

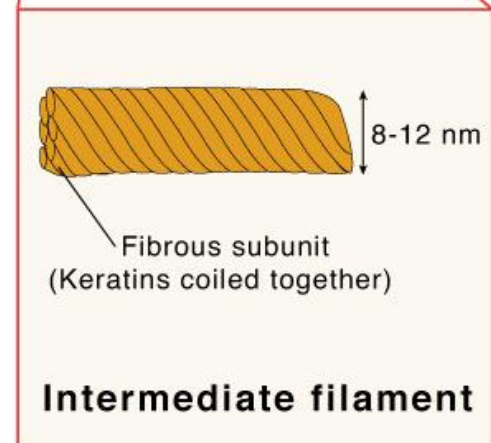
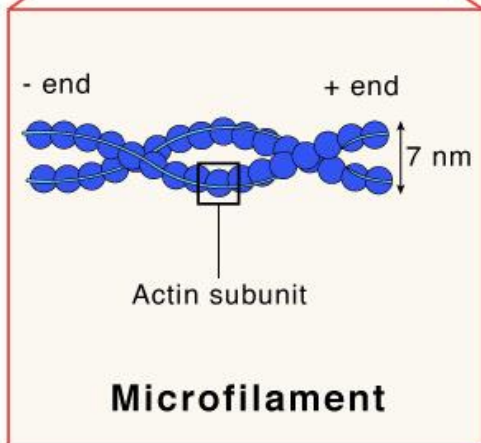
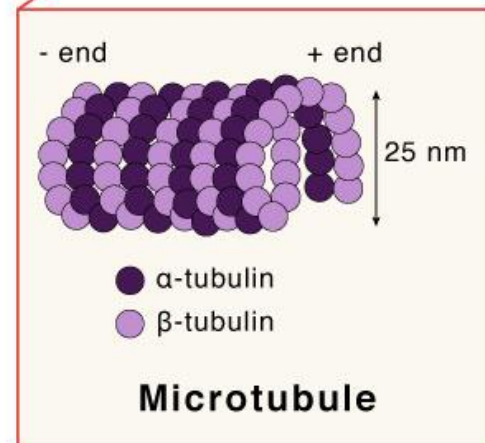
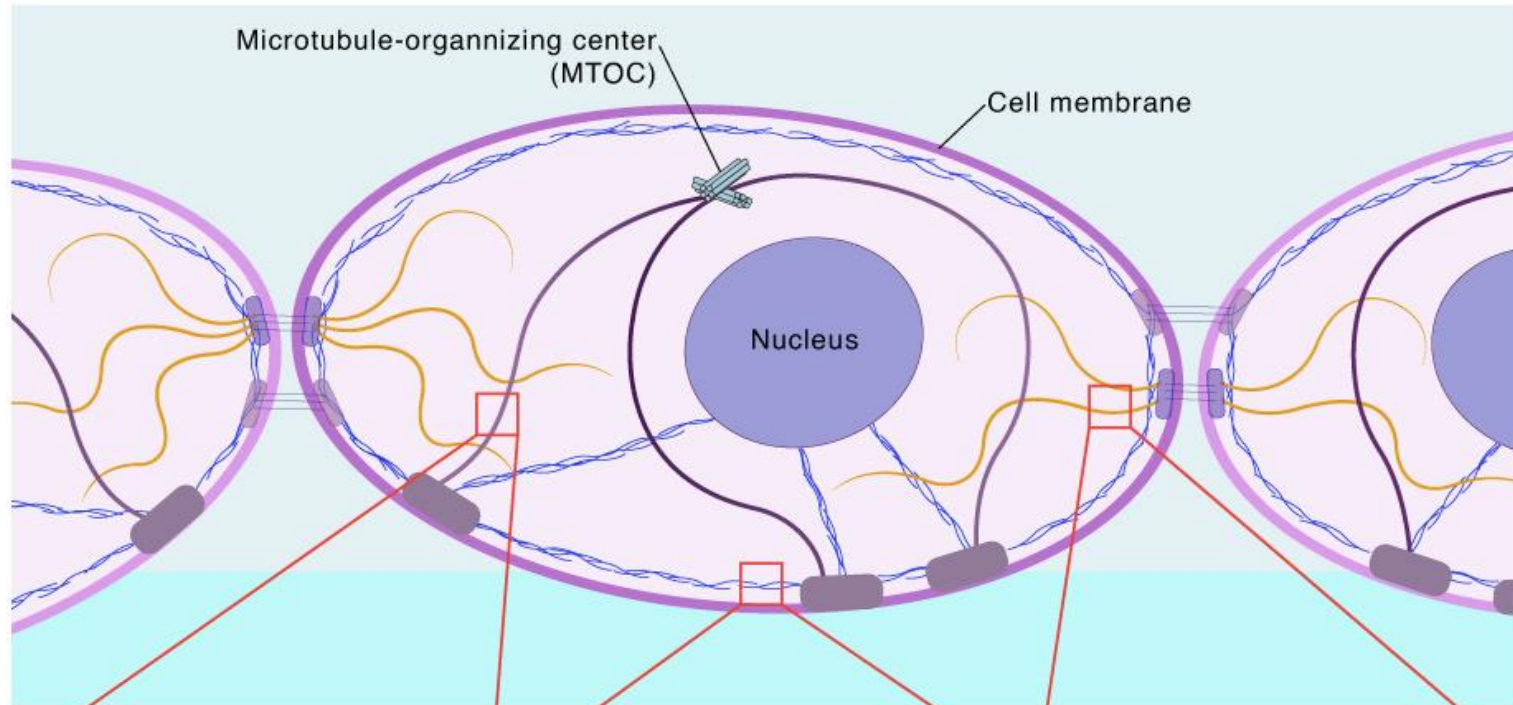
Embryologie I OOGENESIS

autumn 2024

Oocyte cytoskeleton

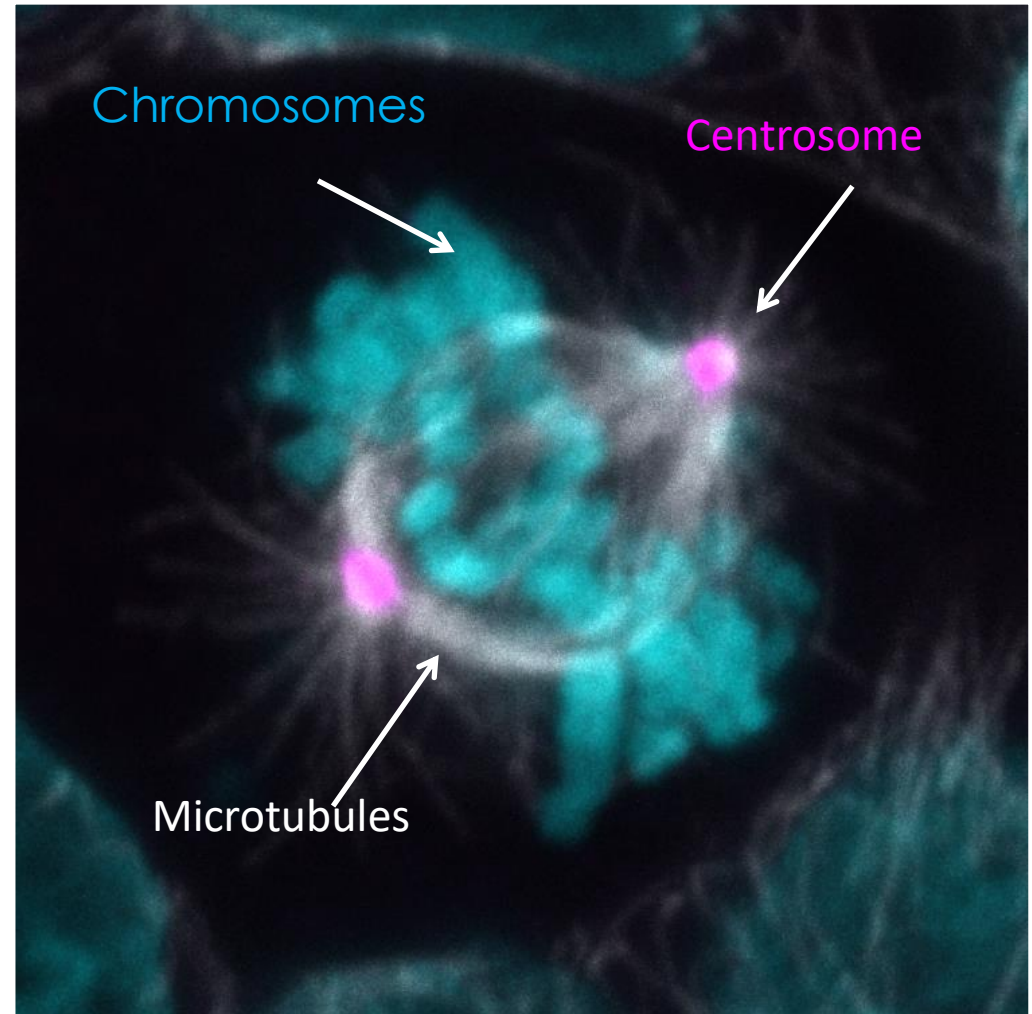
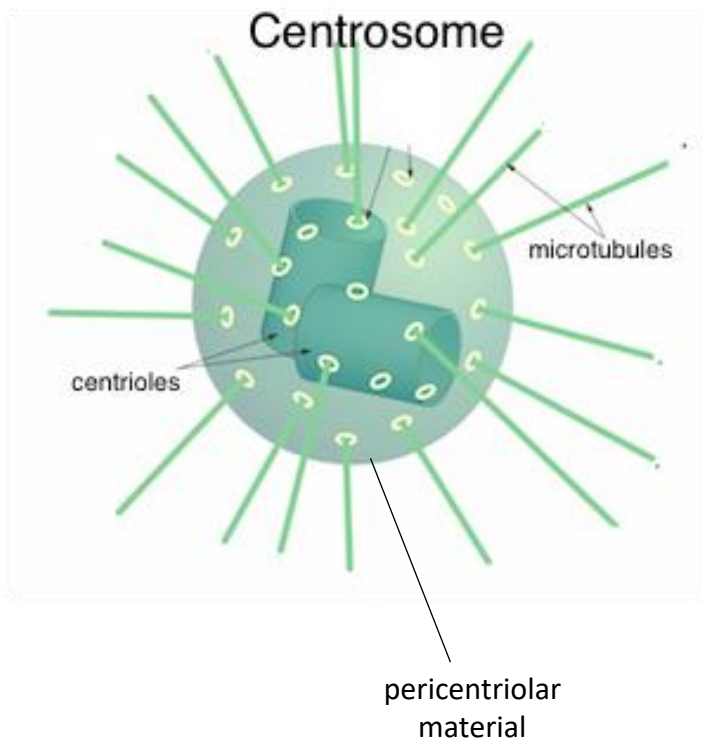
Zuzana Holubcová
Department of Histology and Embryology
zholub@med.muni.cz

Cytoskeleton



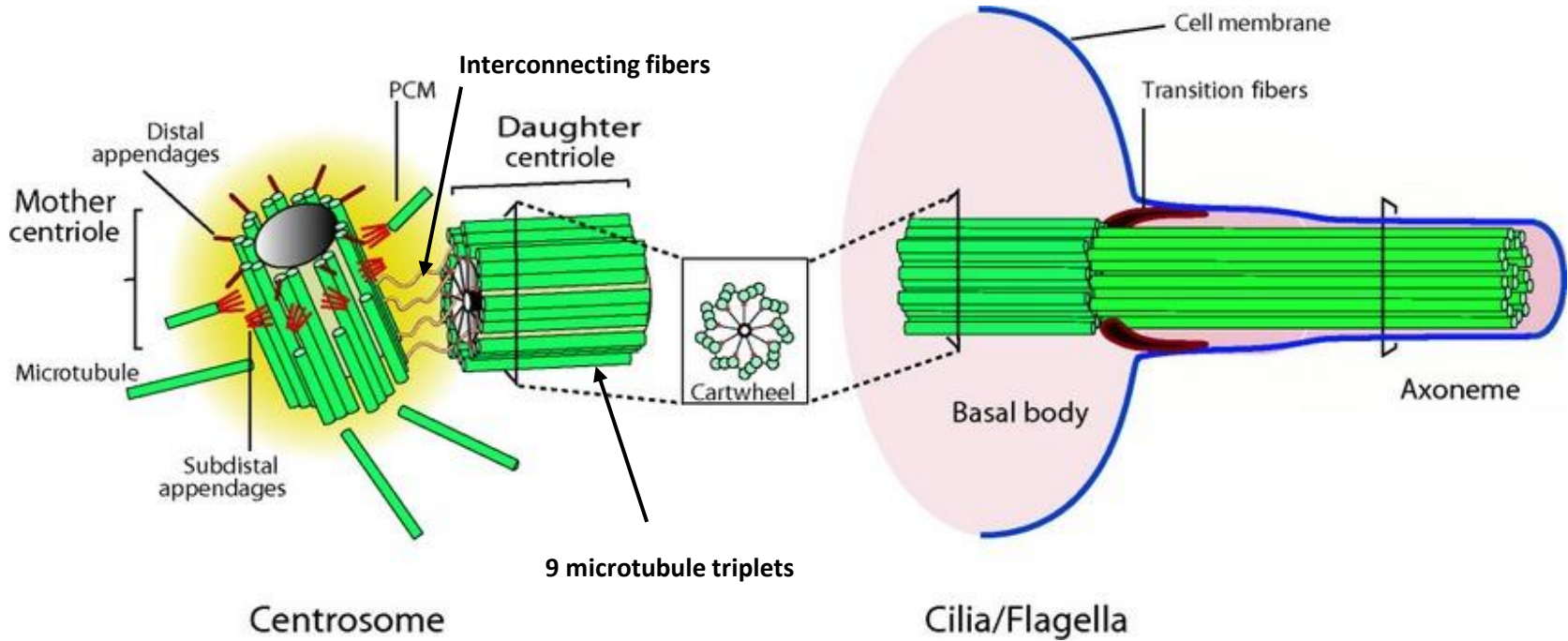
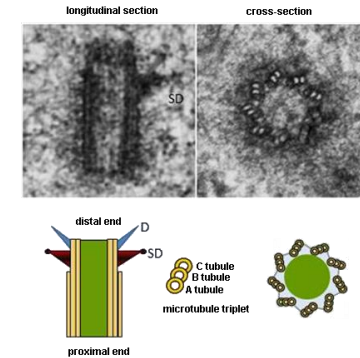
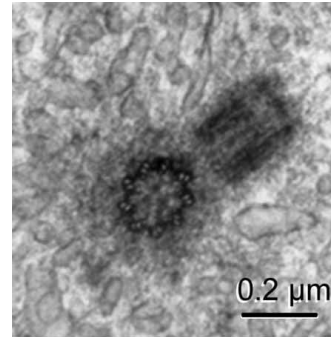
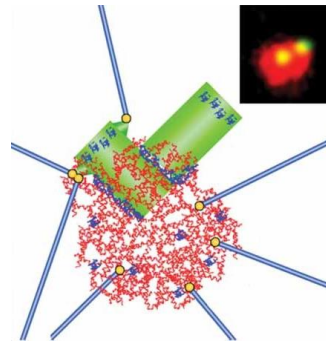
Centrosome

Centrosome = major Microtubule Organising Center (MTOC) in animal cells



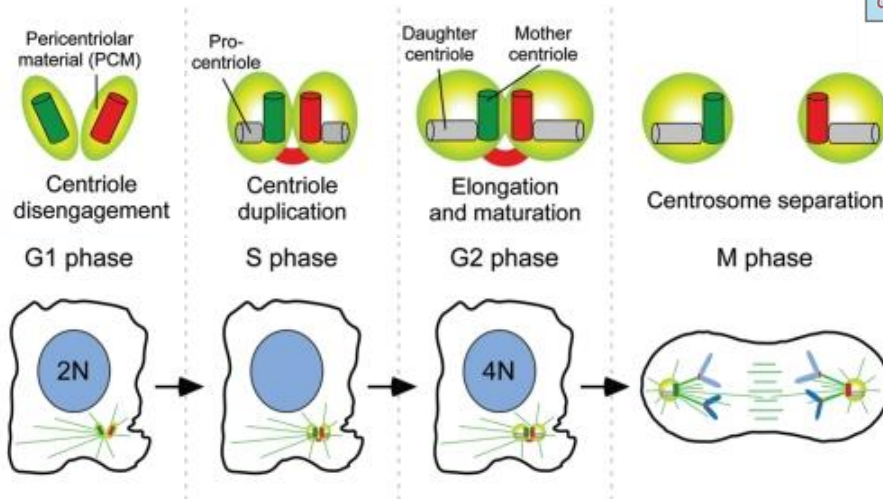
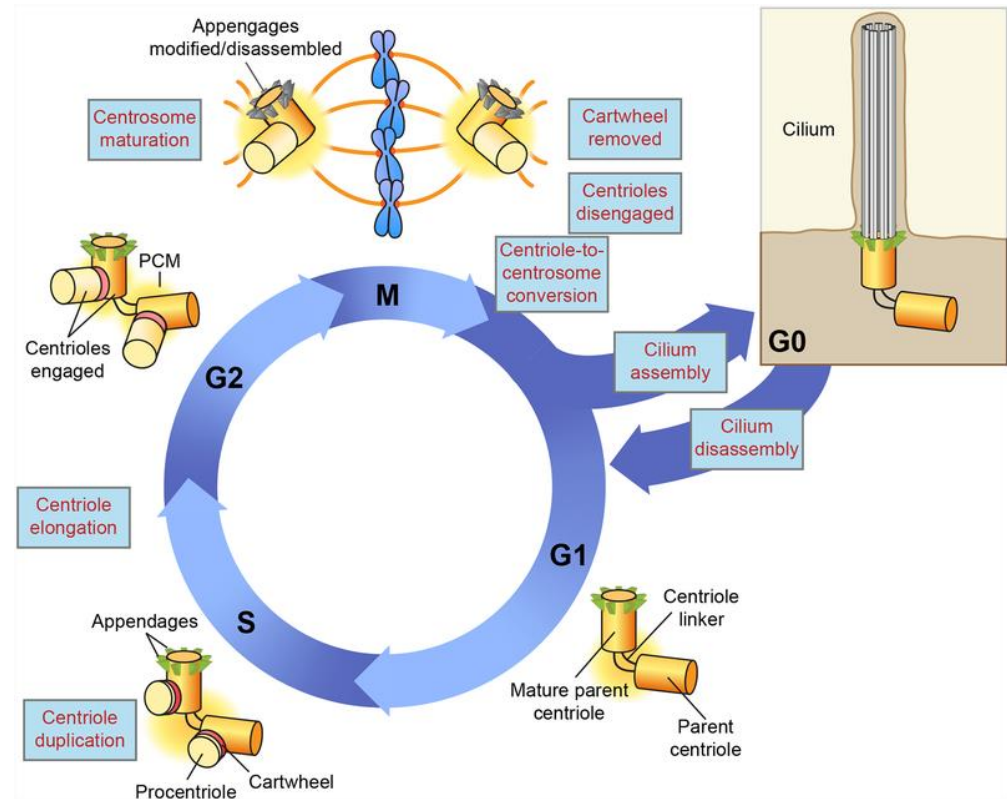
Centrosome

➤ Centrioles



Centrosome

➤ Centrosome duplication cycle

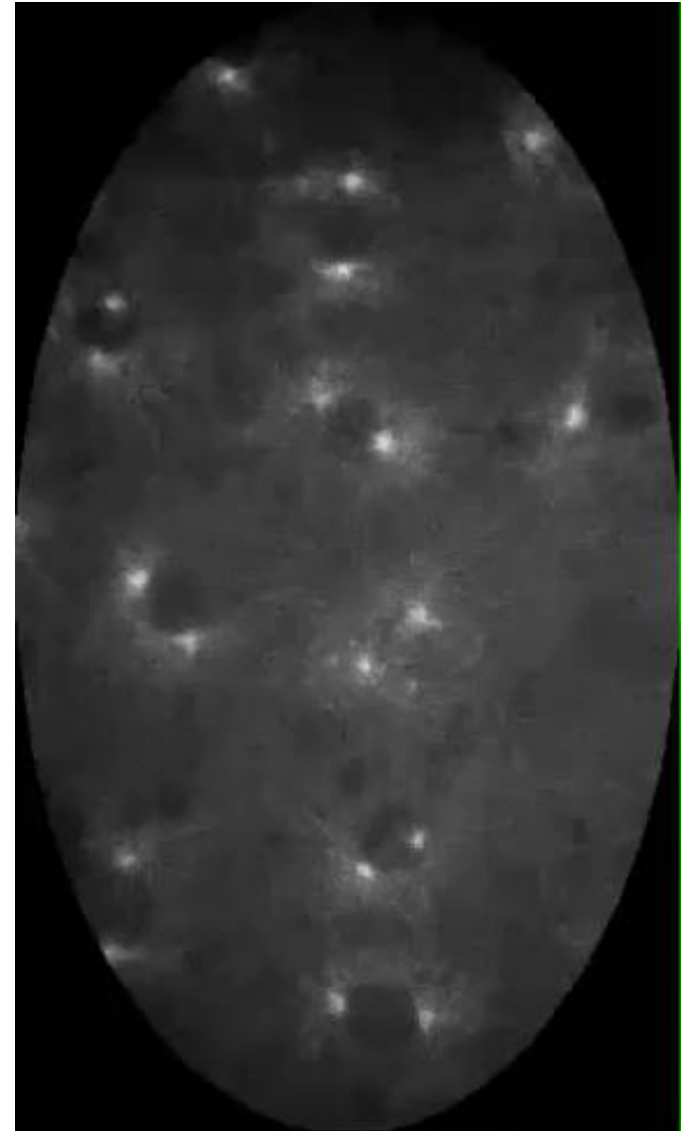
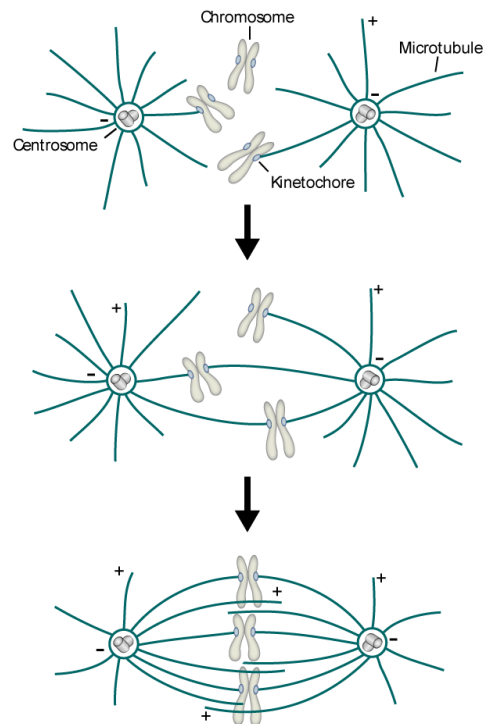


Centrosomes duplicate in coordination with DNA synthesis

Centrosome

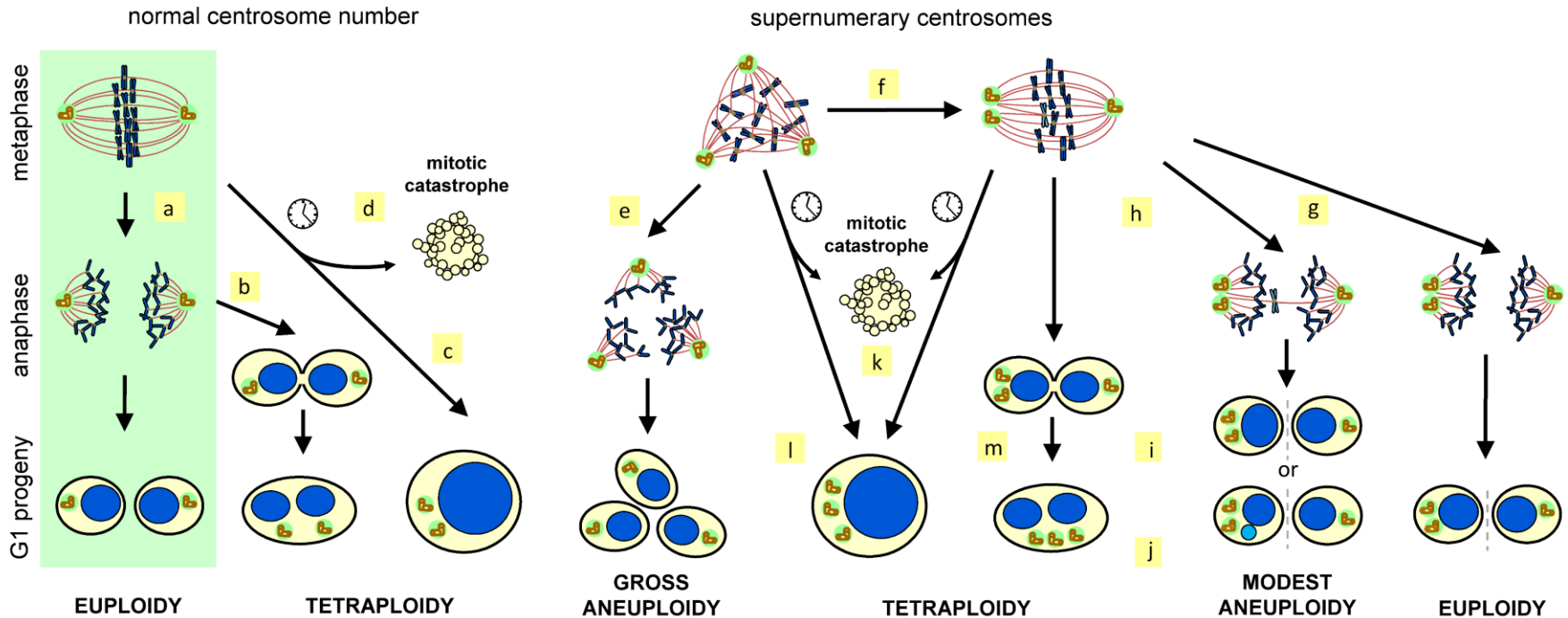
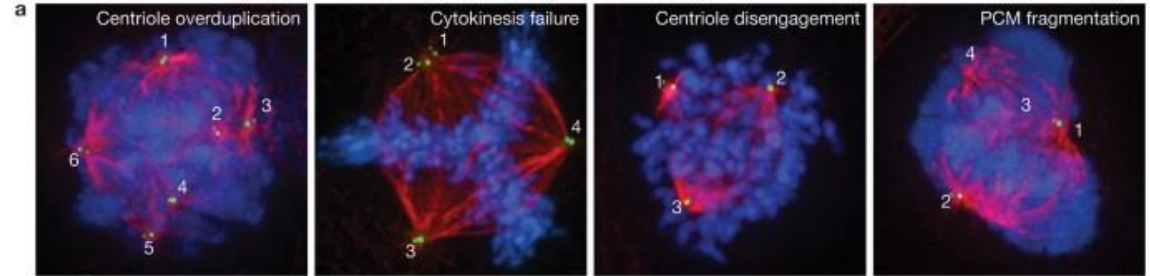
In animal somatic cells, centrosomes

- drive microtubule (MT) nucleation
- focus microtubule (-)ends at spindle poles and stabilize spindle poles
- assemble central bipolar spindle that evenly segregate sister chromatids during mitosis



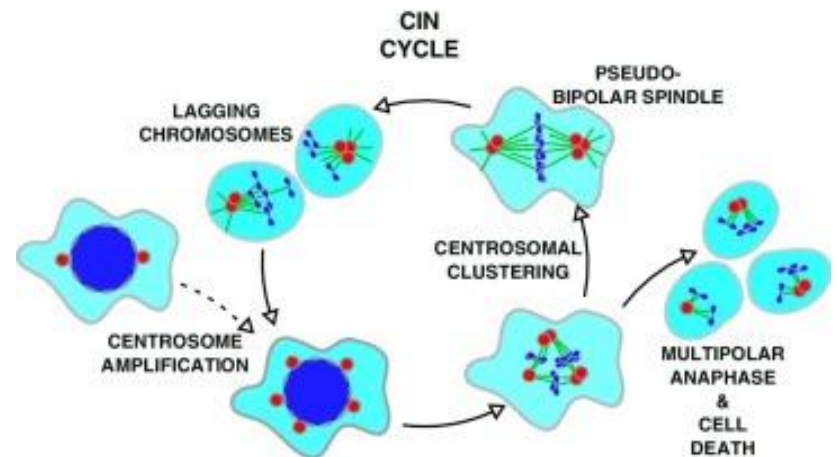
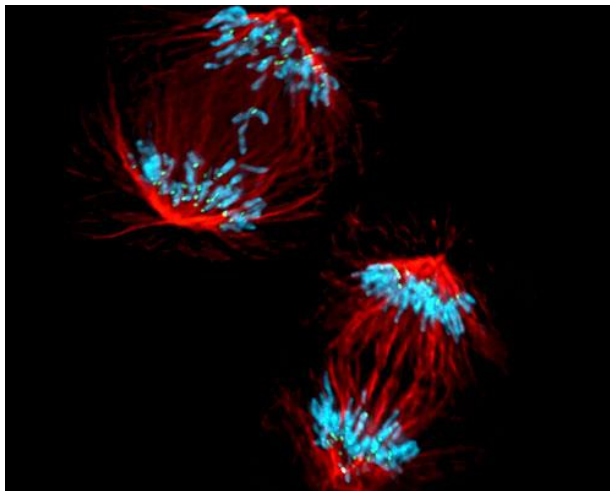
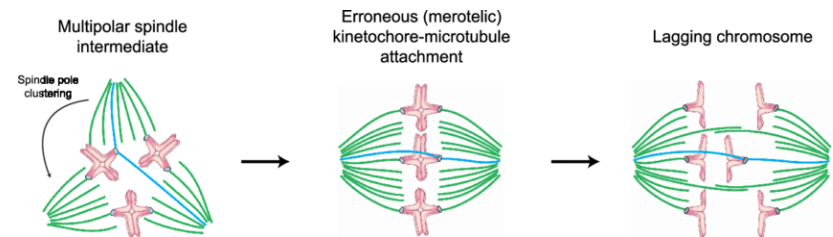
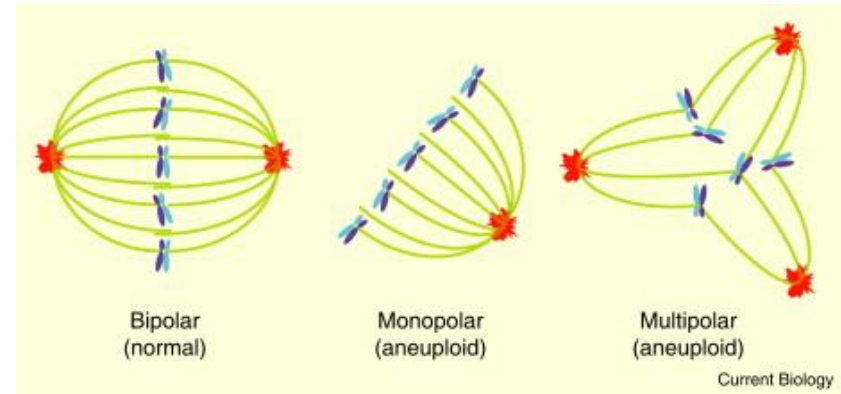
Centrosome overamplification

