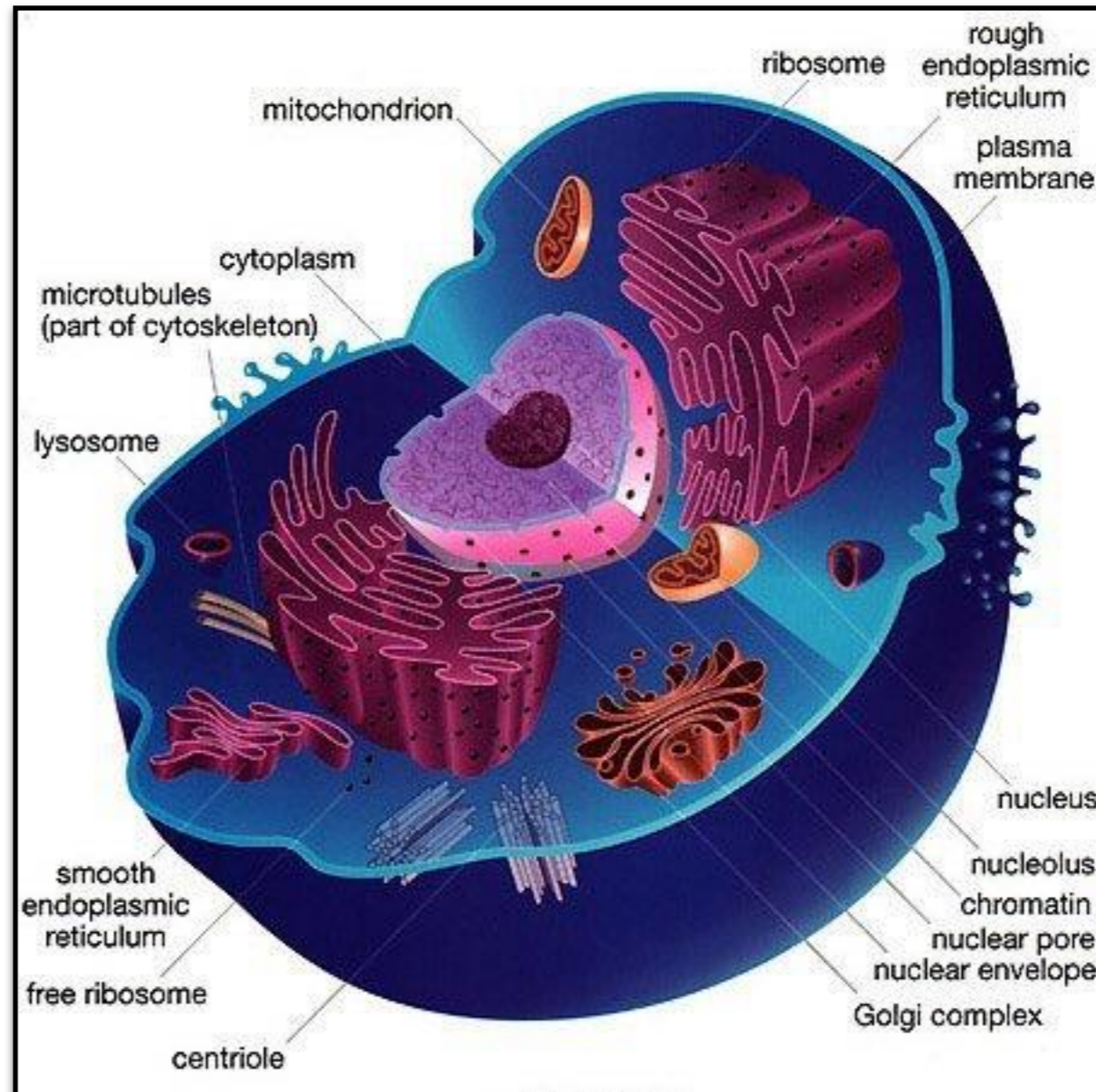


# Cancer as a metabolic disease

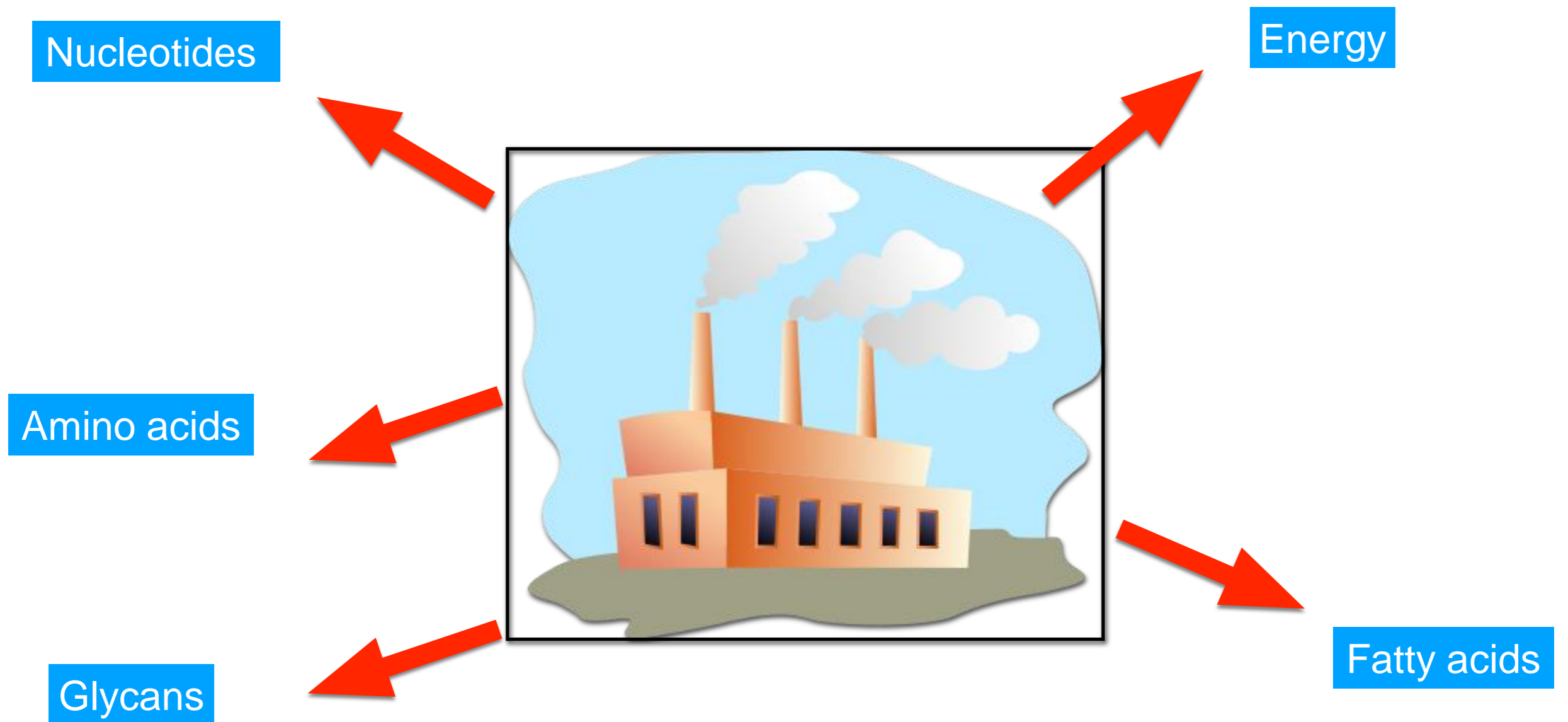
Yoav Shaul

Department of Biochemistry and Molecular Biology  
The Institute for Medical Research Israel-Canada  
The Hebrew University Medical School  
Jerusalem Israel

# Mammalian cell

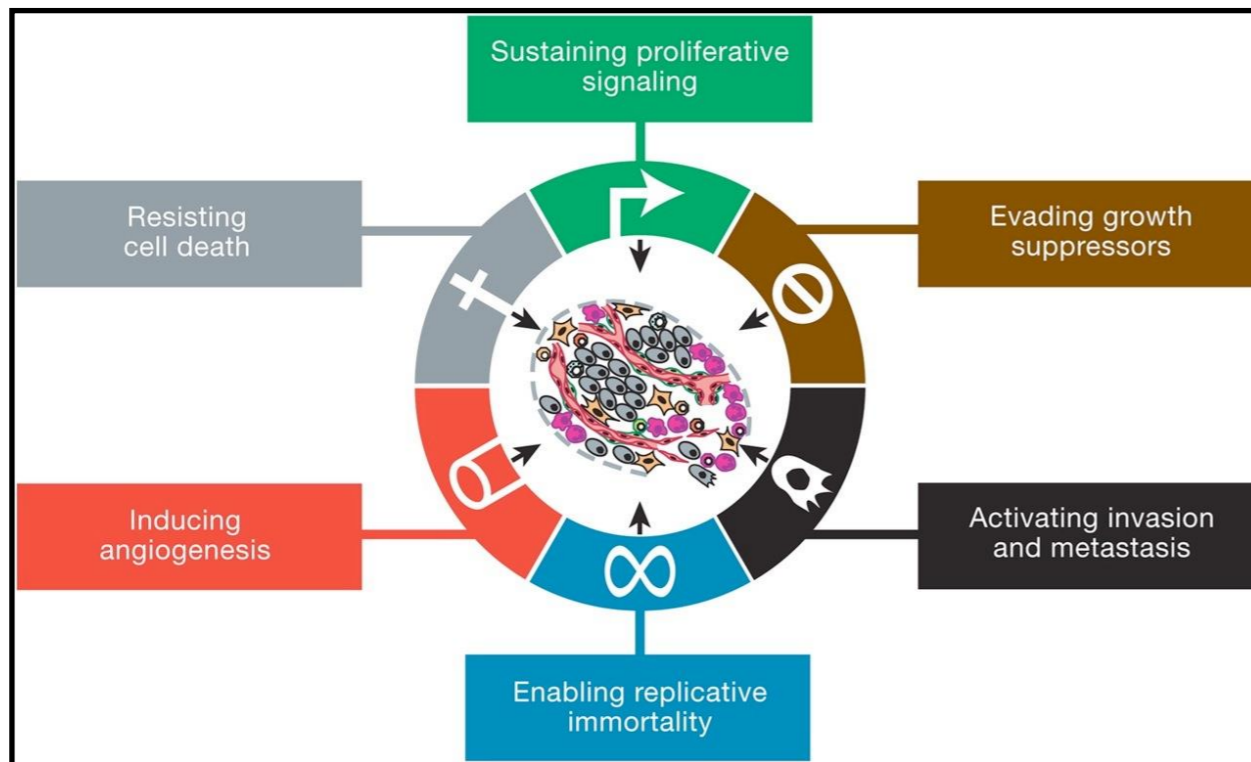


# Mammalian cell as a factory



# Hallmark of cancer

The most fundamental trait of cancer cells involves their ability to sustain chronic proliferation.



Cell, Vol. 100, 57–70, January 7, 2000, Copyright ©2000 by Cell Press

## The Hallmarks of Cancer

Douglas Hanahan\* and Robert A. Weinberg†

\*Department of Biochemistry and Biophysics and  
Hormone Research Institute  
University of California at San Francisco  
San Francisco, California 94143

†Whitehead Institute for Biomedical Research and  
Department of Biology  
Massachusetts Institute of Technology  
Cambridge, Massachusetts 02142

# Emerging hallmark of cancer

Cancer metabolism

## Hallmarks of Cancer: The Next Generation

Douglas Hanahan<sup>1,2,\*</sup> and Robert A. Weinberg<sup>3,\*</sup>

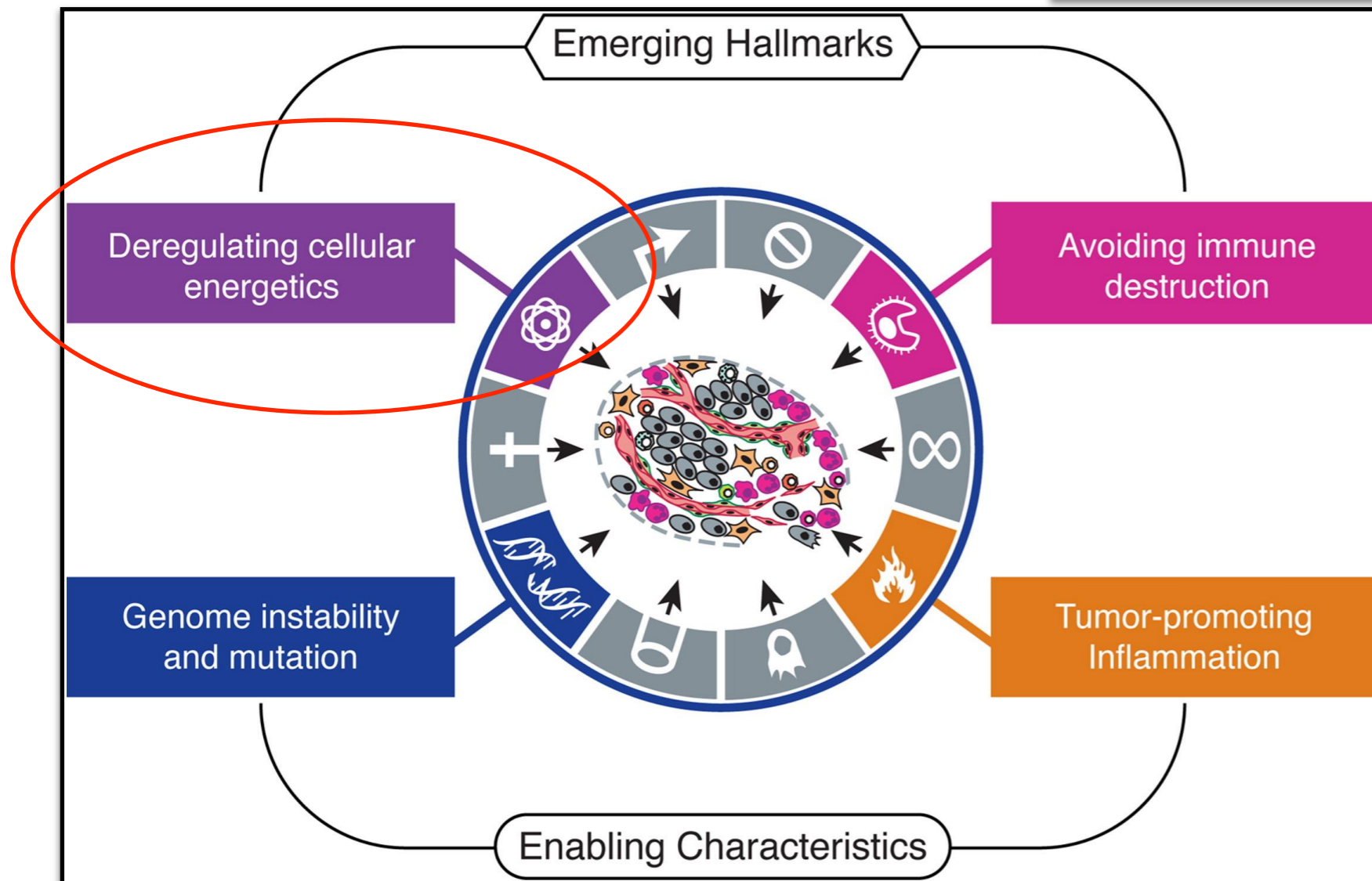
<sup>1</sup>The Swiss Institute for Experimental Cancer Research (ISREC), School of Life Sciences, EPFL, Lausanne CH-1015, Switzerland

<sup>2</sup>The Department of Biochemistry & Biophysics, UCSF, San Francisco, CA 94158, USA

<sup>3</sup>Whitehead Institute for Biomedical Research, Ludwig/MIT Center for Molecular Oncology, and MIT Department of Biology, Cambridge, MA 02142, USA

