

# Embryologie I OOGENESIS

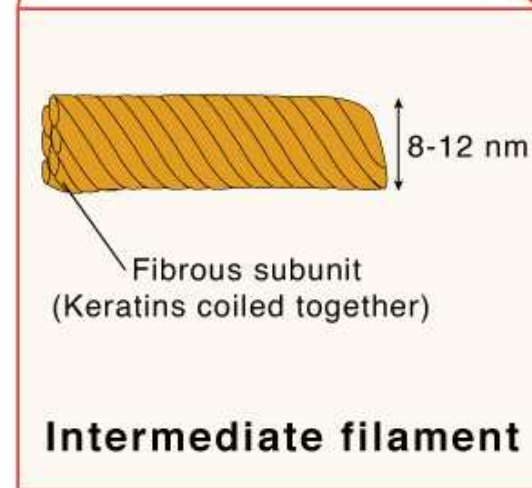
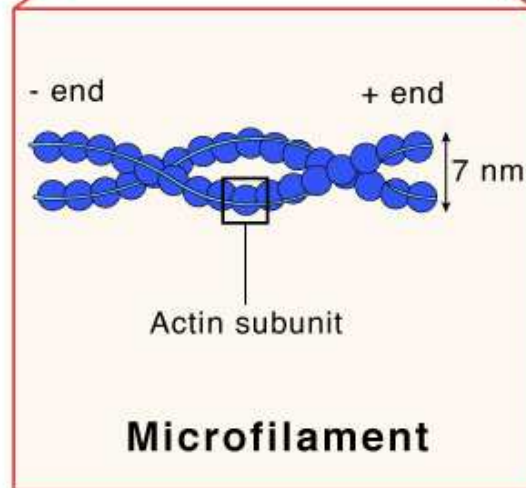
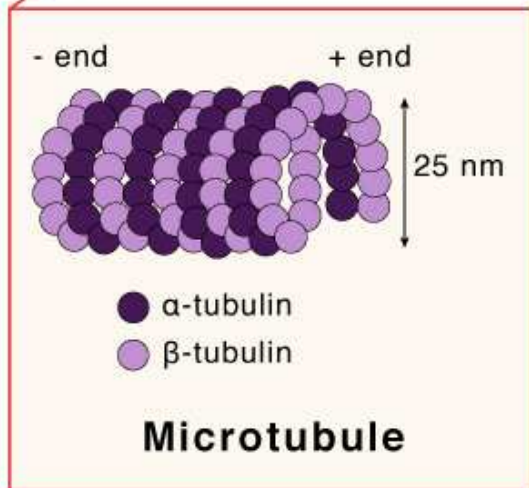
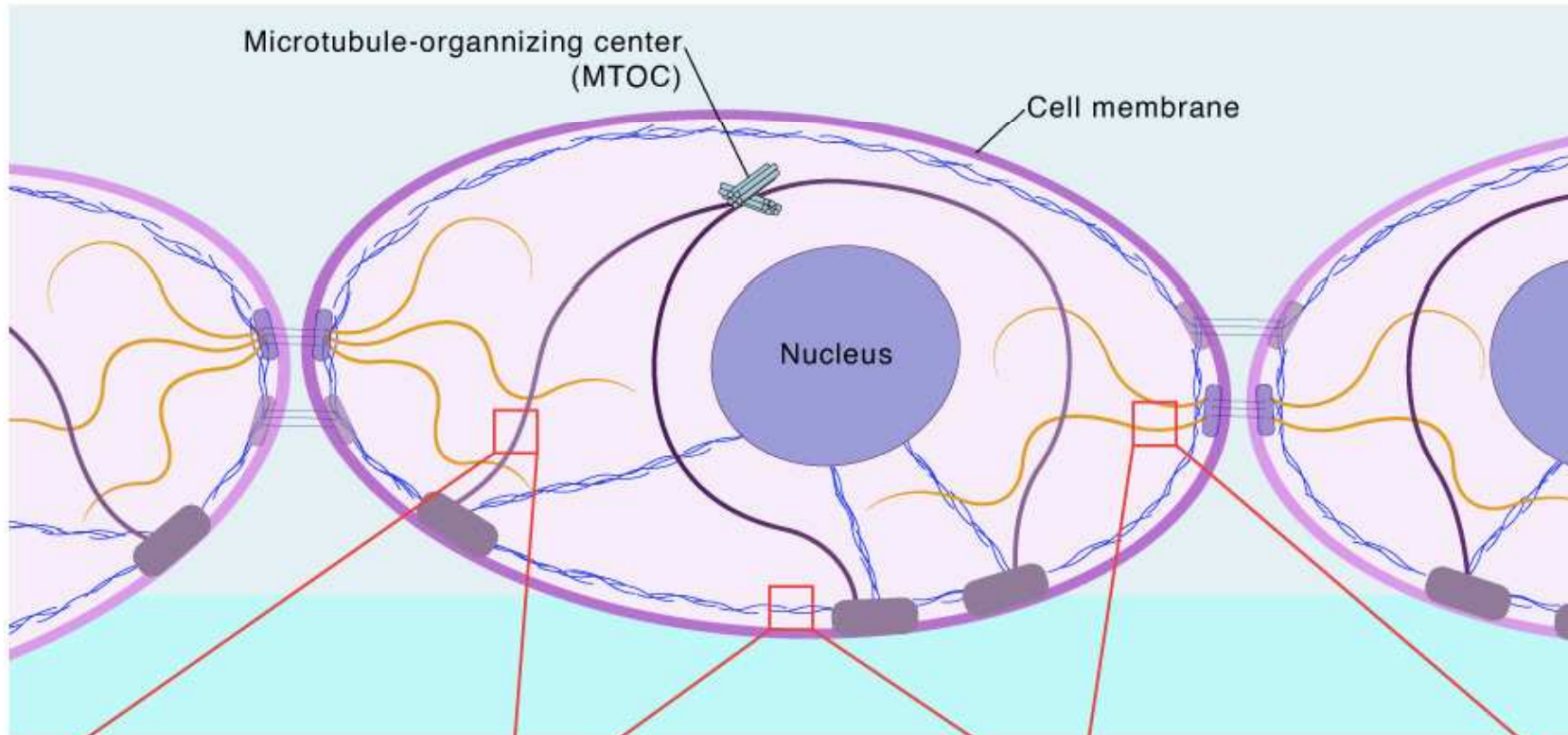
autumn 2024

## Oocyte spindle

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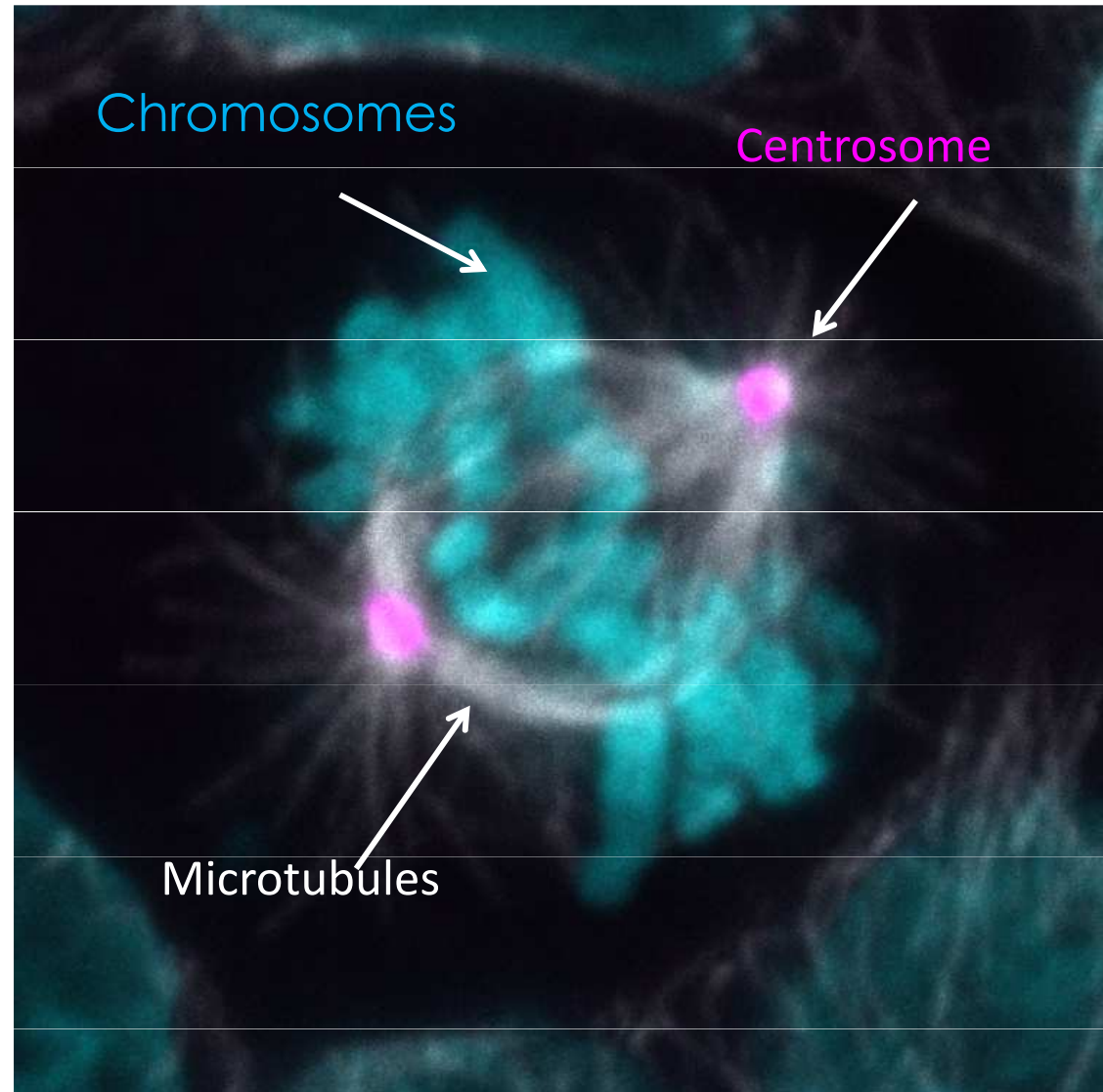
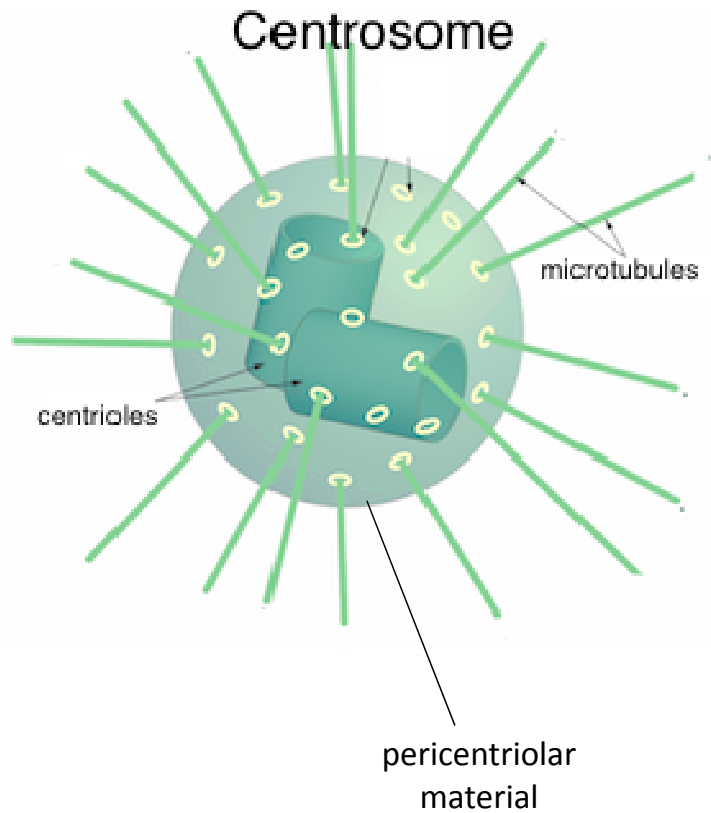


# Cytoskeleton



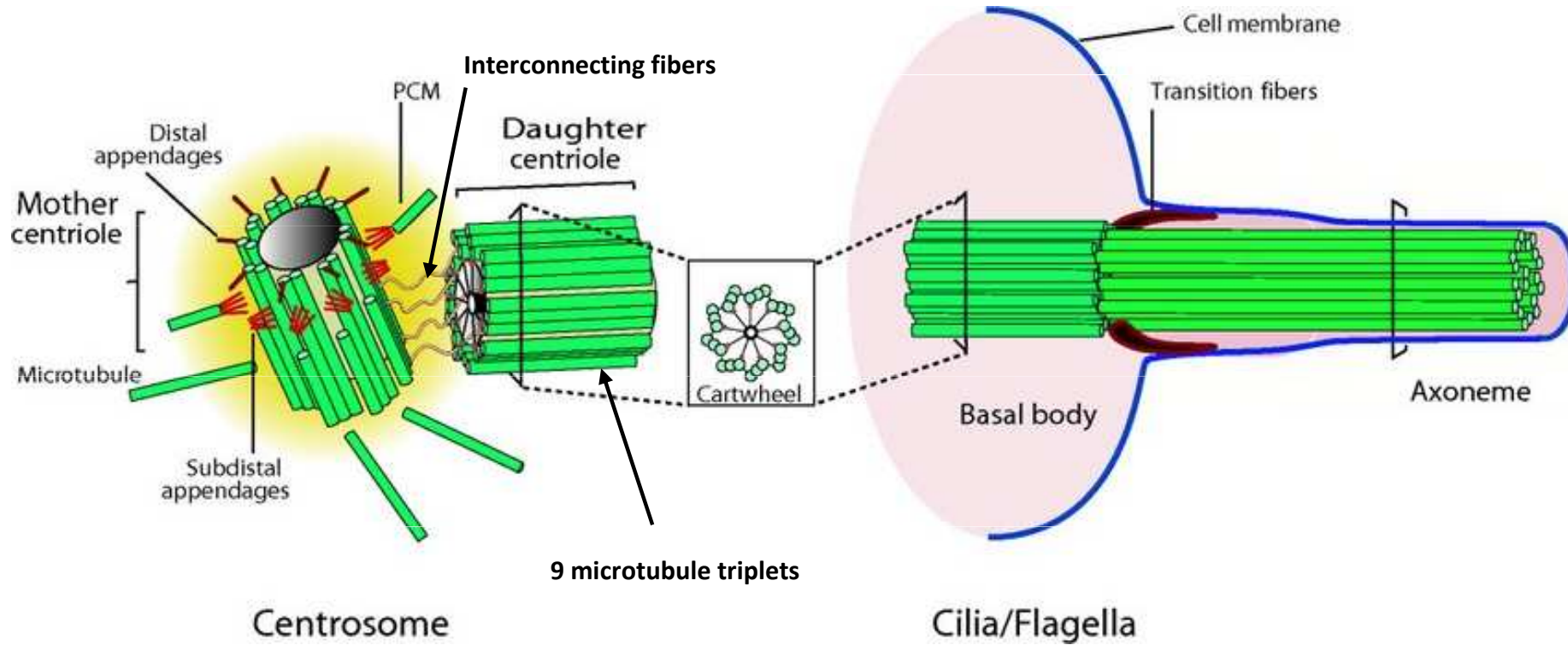
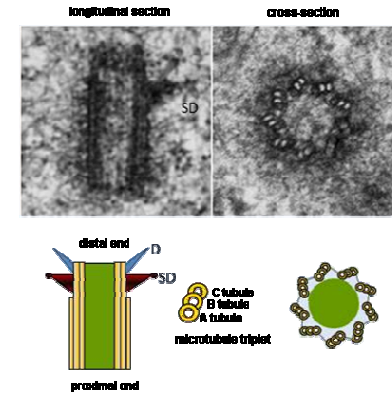
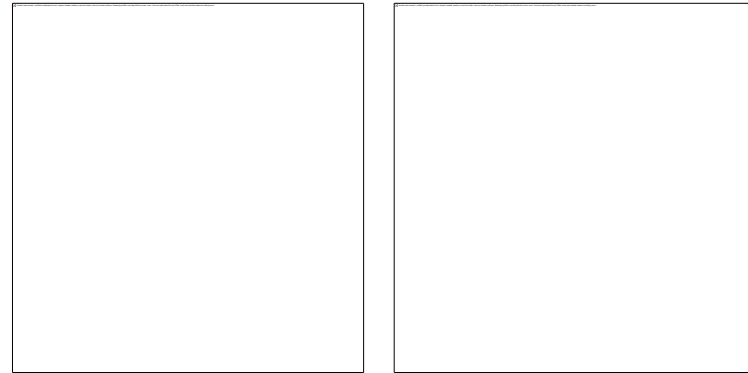
# Centrosome

**Centrosome = major Microtubule Organising Center (MTOC) in animal cells**



# Centrosome

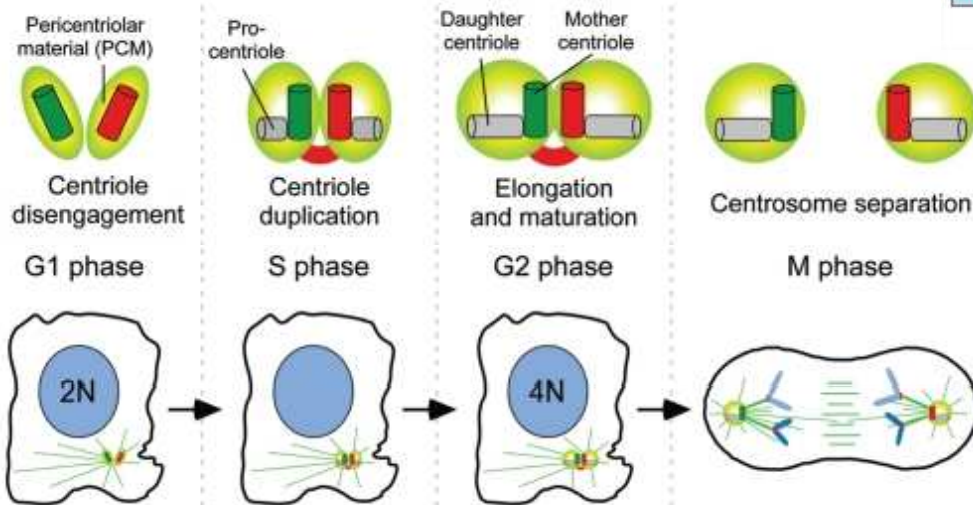
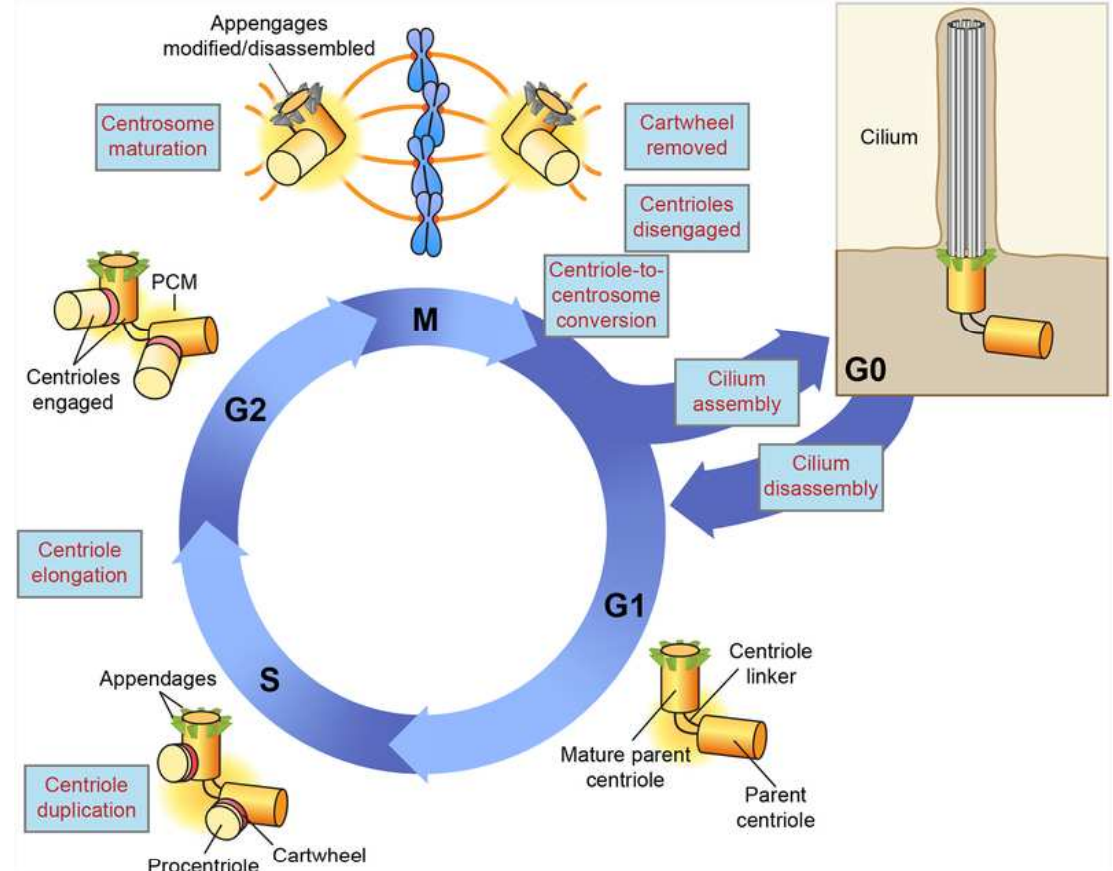
## ➤ Centrioles





# Centrosome

## ➤ Centrosome duplication cycle

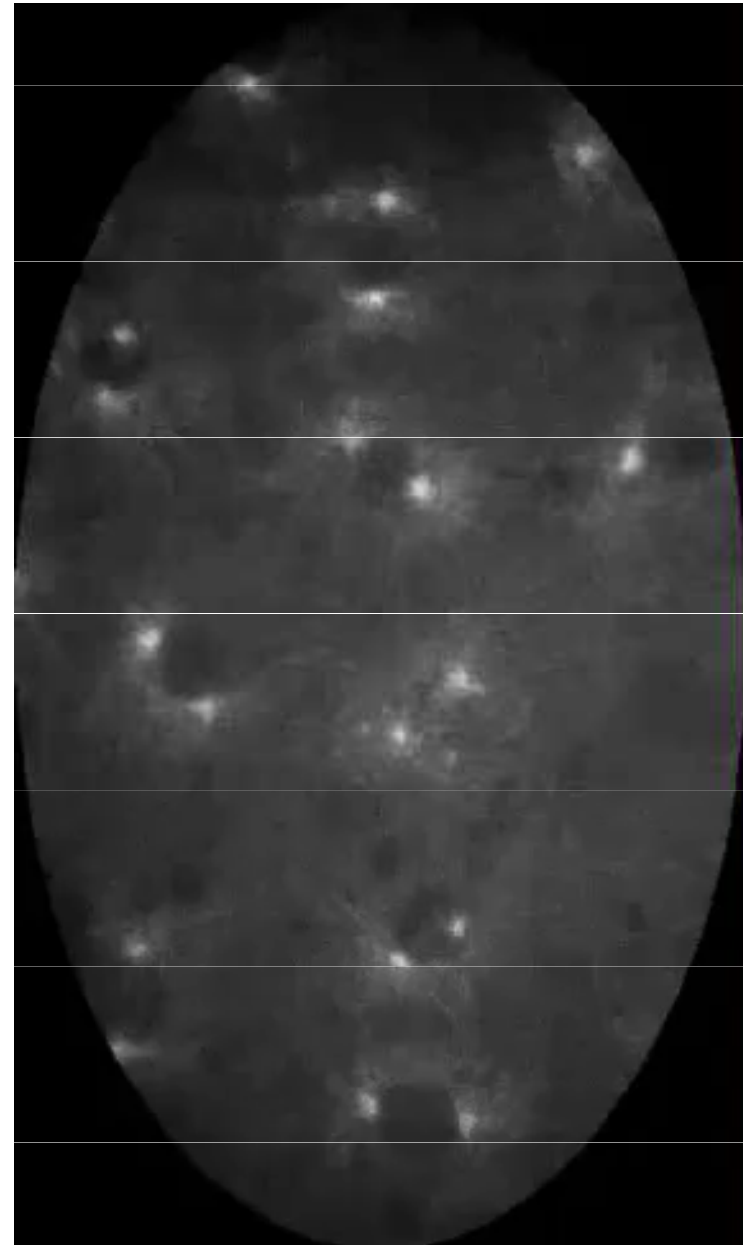
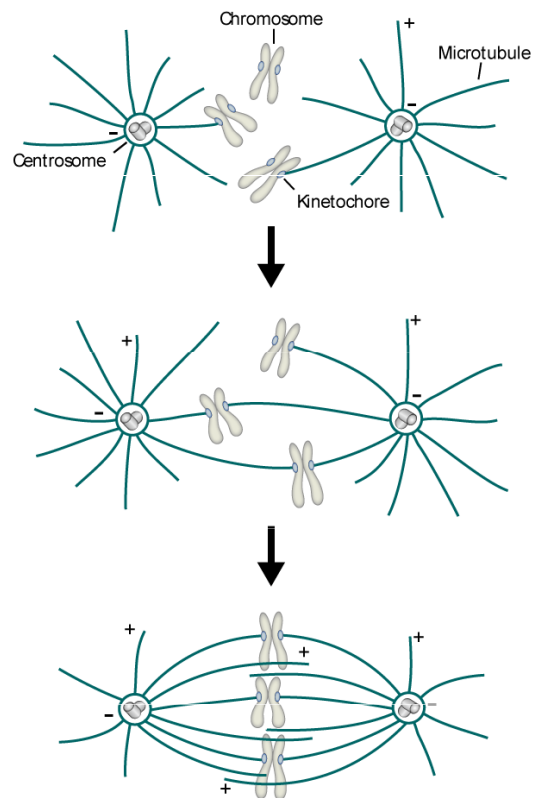


