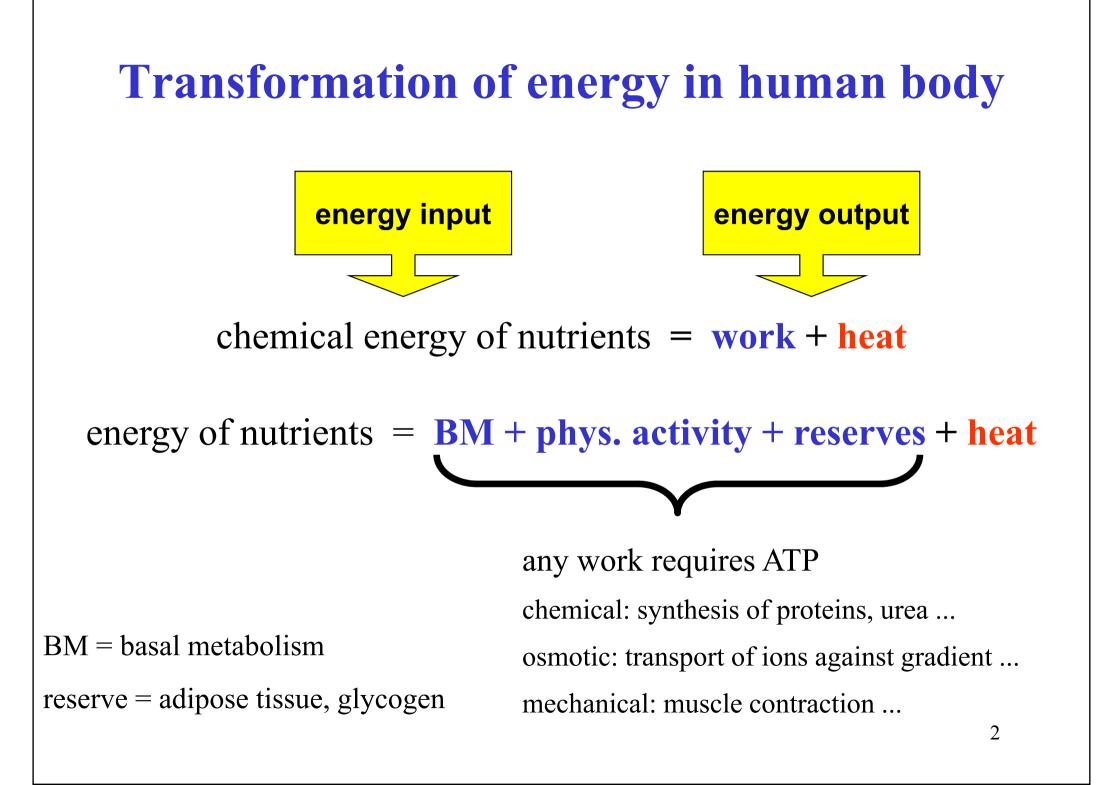
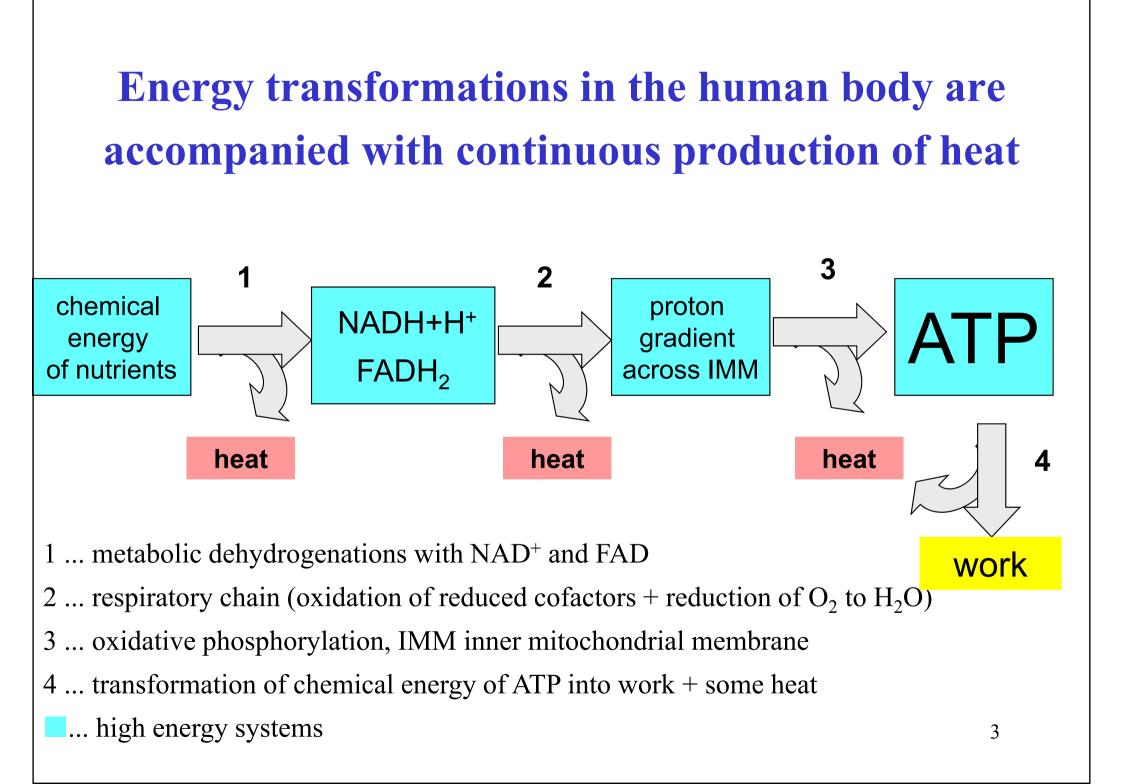
Respiratory chain ~ Reactive oxygen species

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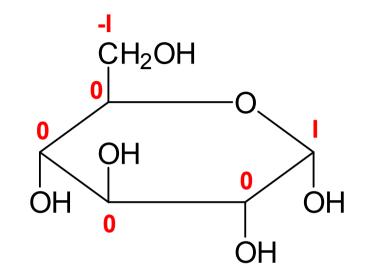




Energetic data about nutrients

Nutrient	Energy (kJ/g)	Thermogenesis	Energy supply/day
Lipids	38	4 %	30 % SAFA 5 %, MUFA 20 %, PUFA 5 %
Saccharides	17	6 %	60 %
Proteins	17	30 %	10 %