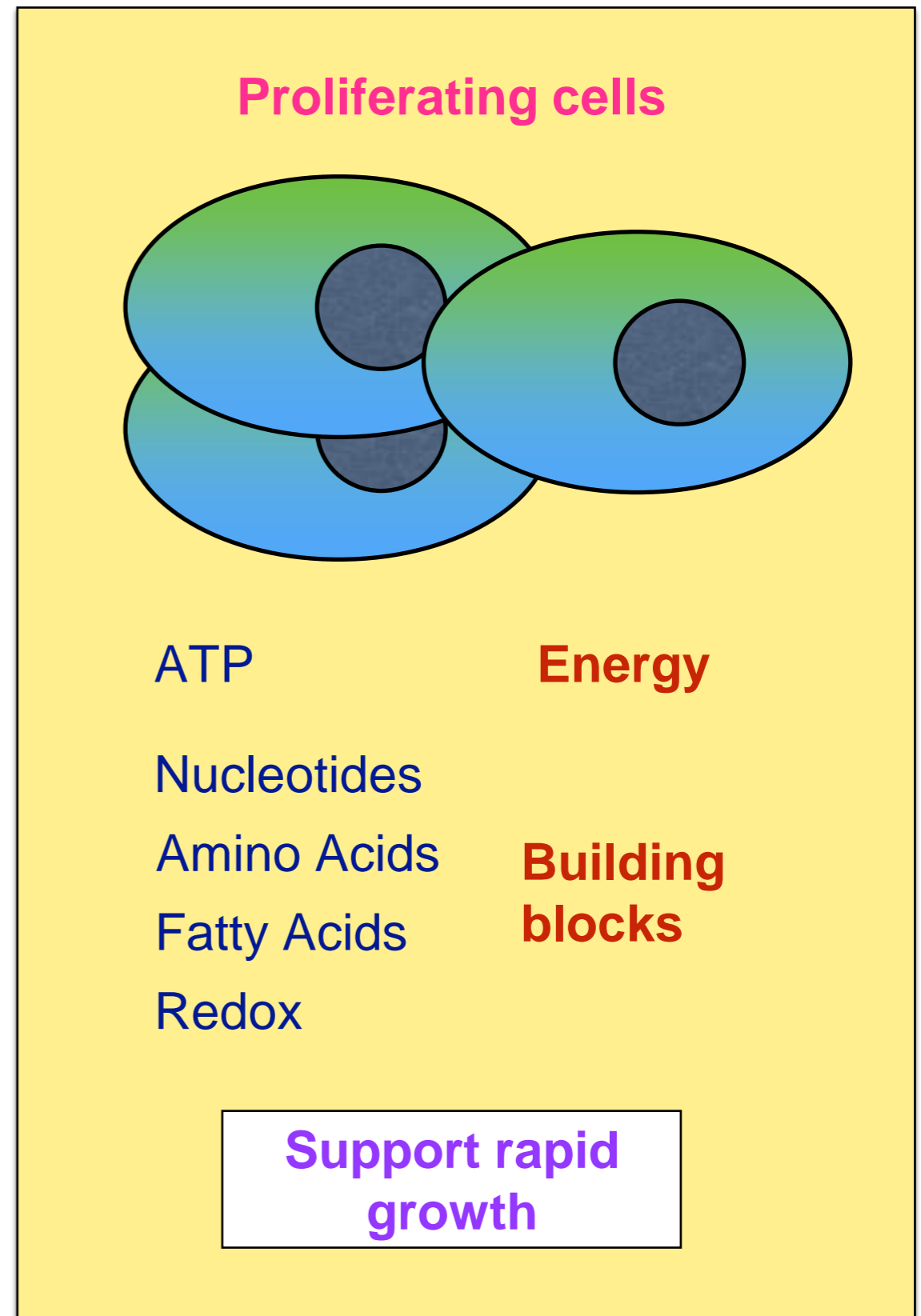
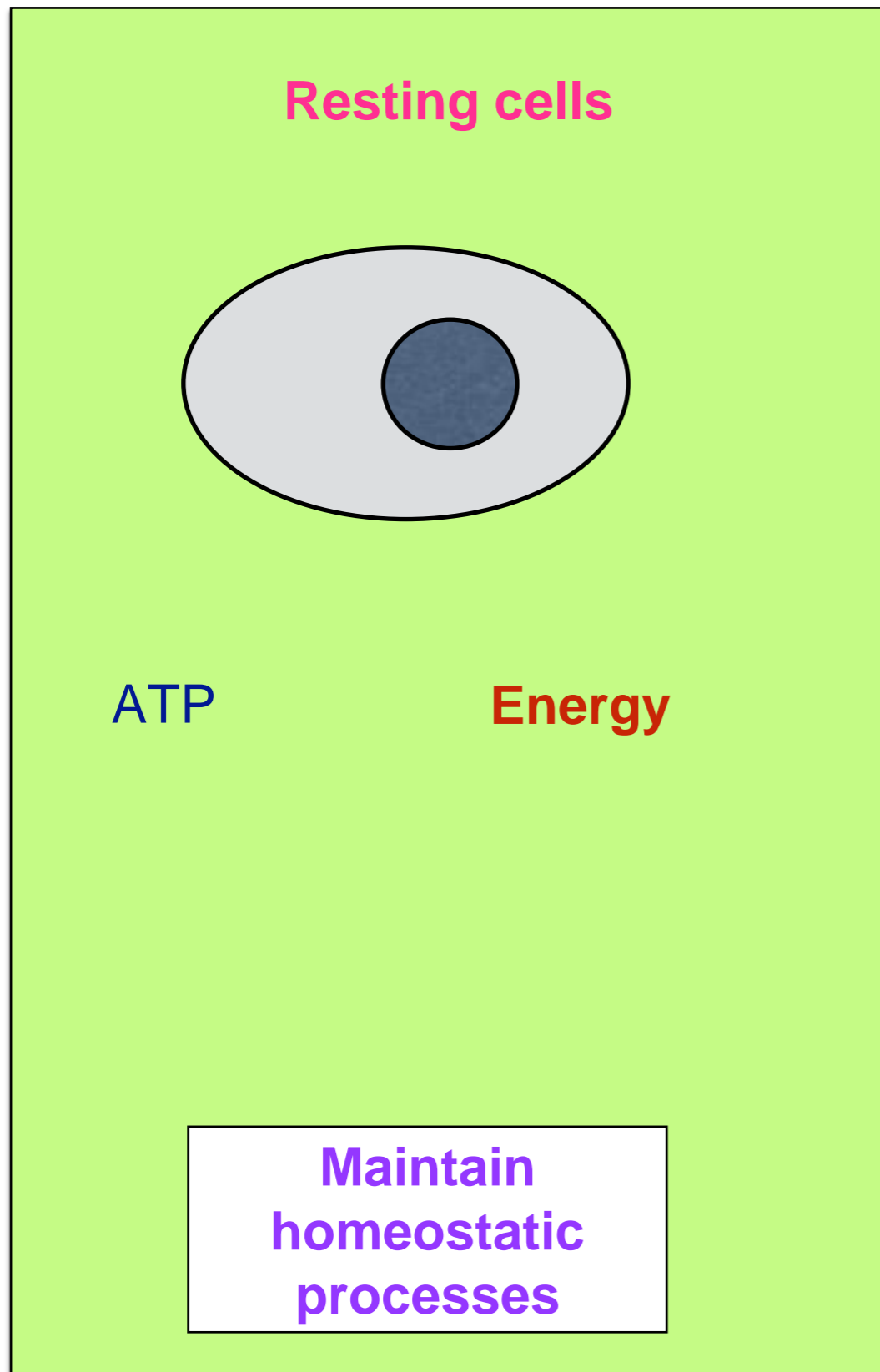
\*Correspondence: dh@epfl.ch (D.H.), weinberg@wi.mit.edu (R.A.W.)

DOI 10.1016/j.cell.2011.02.013



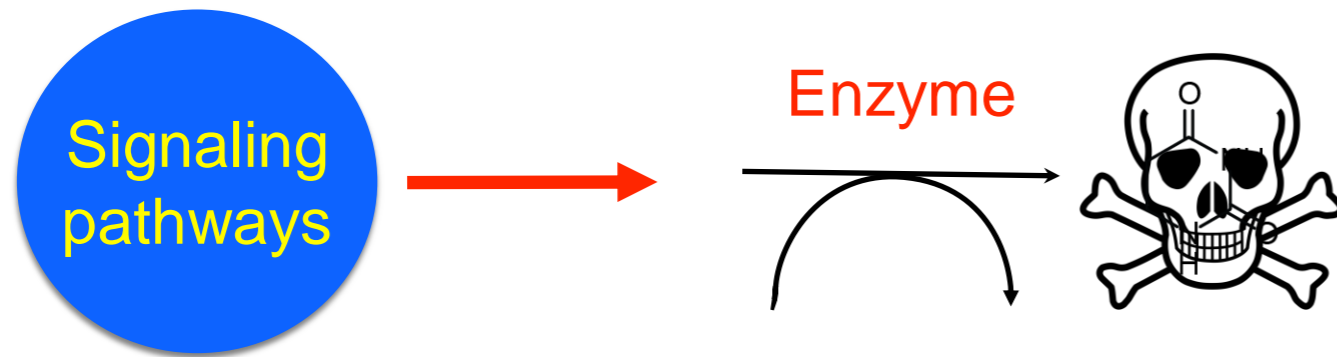


# The metabolism of cell proliferation



# Cancer metabolism: current challenges

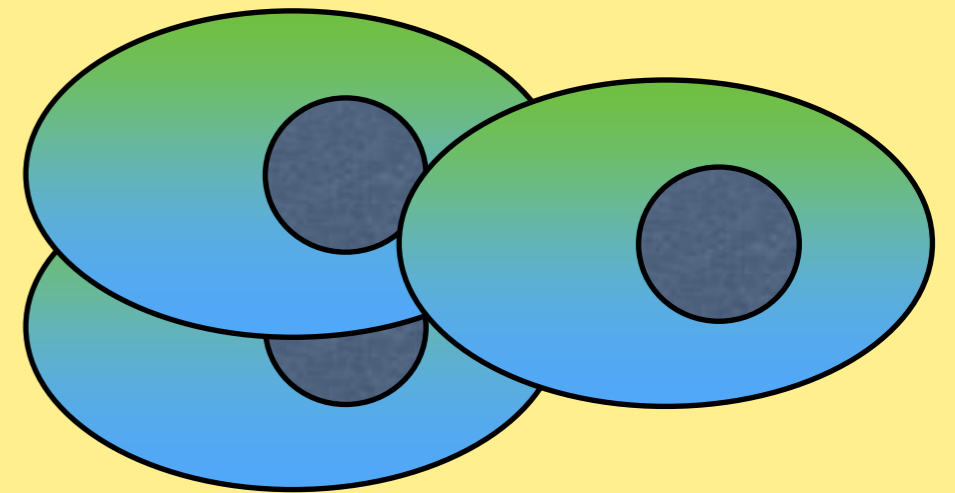
Which metabolic pathways are essential for cancer cells?



Metabolic processes that are essential for cancer cells



Proliferating cells



Building blocks

Support rapid growth

# Leading Questions

## Warburg Effect

What are the molecular mechanisms that regulate the WA?

Is WA a byproduct or an integrated part of the cancer

## Metabolism and cancer

Can metabolism be the drivers of cancers?

## Amino acid metabolism in cancer

What is the role of AA metabolism in cancer

## Oncogenic signaling and metabolic pathways in cancer

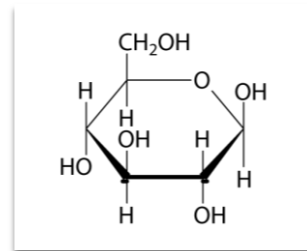
mTOR pathway as an example

## Antimetabolites

How can we exploit metabolic dependency for tumor inhibition?



# Glycolysis



- 6 carbon molecule
- 3 carbon molecule

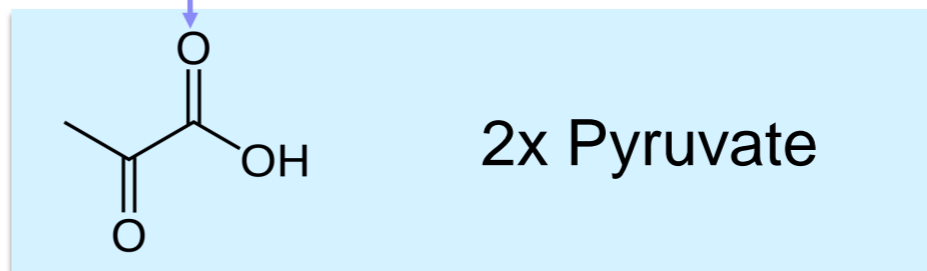
The glycolysis pathways yields 2mol ATP/mol of glucose

-1 ATP

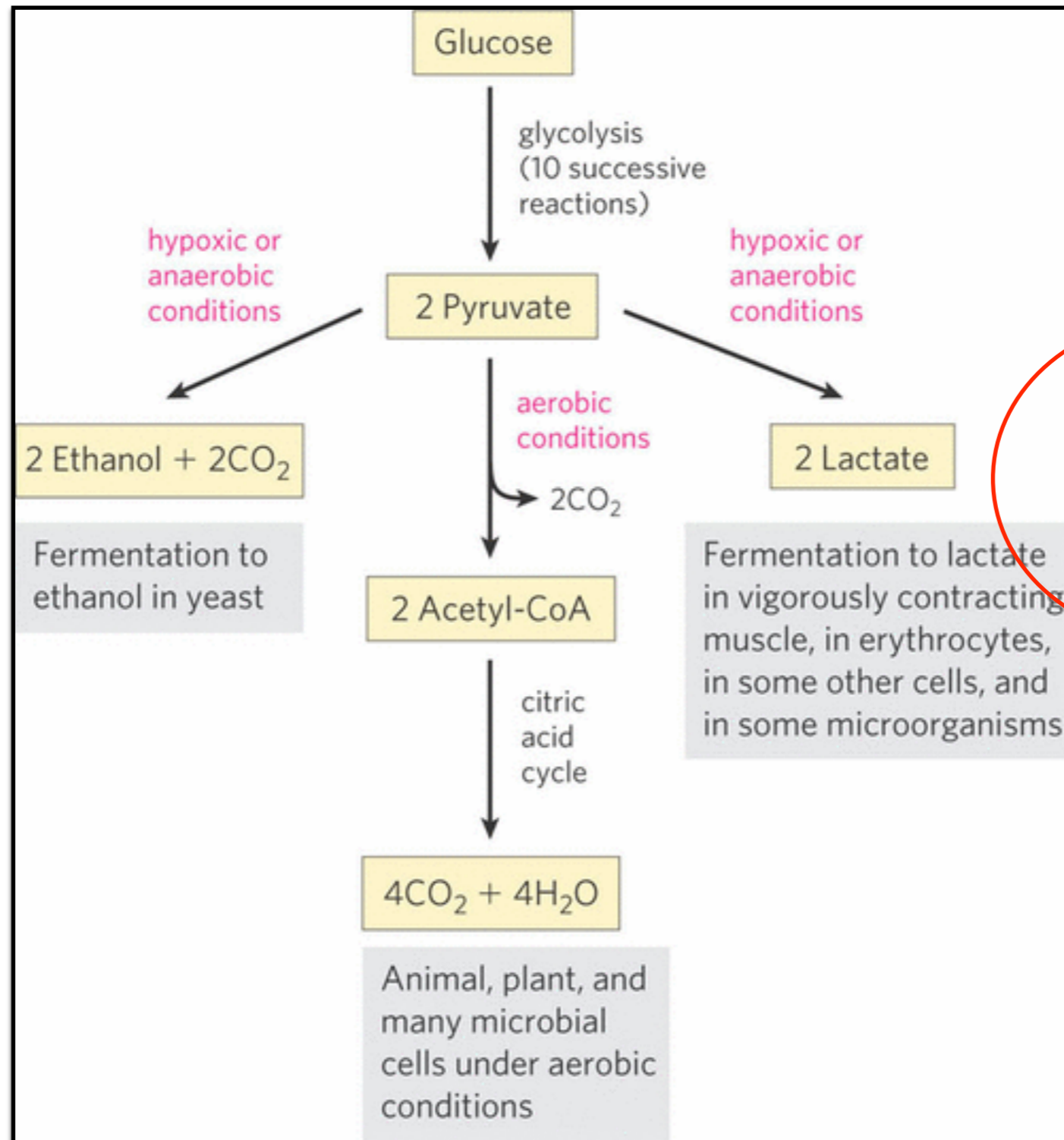
-1 ATP

+2 ATP

+2 ATP



# Catabolic fate of pyruvate



Fermentation

Lactate

TCA cycle

Figure 14-4 Lehninger

# Lactic acid is produced under aerobic conditions

- **Lactic acid** fermentation—occurs in **muscle cells**

Lactic acid is produced in the muscles during rapid **exercise** when the body **cannot** supply enough **oxygen** to the **tissues**—causes **burning sensation** in muscles