**Centrosomes duplicate in coordination with DNA synthesis**

# Centrosome

In animal somatic cells, centrosomes

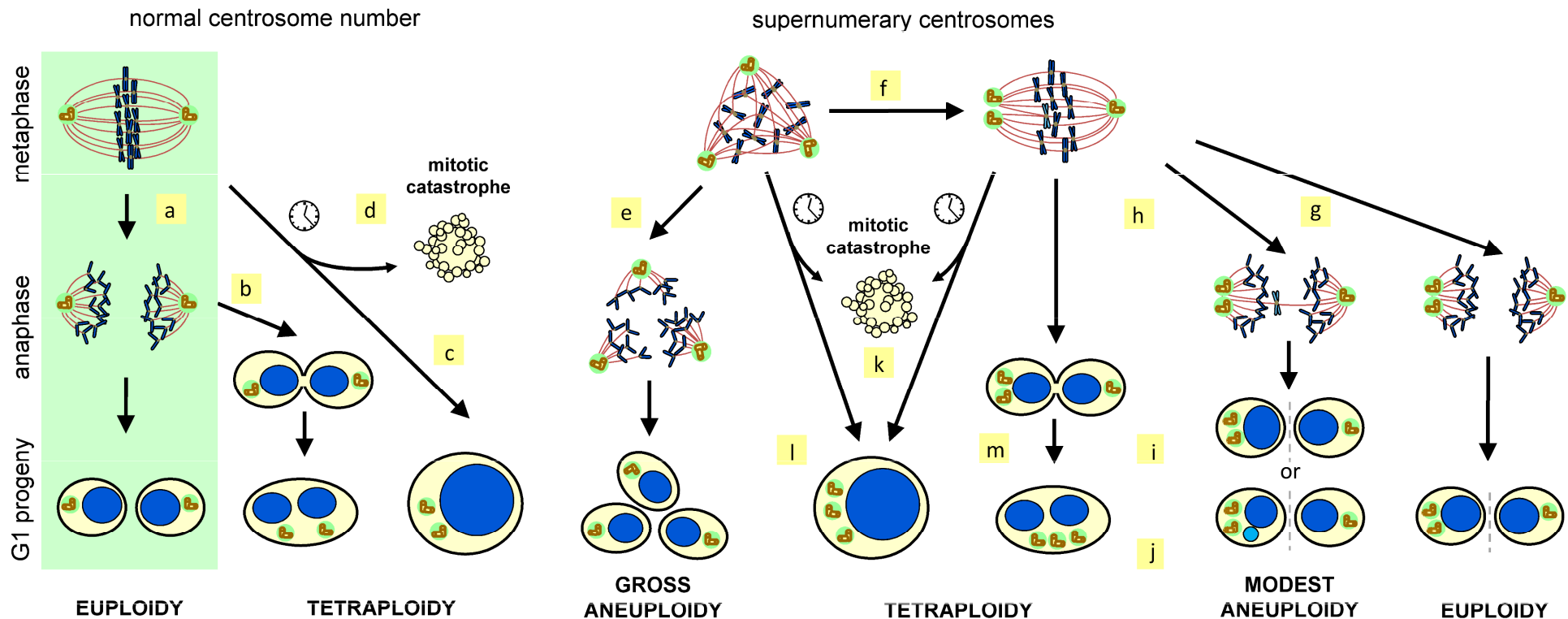
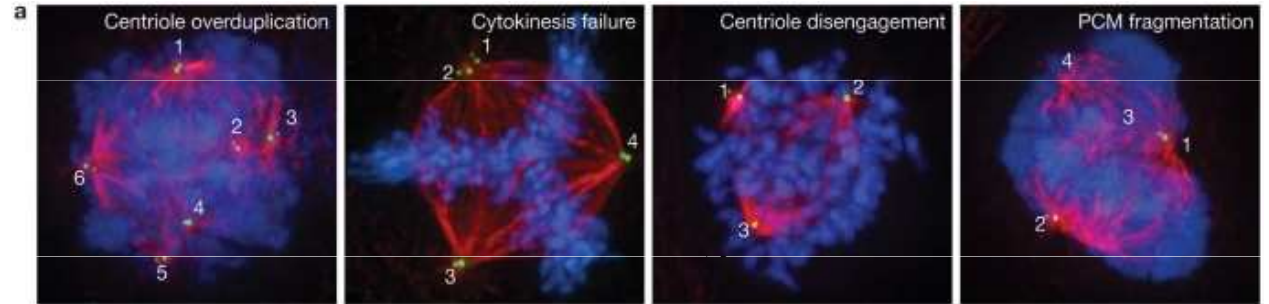
- drive microtubule (MT) nucleation
- focus microtubule (-)ends at spindle poles and stabilize spindle poles
- assemble central bipolar spindle that evenly segregate sister chromatids during mitosis





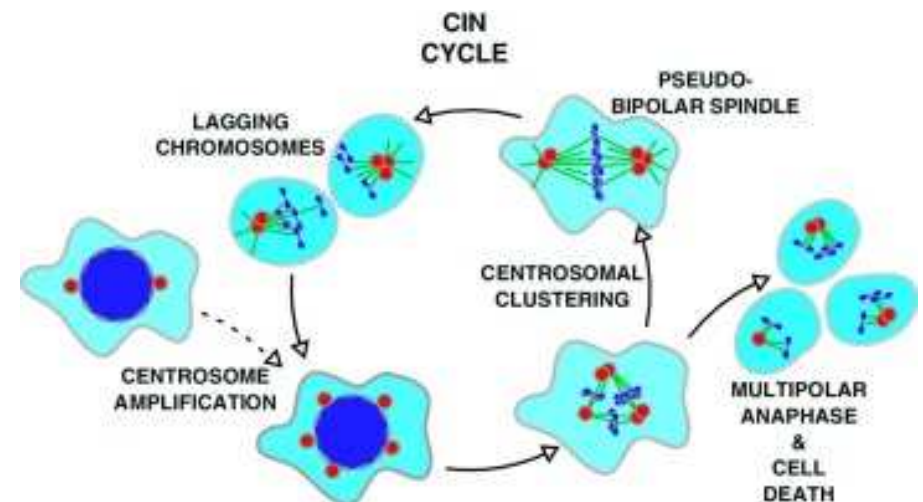
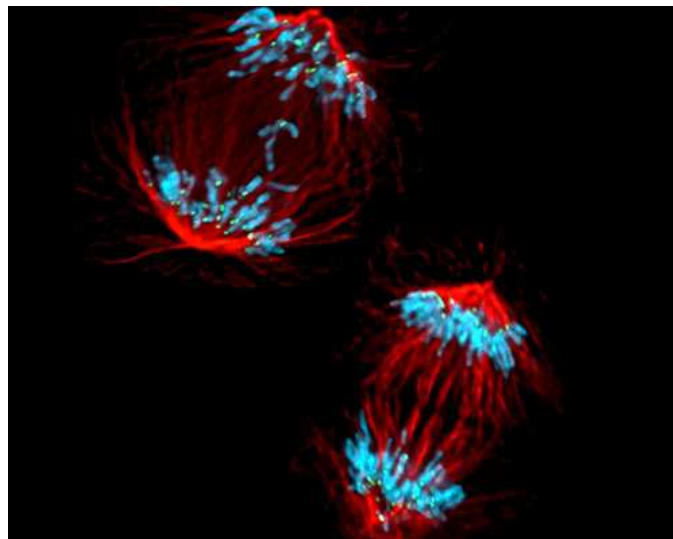
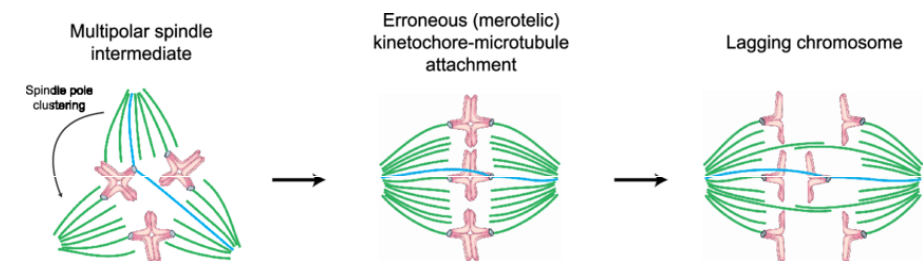
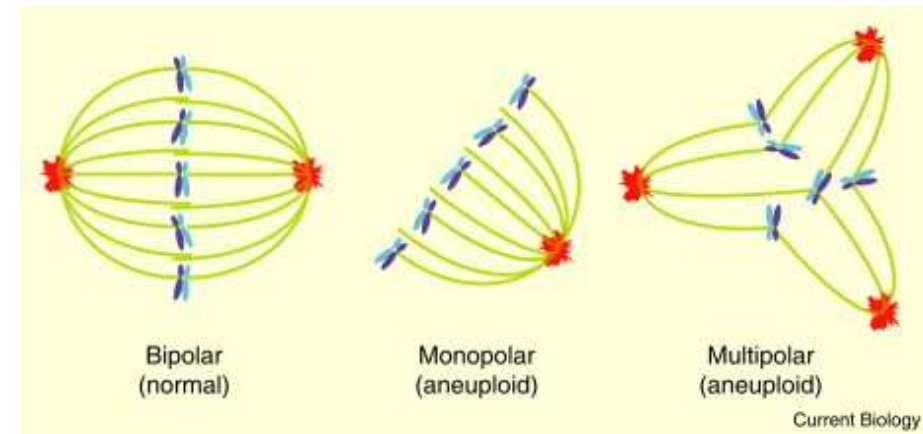
# Centrosome overamplification

- occurs in cancer cells
- promotes genetic instability
- acentriolar centrosomes (only PCM) capable to nucleate and capture microtubules



# Centrosomes define spindle geometry

- overamplification of centrosomes generates multipolar spindle which produces gross aneuploidy
- clustering of centrosomes enables bipolarization but persisting merotelic attachments favour chromosome lagging during anaphase and create risk of chromosome missegregation





# Microtubule nucleation pathways

## Centrosomes

present



absent



present



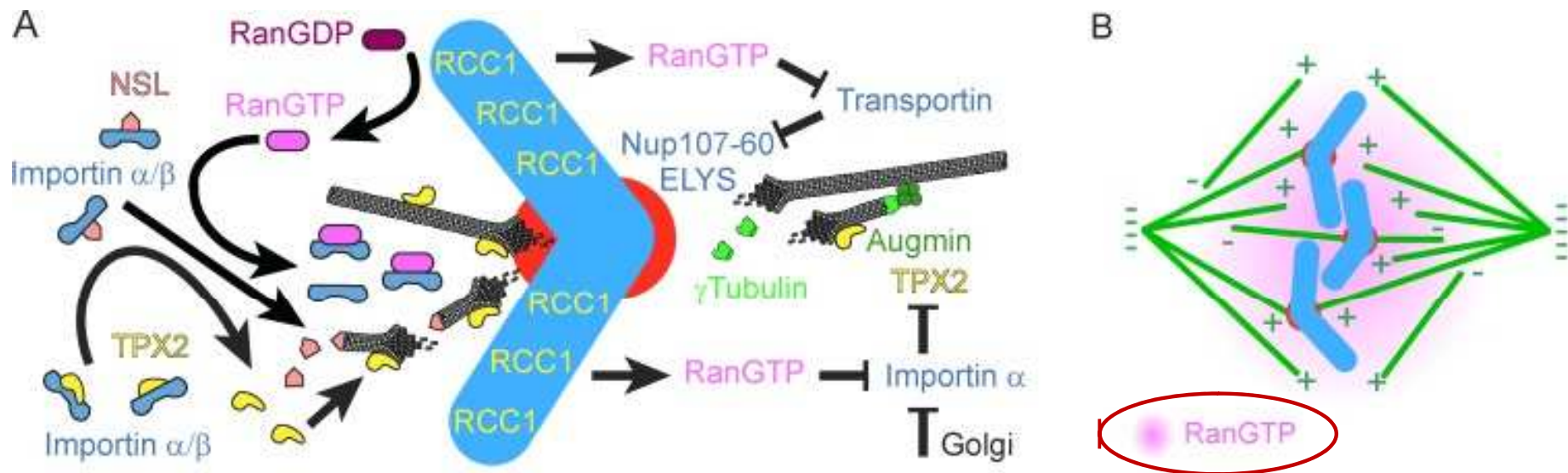
- plant cells

- animal mitotic cells

# Microtubule nucleation pathways

## ❖ Chromatin-driven microtubule (MT) nucleation

- **RanGTP** gradient promotes both de novo MT nucleation near kinetochores and amplification of MT growth toward chromosomes

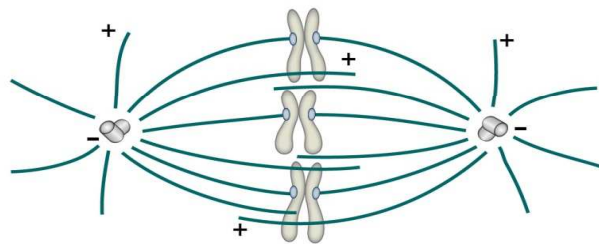
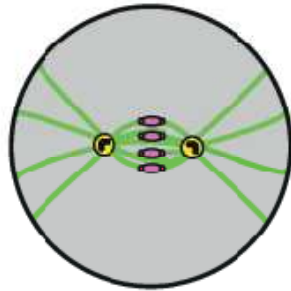




# Female meiotic spindles lack centrosomes

## Mitosis

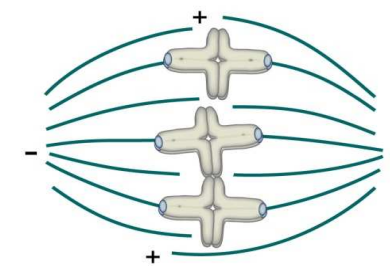
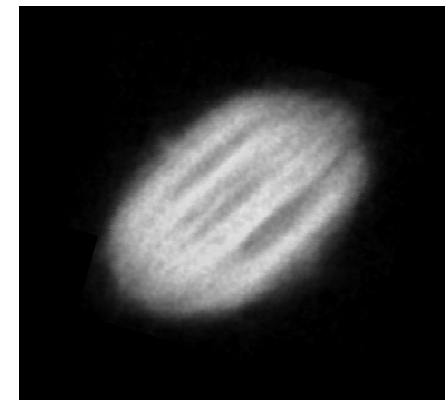
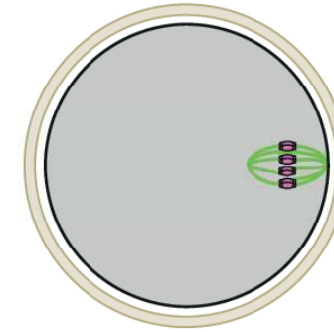
Symmetric division



Centrosomal spindle

## Female meiosis

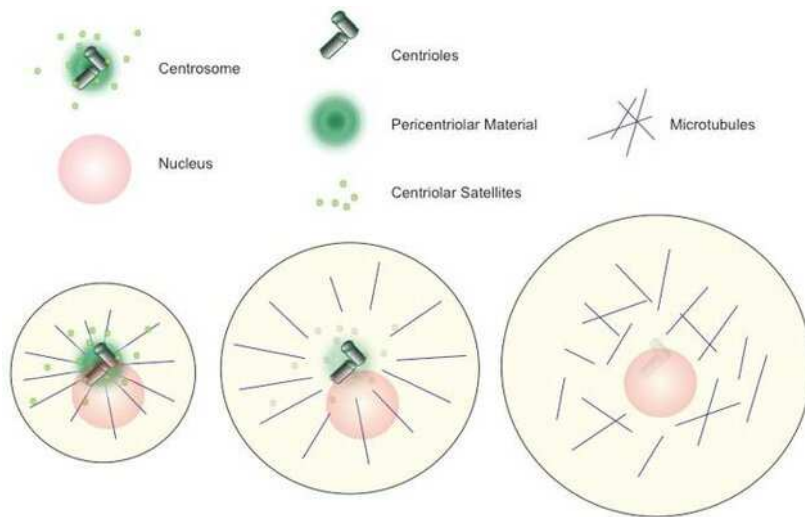
Asymmetric division



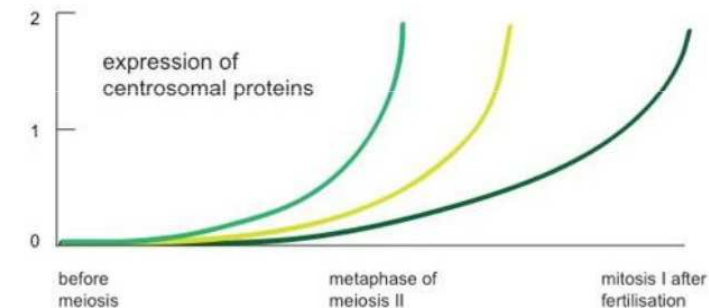
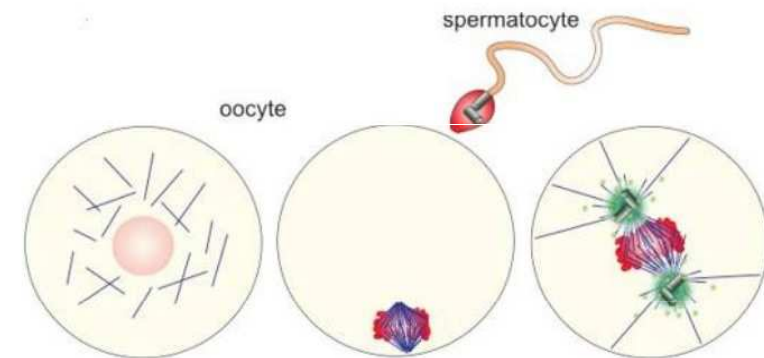
Acentrosomal spindle

# Centrioles are eliminated during oogenesis

- Metazoan oocytes eliminate centrosomes during oogenesis in order to
- (1) ensure highly asymmetric cell division
  - (2) avoid a superior number after fertilisation



- PCM synthesized during oocyte maturation
- Centrioles paternally inherited



- sperm-derived centrioles recruit maternal PCM after fertilization to assemble first mitotic spindle

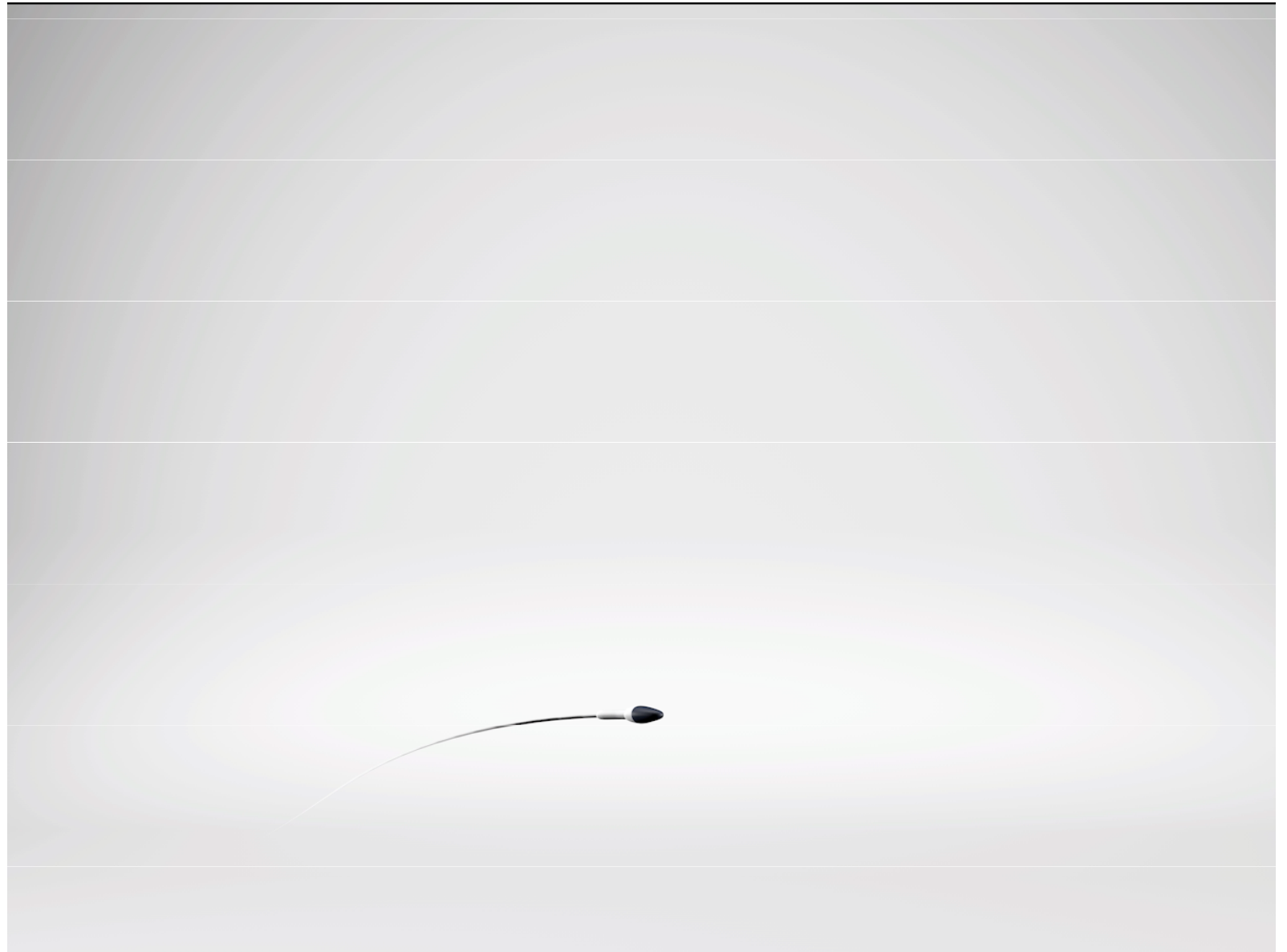
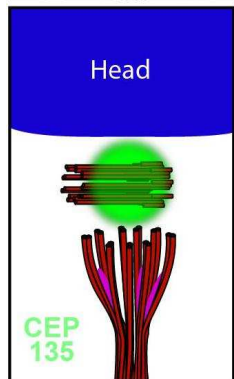
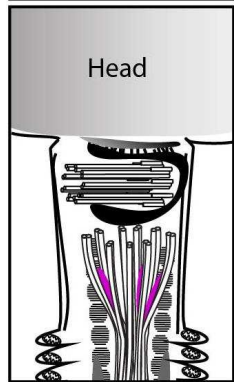
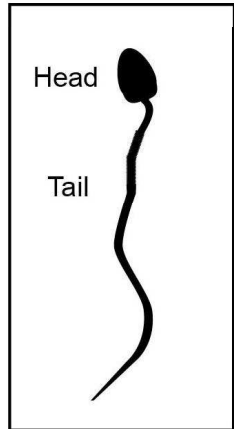


- sperm-derived centrioles are destroyed
- first mitosis with acentrosomal spindle
- de novo centriole assembly during embryo cleavage stage



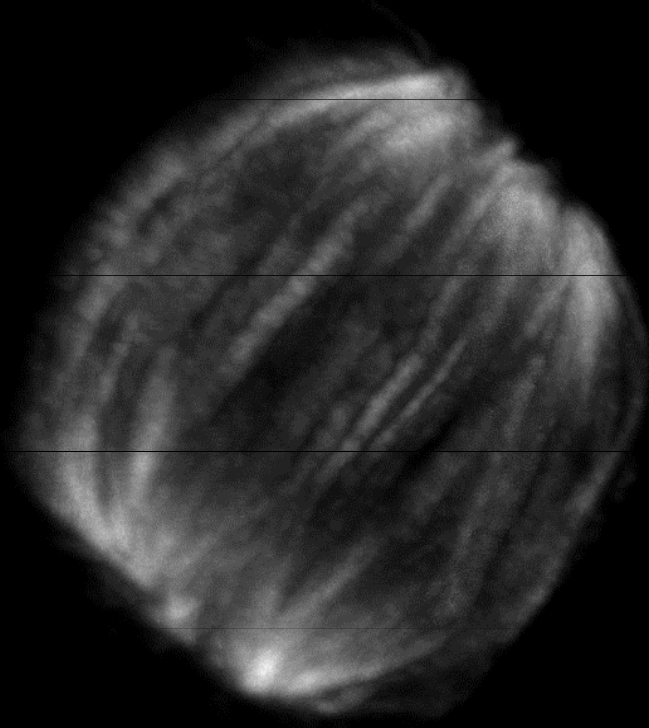
# Centrioles are delivered by sperm during fertilization

Normal sperm

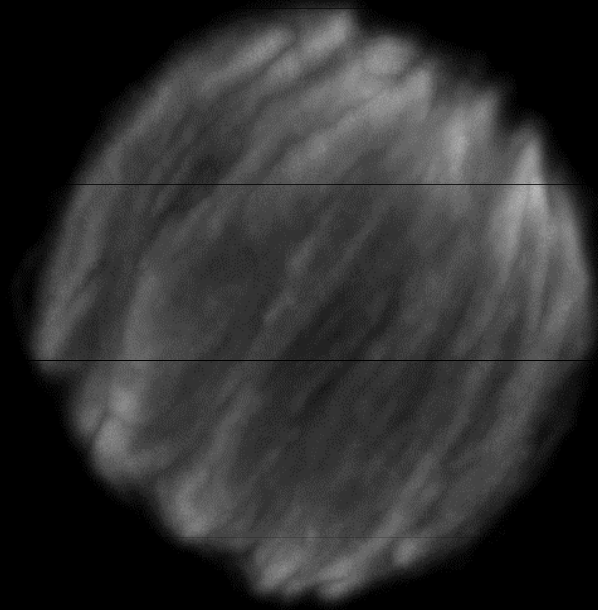


# Human oocyte spindle lacks centrosome

Metaphase I



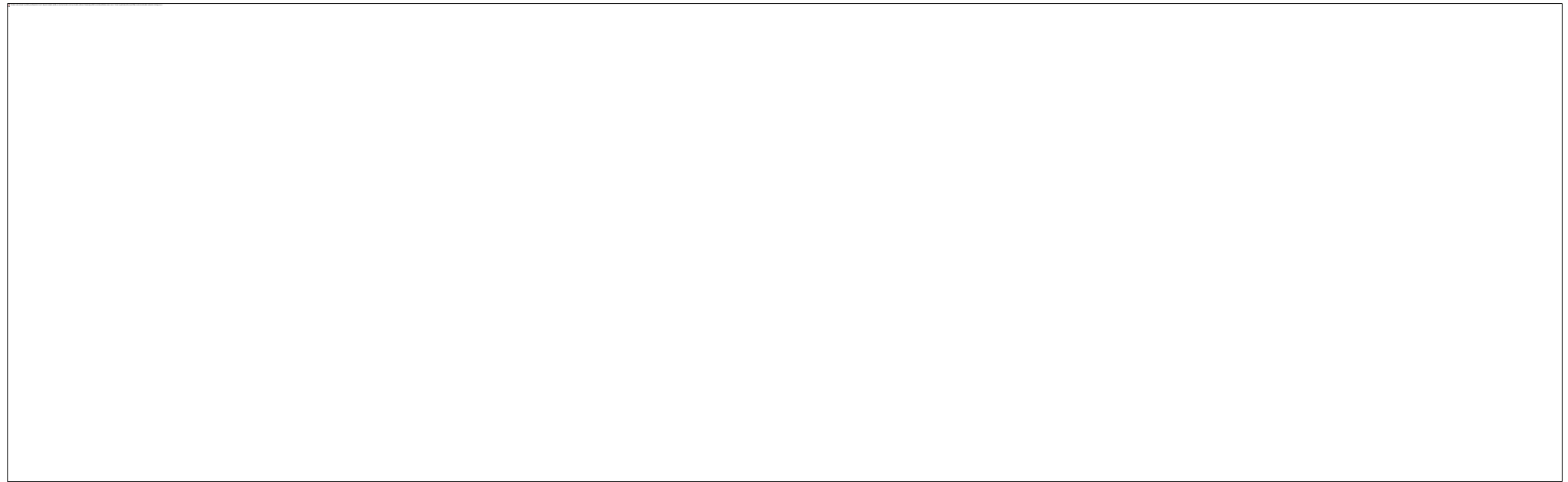
Metaphase II



5  $\mu$ m

How are meiotic spindle poles assembled  
in the absence of centrosomes?