Oxidation numbers of carbon and the content of hydrogen in nutrients



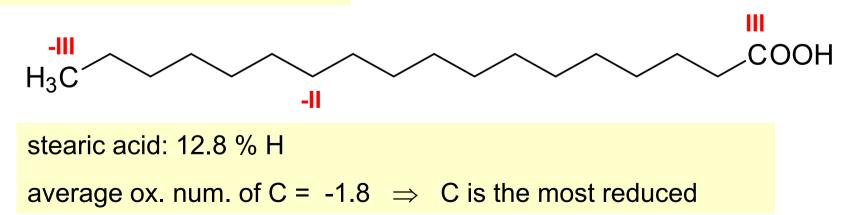
-III 0 III H₃C--CH--COOH

alanine: 7.9 % H

average ox. num. of C = 0.0

glucose: 6.7 % H

average ox. num. of C = 0.0



Two ways of ATP formation in body

1. Oxidative phosphorylation in the presence of O₂ (~ 95 % ATP)

ADP + P_i + energy of H⁺gradient \rightarrow ATP

2. Substrate-level phosphorylation (~ 5 % ATP)

ADP + macroergic phosphate- $P \rightarrow ATP$ + second product

higher energy content than ATP

Compare: Phosphorylation

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substrate-OH + ATP \rightarrow substrate-O-P + ADP
```

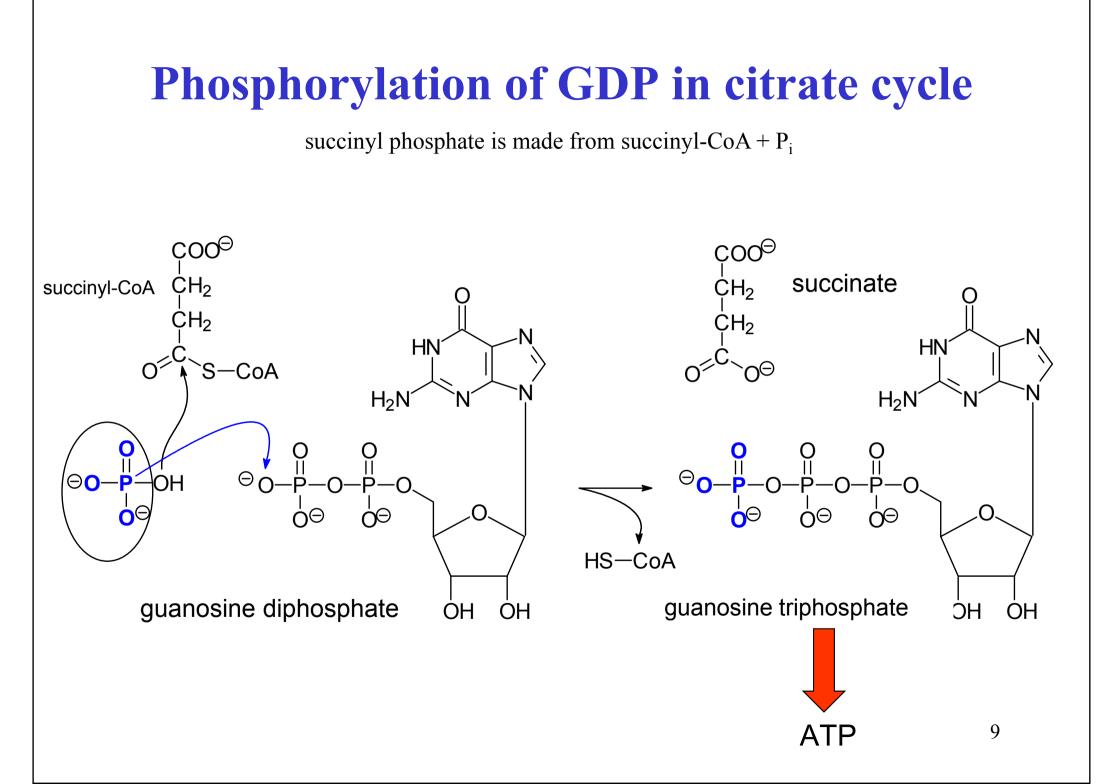
(e.g. phosphorylation of glucose, proteins, etc., catalyze kinases)

Distinguish

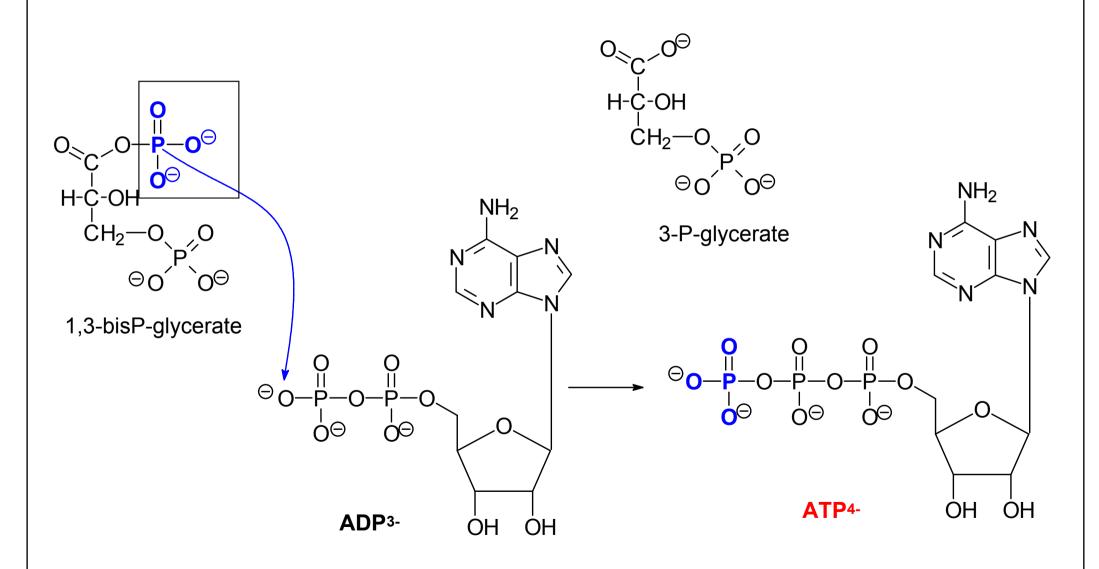
Process	ATP is
Oxidative phosphorylation	produced
Substrate-level phosphorylation	produced
Phosphorylation of a substrate	consumed

Substrate level phosphorylation

- phosphorylation of ADP (GDP) is performed by the high-energy intermediates
- succinyl-CoA (CAC)
- 1,3-bisphosphoglycerate (glycolysis)
- phosphoenolpyruvate (glycolysis)

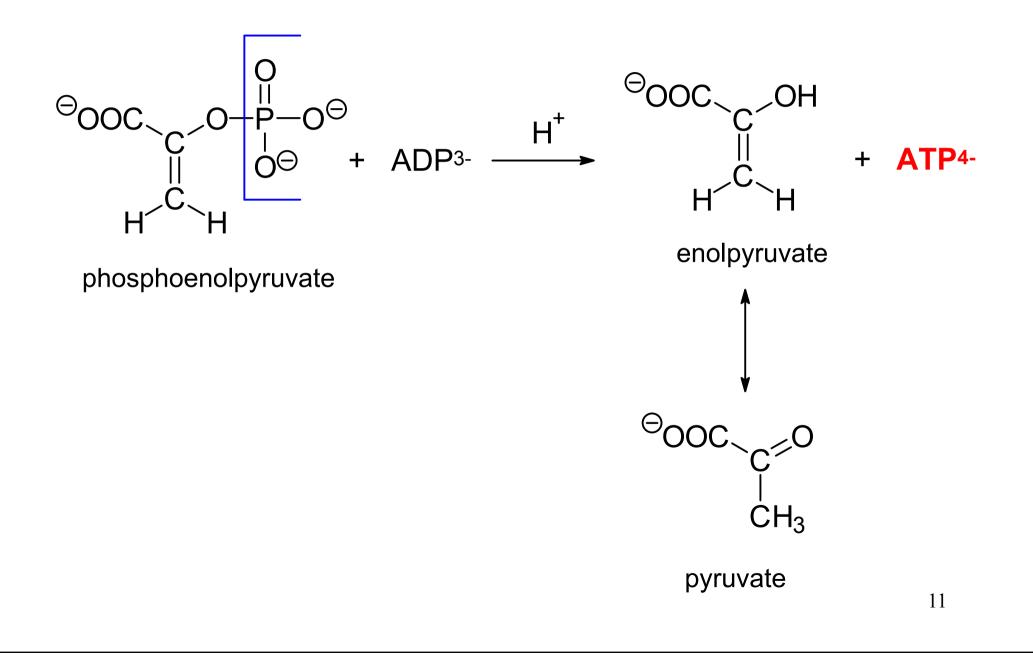


Phosphorylation of ADP by 1,3-bisphosphoglycerate

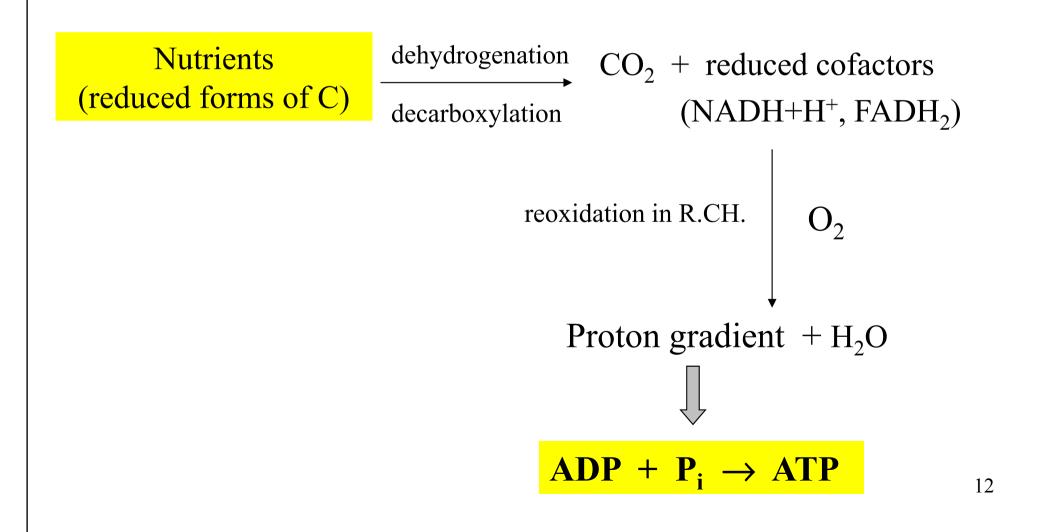


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Phosphorylation of ADP by phosphoenolpyruvate



Aerobic phosphorylation follows the reoxidation of reduced cofactors in R.CH.



NADH formation in mitochondrial matrix

(substrates of the important reactions)

• Citrate cycle

isocitrate 2-oxoglutarate malate

β-oxidation of FA
 β-hydroxyacyl-CoA

Oxidative decarboxylation

pyruvate

2-oxoglutarate

2-oxo acids from Val, Leu, Ile

- Dehydrogenation of KB
 β-hydroxybutyrate
- Oxidative deamination glutamate

NADH formation in cytoplasm

• Glycolysis

(dehydrogenation of glyceraldehyde-3-P)

Gluconeogenesis

(dehydrogenation of lactate to pyruvate)

• **Dehydrogenation of ethanol** (to acetaldehyde)

FADH₂ formation in matrix of mitochondria

• β-Oxidation of FA

(dehydrogenation of saturated acyl-CoA)

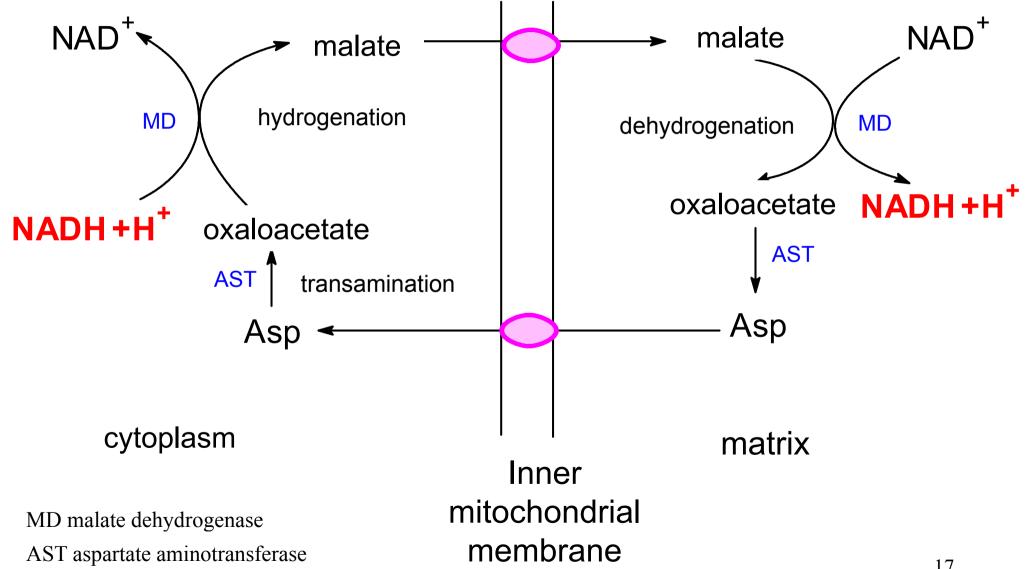
• Citrate cycle

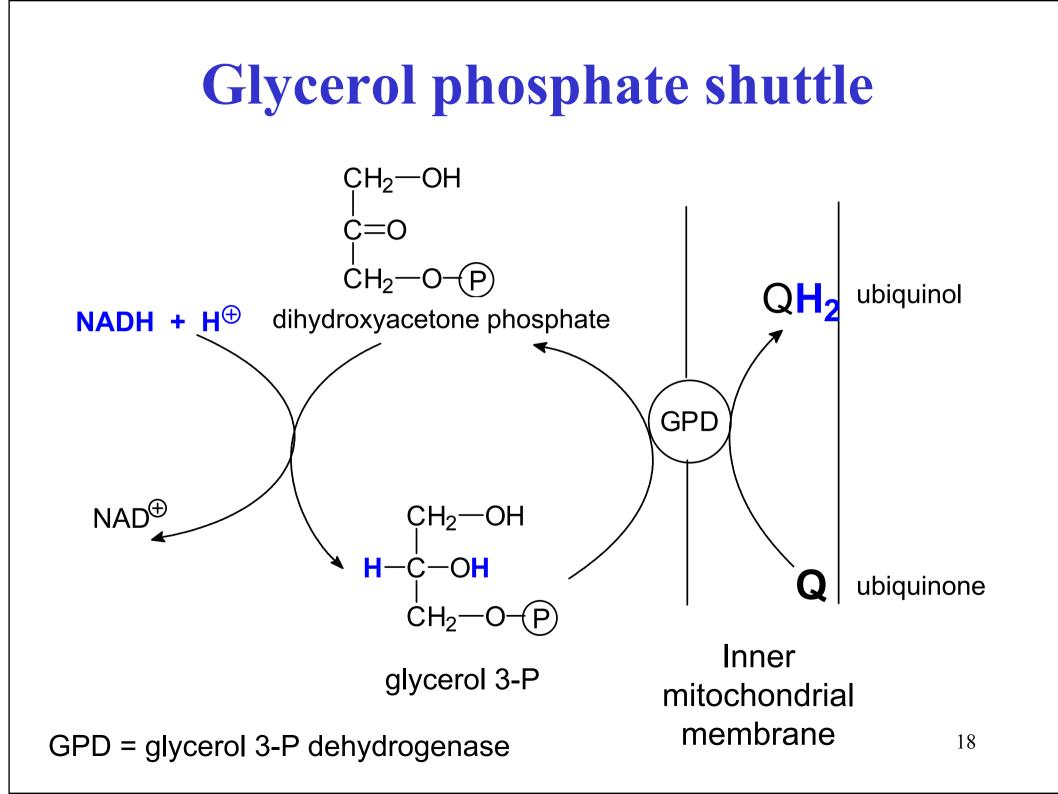
(dehydrogenation of succinate)

Transport of NADH from cytoplasm to matrix

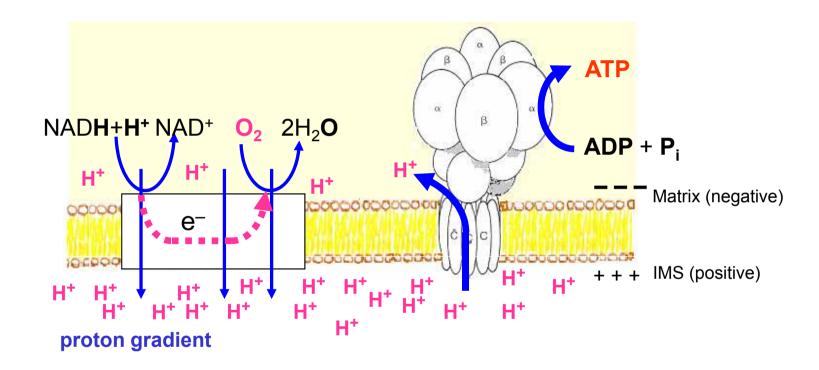
- NADH produced in cytoplasm is transported into matrix to be reoxidized in R.CH.
- inner mitochondrial membrane is impermeable for NADH
- two shuttle systems:
- aspartate/malate shuttle (universal)
- glycerol phosphate shuttle (brain, kidney)

Aspartate/malate shuttle





Respiratory chain is the system of redox reactions in IMM which starts by the NADH oxidation and ends with the reduction of O_2 to water.



The free energy of oxidation of NADH/FADH₂ is utilized for pumping protons to the outside of the inner mitochondrial membrane. **The proton gradient** across the inner mitochondrial membrane represents the energy for ATP synthesis.

Four types of cofactors in R.CH.

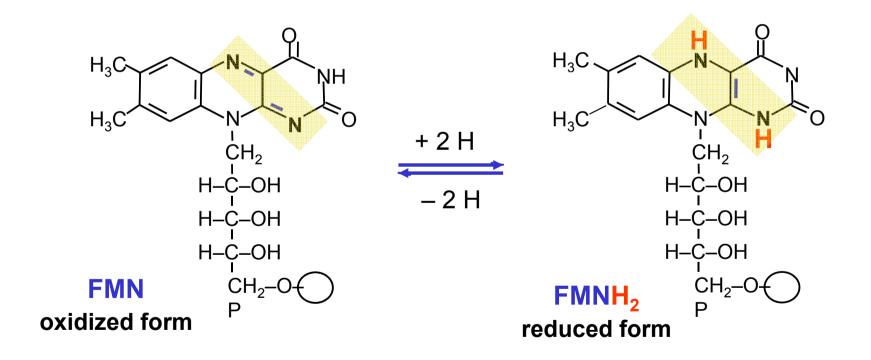
- flavine cofactors (FMN, FAD)
- non-heme iron with sulfur (Fe-S)
- ubiquinone (Q)
- heme (in cytochromes)

Distinguish:

heme (cyclic tetrapyrrol) × cytochrome (hemoprotein)

Flavoproteins

contain flavin prosthetic group as **flavin mononucleotide** (FMN, complex I) or **flavin adenine dinucleotide** (FAD, complex II):

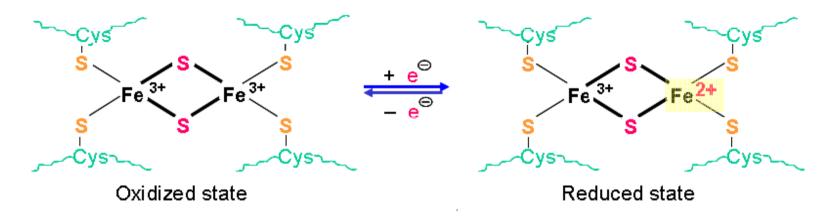


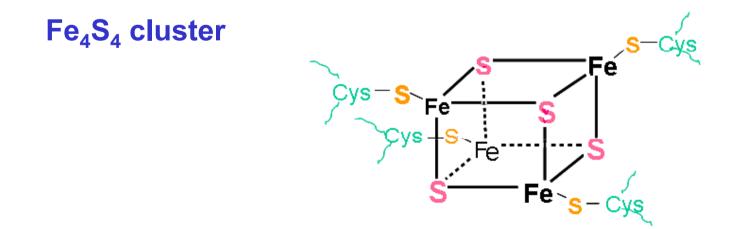
Coenzyme FMN (as well as FAD) transfers two atoms of hydrogen.