glucose  $\longrightarrow$  lactic acid + carbon dioxide + **2 ATP**

**Muscle Cramps**

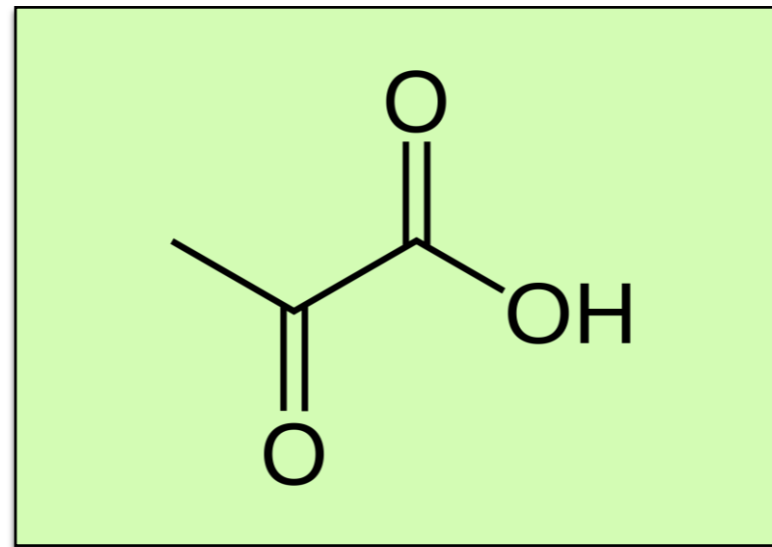


# The Warburg effect



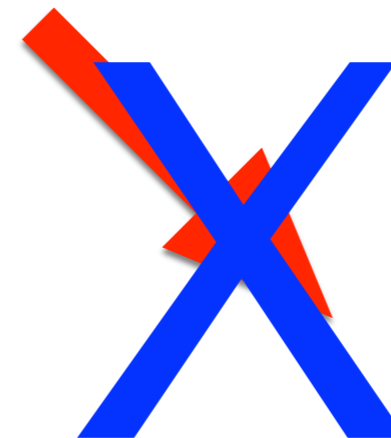
Otto Warburg

Pyruvate



Anaerobic glycolysis

$-O_2$



$+O_2$

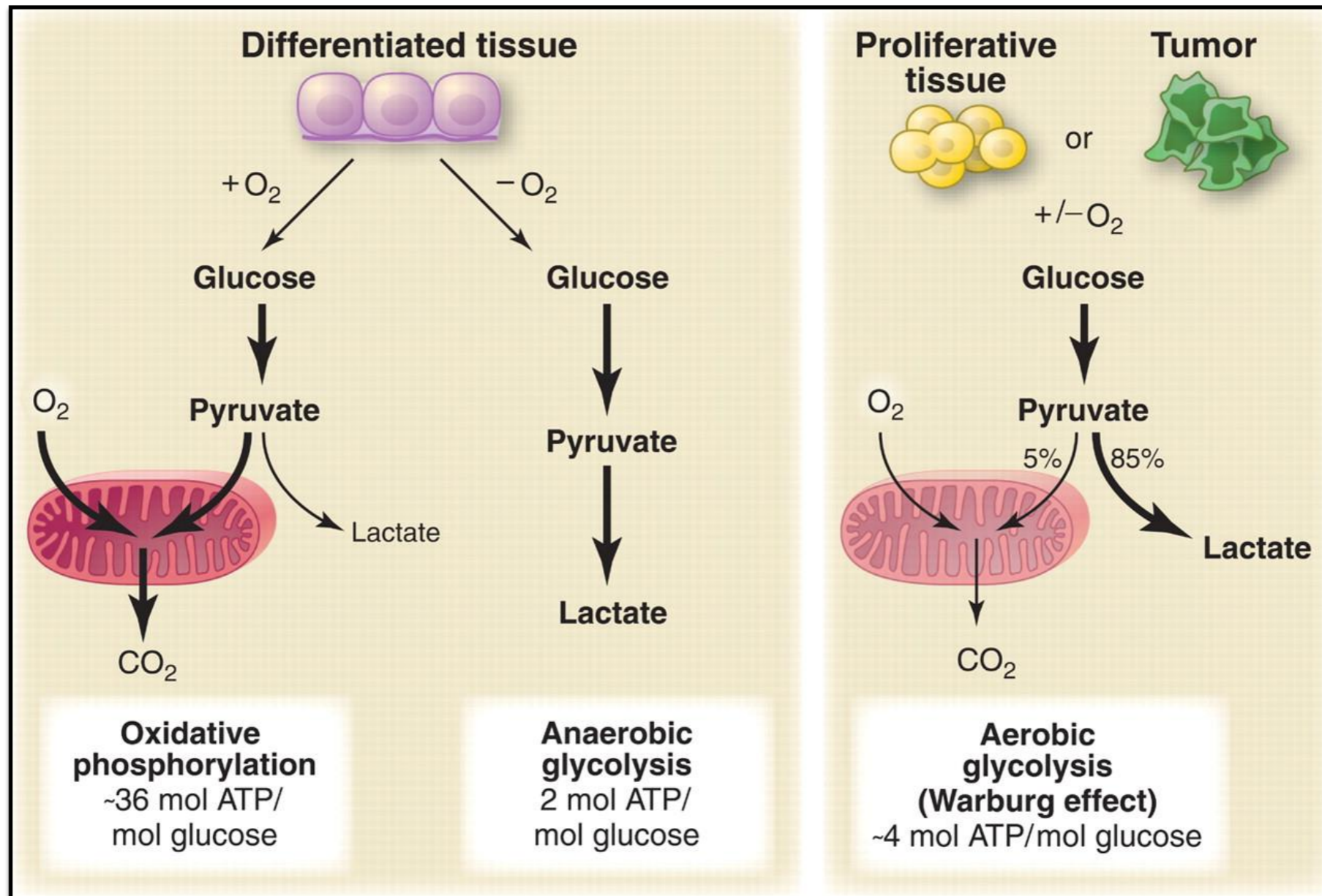
**Aerobic glycolysis**

2 mol ATP/mol of glucose

31 mol ATP/mol of glucose

First indication of cancer dependent metabolic remodelling

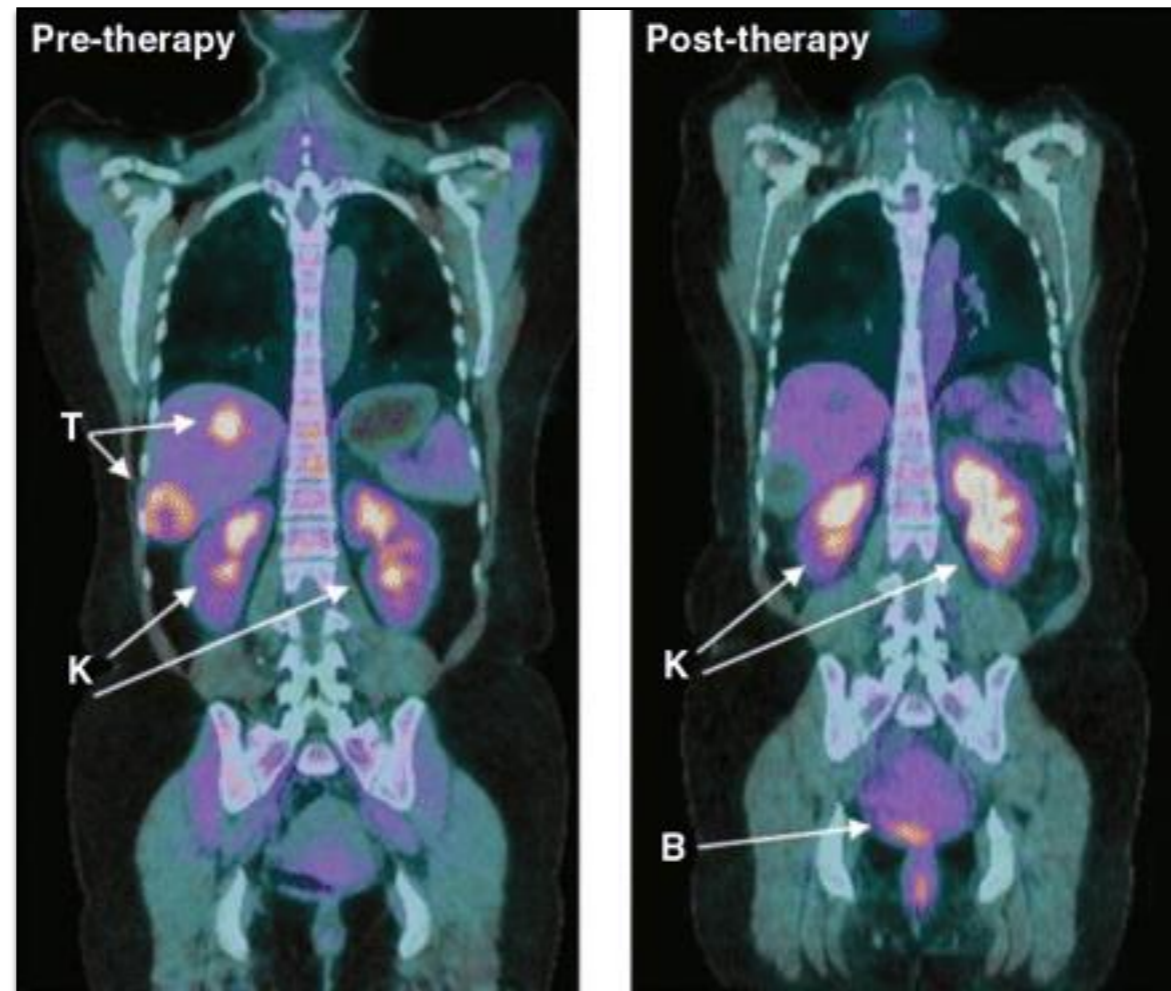
# Aerobic glycolysis



Cell produce large amount of lactate regardless of the availability of oxygen



# Detection of tumors by glucose analogs

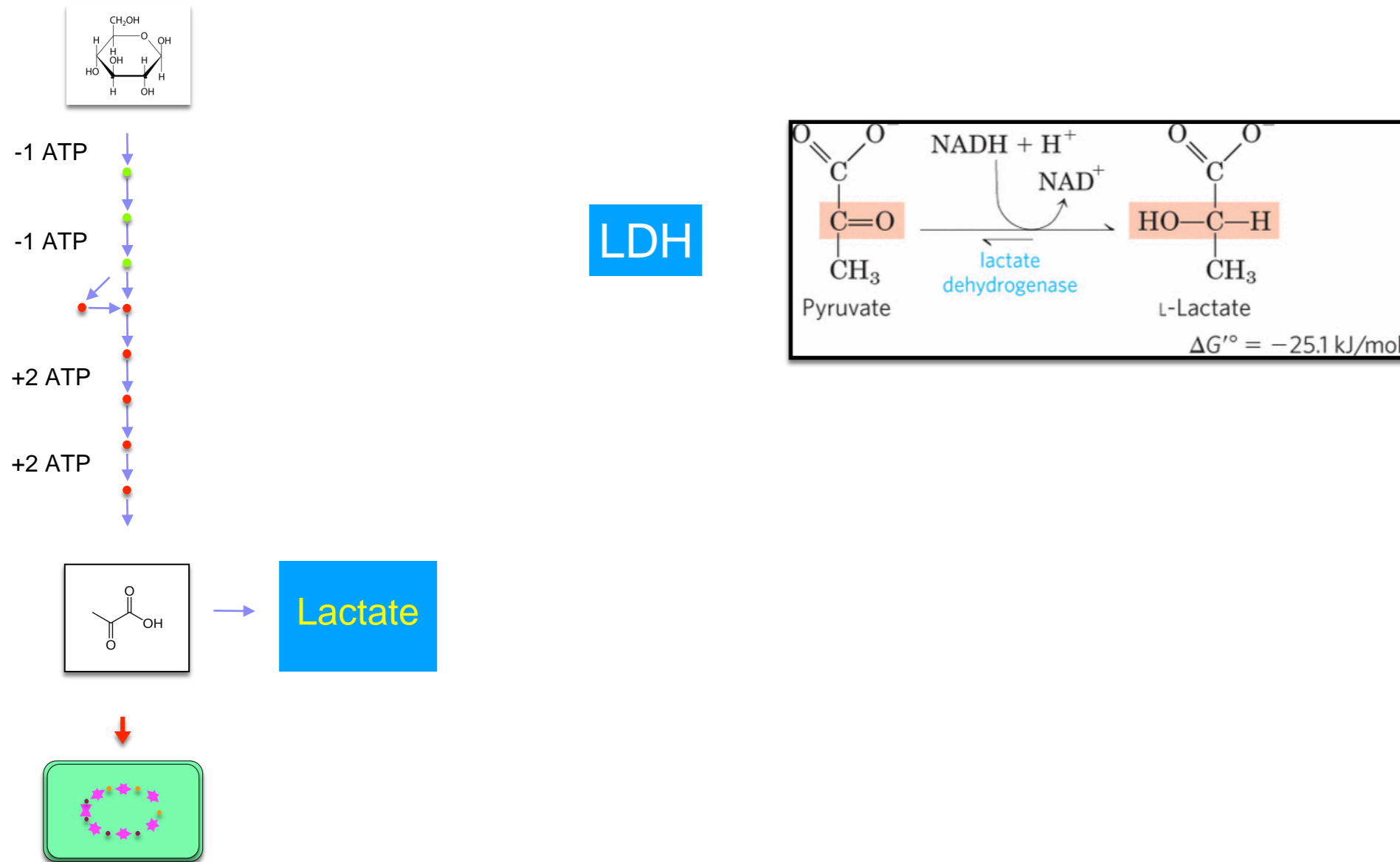


Decreased metabolism of glucose by tumors, visualised by PET with the glucose analog

FDG



# Suggested molecular mechanics for Warburg effect



LDHA (LDH5) functions as a prognosis marker in many tumors including: lymphoma, prostate cancer, renal cell carcinoma (RCC), and melanoma

# Course leading Questions

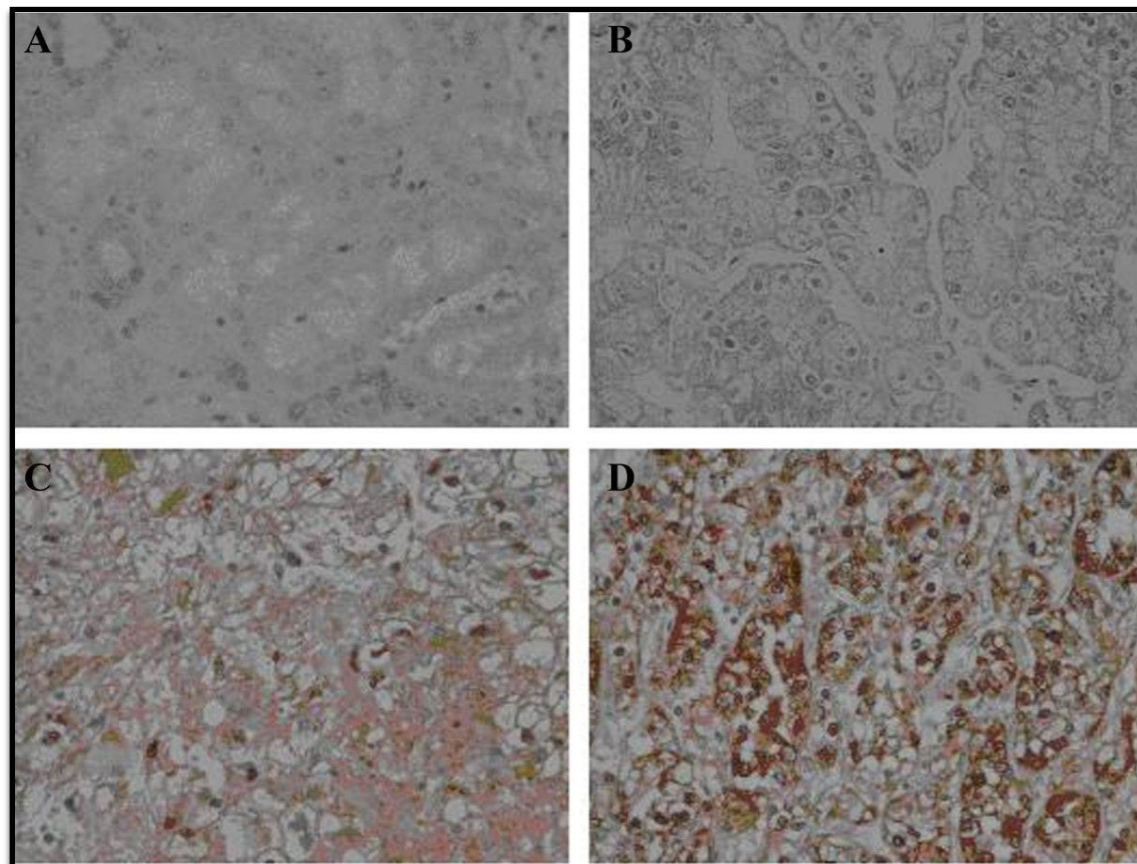
## Warburg Effect

What are the molecular mechanisms that regulate the WA?

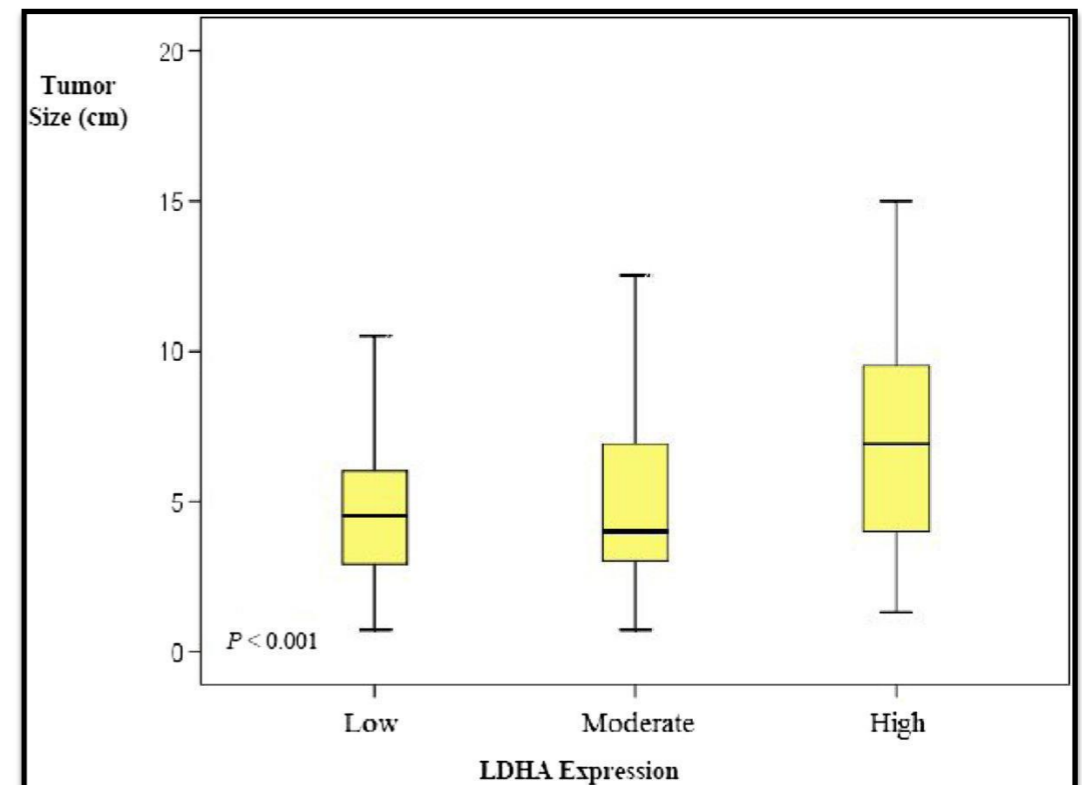
Is WA a byproduct or an integrated part of the cancer

# Lactate Dehydrogenase as a tumor marker

## LDHA protein expression by immunohistochemistry



## Box plot representing LDHA expression levels in relation to tumor size



(A) Normal kidney proximal tubular epithelium,  
(B) weak  
(C) moderate  
(D) strong staining Clear Cell Renal Cell Carcinoma

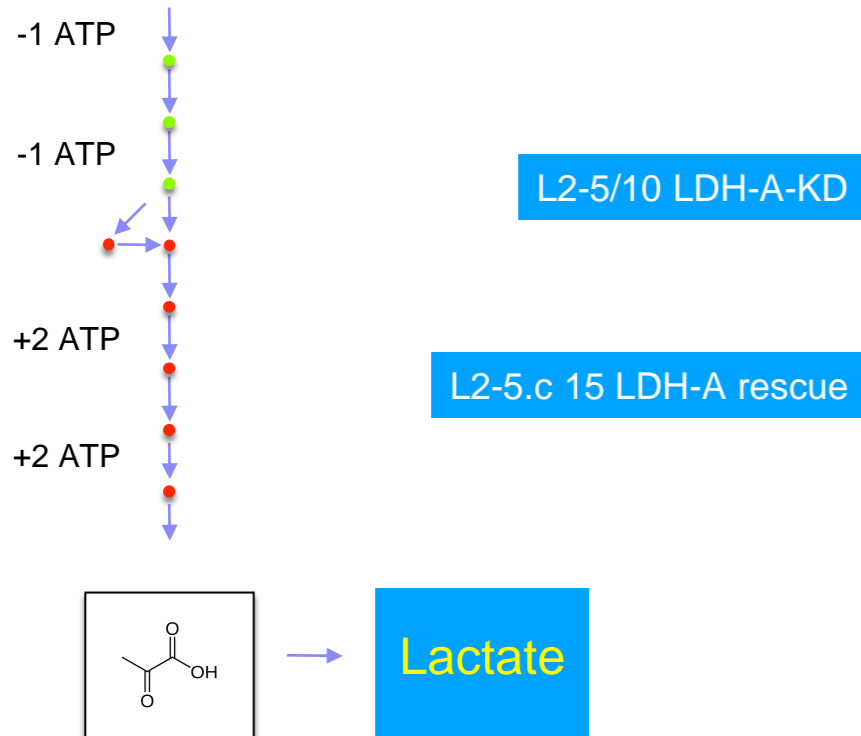
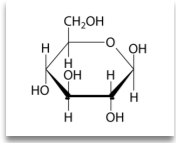
# Lactate Dehydrogenase KD

