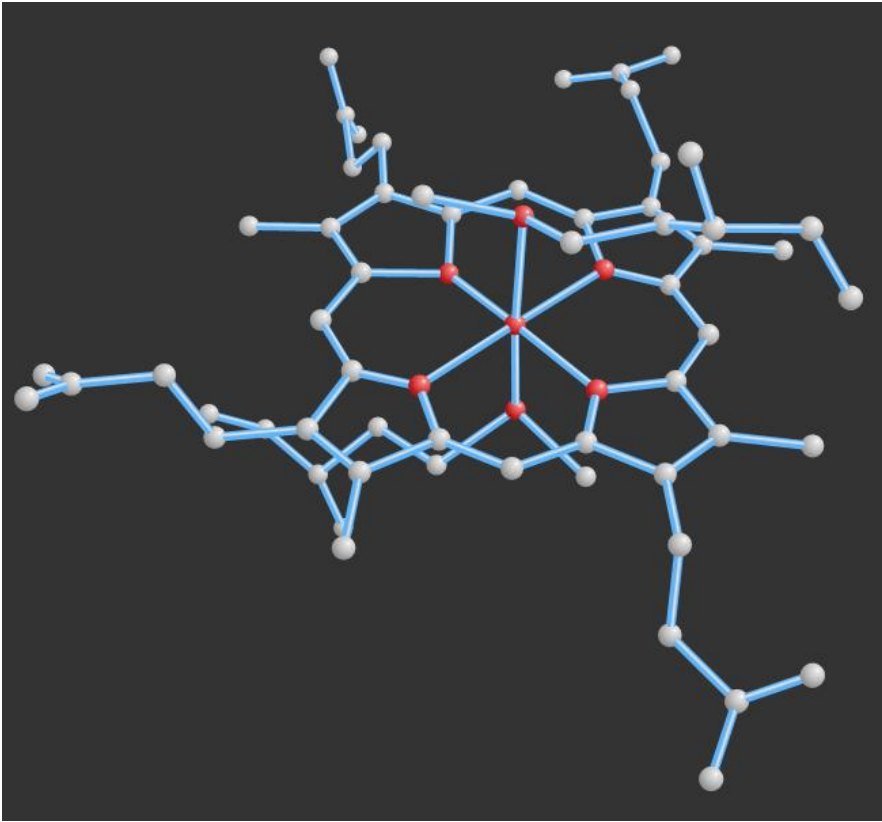
Analysis of the  
model structure



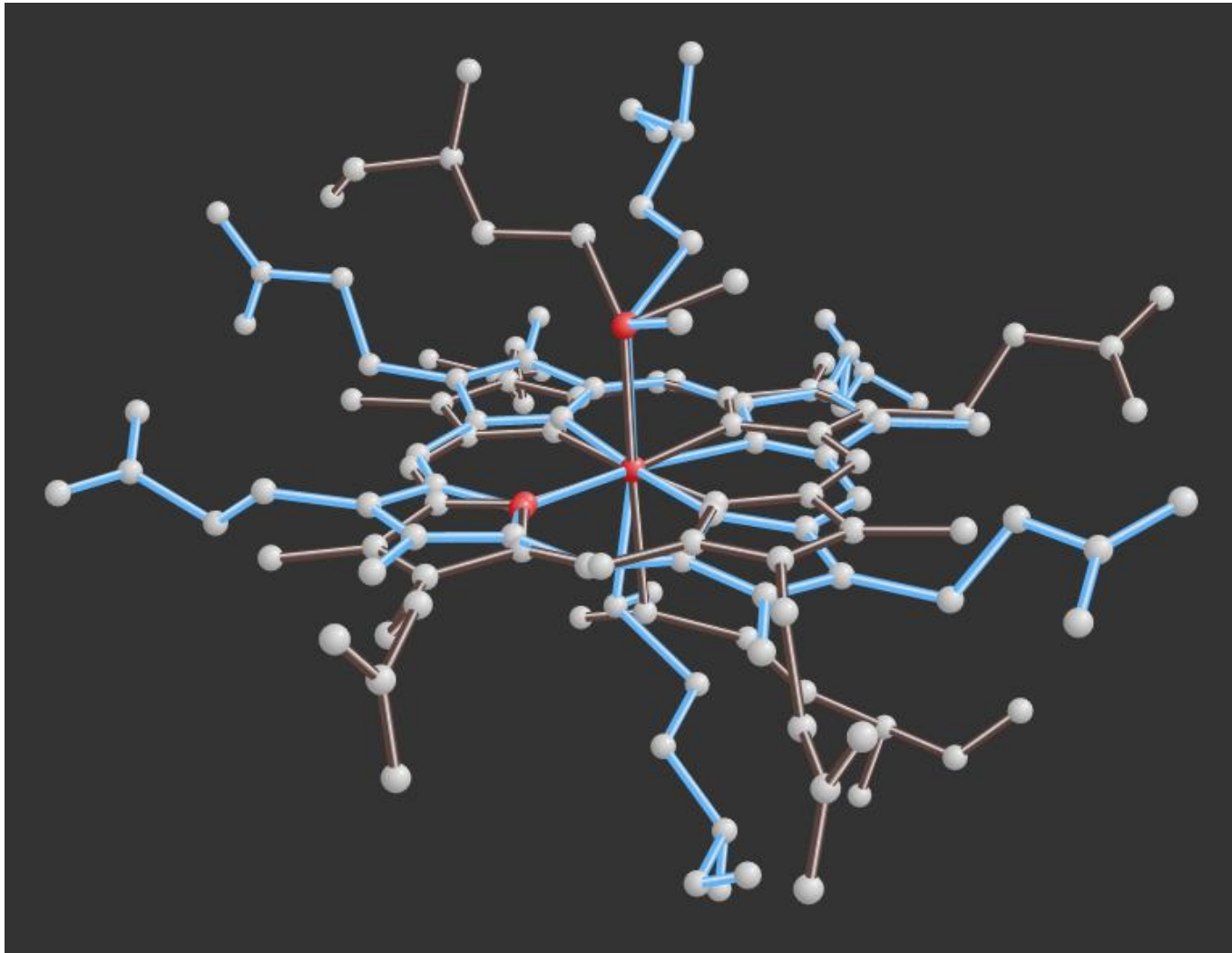
# (Sort of Mine) Better Approach

- Hoppe A., Frommel C.: *NeedleHaystack: a program for the rapid recognition of local structures in large sets of atomic coordinates*, Journal of applied crystallography, 2003.
- Find sets of 3 suitable pivots in each structure
- For each pair of pivot sets
  - Align the structures using these pivots (quaternions)