Iron-sulfur proteins (FeS-proteins, non-heme iron proteins)

Despite the different number of iron atoms present, each cluster accepts or donates **only one electron**.

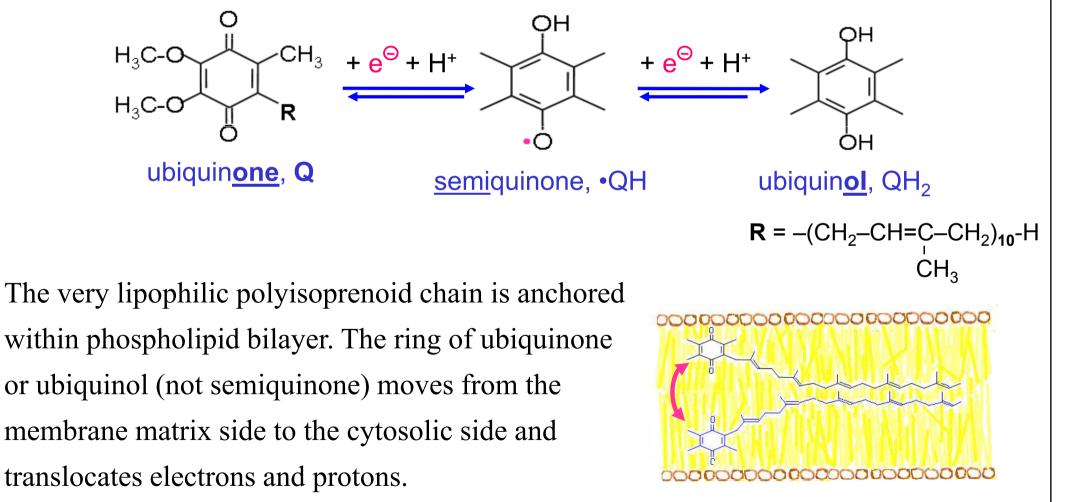
Fe₂S₂ cluster





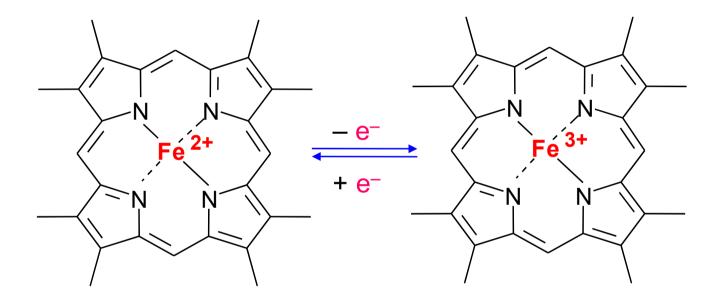
Ubiquinone (coenzyme Q)

It accepts stepwise **two electrons** (one from the complex I or II and the second from the cytochrome *b*) **and two protons** (from the mitochondrial matrix), so that it is fully reduced to ubiquinol:



Cytochromes

are **heme proteins**, which are **one-electron carriers** due to reversible oxidation of the iron atom:



Mammalian cytochromes are of three types -a, b, c. They differ in the substituents attached to the porphin ring. All these types of cytochromes occur in the mitochondrial respiratory chain.

Cytochromes type \boldsymbol{b} (including cytochromes class P-450) occur also in membranes of endoplasmic reticulum and the outer mitochondrial membrane.

Some differences in cytochrome structures

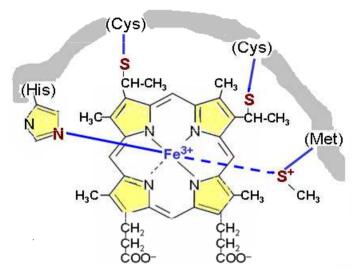
Cytochrome c

central Fe ion is attached by coordination to N-atom of His_{18} and to S-atom of Met_{80} ; two vinyl groups bind covalently S-atoms of Cys_{14} + Cys_{17} . The heme is dived deeply in the protein terciary structure so that it is unable to bind O₂, CO. Cyt *c* is water-soluble, peripheral protein that moves on the outer side of the inner mitochondrial membrane.

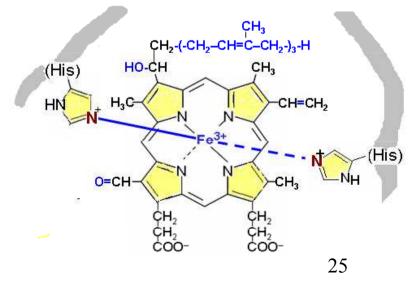
Cytochrome *aa*₃

central Fe ion is attached by coordination to two His residues; one of substituents is a hydrophobic isoprenoid chain, another one is oxidized to formyl. The heme *a* is the accepts an electrons from the copper centre A (two atoms Cu_A). Its function is inhibited by carbon monoxide, CN^- , HS⁻, and N₃⁻ anions.

Heme of cytochrome *c*



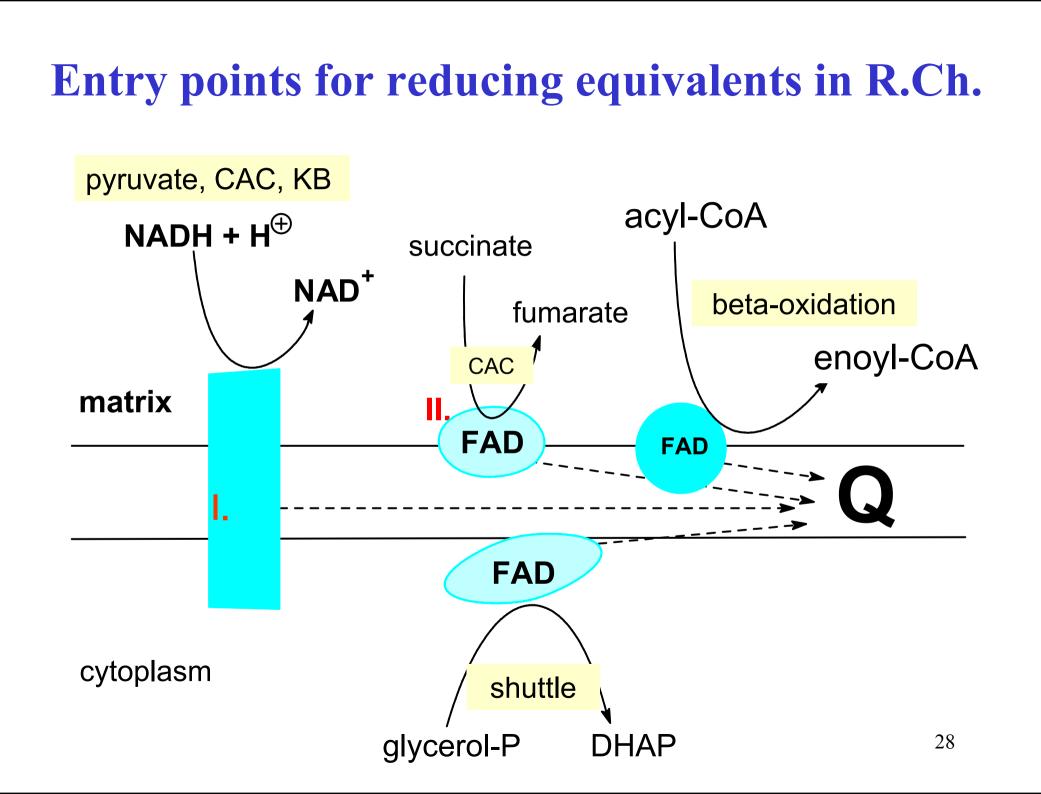




Redox pairs in the respiratory chain

Oxidized / Reduced form	<i>E</i> °′(V)	
NAD ⁺ / NADH+H ⁺	-0,32	
FAD / FADH ₂	0,00	
Ubiquinone (Q) / Ubiquinol (QH ₂)	0,10	
Cytochrome c_1 (Fe ³⁺ / Fe ²⁺)	0,22	
Cytochrome c (Fe ³⁺ / Fe ²⁺)	0,24	
Cytochrome a_3 (Fe ³⁺ / Fe ²⁺)	0,39	
O ₂ / 2 H ₂ O	0,82	

- redox pairs are listed according to increasing $E^{\circ'}$
- they are standard values (1 mol/l), real cell values are different
- the strongest reducing agent in R.CH. is NADH
- the strongest oxidizing agent in R.CH. is O_2
- the value of potential depends on protein molecule (compare cytochromes)



Enzyme complexes in respiratory chain

No.	Name	Cofactors	Oxidation	Reduction
I.	NADH-Q oxidoreductase*	FMN, Fe-S	$NADH \rightarrow NAD^+$	$Q \rightarrow QH_2$
II.	succinate-Q reductase	FAD,Fe-S,cyt b	$FADH_2 \rightarrow FAD$	$Q \rightarrow QH_2$
III.	Q-cytochrome- <i>c</i> -reductase	Fe-S, cyt b , c_1	$QH_2 \rightarrow Q$	$\operatorname{cyt} c_{\operatorname{ox}} \rightarrow \operatorname{cyt} c_{\operatorname{red}}$
IV.	cytochrome- <i>c</i> -oxidase	cyt <i>a</i> , <i>a</i> ₃ , Cu	$\operatorname{cyt} c_{\operatorname{red}} \to \operatorname{cyt} c_{\operatorname{ox}}$	$O_2 \rightarrow 2 H_2O$

* also called NADH dehydrogenase

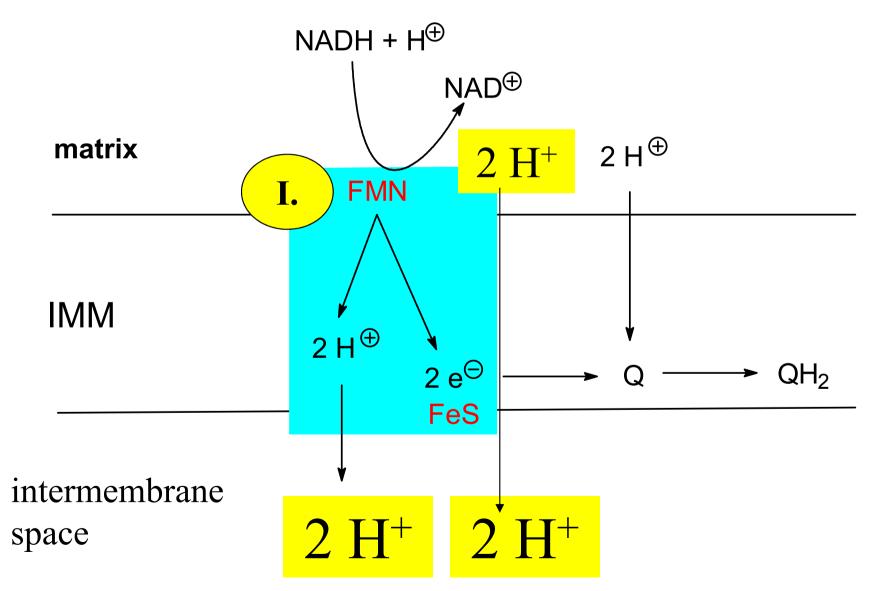
Complex I oxidizes NADH and reduces ubiquinone (Q)

FMN, Fe-S

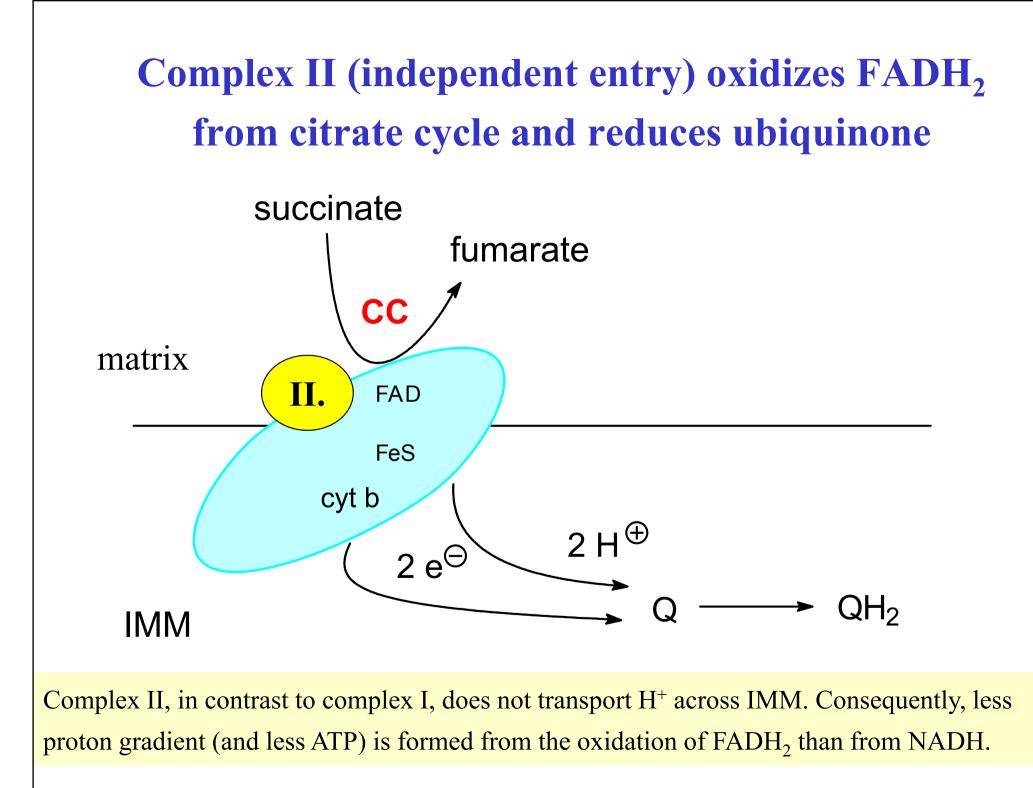
NADH+H⁺ + Q + 4 $H^+_{matrix} \rightarrow NAD^+ + QH_2 + 4 H^+_{ims}$

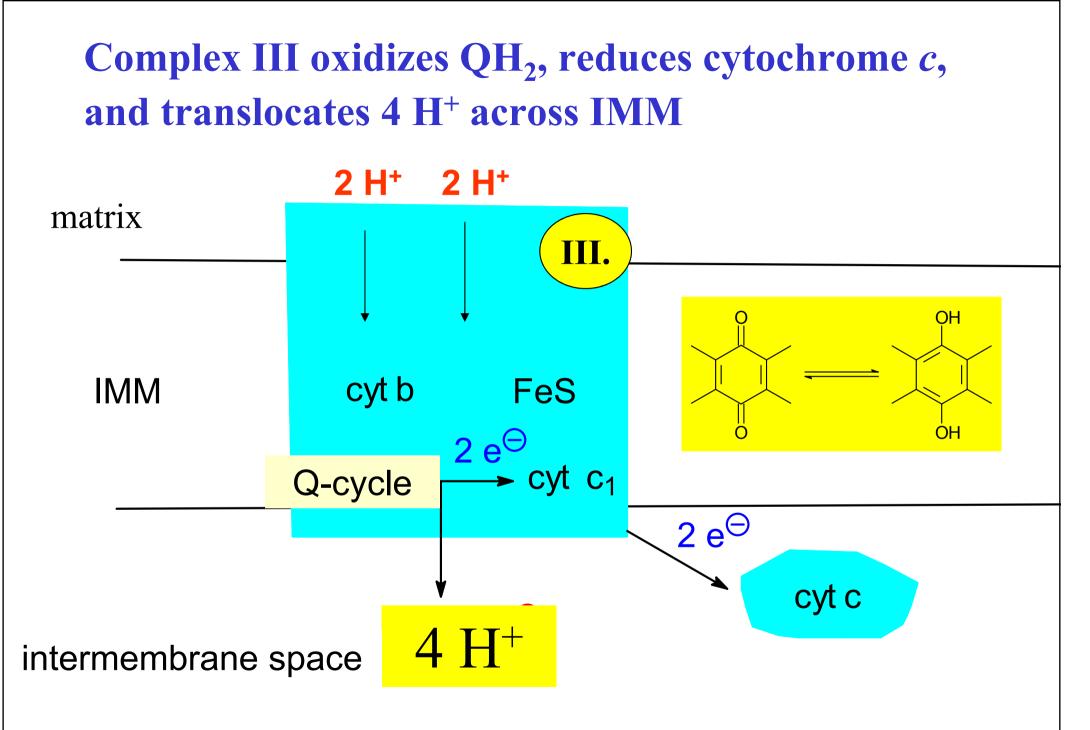
The four H⁺ are translocated from matrix to intermembrane space (ims)

Complex I oxidizes NADH and translocates 4 H⁺ into intermembrane space

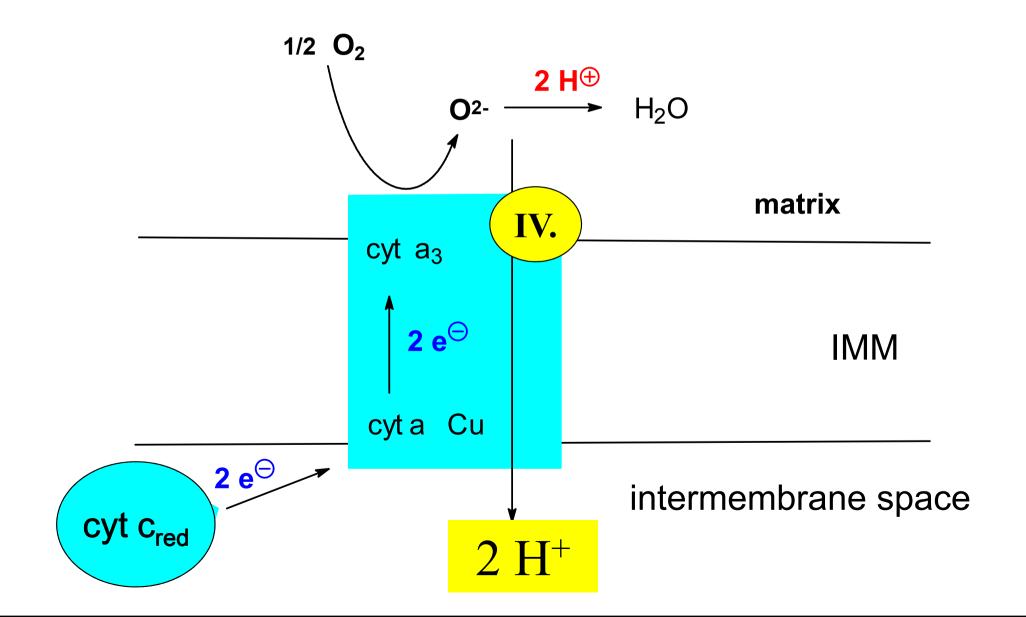