- occurs in cancer cells
- promotes genetic instability
- acentriolar centrosomes (only PCM) capable to nucleate and capture microtubules



Centrosomes define spindle geometry

- overamplification of centrosomes generates multipolar spindle which produces gross aneuploidy
- clustering of centrosomes enables bipolarization but persisting merotelic attachments favour chromosome lagging during anaphase and create risk of chromosome missegregation



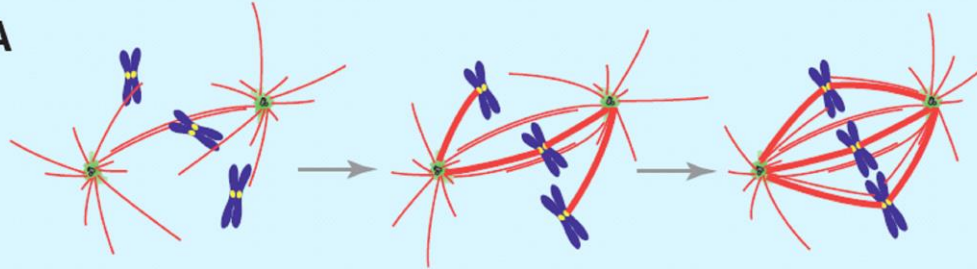
Microtubule nucleation pathways

Centrosomes

present



A

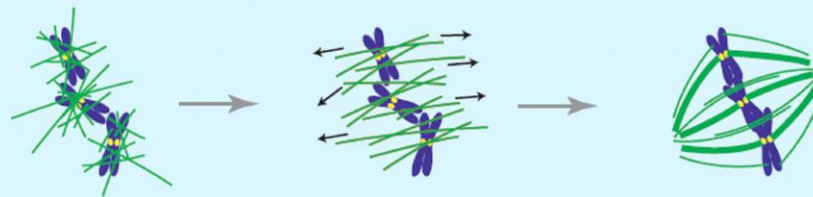


Search and capture

absent



B



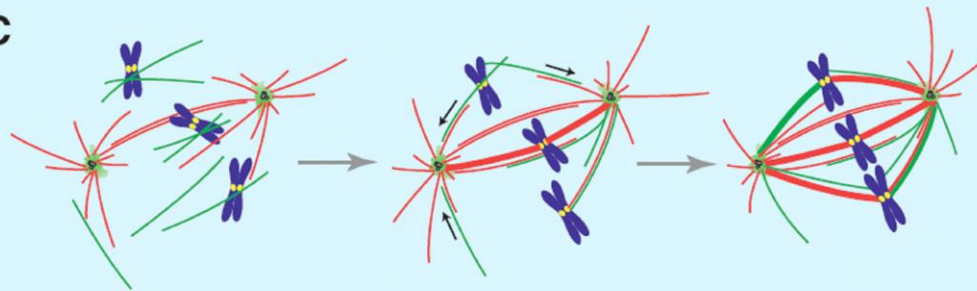
Self organization

- plant cells

present



C



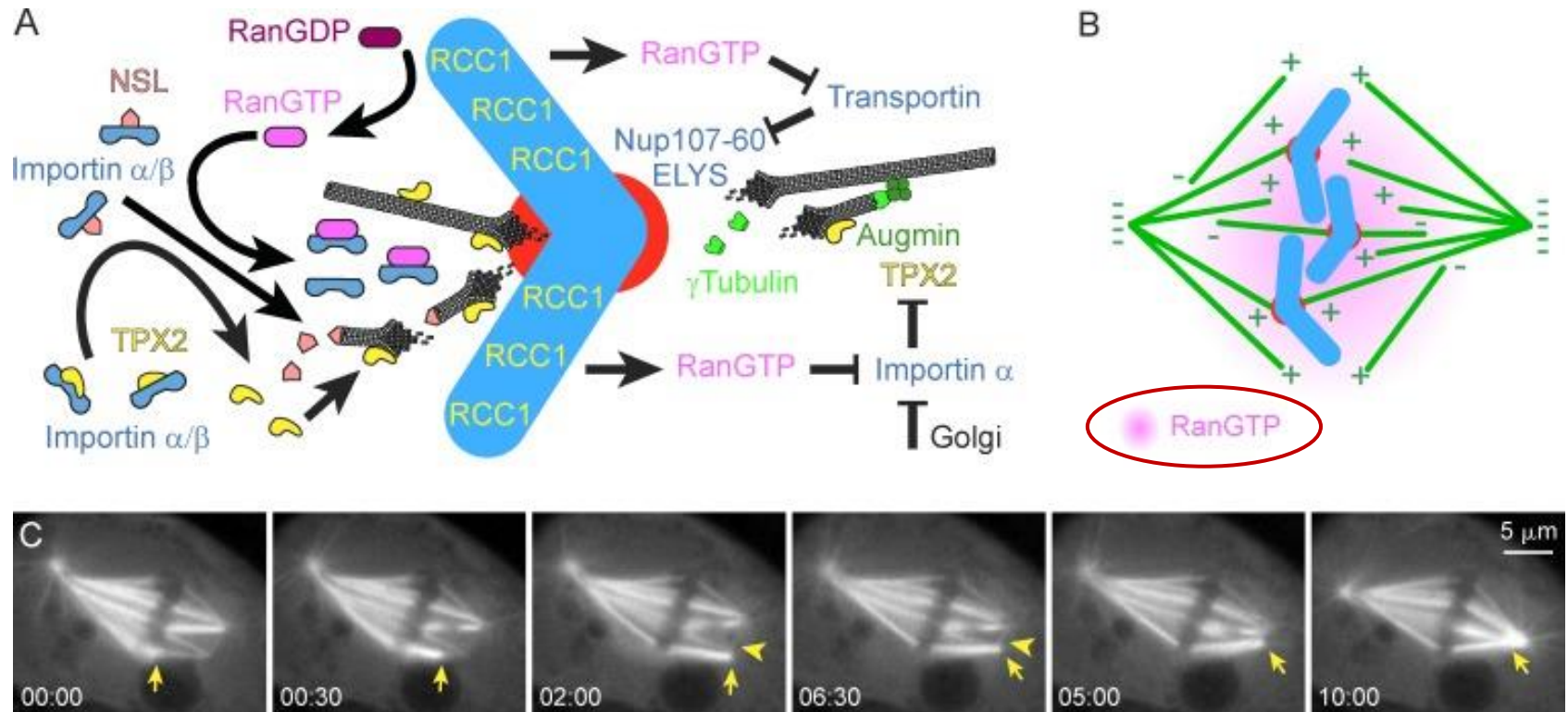
Combined

- animal mitotic cells

Microtubule nucleation pathways

❖ Chromatin-driven microtubule (MT) nucleation

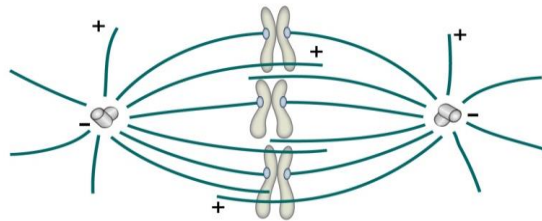
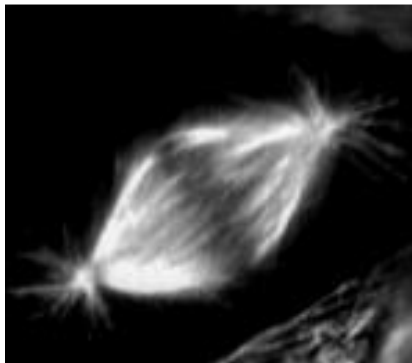
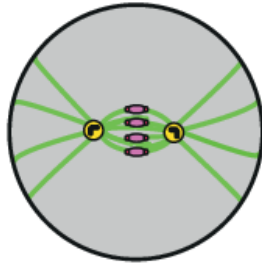
- **RanGTP** gradient promotes both de novo MT nucleation near kinetochores and amplification of MT growth toward chromosomes



Female meiotic spindles lack centrosomes

Mitosis

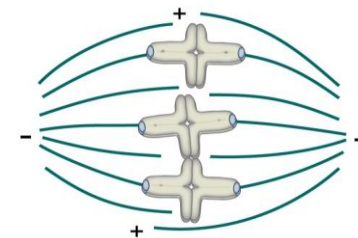
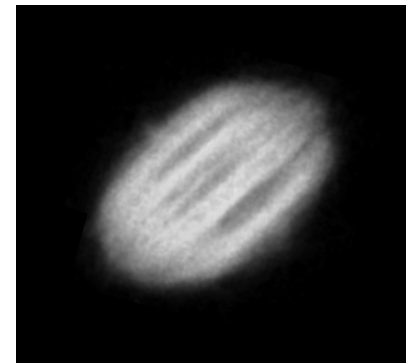
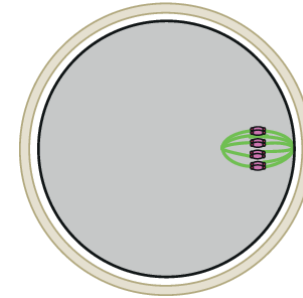
Symmetric division



Centrosomal spindle

Female meiosis

Asymmetric division

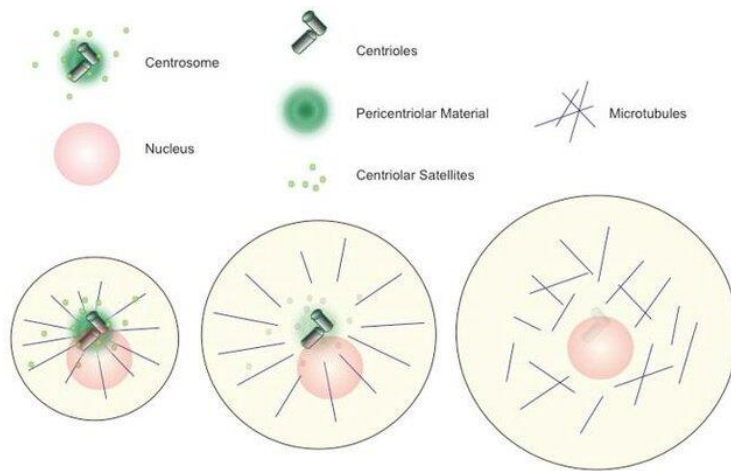


Acentrosomal spindle

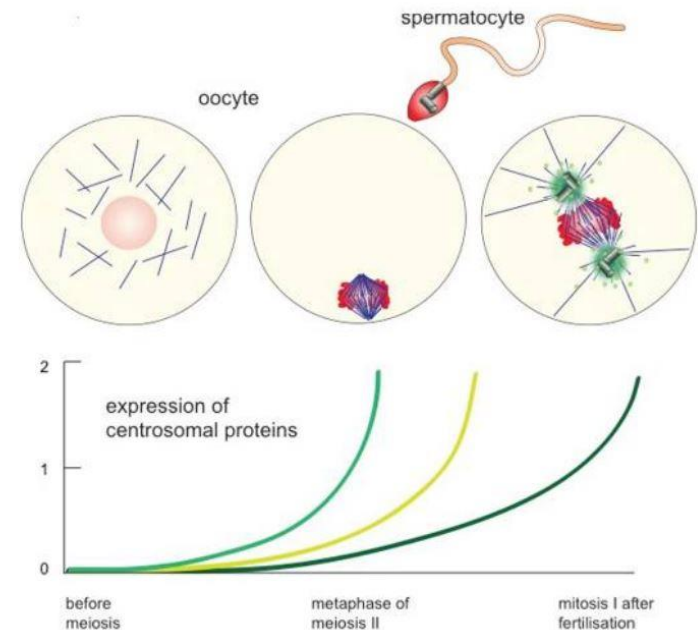
Centrioles are eliminated during oogenesis

Metazoan oocytes eliminate centrosomes during oogenesis in order to

- (1) ensure highly asymmetric cell division
- (2) avoid a superior number after fertilisation



- PCM synthesized during oocyte maturation
- **Centrioles paternally inherited**

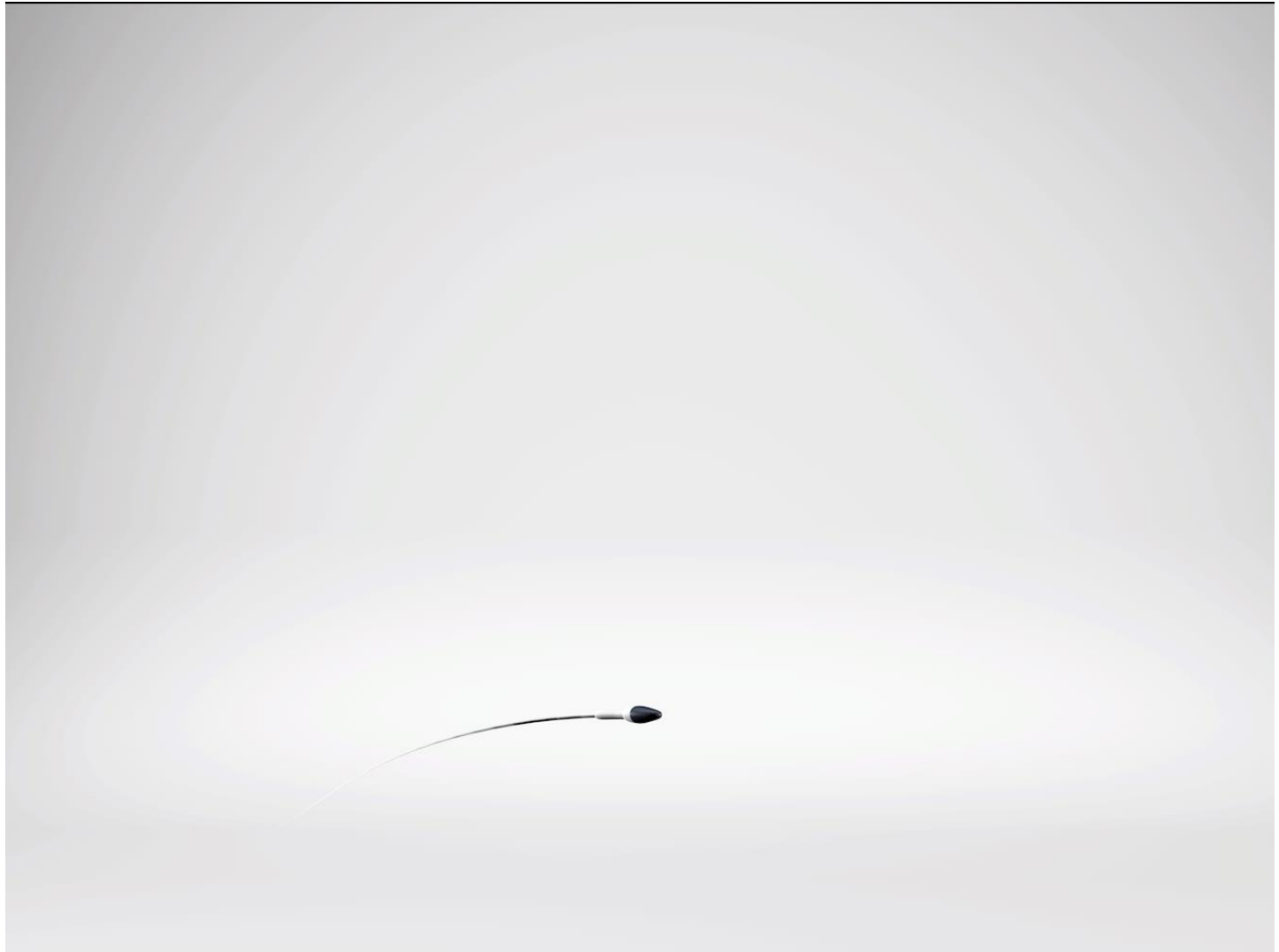
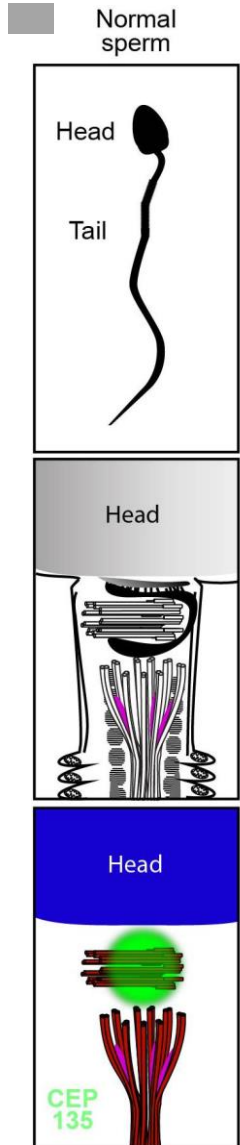


- sperm-derived centrioles recruit maternal PCM after fertilization to assemble first mitotic spindle



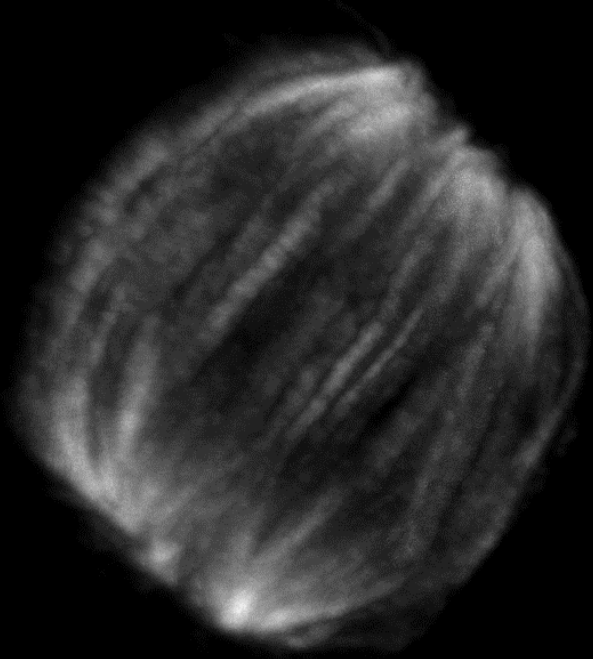
- sperm-derived centrioles are destroyed
- first mitosis with acentrosomal spindle
- de novo centriole assembly during embryo cleavage stage

Centrioles are delivered by sperm during fertilization

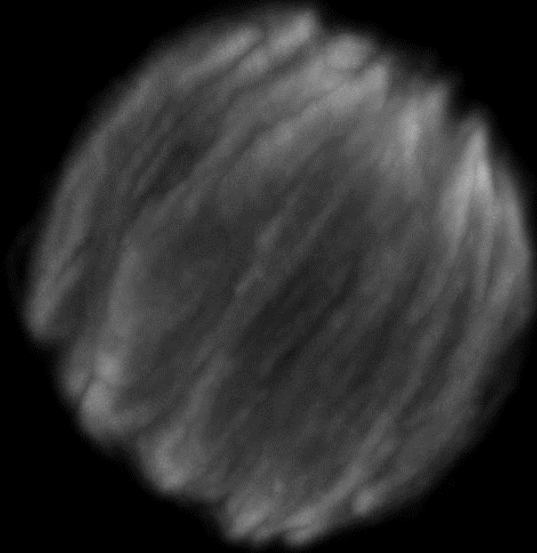


Human oocyte spindle lacks centrosome

Metaphase I



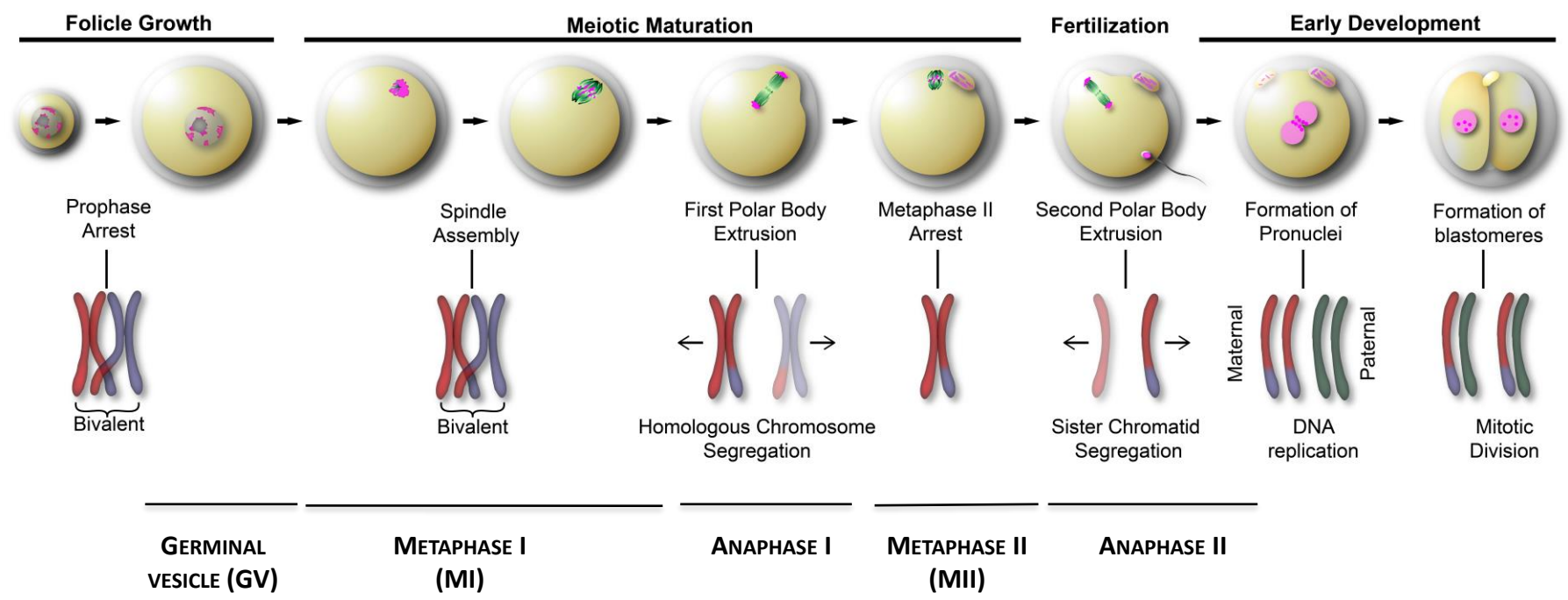
Metaphase II



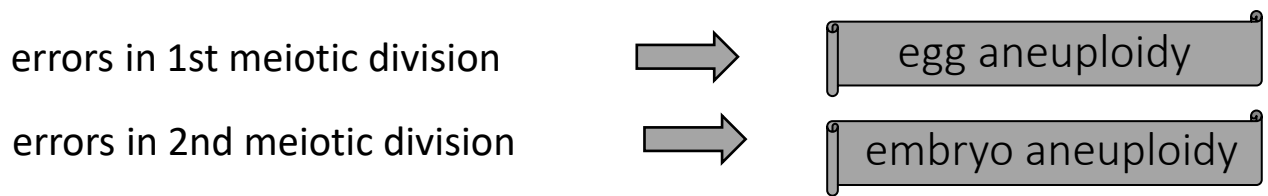
5 μ m

How are meiotic spindle poles assembled
in the absence of centrosomes?

Acentrosomal spindle drives chromosomal segregation during female meiosis



Functional spindle is required for chromosome segregation fidelity



Acentrosomal spindle assembly in mouse oocytes

- high-resolution confocal live cell imaging of mouse oocytes maturing in vitro showed that mouse oocyte spindle is assembled by **multiple small acentriolar MTOCs that functionally replace canonical centrosomes**

Cell

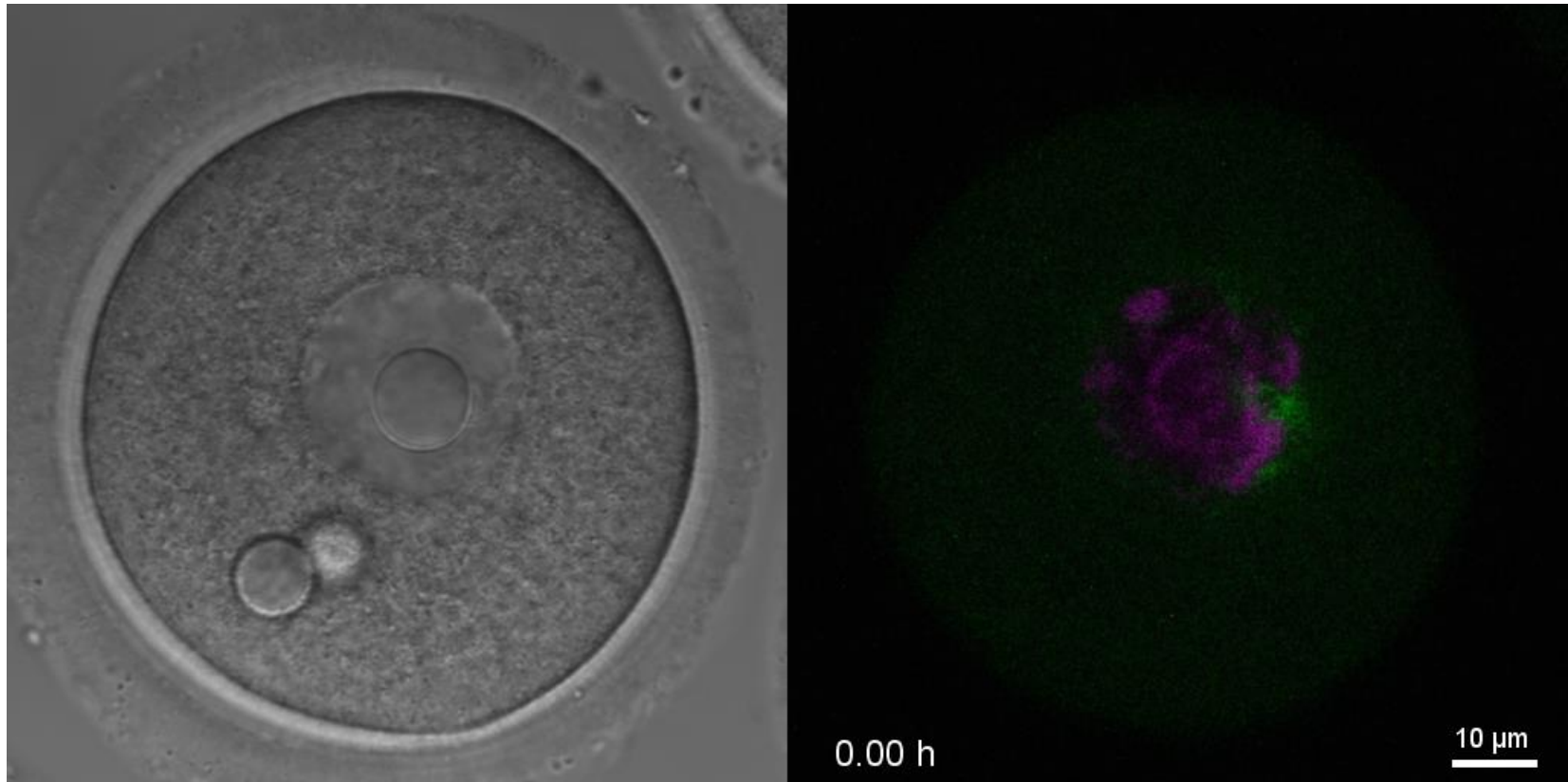
Schuh and Ellenberg, 2007

Self-Organization of MTOCs Replaces Centrosome Function during Acentrosomal Spindle Assembly in Live Mouse Oocytes

Melina Schuh¹ and Jan Ellenberg^{1,*}
¹Gene Expression Unit, European Molecular Biology Laboratory (EMBL), Meyerhofstrasse 1, D-69117 Heidelberg, Germany
*Correspondence: jan.ellenberg@embl.de
DOI: 10.1016/j.cell.2007.06.025



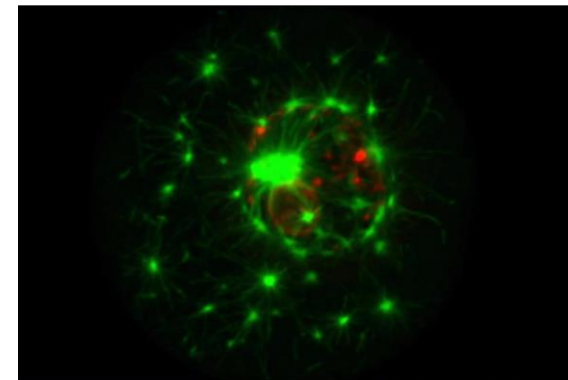
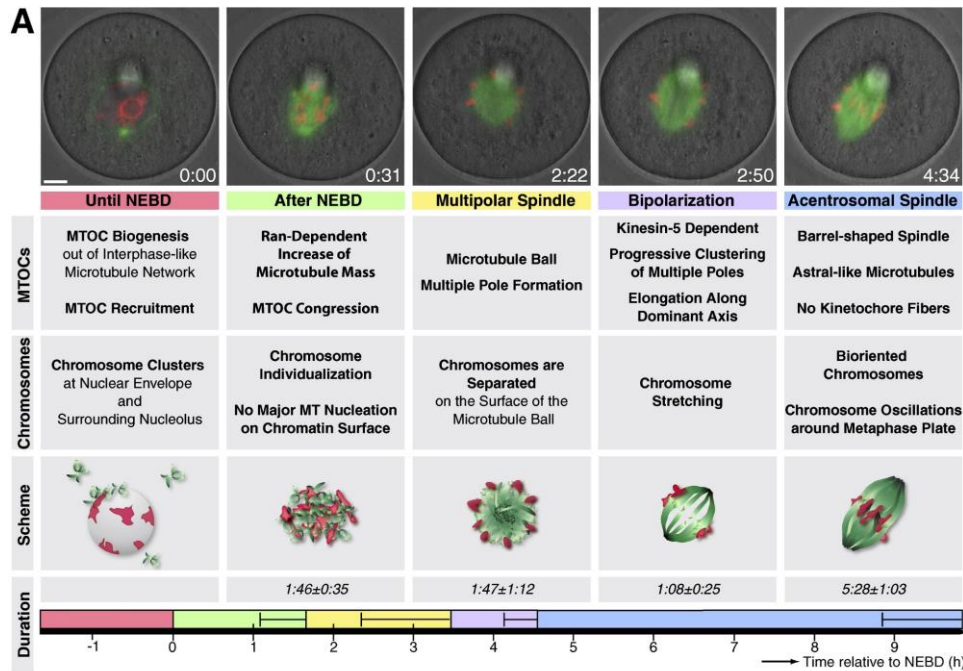
Melina Schuh