LDH-A suppression interferes with tumorigenicity of malignant cell

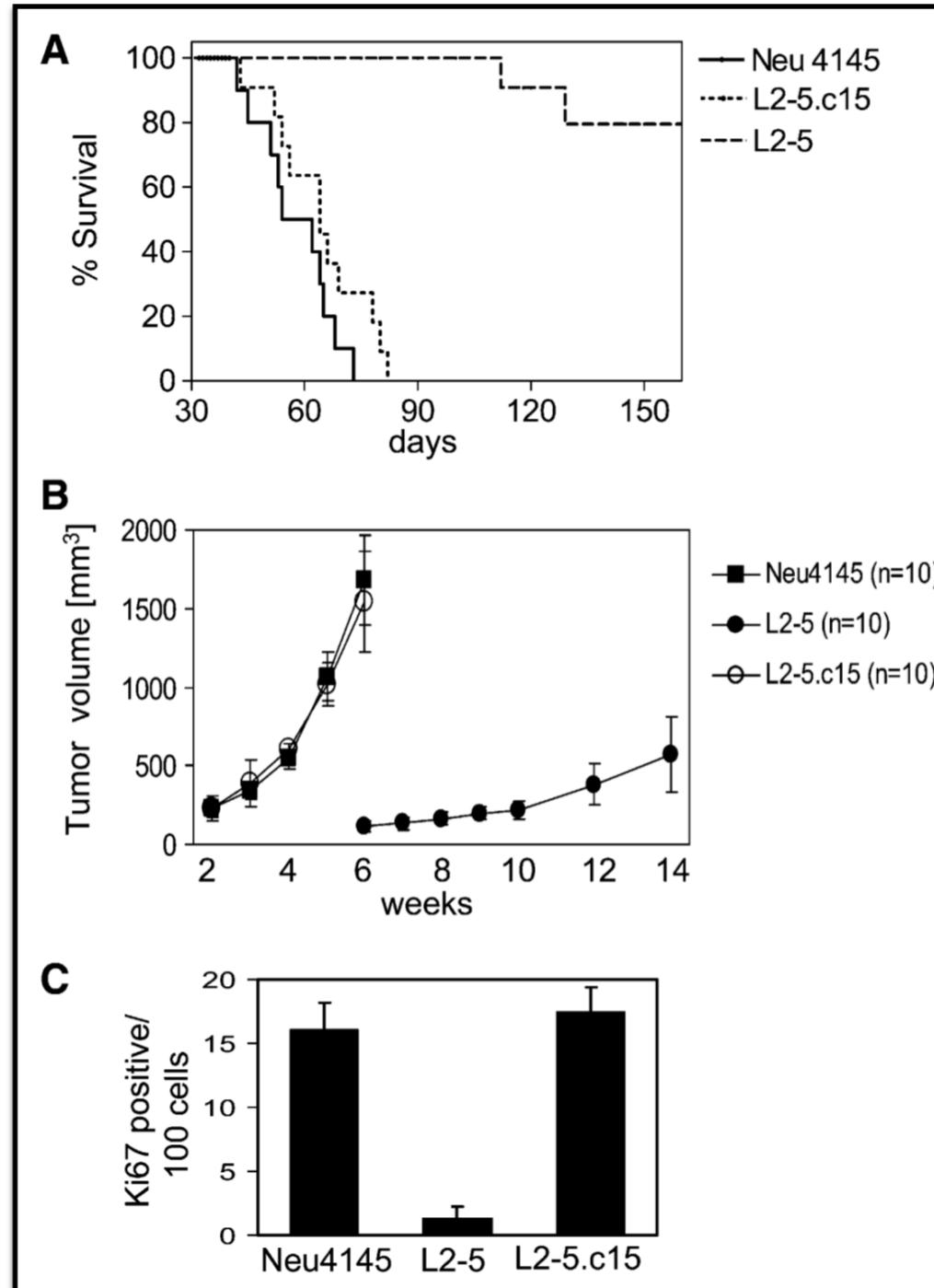
Attenuation of LDH-A expression uncovers a link between glycolysis, mitochondrial physiology, and tumor maintenance

Valeria R. Fantin,<sup>1,3,4</sup> Julie St-Pierre,<sup>2,3,5</sup> and Philip Leder<sup>1,\*</sup>

<sup>1</sup>Department of Genetics, Harvard Medical School and Howard Hughes Medical Institute, Boston, Massachusetts 02115  
<sup>2</sup>Dana-Farber Cancer Institute and Department of Cell Biology, Harvard Medical School, Boston, Massachusetts 02115  
<sup>3</sup>These authors contributed equally to this work.  
<sup>4</sup>Present address: Merck & Co., Inc., Boston, Massachusetts 02115.  
<sup>5</sup>Present address: Institut de Recherche en Immunologie et en Cancérologie (IRIC), Université de Montréal Pavillon Marcelle-Coutu, Montréal, Québec, Canada H3T 1J4.  
<sup>\*</sup>Correspondence: leder@genetics.med.harvard.edu

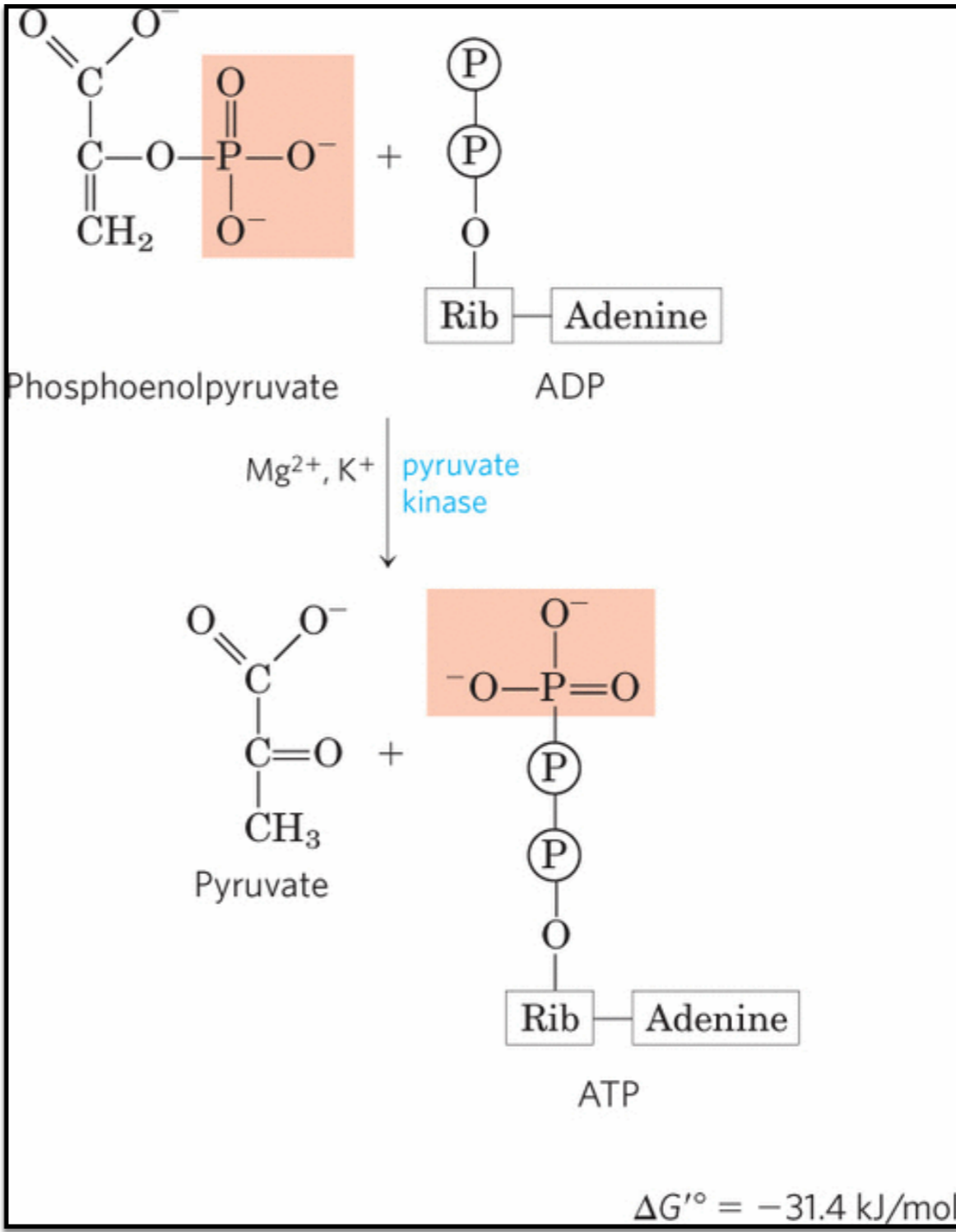
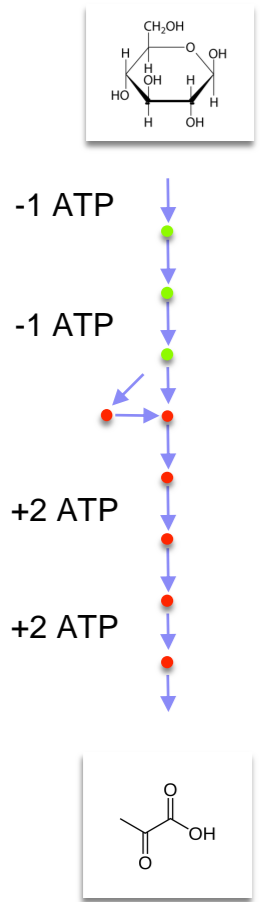


Ki67- mitotic marker



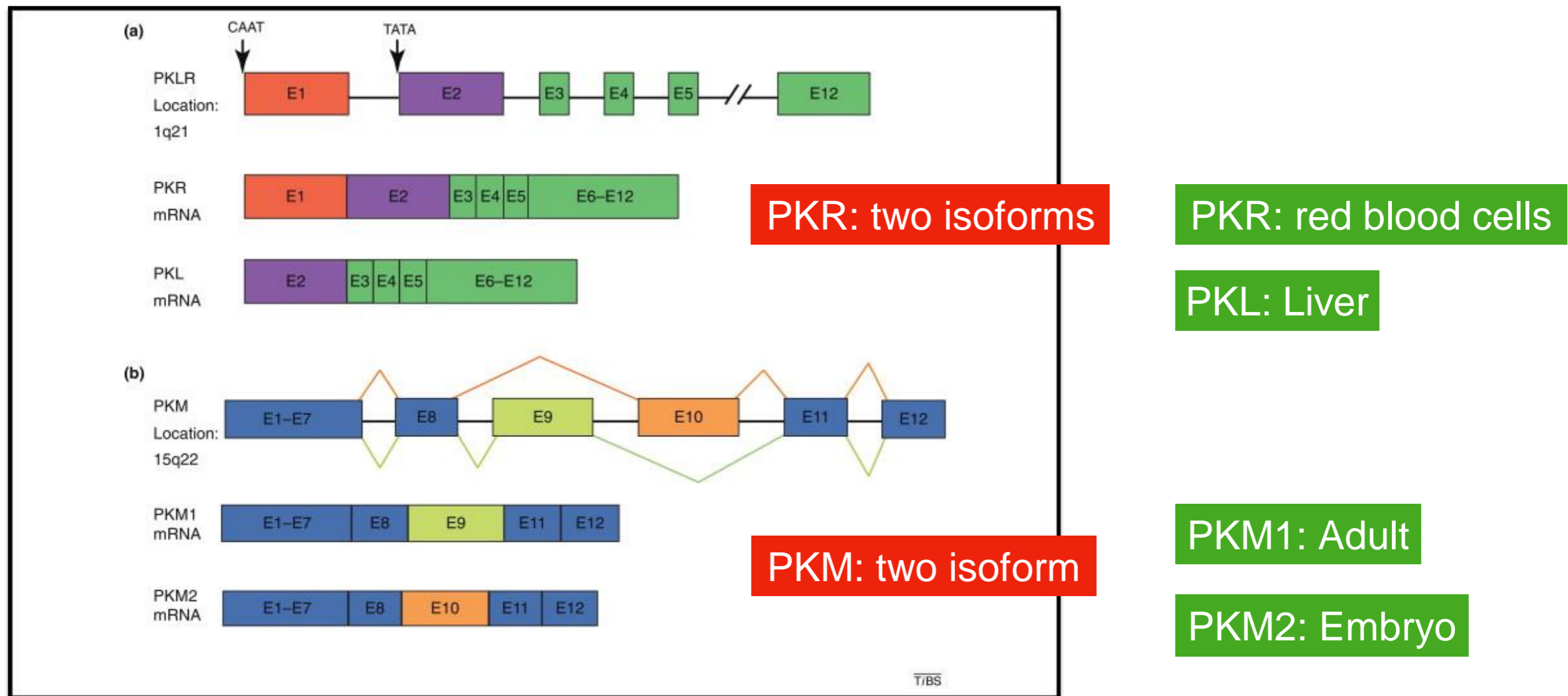
# Pyruvate kinase

## Pyruvate kinase



# Different isoforms of PK

There are four isoforms of Pyruvate kinase  
Encoded by two genes:

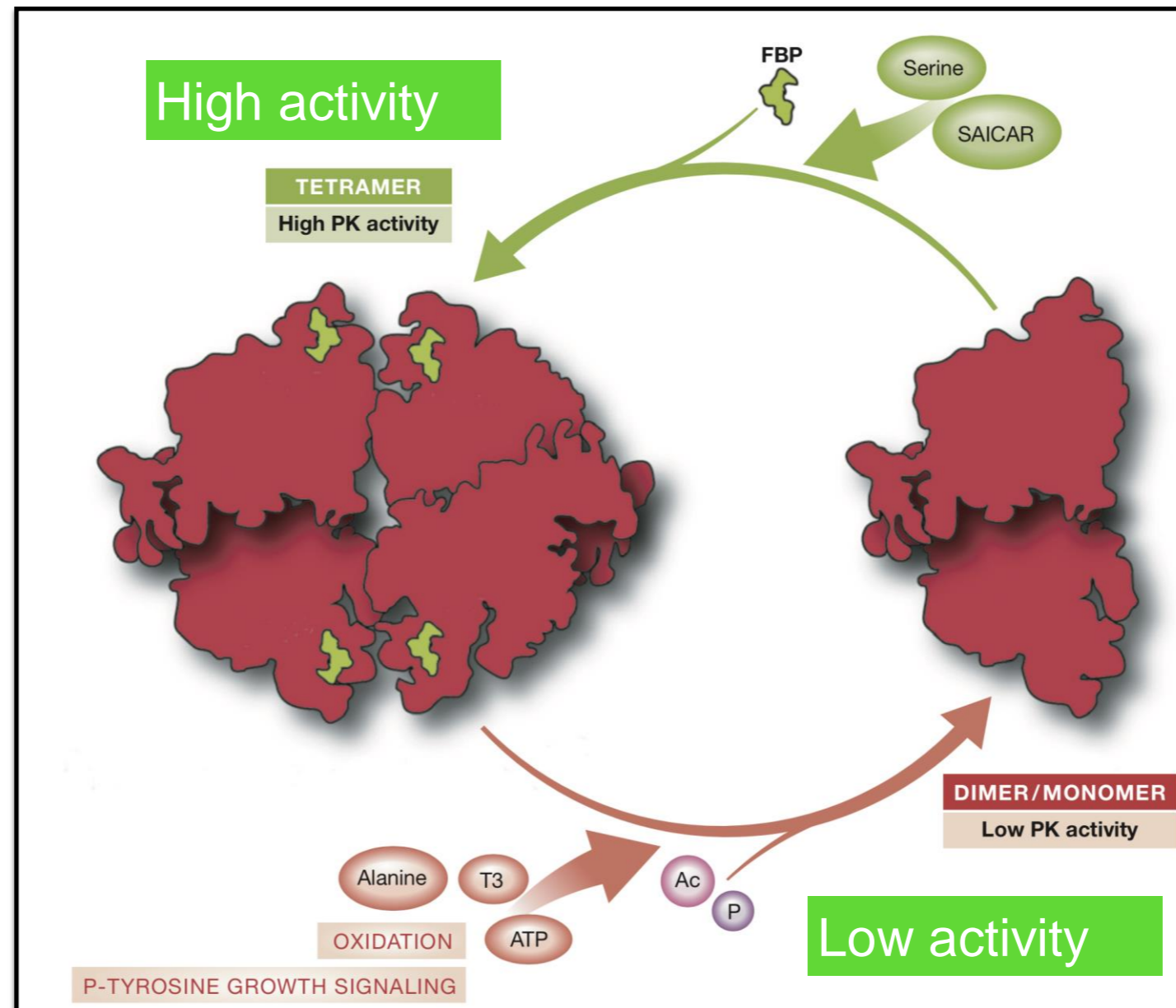


1. Chaneton, B., and Gottlieb, E. (2012) Rocking cell metabolism: revised functions of the key glycolytic regulator PKM2 in cancer. Trends Biochem Sci. 37, 309–316