    1. Match atoms based on distance (with a threshold)
    2. Align again (quaternions)
    3. (Goto 1 until a stable configuration is reached)
  - Score =  $f(A, B, \text{RMSD}_M)$  M is the set of matched atoms, N is the total number of atoms, and  $\text{RMSD}_M$  is the RMSD of matched atoms.
- Return the alignment with the best score

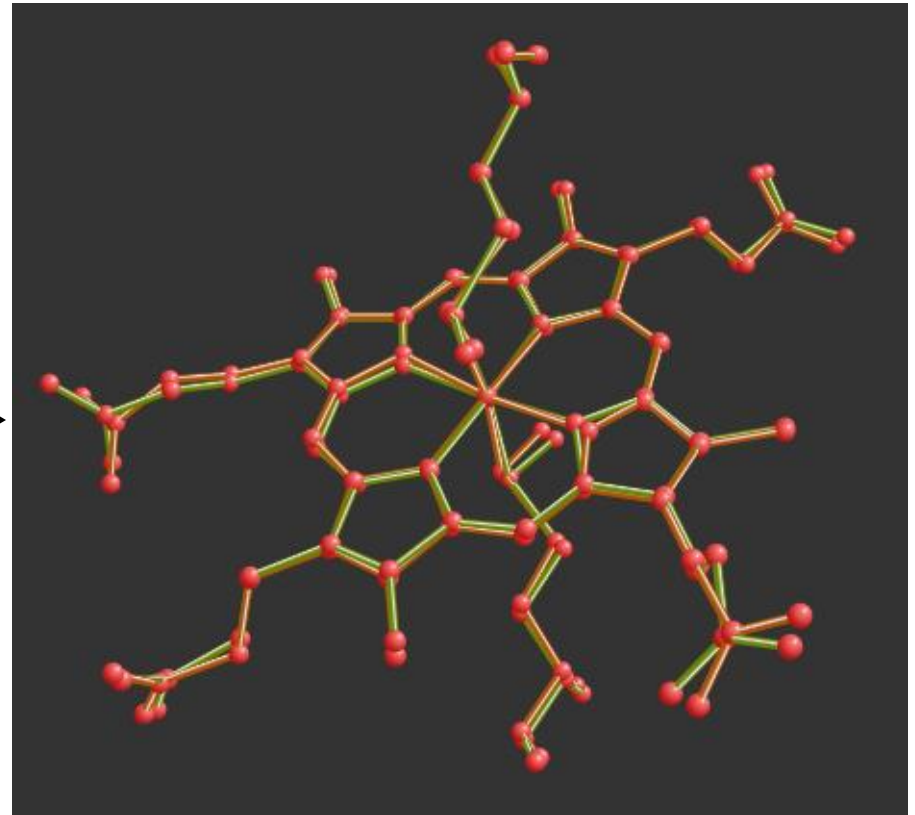
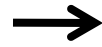
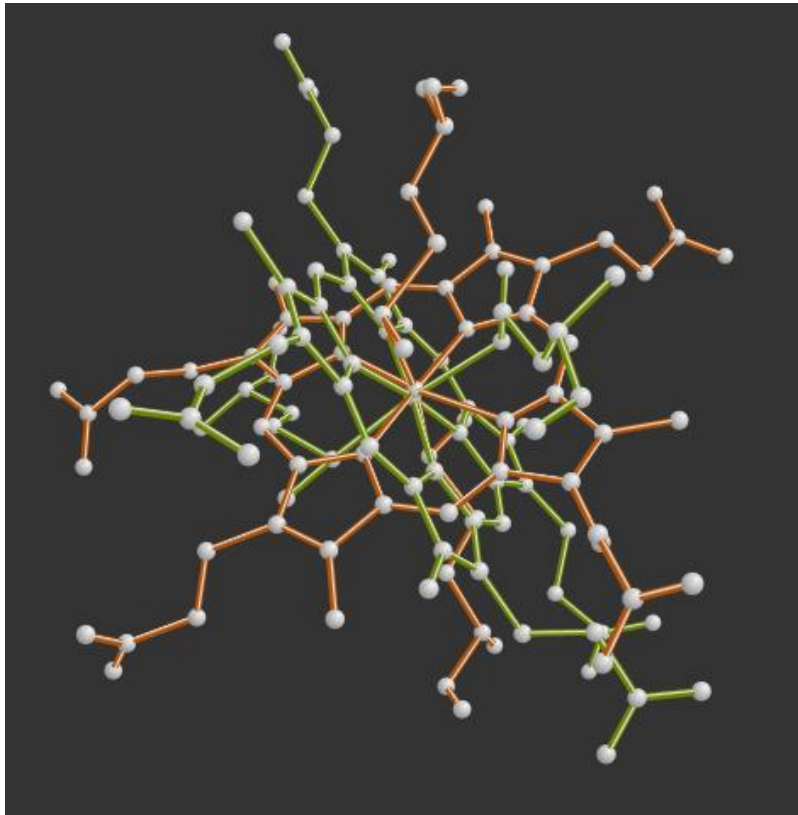
# Example ( $\text{FeN}_4\text{S}_2$ )