DNA

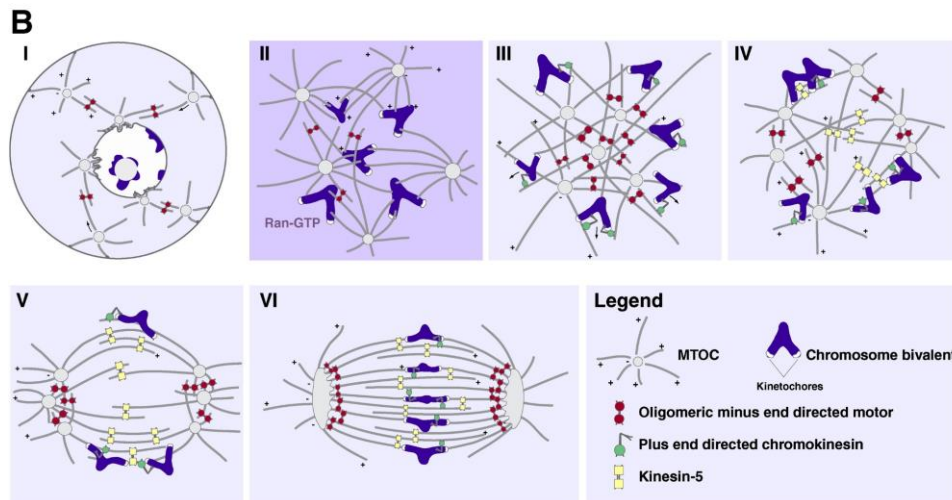
microtubules

Acentrosomal spindle assembly in mouse oocytes

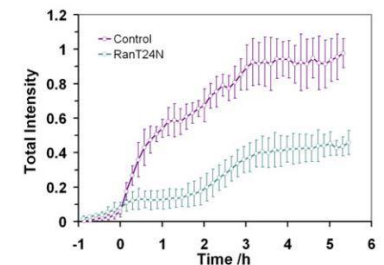


Prophase microtubule network with low dynamics

- MTOC consists of PCM proteins (pericentrin, γ -tubulin, Cep192, Cep120, Cep 125, NEDD1,..)
- MTOCS cluster around nucleus before NEBD
- MTOC nucleate MT „ball“ which carries chromosomes on its surface
- MT mass elongates and chromosome congress
- chromosome alignment after spindle bipolarization
- spindle migration to the cortex

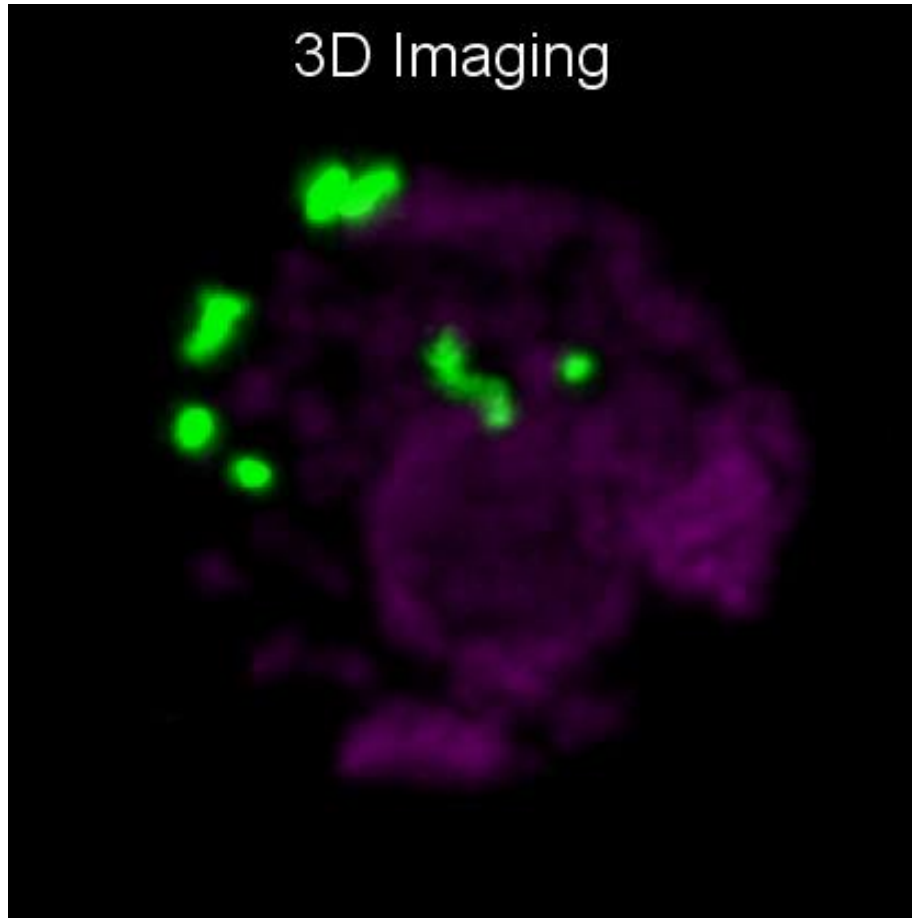


- Ran activity overdriven by coordinated action of MTOCS



Acentrosomal spindle assembly in mouse oocytes

Multipleacentriolar MTOCs converge at spindle poles and stabilize them



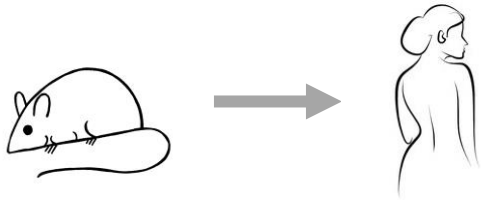
MTOC 3D reconstruction



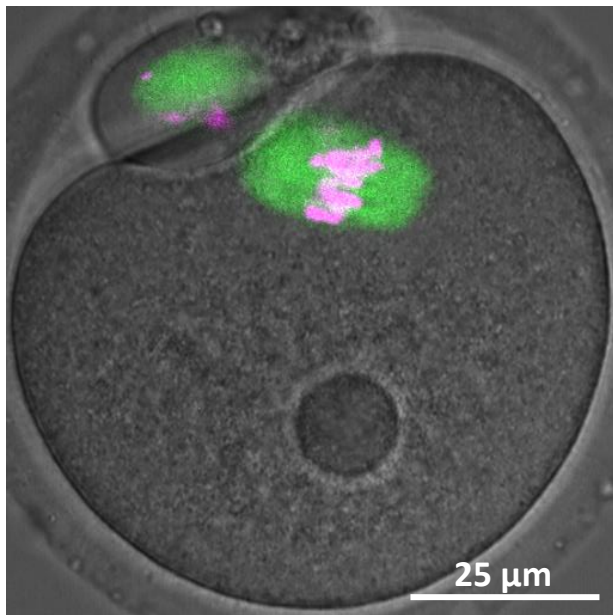
-01:00

DNA
MTOCs

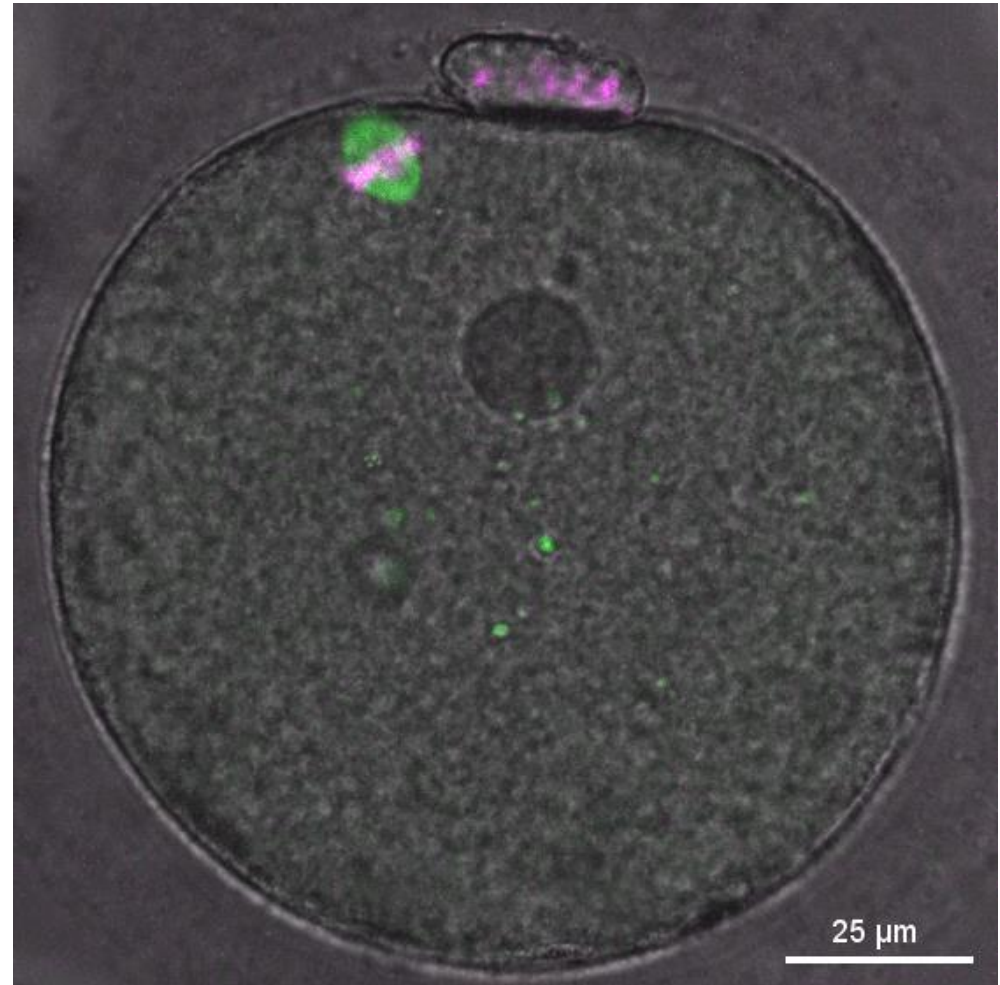
From mice to human



Mouse oocyte



Human oocyte

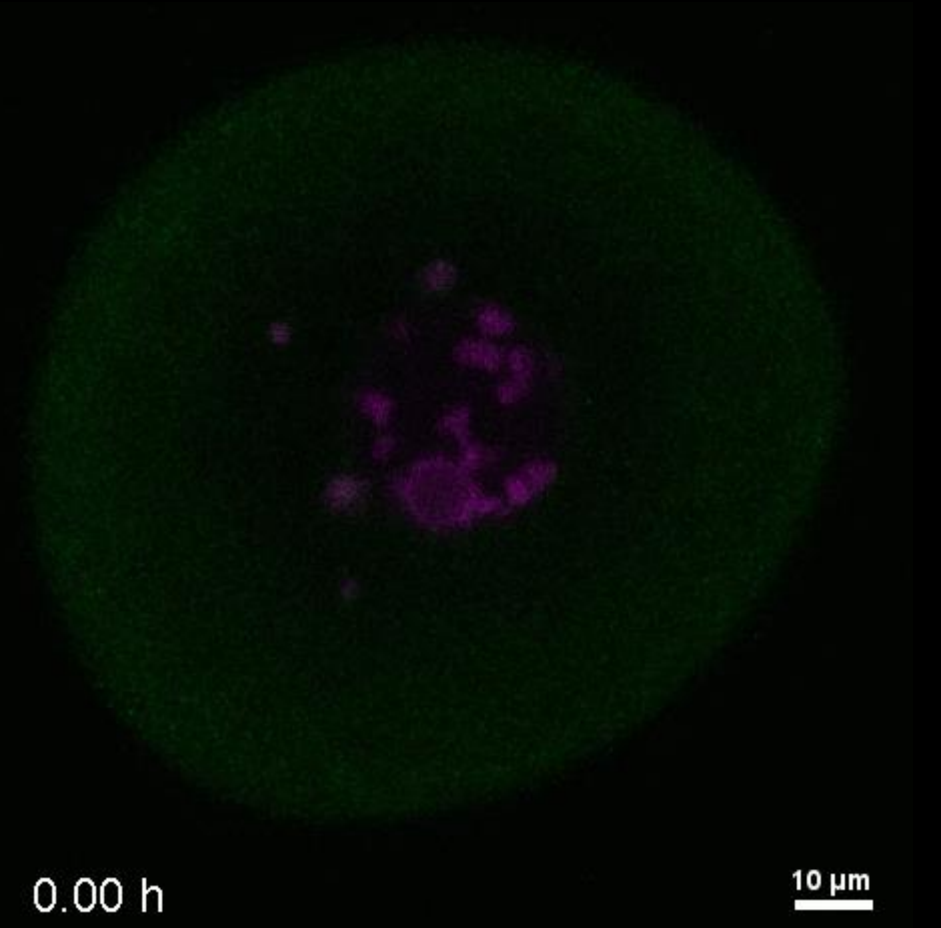


Chromosomes
Microtubules

Acentrosomal spindle assembly in human oocytes



DIC (transmitted light)



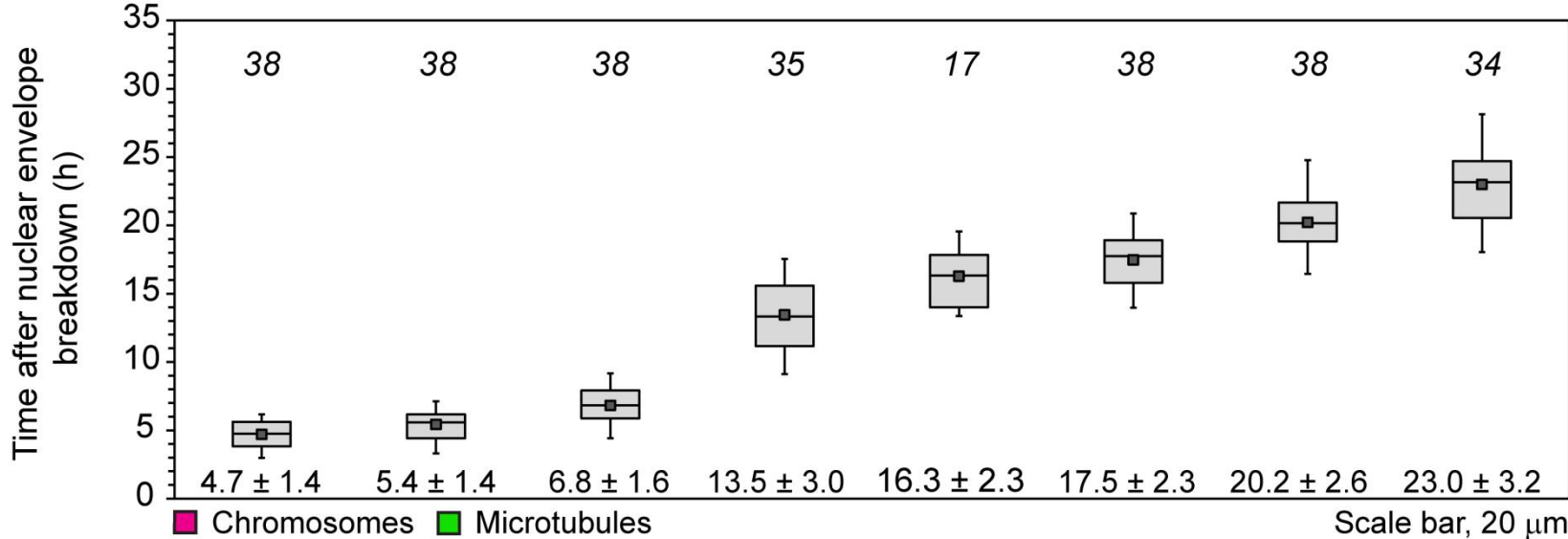
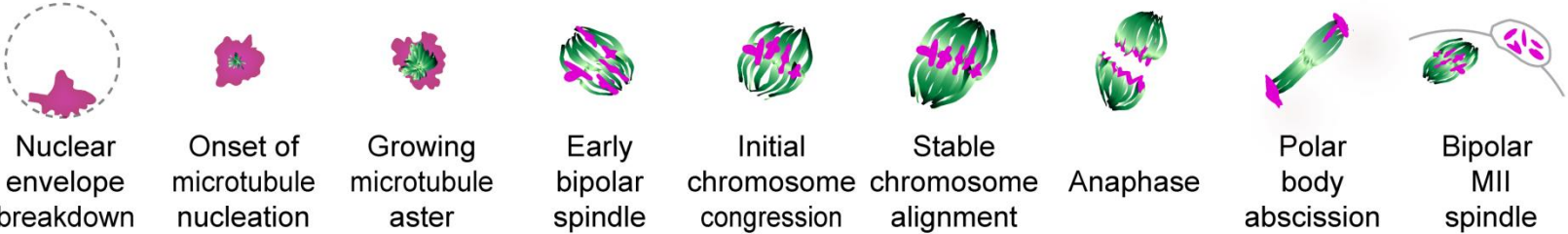
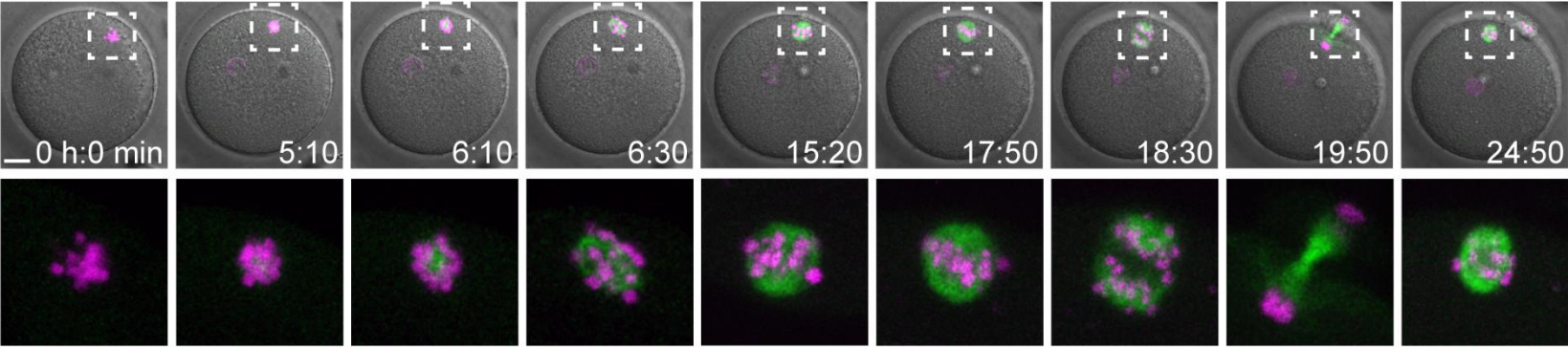
0.00 h

10 μm

Chromosomes (H2B-mRFP)

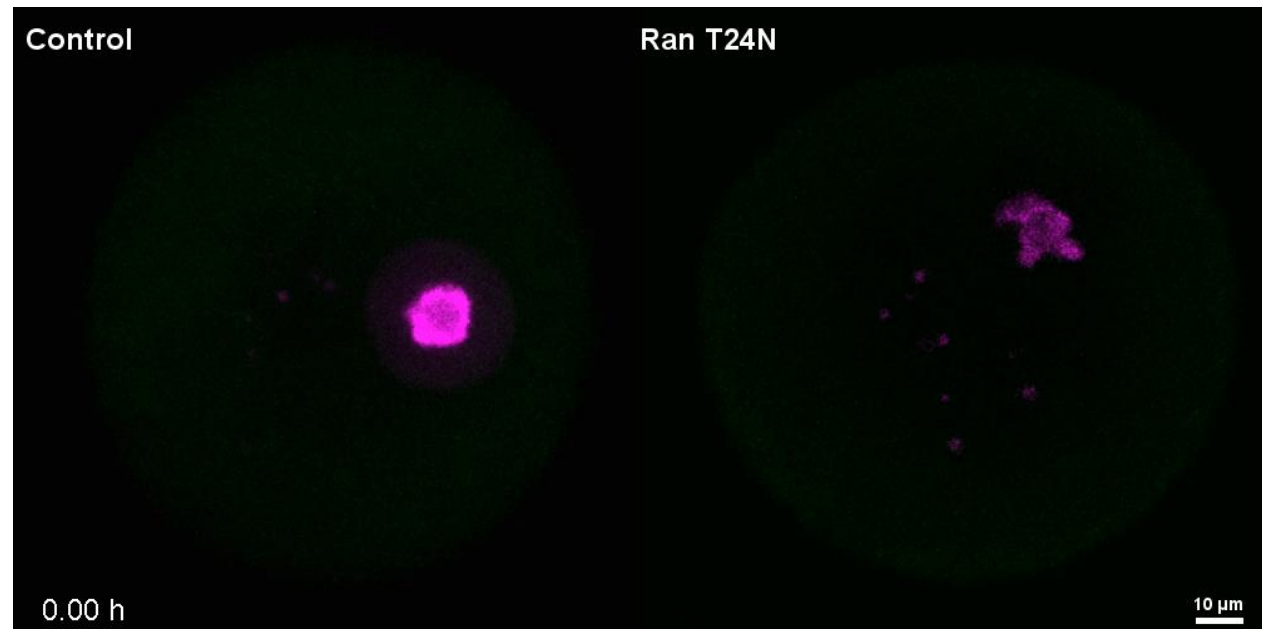
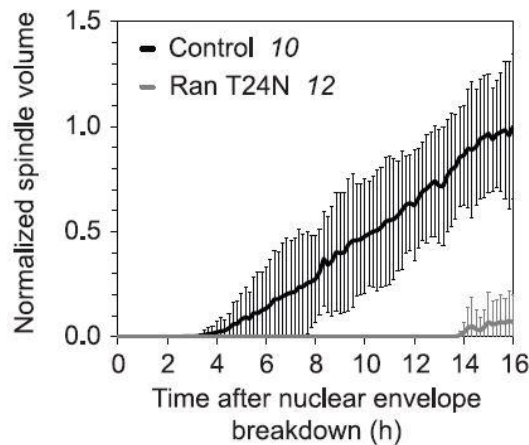
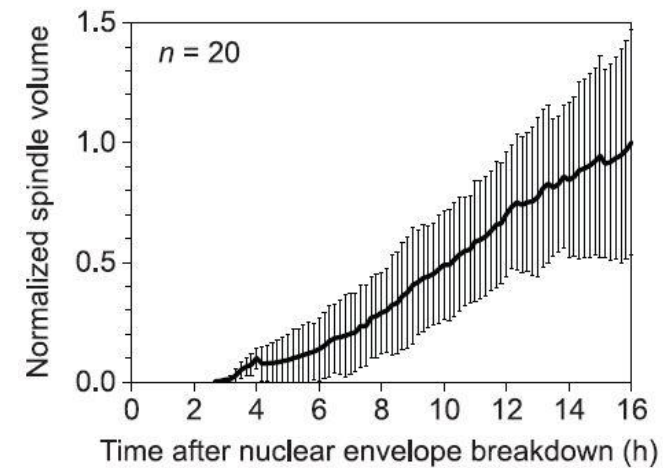
Microtubules (MAP4-EGFP)

Acentrosomal spindle assembly in human oocytes



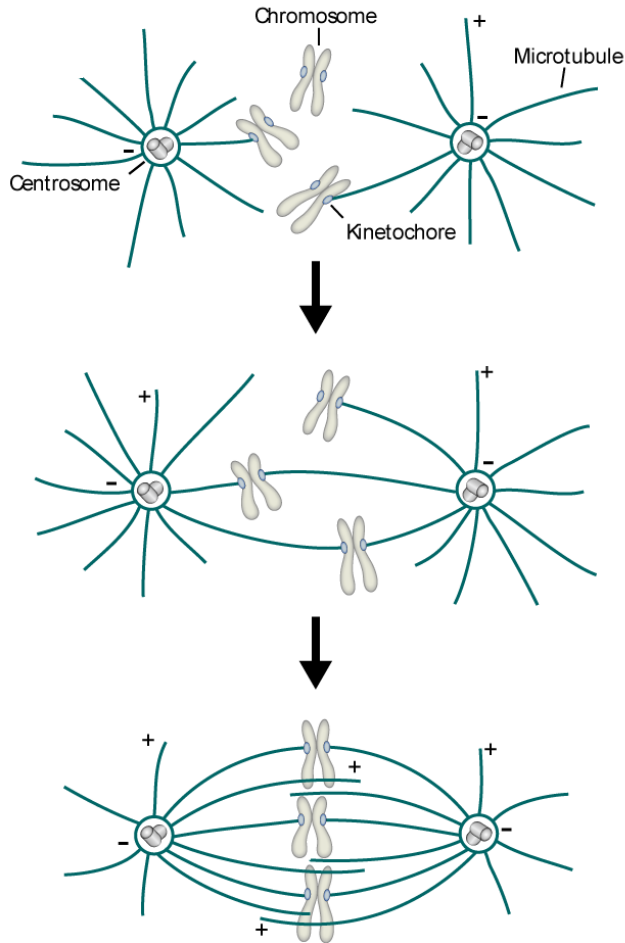
Acentrosomal spindle assembly in human oocytes

- human oocytes assemble a meiotic spindle independently of either centrosomes or other MTOCs
- spindle assembly is mediated by chromosomes and the small guanosine triphosphatase **Ran**
- spindle assembly is unusually long, requiring ~16 hours

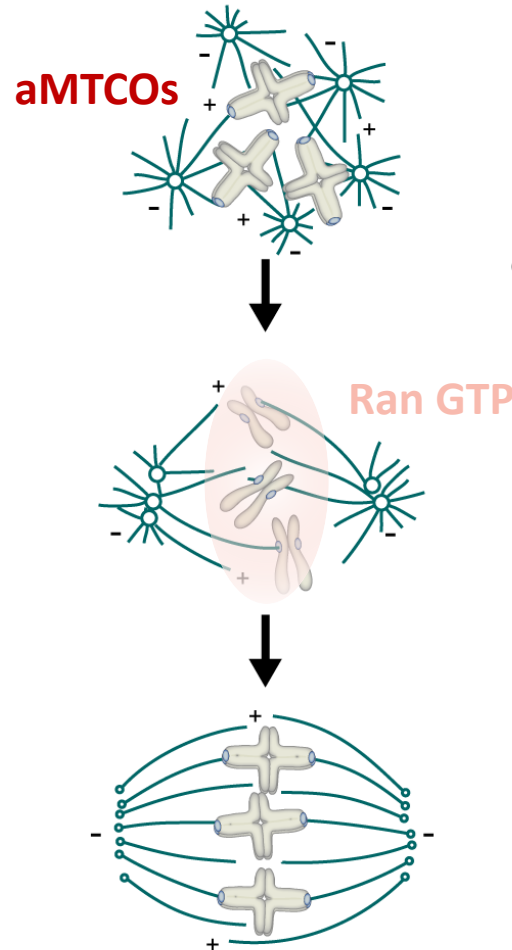


Spindle assembly strategies

Centrosomal Spindle *Mitotic cells*

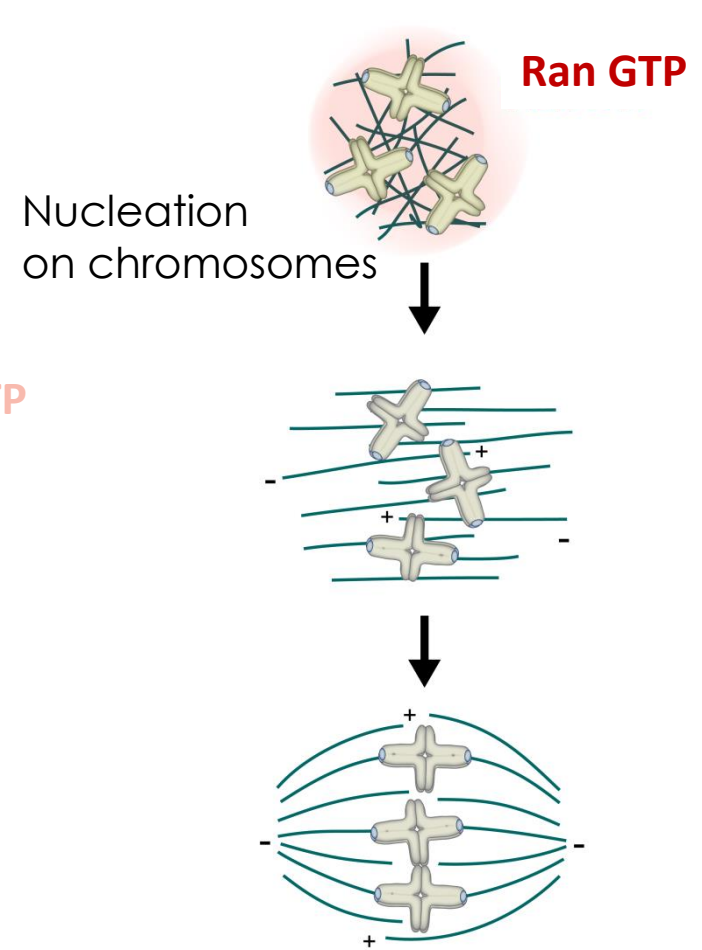


Acentrosomal Spindle *Mouse Oocyte*



Schuh and Ellenberg, Cell 2007.