# Acentrosomal spindle drives chromosomal segregation during female meiosis



GERMINAL  
VESICLE (GV)

METAPHASE I  
(MI)

ANAPHASE I

METAPHASE II  
(MII)

ANAPHASE II

**Functional spindle is required for chromosome segregation fidelity**

errors in 1st meiotic division



egg aneuploidy

errors in 2nd meiotic division



embryo aneuploidy



# Acentrosomal spindle assembly in mouse oocytes

- high-resolution confocal live cell imaging of mouse oocytes maturing in vitro showed that mouse oocyte spindle is assembled by **multiple small acentriolar MTOCs that functionally replace canonical centrosomes**

Cell

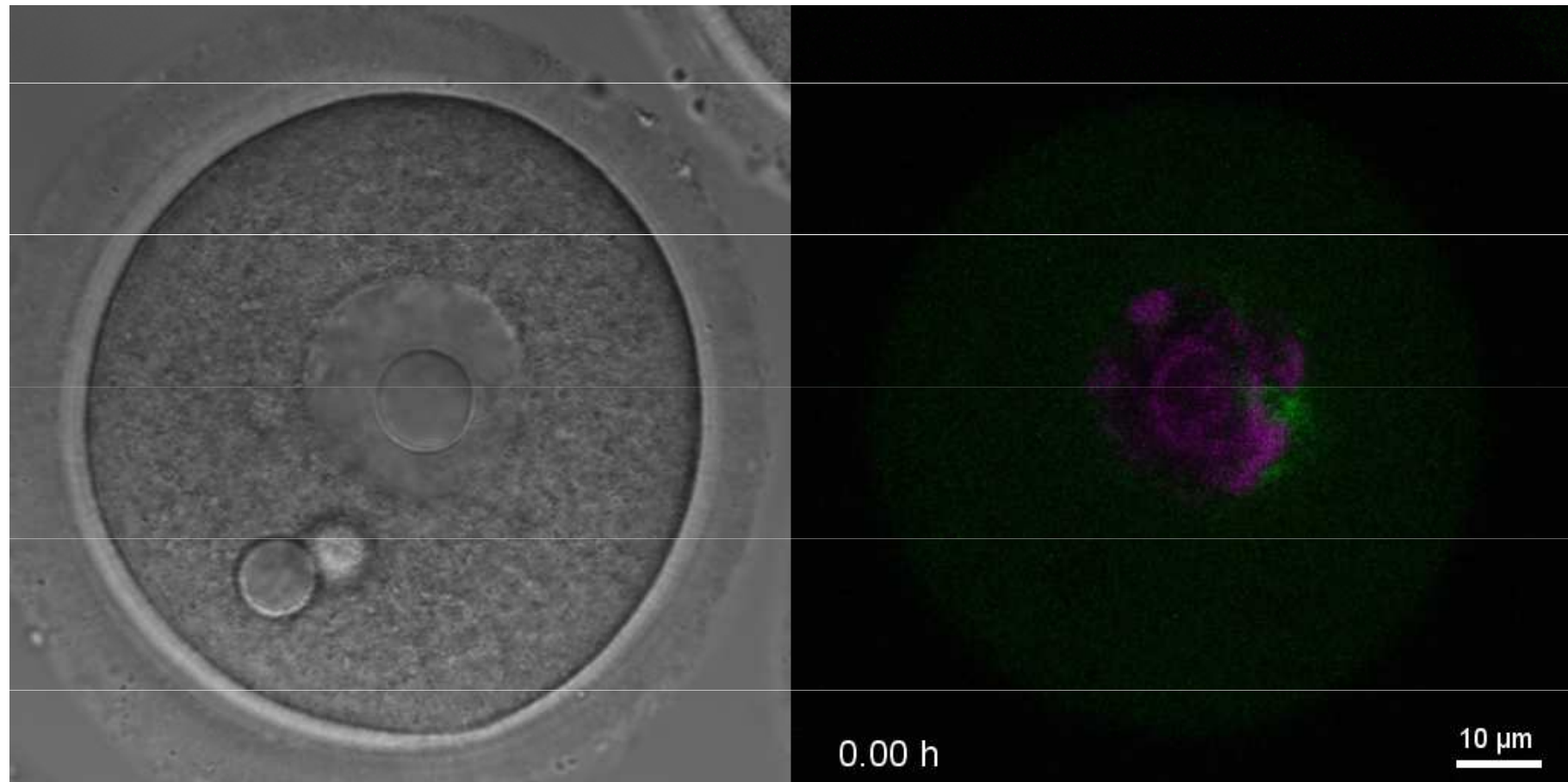
Schuh and Ellenberg, 2007

## Self-Organization of MTOCs Replaces Centrosome Function during Acentrosomal Spindle Assembly in Live Mouse Oocytes

Melina Schuh<sup>1</sup> and Jan Ellenberg<sup>1,\*</sup>  
<sup>1</sup>Gene Expression Unit, European Molecular Biology Laboratory (EMBL), Meyerhofstrasse 1, D-69117 Heidelberg, Germany  
\*Correspondence: jan.ellenberg@embl.de  
DOI 10.1016/j.cell.2007.06.025



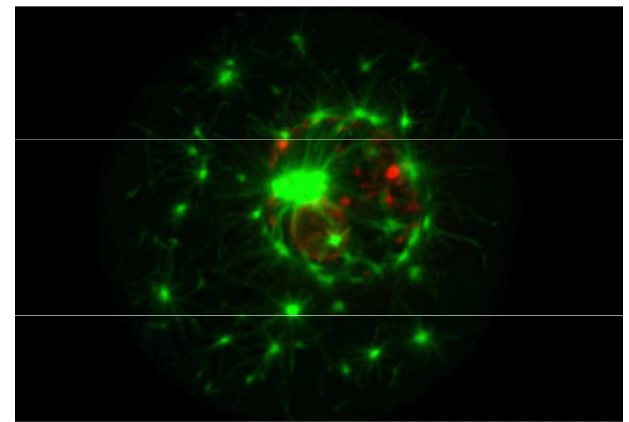
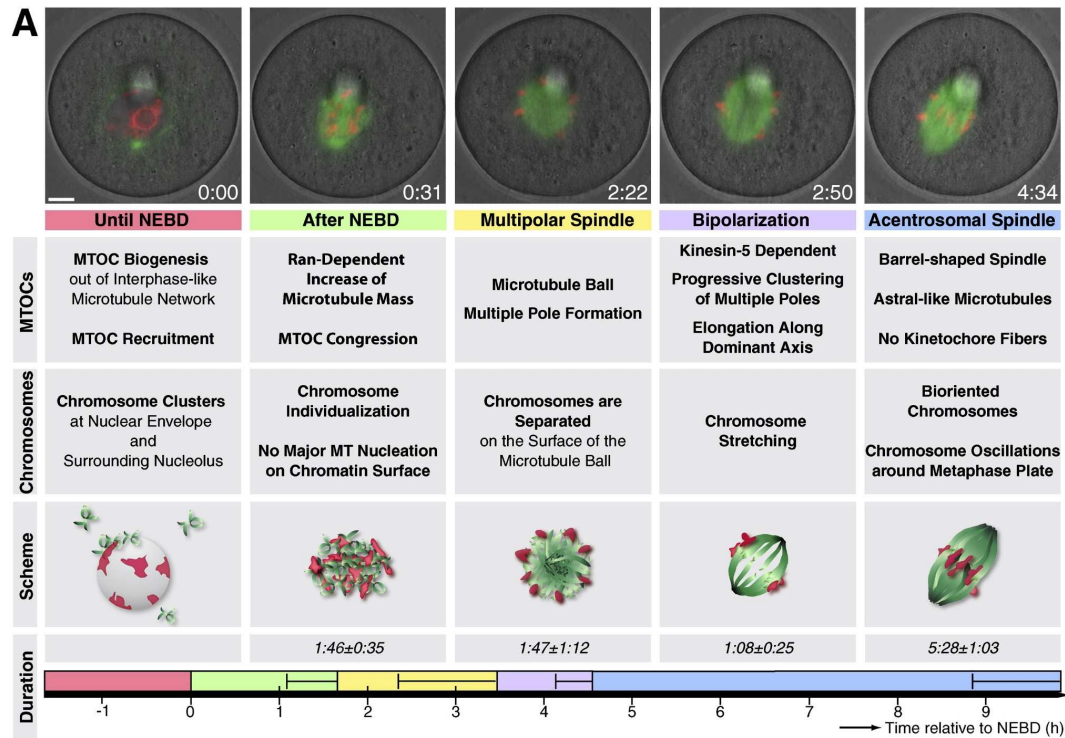
Melina Schuh



DNA

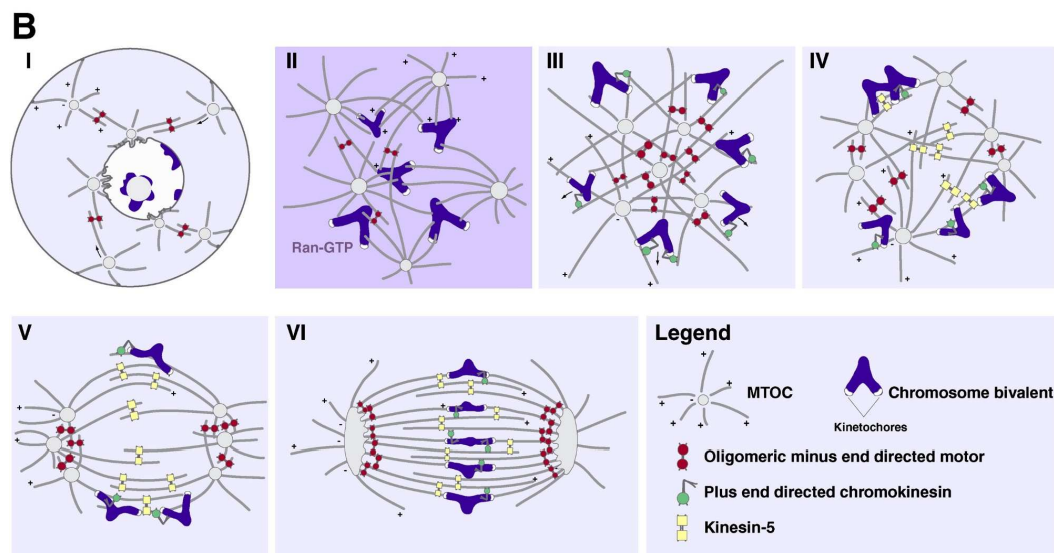
microtubules

# Acentrosomal spindle assembly in mouse oocytes

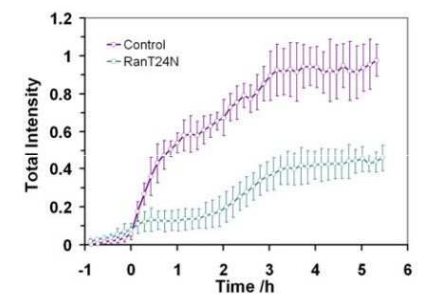


Prophase microtubule network with low dynamics

- MTOC consists of PCM proteins (pericentrin,  $\gamma$ -tubulin, Cep192, Cep120, Cep 125, NEDD1,..)
- MTOCS cluster around nucleus before NEBD
- MTOC nucleate MT „ball“ which carries chromosomes on its surface
- MT mass elongates and chromosome congress
- chromosome alignment after spindle bipolarization
- spindle migration to the cortex

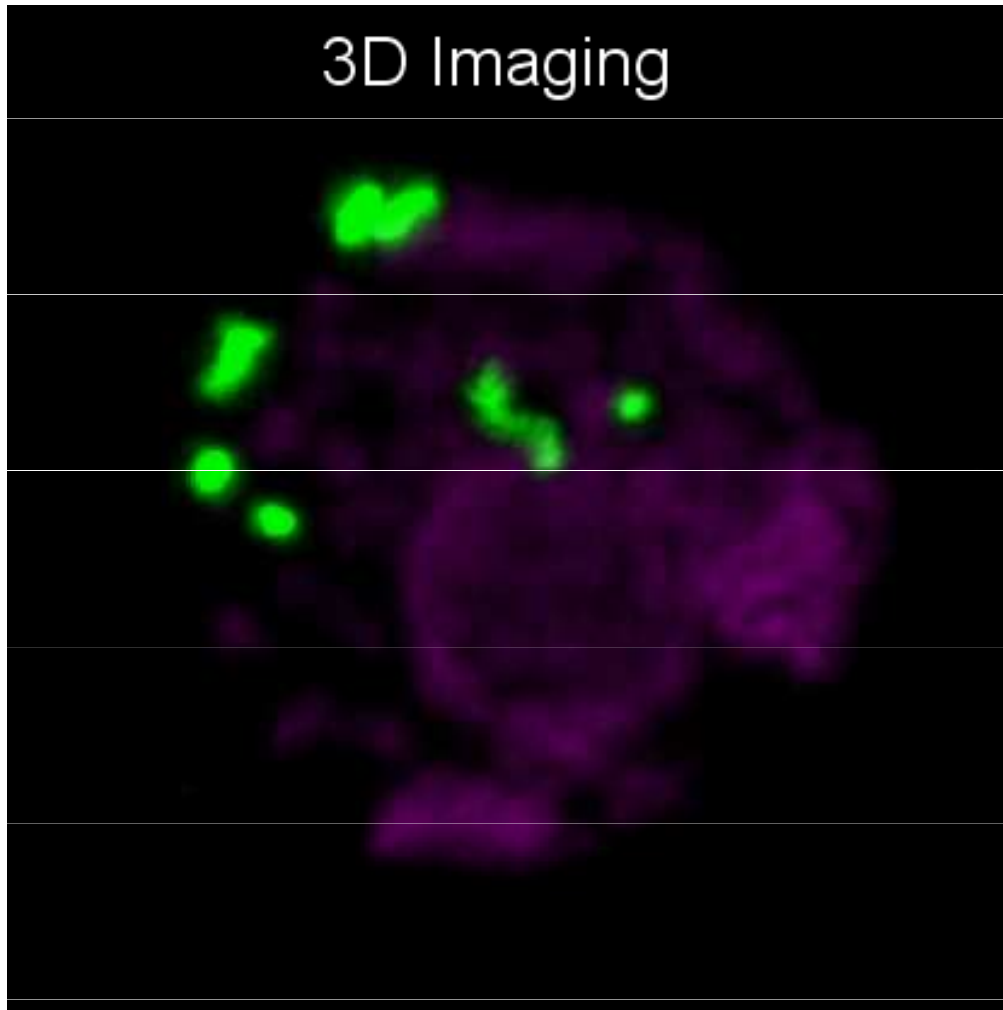


- Ran activity overdriven by coordinated action of MTOCS



# Acentrosomal spindle assembly in mouse oocytes

Multipleacentriolar MTOCs converge at spindle poles and stabilize th



### MTOC 3D reconstruction

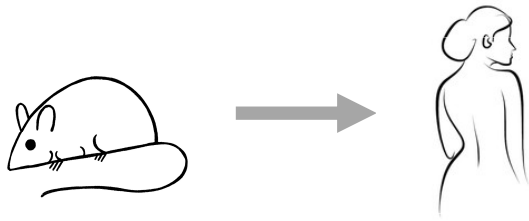


-01:00

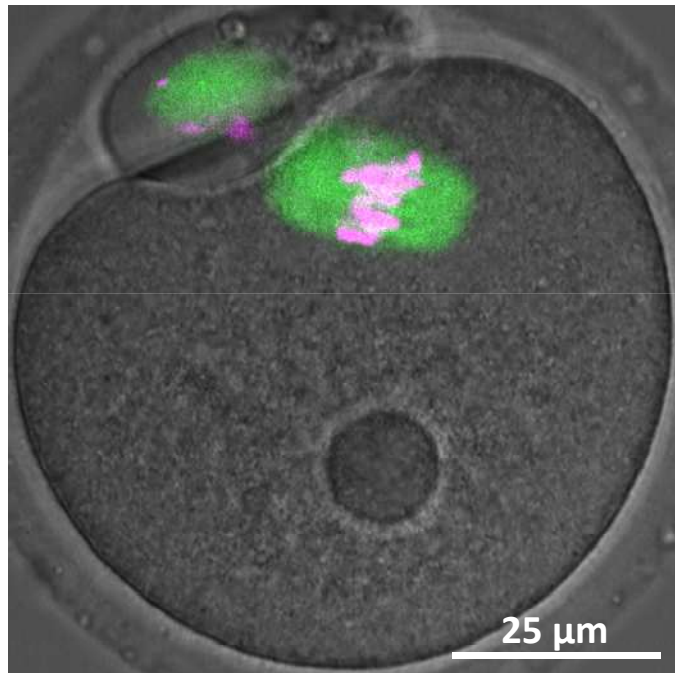
DNA  
MTOCs



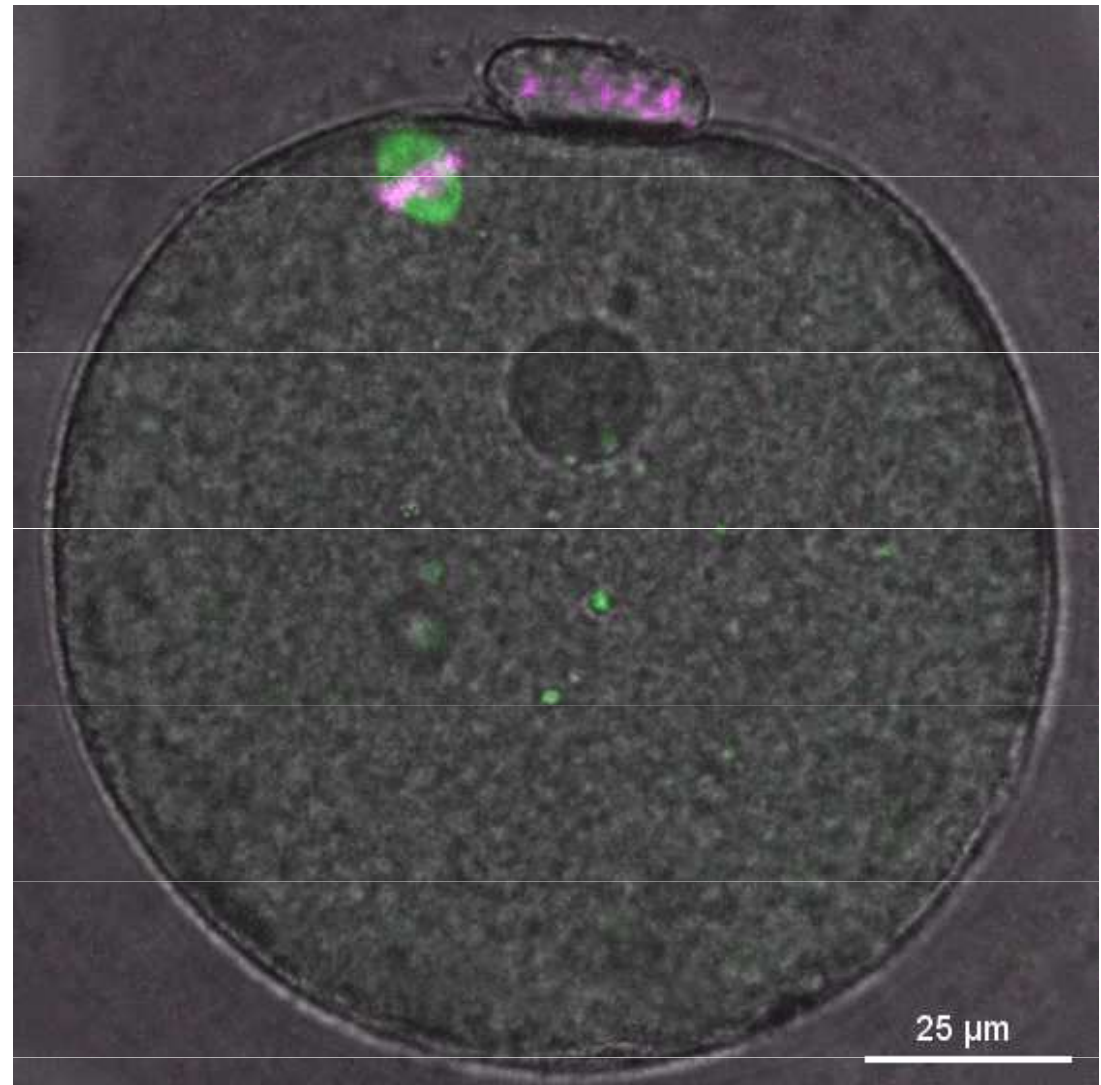
# From mice to human



**Mouse oocyte**



**Human oocyte**



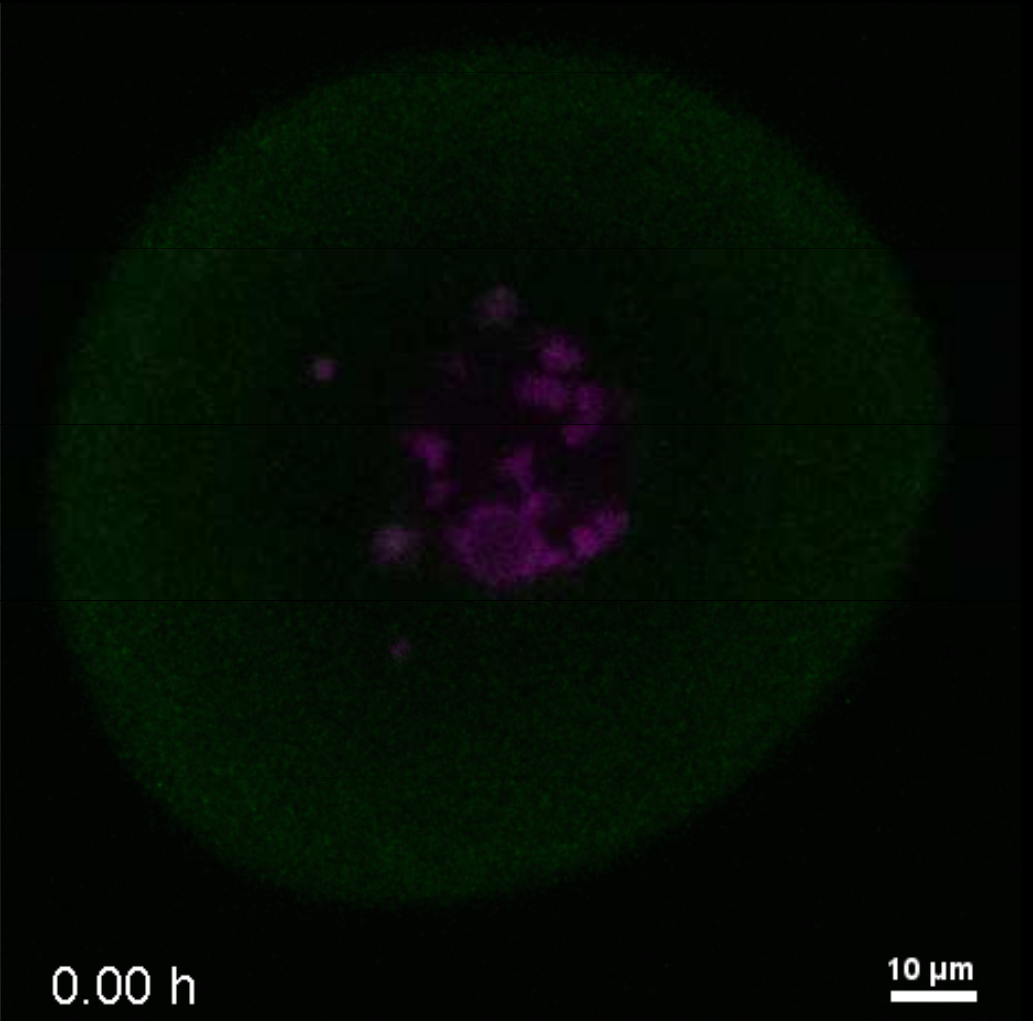
Chromosomes  
Microtubules

# Acentrosomal spindle assembly in human oocytes



DIC (transmitted light)

*Holubcova et al. Science 2015.*



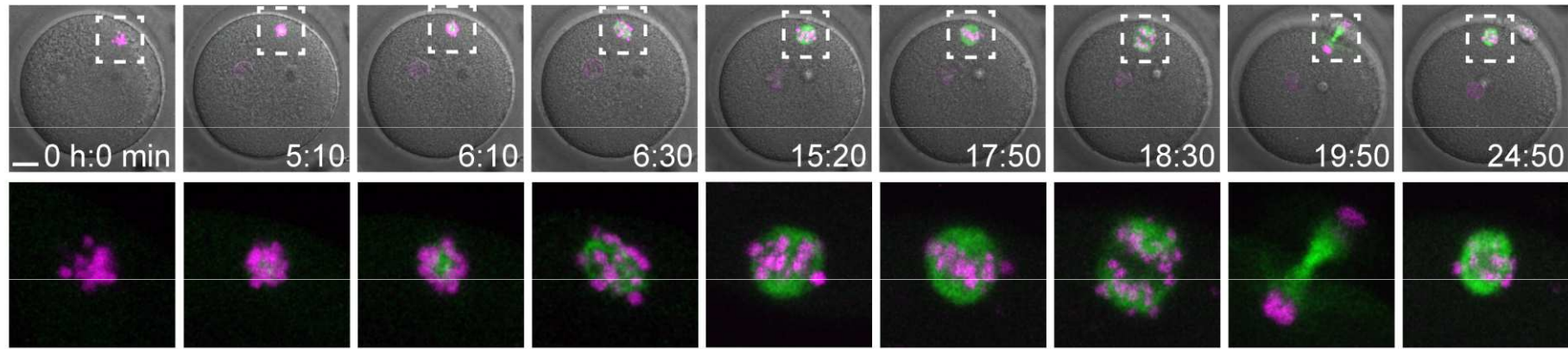
0.00 h

10 μm

Chromosomes (H2B-mRFP)

Microtubules (MAP4-EGFP)