# Example ( $\text{FeN}_4\text{S}_2$ )



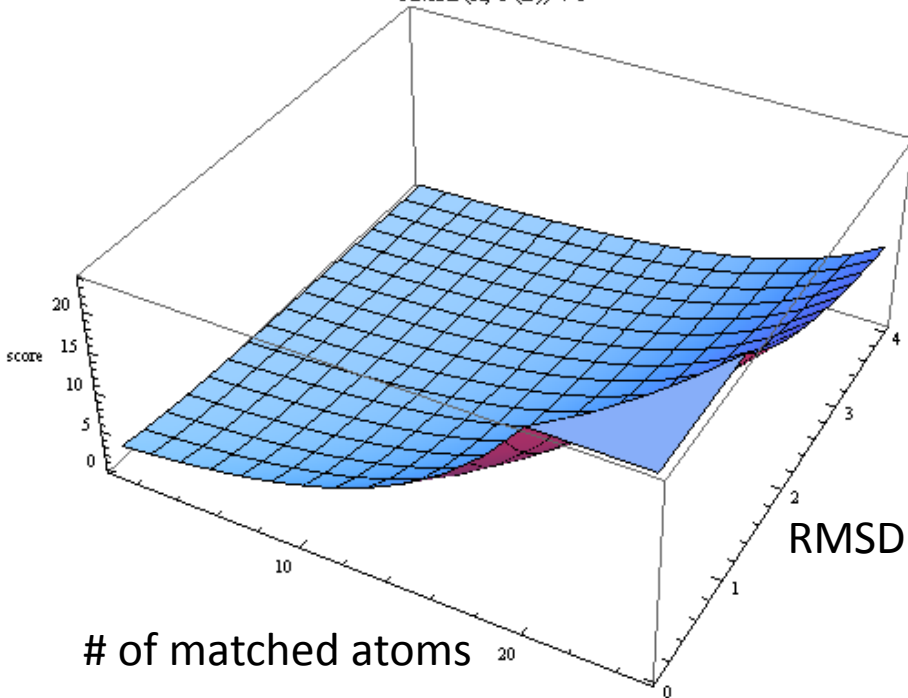
# Example ( $\text{FeN}_4\text{S}_2$ )



# Scoring

## SiteBinder

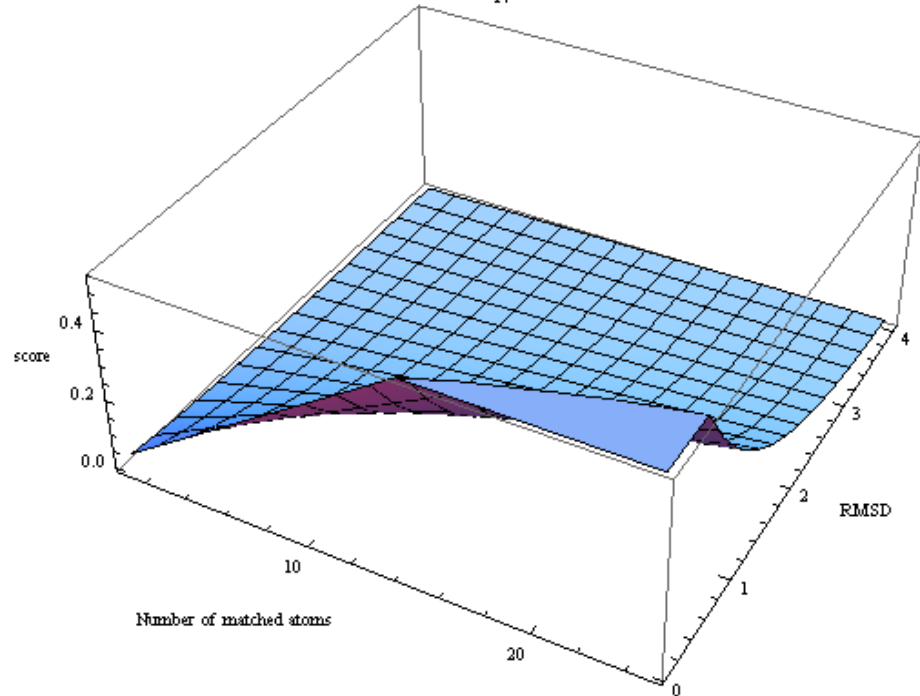
$$\frac{e^{\left(\frac{|M|}{N} + 1\right)^2}}{\text{RMSD}(A, T(B)) + 1}$$