# PKM2 activity is tightly regulated

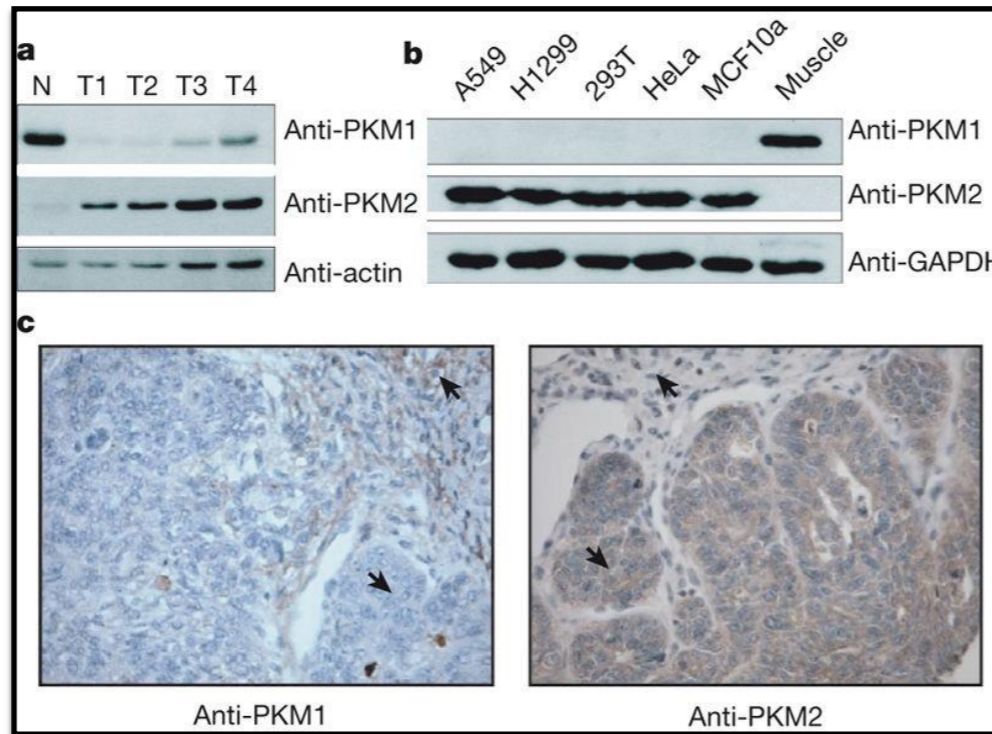
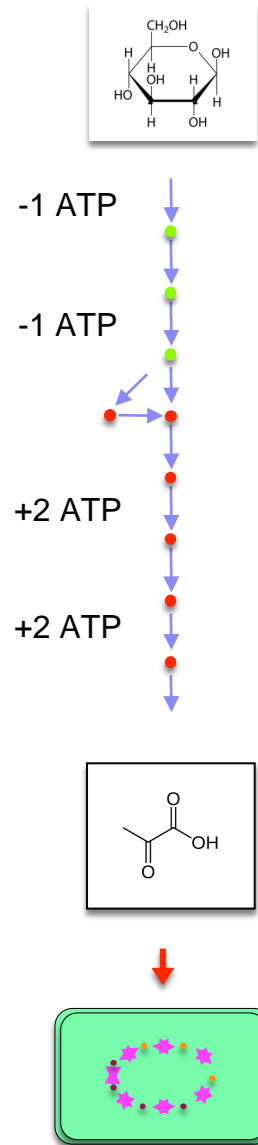
PKM2 enzymatic activity can be inhibited by a variety of mechanisms



Ability to inhibit PKM2 provides advantage to proliferating cells

# PKM

There are two isoforms of PKM2



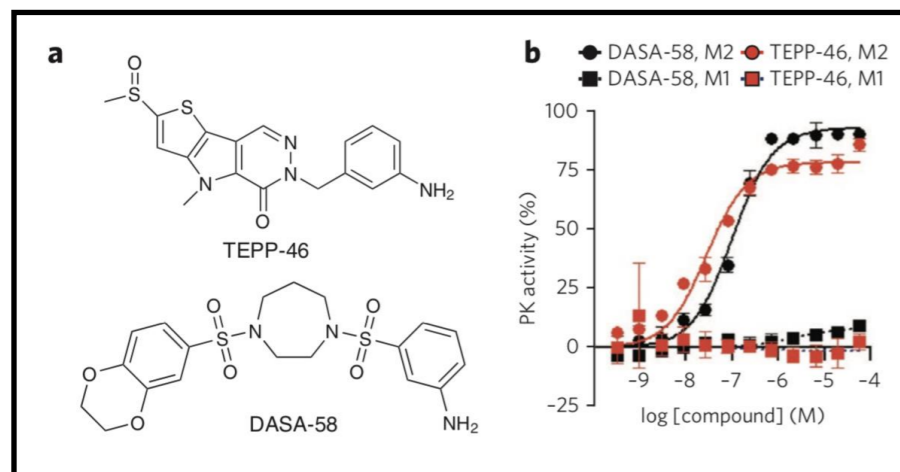
PKM1 expressed in normal tissues

PKM2 expressed in cancer

PKM1 is a more active enzyme than PKM2

# Activating PKM2 suppress tumorigenesis

## TEPP-46 activates PKM2



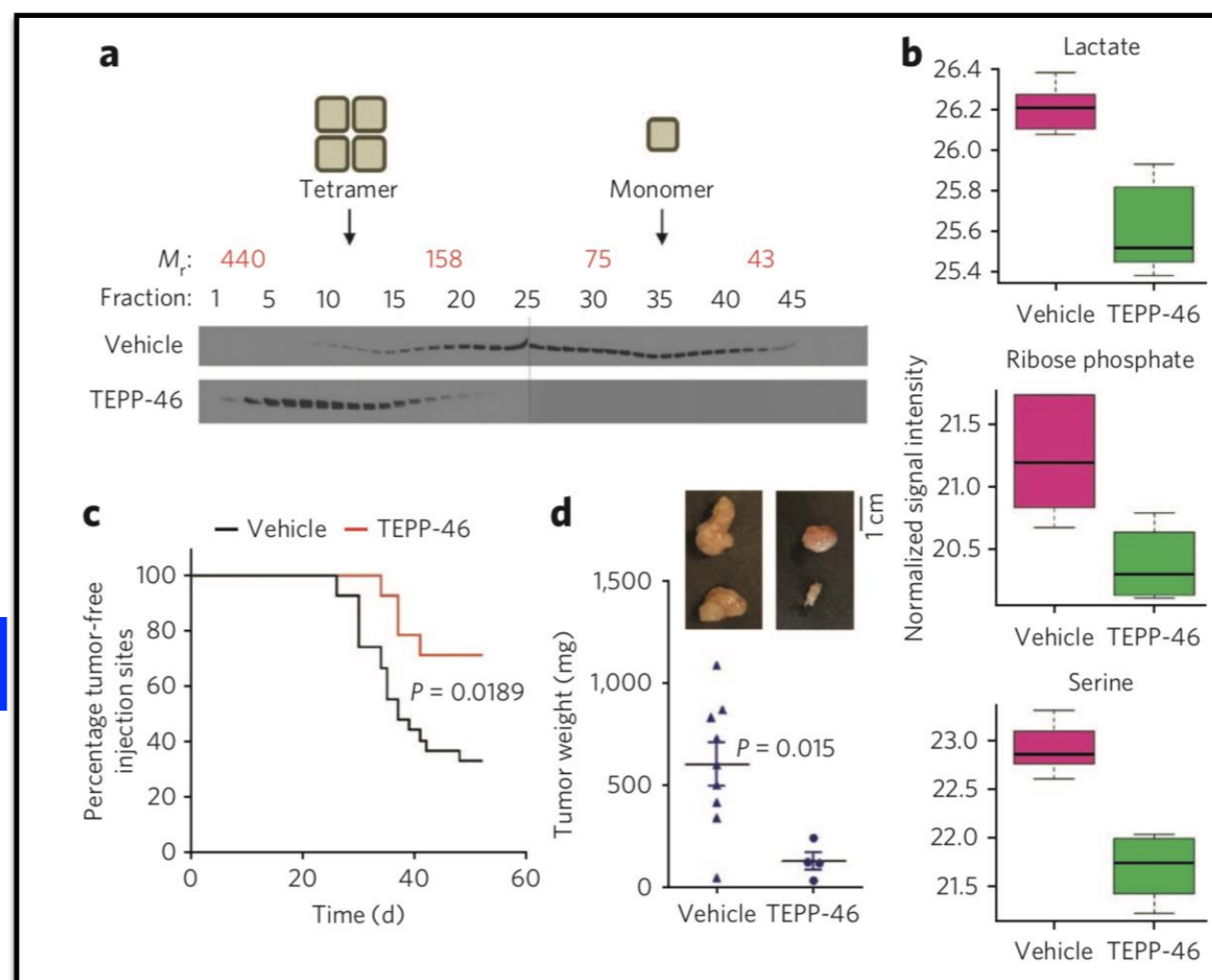
nature  
chemical biology

ARTICLE  
PUBLISHED ONLINE: 26 AUGUST 2012 | DOI: 10.1038/NCHEM.1060

**Pyruvate kinase M2 activators promote tetramer formation and suppress tumorigenesis**

Dimitrios Anastasiou<sup>1,2,3</sup>, Yimin Yu<sup>1,3</sup>, William J Israelsen<sup>1,3</sup>, Jian-Kang Jiang<sup>4</sup>, Matthew B Boxer<sup>4</sup>, Bum Soo Hong<sup>5</sup>, Wolfram Tempel<sup>5</sup>, Svetoslav Dimov<sup>5</sup>, Min Shen<sup>6</sup>, Abhishek Jha<sup>6</sup>, Hua Yang<sup>7</sup>, Katherine R Mattaini<sup>8</sup>, Christian M Metallo<sup>8</sup>, Brian P Fiske<sup>8</sup>, Kevin D Courtney<sup>1,2,9</sup>, Scott Malstrom<sup>9</sup>, Tahsin M Khan<sup>10</sup>, Charles Kung<sup>11</sup>, Amanda P Skoumbourdis<sup>12</sup>, Henrike Veith<sup>13</sup>, Noel Southall<sup>14</sup>, Martin J Walsh<sup>15</sup>, Kyle R Brimacombe<sup>16</sup>, William Leister<sup>17</sup>, Sophia Y Lunt<sup>18</sup>, Zachary R Johnson<sup>19</sup>, Katharine E Yen<sup>20</sup>, Kaiko Kunii<sup>21</sup>, Shawn M Davidson<sup>22</sup>, Heather R Christofk<sup>23</sup>, Christopher P Austin<sup>24</sup>, James Inglese<sup>25</sup>, Marian H Harris<sup>26</sup>, John M Asara<sup>10</sup>, Gregory Stephanopoulos<sup>27</sup>, Francesco G Salituro<sup>28</sup>, Shengfang Jin<sup>29</sup>, Lenny Dang<sup>30</sup>, Douglas S Auld<sup>31</sup>, Hee-Won Park<sup>32</sup>, Lewis C Cantley<sup>1,2</sup>, Craig J Thomas<sup>33</sup> & Matthew G Vander Heiden<sup>1,2,3\*</sup>

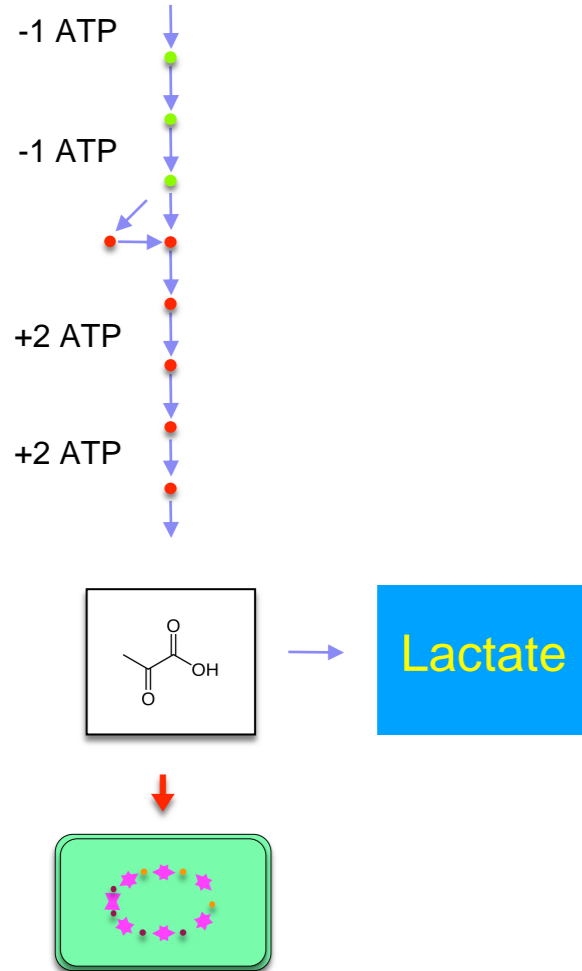
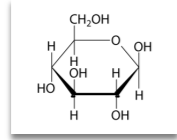
## TEPP-46 affects glycolysis intermediates



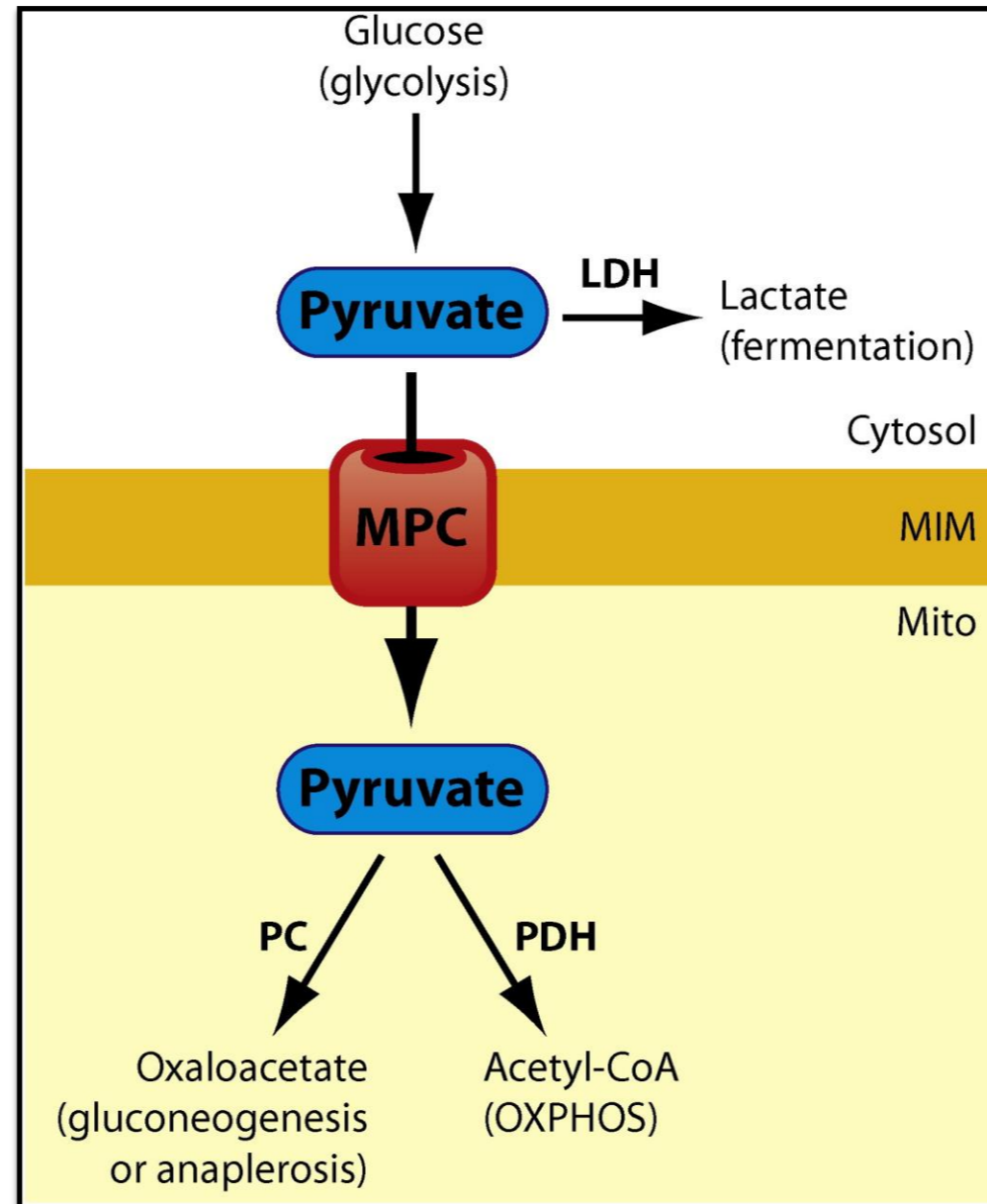
## TEPP-46 decrease tumor growth

Anastasiou, D., Yu, Y., Israelsen, W. J., Jiang, J.-K., Boxer, M. B., Hong, B. S., Tempel, W., Dimov, S., Shen, M., Jha, A., Yang, H., Mattaini, K. R., Metallo, C. M., Fiske, B. P., Courtney, K. D., Malstrom, S., Khan, T. M., Kung, C., Skoumbourdis, A. P., Veith, H., Southall, N., Walsh, M. J., Brimacombe, K. R., Leister, W., Lunt, S. Y., Johnson, Z. R., Yen, K. E., Kunii, K., Davidson, S. M., Christofk, H. R., Austin, C. P., Inglese, J., Harris, M. H., Asara, J. M., Stephanopoulos, G., Salituro, F. G., Jin, S., Dang, L., Auld, D. S., Park, H.-W., Cantley, L. C., Thomas, C. J., and Vander Heiden, M. G. (2012) Pyruvate kinase M2 activators promote tetramer formation and suppress tumorigenesis. *Nat Chem Biol.* 8, 839–847