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Complex IV oxidizes cyt c_{red} and two electrons reduce monooxygen ($\frac{1}{2}O_2$)



Complex IV: real process is four-electrone reduction of dioxygen

partial reaction (redox pair):

$$O_2 + 4 e^- + 4 H^+ \rightarrow 2 H_2O$$

complete reaction:

 $4 \text{ cyt-Fe}^{2+} + \text{O}_2 + 8 \text{ H}^+_{\text{matrix}} \rightarrow 4 \text{ cyt-Fe}^{3+} + 2 \text{ H}_2\text{O} + 4 \text{ H}^+_{\text{ims}}$

metabolic water

For every 2 electrons, 2 H⁺ are pumped into intermembrane space

Three times translocated protons create electrochemical H⁺ gradient across IMM

It consists of two components:

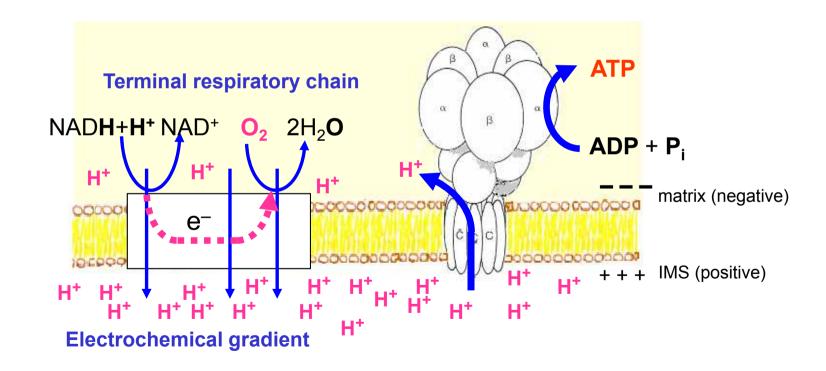
1) difference in **pH**, $\Delta G = \text{RT} \ln ([\text{H}^+]_{\text{out}}/[\text{H}^+]_{\text{in}}) = 2.3 \text{ RT}(\text{pH}_{\text{out}} - \text{pH}_{\text{in}})$ 2) difference in **electric potential** ($\Delta \Psi$, negative inside), depends not only on protons, but also on concentrations of other ions, $\Delta G = -nF\Delta \Psi$ The **proton motive force** Δp is the quantity expressed in the term of potential (milivolts per mole of H⁺ transferred): $\Delta p = -\Delta G / nF = \Delta \Psi + 60 \Delta \text{ pH}$.

Utilization of proton motive force

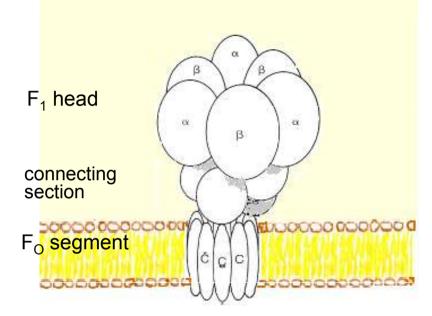
- synthesis of ATP = aerobic phosphorylation
- heat production especially in brown adipose tissue
- active transport of ions and metabolites across IMM

Aerobic phosphorylation of ADP by ATP synthase

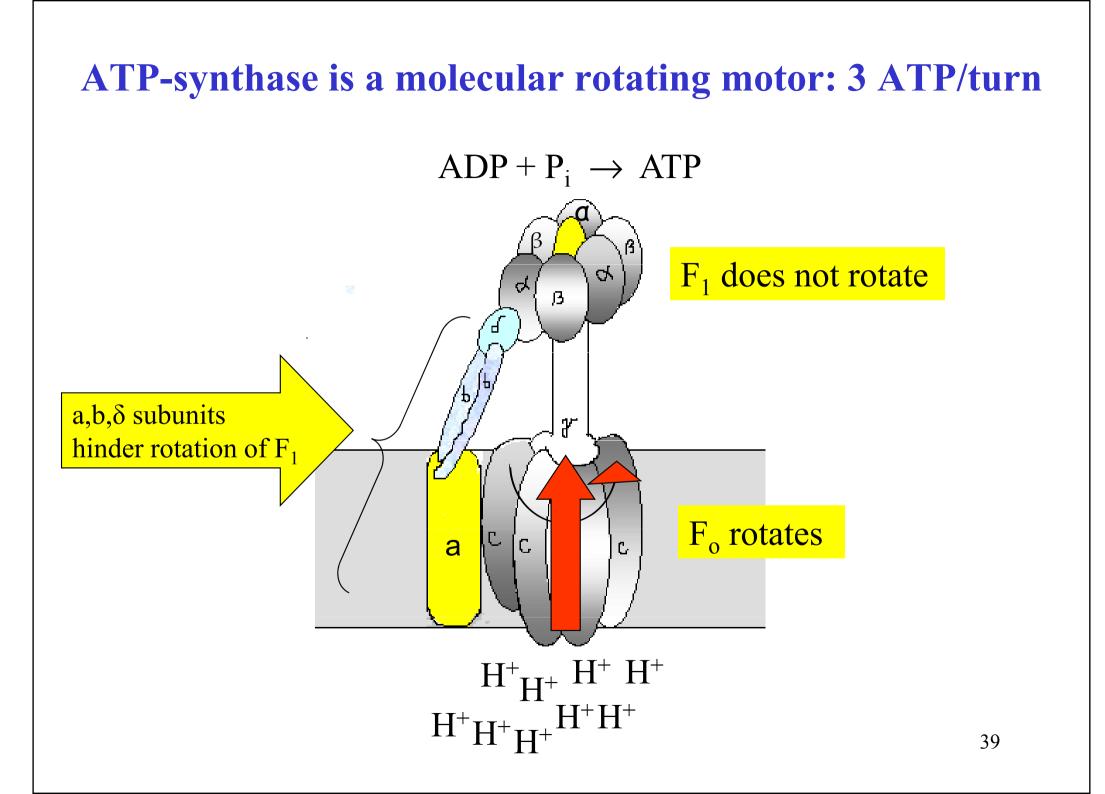
The endergonic phosphorylation of ADP is driven by the flux of protons back into the matrix along the electrochemical gradient through ATP synthase.



ATP synthase consists of three parts



- 1) F_1 complex projects into the matrix, 5 subunit types (α_3 , β_3 , γ , δ , ϵ) catalyze the ATP synthesis
- 2) connecting section
- 3) F_o inner membrane component, several *c*-units form a rotating proton channel



Stoichiometry of ATP synthesis is not exactly recognized

- transfer of 2 e⁻ from NADH to $\frac{1}{2}$ O₂ 3 ATP
- transfer of 2 e⁻ from FADH₂ to $\frac{1}{2}$ O₂ 2 ATP

new research data indicate somewhat lower values (see Harper)

- transfer of 2 e⁻ from NADH to $\frac{1}{2}$ O₂ 2.5 ATP
- transfer of 2 e⁻ from FADH₂ to $\frac{1}{2}$ O₂ 1.5 ATP

Control of the oxidative phosphorylation

Production of ATP is strictly coordinated so that ATP is never produced more rapidly than necessary.

Synthesis of ATP depends on:

- supply of substrates (mainly NADH+H⁺)
- supply of dioxygen
- the energy output of the cell; hydrolysis of ATP increases the concentration of ADP in the matrix, which activates ATP production.

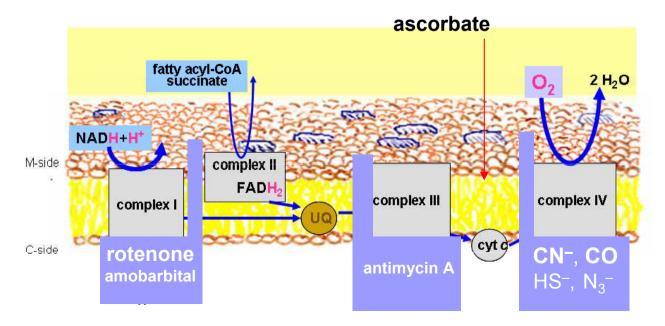
This mechanism is called respiratory control.

Inhibitors of the terminal respiratory chain

Complex I is blocked by an insecticide <u>rotenone</u>. A limited synthesis of ATP exists due to electrons donated to ubiquinone through complex II.

Complex III is inhibited by <u>antimycin A</u> – complexes I and II become reduced, complexes III and IV remain oxidized. Ascorbate restores respiration, because it reduces cyt c.

Complex IV is blocked by <u>carbon monoxide</u>, <u>cyanide</u> ion, <u>HS</u>⁻ (sulfane intoxication), <u>azide</u> ion N₃⁻. Respiration is disabled.



Cyanide poisoning

occurs after ingestion of alkali cyanides or inhalation of hydrogen cyanide. Bitter almonds or apricot kernels contain amygdalin, which can release HCN.

Cyanide ion, besides inhibition of cytochrome c oxidase, binds with high affinity onto methemoglobin (Fe³⁺).