$$\frac{e^{(1+|M|/N)^2}}{1 + \text{RMSD}_M(A, B)}$$

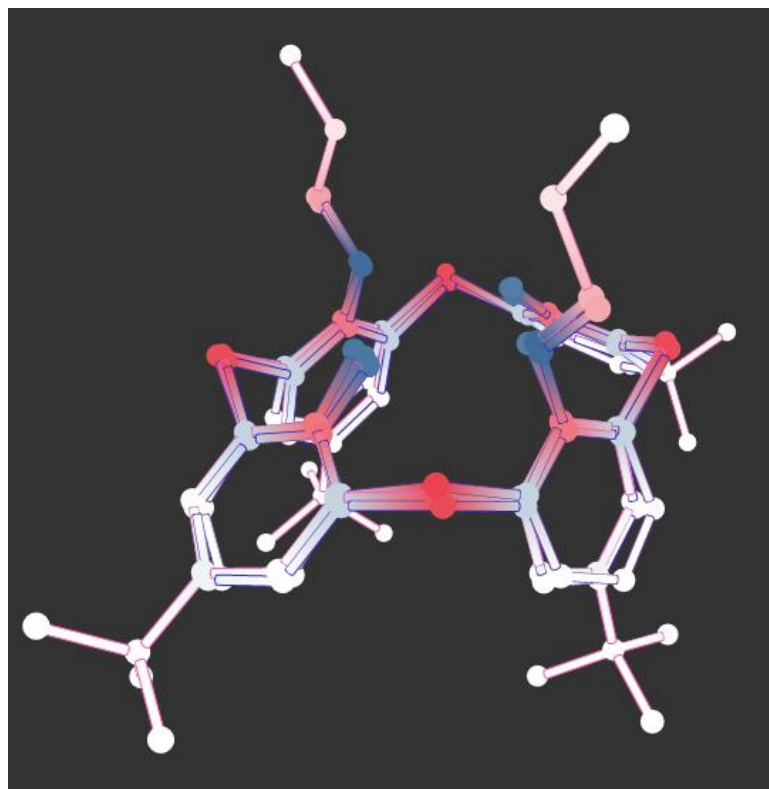
## Needle-Haystack

$$\frac{|M| e^{-\text{RMSD}(A, T(B))}}{N}$$



M is the set of matched atoms, N is the total number of atoms, and  $\text{RMSD}_M$  is the RMSD of matched atoms.

# Different Pivot Criteria (Charges)



$$\text{score}^*(M, N, A, B) = R_M^2 \cdot \text{score}'(M, N, A, B) = R_M^2 \cdot \frac{e^{(1+|M|/N)^2}}{1 + \text{RMSD}_M(A, B)}$$

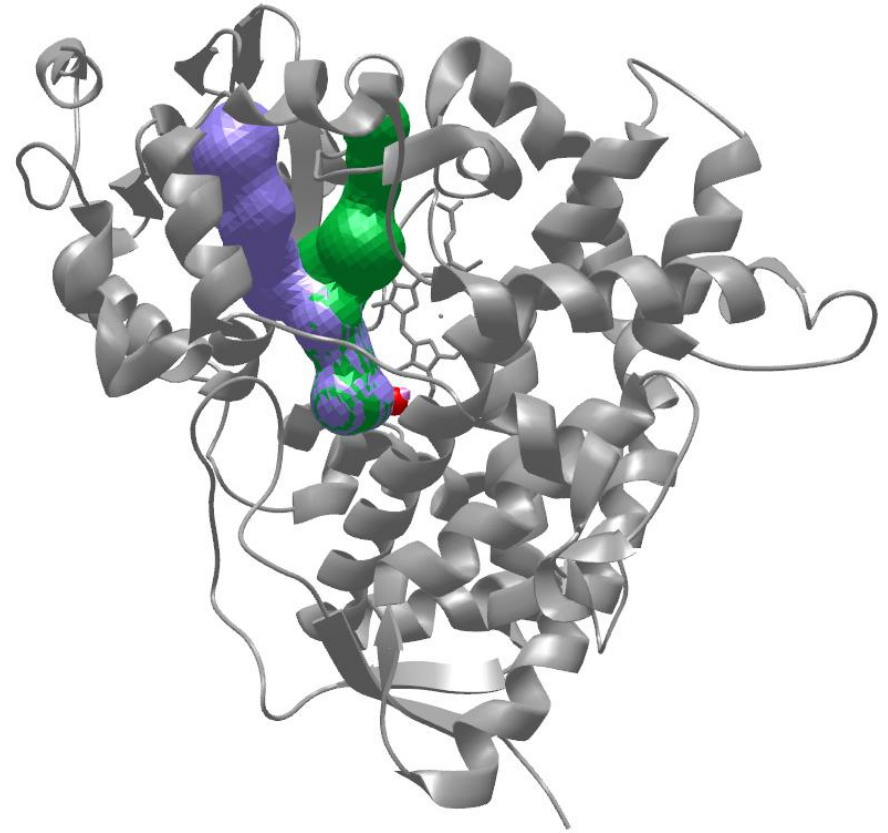
$R_M^2$  says how well the matched charges correlate.