# Mitochondrial Pyruvate Carrier (MPC)



## Gatekeeper of pyruvate entry to the mitochondria



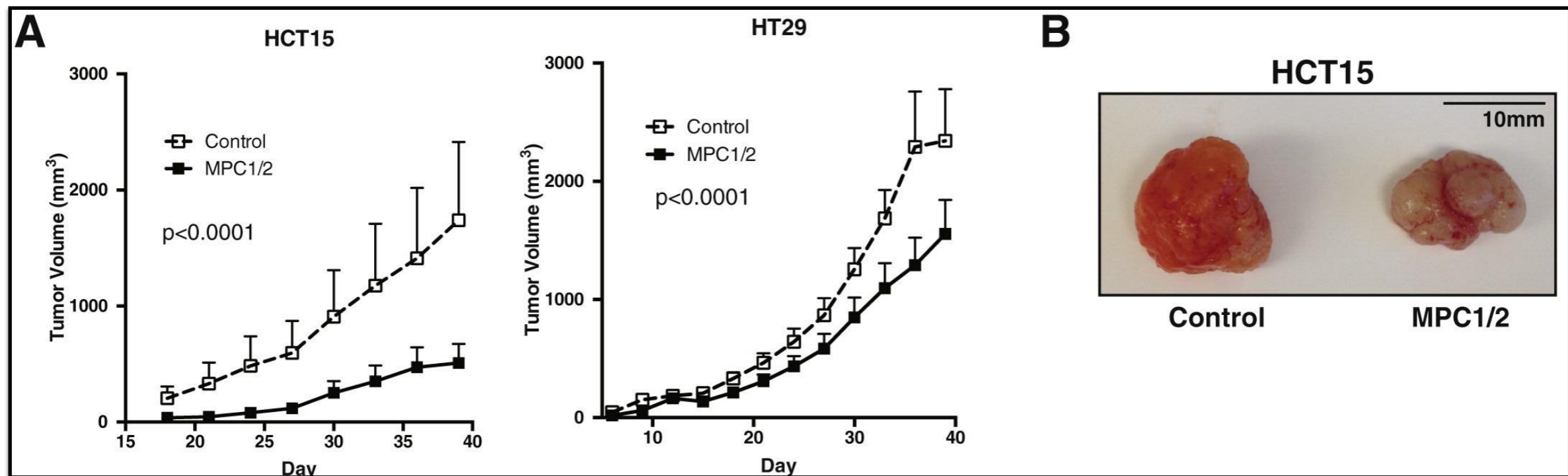
Two inform MPC1 and 2

# MPC1/2 re-expression decreases tumor size

## A Role for the Mitochondrial Pyruvate Carrier as a Repressor of the Warburg Effect and Colon Cancer Cell Growth

John C. Schell,<sup>1,4</sup> Kristofer A. Olson,<sup>1,4</sup> Lei Jiang,<sup>2</sup> Amy J. Hawkins,<sup>1</sup> Jonathan G. Van Vranken,<sup>1</sup> Jianxin Xie,<sup>3</sup> Robert A. Egnatchik,<sup>2</sup> Espen G. Earl,<sup>1</sup> Ralph J. DeBerardinis,<sup>2</sup> and Jared Rutter<sup>1,\*</sup>  
<sup>1</sup>Department of Biochemistry, University of Utah School of Medicine, Salt Lake City, UT 84112-5650, USA  
<sup>2</sup>Children's Medical Center Research Institute, UT Southwestern Medical Center, Dallas, TX 75390-8502, USA  
<sup>3</sup>Cell Signaling Technology, Inc., Danvers, MA 01923, USA  
<sup>4</sup>Co-first Authors  
\*Correspondence: rutter@biochem.utah.edu  
<http://dx.doi.org/10.1016/j.molcel.2014.09.026>

MPC1/2 slowed tumor growth



Schell, J. C., Olson, K. A., Jiang, L., Hawkins, A. J., Van Vranken, J. G., Xie, J., Egnatchik, R. A., Earl, E. G., Deberardinis, R. J., and Rutter, J. (2014) A Role for the Mitochondrial Pyruvate Carrier as a Repressor of the Warburg Effect and Colon Cancer Cell Growth. *Mol Cell*. 56, 400–413



# Leading Questions

## Metabolism and cancer

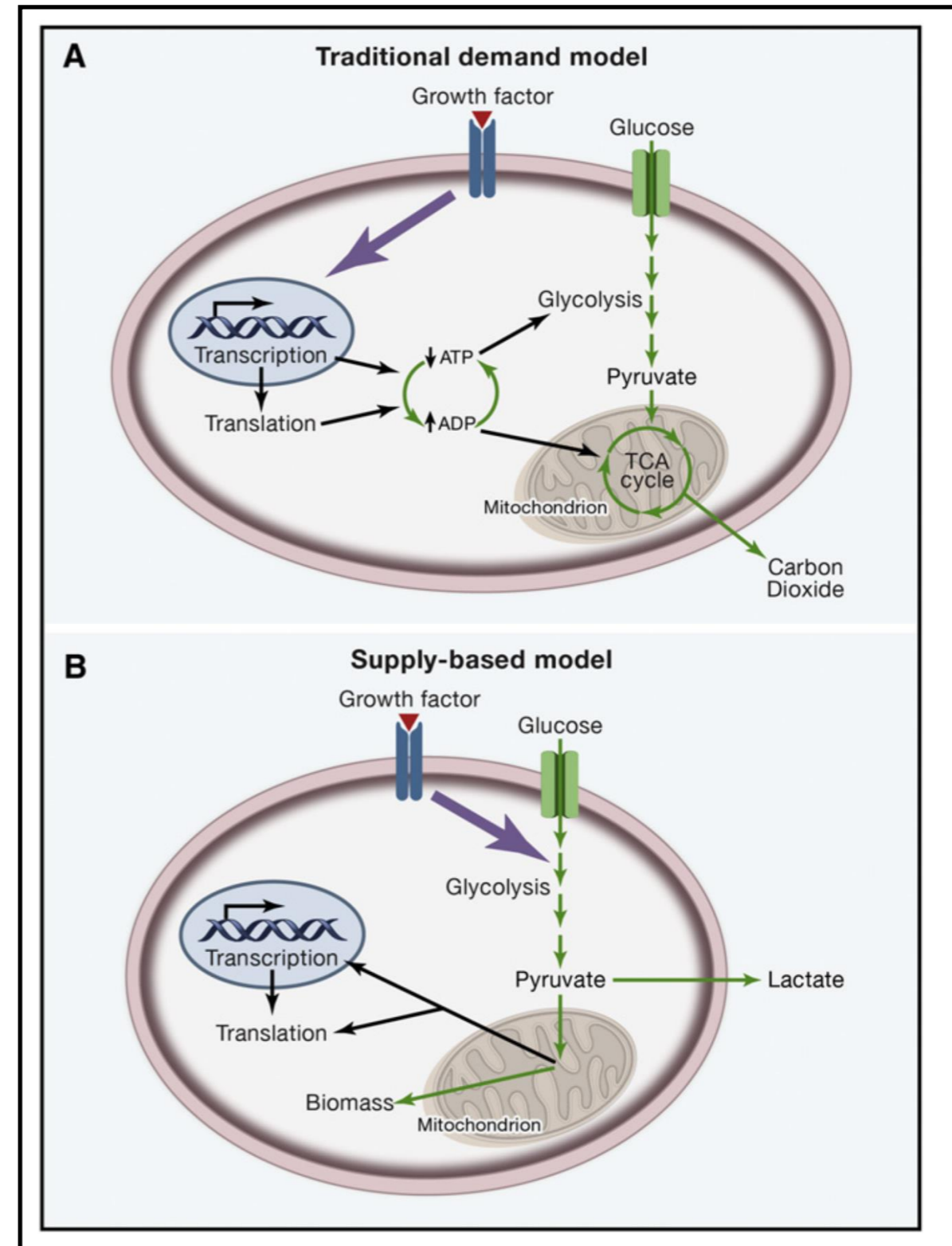
Can metabolism be the drivers of cancers?

### Traditional model

Metabolism is altered in response to growth factor

### Supply-based model

Growth factor signaling directly reprograms nutrient uptake and metabolism





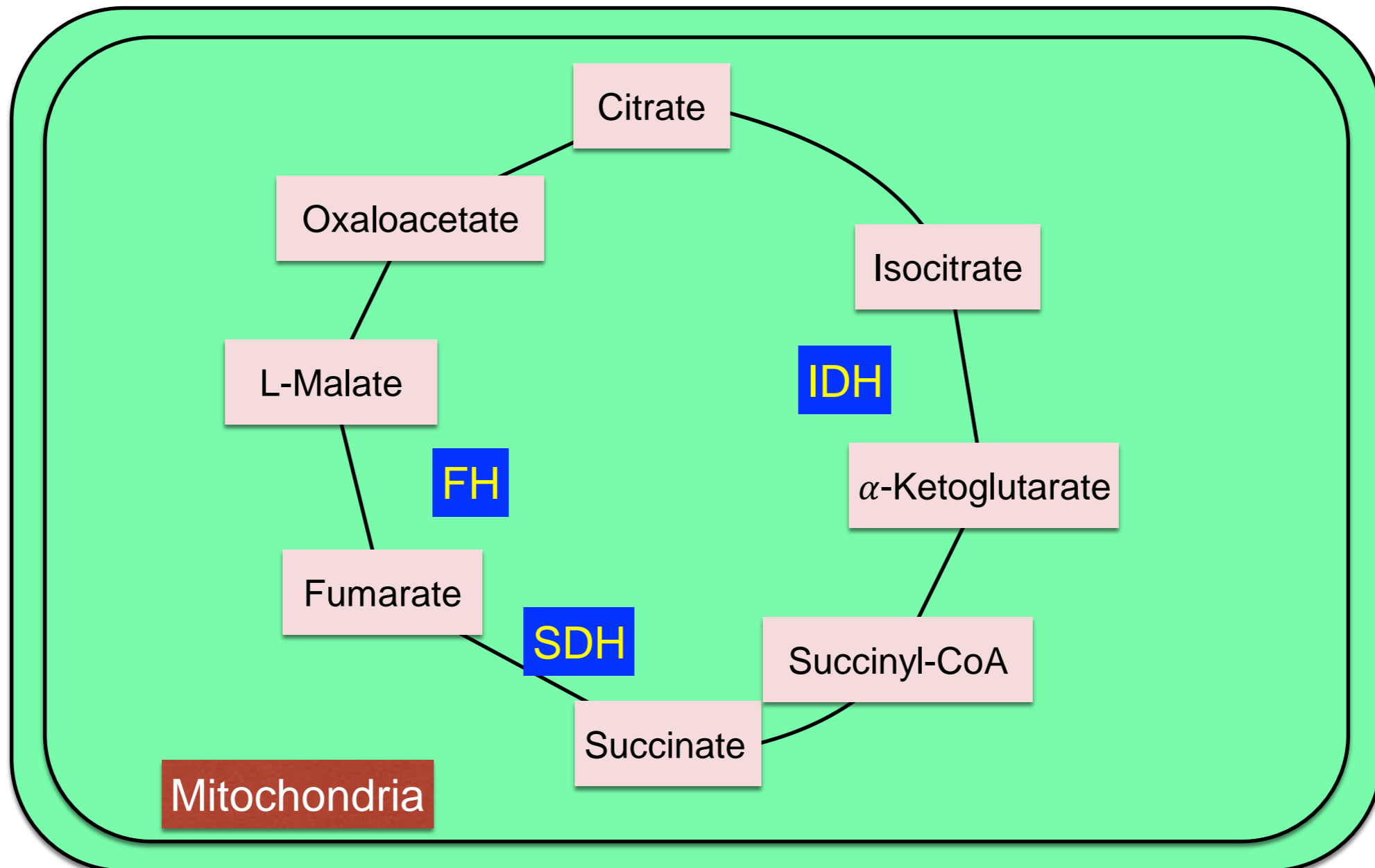
# Human tumor suppressor genes that have been cloned

## Evading growth suppressors

Metabolic gene

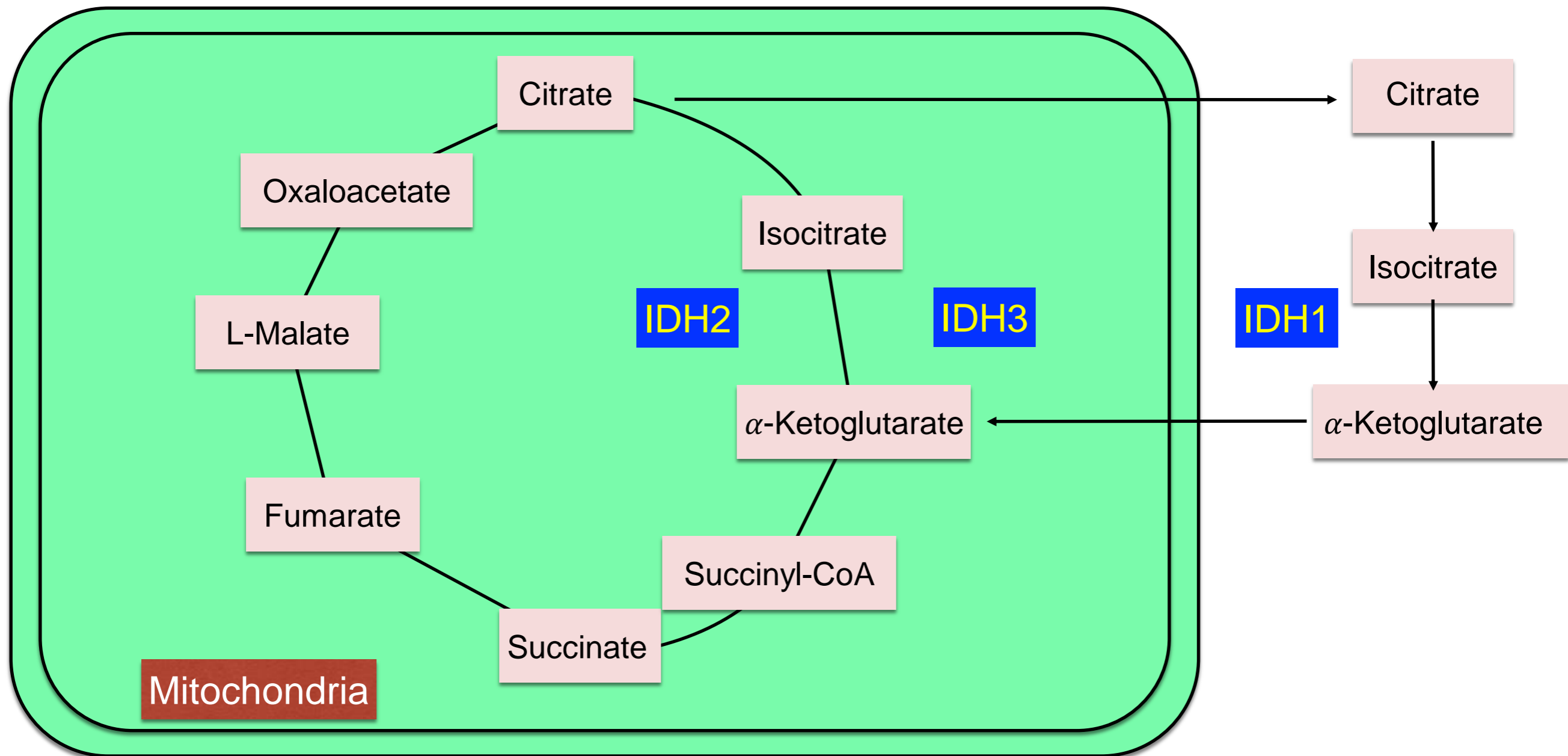
Name of gene	Chromosomal location	Familial cancer syndrome	Sporadic cancer	Function of protein
<i>RUNX3</i>	1p36	—	gastric carcinoma	TF co-factor
<i>HRPT2</i>	1q25–32	parathyroid tumors, jaw fibromas	parathyroid tumors	chromatin protein
<i>FH</i>	1q42.3	familial leiomyomatosis <sup>a</sup>	—	fumarate hydratase
<i>FHIT</i>	3p14.2	—	many types	diadenosine triphosphate hydrolase
<i>RASSF1A</i>	3p21.3	—	many types	multiple functions
<i>TGFBR2</i>	3p2.2	HNPCC	colon, gastric, pancreatic carcinomas	TGF-β receptor
<i>VHL</i>	3p25	von Hippel–Lindau syndrome	renal cell carcinoma	ubiquitylation of HIF
<i>hCDC4</i>	4q32	—	endometrial carcinoma	ubiquitin ligase
<i>APC</i>	5p21	familial adenomatous polyposis coli	colorectal, pancreatic, and stomach carcinomas; prostate carcinoma	β-catenin degradation
<i>NKX3.1</i>	8p21	—	prostate carcinoma	homeobox TF
<i>p16<sup>INK4A</sup> b</i>	9p21	familial melanoma	many types	CDK inhibitor
<i>p14<sup>ARF</sup> c</i>	9p21	—	all types	p53 stabilizer
<i>PTC</i>	9q22.3	nevroid basal cell carcinoma syndrome	medulloblastomas	receptor for hedgehog GF
<i>TSC1</i>	9q34	tuberous sclerosis	—	inhibitor of mTOR <sup>f</sup>
<i>BMPR1</i>	10q21–22	juvenile polyposis	—	BMP receptor
<i>PTEN<sup>d</sup></i>	10q23.3	Cowden's disease, breast and gastrointestinal carcinomas	glioblastoma; prostate, breast, and thyroid carcinomas	PIP <sub>3</sub> phosphatase
<i>WT1</i>	11p13	Wilms tumor	Wilms tumor	TF
<i>MEN1</i>	11p13	multiple endocrine neoplasia	—	histone modification, transcriptional repressor
<i>BWS/CDKN1C</i>	11p15.5	Beckwith–Wiedemann syndrome	—	p57 <sup>Kip2</sup> CDK inhibitor
<i>SDHD</i>	11q23	familial paraganglioma	pheochromocytoma	mitochondrial protein <sup>e</sup>
<i>RB</i>	13q14	retinoblastoma, osteosarcoma	retinoblastoma; sarcomas; bladder, breast, esophageal, and lung carcinomas	transcriptional repression; control of E2Fs
<i>TSC2</i>	16p13	tuberous sclerosis	—	inhibitor of mTOR <sup>f</sup>
<i>CBP</i>	16p13.3	Rubinstein–Taybi	AML <sup>g</sup>	TF co-activator
<i>CYLD</i>	16q12–13	cylindromatosis	—	deubiquitinating enzyme
<i>CDH1</i>	16q22.1	familial gastric carcinoma	invasive cancers	cell–cell adhesion
<i>BHD</i>	17p11.2	Birt–Hogg–Dube syndrome	kidney carcinomas, hamartomas	unknown
<i>TP53</i>	17p13.1	Li–Fraumeni syndrome	many types	TF
<i>NF1</i>	17q11.2	neurofibromatosis type 1	colon carcinoma, astrocytoma	Ras-GAP
<i>BECN1</i>	17q21.3	—	breast, ovarian, prostate	autophagy
<i>PRKAR1A</i>	17.q22–24	multiple endocrine neoplasia <sup>h</sup>	multiple endocrine tumors	subunit of PKA
<i>DPC4<sup>i</sup></i>	18q21.1	juvenile polyposis	pancreatic and colon carcinomas	TGF-β TF
<i>LKB1/STK11</i>	19p13.3	Peutz–Jegher syndrome	hamartomatous colonic polyps	serine/threonine kinase
<i>RUNX1</i>	21q22.12	familial platelet disorder	AML	TF
<i>SNF5<sup>j</sup></i>	22q11.2	rhabdoid predisposition syndrome	malignant rhabdoid tumors	chromosome remodeling
<i>NF2</i>	22q12.2	neurofibroma-position syndrome	schwannoma, meningioma; ependymoma	cytoskeleton–membrane linkage

# Mutated metabolic enzymes in cancer



# Isocitrate dehydrogenase (IDH)

Exists as three isoform:



# IDH mutation in cancer

IDH1 is mutated in 80% of grade II–III gliomas

Vol 462 | 10 December 2009 | doi:10.1038/nature08617 nature

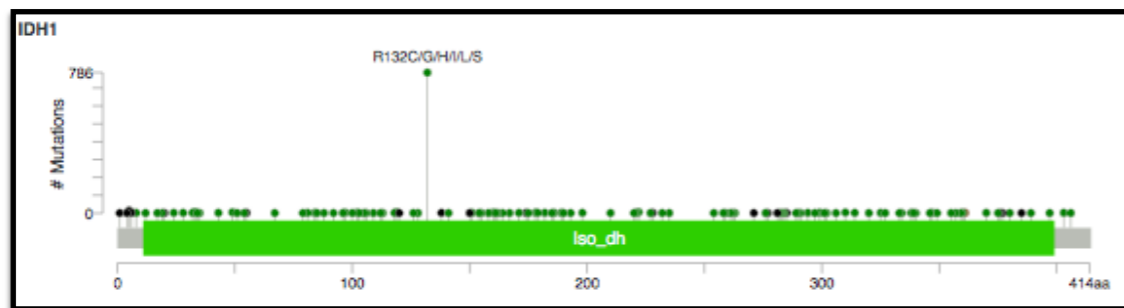
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ARTICLES

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## Cancer-associated IDH1 mutations produce 2-hydroxyglutarate

Lenny Dang<sup>1</sup>, David W. White<sup>1</sup>, Stefan Gross<sup>1</sup>, Bryson D. Bennett<sup>2</sup>, Mark A. Bittinger<sup>1</sup>, Edward M. Driggers<sup>1</sup>, Valeria R. Fantin<sup>1</sup>, Hyun Gyung Jang<sup>1</sup>, Shengfang Jin<sup>1</sup>, Marie C. Keenan<sup>1</sup>, Kevin M. Marks<sup>1</sup>, Robert M. Prins<sup>3</sup>, Patrick S. Ward<sup>4</sup>, Katharine E. Yen<sup>1</sup>, Linda M. Liao<sup>3</sup>, Joshua D. Rabinowitz<sup>2</sup>, Lewis C. Cantley<sup>5</sup>, Craig B. Thompson<sup>4</sup>, Matthew G. Vander Heiden<sup>1†</sup> & Shinsan M. Su<sup>1</sup>



IDH mutation are selected at early stages of tumorigenesis

What does this mutation do to the enzymatic activity?