The lethal dose LD_{50} of alkali cyanide is about 250 mg. Symptoms - dizziness, gasping for breath, cramps, and unconsciousness follow rapidly.

Antidotes may be effective, when applied without any delay:

Hydroxycobalamin (a semisynthetic compound) exhibits high affinity to CN^- ions, binds them in the form of harmless cyanocobalamin (B₁₂).

Sodium nitrite NaNO₂ or amyl nitrite oxidize hemoglobin (Fe^{II}) to methemoglobin (Fe^{III}), which is not able to transport oxygen, but binds CN^- and may so prevent inhibition of cytochrome *c* oxidase.

Sodium thiosulfate $Na_2S_2O_3$, administered intravenously, can convert cyanide to the relatively harmless thiocyanate ion: $CN^- + S_2O_3^{2-} \rightarrow SCN^- + SO_3^{2-}$.

Carbon monoxide poisoning

CO binds primarily to hemoglobin (Fe^{II}) and inhibits oxygen transport, but it also blocks the respiratory chain by inhibiting cytochrome oxidase (complex IV). **Oxygenotherapy** improves blood oxygen transport, administered **methylene blue** serves as acceptor of electrons from complex III so that limited ATP synthesis can continue. 43

Uncoupling of the respiratory chain and phosphorylation

is the **wasteful oxidation of substrates without concomitant ATP synthesis:** protons are pumped across the membrane, but they re-enter the matrix using some other way than that represented by ATP synthase.

The free energy derived from oxidation of substrates appears as heat..

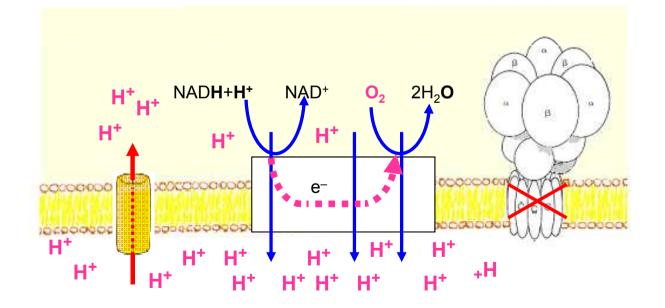
There are **four types** of artificial or natural uncouplers:

- 1 "True" uncouplers compounds that transfer protons through the membrane. A typical uncoupler is 2,4-dinitrophenol (DNP): DNP is very toxic, the lethal dose is about 1 g. More than 80 years ago, the long-term application of small doses (2.5 mg/kg) was recommended as a "reliable" drug in patients seeking to lose weight. Its use has been banned, because hyperthermia and toxic side effect (with fatal results) were excessive.
- 2 **Ionophors** that do not disturb the chemical potential of protons, but diminish the electric potential $\Delta \Psi$ by enabling free re-entry of K⁺ (e.g. valinomycin) or both K⁺ and Na⁺ (e.g. gramicidin A).
- **3** Inhibitors of ATP synthase oligomycin.
- **4 Inhibitors of ATP/ADP translocase** like unusual plant and mould toxins bongkrekic acid (irreversibly binds ADP onto the translocase) and atractylate (inhibits binding of ATP to the translocase). ATP synthase then lacks its substrate.

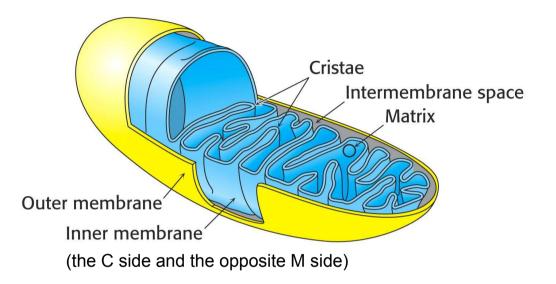
Thermogenin is a natural uncoupler

is a inner mitochondrial membrane protein that transports protons back into the matrix, bypassing so ATP synthase.

It occurs in **brown adipose tissue** of newborn children and hibernating animals, which spend the winter in a dormant state.



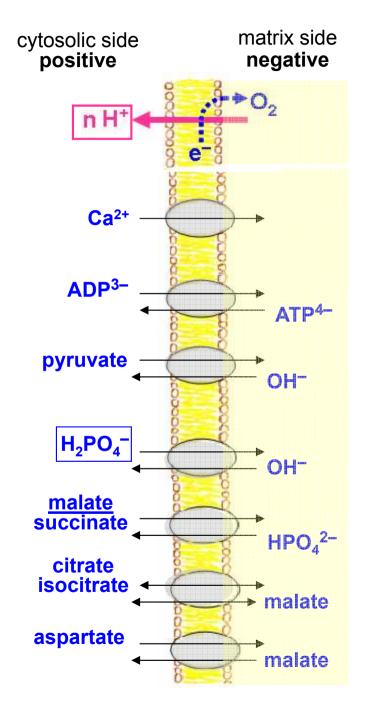
Mitochondrial metabolite transport



The outer membrane is quite permeable for small molecules and ions – it contains many copies of **mitochondrial porin** (voltage-dependent anion channel, VDAC).

The inner membrane is intrinsically impermeable to nearly all ions and polar molecules, but there are many **specific transporters** which shuttles **metabolites** (e.g. pyruvate, malate, citrate, ATP) and **protons** (terminal respiratory chain and ATP synthase) across the membrane.

Transport through the inner mitochondrial membrane



Free diffusion of O₂, CO₂, H₂O, NH₃

Primary active H⁺ transport forms the proton motive force (the primary gradient)