# 3D Alignment Summary

**3D structural alignment using pivot elements and improved scoring function to identify the correct result.**

# Tunnels

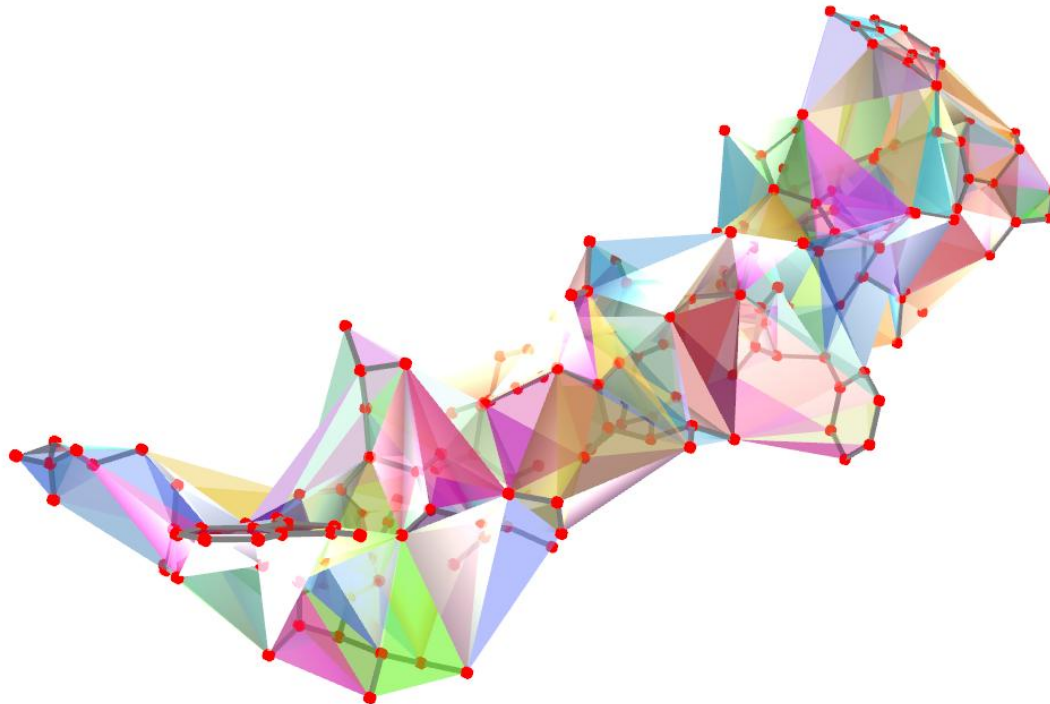
- **MOLE**, Caver, Hollow, MolAxis
- Static analysis vs. dynamics
- Grid/Triangulation Based





# Protein (Delaunay) Triangulation

- Edelsbrunner H., Liang J.: Anatomy of protein pockets and cavities: measurement of binding site geometry and implications for ligand design, Protein Science, 1998.





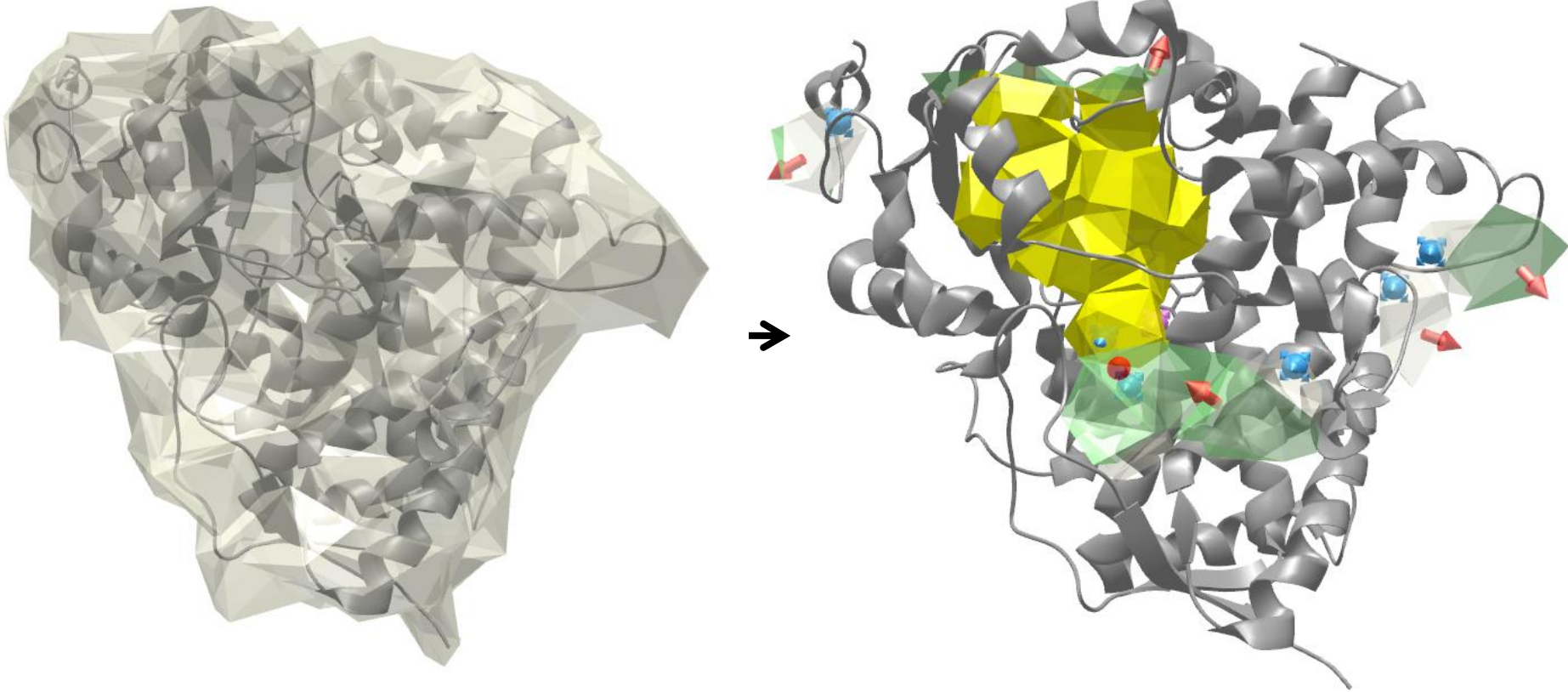
# Tunnel Finding Algorithm

1. Triangulate the molecule (Voronoi diagram)
2. Specify starting point (from a database/user)
3. Do depth-first ( $A^*$ ) search
  - Each time a boundary tetrahedron is visited, report a tunnel
  - Edge weight  $\sim$  amount off space around the edge
4. Post-process the tunnels (remove duplicate tunnels, ...)