# Acentrosomal spindle assembly in human oocytes



Nuclear envelope breakdown



Onset of microtubule nucleation



Growing microtubule aster



Early bipolar spindle



Initial chromosome congression



Stable chromosome alignment



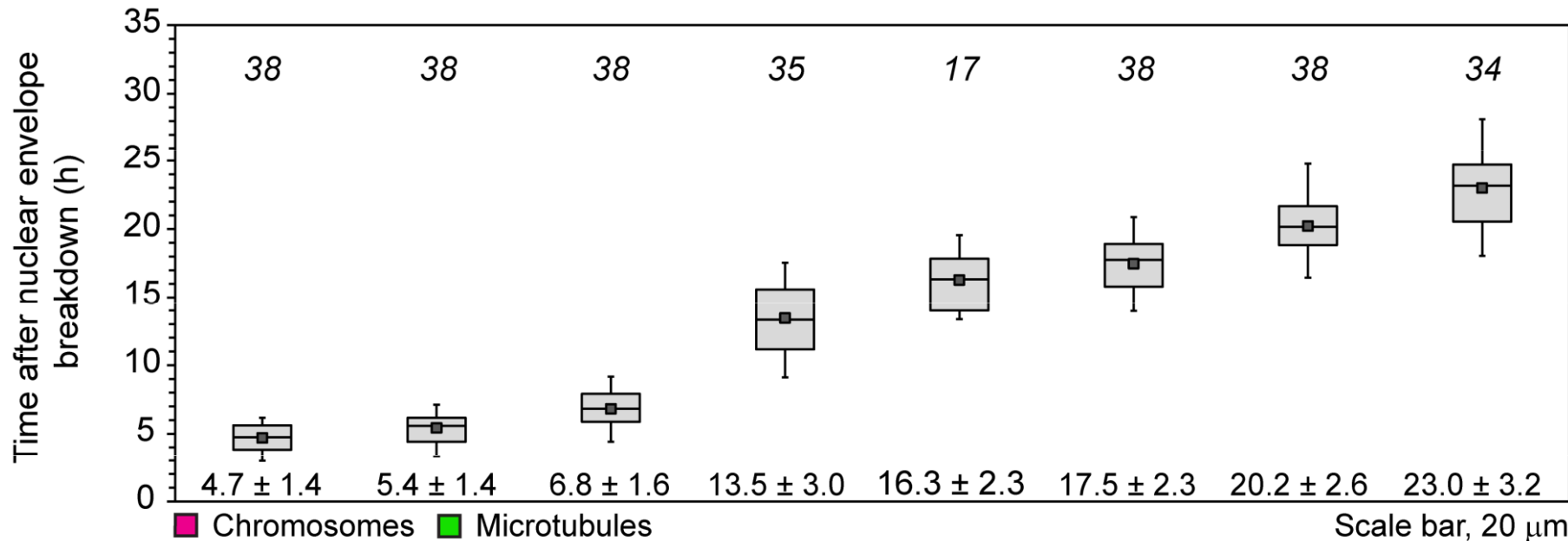
Anaphase



Polar body abscission



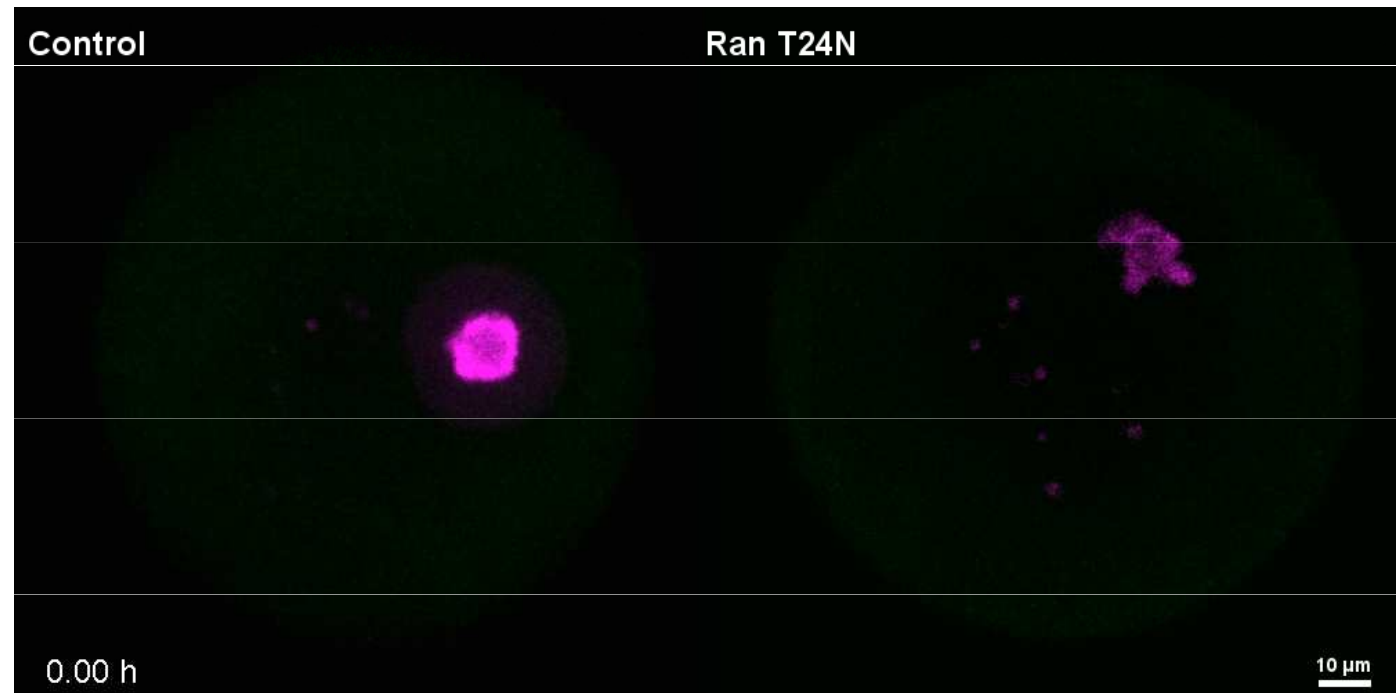
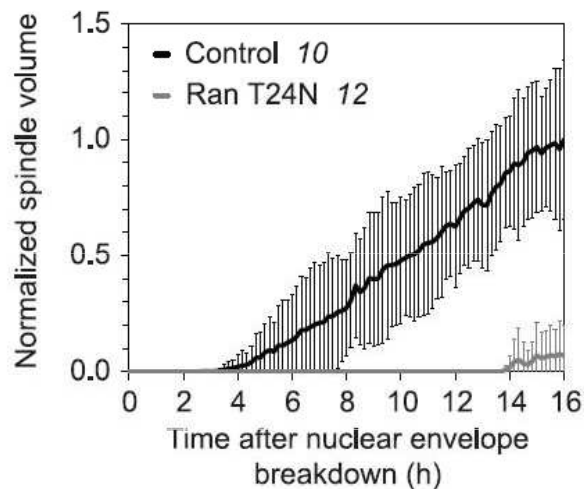
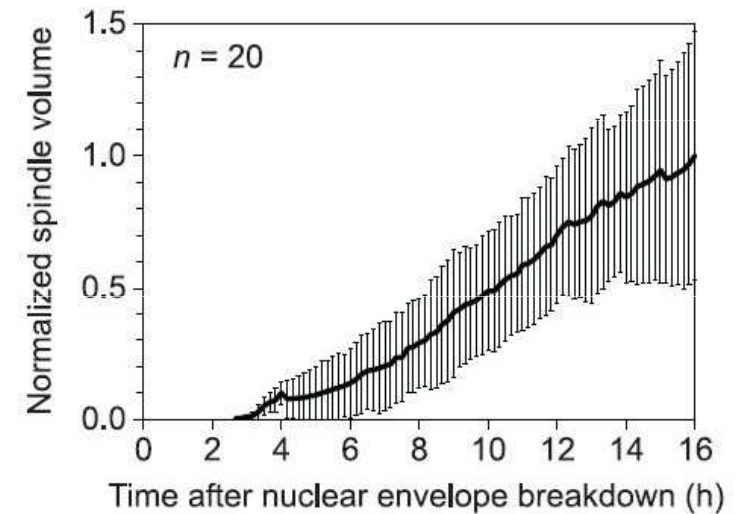
Bipolar MII spindle





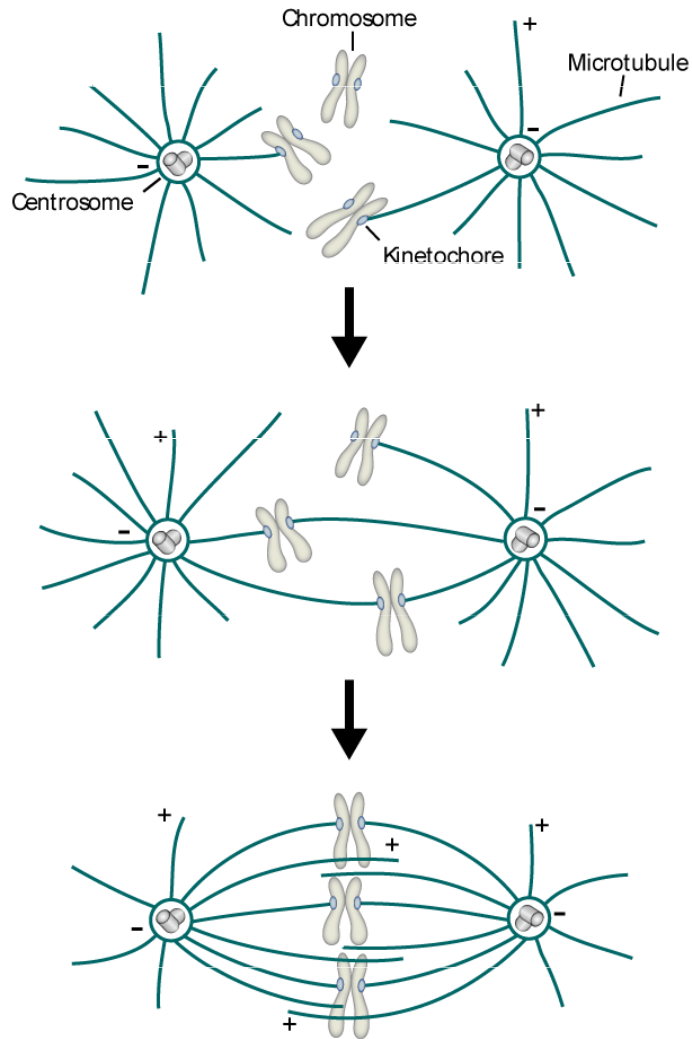
# Acentrosomal spindle assembly in human oocytes

- human oocytes assemble a meiotic spindle independently of either centrosomes or other MTOCs
- spindle assembly is mediated by chromosomes and the small guanosine triphosphatase **Ran**
- spindle assembly is unusually long, requiring ~16 hours

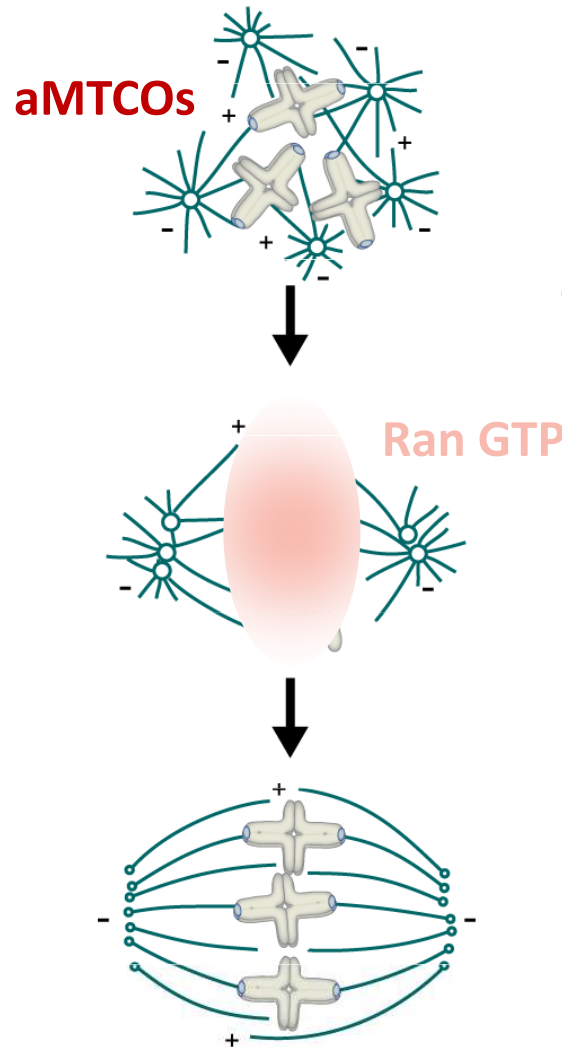


# Spindle assembly strategies

Centrosomal Spindle  
*Mitotic cells*

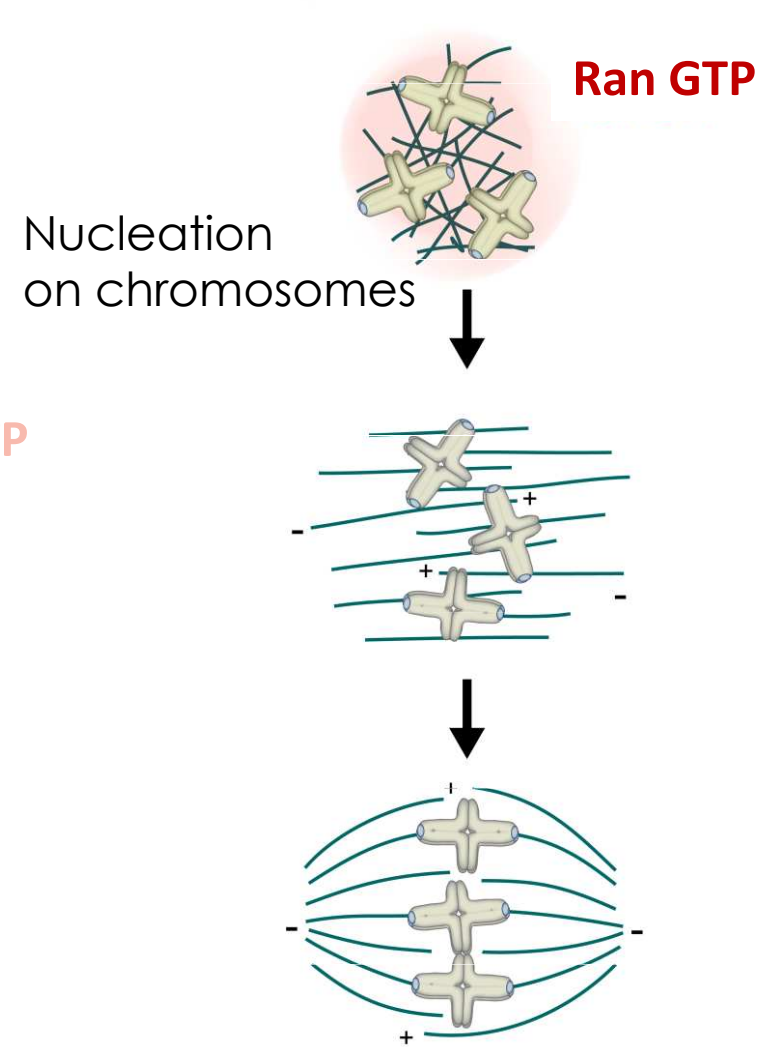


Acentrosomal Spindle  
*Mouse Oocyte*



Schuh and Ellenberg, Cell 2007.

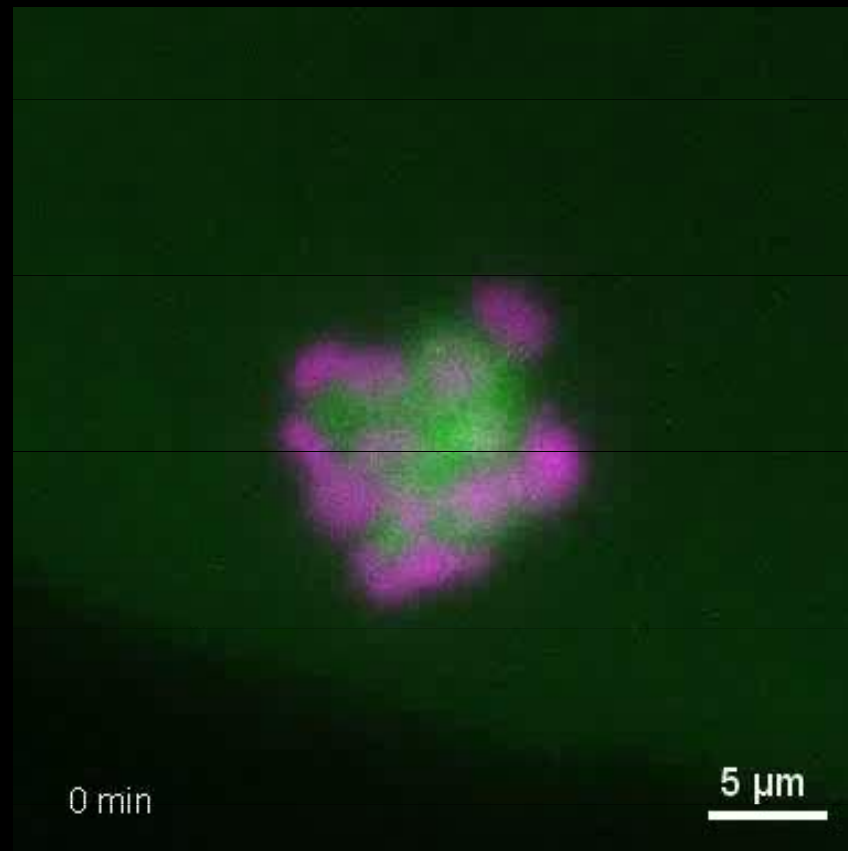
Acentrosomal Spindle  
*Human oocytes (and plant cells)*



Holubcova et al., Science 2015.

# Human oocyte spindle is unstable

## Moderate spindle instability

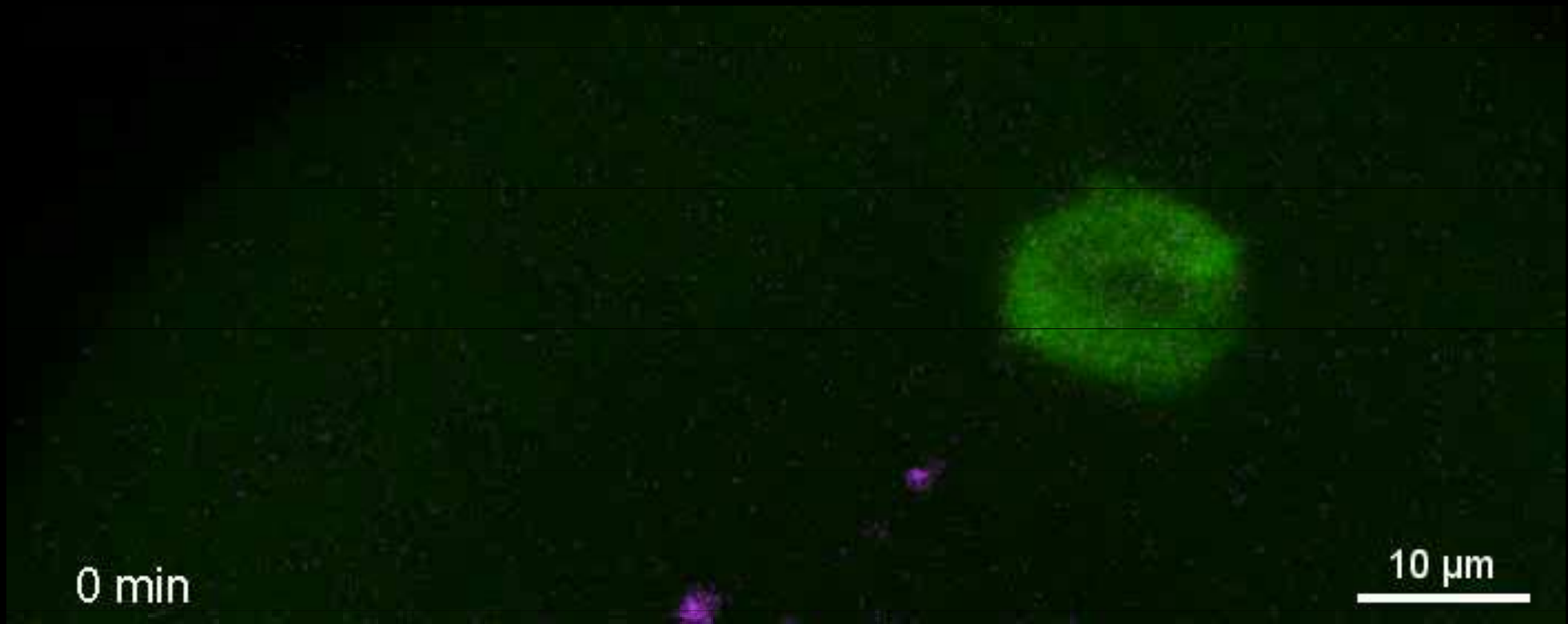


Chromosomes (H2B-mRFP)  
Microtubules (MAP4-EGFP)



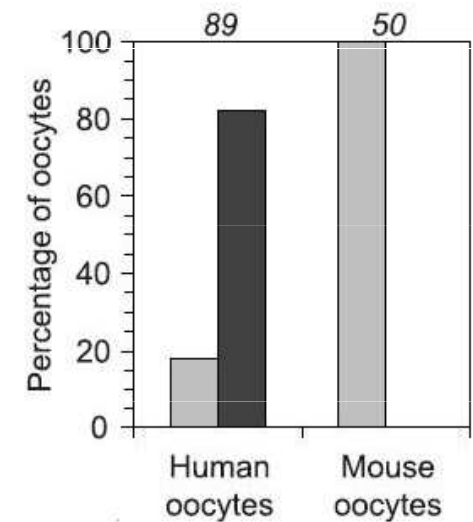
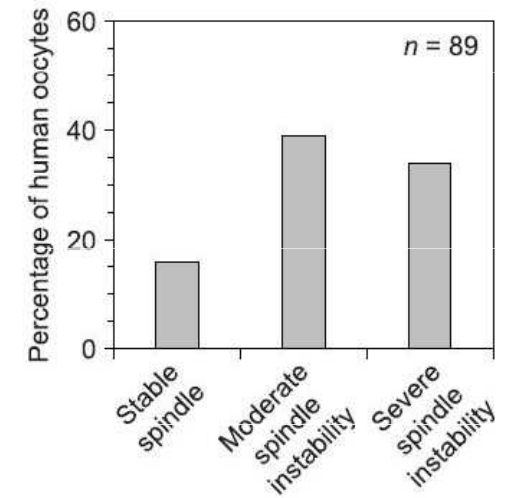
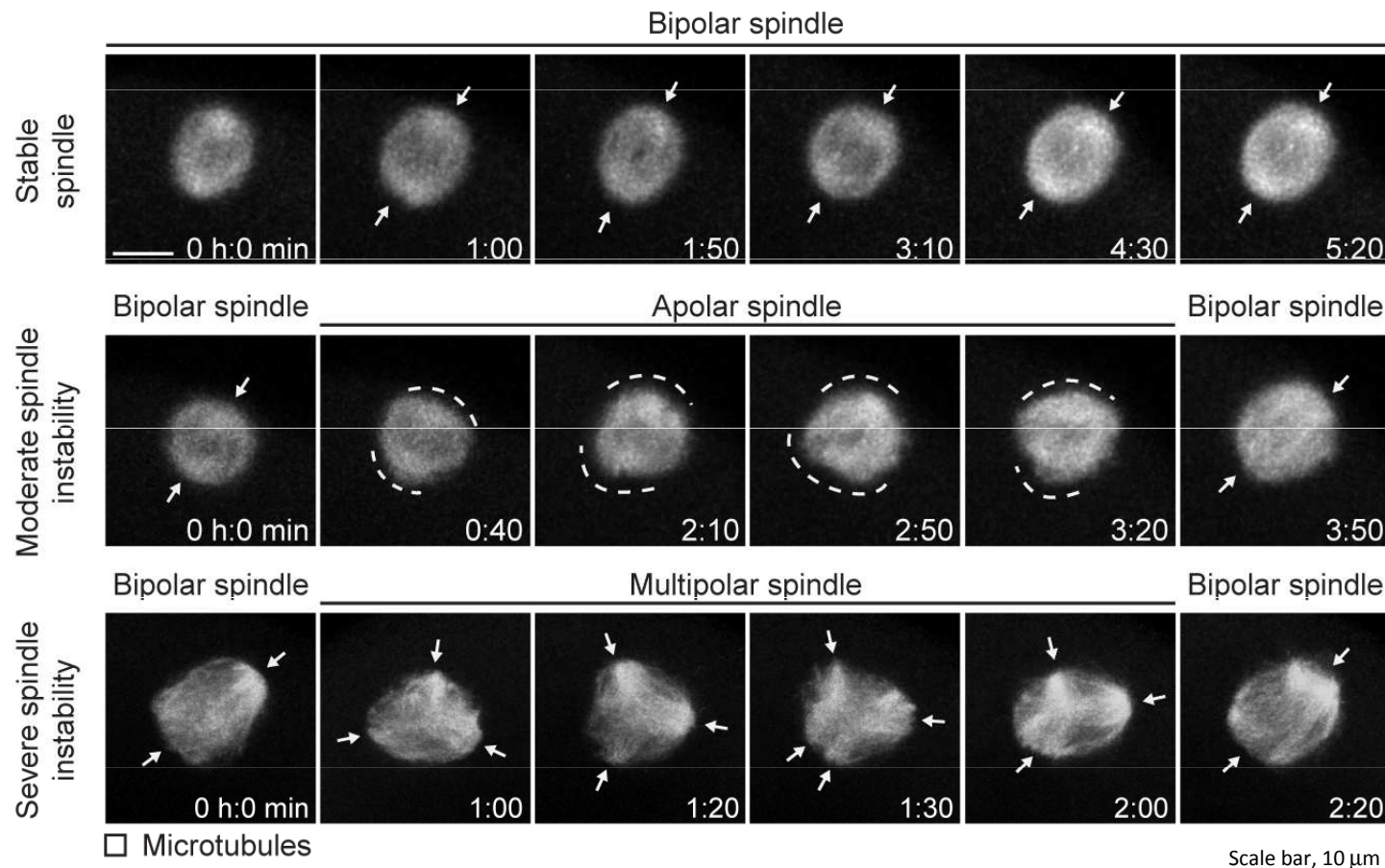
# Human oocyte spindle is unstable

## Severe spindle instability



Chromosomes (H2B-mRFP)  
Microtubules (MAP4-EGFP)

# Human oocyte spindle is unstable

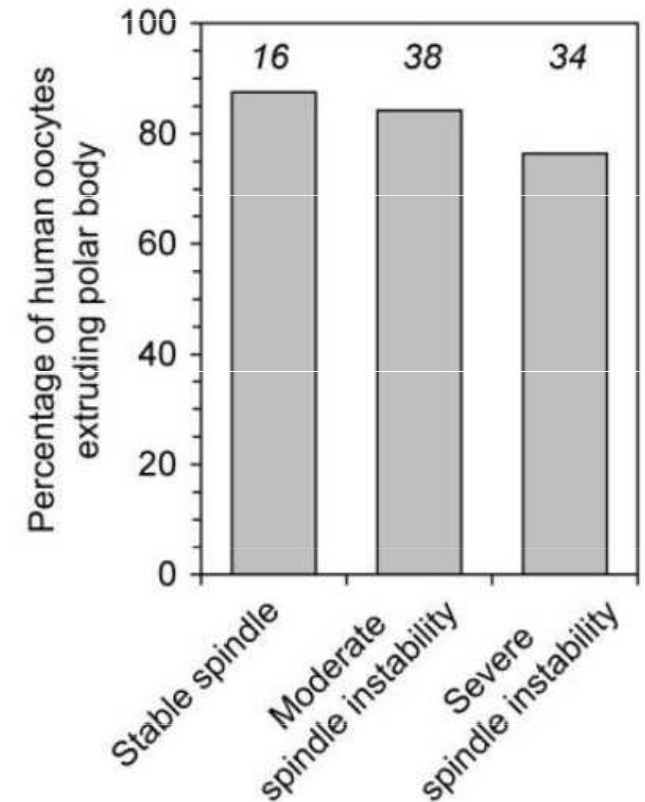
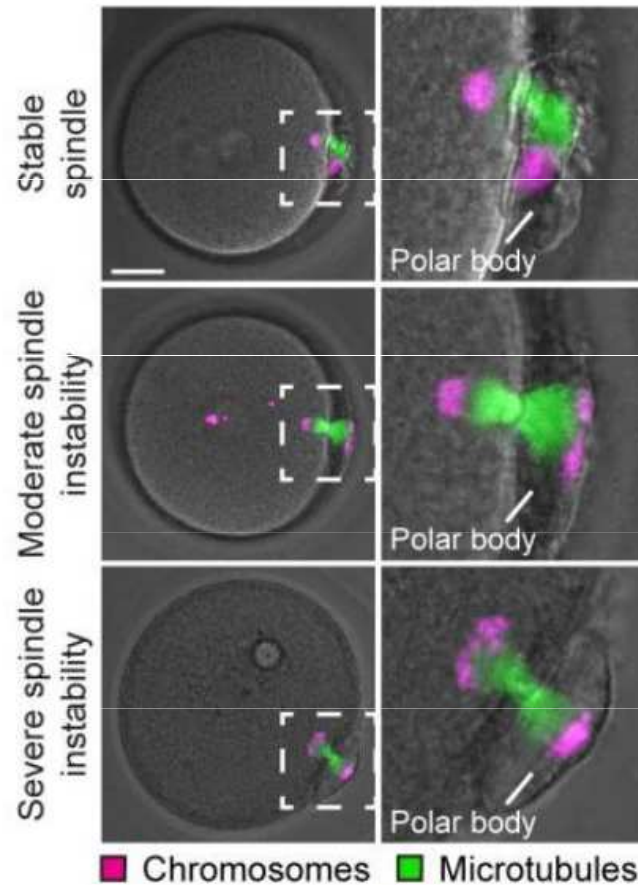
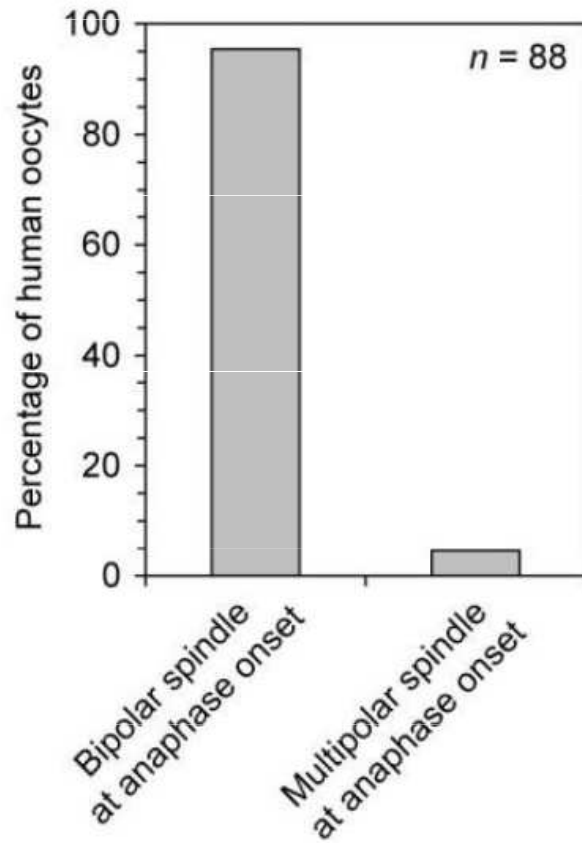


❖ **Prolonged spindle instability was observed in ~80% of human oocytes\* but no mouse oocytes**

\*Surplus oocytes from stimulated IVF cycles matured *in vitro* !