Secondary active transports driven by a H⁺ gradient and dissipating it:

ATP/ADP translocase

pyruvate transporter

phosphate permease – forms a (<u>secondary</u>) phosphate gradient

dicarboxylate carrier

tricarboxylate carrier

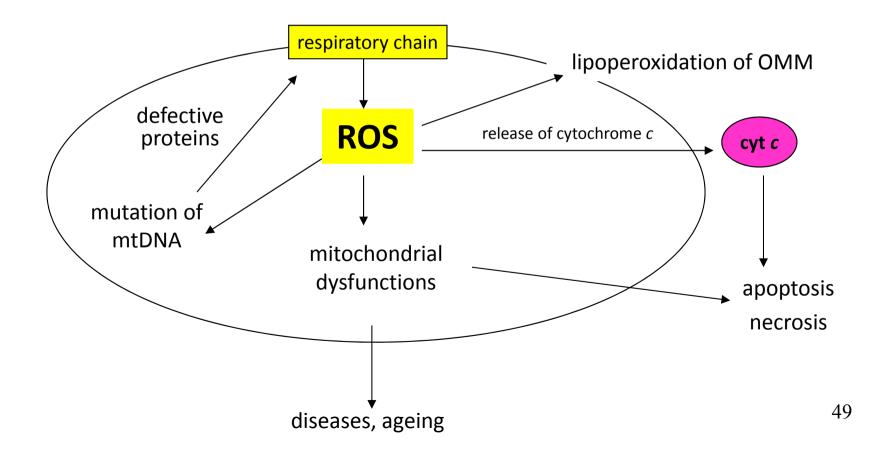
the malate shuttle for NADH + H^+

Mitochondria and apoptosis

- apoptosis is a controlled process of cell death with minimal effect on surrounding tissue
- apoptosis is important for physiological tissue turnover
- apoptosis is regulated by a number of cell signals
- regulatory protein family Bcl-2 (B-cell lymphoma 2)
- some proteins are anti-apoptotic (Bcl-xl), other pro-apoptotic (Bax, Bak)
- Bax and Bak proteins oligomerize and make a pore in outer mitochondrial membrane
- cytochrome c is released into cytosol, binds with inactive caspases and other pro-apoptotic factors creates **apoptosome** and triggers the executive phase of apoptosis (caspase cascade)

Mitochondria and oxidative stress

- about 98 % of O₂ is consumed in respiratory chain for the complete reduction to water (cytochrome-*c*-oxidase)
- however, other partly reduced oxygen species are also produced
- they are called reactive oxygen species (ROS)
- mainly in compl. I, III, especially, if electron trasport is slowned down or reversed
- mitochondria contains a number of antioxidants (GSH, QH₂, superoxide dismutase)



Reactive oxygen species in human body

Radicals	Neutral / Anion / Cation
Superoxide $\cdot O_2^-$	Hydrogen peroxide H-O-O-H
Hydroxyl radical ·OH	Hydroperoxide* R-O-O-H
Peroxyl radical* ROO·	Hypochlorous acid HClO
Alkoxyl radical RO·	Singlet oxygen ¹ O ₂
Hydroperoxyl radical HOO·	Peroxynitrite ONOO-
Nitric oxide NO·	Nitronium NO ₂ ⁺

* Typically phospholipid-PUFA derivatives during lipoperoxidation: PUFA-OO·, PUFA-OOH

Superoxide anion-radical •O₂

• <u>One-electrone</u> reduction of dioxygen

 $O_2 + e^- \rightarrow O_2^-$

[one redox pair]

Superoxide formation

- **Respiratory burst** (in neutrophils) $2 O_2 + NADPH \longrightarrow 2 \cdot O_2^- + NADP^+ + H^+$
- Spontaneous oxidation of heme proteins

heme-Fe²⁺ + $O_2 \longrightarrow heme-Fe^{3+} + O_2^-$

[complete redox reactions, combinations of two redox pairs]

Radical •OH is the most reactive species; it is formed from superoxide and hydrogen peroxide

$\bullet O_2^- + H_2O_2 \rightarrow O_2 + OH^- + \bullet OH$

Catalyzed by reduced metal ions (Fe²⁺, Cu⁺) (Fenton reaction)

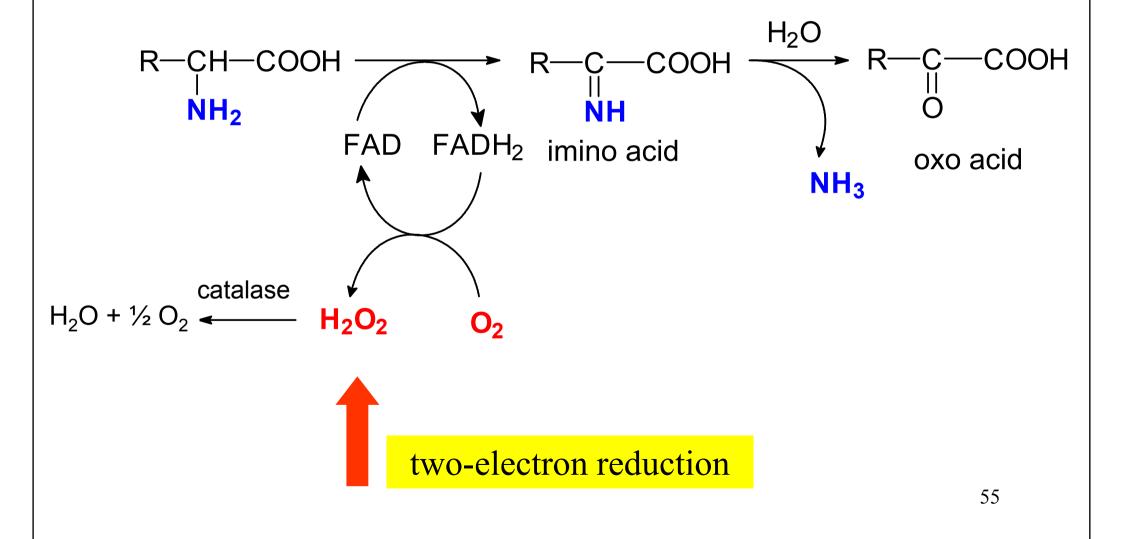
Singlet oxygen ¹O₂

- excited form of triplet dioxygen
- formed after absorption of light by some compounds (porphyrins)

$$^{3}O_{2} \rightarrow ^{1}O_{2}$$

for electron configuration see Medical Chemistry I, p. 18

Hydrogen peroxide H_2O_2 is a side product in the deamination of certain amino acids



Xanthin oxidase reaction produces hydrogen peroxide

hypoxanthin + O_2 + $H_2O \rightarrow xanthin + H_2O_2$

xanthin + O_2 + $H_2O \rightarrow uric acid + H_2O_2$

most tissues, mainly liver

Compare: reduction of dioxygen

Type of reduction	Redox pair
Four-electron	$O_2 + 4 e^- + 4 H^+ \rightarrow 2 H_2O$
One-electron	$O_2 + e^- \rightarrow \cdot O_2^-$
Two-electron	$O_2 + 2 e^- + 2 H^+ \rightarrow H_2O_2$

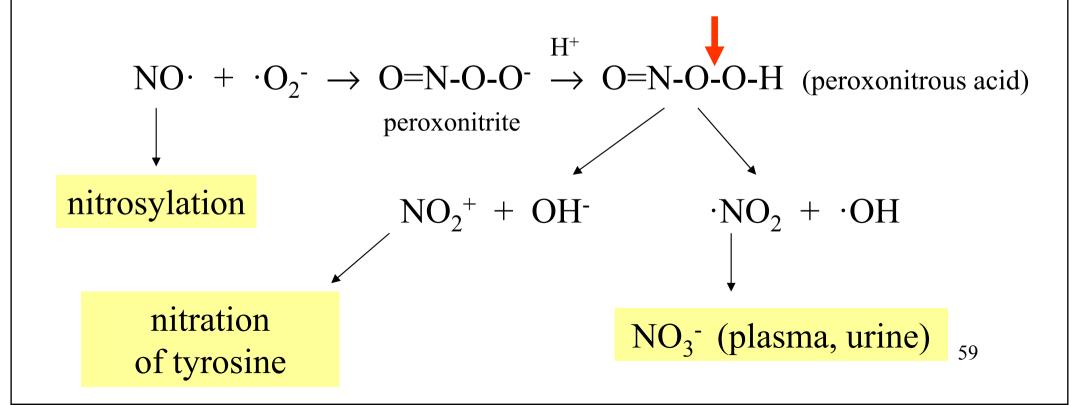
Hypochlorous acid HClO

- in some neutrophils
- myeloperoxidase reaction
- HClO has strong oxidative and bactericidal effects

$H_2O_2 + Cl^- + H^+ \rightarrow HClO + H_2O$

Nitric oxide NO \cdot is released from arginine

- exogenous sources: drugs vasodilators
- NO· activates guanylate cyclase \Rightarrow cGMP \Rightarrow relaxation of smooth muscles