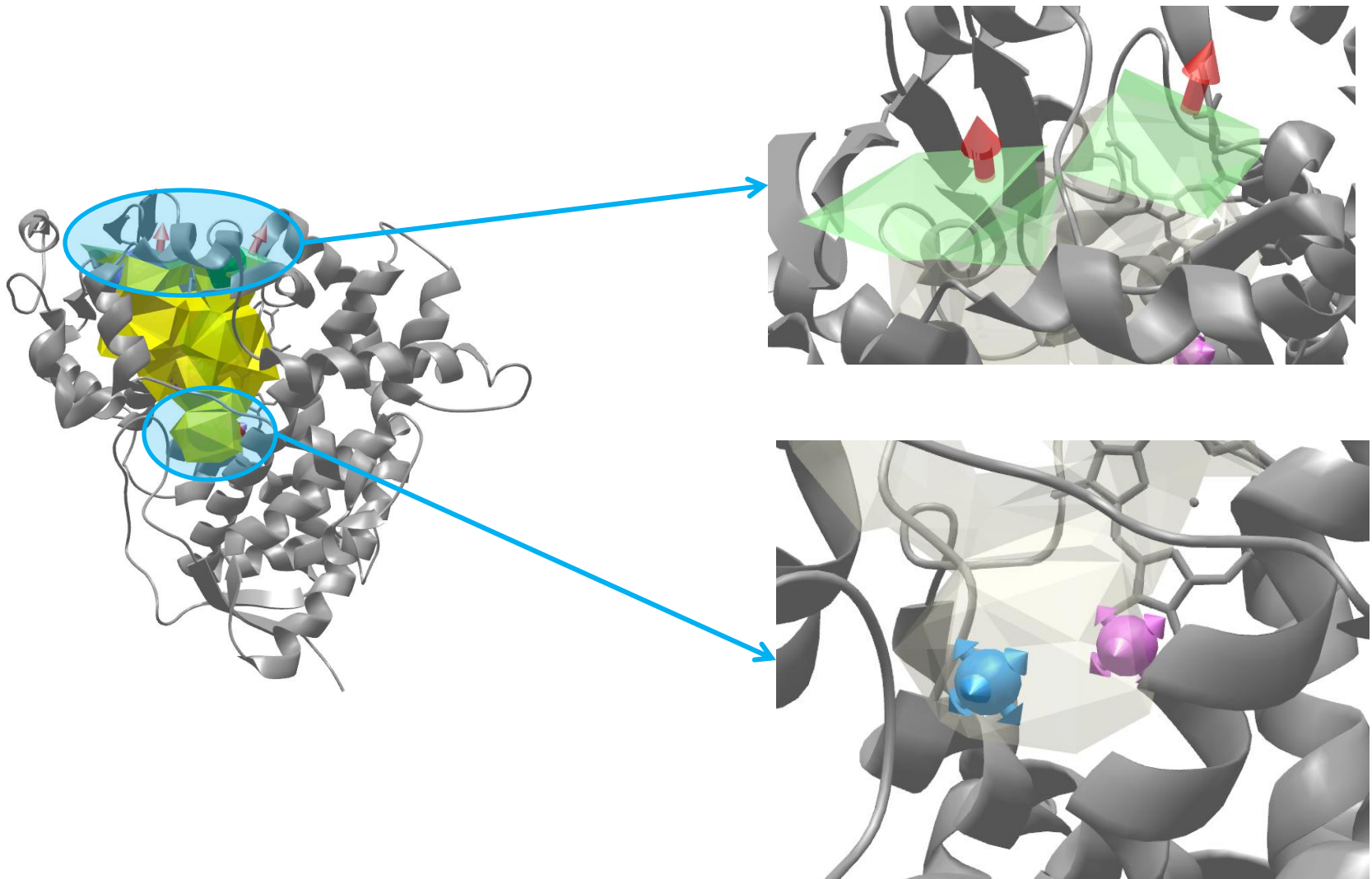
# (My) Modified Approach

1. Triangulate the molecule (Voronoi diagram)
2. Identify cavities (“empty space”)
  - Remove tetrahedrons that are too big or too small
  - Find connected components in the new graph
3. Identify start points within cavities
  - Deep point(s) in each cavity
  - Database/user
4. Identify end points within cavities
  - Connected components of the “boundary”
5. For each pair of start and end points in the same cavity use Dijkstra’s algorithm to find the tunnel
  1. Edge weight  $\sim$  amount of space around the edge
6. Post-process the tunnels (remove duplicate tunnels, ...)

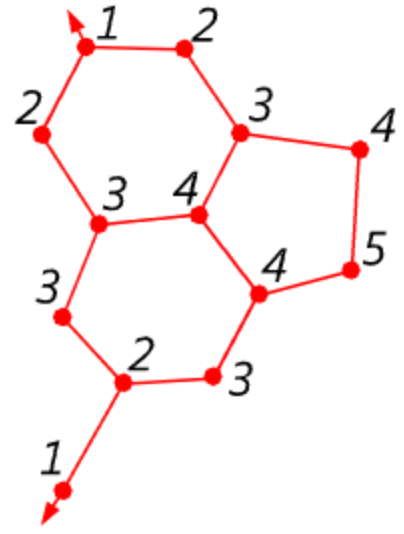
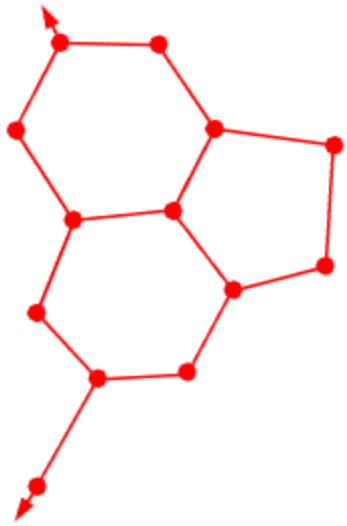
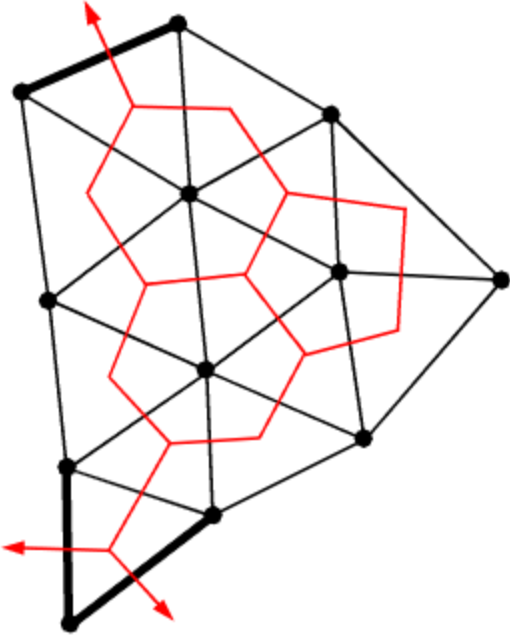
# The Cavities