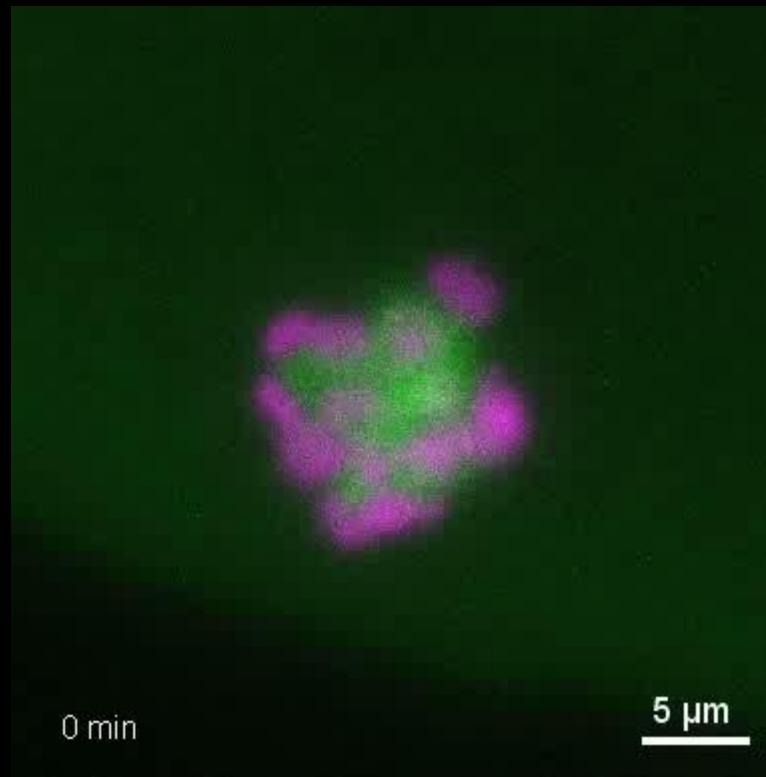
Acentrosomal Spindle *Human oocytes (and plant cells)*



Holubcova et al., Science 2015.

Human oocyte spindle is unstable

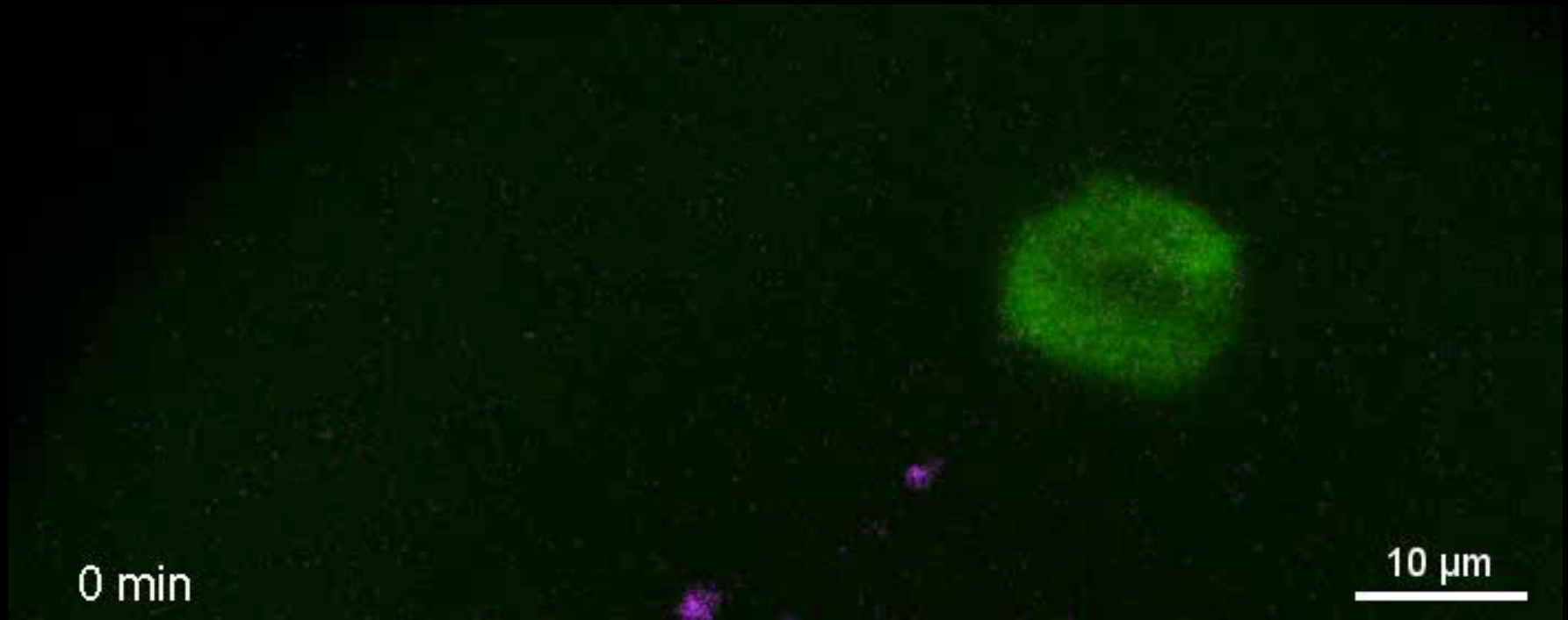
Moderate spindle instability



Chromosomes (H2B-mRFP)
Microtubules (MAP4-EGFP)

Human oocyte spindle is unstable

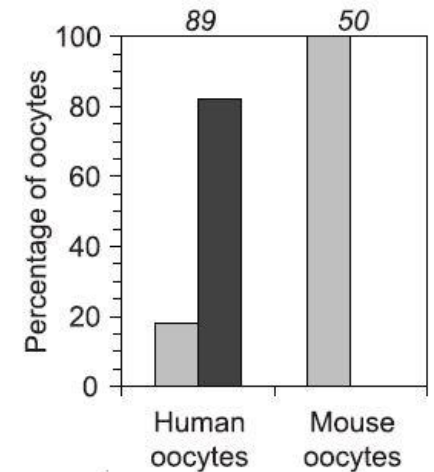
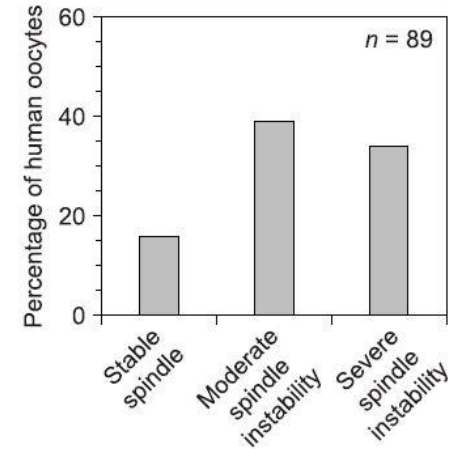
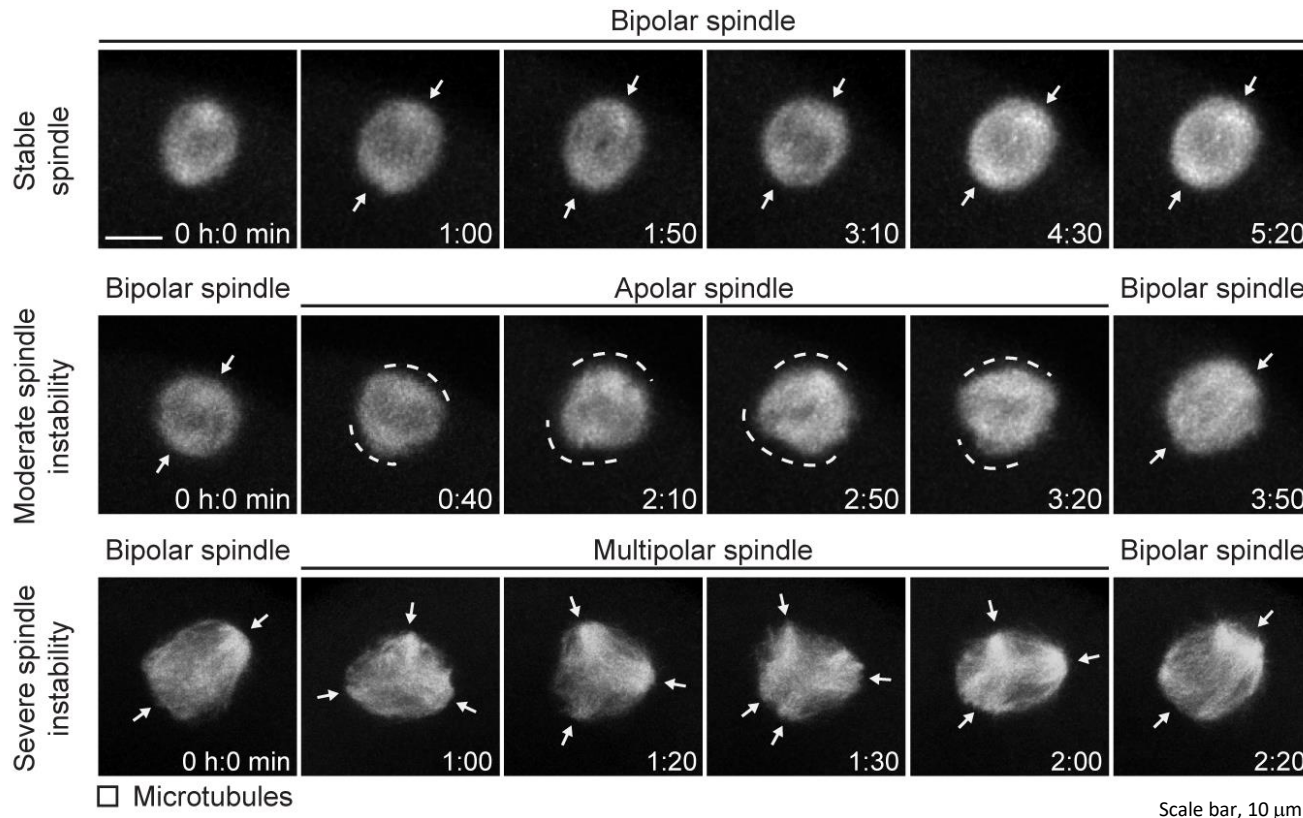
Severe spindle instability



Chromosomes (H2B-mRFP)

Microtubules (MAP4-EGFP)

Human oocyte spindle is unstable

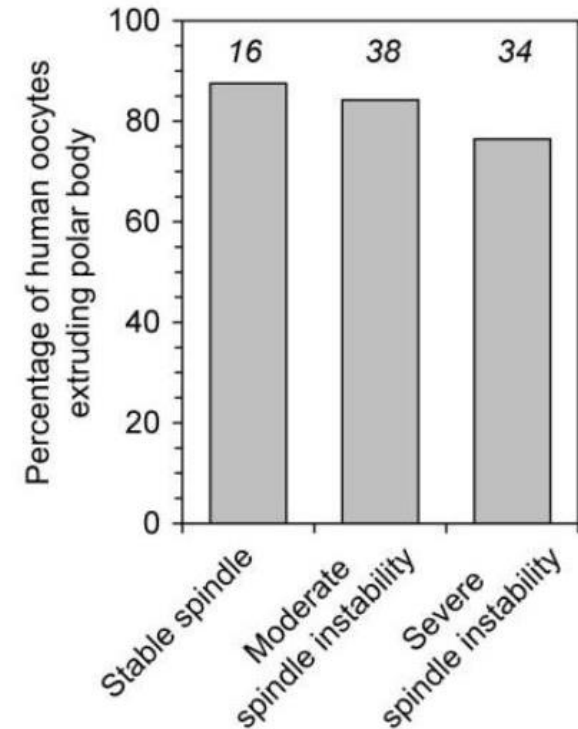
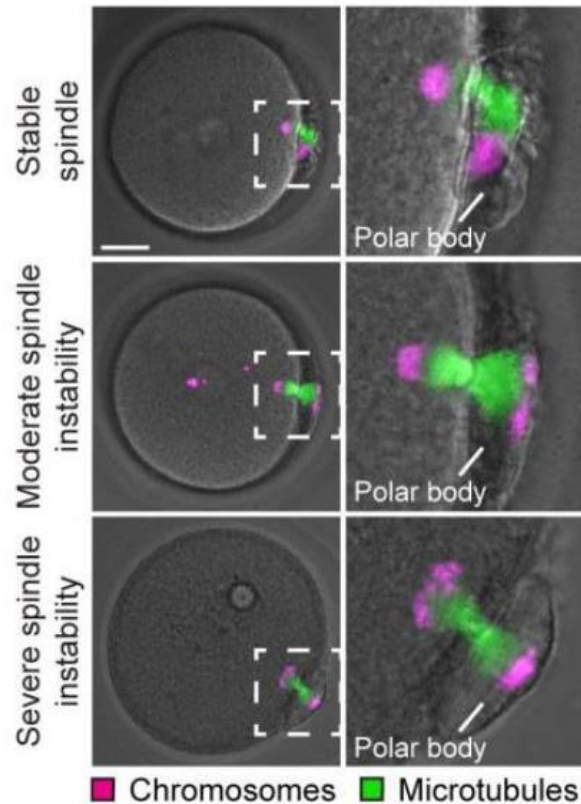
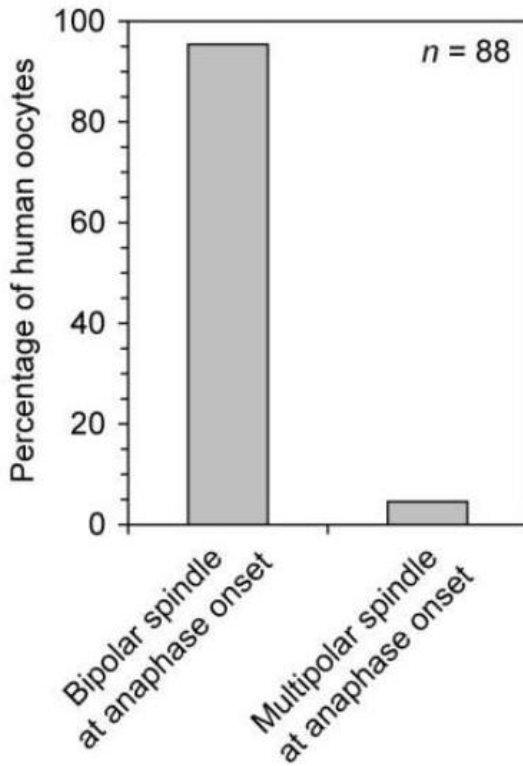


❖ **Prolonged spindle instability was observed in ~80% of human oocytes* but no mouse oocytes**

*Surplus oocytes from stimulated IVF cycles matured *in vitro* !

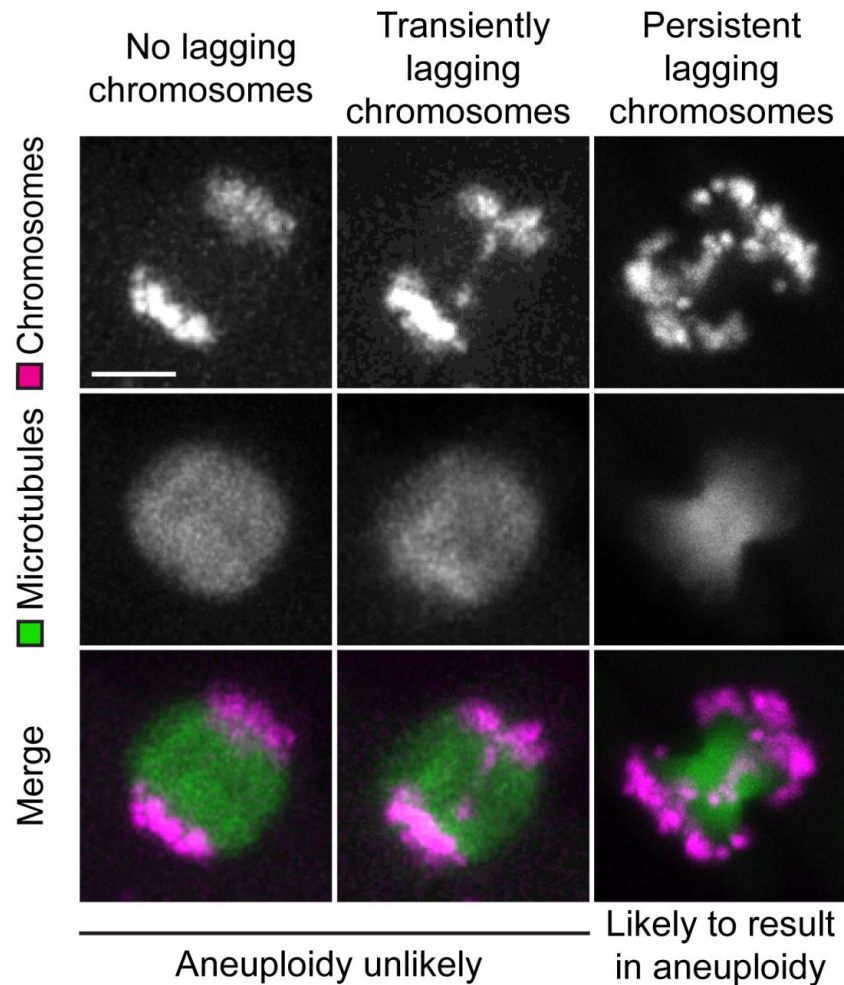
Human oocyte spindle is unstable

- ❖ Majority of human oocytes recovered from spindle instability before anaphase and extruded a polar body

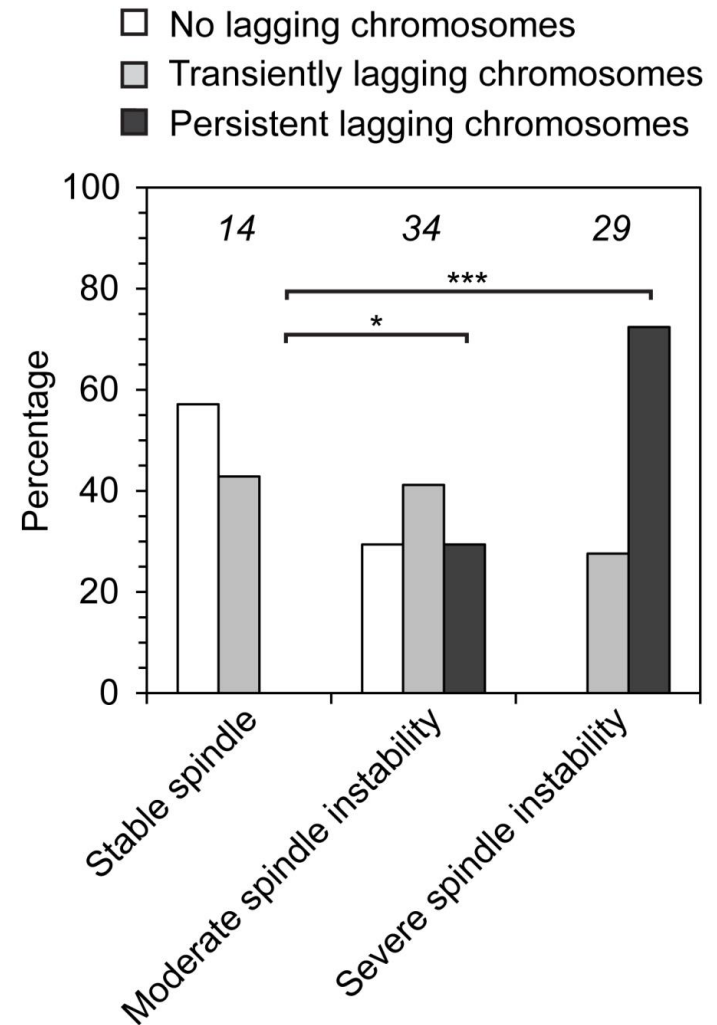


Spindle instability favours chromosome missegregation

❖ Spindle instability correlates with chromosome segregation errors

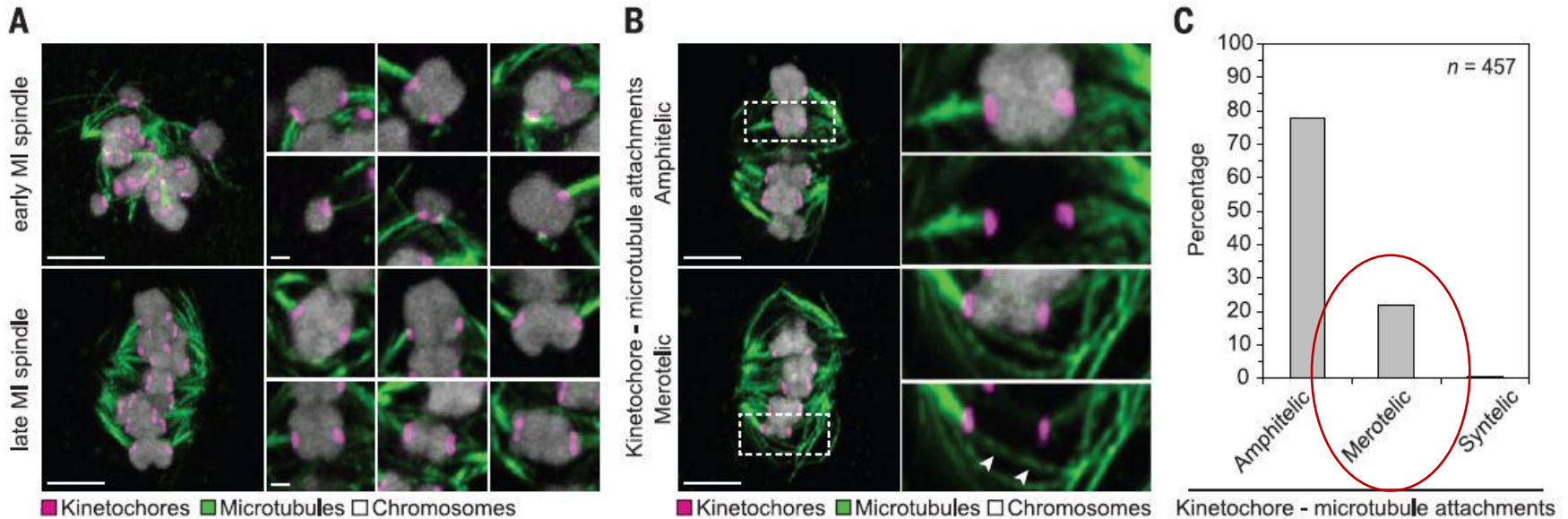


Scale bar, 10 μ m



Spindle instability favours chromosome missegregation

- ❖ Correction of kinetochore-microtubule attachments is incomplete close to anaphase

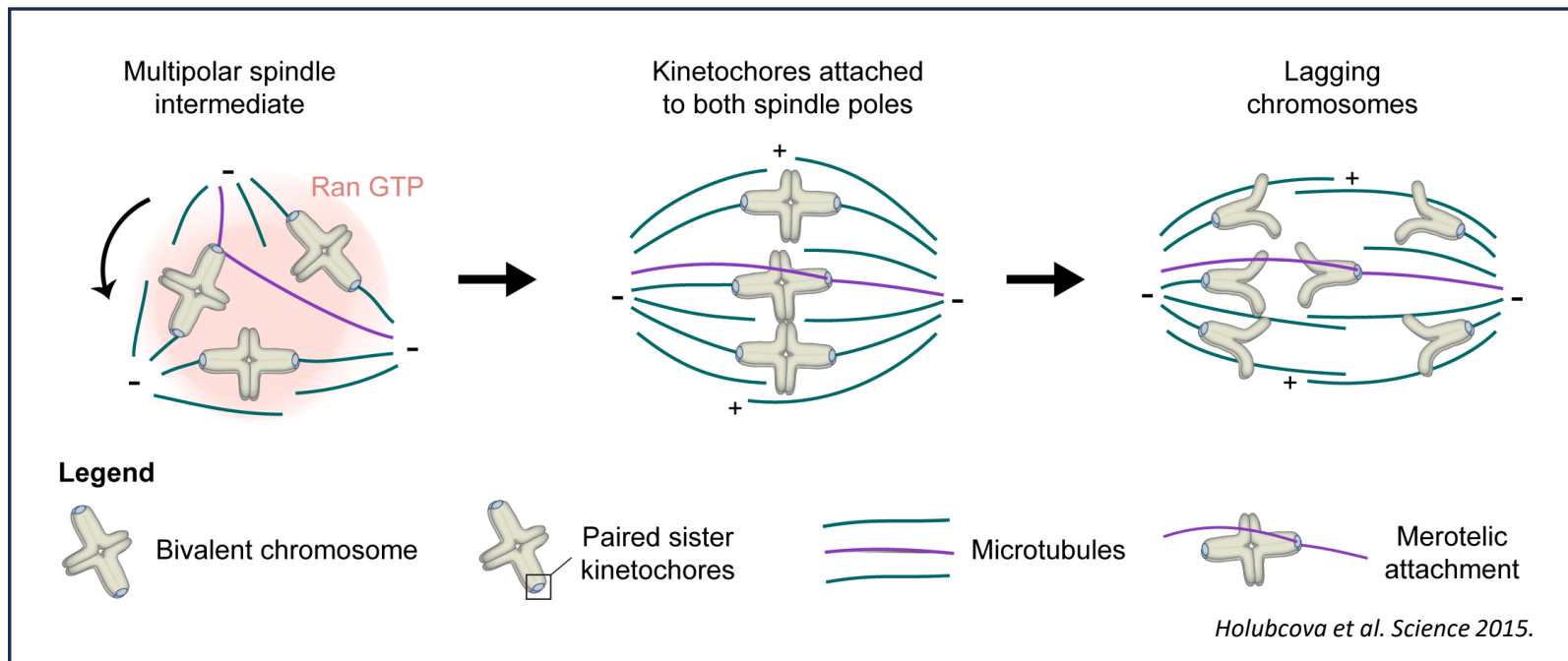


Scale bar, 5 μ m



Spindle instability favours chromosome missegregation

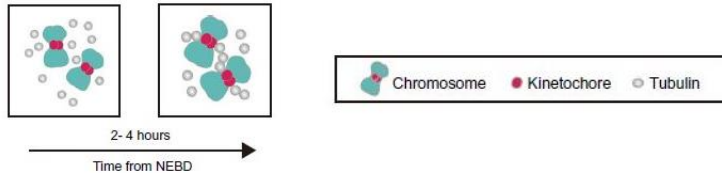
- ❖ at the absence of centrosomes, human oocytes rely on MT nucleation from chromatin
- ❖ chromosome-mediated spindle assembly is slow process and formed spindle is inherently unstable



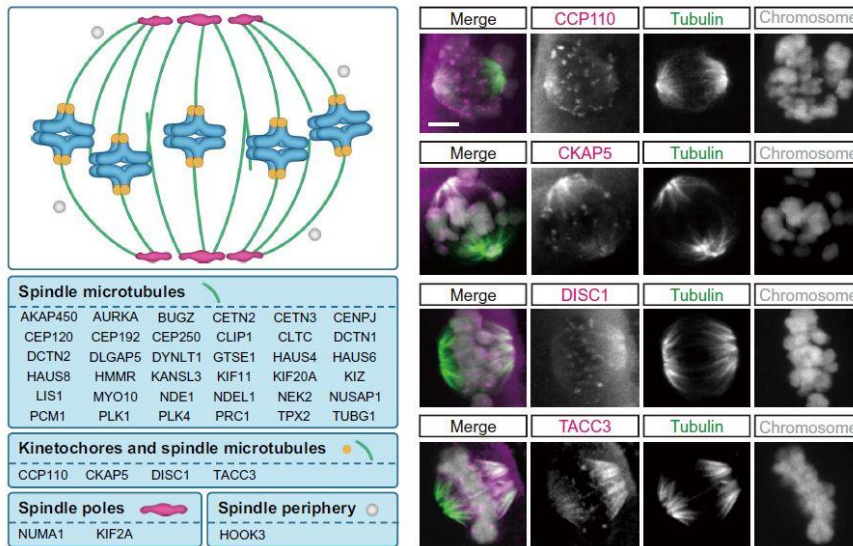
- ❖ improper microtubule-kinetochore attachments established during spindle build-up and remodelling persist to anaphase causing chromosome lagging that is likely to result in aneuploidy

Acentrosomal spindle assembly

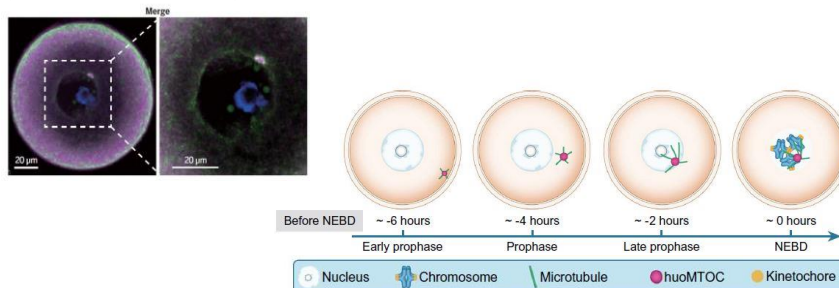
- MT nucleation initiated at kinetochores



- molecular composition of human oocyte spindle



- putative human specific MT nucleators (huMTOC)?



