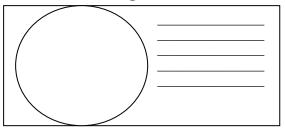
Topic P08: Laboratory diagnostics of tuberculosis, actinomycetes and nocardiae

Task 1: Microscopy of acid-fast and filamentous microoorganisms

While entirely acid-fast microorganisms (*Mycobacterium*) cannot be stained at all according to Gram, only partially acid-fast ones (*Nocardia*) or not at all (*Actinomyces*) can be Gram-stained. Typical morphology of *Nocardia* and *Actinomyces* is that of branched filaments, but sometimes they might be shorter, or even coccoid.

a) Staining of (negative) clinical sample using Ziehl-Neelsen staining method

Ziehl-Neelsen staining is used for mycobacteria (*M. tuberculosis, M. leprae*), but also for some parasites (*Cryptosporidium parvum, Cyclospora cayetanensis*). The acid-fast organisms are stained only when heated during staining or in the "cold" variant (according to Kiyoun) at use of concentrated carbolfuchsin and concentrated phenol. On the other hand, after that, they are not decolorized even by so called "acid alcohol" (solution of alcohol with HCl or H₂SO₄). Decolorized background is then counterstained by a blue or green contrast dye.



Stain the negative sputum according to the Ziehl-Neelsen method (classic "hot" method, methylene blue variant). It is not likely that acid-fast rods would be present. Observe in microscope (immersion). Draw the results; at least, you will see the background, e. g. leucocytes, epithelia and other objects. Do not forget do **describe** your picture (use lines)!

Describe also the staining procedure – fill in the following table with names of used reagents

Describe this the standing procedure in in the following there with names of used reagents.					
1.	During the staining the preparation is		until		
2.	This reagent is made of		and		
3.	Instead of this reagent, it is also possible to use				

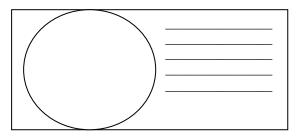
b) Microscopy of a mycobacterial culture

Examine microscopically (immersion oil, immersion 100× objective) the preparation of mycobacterial culture stained by Ziehl-Neelsen staining method.

Evaluate presence of red acid-fast rods.

Draw observed structures.

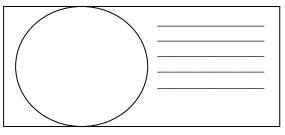
Do not forget do **describe** your picture (use lines)!



c) Microscopic examination of actinomycetes and nocardia strains

Examine microscopically the slide stained by Gram. Describe and draw observed formations. Observe high polymorphism of the microorganisms (from coccal shape, through rods to fibre/strings, often branched; Grampositive, but often staining half Gram-negative).

Do not forget do **describe** your picture (use lines)!



Task 2: Culture of mycobacteria, Actinomyces and Nocardia.

The culture requests of acid fast and partialy bacteria are very different.

- ❖ For *Mycobacterium tuberculosis* we use special media: liquid media (Šula, Banič) and solid media (Ogawa, Löwenstein-Jenssen). The solid media are different from majority of other solid media used in medical microbiology; they do not contain agar, they are "solid" because of coagulated egg proteins. Before culturing, the examined specimens should be rid of other microbes, usually by NaOH ("pickling")
- ❖ For *Nocardia* a current blood agar is sufficient.
- For *Actinomyces* we need VL-agar and culture in anaerostat/anaerobic jar (see P07), as this organism is microaerophilic with so low need for oxygen that anaerobic conditions are optimal for it.

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Medium name		Liquid/solid	Colou	r	Notes
b) Describe a	and drav	w the growth	of Mycobacterium,	Actinomyces and	d <i>Nocardia</i> on/in differ
Bacterium	Mediu	m name		of growth, possibly ords to characterize	also growth character
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		bic WCHA aga ed "WCHA")	ır		
Nocardia		ed "KA")			
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		test is used for te			
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a) Determina By comparing	ntion of s g with a ntuberculo	susceptibility to a control test-to	antituberculotics	lts of antitubercul	lotics susceptibility tests Growth control
a) Determina By comparing Mycobacterium Antituberculoti Growth Y/N	ntion of s g with a ntuberculo	susceptibility to a control test-to	antituberculotics	lts of antitubercul	• •
a) Determina By comparing Mycobacterium Antituberculoti	ntion of s g with a ntuberculo	susceptibility to a control test-to	antituberculotics	lts of antitubercul	• •
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Task 4: PCR in the TB diagnostics

As the culture of mycobacteria is rather prolonged (on average 6 weeks), PCR becomes a very important method in the diagnostics of TB.

Read a result of PCR TB diagnostics (from the slideshow), write down the results and interpret them.

Patient No.	Sample band	Control band	Interpretation
1			
2			
3			
4			

Task 5: Diagnostics of leprosy

Leprosy is a disease that still affects millions of people in underdeveloped countries. Its laboratory diagnostics is difficult because *Mycobacterium leprae* does not grow on artificial media. Fill in the following table.

The name of this animal is	
It is used to produce	
and this substance is used for	

Picture source: http://www.1-costaricalink.com/costa_rica_fauna/nine_banded_armadillo.htm

Task 6: Indirect TB detection by means of QUANTIFERON[©]-TB Gold test

It is a test of induced interferon gamma release checking and by means of this, checking of the cell-mediated immunity. **Test principle:** It was proven that in TB, including latent TB, tuberculosis antigens activate T-lymphocytes and they produce big amounts of interferon gamma. Similarly those T-lymphocytes may be activated non-specifically by so called mitogene; that is why mitogene is used as a positive control (MIT). As a negative control we use a test tube containing nothing (NIL). The test tube with proper TB antigen is labeled "TB". Interferon itself is detected by ELISA reaction.

Interpret the Quantiferon-TB Gold examination in four patients with use of interpretation table.

Anna:	MIT = 4.8	TB = 1.2	NIL = 1.1	Your interpretation:
Berta:	MIT = 5.3	TB = 4.8	NIL = 2.1	Your interpretation:
Cecil:	MIT = 0.9	TB = 0.9	NIL = 0.8	Your interpretation:
Dimos:	MIT = 8.4	TB = 8.3	NIL = 8.2	Your interpretation:

(all values are in IU/ml)

Interpretation table (according to test recommendations; simplified!)

NIL	TB minus NIL	MIT minus NIL	Final test interpretation	Presence of infectionM. tuberculosis	
	< 0.35	≥ 0.5	negative	Not likely	
≤ 8,0	≥ 0.35	any value	positive	Likely	
	< 0.35	< 0.5	uncuro	Connot be determined	
> 8,0	any value	any value	unsure	Cannot be determined	

Note: Updated variant of QUANTIFERON test contains four (and not three) test tubes, as "TB" is replaced by two types of antigens. Nevertheless, for simplification, we count here with the classic variant of the test.

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