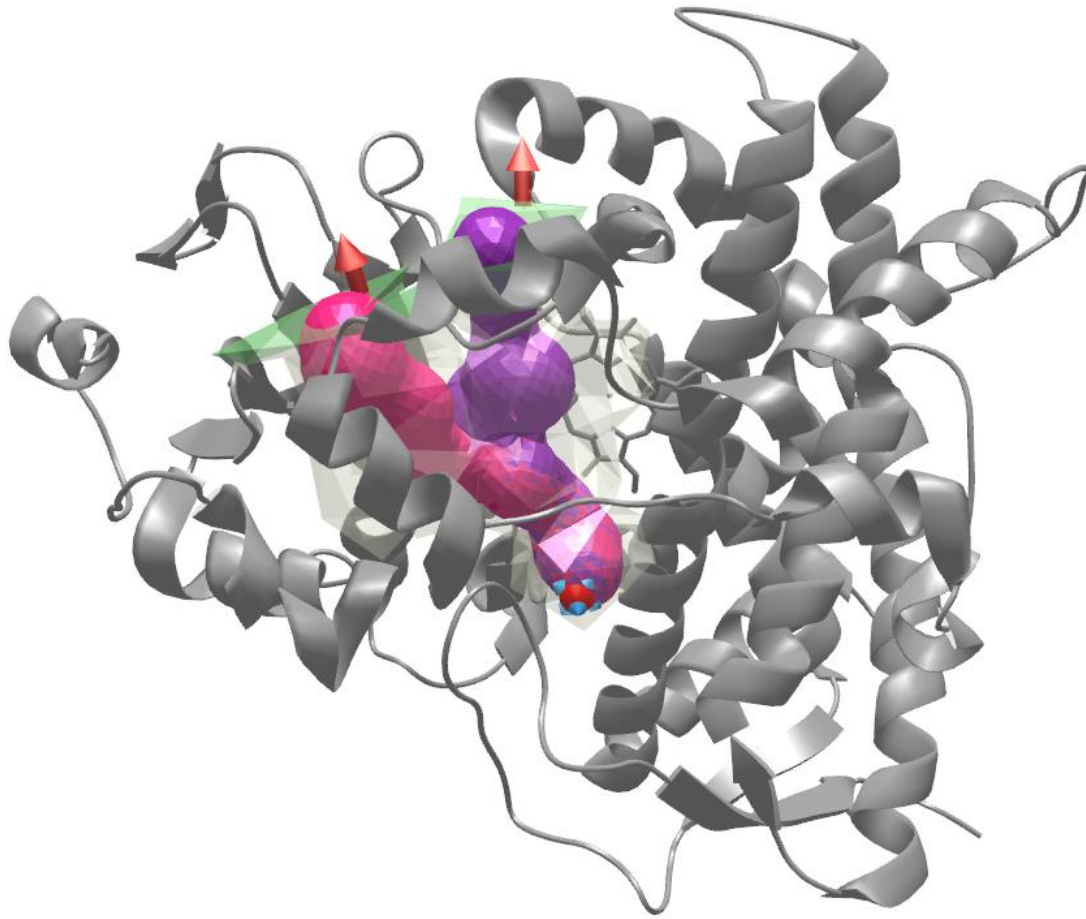
# Start and End Points



# Depth Computation



# The Tunnels

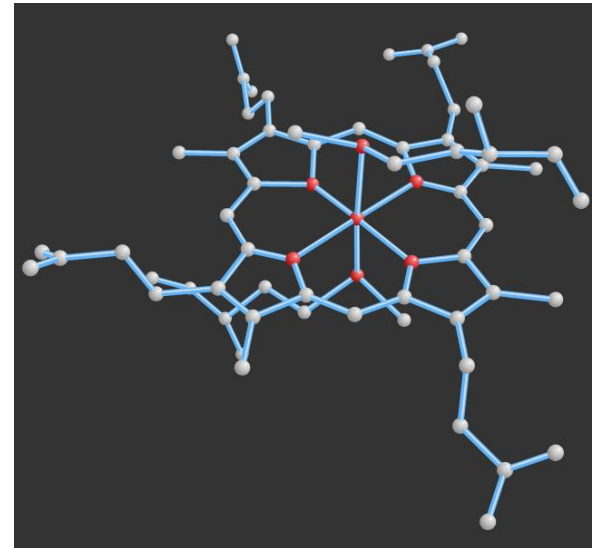
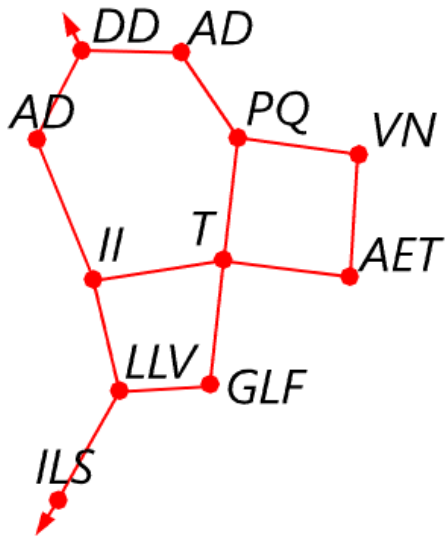
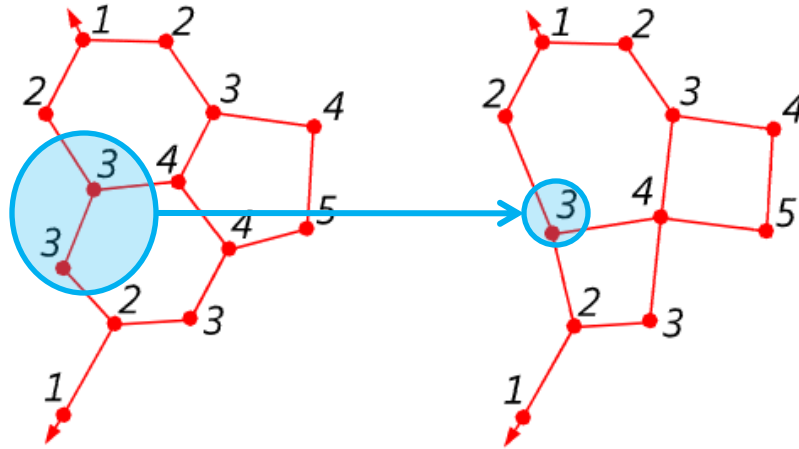




# Tunnel Computation Summary

**Triangulate the protein, split it into smaller graphs and use Dijkstra's