- NO \cdot is a radical and affords other reactive metabolites:



Compounds releasing NO

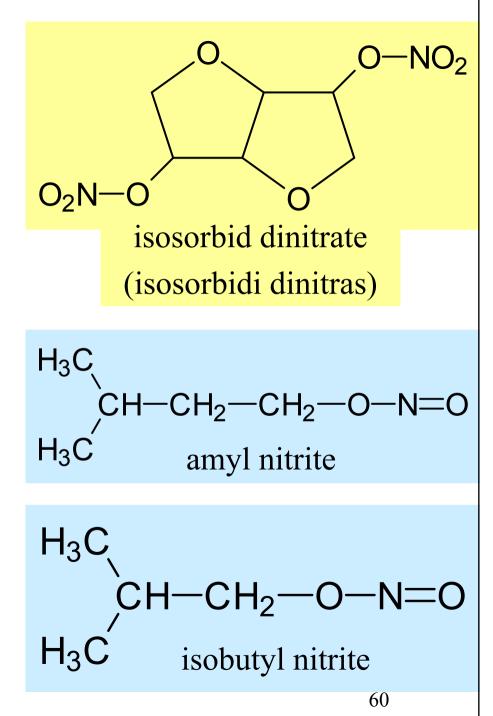
$$CH_{2} - O - NO_{2}$$

 $CH - O - NO_{2}$
 $CH_{2} - O - NO_{2}$

glycerol trinitrate (glyceroli trinitras)

Na₂[Fe(CN)₅NO]

sodium nitroprusside (natrii nitroprussias) sodium pentacyanonitrosylferrate(III)



Good effects of ROS

- intermediates of oxidase and oxygenase reactions
 (cyt P-450), during reactions the radicals are trapped in enzyme molecule so that they are not harmful
- bactericidal effect fagocytes, respiratory burst (NADPH-oxidase)
- **signal molecules** clearly proved in NO·, perhaps other radical species can have similar action

Bad effects of ROS

Substrate	Damage	Consequences
PUFA	formation of aldehydes (MDA) and peroxides	changes in membrane permeability, damage of membrane enzymes
Proteins	aggregation, cross-linkage fragmentation oxidation of –SH, phenyl	changes in ion transport influx of Ca ²⁺ into cytosol altered enzyme activity
DNA	deoxyribose decomposition modification of bases chain breaks	mutations translations errors inhibition of proteosynthesis

Antioxidant systems in the body

Enzymes

• superoxide dismutase, catalase, glutathione peroxidase

Low molecular antioxidants = reducing compounds with

- phenolic -OH (tocopherol, flavonoids, urates)
- enolic -OH (ascorbate)
- -SH (glutathione GSH, dihydrolipoate)
- or compounds with extended system of conjugated double bonds (carotenoids)

Elimination of superoxide

- Superoxide dismutase
- Catalyzes the dismutation of superoxide

 $2 \cdot O_2^- + 2 H^+ \longrightarrow O_2^- + H_2O_2^-$

• Oxidation numbers of oxygen

 $-\frac{1}{2} \longrightarrow 0 -I$

• two forms:

SOD1 (Cu, Zn, cytosol), SOD2 (Mn, mitochondria)

Dismutation is a special type of redox reaction in which an element is simultaneously reduced and oxidized so as to form two different products.

Elimination of H₂O₂

• **catalase** - in erythrocytes and other cells

 $H_2O_2 \rightarrow \frac{1}{2}O_2 + H_2O_2$

• glutathione peroxidase

3% H₂O₂ aplied to a wound releases bubbles

- contains selenocystein, reduces H_2O_2 and hydroperoxides of phospholipids (ROOH)
 - $2 \text{ G-SH} + \text{H-O-O-H} \rightarrow \text{G-S-S-G} + 2 \text{H}_2\text{O}$
 - $2 \text{ G-SH} + \text{R-O-O-H} \rightarrow \text{G-S-S-G} + \text{R-OH} + \text{H}_2\text{O}$

Lipophilic antioxidants

Antioxidant Sources

Tocopherol Plant oils, nuts, seeds, germs

Carotenoids Fruits, vegetables (most effective is lycopene)

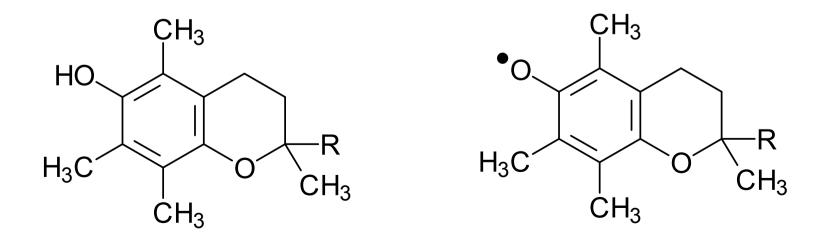
Ubiquinol Formed in the body from tyrosine

Hydrophilic antioxidants

Antioxidant	Sources
L-ascorbate	Fruits, vegetables, potatoes
Flavonoids	Fruits, vegetables, tea, wine
Dihydrolipoate	Made in the body from cysteine
Uric acid	Catabolite of purine bases
Glutathione	Made in the body from cysteine

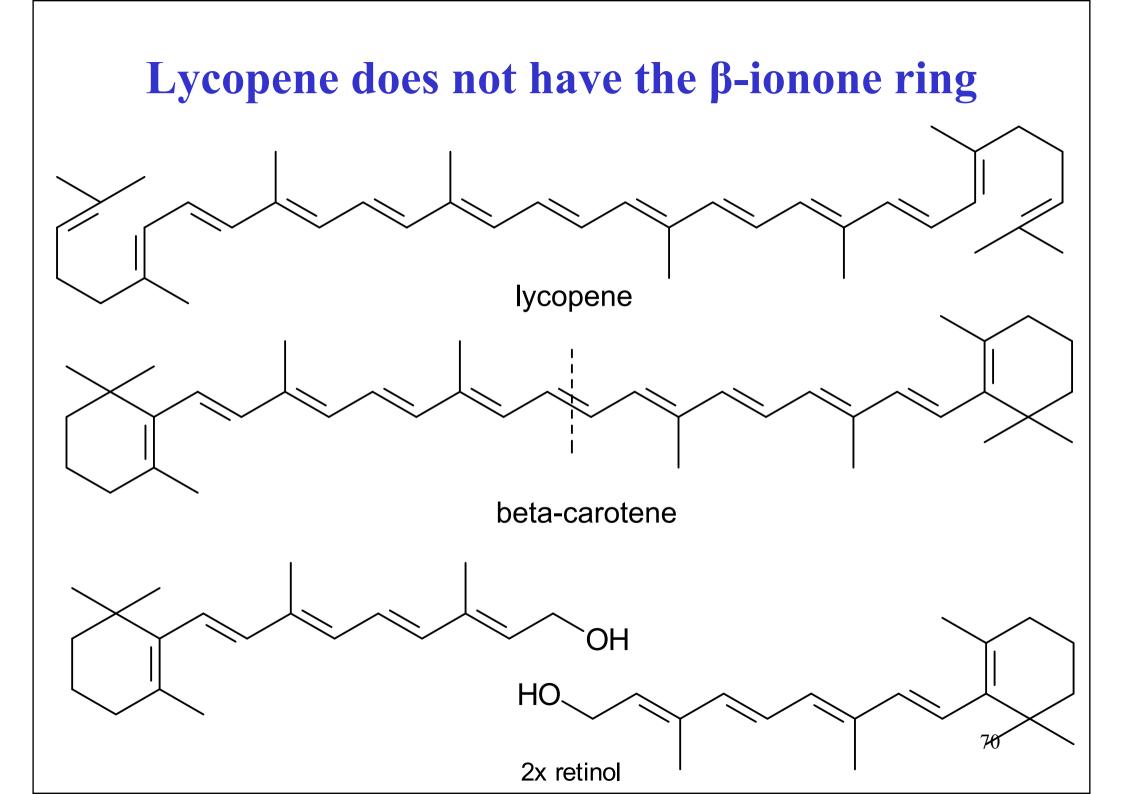
Tocopherol (Toc-OH)

- Lipophilic antioxidant of cell membranes and lipoproteins
- Reduces peroxyl radicals of phospholipids to hydroperoxides which are further reduced by GSH, tocopherol is oxidized to stable radical Toc-O· $PUFA-O-O· + Toc-OH \rightarrow PUFA-O-O-H + Toc-O·$
- Toc-O \cdot is partially reduced to Toc-OH by ascorbate or GSH



Carotenoids

- polyisoprenoid hydrocarbons (tetraterpens)
- eliminate peroxyl radicals
- they can quench singlet oxygen
- food sources: green leafy vegetables, yellow, orange, red vegetables and fruits
- very potent antioxidant is **lycopene** (tomatoes, more available from cooked tomatoes, ketchup etc.)