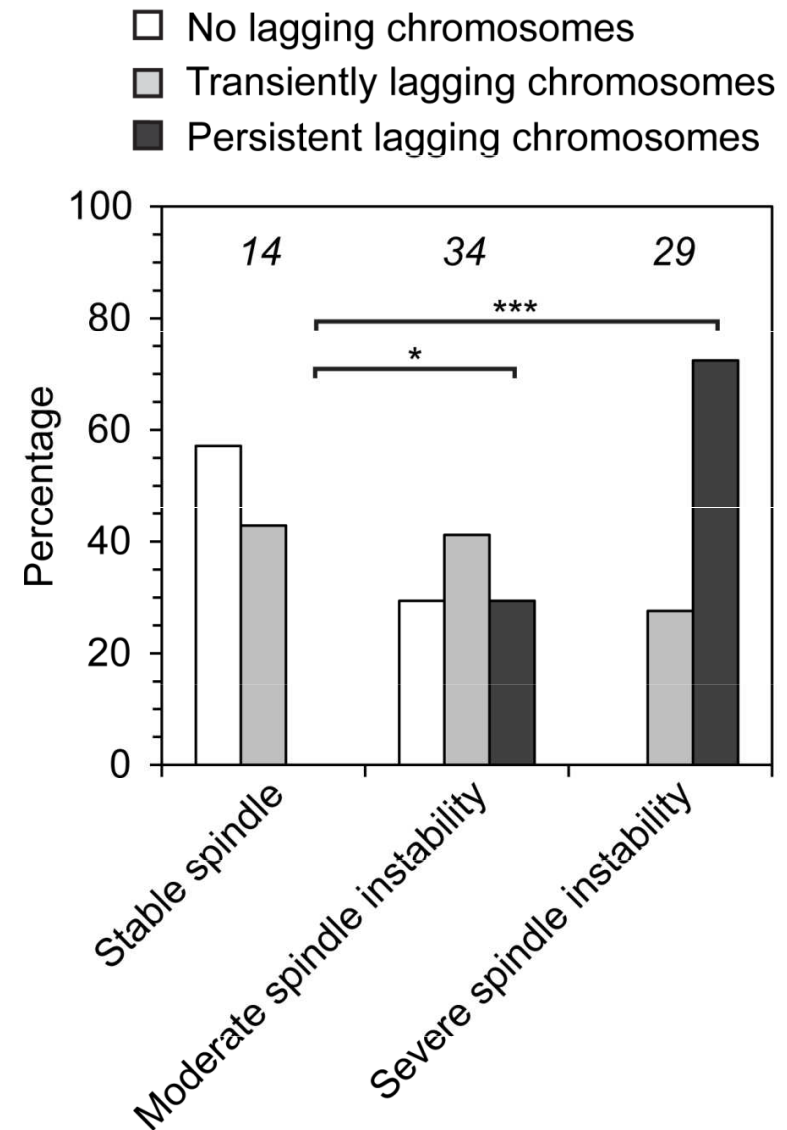
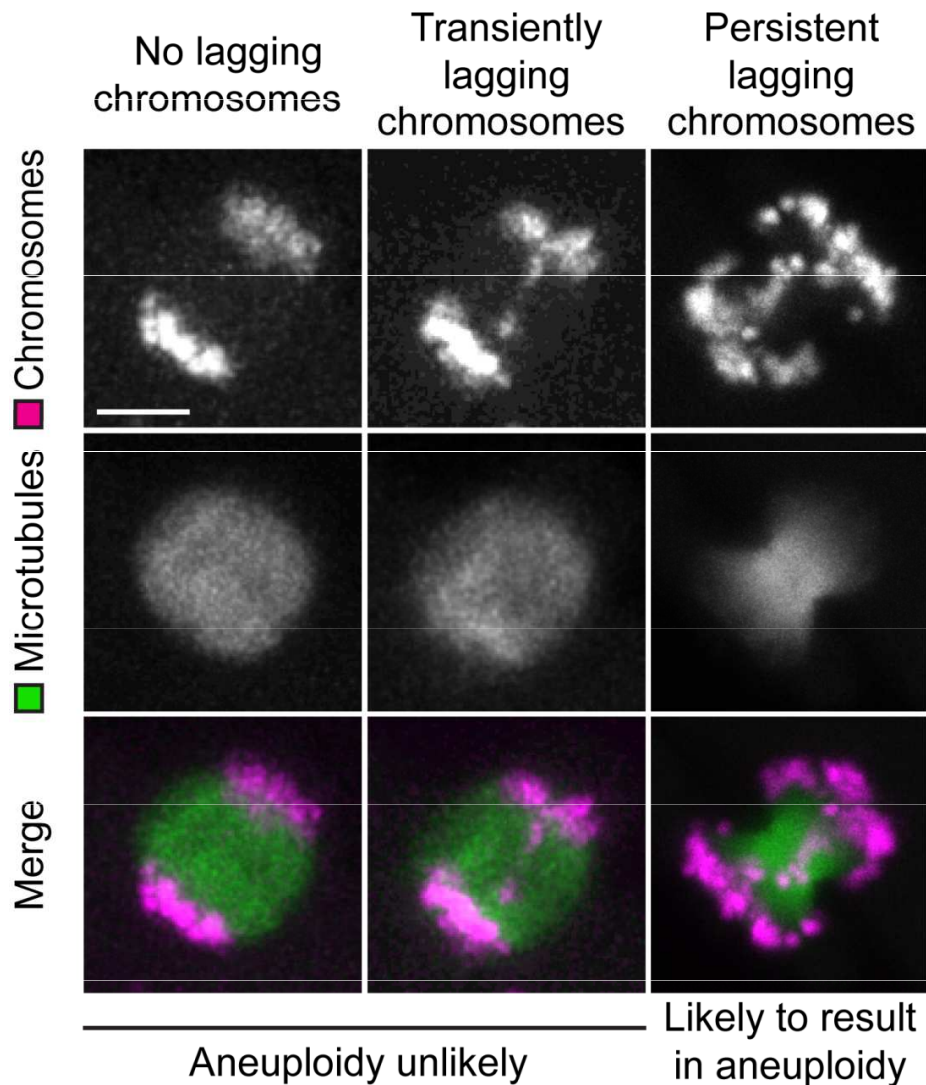
# Human oocyte spindle is unstable

- ❖ Majority of human oocytes recovered from spindle instability before anaphase and extruded a polar body



# Spindle instability favours chromosome missegregation

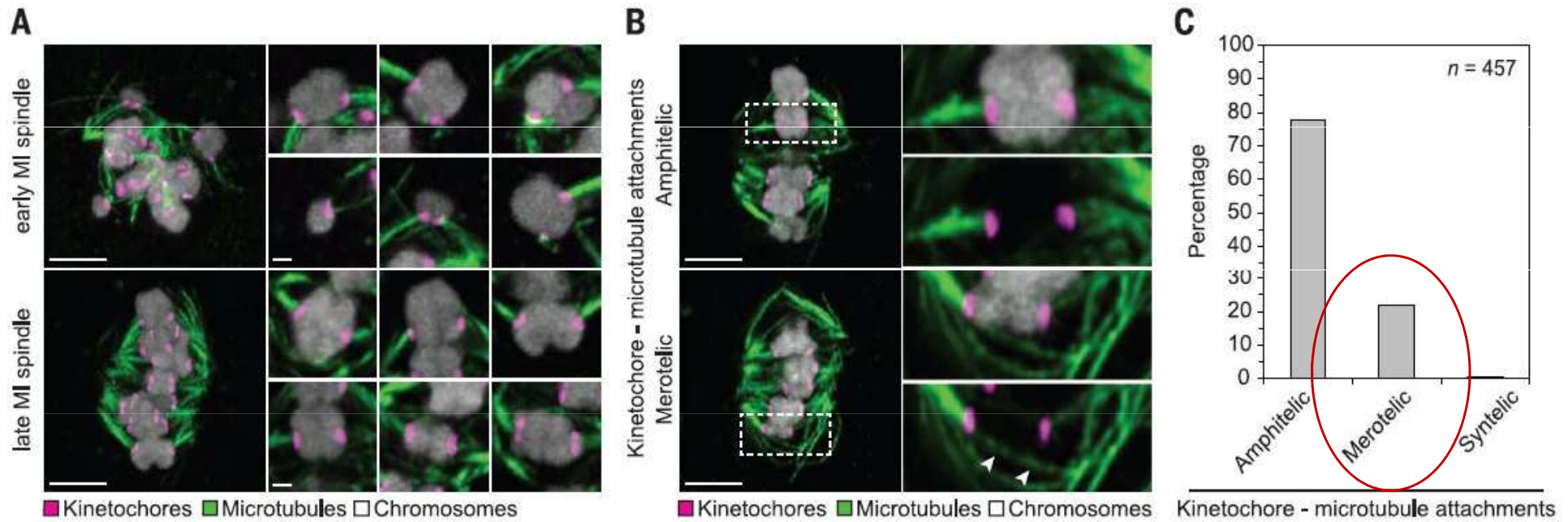
## ❖ Spindle instability correlates with chromosome segregation errors





# Spindle instability favours chromosome missegregation

- ❖ Correction of kinetochore-microtubule attachments is incomplete close to anaphase

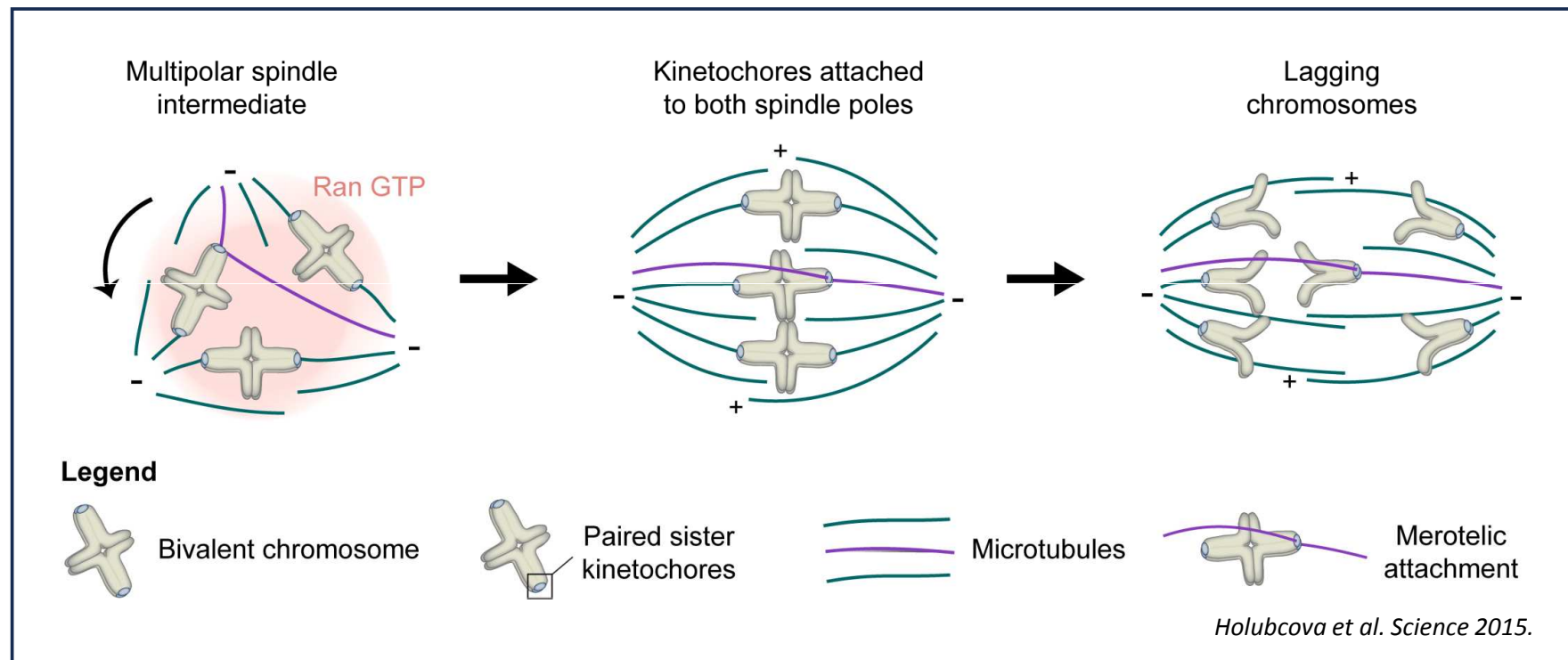


Scale bar, 5  $\mu$ m



# Spindle instability favours chromosome missegregation

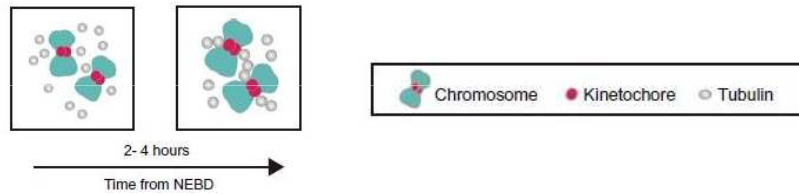
- ❖ at the absence of centrosomes, human oocytes rely on MT nucleation from chromatin
- ❖ chromosome-mediated spindle assembly is slow process and formed spindle is inherently unstable



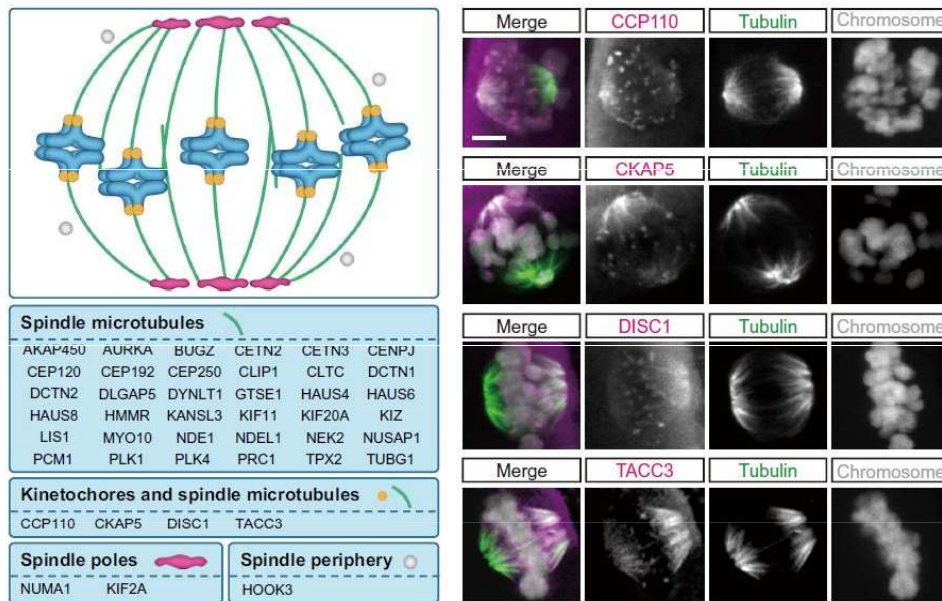
- ❖ improper microtubule-kinetochore attachments established during spindle build-up and remodelling persist to anaphase causing chromosome lagging that is likely to result in aneuploidy

# Acentrosomal spindle assembly

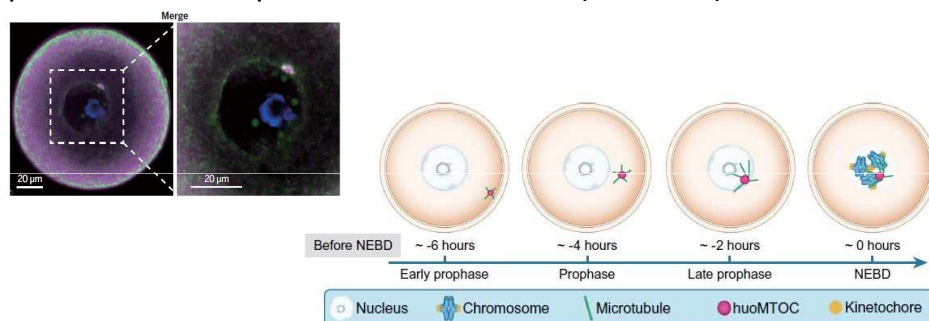
- MT nucleation initiated at kinetochores



- molecular composition of human oocyte spindle



- putative human specific MT nucleators (huMTOC)?



RESEARCH

Wu et al., Science 2022

## RESEARCH ARTICLE SUMMARY

HUMAN FERTILITY

### The mechanism of acentrosomal spindle assembly in human oocytes

Tianyu Wu†, Jie Dong†, Jing Fu†, Yanping Kuang†, Biaobang Chen, Hao Gu, Yuxi Luo, Ruihuan Gu, Meiling Zhang, Wen Li, Xi Dong, Xiaoxi Sun\*, Qing Sang\*, Lei Wang\*



## RESEARCH ARTICLE SUMMARY

Wu et al., Science 2024

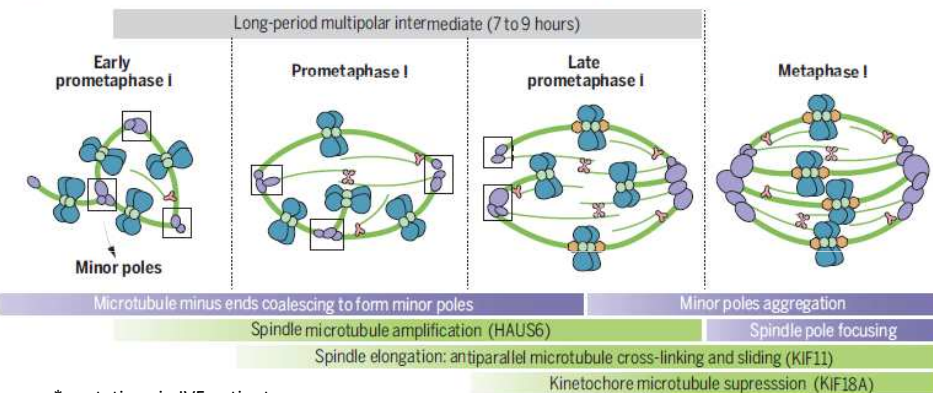
REPRODUCTION

### Mechanisms of minor pole-mediated spindle bipolarization in human oocytes

Tianyu Wu†, Yuxi Luo†, Meiling Zhang†, Biaobang Chen†, Xingzhu Du, Hao Gu, Siyuan Xie, Zhiqi Pan, Ran Yu, Ruiqi Hai, Xiangli Niu, Guimin Hao, Liping Jin, Juanzi Shi, Xiaoxi Sun, Yanping Kuang, Wen Li\*, Qing Sang\*, Lei Wang\*



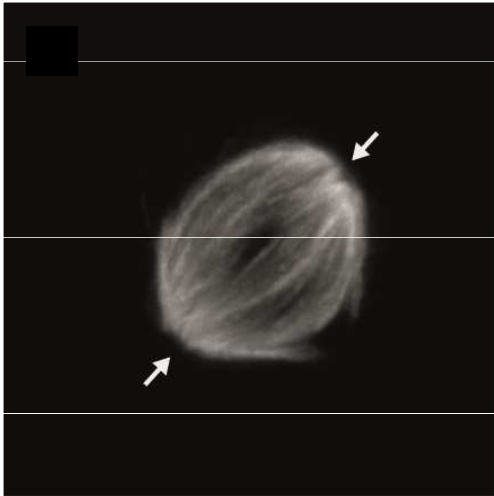
- nascent MT (-)ends coalesce into minor spindle poles which later aggregate to generate opposite spindle poles
- MT amplification, cross-linking and sliding that is required for spindle elongation and bipolarization



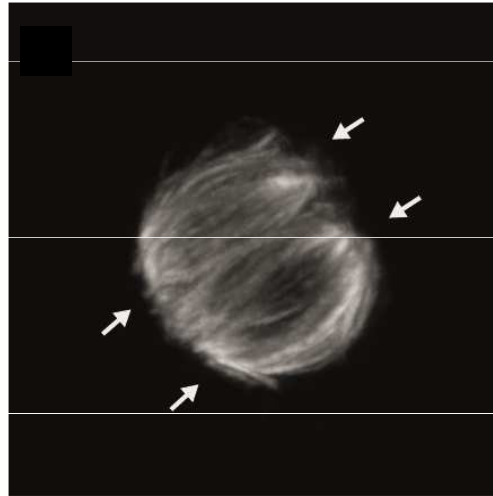
\* mutations in IVF patients

# Human oocyte spindle poles are not stabilized

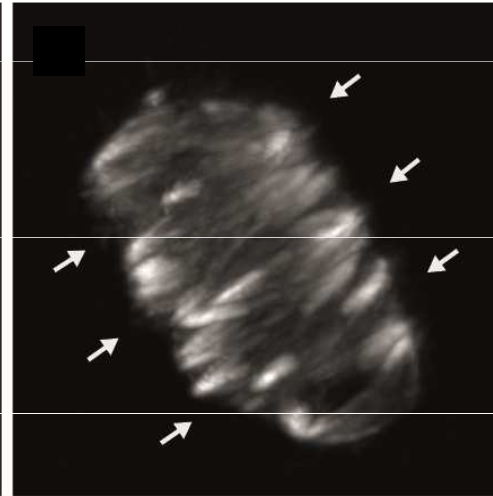
- ❖ Established spindle poles in human oocytes are prone to loosening and disintegration



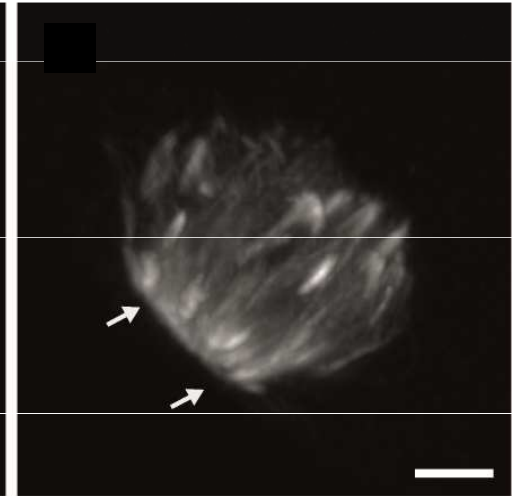
Focused spindle poles



Broad spindle poles

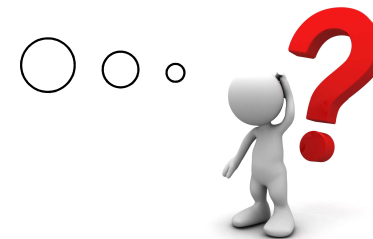


Loosen spindle poles



Disintegrated spindle pole

How are spindle poles organized at the absence of centrosomes?