RESEARCH

Wu et al., Science 2022

RESEARCH ARTICLE SUMMARY

HUMAN FERTILITY

The mechanism of acentrosomal spindle assembly in human oocytes

Tianyu Wu†, Jie Dong†, Jing Fu†, Yanping Kuang†, Biaobang Chen, Hao Gu, Yuxi Luo, Ruihan Gu, Meiling Zhang, Wen Li, Xi Dong, Xiaoxi Sun*, Qing Sang*, Lei Wang*



RESEARCH ARTICLE SUMMARY

Wu et al., Science 2024

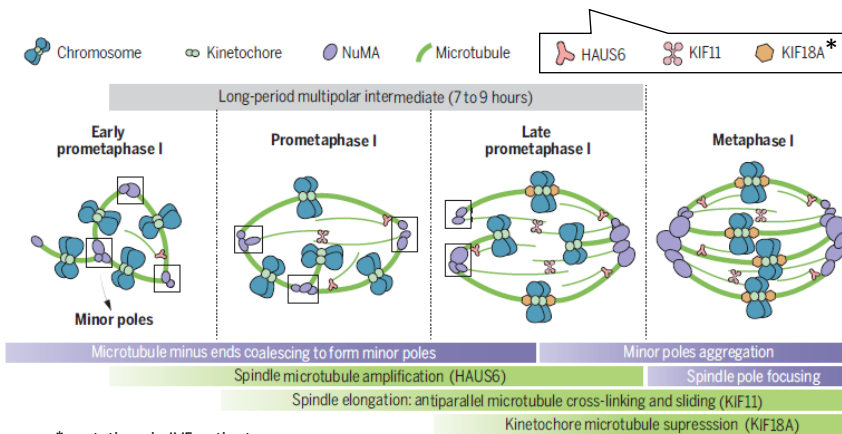
REPRODUCTION

Mechanisms of minor pole-mediated spindle bipolarization in human oocytes

Tianyu Wu†, Yuxi Luo†, Meiling Zhang†, Biaobang Chen†, Xingzhu Du, Hao Gu, Siyuan Xie, Zhiqi Pan, Ran Yu, Ruiqi Hai, Xiangli Niu, Guimin Hao, Liping Jin, Juanzi Shi, Xiaoxi Sun, Yanping Kuang, Wen Li*, Qing Sang*, Lei Wang*



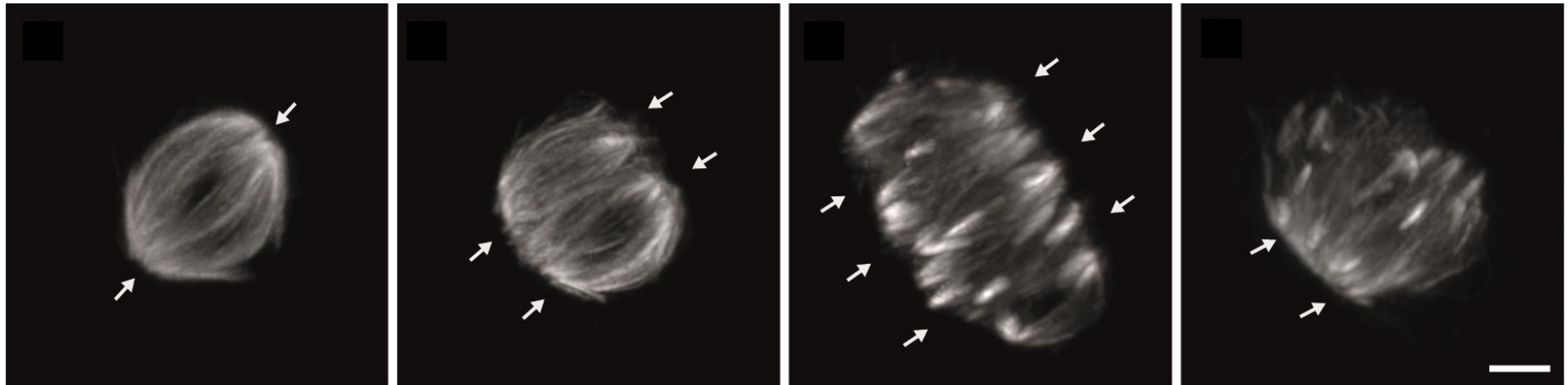
- nascent MT (-)ends coalesce into minor spindle poles which later aggregate to generate opposite spindle poles
- MT amplification, cross-linking and sliding that is required for spindle elongation and bipolarization



* mutations in IVF patients

Human oocyte spindle poles are not stabilized

- ❖ Established spindle poles in human oocytes are prone to loosening and disintegration



Focused spindle poles

Broad spindle poles

Loosen spindle poles

Disintegrated spindle pole

How are spindle poles organized at the absence of centrosomes?



Incidence of unstable acentrosomal spindles



82 %

How are spindles in non-human mammalian oocytes stabilized



0 %



4.4 %



6 %



Mammalian oocyte spindle pole organization



RESEARCH ARTICLE

OOCYTE DIVISION

A liquid-like spindle domain promotes acentrosomal spindle assembly in mammalian oocytes

Chun So^{1*}, K. Bianka Seres^{1,2,3*}, Anna M. Steyer^{4,5}, Eike Mönlich¹, Dean Clift², Anastasija Pejkovska¹, Wiebke Möbius^{4,5}, Melina Schuh^{1,2,†}



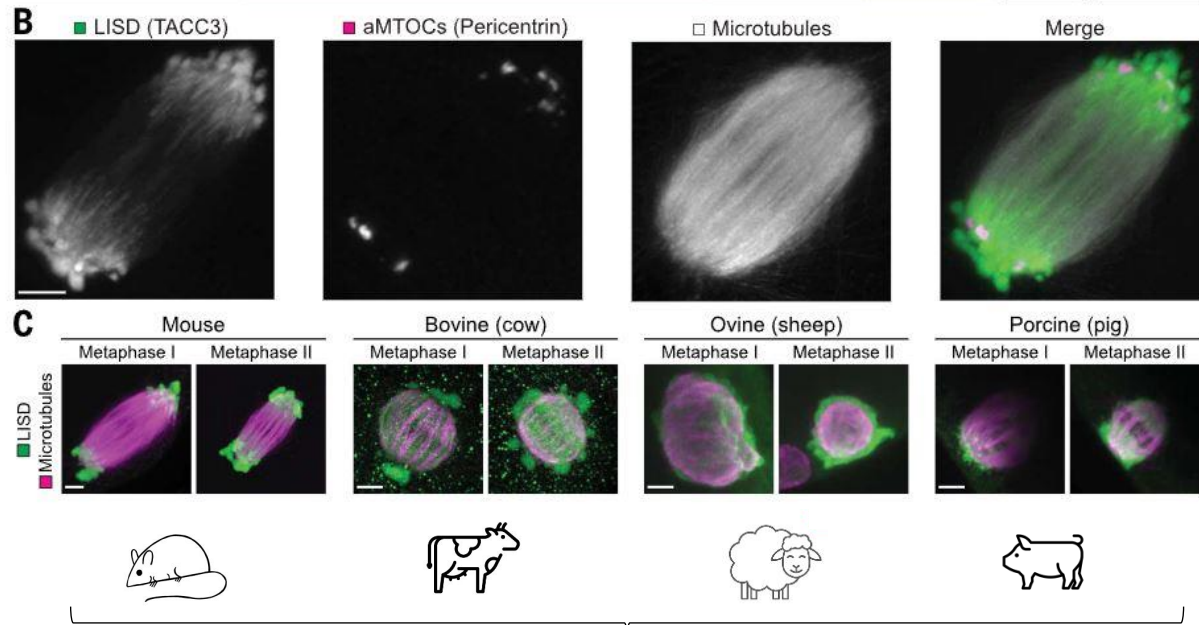
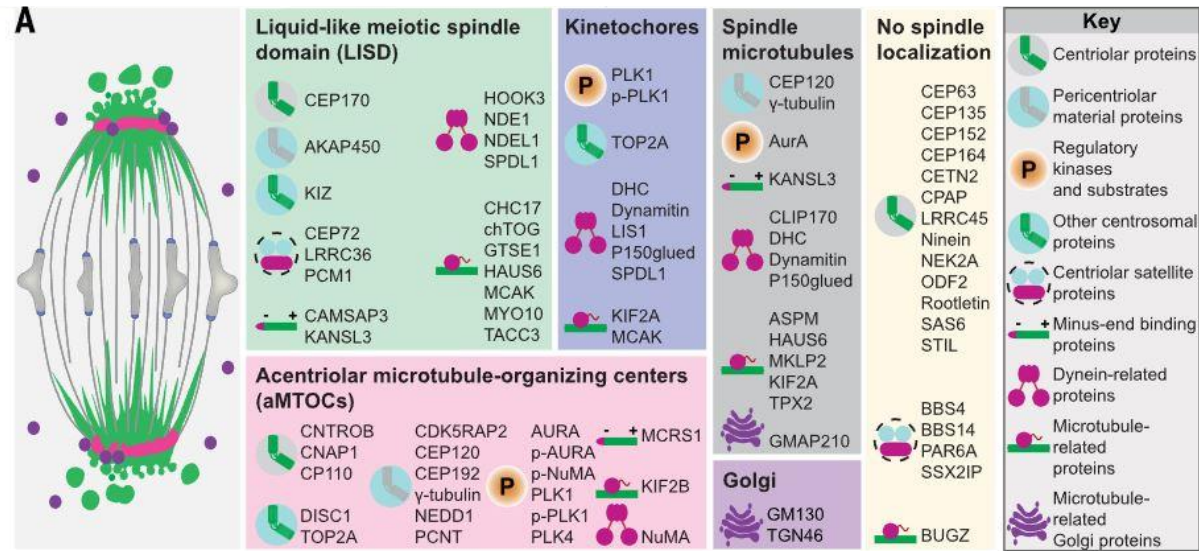
Chun So



Melina Schuh

❖ Liquid-like meiotic spindle domain (LISD)

- localized at poles and permeates the MT mass of mammalian oocytes
- selectively concentrates multiple centrosomal and MT-associated proteins
- allows rapid diffusion within the spindle volume
- disruption of the LISD disperses spindle regulatory factors and leads to severe spindle assembly defects

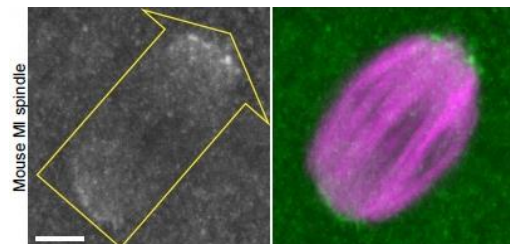
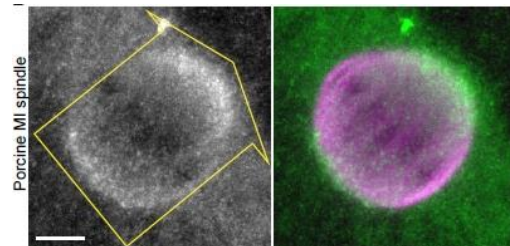
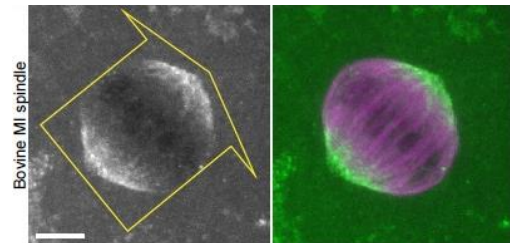
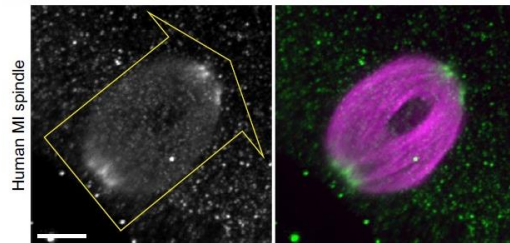


stable spindles

So et al., Science 2019

Human oocyte spindle pole organization

❖ NuMA decorates spindle poles in mammalian oocytes



■ NuMA ■ NuMA ■ Microtubules

RESEARCH ARTICLE

So et al., Science 2022

CELL BIOLOGY

Mechanism of spindle pole organization and instability in human oocytes



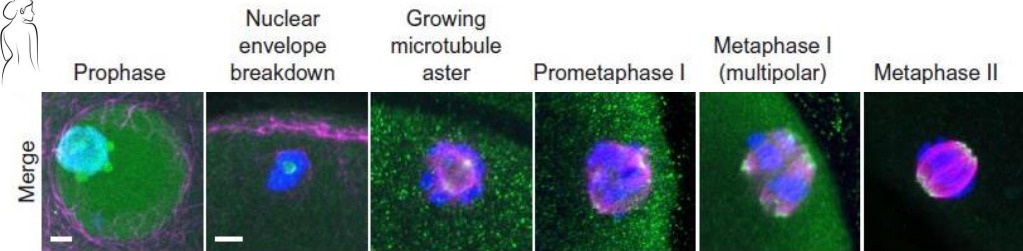
Chun So¹, Katerina Menelaou^{1,2†}, Julia Uraj^{1,2†}, Katarina Harasimov¹, Anna M. Steyer³, K. Bianka Seres^{1,2}, Jonas Bucevičius⁴, Gražvydas Lukinavičius⁴, Wiebke Möbius^{3,5}, Claus Sibold⁶, Andreas Tandler-Schneider⁶, Heike Eckel⁷, Rüdiger Moltrecht⁷, Martyn Blayney², Kay Elder², Melina Schuh^{1,5*}



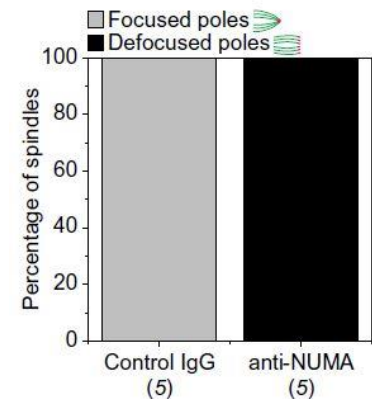
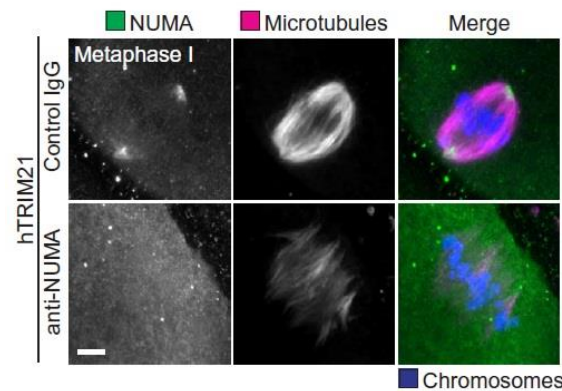
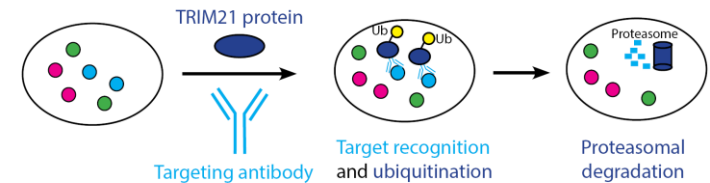
Chun So



Melina Schuh

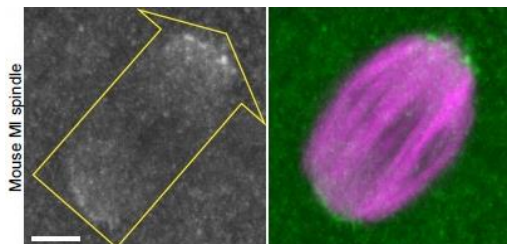
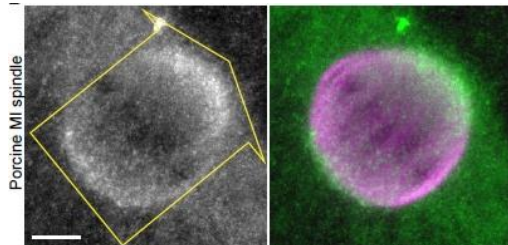
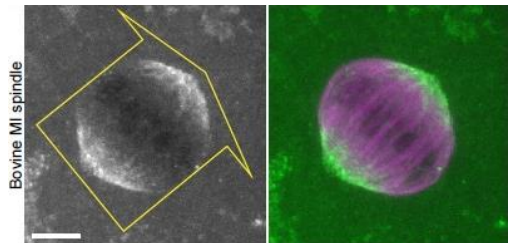
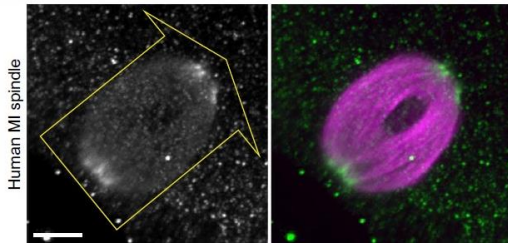


- NuMA is required for MT focusing at spindle poles in human oocytes



Human oocyte spindle pole organization

❖ NuMA decorates spindle poles in mammalian oocytes



■ NuMA

■ NuMA ■ Microtubules

RESEARCH ARTICLE

So et al., Science 2022

CELL BIOLOGY

Mechanism of spindle pole organization and instability in human oocytes



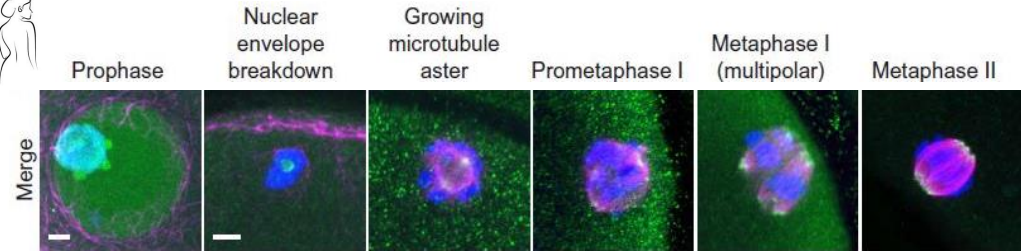
Chun So¹, Katerina Menelaou^{1,2†}, Julia Uraji^{1,2†}, Katarina Harasimov¹, Anna M. Steyer³, K. Bianka Seres^{1,2}, Jonas Bucevičius⁴, Gražvydas Lukinavičius⁴, Wiebke Möbius^{3,5}, Claus Sibold⁶, Andreas Tandler-Schneider⁶, Heike Eckel⁷, Rüdiger Moltrecht⁷, Martyn Blayney², Kay Elder², Melina Schuh^{1,5*}



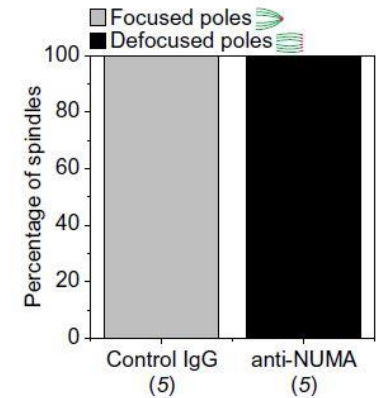
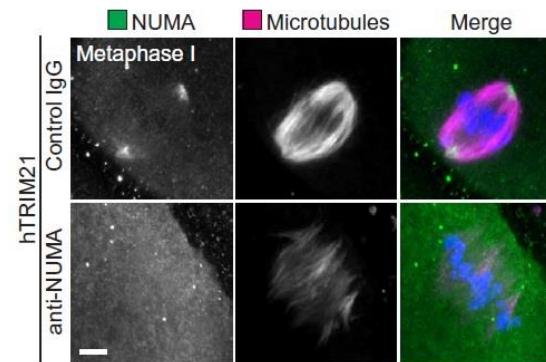
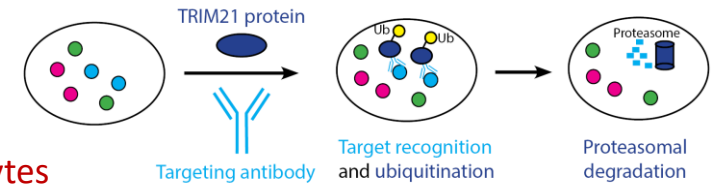
Chun So



Melina Schuh

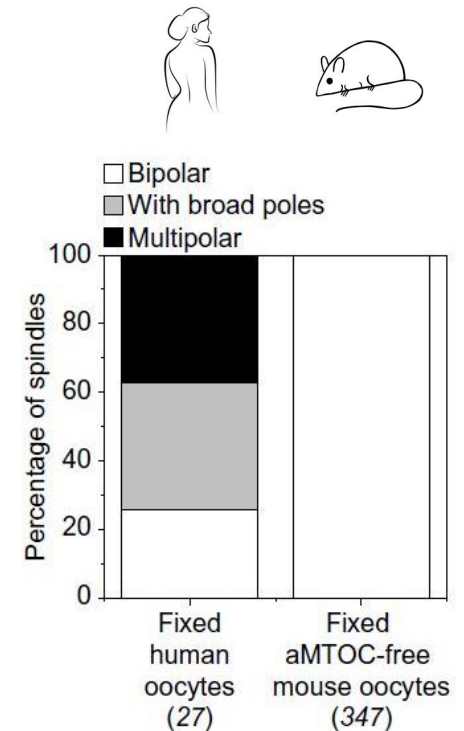
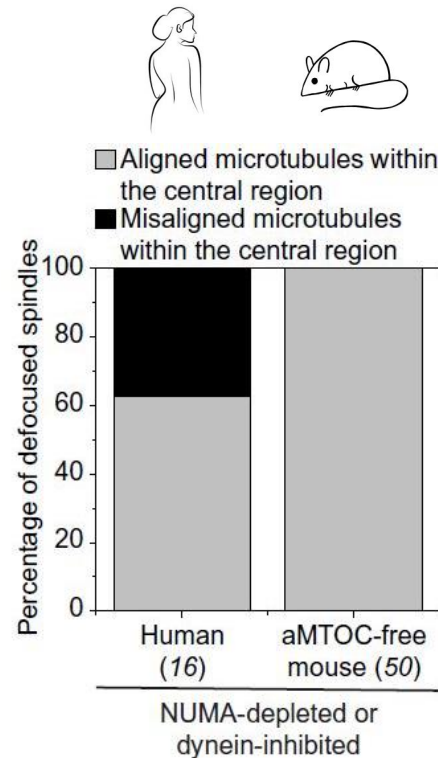
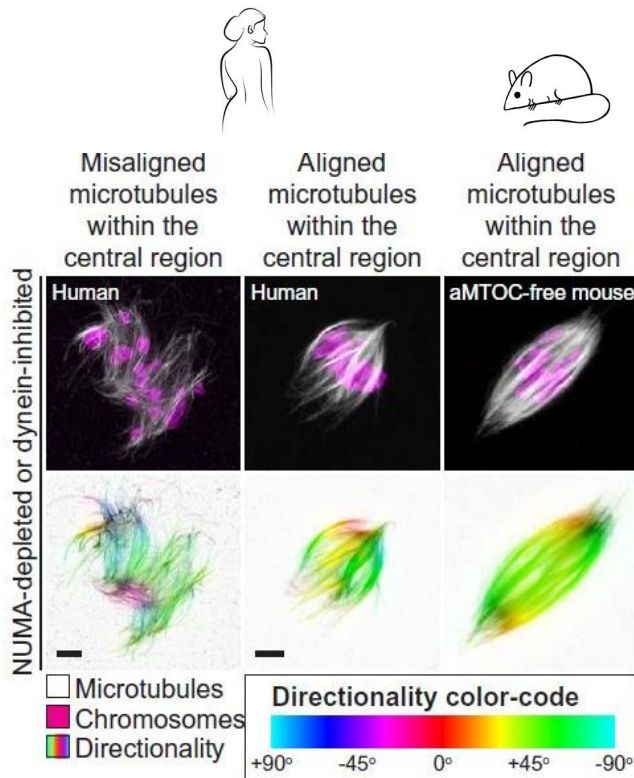


- acute depletion of NuMA → NuMA is required for MT focusing at spindle poles in human oocytes



Search for oocyte spindle stabilizing factor

Misalignment of microtubules in central region of human oocyte spindle



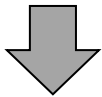
Human oocytes must lack a stabilizing protein that protects mouse, porcine and bovine oocytes from spindle instability

Search for oocyte spindle stabilizing factor

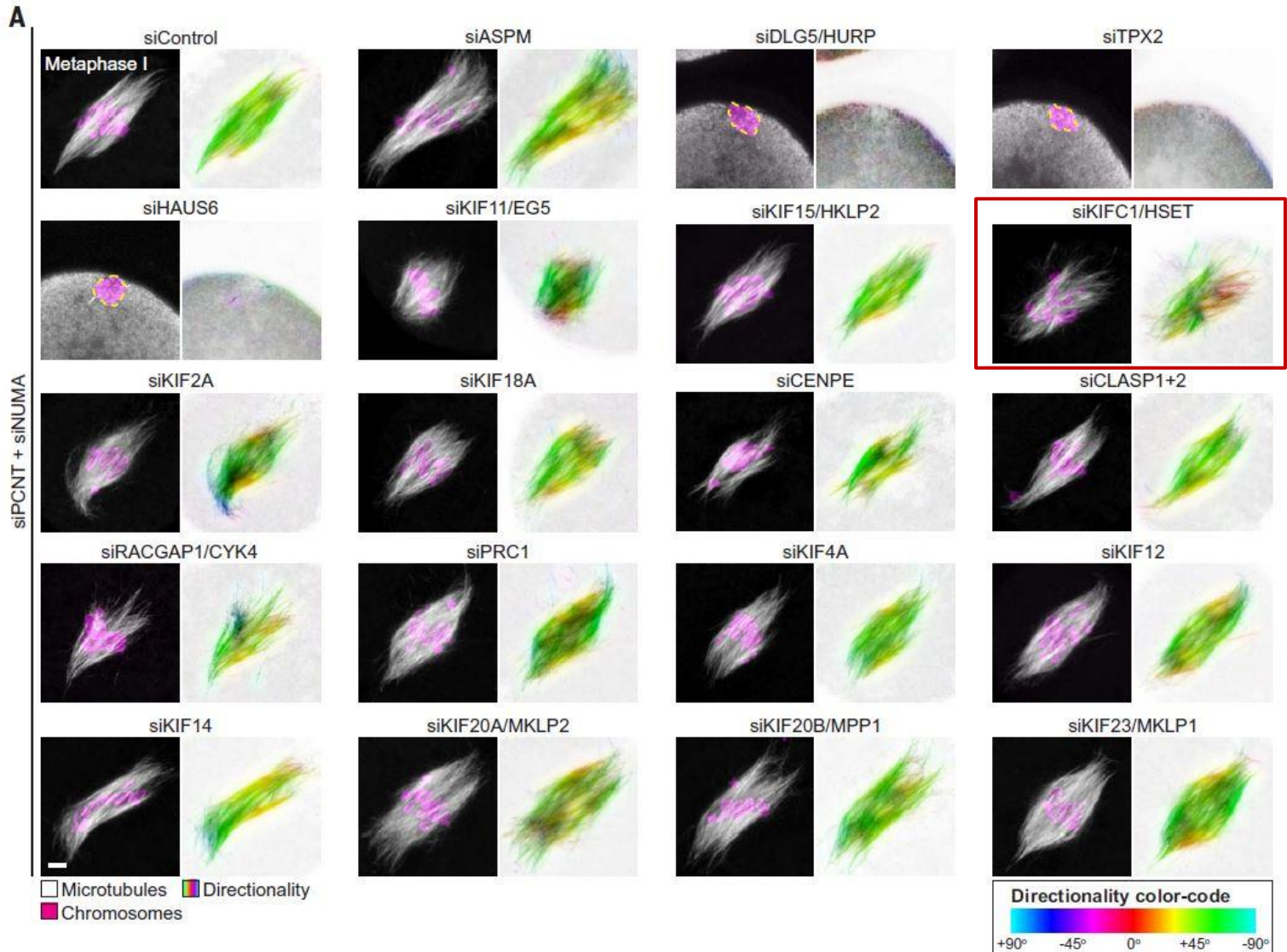
si RNA screen



co-depletion of
NuMA and
candidate
spindle-
associated
proteins

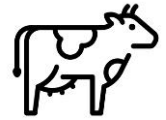


phenotypes
analyzed



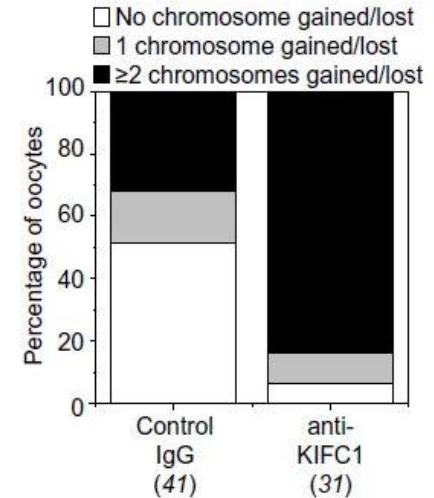
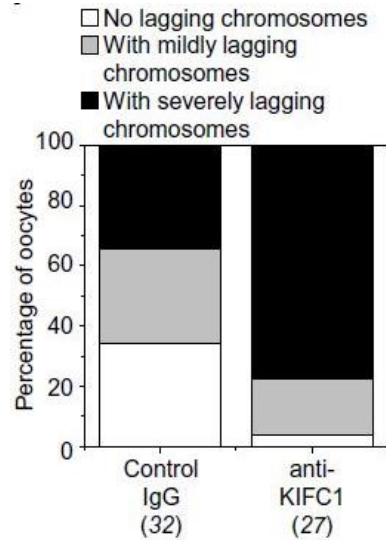
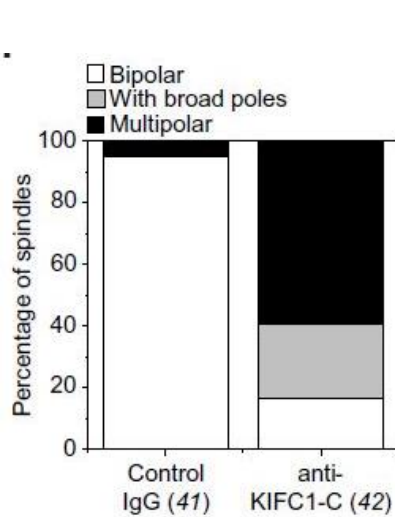
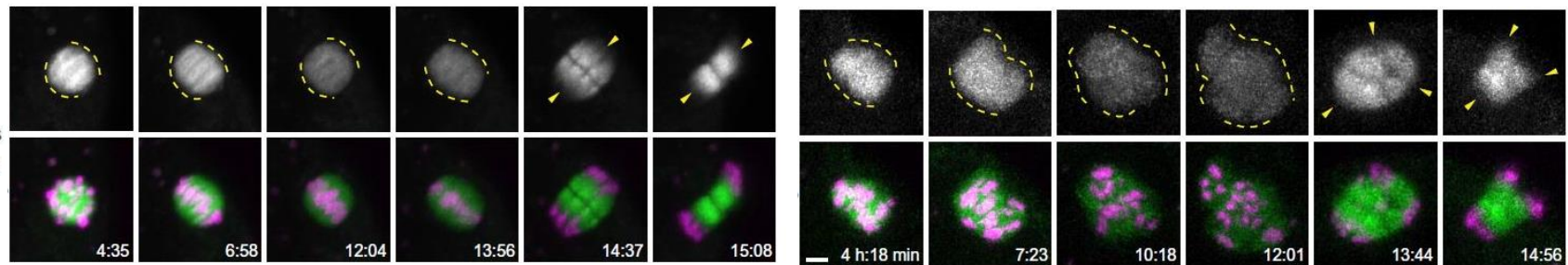
Search for oocyte spindle stabilizing factor