Lycopene in food (mg/100 g)

Tomato purée	10-150
Ketchup	10-14
Tomato juice/sauce	5-12
Watermelon	2-7
Papaya	2-5
Tomatoes fresh	1-4
Apricots canned	~ 0.06
Apricots fresh	~ 0.01

In order to effectively

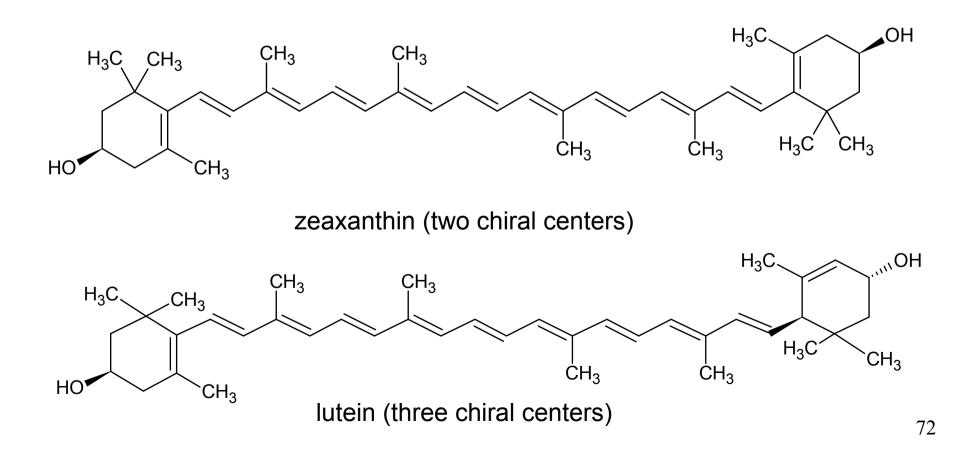
absorb lycopene,

tomatoes should be

- chopped and mashed
- stewed slowly
- combined with oil

Zeaxanthin and lutein

- belong to xanthophylls oxygen derivatives of carotenoids
- they differ in the position of double bond and in the number of C*
- occur mainly in green leafy vegetables (spinach, cabbage, kale)
- contained in macula lutea, prevents it against degeneration
- many pharmaceutical preparations available



Ubiquinol (QH₂)

- occurs in all membranes
- Endogenous synthesis by intestinal microflora from tyrosine and farnesyl diphosphate (biosyntheis of cholesterol)
- Exogenous sources: liver, meat and other foods
- Reduced form QH₂ regenerates to copherol
- Toc-O· + $QH_2 \rightarrow Toc-OH + \cdot QH$

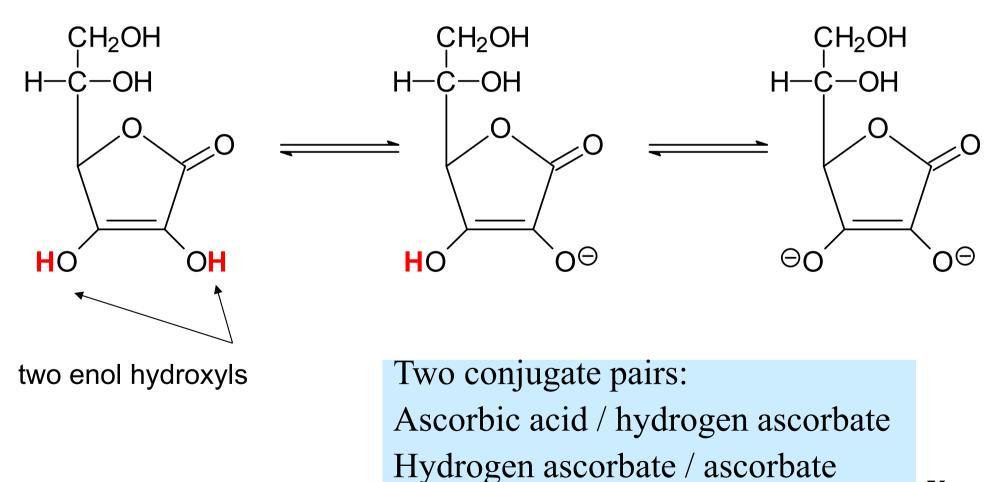
L-Ascorbate (vitamin C)

- cofactor of proline hydroxylation (maturation of collagen)
- cofactor of dopamine hydroxylation (to noradrenaline)
- potent reducing agent (Fe³⁺ \rightarrow Fe²⁺, Cu²⁺ \rightarrow Cu⁺)
- supports intestinal absorption of iron
- Reduces many radicals: $\cdot OH$, $\cdot O_2^-$, HO_2^- , ROO^-
- Regenerates tocopherol
- It is catabolized to oxalate!! (high doses are not recomended)
- excess of ascorbate has pro-oxidative effects:

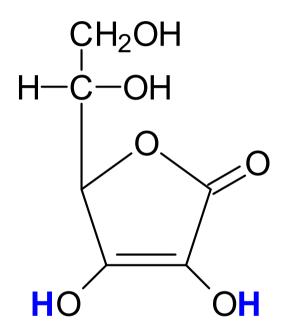
Fe²⁺ and Cu⁺ catalyze the formation of hydroxyl radical ascorbate + $O_2 \rightarrow O_2^-$ + ·monodehydroascorbate

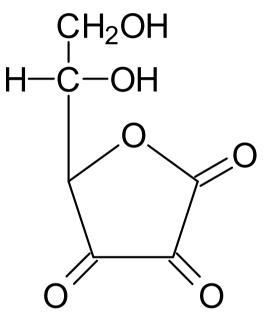
L-Ascorbic acid is a weak diprotic acid

 $pK_{A1} = 4.2$ $pK_{A2} = 11.6$



L-Ascorbic acid has reducing properties (antioxidant)





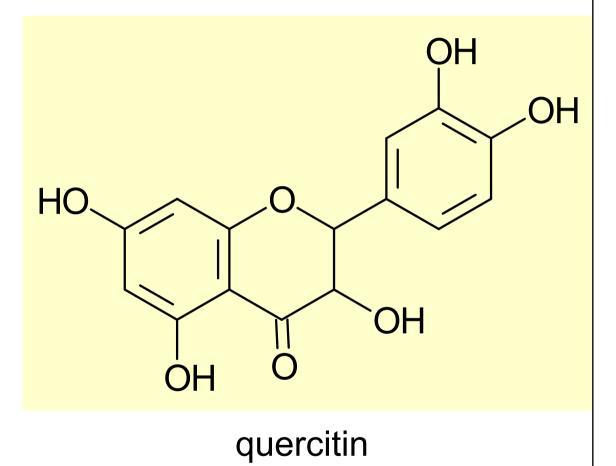
ascorbic acid (reduced form) dehydroascorbic acid (oxidized form)

Flavonoids and other polyphenols

- commonly spread in plant food
- total intake about 1 g (higher than in vitamins)
- derivatives of chromane (benzopyrane), many phenolic hydroxyls
- a main example: **quercitin** (see also Med. Chem. II, p. 76)
- they reduce free radicals, themselves are converted to unreactive phenoxyl radicals
- they chelate free metal ions (Fe²⁺, Cu⁺) blocking them to catalyze
 Fenton reaction and lipoperoxidation

Main sources of flavonoids

- vegetable (onion)
- fruits (apples, grapes)
- green tea
- cocoa, quality chocolate
- olive oil (Extra Virgin)
- red wine



Glutathione (GSH)

 NH_2

- tripeptide
- γ -glutamylcysteinylglycine HOOC α
- made in all cells
- reducing agent (-SH)
- reduces H_2O_2 and ROOH (glutathione peroxidase)
- reduces many ROS
- regenerates -SH groups of proteins and coenzyme A
- regenerates tocopherol and ascorbate

COOH

CH₂

SH

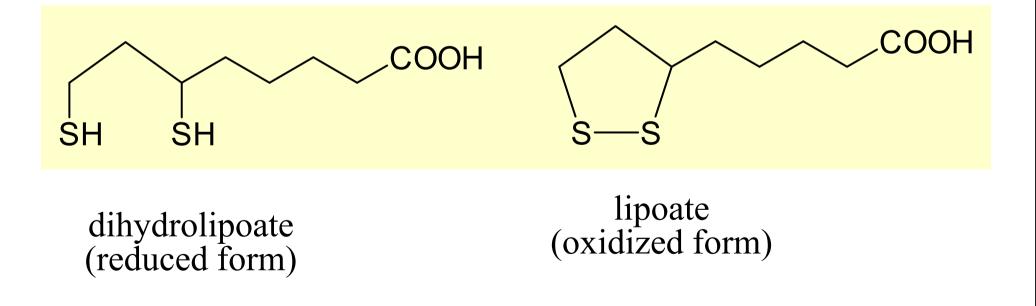
Regeneration of reduced form of GSH

- continuous regeneration of GSH proceeds in many cells
- glutathione reductase, esp. in erythrocytes
- GSSG + NADPH + H⁺ \rightarrow 2 GSH + NADP⁺

pentose cycle

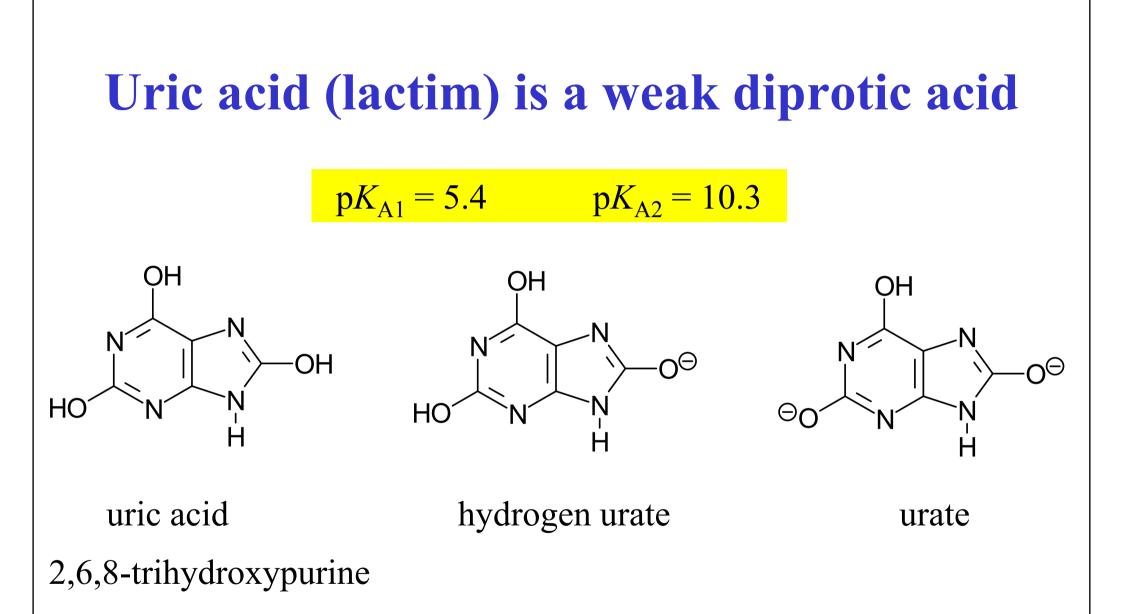
Dihydrolipoate

- cofactor of oxidative decarboxylation of pyruvate, 2-OG
- reduces many radicals (mechanism not well understood)



Uric acid

- final catabolite of purine bases
- in kidney, tubular cells, 90 % of urates are resorbed
- the most abundant antioxidant of blood plasma
- reducing properties, reduces various radicals
- has ability to chelate iron and copper ions



Uric acid is the most abundant plasma antioxidant