# Incidence of unstable acentrosomal spindles



How are spindles in non-human mammalian oocytes stabilized

**82 %**



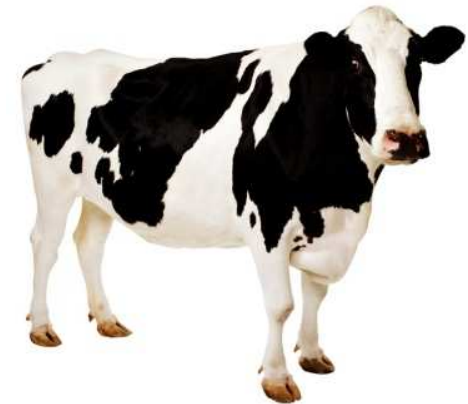
0 %



4.4 %



6%



# Mammalian oocyte spindle pole organization



## RESEARCH ARTICLE

OOCYTE DIVISION

### A liquid-like spindle domain promotes acentrosomal spindle assembly in mammalian oocytes

Chun So<sup>1\*</sup>, K. Bianka Seres<sup>1,2,3\*</sup>, Anna M. Steyer<sup>4,5</sup>, Eike Mönlich<sup>1</sup>, Dean Clift<sup>2</sup>, Anastasija Pejkovska<sup>1</sup>, Wiebke Möbius<sup>4,5</sup>, Melina Schuh<sup>1,2,†</sup>



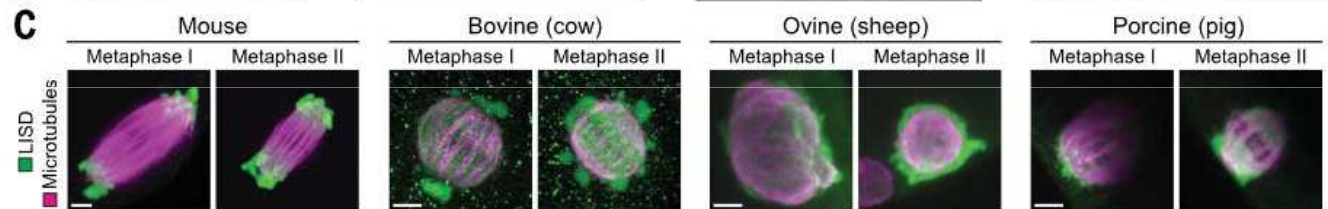
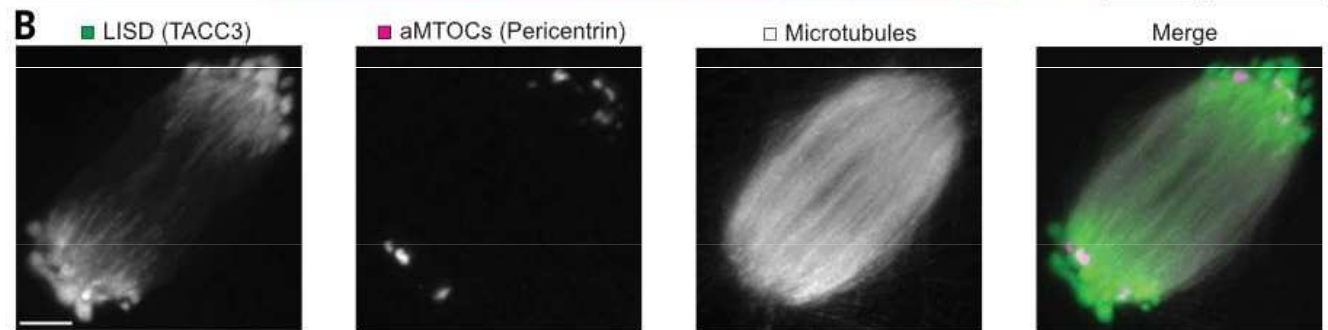
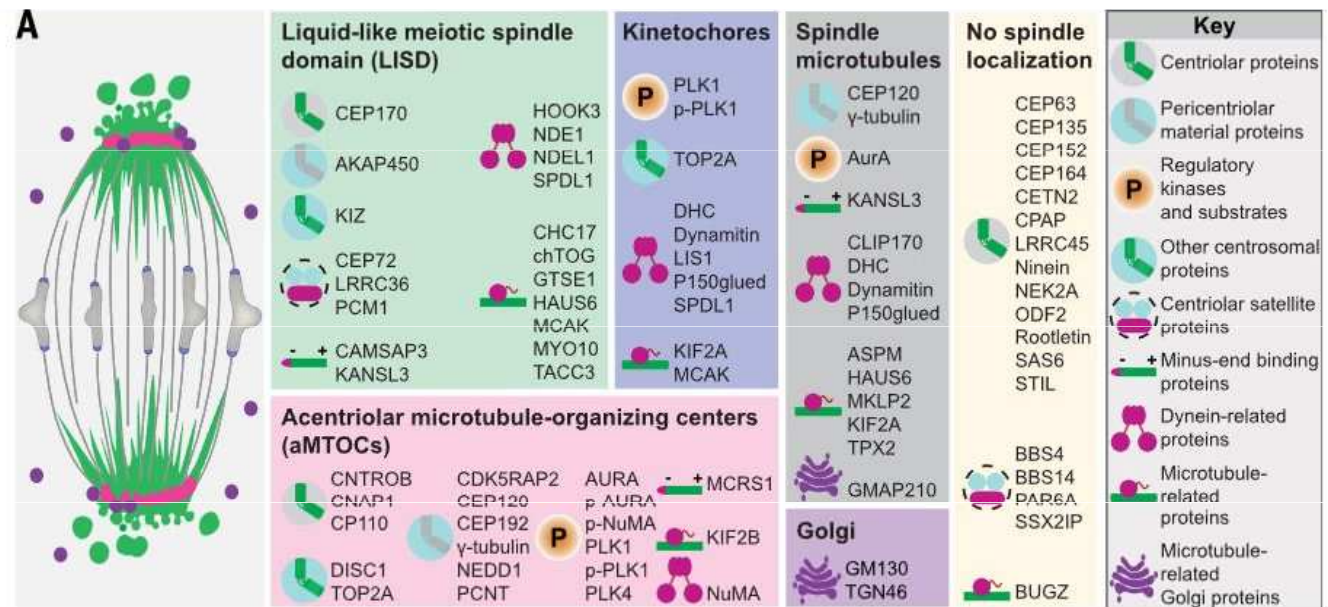
Chun So



Melina Schuh

#### ❖ Liquid-like meiotic spindle domain (LISD)

- localized at poles and permeates the MT mass of mammalian oocytes
- selectively concentrates multiple centrosomal and MT-associated proteins
- allows rapid diffusion within the spindle volume
- disruption of the LISD disperses spindle regulatory factors and leads to severe spindle assembly defects



stable spindles

So et al., Science 2019



# Human oocyte spindle pole organization

## ❖ NuMA decorates spindle poles in mammalian oocytes

RESEARCH ARTICLE

So et al., Science 2022

CELL BIOLOGY

### Mechanism of spindle pole organization and instability in human oocytes

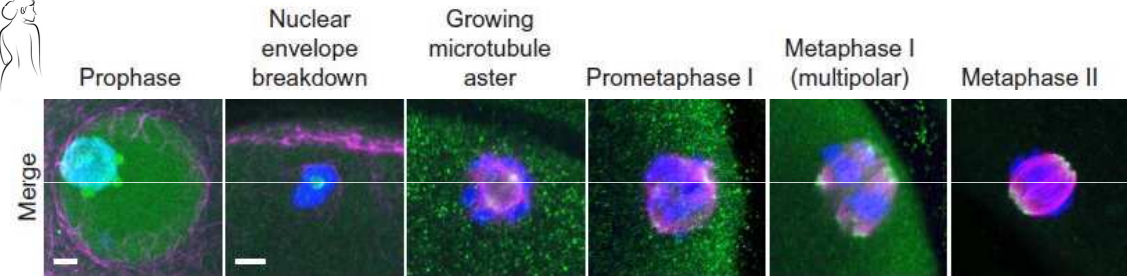


Chuh So

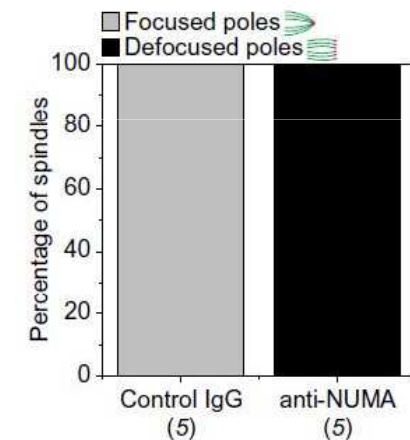
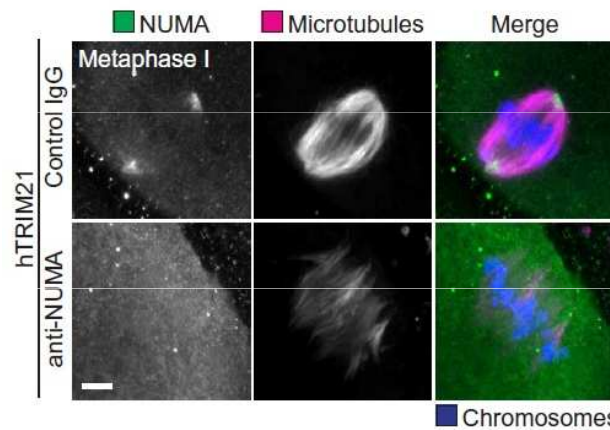
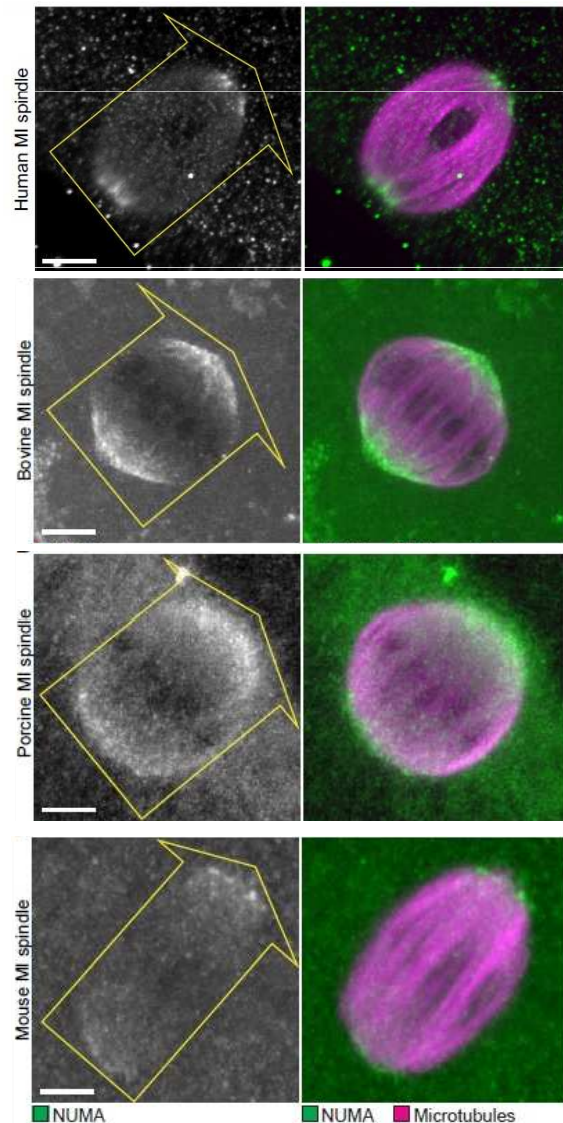
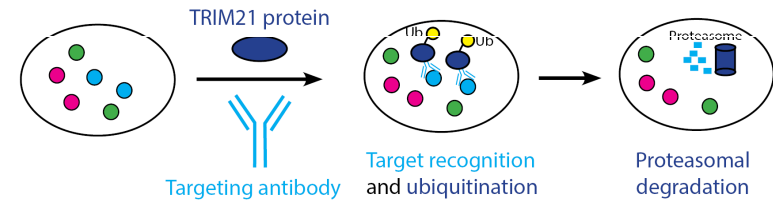


Melina Schuh

Chun So<sup>1</sup>, Katerina Menelaou<sup>1,2†</sup>, Julia Uraji<sup>1,2†</sup>, Katarina Harasimov<sup>1</sup>, Anna M. Steyer<sup>3</sup>, K. Bianka Seres<sup>1,2</sup>, Jonas Bucevičius<sup>4</sup>, Gražvydas Lukinavičius<sup>4</sup>, Wiebke Möbius<sup>3,5</sup>, Claus Sibold<sup>6</sup>, Andreas Tandler-Schneider<sup>6</sup>, Heike Eckel<sup>7</sup>, Rüdiger Moltrecht<sup>7</sup>, Martyn Blayney<sup>2</sup>, Kay Elder<sup>2</sup>, Melina Schuh<sup>1,5\*</sup>



- NuMA is required for MT focusing at spindle poles in human oocytes



# Human oocyte spindle pole organization

## ❖ NuMA decorates spindle poles in mammalian oocytes

RESEARCH ARTICLE

So et al., Science 2022

CELL BIOLOGY

### Mechanism of spindle pole organization and instability in human oocytes

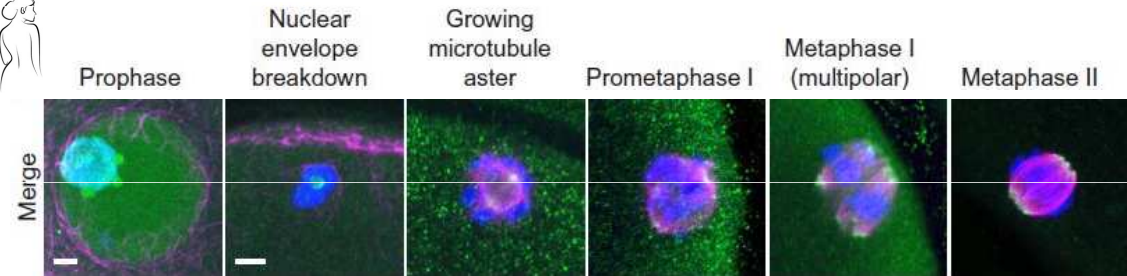


Chuh So

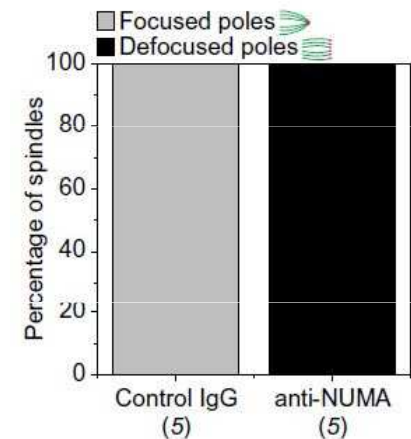
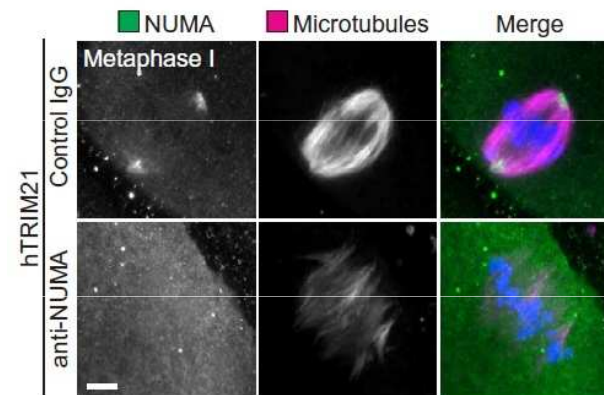
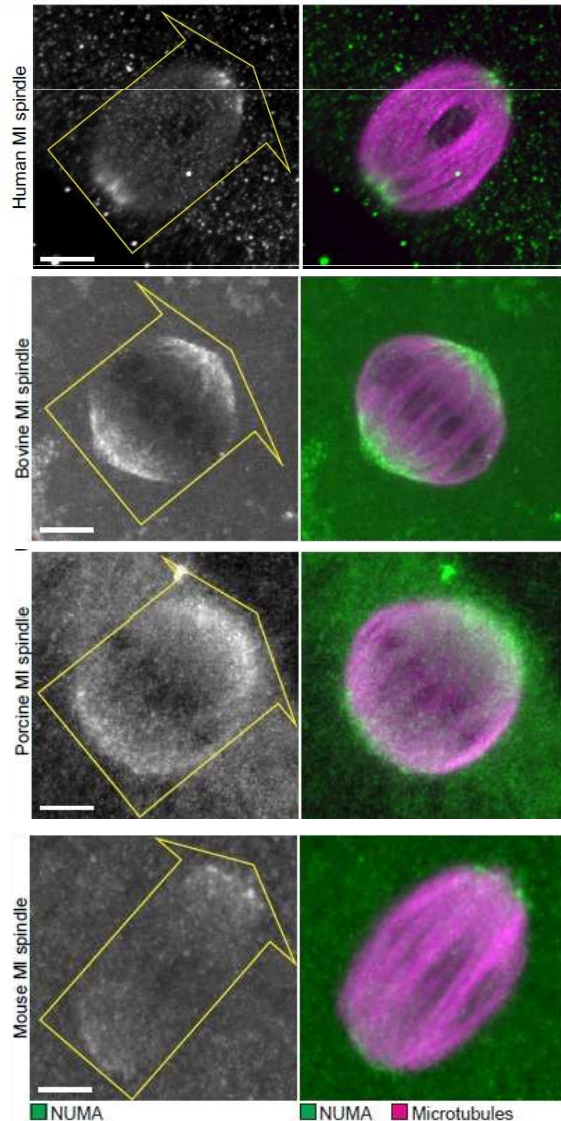
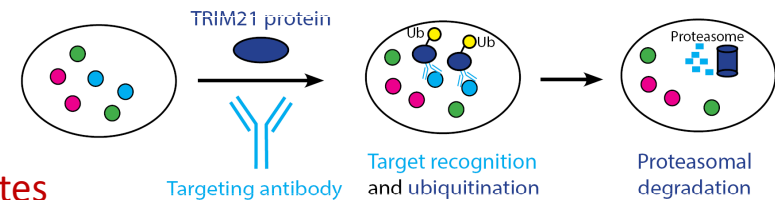


Melina Schuh

Chun So<sup>1</sup>, Katerina Menelaou<sup>1,2†</sup>, Julia Uraji<sup>1,2†</sup>, Katarina Harasimov<sup>1</sup>, Anna M. Steyer<sup>3</sup>, K. Bianka Seres<sup>1,2</sup>, Jonas Bucevičius<sup>4</sup>, Gražvydas Lukinavičius<sup>4</sup>, Wiebke Möbius<sup>3,5</sup>, Claus Sibold<sup>6</sup>, Andreas Tandler-Schneider<sup>6</sup>, Heike Eckel<sup>7</sup>, Rüdiger Moltrecht<sup>7</sup>, Martyn Blayney<sup>2</sup>, Kay Elder<sup>2</sup>, Melina Schuh<sup>1,5\*</sup>



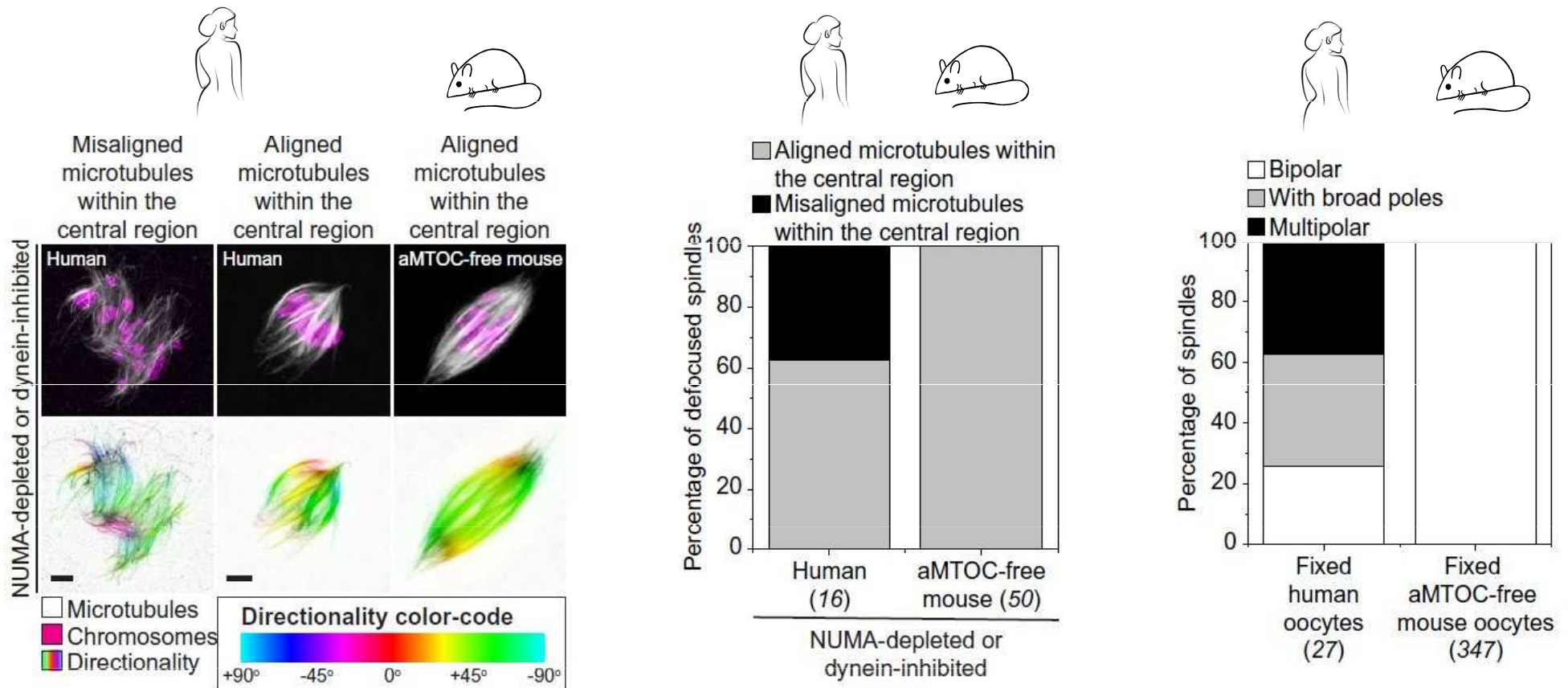
- acute depletion of NuMA → NuMA is required for MT focusing at spindle poles in human oocytes





# Search for oocyte spindle stabilizing factor

## Misalignment of microtubules in central region of human oocyte spindle



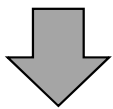
**Human oocytes must lack a stabilizing protein that protects mouse, porcine and bovine oocytes from spindle instability**

# Search for oocyte spindle stabilizing factor

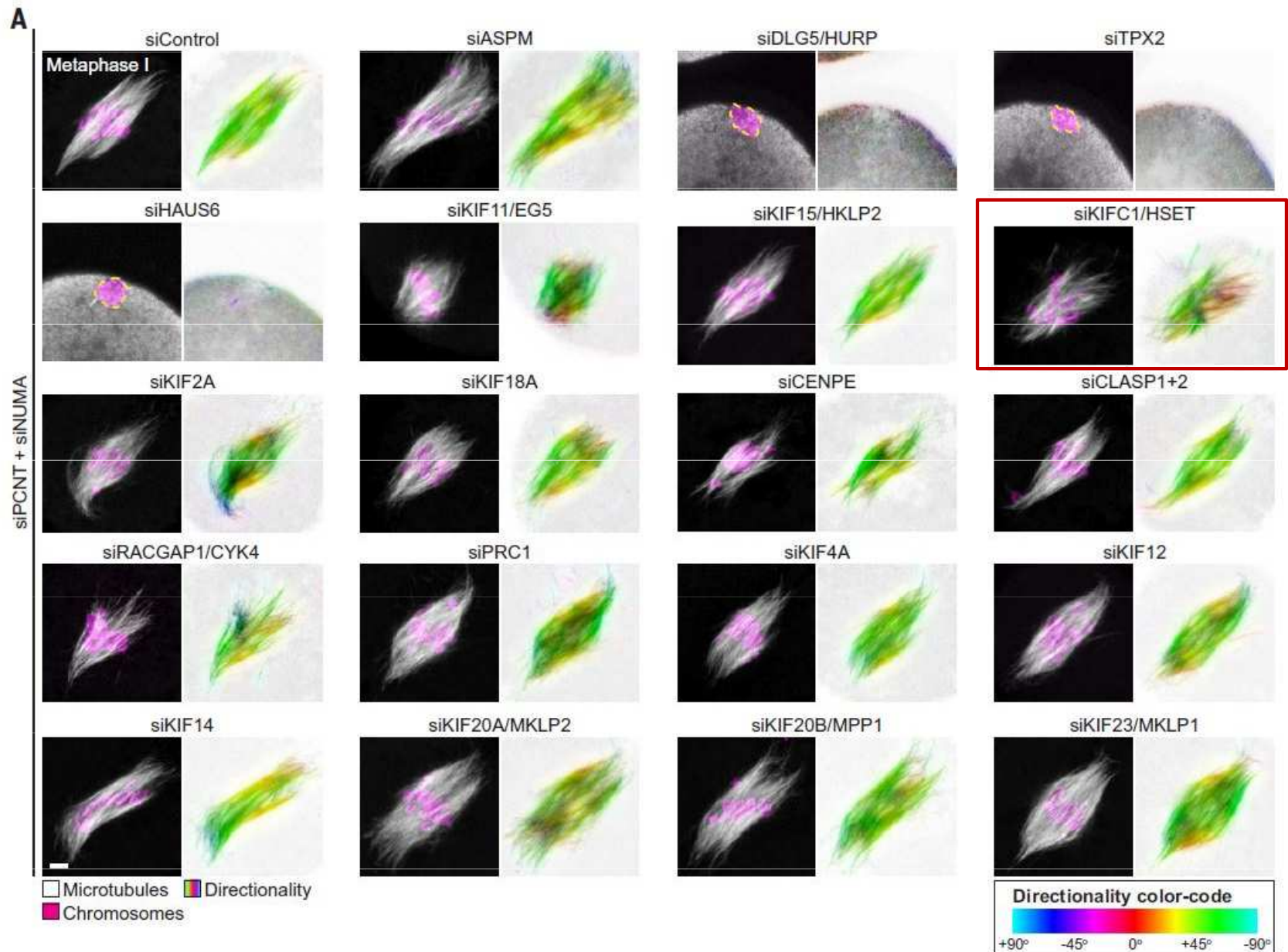
## si RNA screen



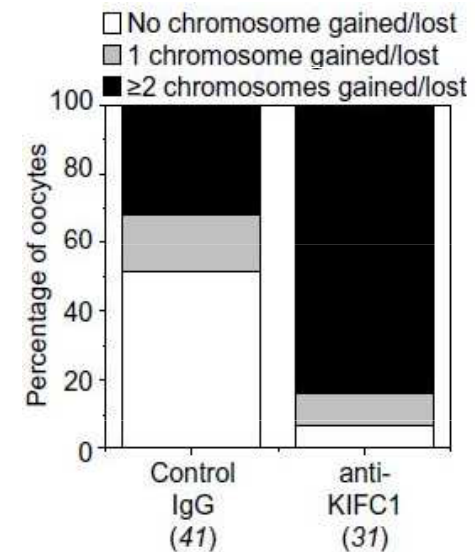
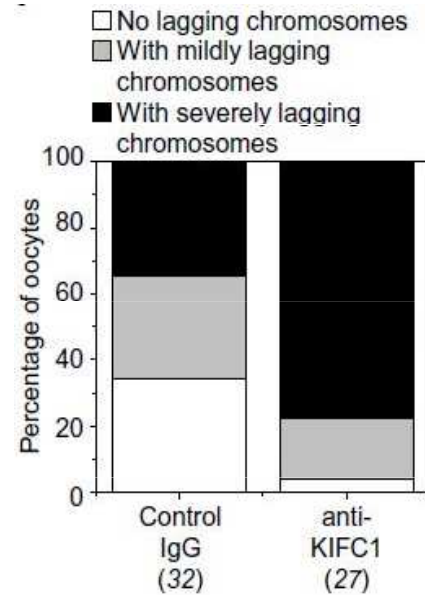
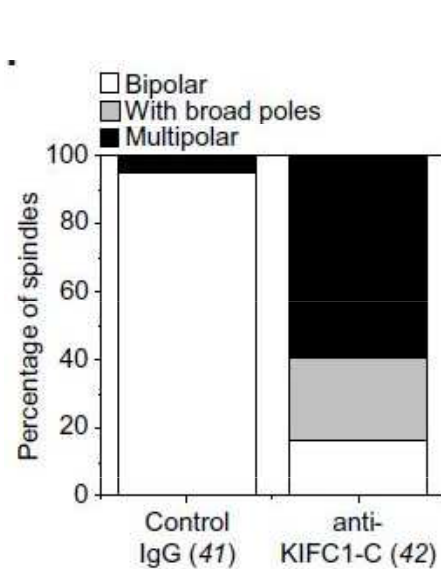
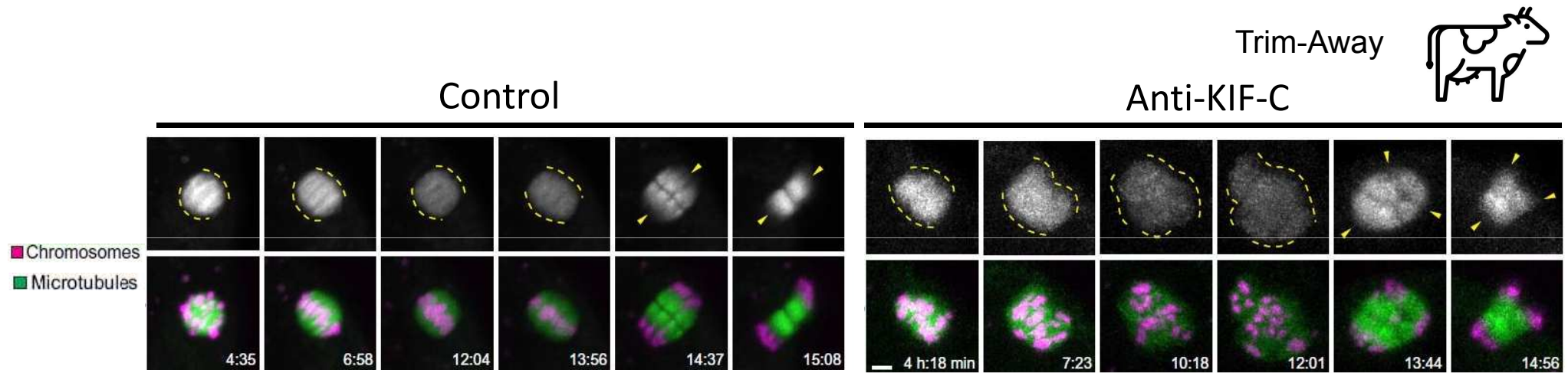
co-depletion of  
NuMA and  
candidate  
spindle-  
associated  
proteins