# IDH1 mutations produce 2-hydroxyglutarate

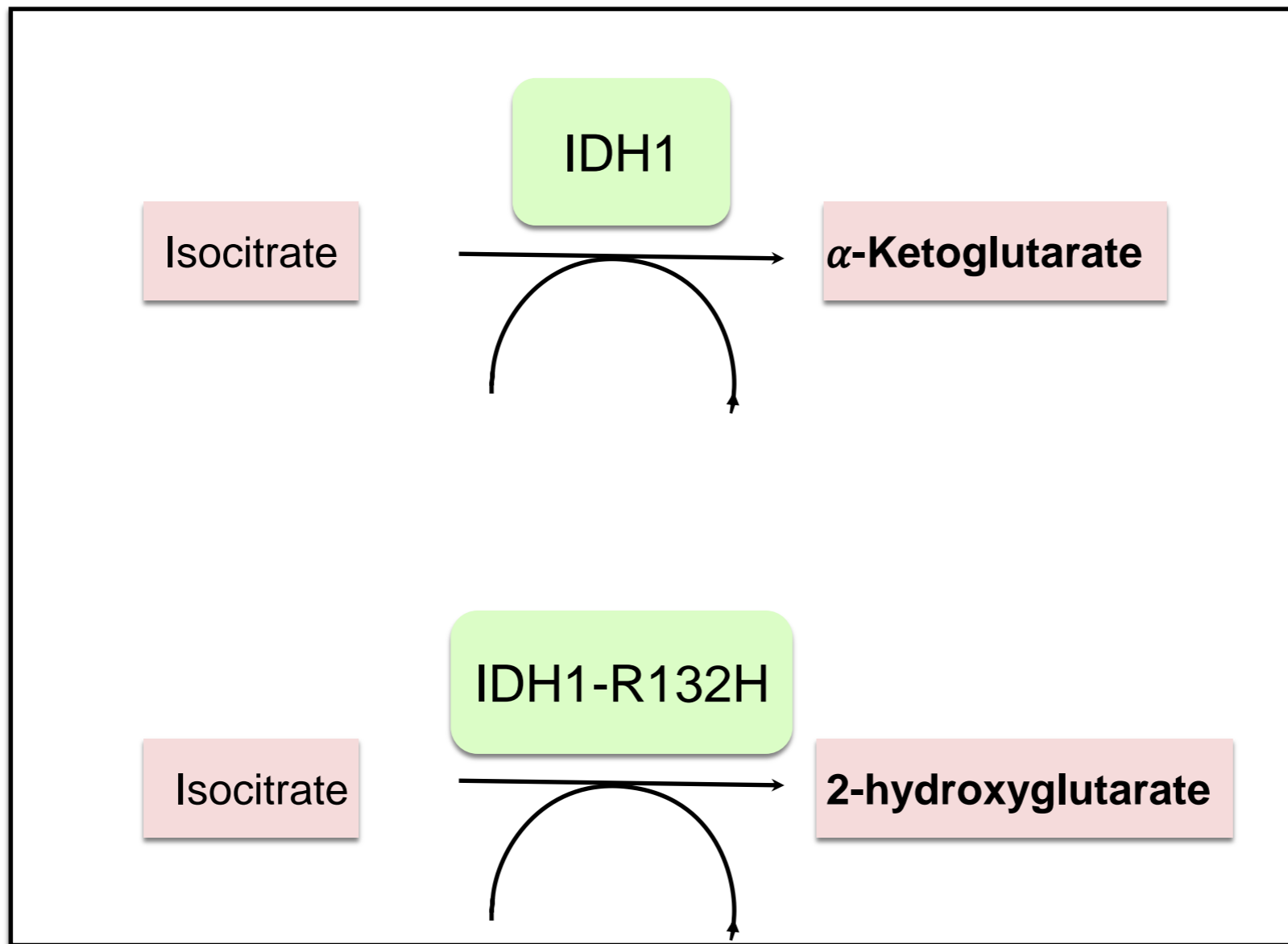
Vol 462 | 10 December 2009 | doi:10.1038/nature08617 nature

ARTICLES

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**Cancer-associated IDH1 mutations produce 2-hydroxyglutarate**

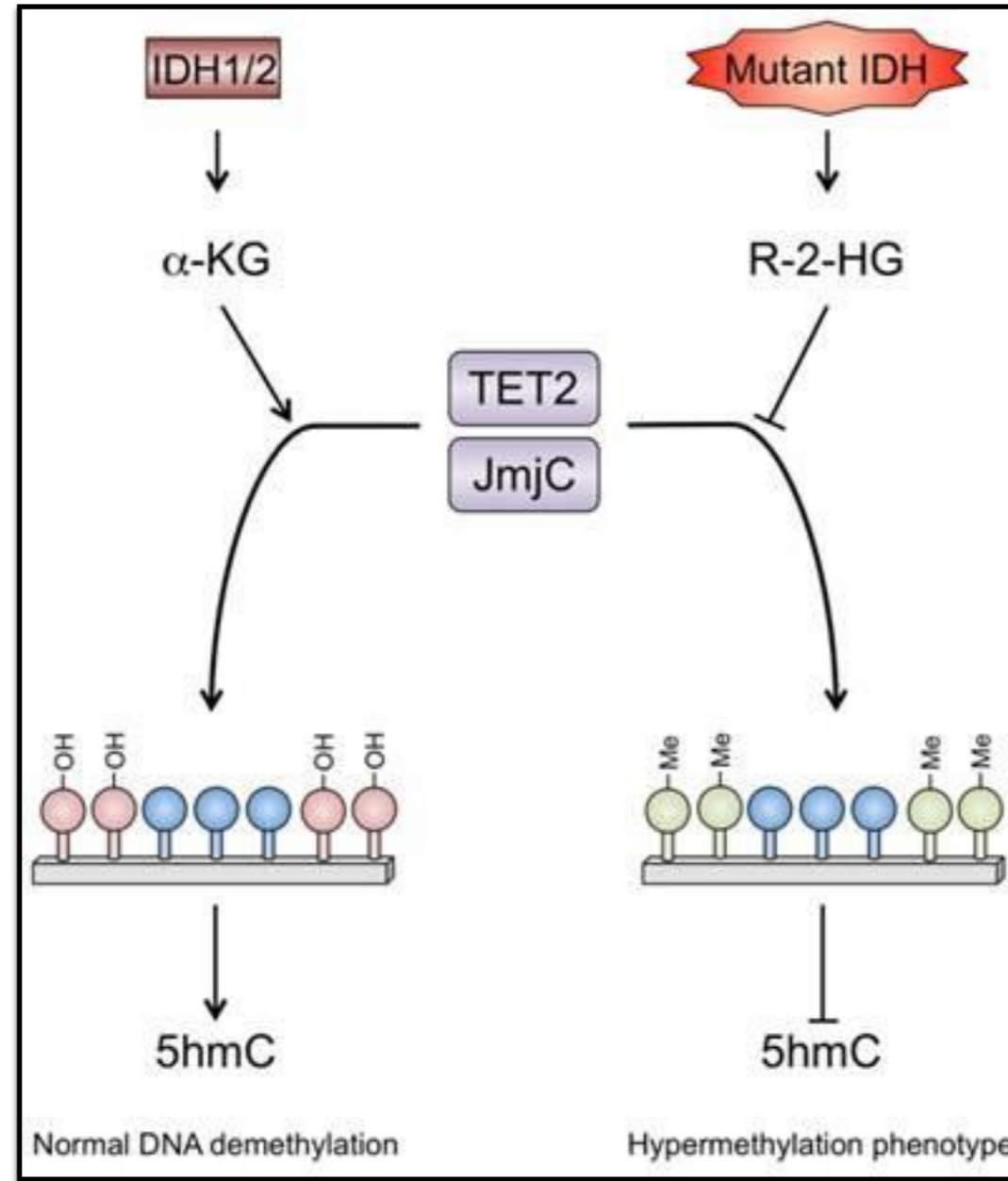
Lenny Dang<sup>1</sup>, David W. White<sup>1</sup>, Stefan Gross<sup>1</sup>, Bryson D. Bennett<sup>2</sup>, Mark A. Bittinger<sup>1</sup>, Edward M. Driggers<sup>1</sup>, Valeria R. Fantin<sup>1</sup>, Hyun Gyung Jang<sup>1</sup>, Shengfang Jin<sup>1</sup>, Marie C. Keenan<sup>1</sup>, Kevin M. Marks<sup>1</sup>, Robert M. Prins<sup>3</sup>, Patrick S. Ward<sup>1</sup>, Katharine E. Yen<sup>1</sup>, Linda M. Liaw<sup>3</sup>, Joshua D. Rabinowitz<sup>2</sup>, Lewis C. Cantley<sup>5</sup>, Craig B. Thompson<sup>4</sup>, Matthew G. Vander Heiden<sup>1†</sup> & Shinsan M. Su<sup>1</sup>



How do 2HG affect the cells?

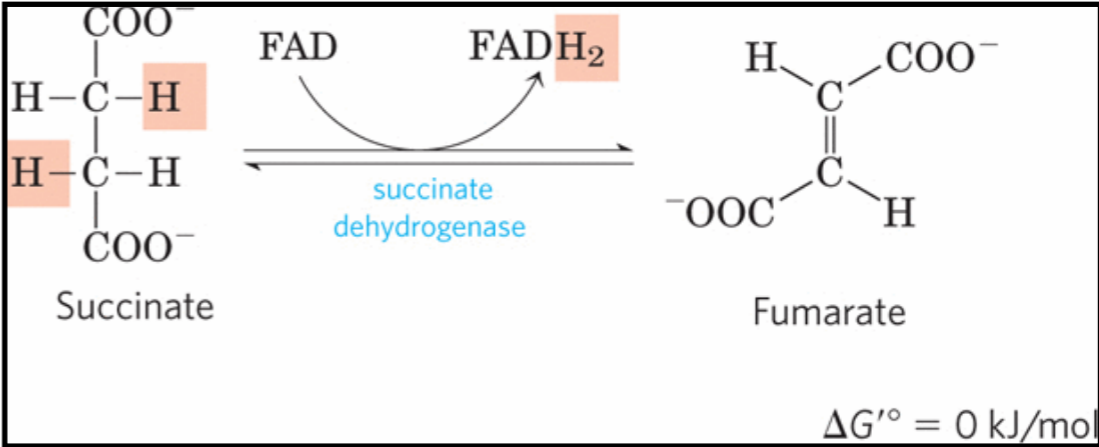
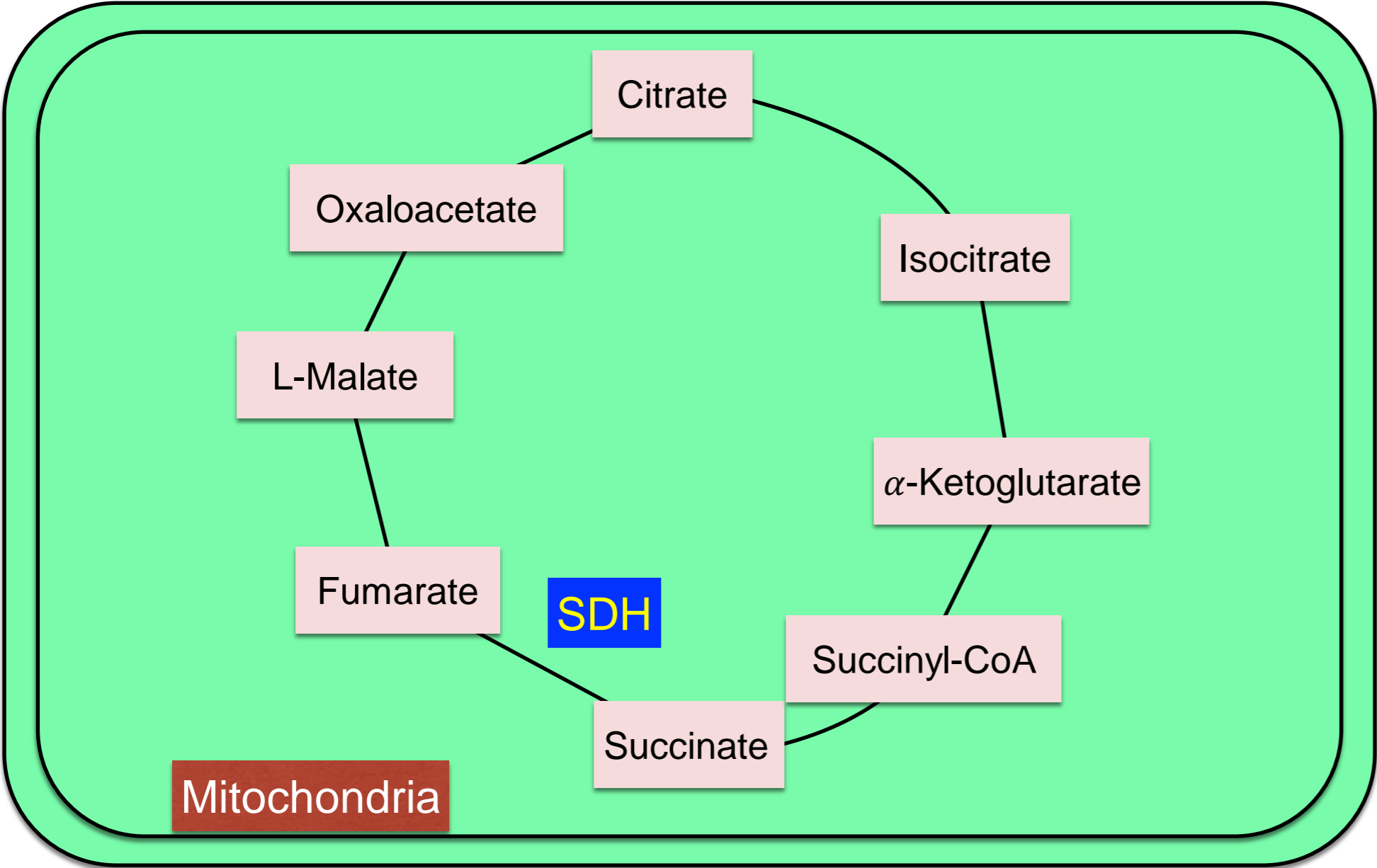
# 2-hydroxyglutarate