Trim-Away



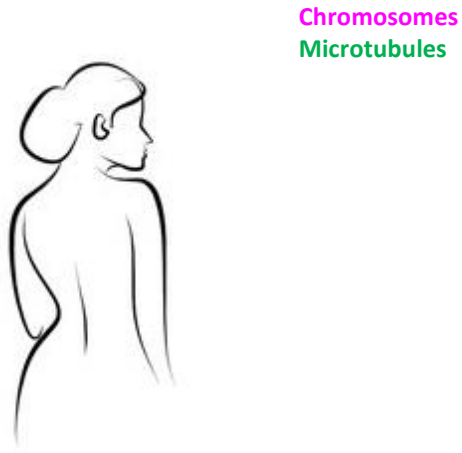
Control

Anti-KIF-C

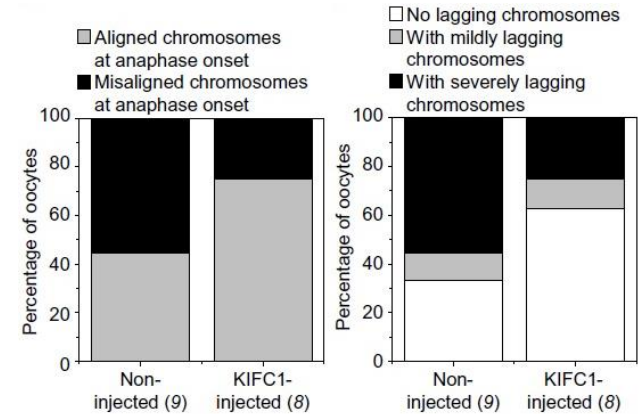
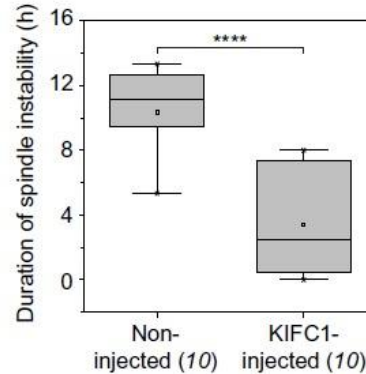
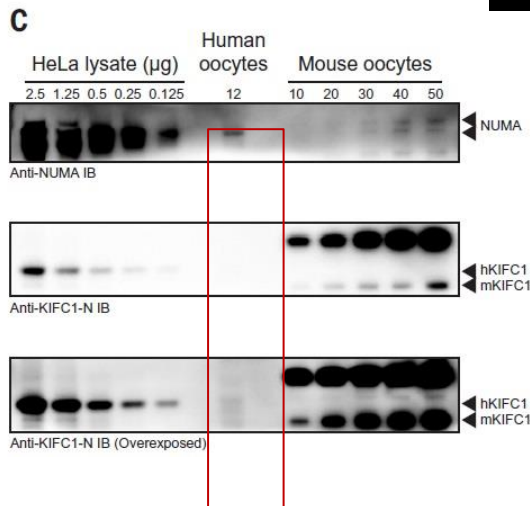
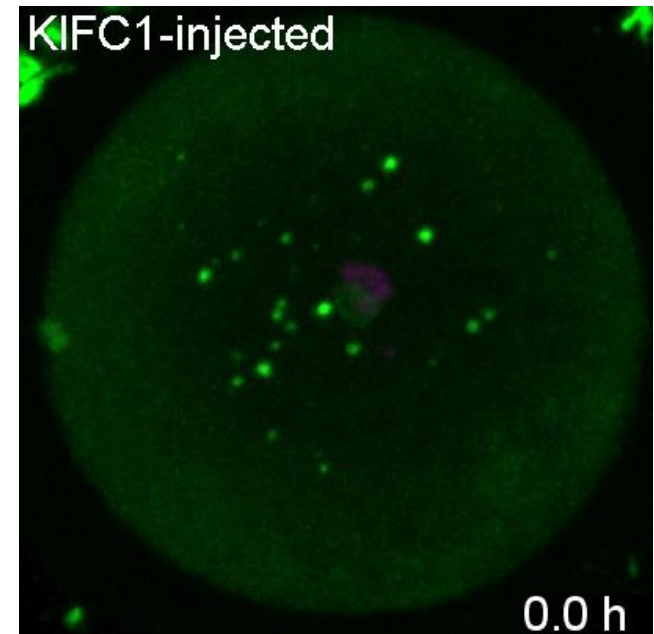
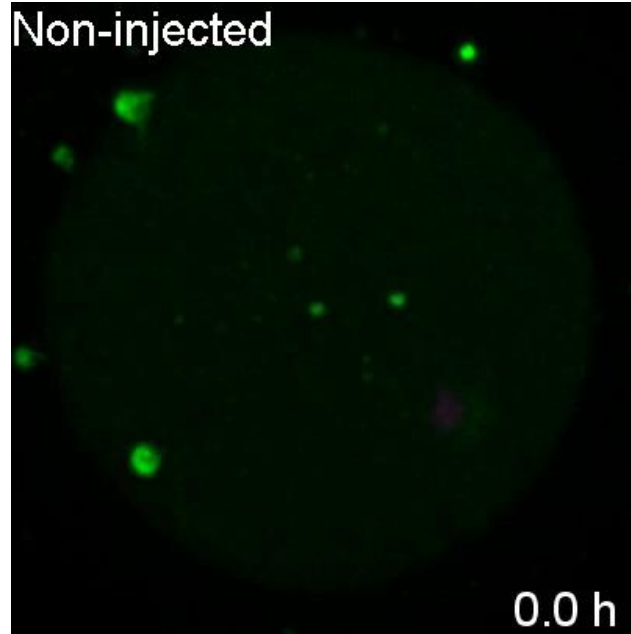


Depletion of KIFC1/HSET induces spindle instability and promotes aneuploidy in bovine oocytes

Search for oocyte spindle stabilizing factor



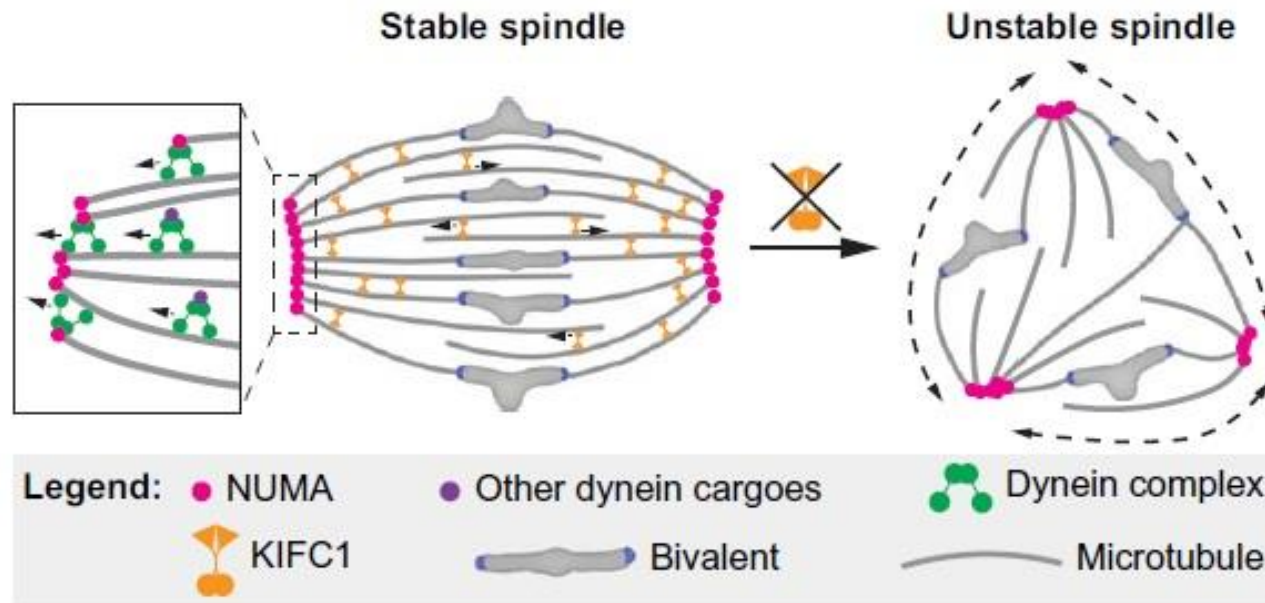
❖ Human oocytes are deficient in KIFC1



❖ Exogenous KIFC1 rescues spindle instability in human oocytes

Prevention of oocyte spindle instability

- ❖ **KIFC1** ensures the spindle stability and prevents fragmentation of spindle poles by **ensuring alignment of MT at central region and crosslinking MT minus ends at spindle poles**

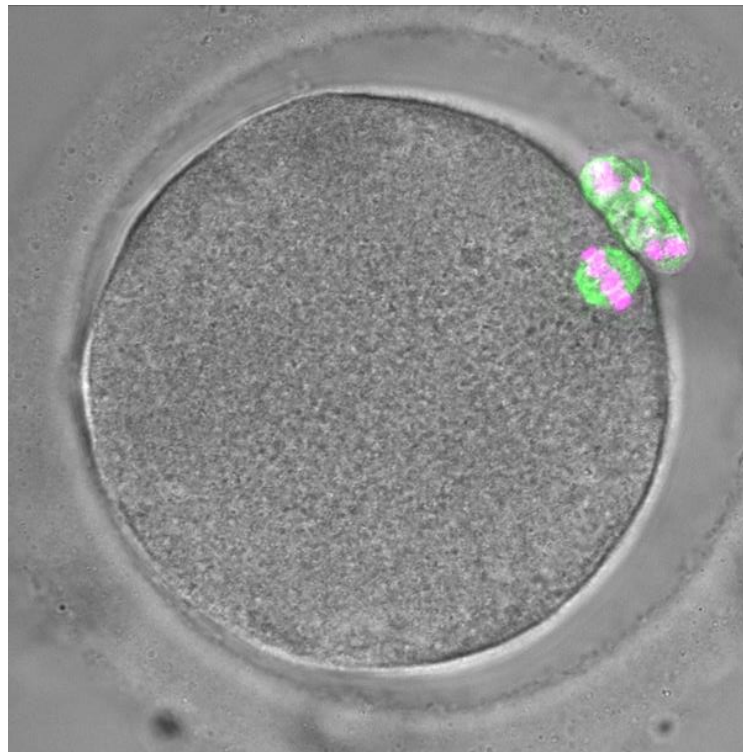


- ❖ **NuMa** organize acentrosomal spindle poles by ensuring coalescence of crosslinked MT-minus ends

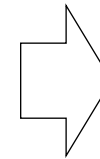
Egg maturity

❖ Mature egg

= metaphase II arrested oocytes with PB extruded and chromosomes aligned in MII spindle



DNA, microtubules

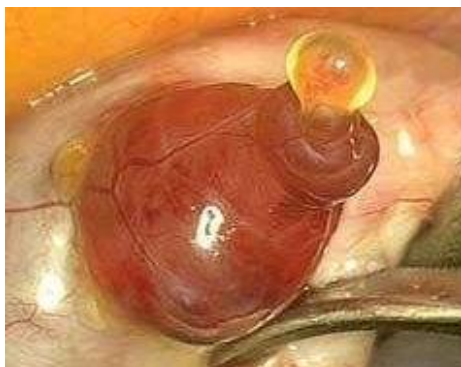


In IVF practice,
all PB-displaying oocytes
are regarded as MIIs
and subjected to ICSI

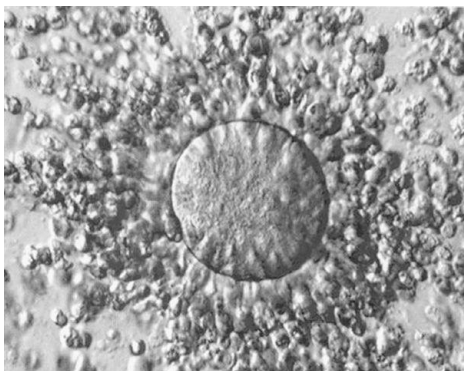
Oocyte maturity

IN VIVO

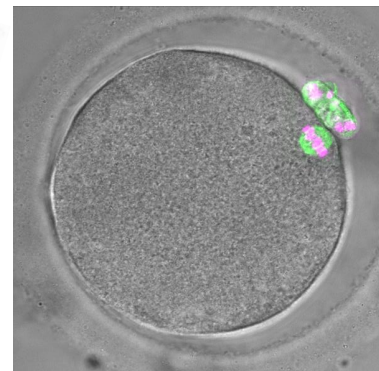
NATURAL CYCLE



OVULATION



SINGLE COC

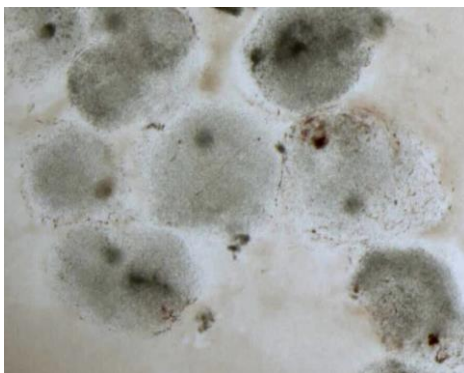


MATURE EGG

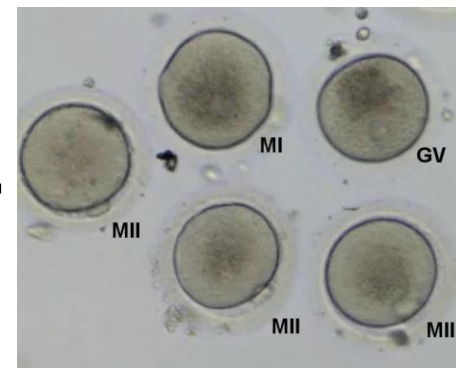
CONTROLLED OVARIAN STIMULATION



PREOVULATORY FOLLICLES



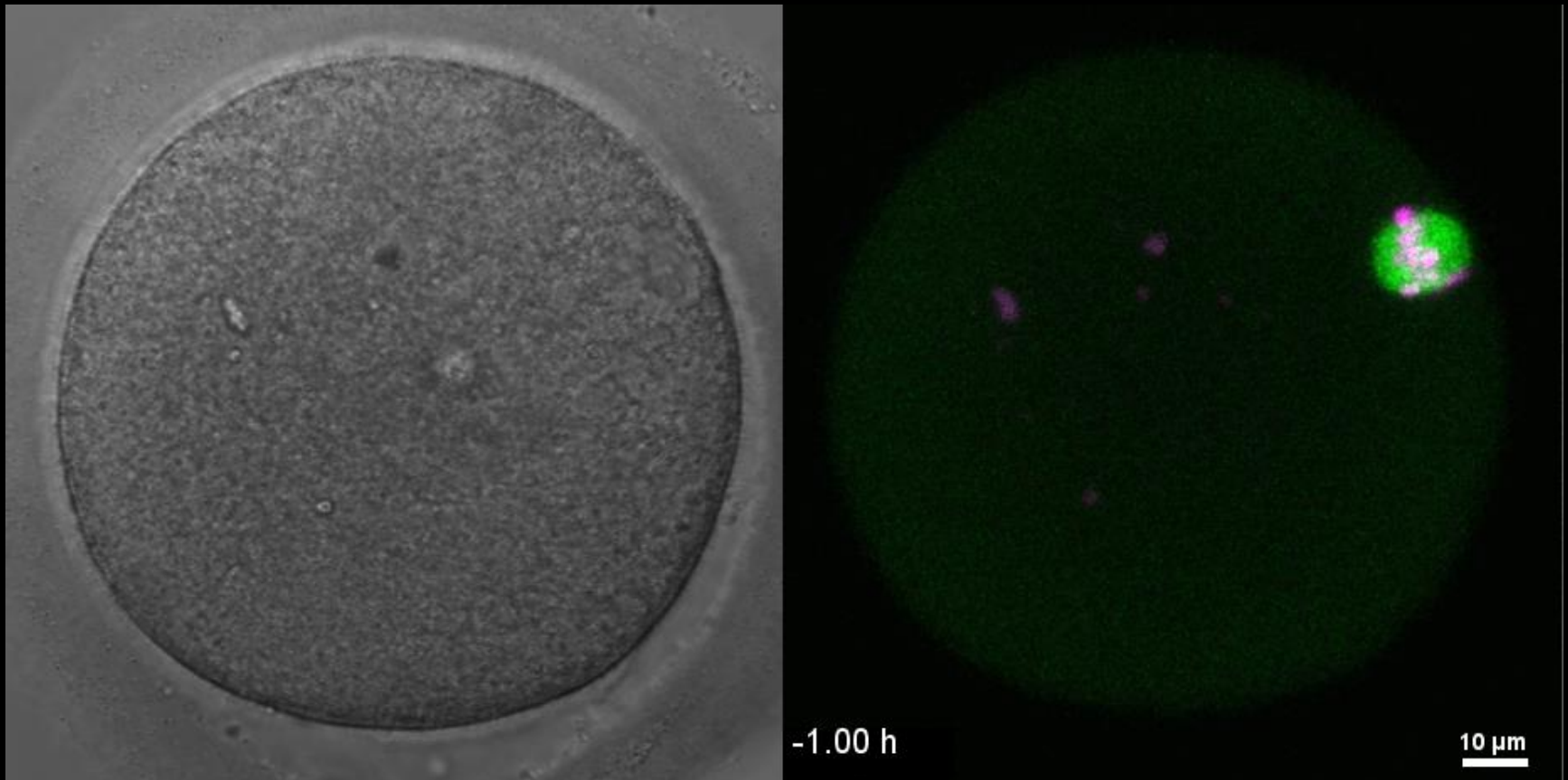
MULTIPLE COCS



MATURE + IMMATURE OOCYTES

MI to MII transition and MII spindle assembly

- MII spindle formation is rapid compared to MI

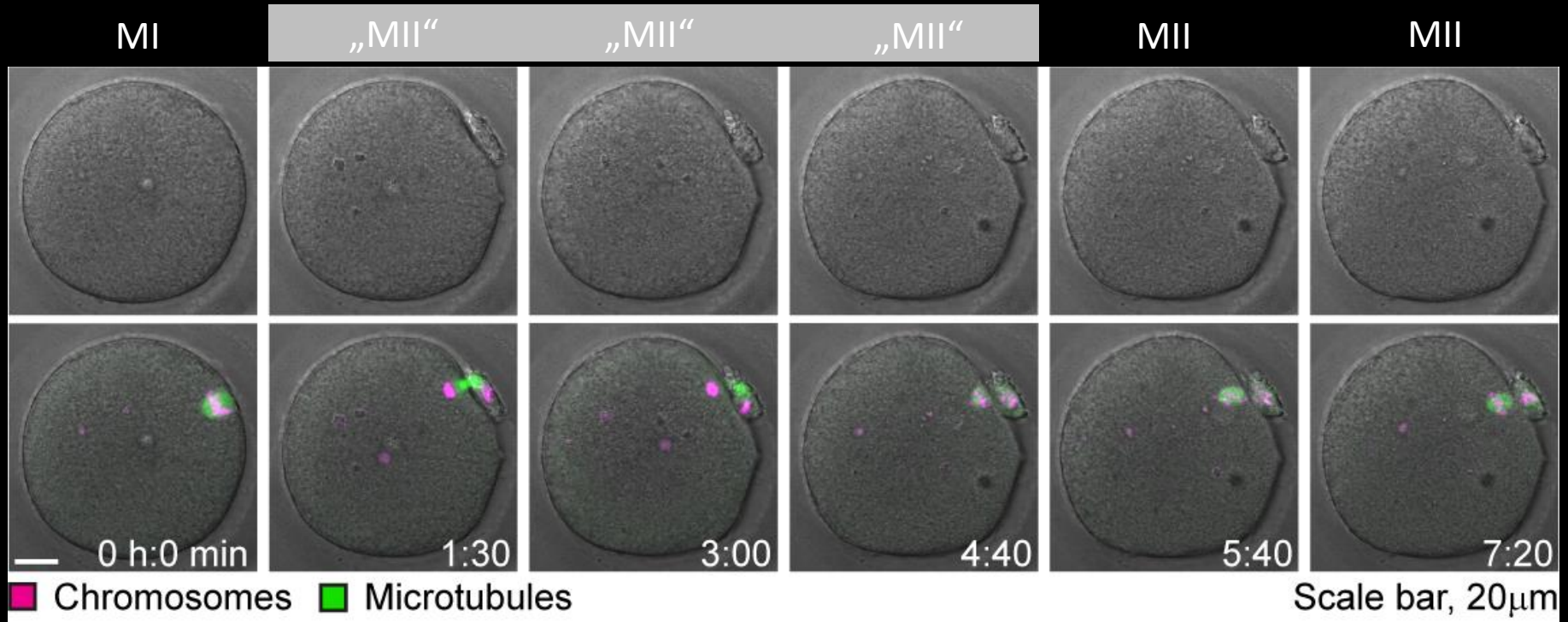


Chromosomes
(H2B-mRFP)
Microtubules
(MAP4-EGFP)

- asynchrony between PB extrusion and MII arrest !

MI to MII transition and MII spindle assembly

- Emergence of PB precedes MII arrest



→ risk of untimely fertilization (ICSI)



Non-invasive spindle visualization

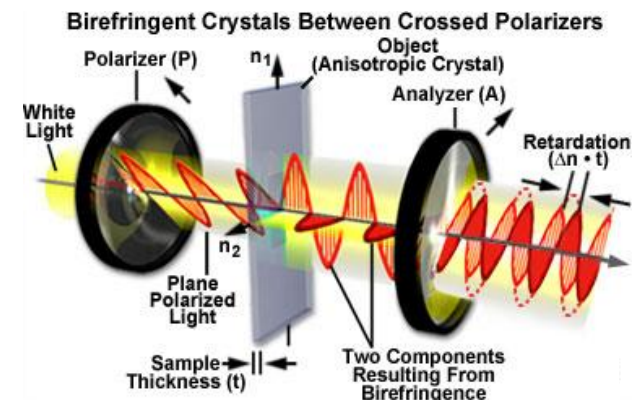
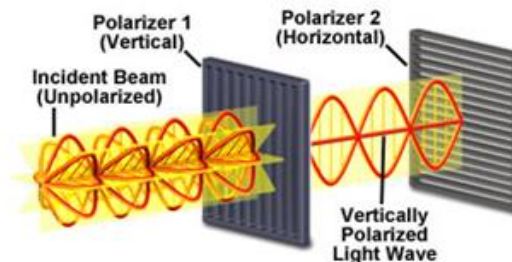
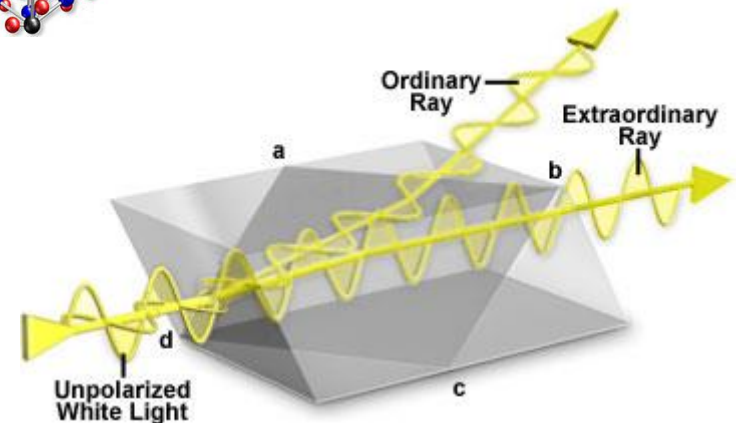
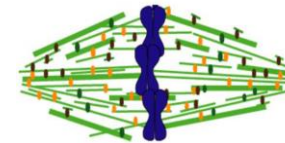
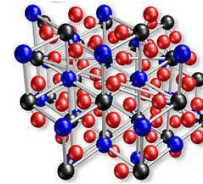
❖ Polarized Light Microscopy (PLM)

- based on interference of **polarized light** with **anisotropic** substances e.g. axial crystals, liquid crystals and **oriented (bio)polymers**

BIREFRINGENCE

- property of certain materials to split a light beam to two rays (ordinary/extraordinary)

- polarized light is refracted by these anisotropic materials and divided to separate components vibrating perpendicularly
- both polarized light ray then pass through the analyzer and the relative retardance of one ray to the other is calculated



Non-invasive spindle visualization

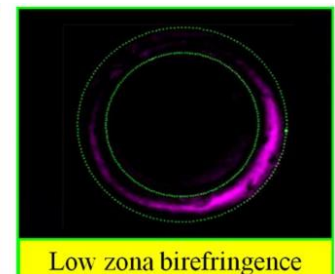
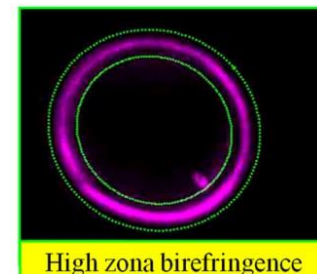
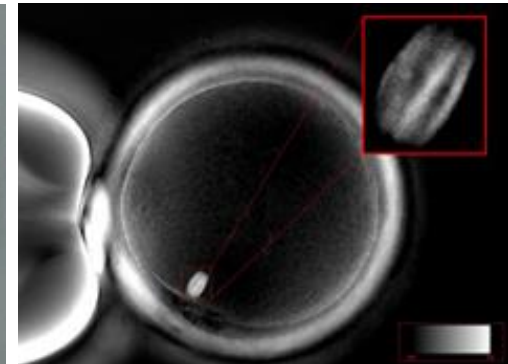
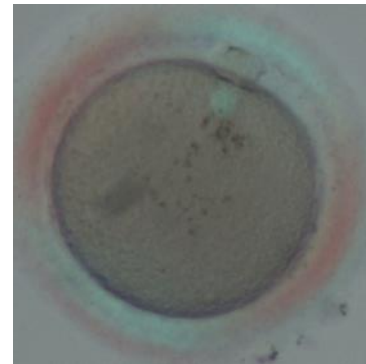
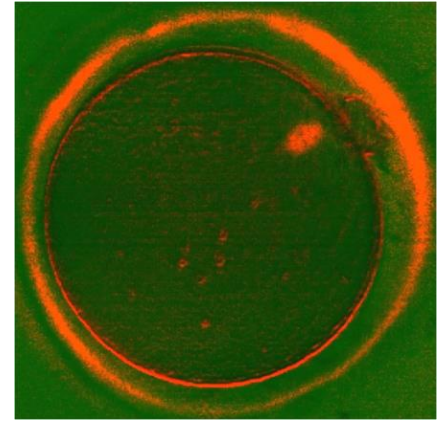
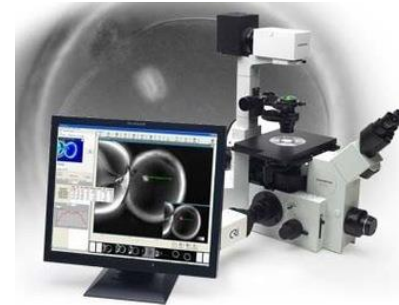
❖ Polarized Light Microscopy (PLM)

- enables **non-invasive** imaging of **birefringent** structures in living cells



- presence and positioning of MII spindle
- pattern of zona pellucida

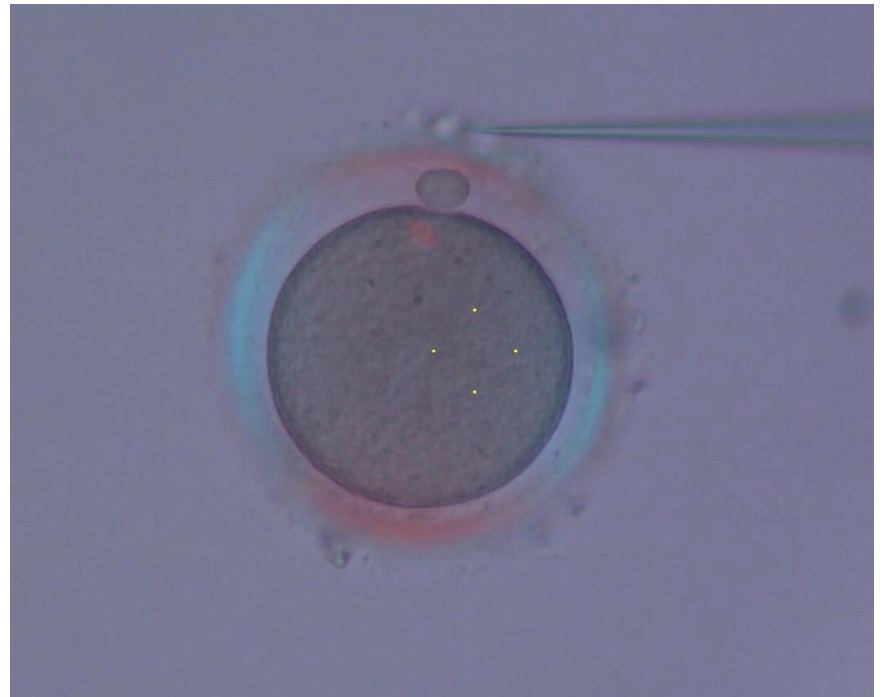
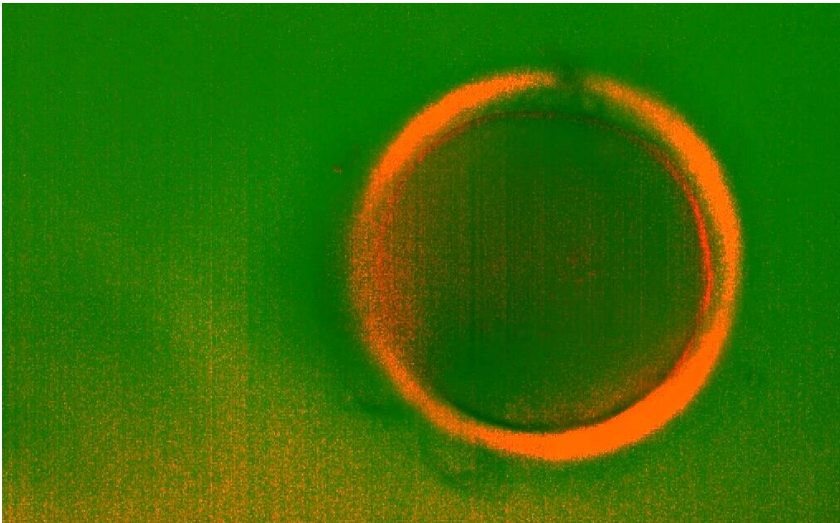
- presence of PLM-detectable MII spindle is a **positive marker of egg's fertilization and developmental competence**