phenotypes  
analyzed



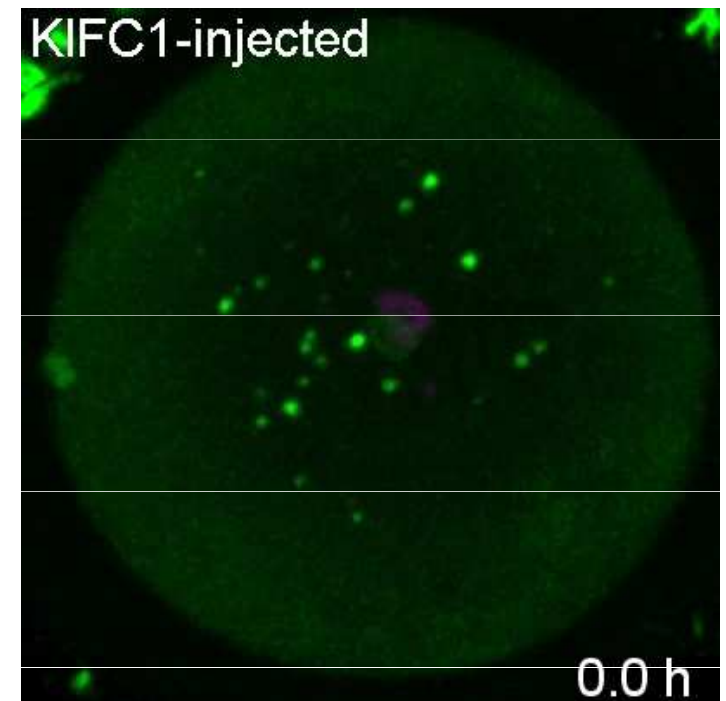
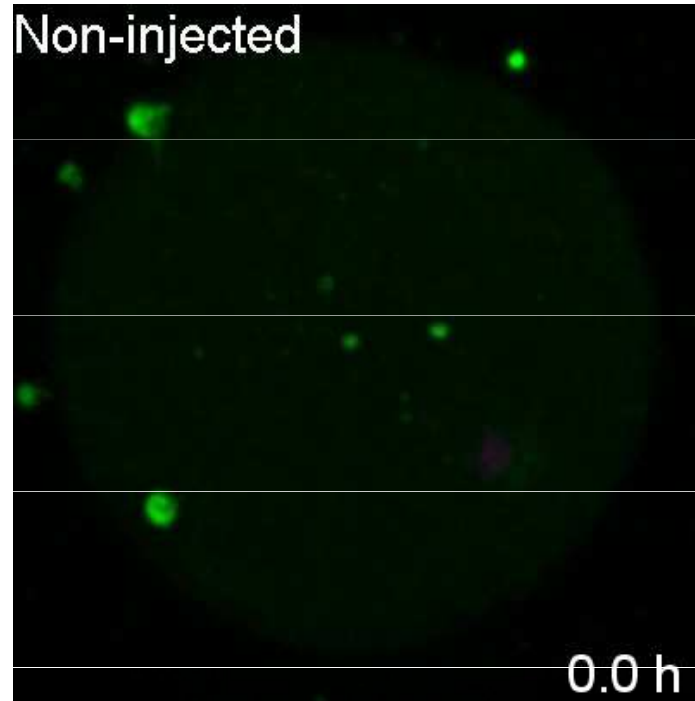
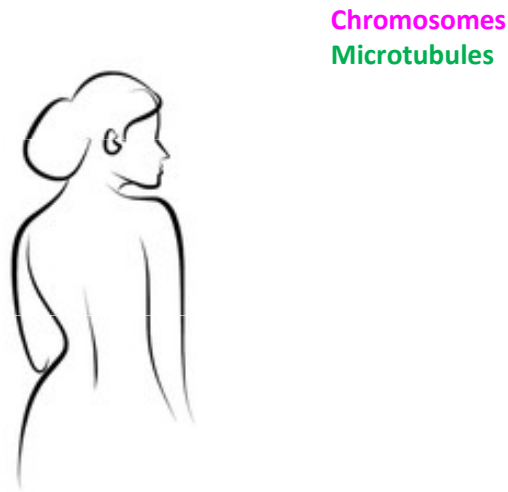
# Search for oocyte spindle stabilizing factor



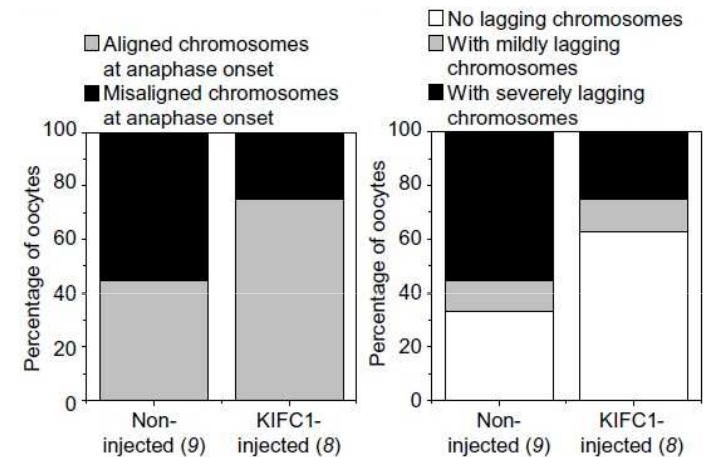
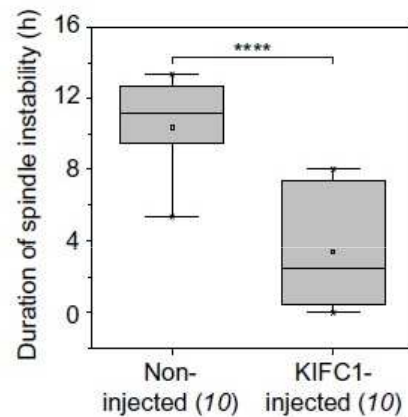
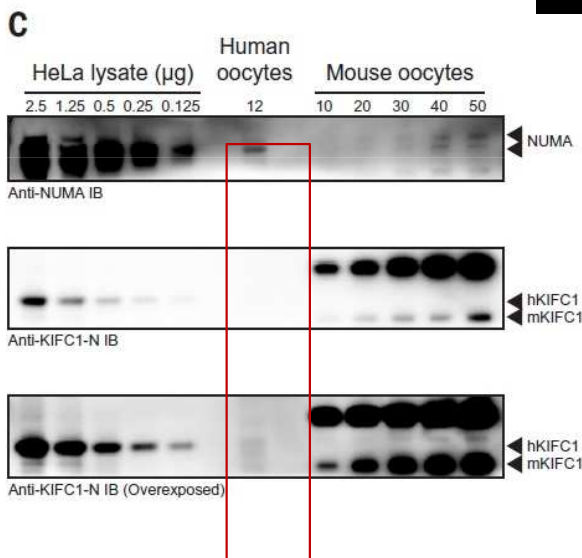
Depletion of KIFC1/HSET induces spindle instability and promotes aneuploidy in bovine oocytes



# Search for oocyte spindle stabilizing factor



❖ Human oocytes are deficient in KIFC1

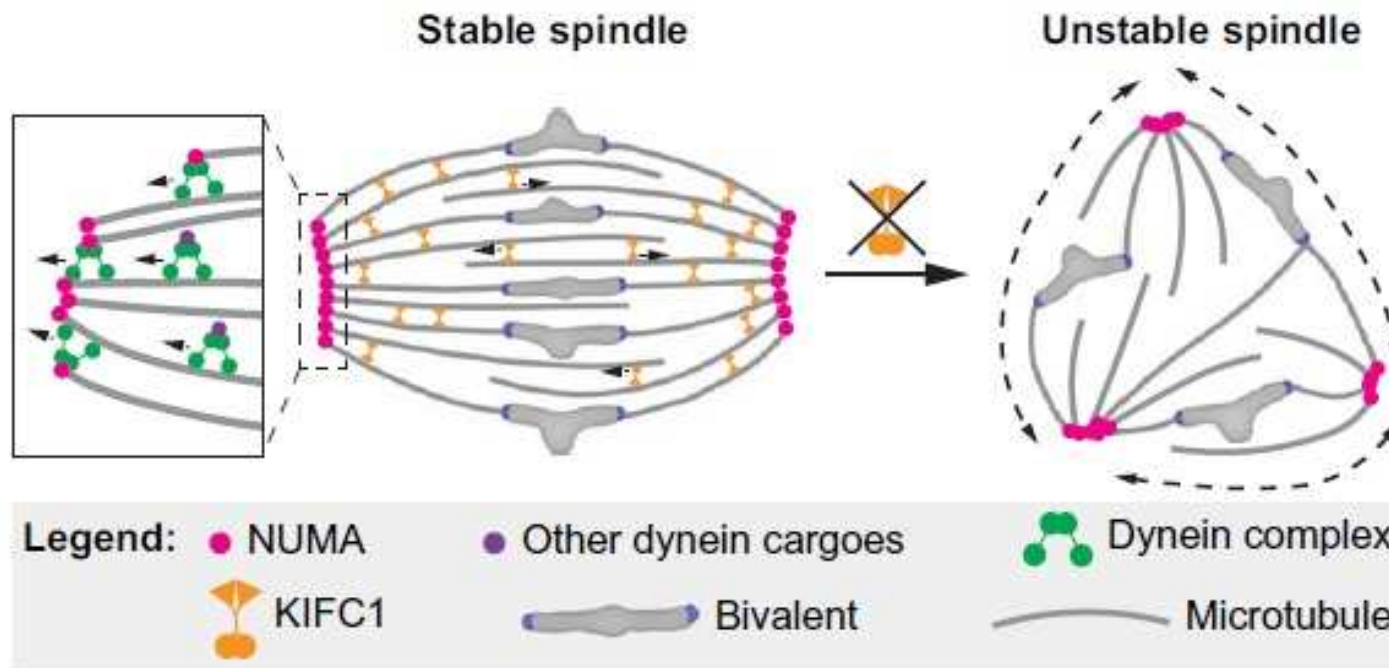


❖ Exogenous KIFC1 rescues spindle instability in human oocytes



# Prevention of oocyte spindle instability

- ❖ **KIFC1** ensures the spindle stability and prevents fragmentation of spindle poles by **ensuring alignment of MT at central region and crosslinking MT minus ends at spindle poles**

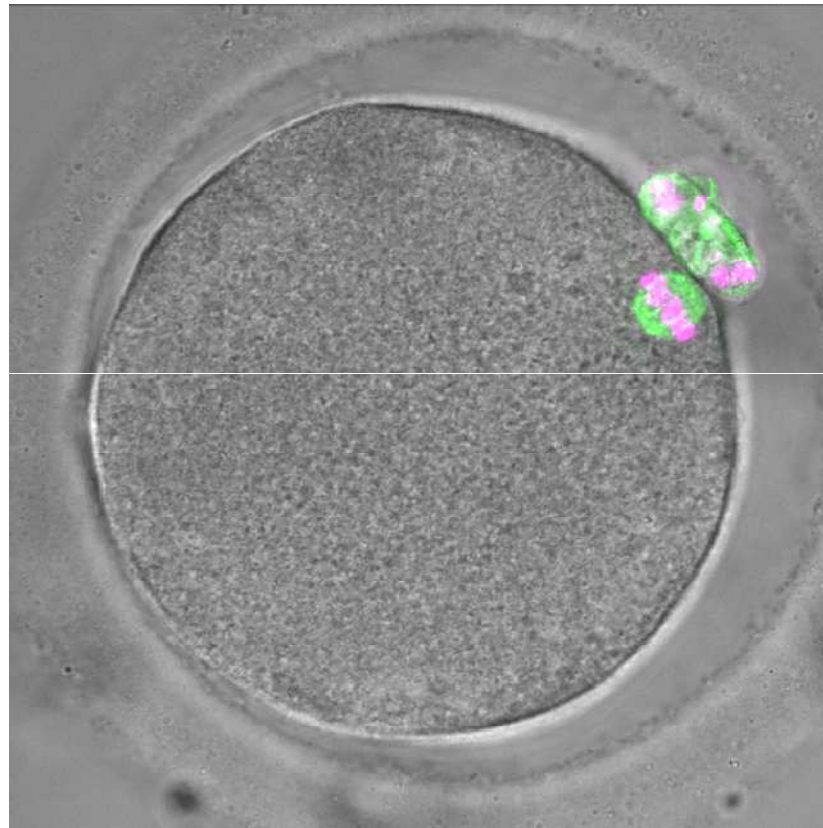


- ❖ **NuMa** organize acentrosomal spindle poles by ensuring coalescence of crosslinked MT-minus ends

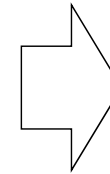
# Egg maturity

## ❖ Mature egg

= metaphase II arrested oocytes with PB extruded and chromosomes aligned in MI



DNA, microtubules

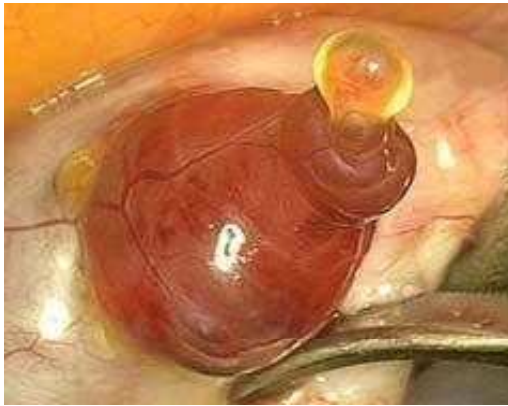


In IVF practice,  
all PB-displaying oocytes  
are regarded as MIIs  
and subjected to ICSI

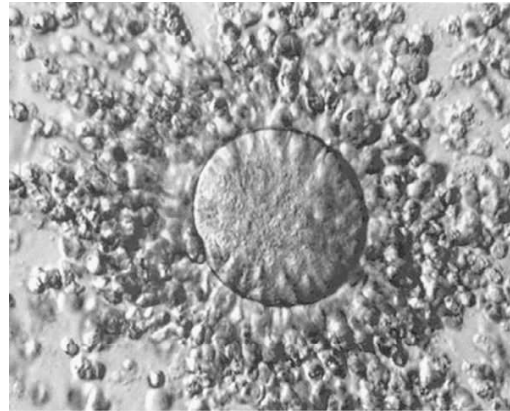
# Oocyte maturity

## IN VIVO

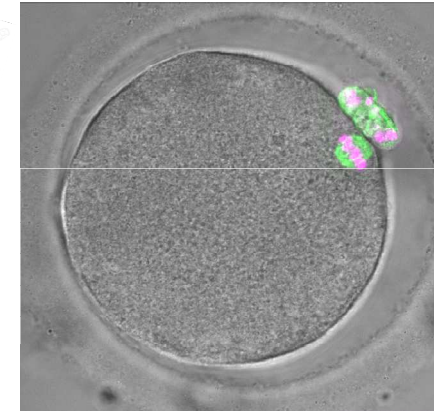
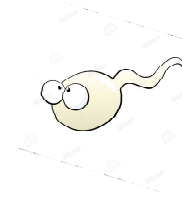
### NATURAL CYCLE



OVULATION

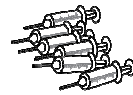


SINGLE COC

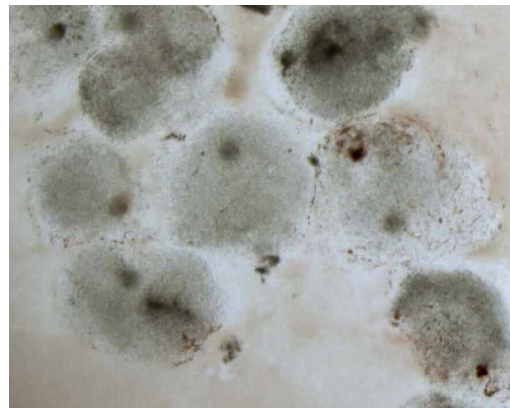


MATURE EGG

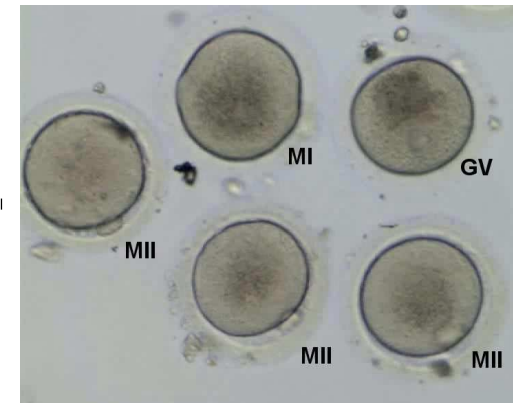
### CONTROLLED OVARIAN STIMULATION



PREOVULATORY FOLLICLES



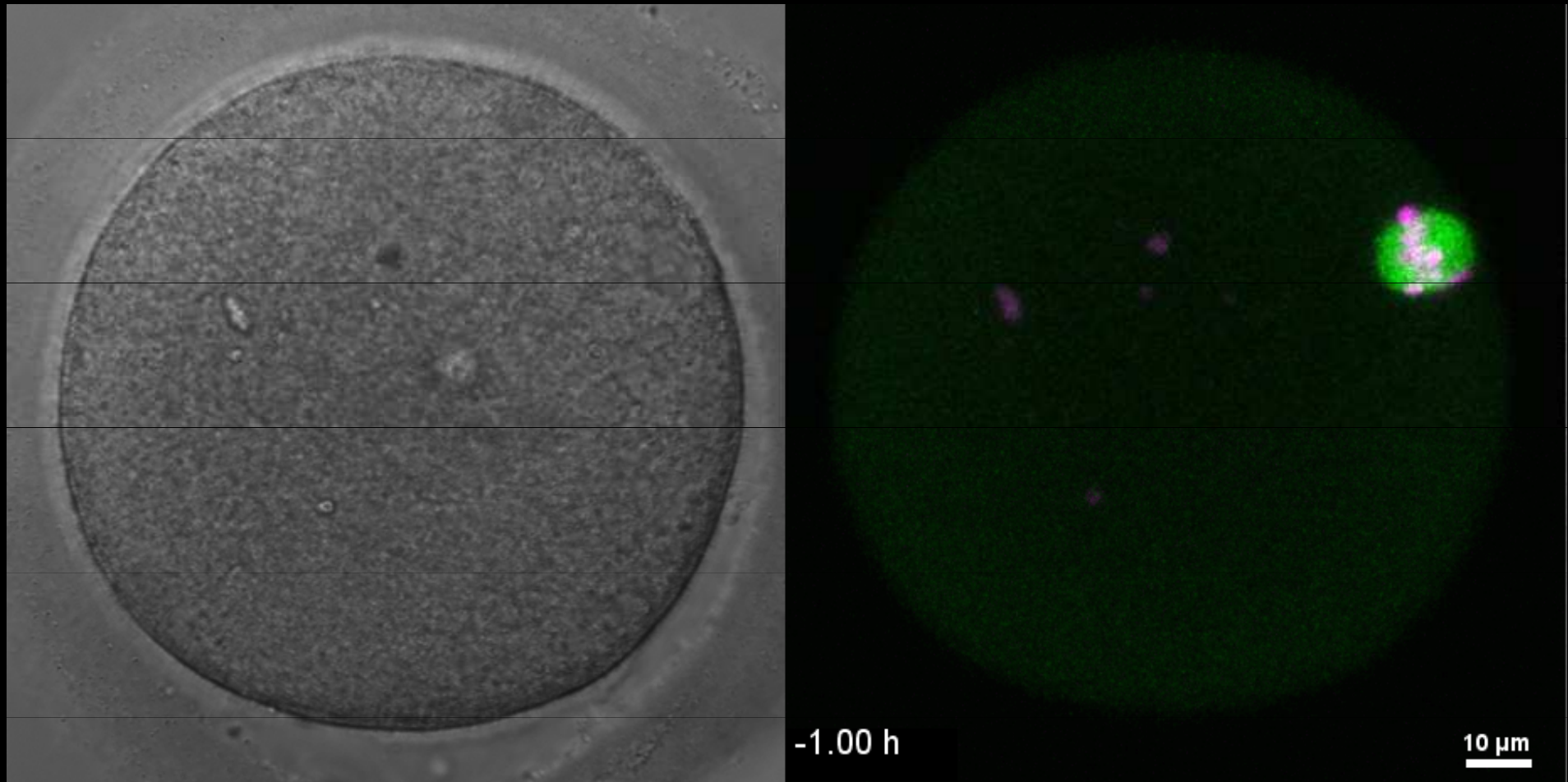
MULTIPLE COCS



MATURE + IMMATURE OOCYTES

# MI to MII transition and MII spindle assembly

- MII spindle formation is rapid compared to MI



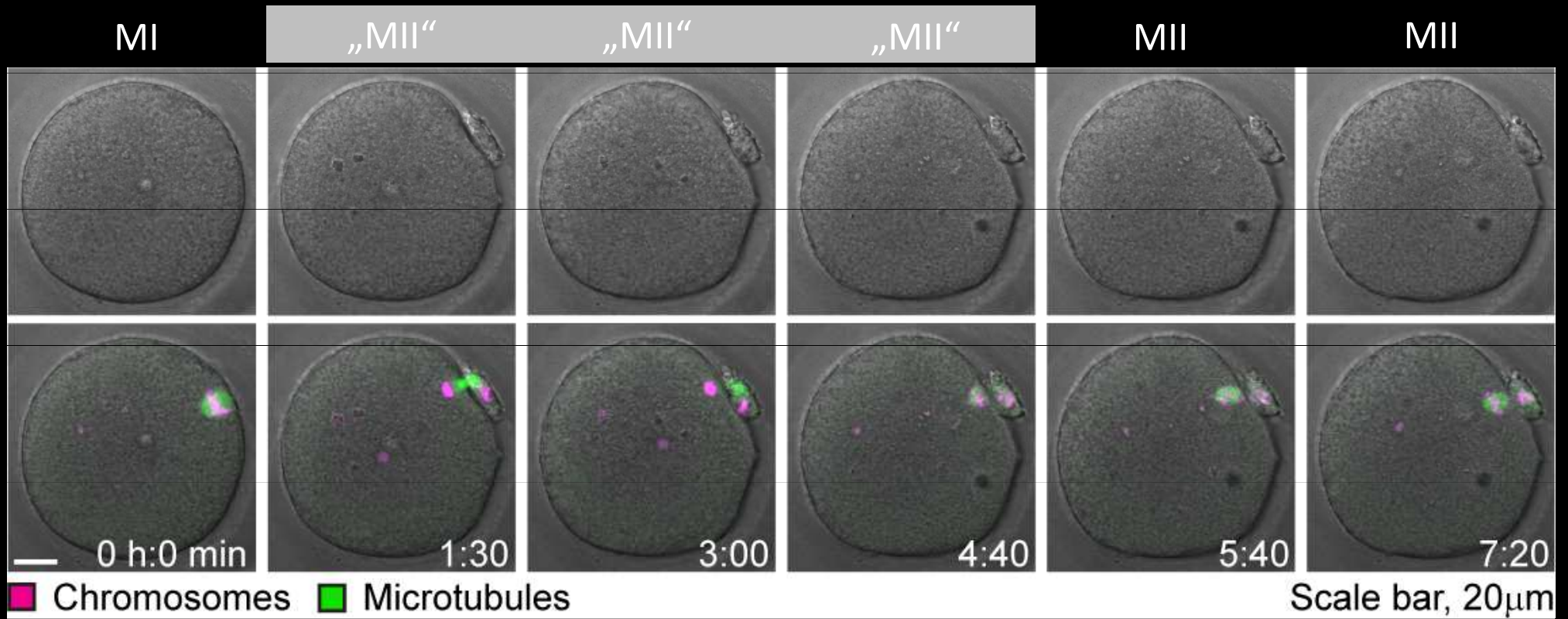
Chromosomes  
(H2B-mRFP)  
Microtubules  
(MAP4-EGFP)

- asynchrony between PB extrusion and MII arrest !