algorithm to find interesting paths (= tunnels) in them.**

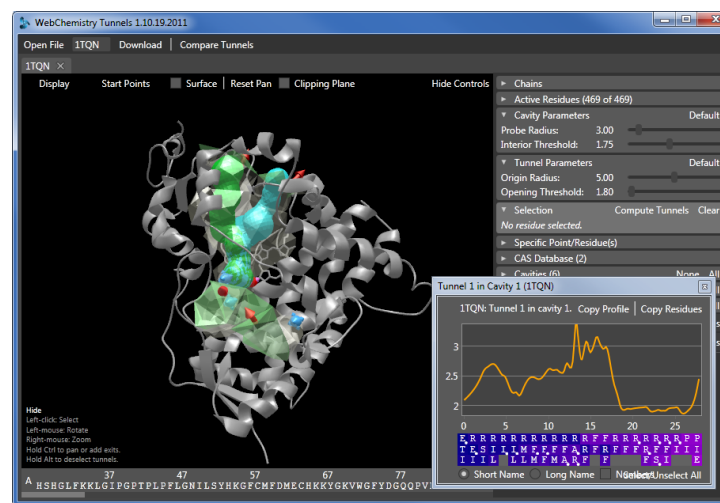
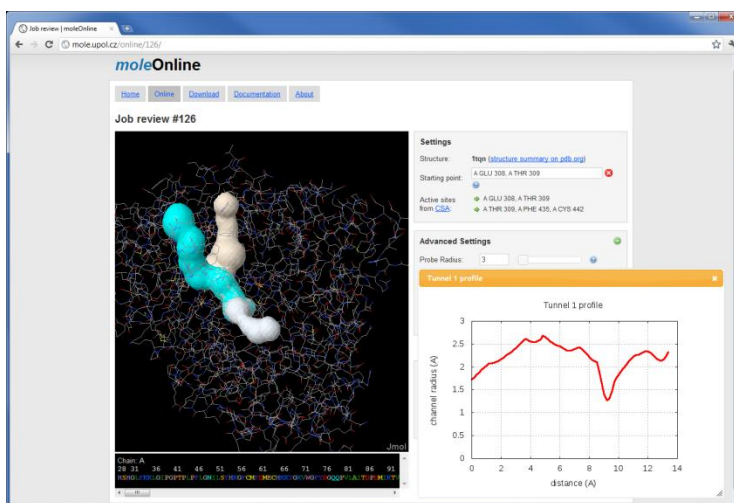
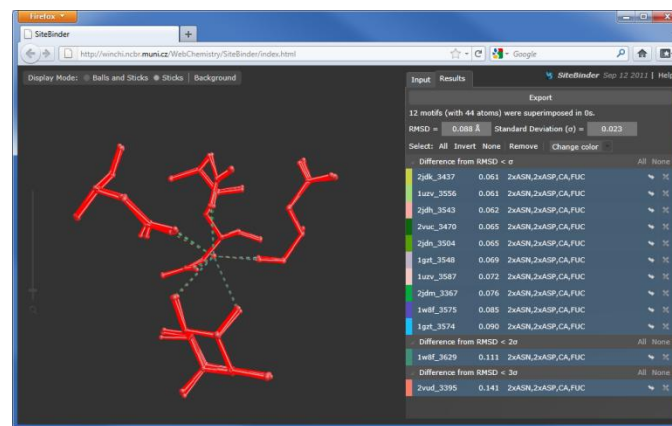
# Aligning the Tunnels/Cavities



Reduced Cavity Graph

# Summary

- 3D Alignment Algorithm
- Tunnel Finding Algorithm





**Thank you for your attention.**