Non-invasive spindle visualization

❖ Polarized Light Microscopy (PLM)

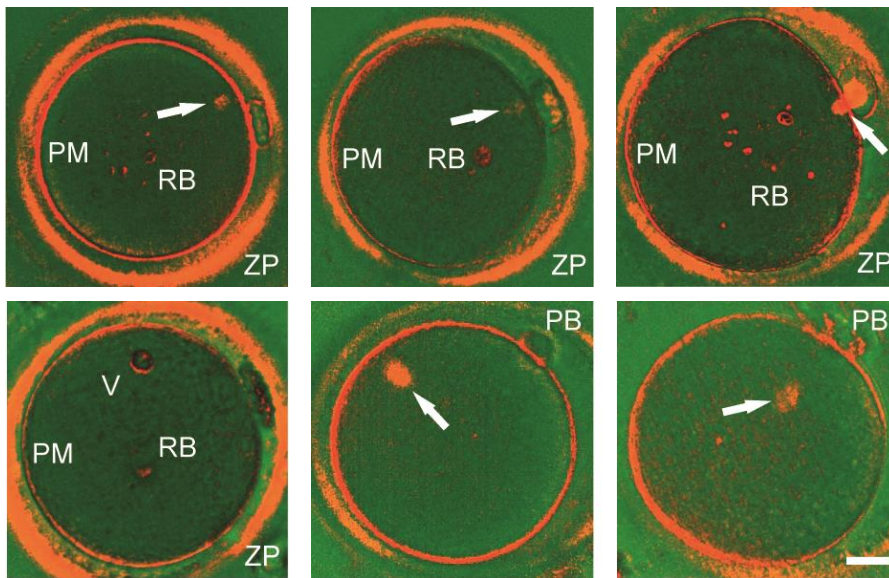
- PLM signal is orientation-dependent
- spindle imaging requires oocyte orientation



Non-invasive spindle visualization

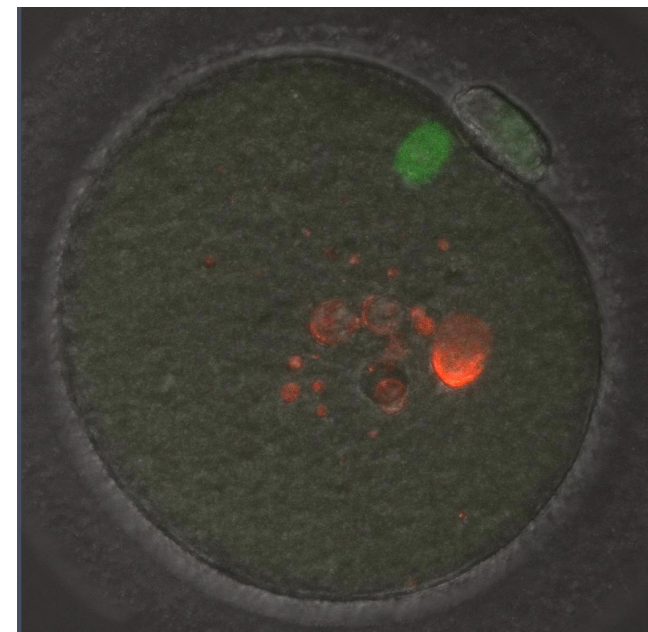
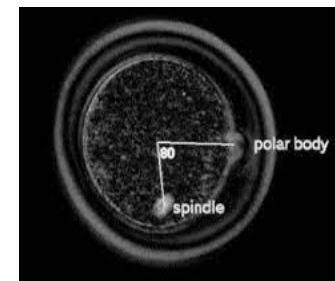
❖ Polarized Light Microscopy (PLM)

- birefringent structures in human oocytes



PM.... Plasma mebrane
ZP.....Zona pellucida
RB....Refractile body
V.....Vacuole

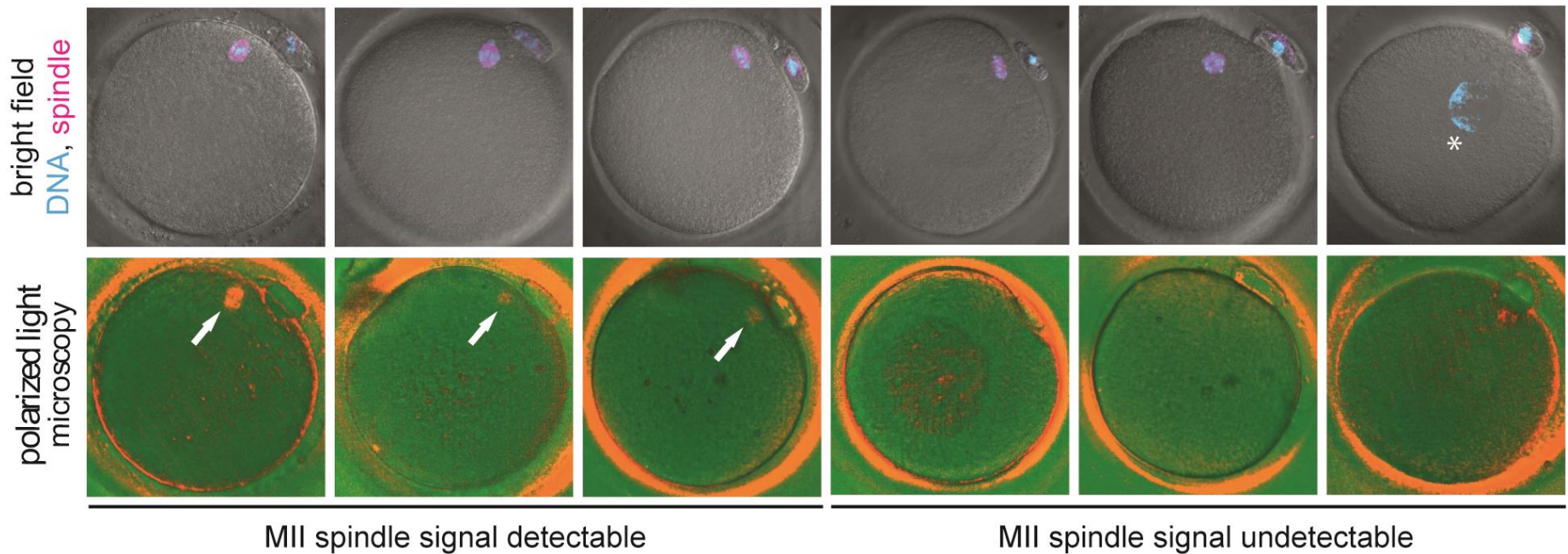
- relative position of spindle and PB



Non-invasive spindle visualization

❖ Polarized Light Microscopy (PLM)

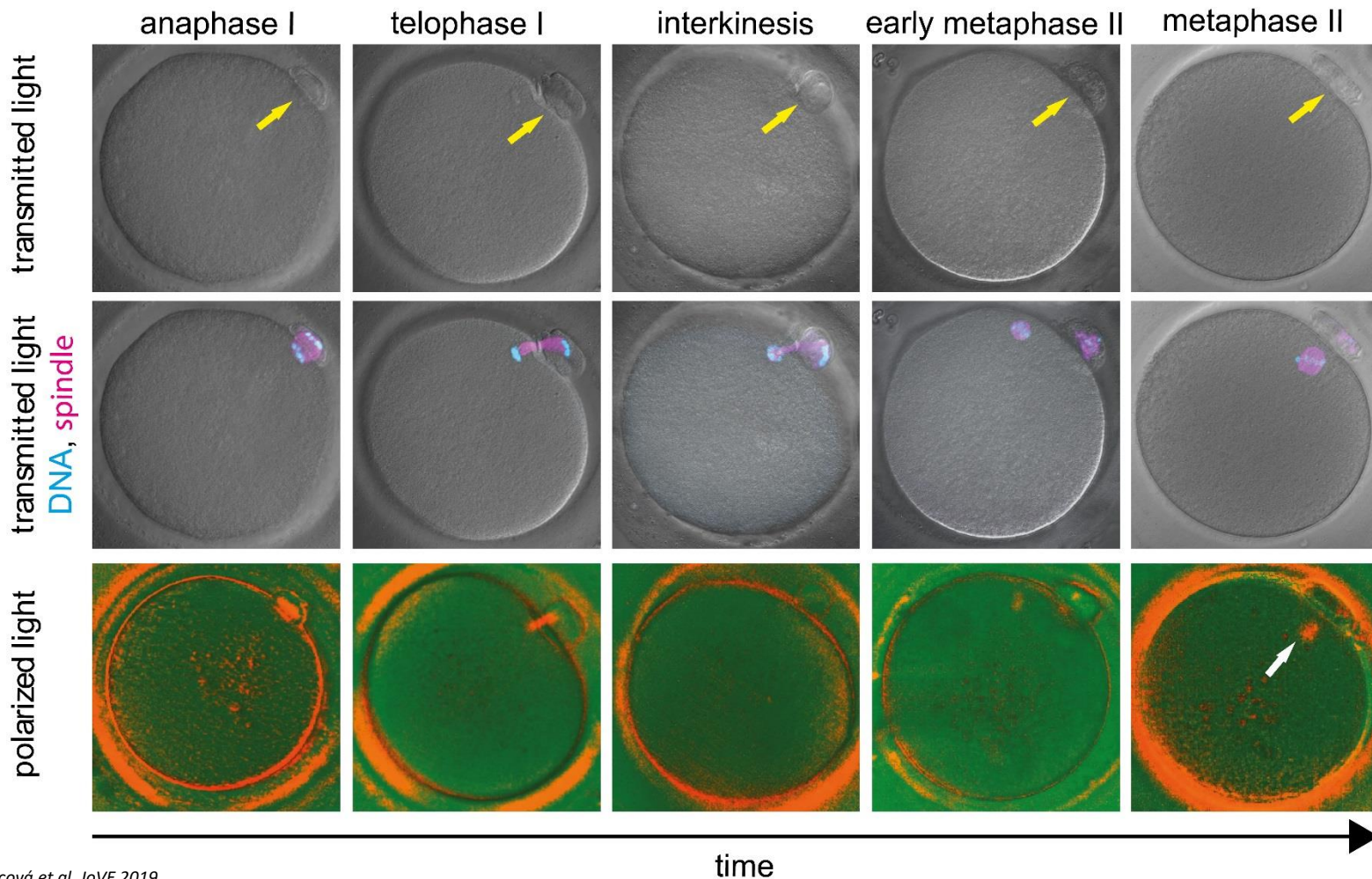
- the strength of the signal reflect the material ordering
- sufficient mass of paralely oriented MT required to produce noticeable signal



Non-invasive spindle visualization

❖ Polarized Light Microscopy (PLM)

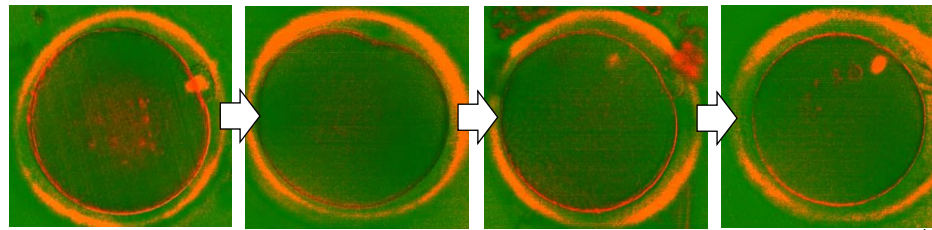
- enables monitoring of MI/MII transition and ICSI time optimisation in clinical practice



Non-invasive spindle visualization

❖ Polarized Light Microscopy (PLM)

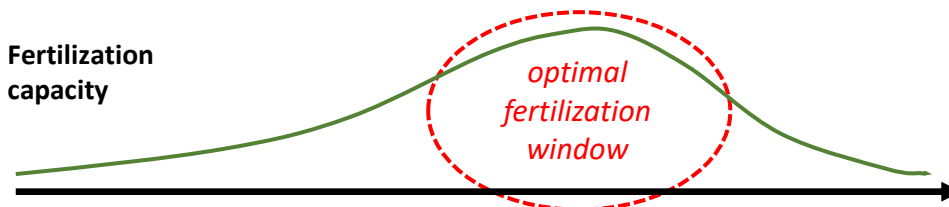
- enables monitoring of MI/MII transition and ICSI time optimisation in clinical practice



Postponing ICSI

time

Fertilization capacity



time

ANAPHASE I

INTERKINESIS

METAPHASE II

EGG IN VITRO AGING



Factor affecting human oocyte spindle stability in vitro

❖ MII spindle is sensitive

➤ temperature

- optimal 37°C

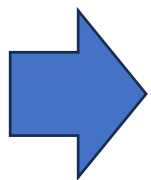


➤ osmolarity alterations

- avoid evaporation
- parafine/mineral oil overlay
- humid conditions

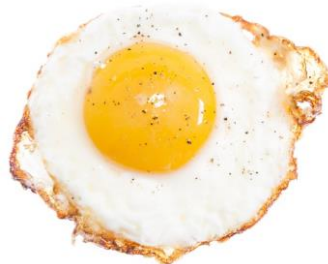
➤ pH fluctuation

- MOPS/HEPES buffered medium for work in ambient conditions



avoid excessive manipulation !

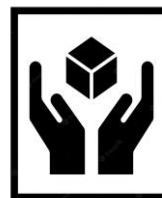
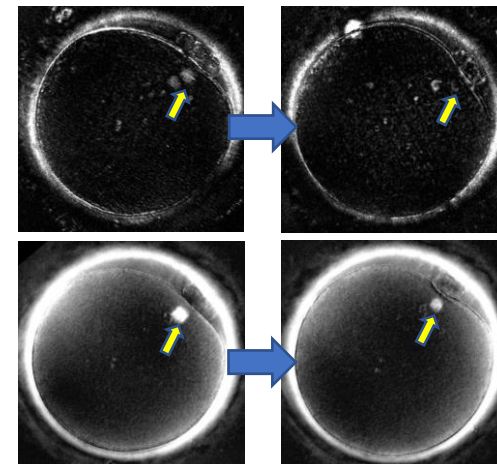
OVERHEATING
→ irreversible denaturation



COOLING
→ spindle desintegration



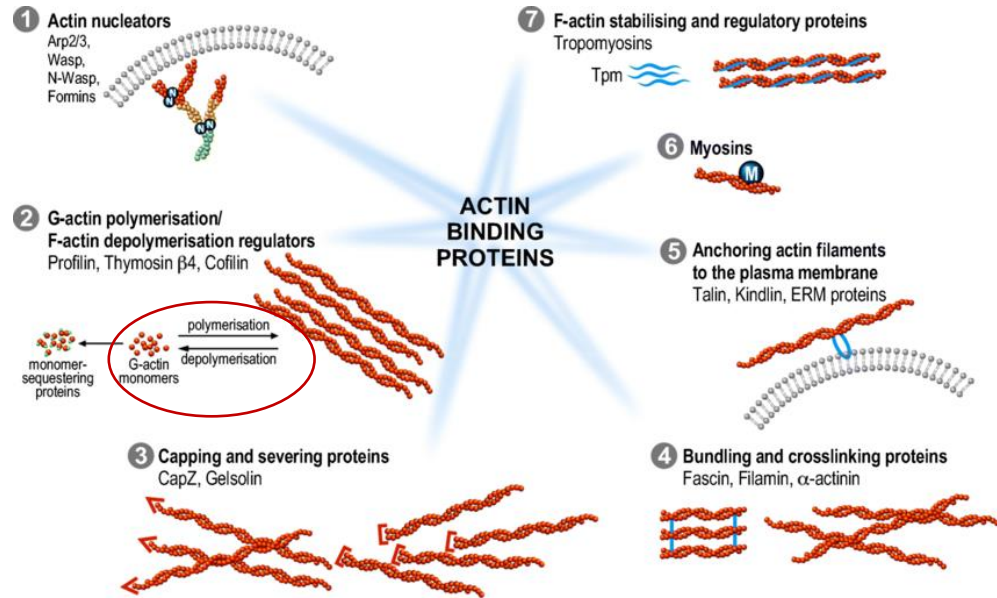
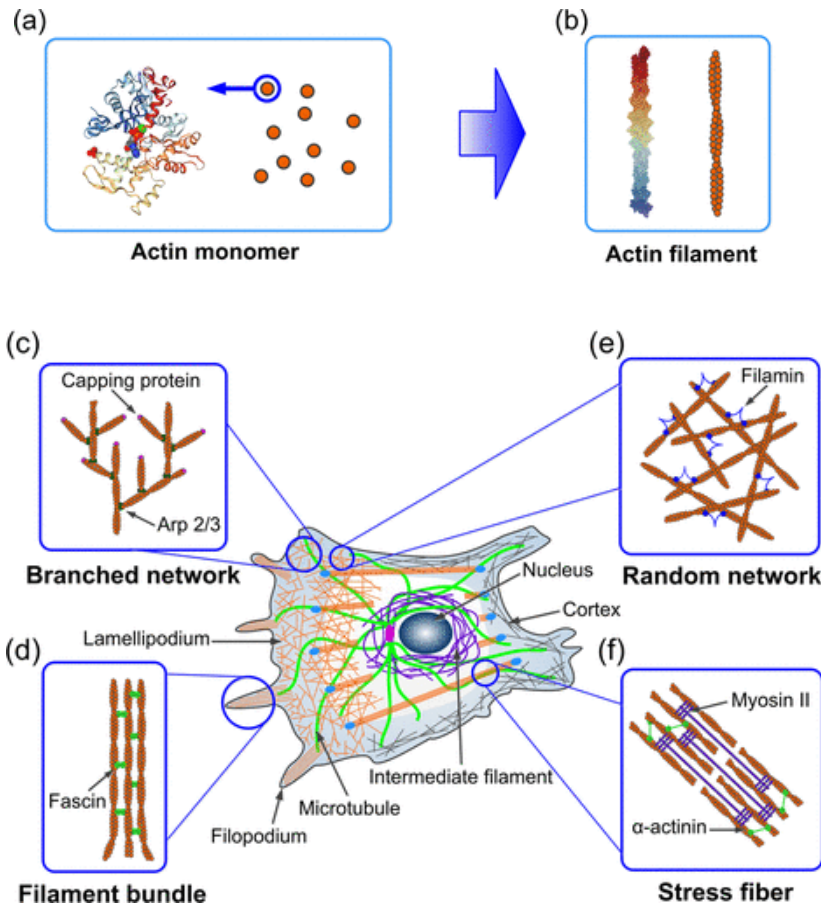
10 min RT



HANDLE WITH CARE

Actin network

- actin network consists of F-actin fibers (microfilaments), formed by dynamic (de)polymerization of globular G-actin monomers



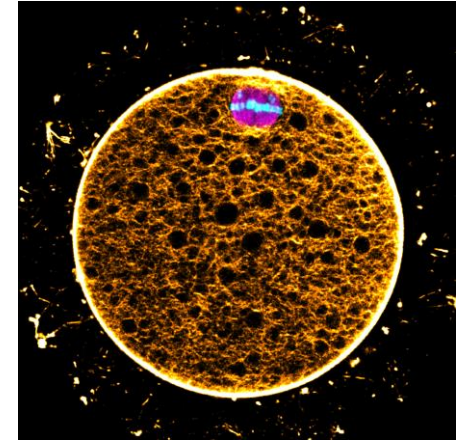
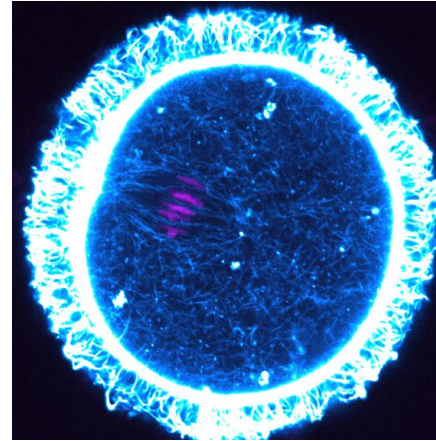
- in association with its binding proteins play versatile roles
(e.g. mechanical support, migration, signalling, trafficking, adhesion, division, contraction,...)

Oocyte actin network

- large oocyte cytoplasm contains **network of longed branched microfilaments** and **cortical actin**

❑ Roles of actin:

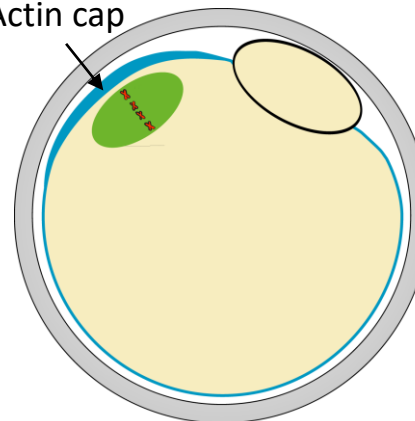
- mechanical support
- vesicular trafficking
- arrangement of cytoplasmic organelles
- spindle formation (with MTs)
- spindle migration
- chromosome alignment promotion
- PB extrusion
- MII spindle anchorage
- membrane polarization



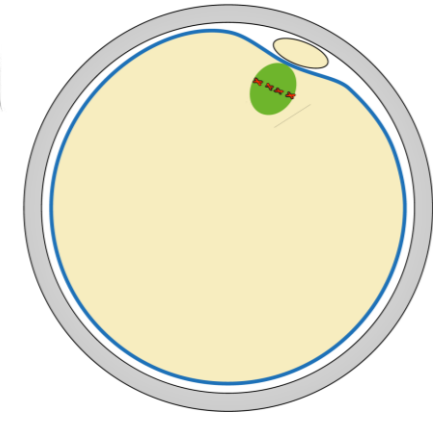
Large volume
Highly asymmetric cell division



Actin cap



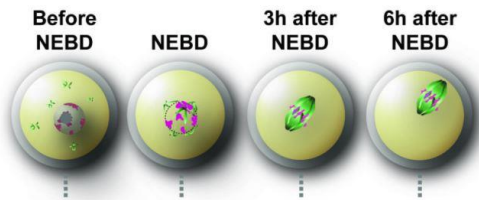
70-80 um



110-120 um

Spindle migration for asymmetric spindle positioning

- in mouse, bipolar spindle is formed centrally

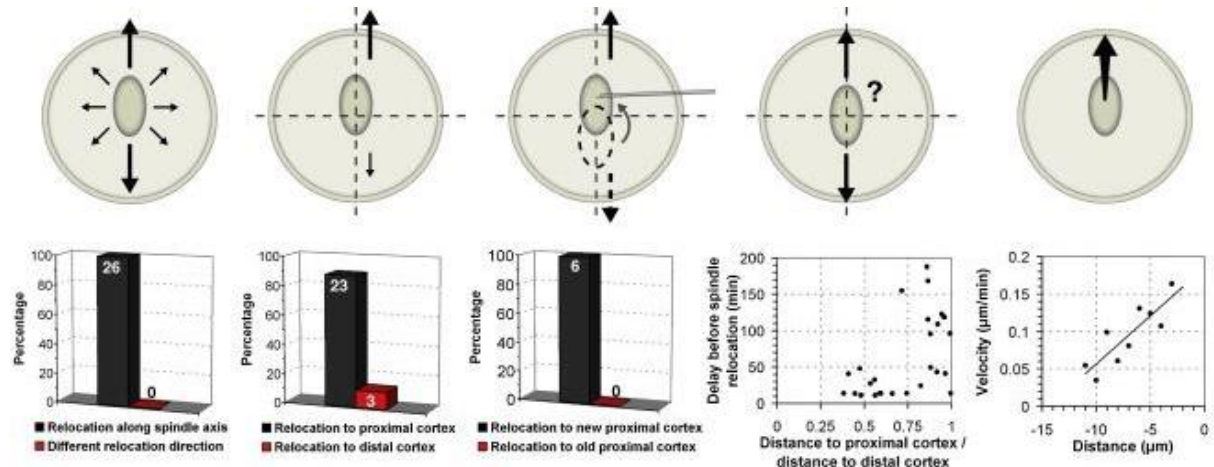
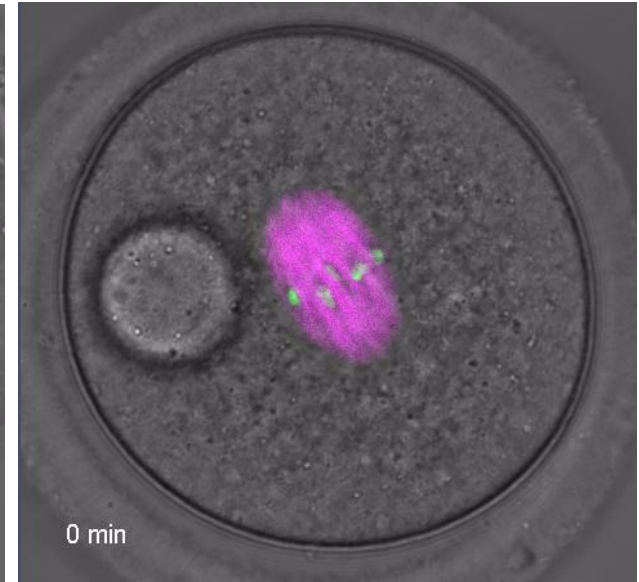


- spindle relocation to cortex ensures high asymmetry of female meiotic division
- spindle migrates along its long axis towards the closest cortex
- spindle accelerates during migration

Spindle relocation

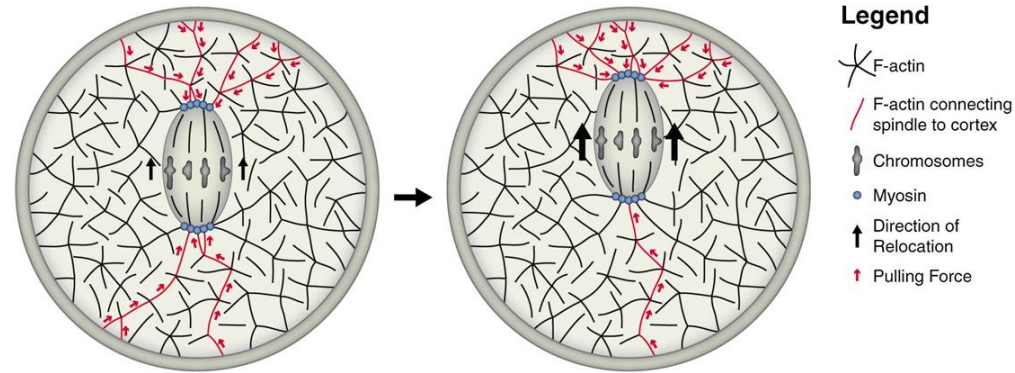


Spindle relocation failure

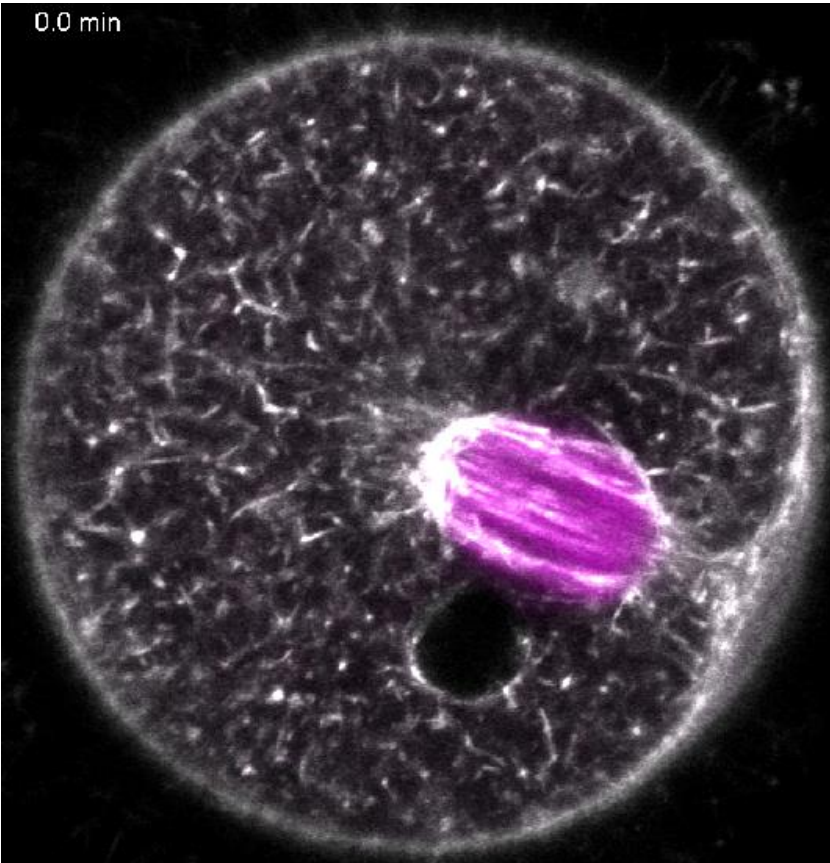


Actin dynamics is required for asymmetric spindle positioning

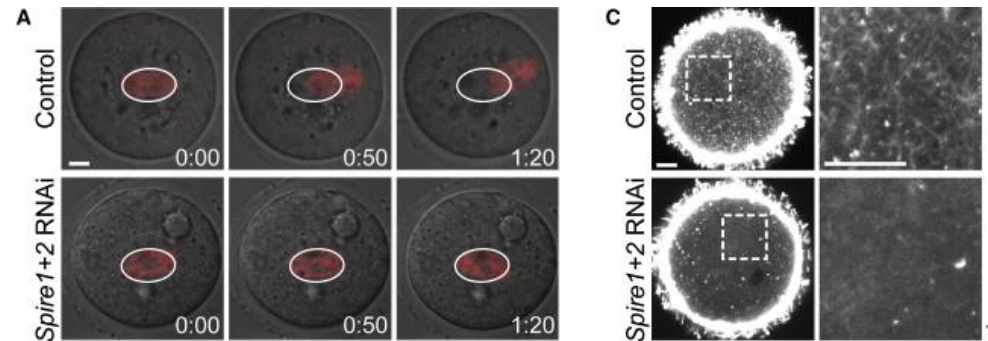
- spindle relocating to the cortex is driven by actin
- spindle pole-associated myosin II pulls the actin filaments against the cortex



0.0 min

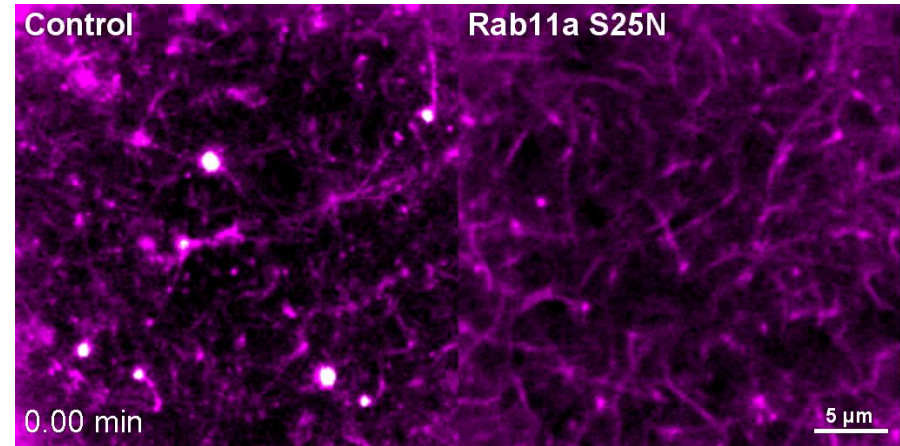
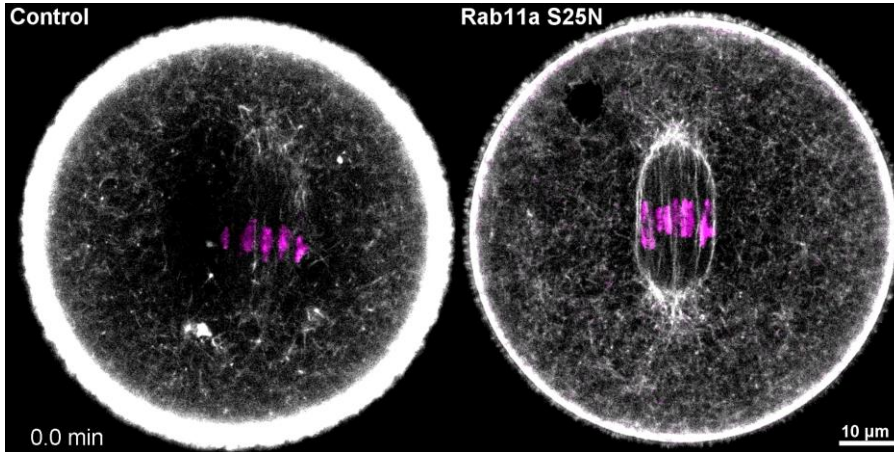


- Active actin nucleation is required for spindle relocation

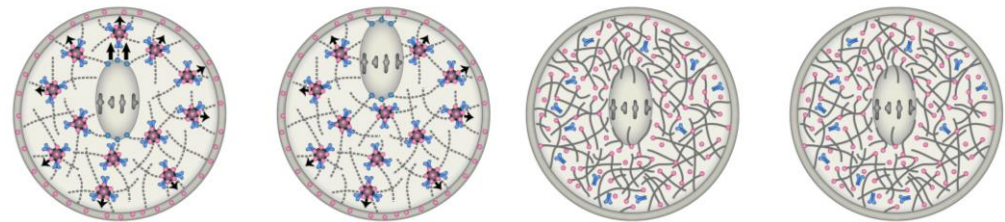


Actin dynamics is required for asymmetric spindle positioning

- vesicle-mediated actin network dynamics and myosin force are required for spindle migration to the cortex



- density and outward directed dynamics of actin network is regulated by size of vesicles sequestering actin nucleators



1. Rab11a-positive vesicles drive the dynamics of the actin network in a Myosin Vb dependent manner.
2. The actin network density is low, because actin nucleators are sequestered at vesicles.
3. The outward directed dynamics of the vesicle-actin network drive asymmetric spindle positioning.