# MI to MII transition and MII spindle assembly

- Emergence of PB precedes MII arrest



→ risk of untimely fertilization (ICSI)



# Non-invasive spindle visualization

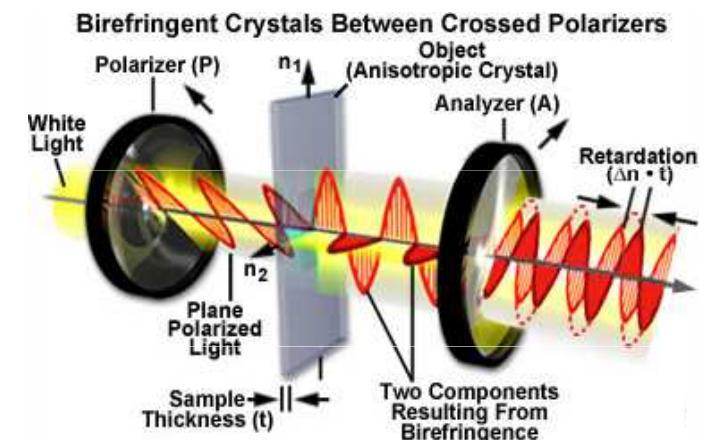
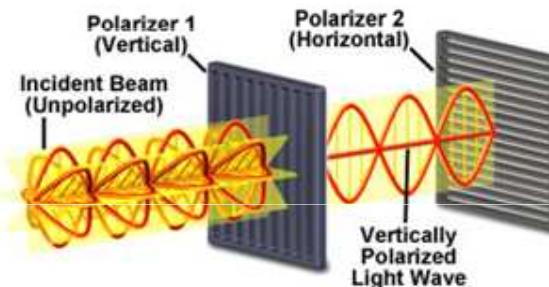
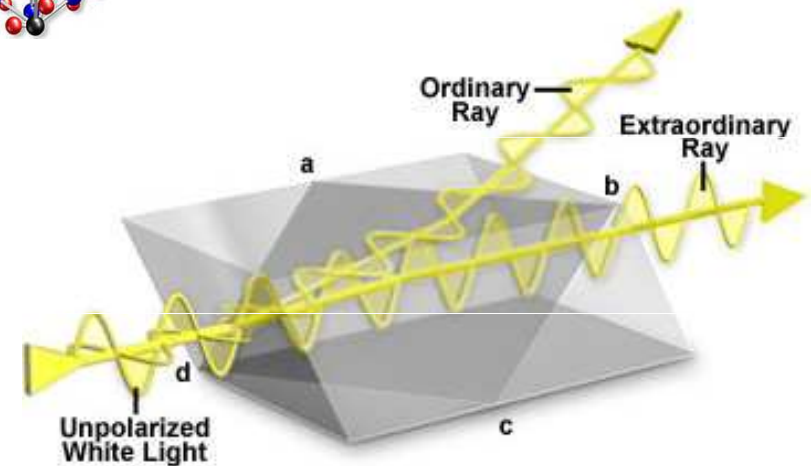
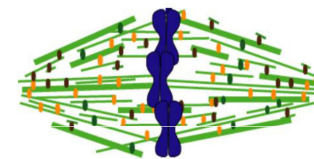
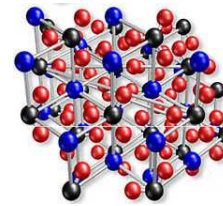
## ❖ Polarized Light Microscopy (PLM)

- based on interference of **polarized light** with **anisotropic** substances e.g. axial crystals, liquid crystals and **oriented (bio)polymers**

### BIREFRINGENCE

- property of certain materials to split a light beam to two rays (ordinary/extraordinary)

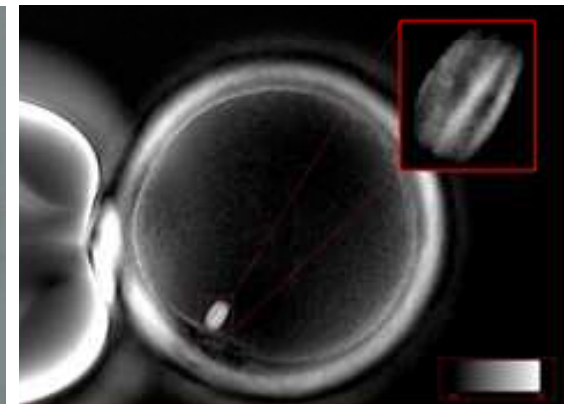
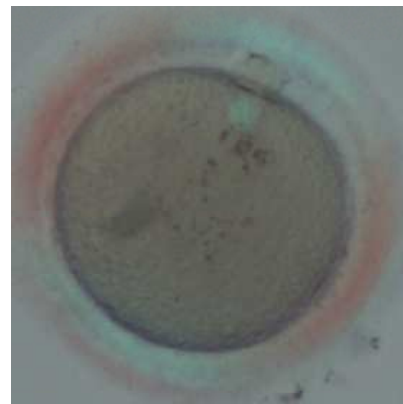
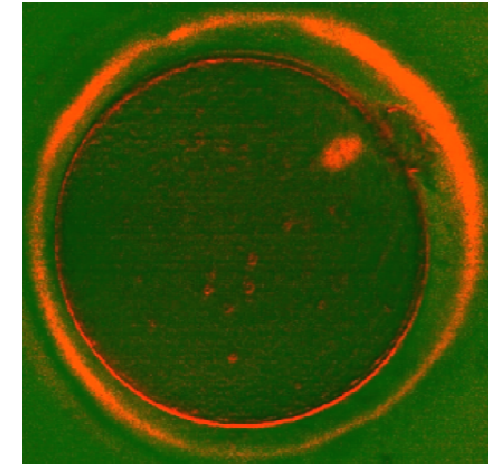
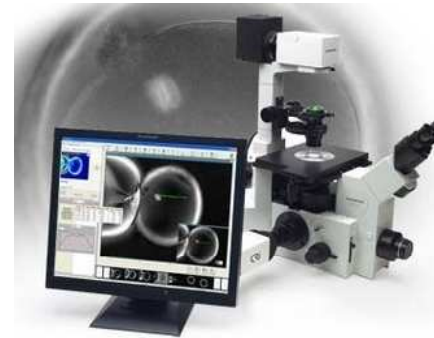
- polarized light is refracted by these anisotropic materials and divided to separate components vibrating perpendicularly
- both polarized light ray then pass through the analyzer and the relative retardance of one ray to the other is calculated



# Non-invasive spindle visualization

## ❖ Polarized Light Microscopy (PLM)

- enables **non-invasive** imaging of **birefringent** structures in living cells
- ↓
- presence and positioning of MII spindle
  - pattern of zona pellucida
- 
- presence of PLM-detectable MII spindle is a **positive marker of egg's fertilization and developmental competence**



jove Journal of Visualized Experiments

www.jove.com

Video Article

### Human Egg Maturity Assessment and Its Clinical Application

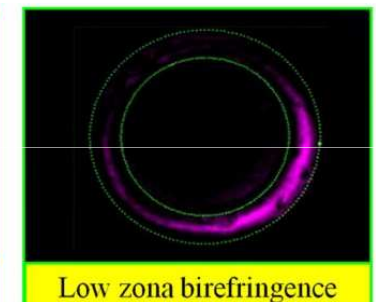
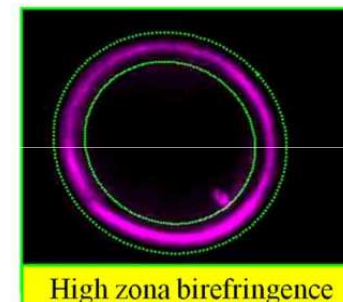
Zuzana Holubcová<sup>1,2</sup>, Drahomíra Kyjovská<sup>1</sup>, Martina Martonová<sup>1</sup>, Darja Páralová<sup>1</sup>, Tereza Klenková<sup>1</sup>, Soňa Kloudová<sup>1</sup>

<sup>1</sup>Reprofit International, Clinic of Reproductive Medicine

<sup>2</sup>Department of Histology and Embryology, Faculty of Medicine, Masaryk University



<https://www.youtube.com/watch?v=tIKo9nqmqGY>

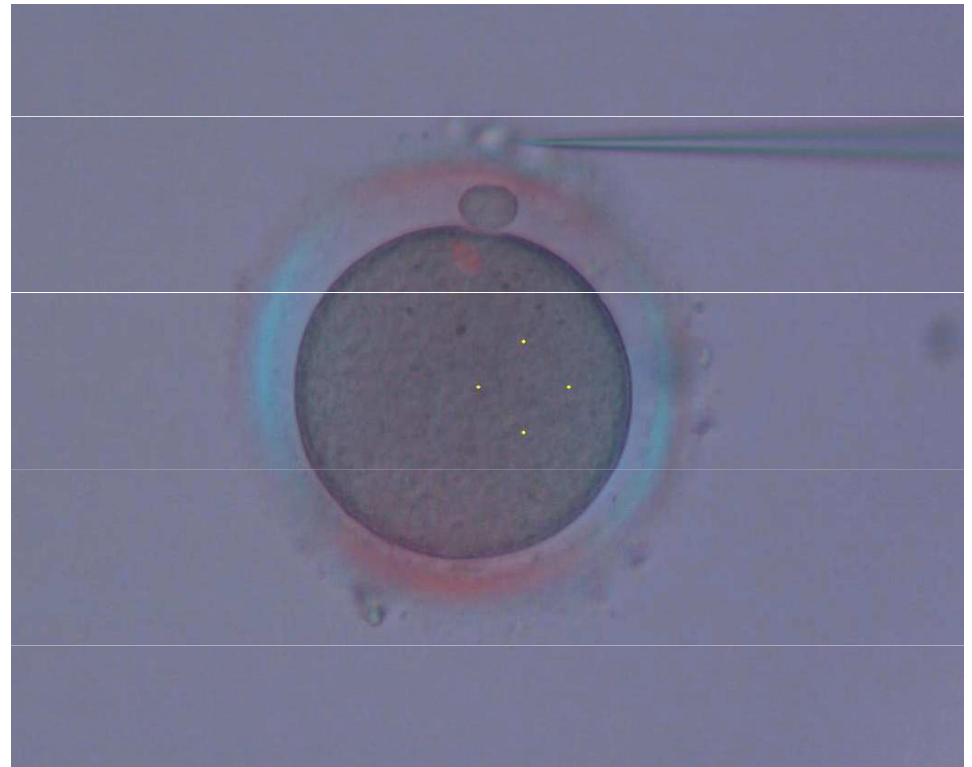
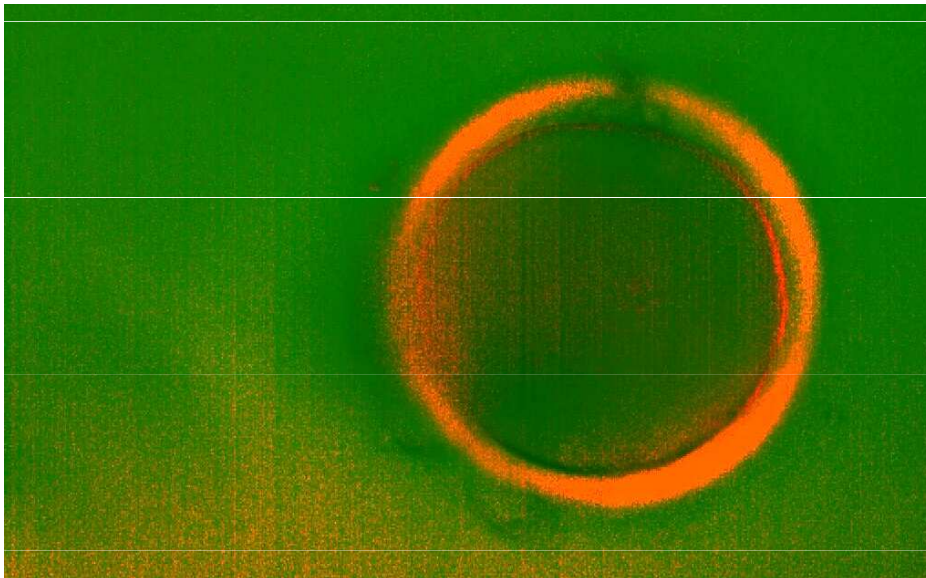




# Non-invasive spindle visualization

## ❖ Polarized Light Microscopy (PLM)

- PLM signal is orientation-dependent
- spindle imaging requires oocyte orientation

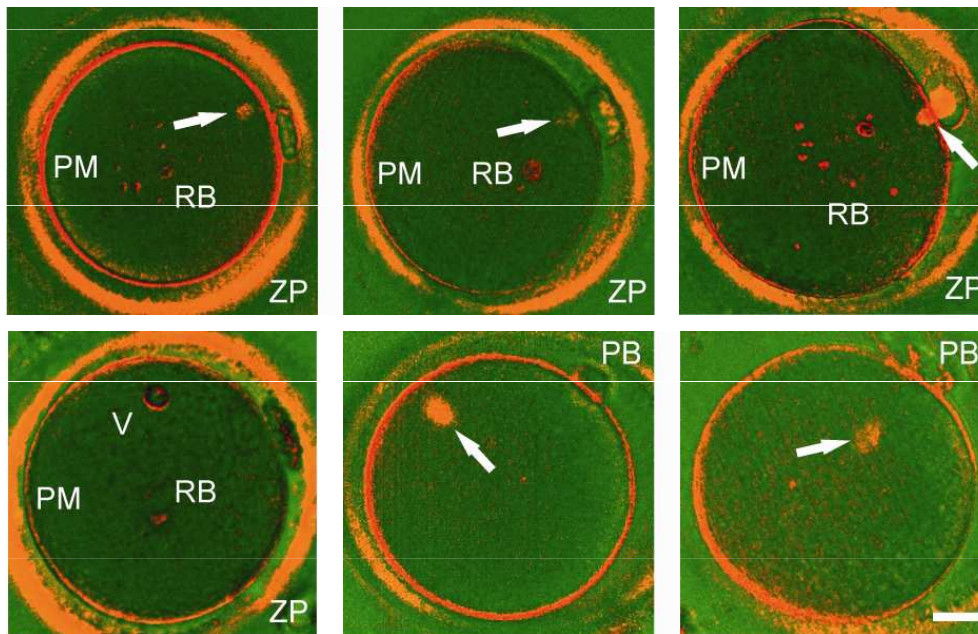




# Non-invasive spindle visualization

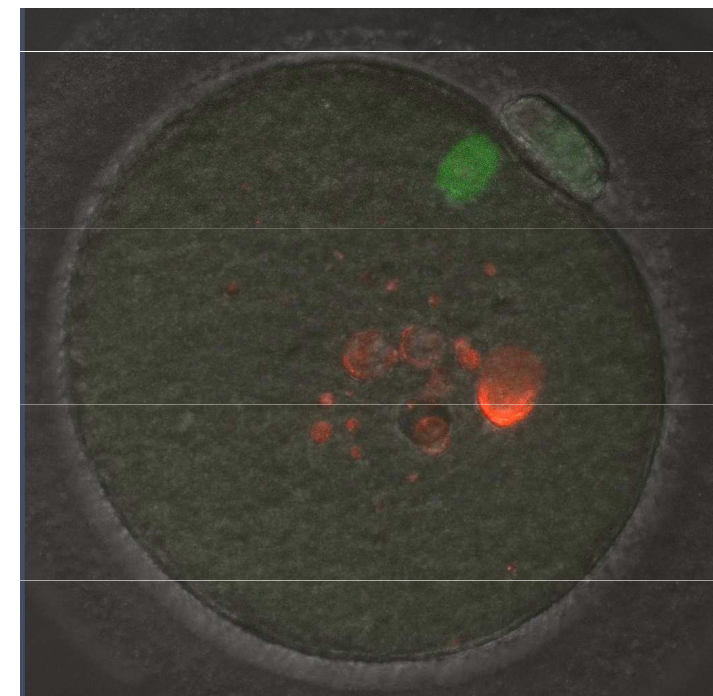
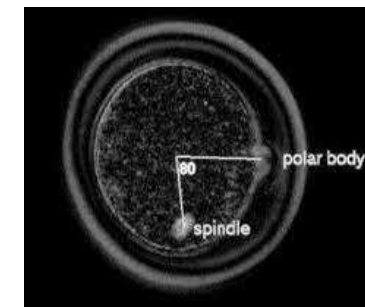
## ❖ Polarized Light Microscopy (PLM)

- birefringent structures in human oocytes



PM.... Plasma mebrane  
ZP.....Zona pellucida  
RB.....Refractile body  
V.....Vacuole

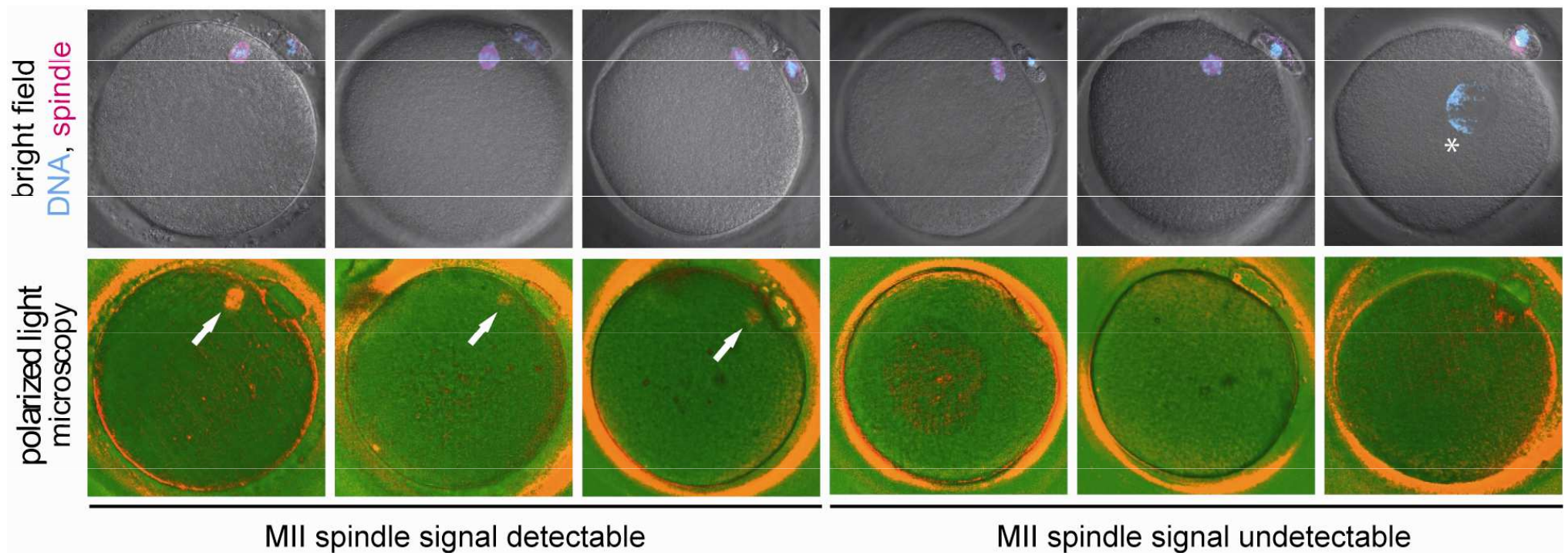
- relative position of spindle and PB



# Non-invasive spindle visualization

## ❖ Polarized Light Microscopy (PLM)

- the strength of the signal reflect the material ordering
- sufficient mass of paralelely oriented MT required to produce noticeable signal

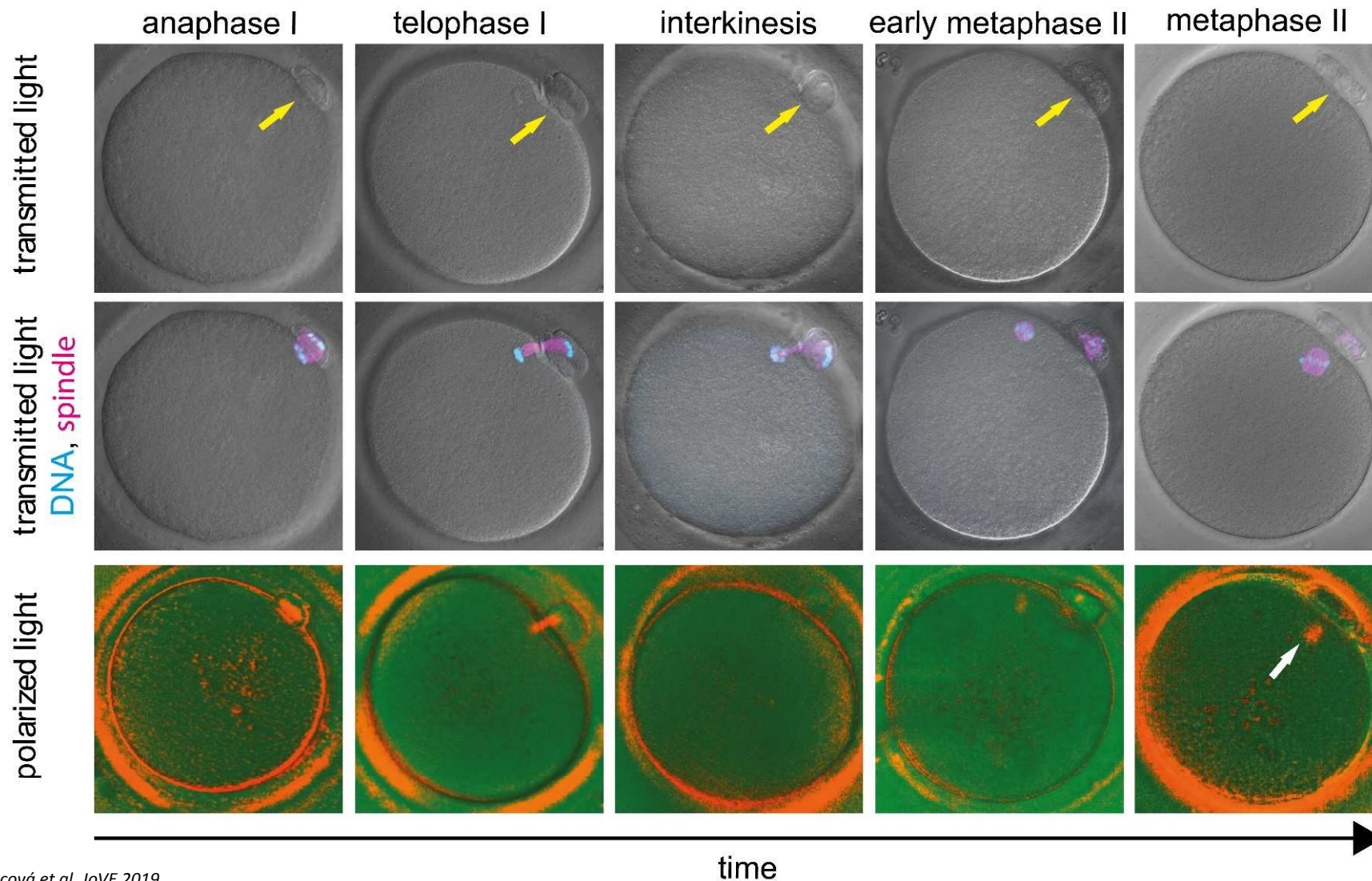




# Non-invasive spindle visualization

## ❖ Polarized Light Microscopy (PLM)

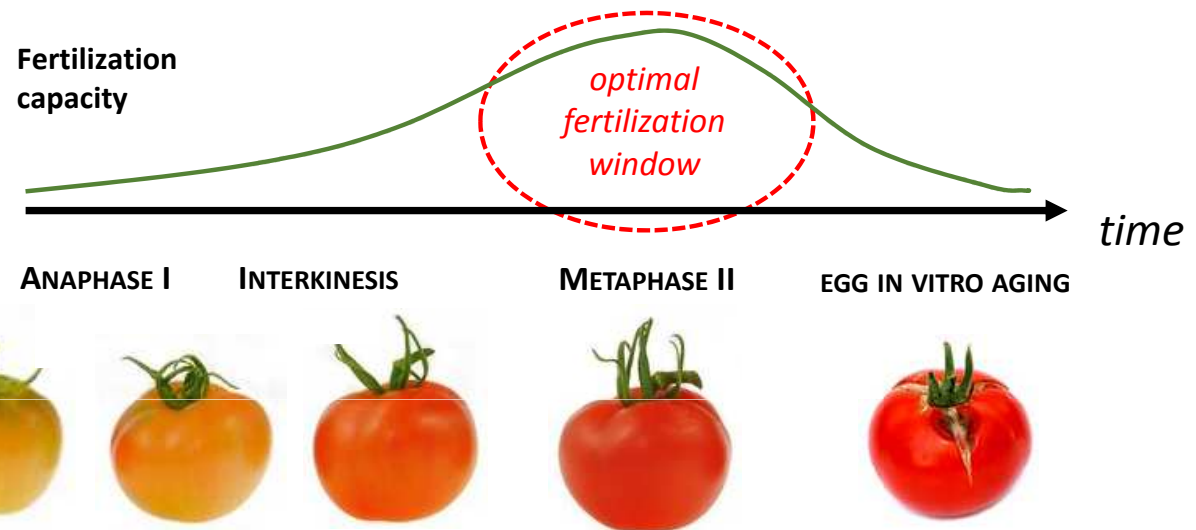
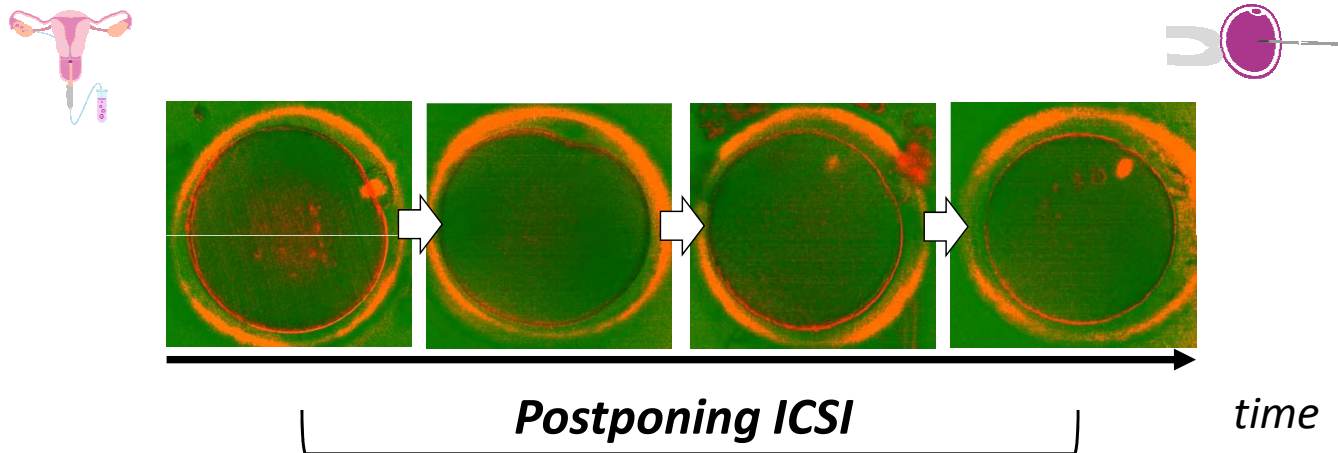
- enables monitoring of MI/MII transition and ICSI time optimisation in clinical practice



# Non-invasive spindle visualization

## ❖ Polarized Light Microscopy (PLM)

- enables monitoring of MI/MII transition and ICSI time optimisation in clinical practice





# Factor affecting human oocyte spindle stability in vitro

## ❖ MII spindle is sensitive

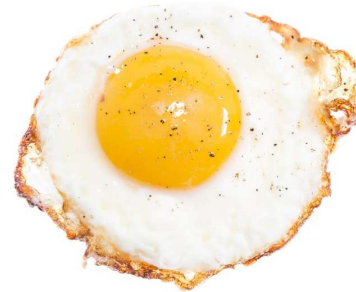
### ➤ temperature

- optimal 37°C



### OVERHEATING

→ irreversible denaturation

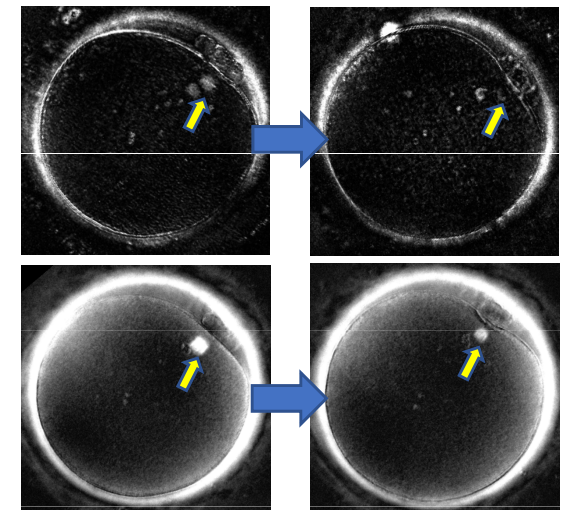


### COOLING

→ spindle desintegration



10 min RT

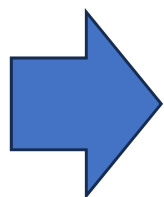


### ➤ osmolarity alterations

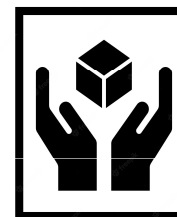
- avoid evaporation
- parafine/mineral oil overlay
- humid conditions

### ➤ pH fluctulation

- MOPS/HEPES buffered medium for work in ambient conditions



avoid excessive manipulation !



HANDLE WITH CARE