1. The actin network is static, because Rab11a-positive vesicles are missing.
2. The actin network density is increased, because actin nucleators are released from vesicles.
3. The static actin network prevents asymmetric spindle positioning.

Legend

- ✕ Static F-actin
- Vesicle
- ⚡ Myosin Vb
- ⊕ Spindle
- ⊗ Dynamic F-actin
- Actin Nucleator
- ⚡ Myosin-dependent pulling from spindle poles

Actin dynamics is required for asymmetric spindle positioning

Developmental Cell

CellPress

Article

Microtubule organizing centers regulate spindle positioning in mouse oocytes

Daniela Londoño-Vásquez,¹ Katherine Rodriguez-Lukey,¹ Susanta K. Behura,¹ and Ahmed Z. Balboula^{1,2,3,*}

¹Animal Sciences Research Center, University of Missouri, Columbia, MO 65211, USA

²University of Cambridge, Department of Genetics, Downing Street, Cambridge, CB2 3EH, UK

³Lead contact

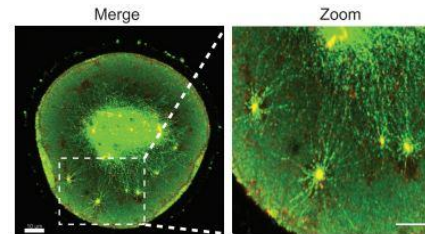
*Correspondence: abalboula@missouri.edu

<https://doi.org/10.1016/j.devcel.2021.12.011>

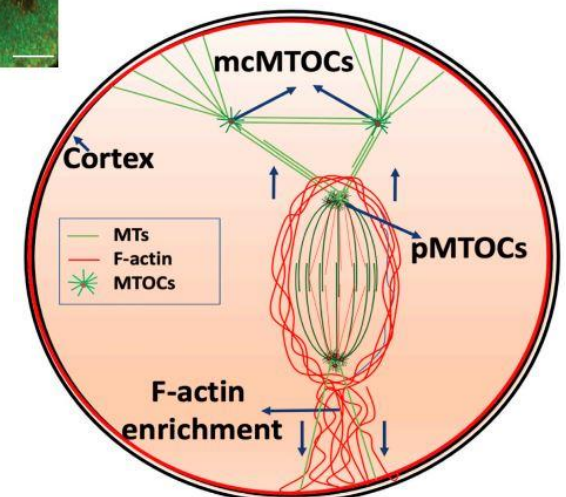


- identification of subset of metaphase cytoplasmic MTOCs (**mcMTOCs**) that do not contribute to spindle assembly and localize opposite to PBE side

- mcMTOCs are interconnected with polar MTOCs and regulate spindle positioning by anchoring the spindle to the oocyte cortex

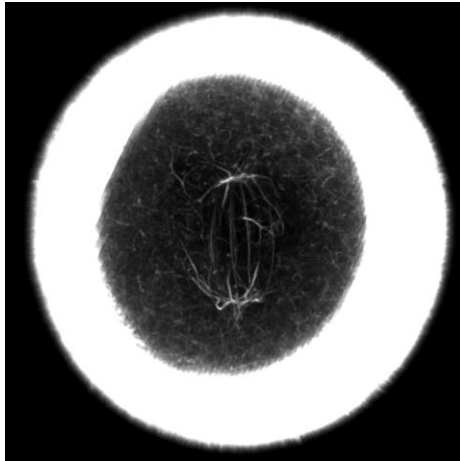


- actin-mediated movement of the meiotic spindle to the cortex is balanced by forces exerted from mcMTOCs to ensure the timely migration and asymmetric division

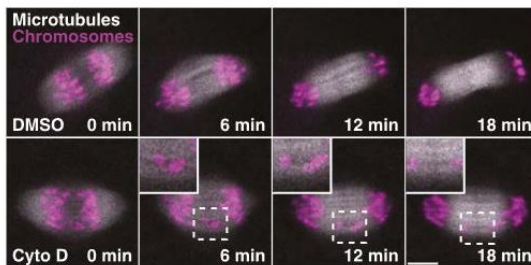
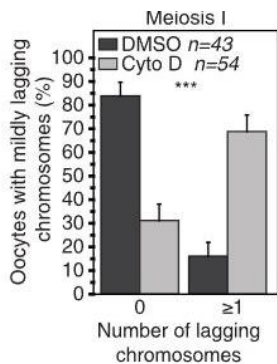


Spindle actin promote chromosome alignment

- actin filaments permeate meiotic spindle in mammalian oocytes



- actin prevents lagging chromosomes and promotes chromosome congression and alignment



RESEARCH ARTICLE

CELL BIOLOGY

Actin protects mammalian eggs against chromosome segregation errors

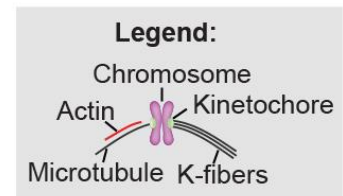
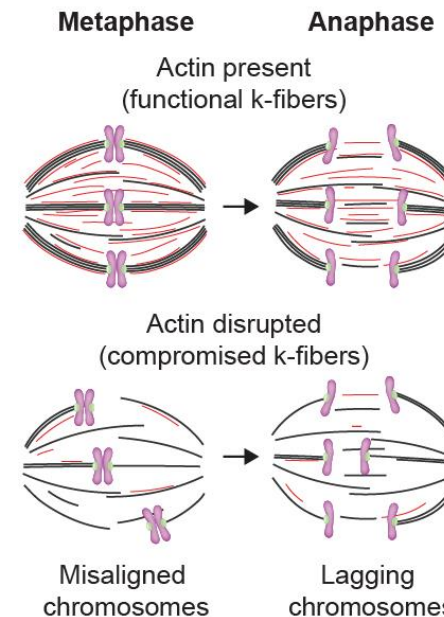
Binyam Mogessie and Melina Schuh*



Binyam Mogessie



Melina Schuh



- dynamic actin promotes formation of kinetochores fibers

Actin and microtubule cooperate to insure spindle integrity



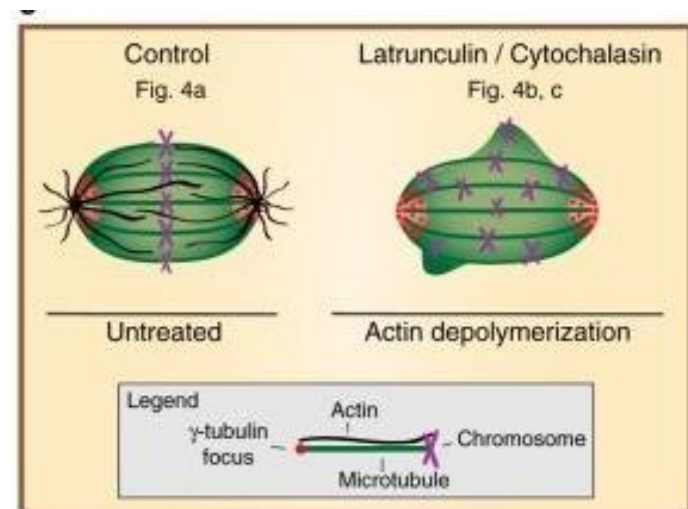
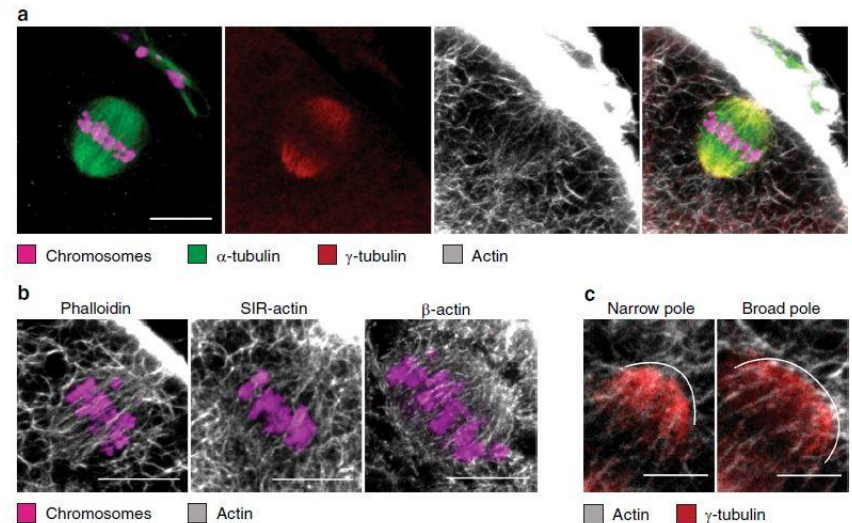
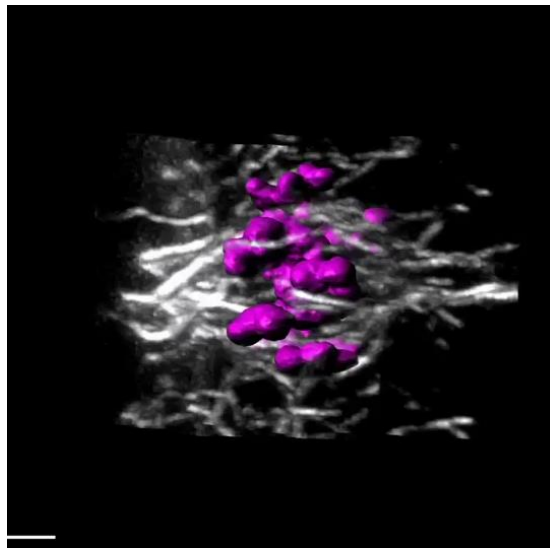
ARTICLE

<https://doi.org/10.1038/s41467-019-12674-9> OPEN

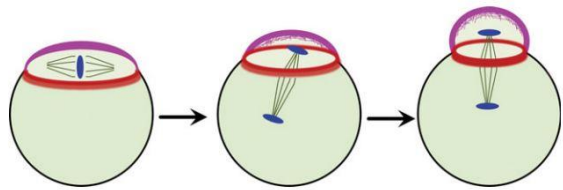
Actin-microtubule interplay coordinates spindle assembly in human oocytes

Johannes Roeles¹ & Georgios Tsiavaliaris^{1*}

- actin fibers permeate spindle and structural integrity of actin spindle is dependent on microtubules
- actin and microtubules co-operate to assemble functional spindle and help to align chromosomes in human oocytes



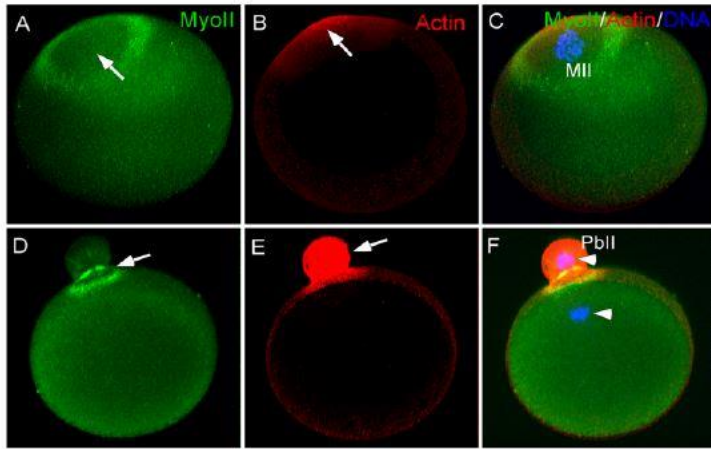
PB extrusion



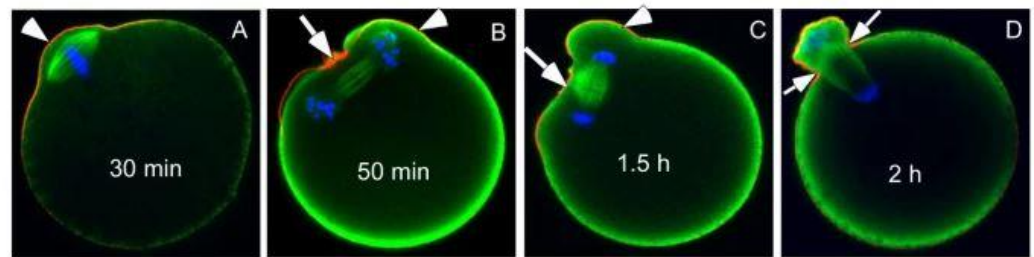
in mice PBE requires spindle rotation



- (1) cortical membrane protrusion (**actin cap**)
- (2) spindle midzone induced membrane furrowing
- (3) **actomyosin ring** constriction
- (4) abscission

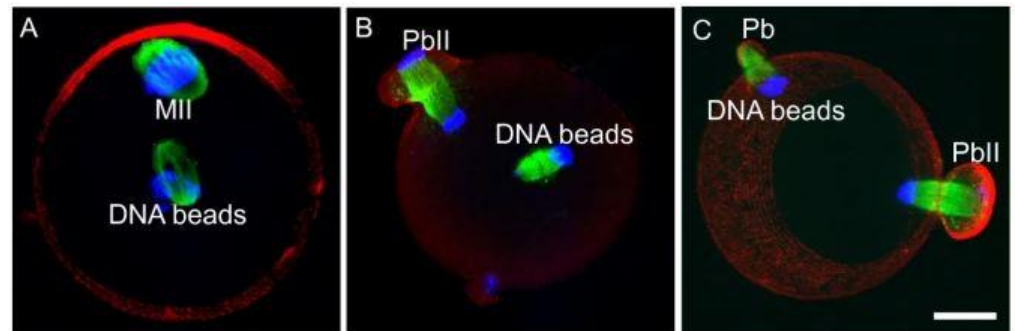


Deng et al 2007



Wang et al 2011

- Furrow induction by spindle midzone is distance-dependent

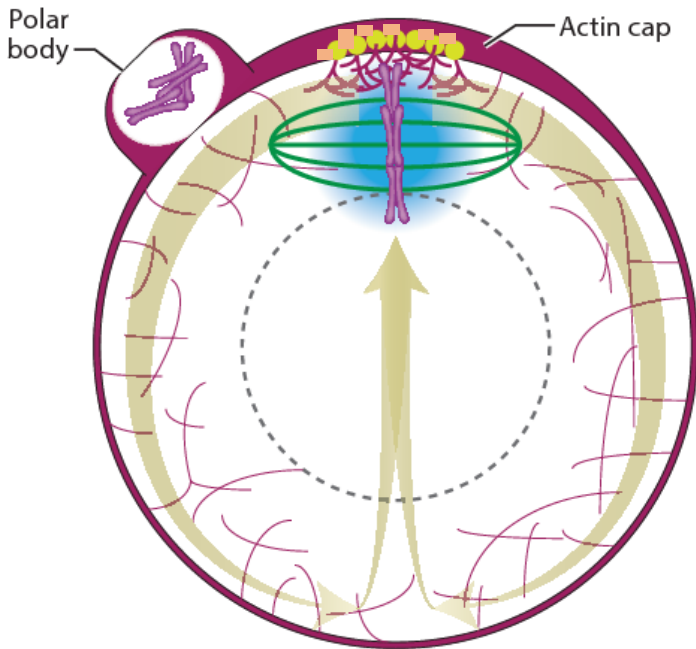


* Injection of DNA beads to MII oocytes

Wang et al 2011

MII spindle anchorage

Meiosis II spindle anchorage



Dynamic maintenance of asymmetric meiotic spindle position through Arp2/3 complex-driven cytoplasmic streaming in mouse oocytes

Kexi Yi^a, Jay R. Unruh^a, Manqi Deng^b, Brian D. Slaughter^a, Boris Rubinstein^a, and Rong Li^{a,c}



Rong Li

Ran gradient



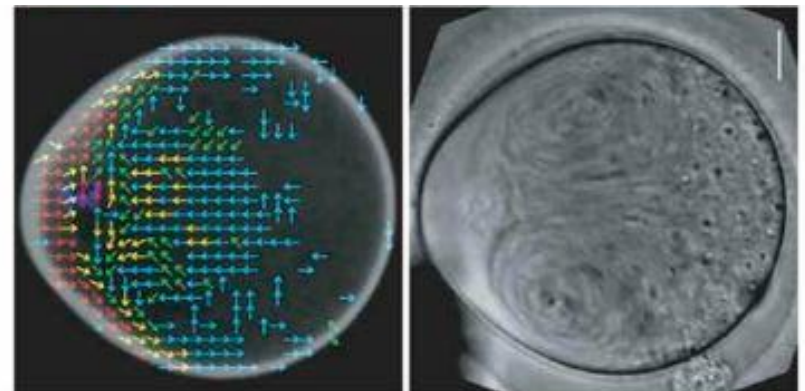
local activation of N-WASP



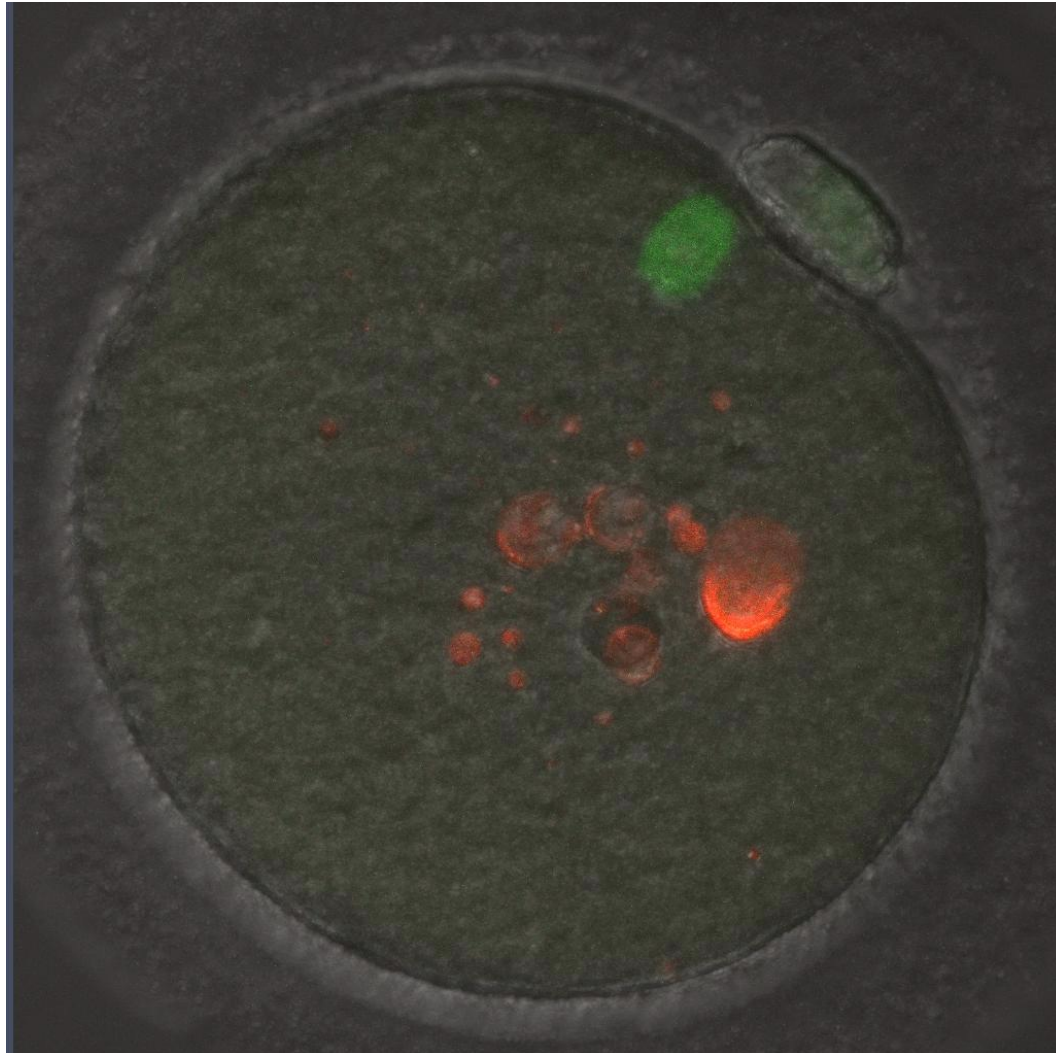
- Arp2/3 complex localization
 - promotes local actin nucleation → actin cap
 - ensures spindle anchoring during prolonged MII stage

— Actin
— Microtubule
● Ran-GTP
➔ Cytoplasmic streaming
■ N-WASP
● Arp2/3

- Arp2/3 nucleation activity initiates retrograde flow of F-actin provoking cytoplasmic streaming that further pushes on spindle to maintain its cortical position



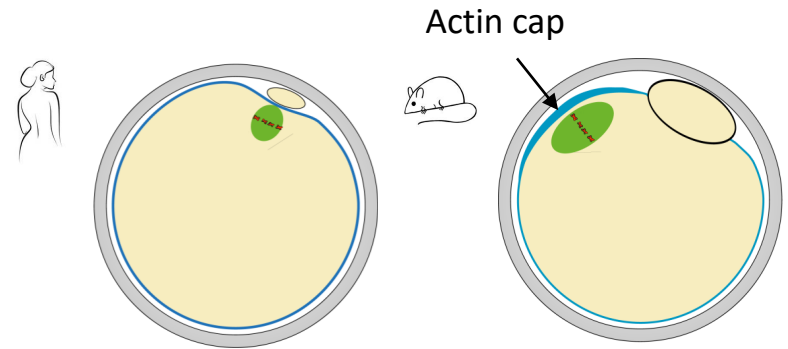
MII spindle anchorage



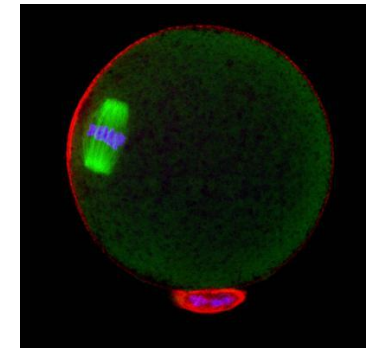
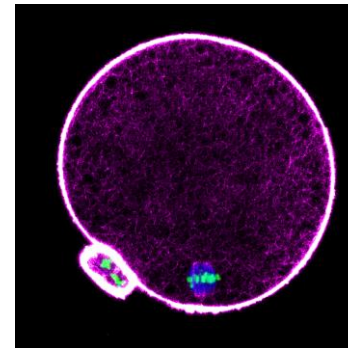
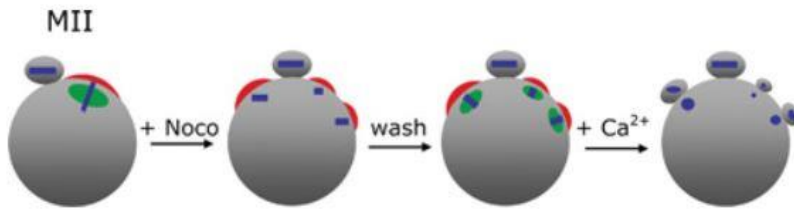
(unpublished)

Actin polarization

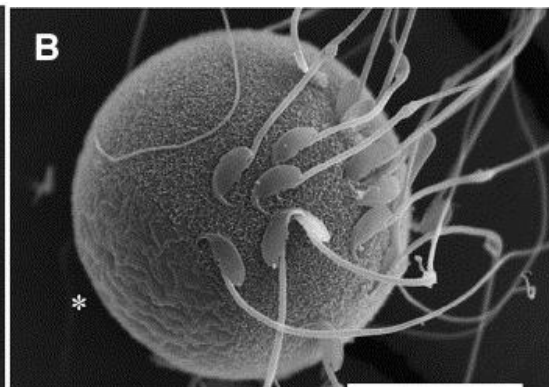
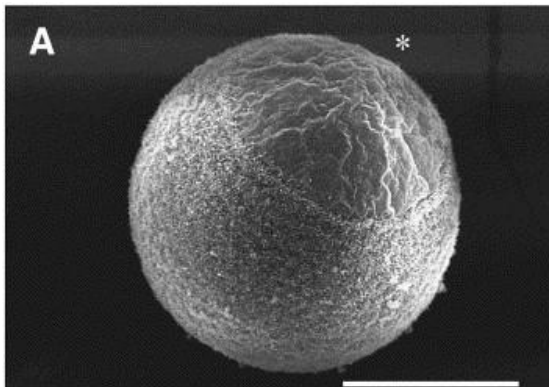
- actin thickening - „actin cap“
 - region overlying MII spindle
 - actin-enriched microvilli-free zone devoid of cortical granules



- induced by spindle chromatin underneath the oolema



Longo and Chen, 1985; Maro et al., 1986

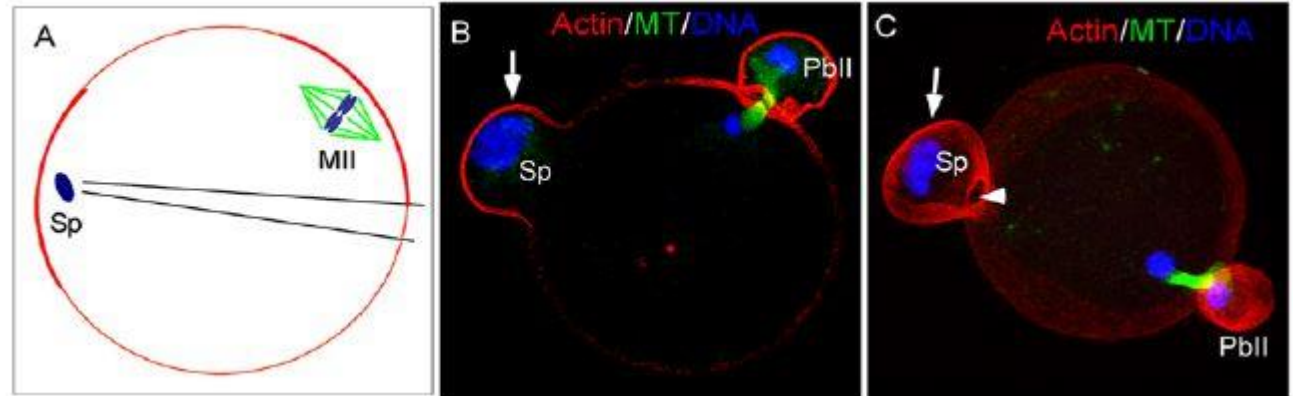


- prominent in mouse but not human oocytes!

Actin polarization

- etopic actin polarization
 - induced artificially by DNA beads or by **sperm chromatin during ICSI**

„fertilization cone“



Deng and Li 2009

Transient phenomenon
or can lead to „3rd PB
extrusion“