IDH movie





# Succinate Dehydrogenase



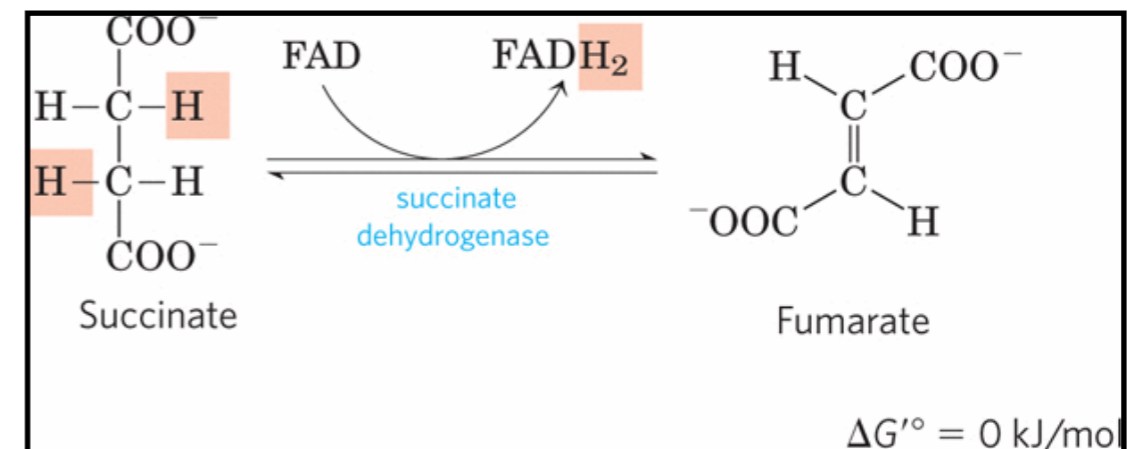
# SDH mutation

SDH was the first mitochondrial enzyme found mutated in cancer

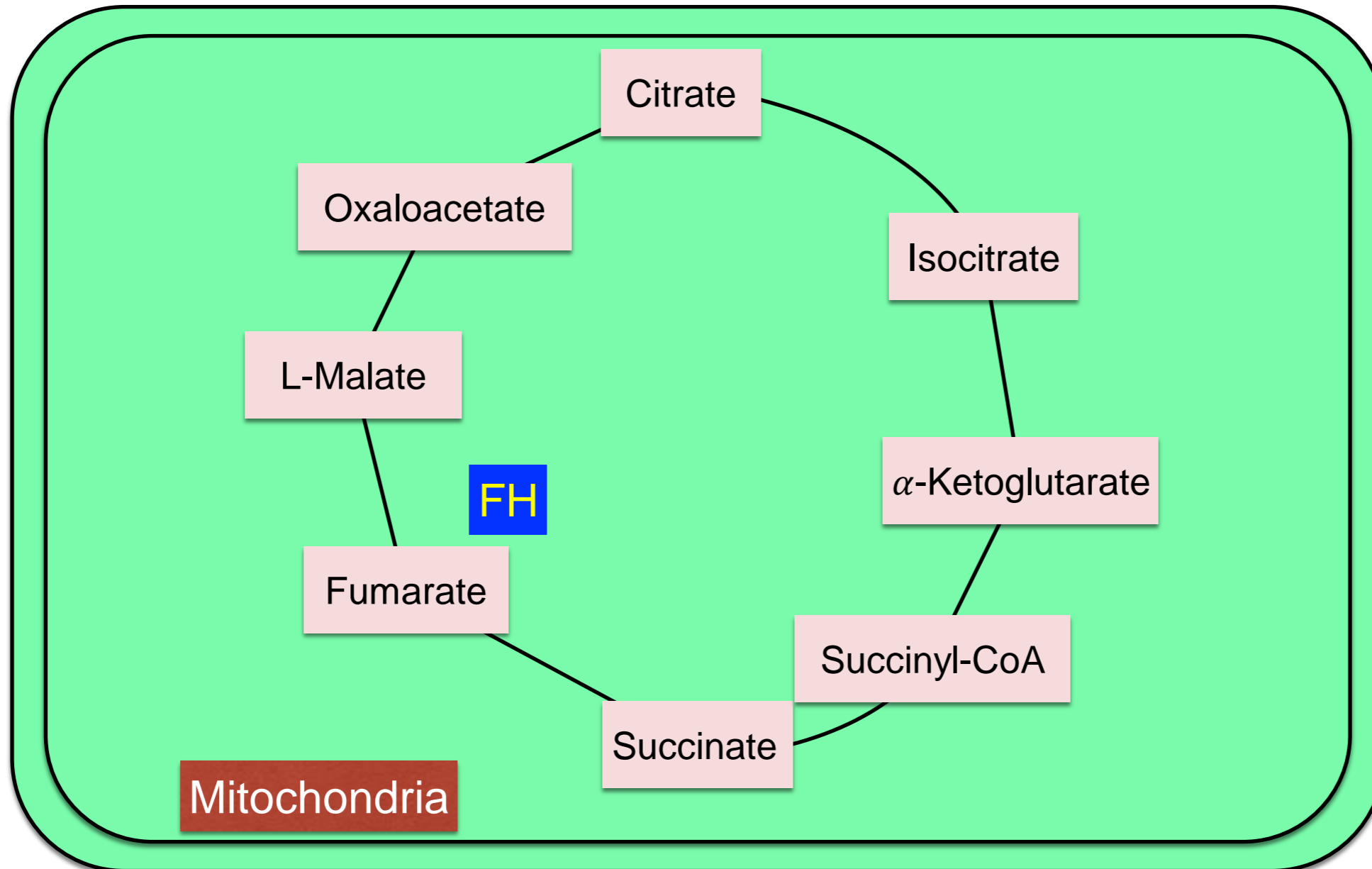
Mutations in SDH are found in familial paraganglioma and pheochromocytoma, renal carcinomas, T-Cell leukaemia, and gastrointestinal stromal tumours

SDH mutants leads to Succinate accumulation

What is the function of succinate in cancer?



# Fumarate hydratase



FH converts fumarate to malate

# FH mutation

heterozygous FH mutations followed by the loss of heterozygosity of the second allele cause Hereditary Leiomyomatosis and Renal Cell Cancer (HLRCC).

FH mutants leads to fumarate accumulation

What is the function of fumarate in cancer?

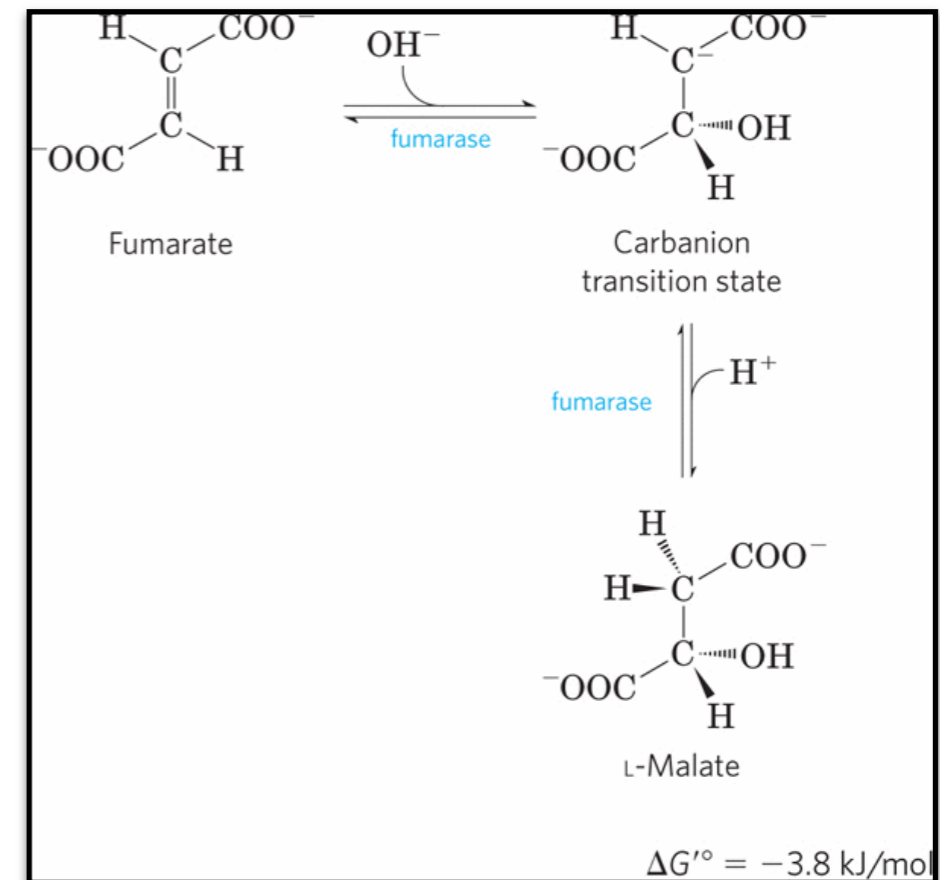
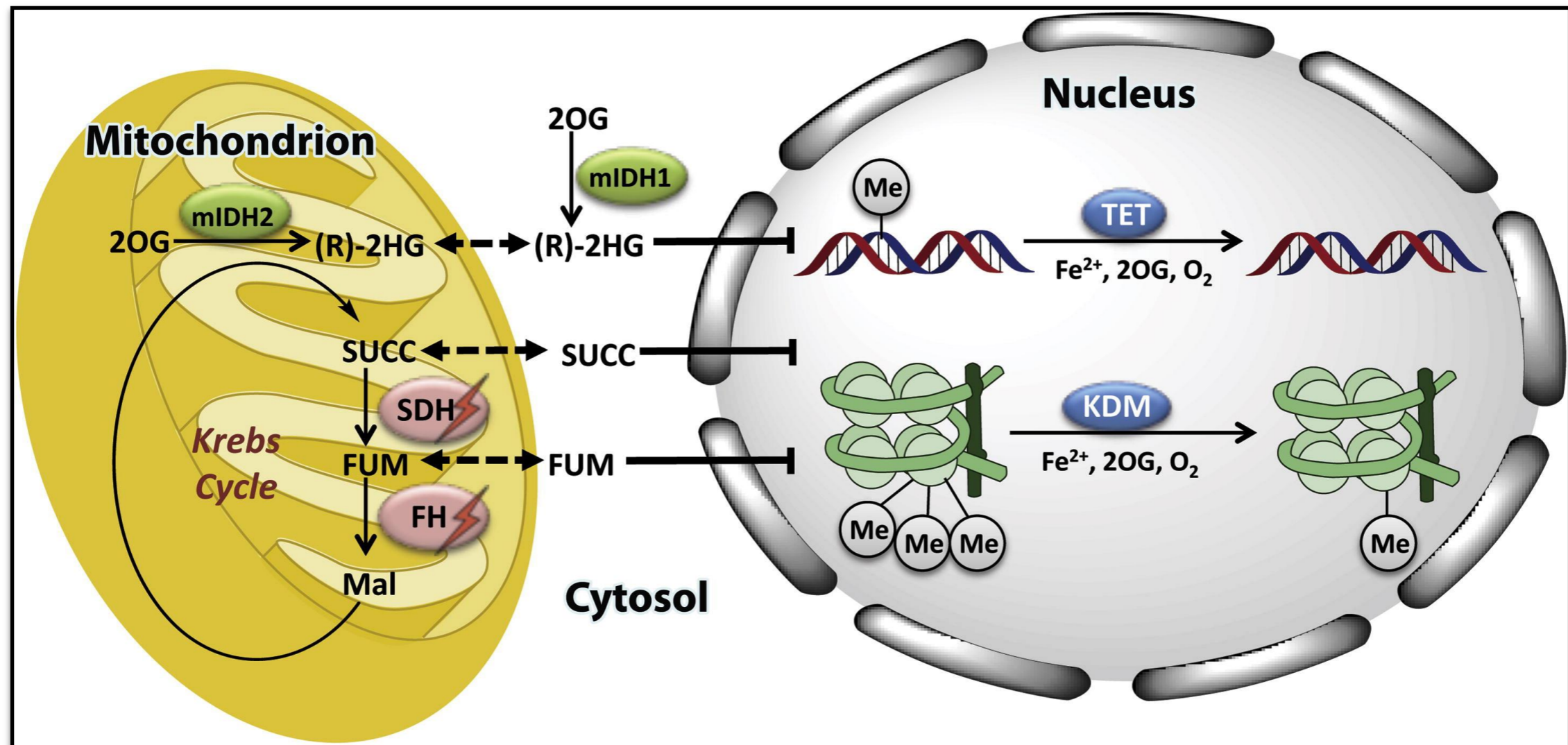


Figure 16 Lehninger

# Epigenetic Reprogramming by Oncometabolites

Modification in the function of all three enzymes leads to epigenetic changes





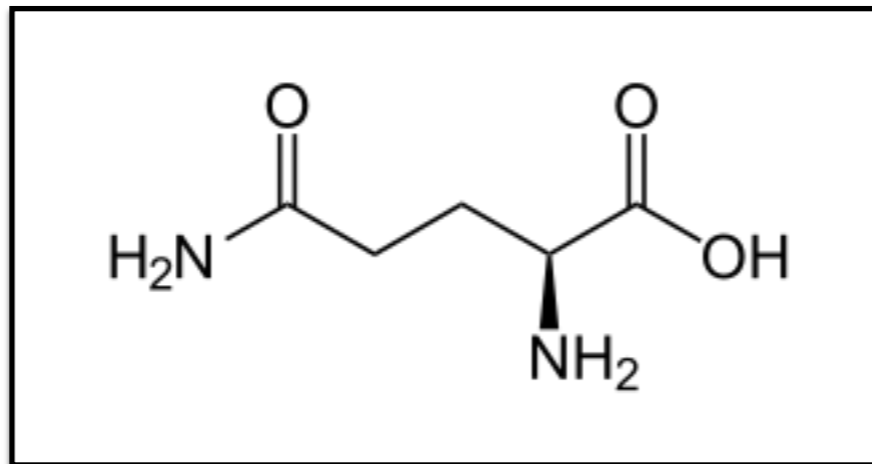
# Amino acid and cancer

## Glutamine

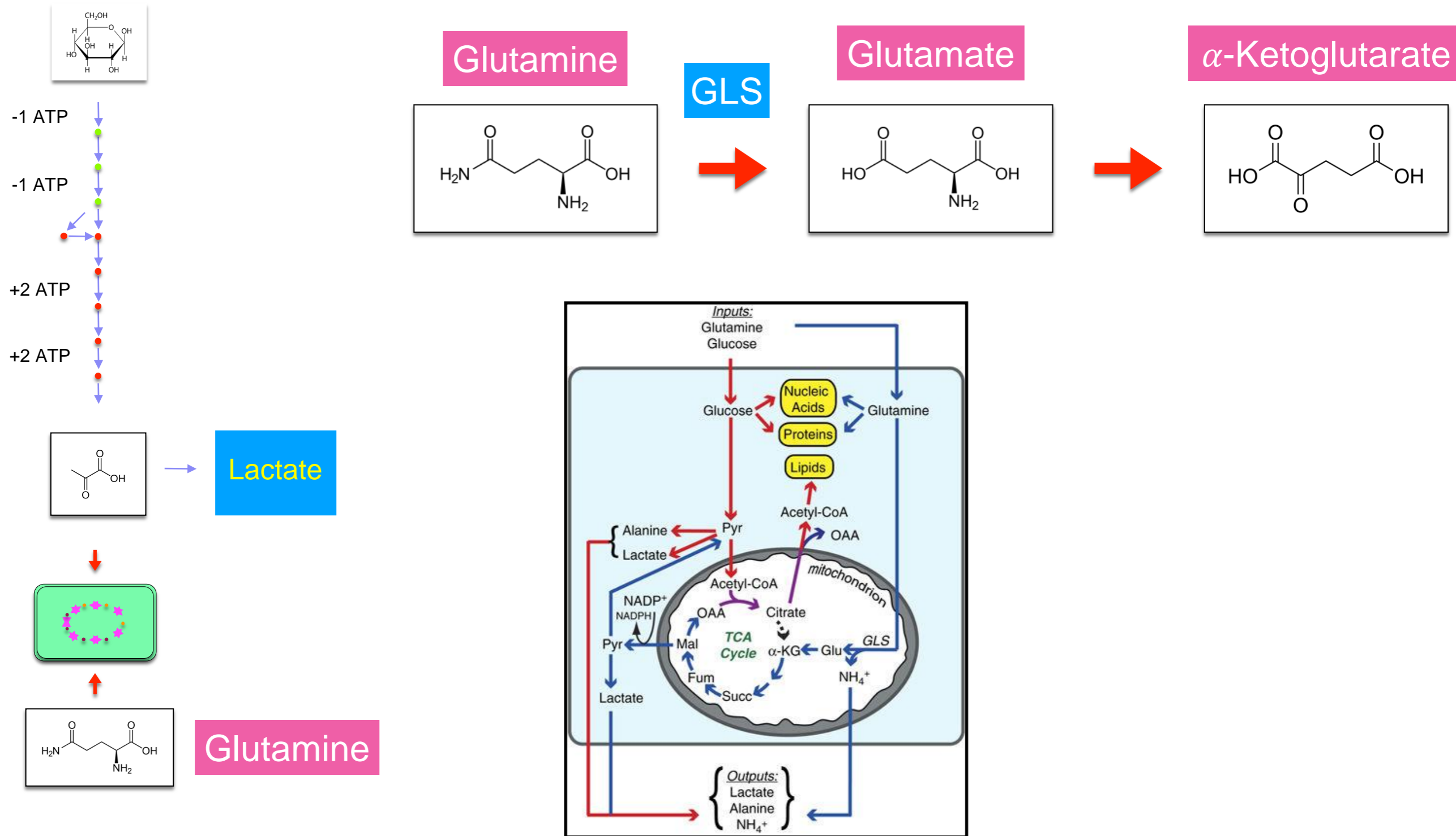
The most abundant amino acid in the plasma

Major carrier of nitrogen between the organs

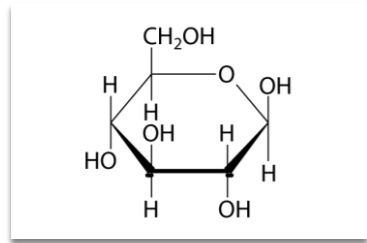
Proliferating cells use glutamine as the nitrogen donor for biosynthesis of nucleotides, nonessential amino acids, and hexosamines



# Glutamine can fuel the TCA cycle

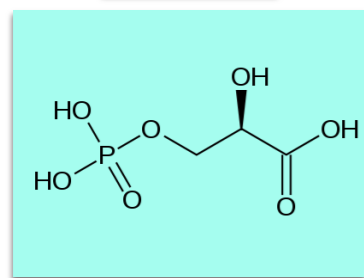


# Serine Synthesis Pathway (SSP)

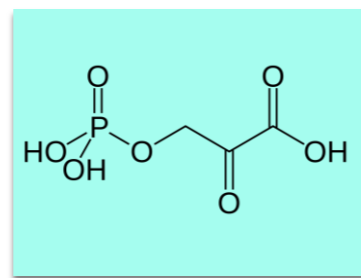


Three step reaction

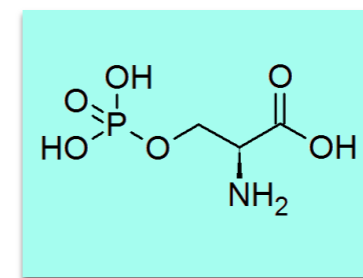
3-PG



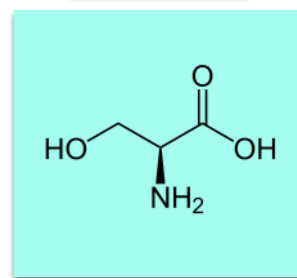
**PHGDH**



**PSAT1**



**PSPH**

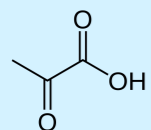


Serine

NAD<sup>+</sup>    NADH

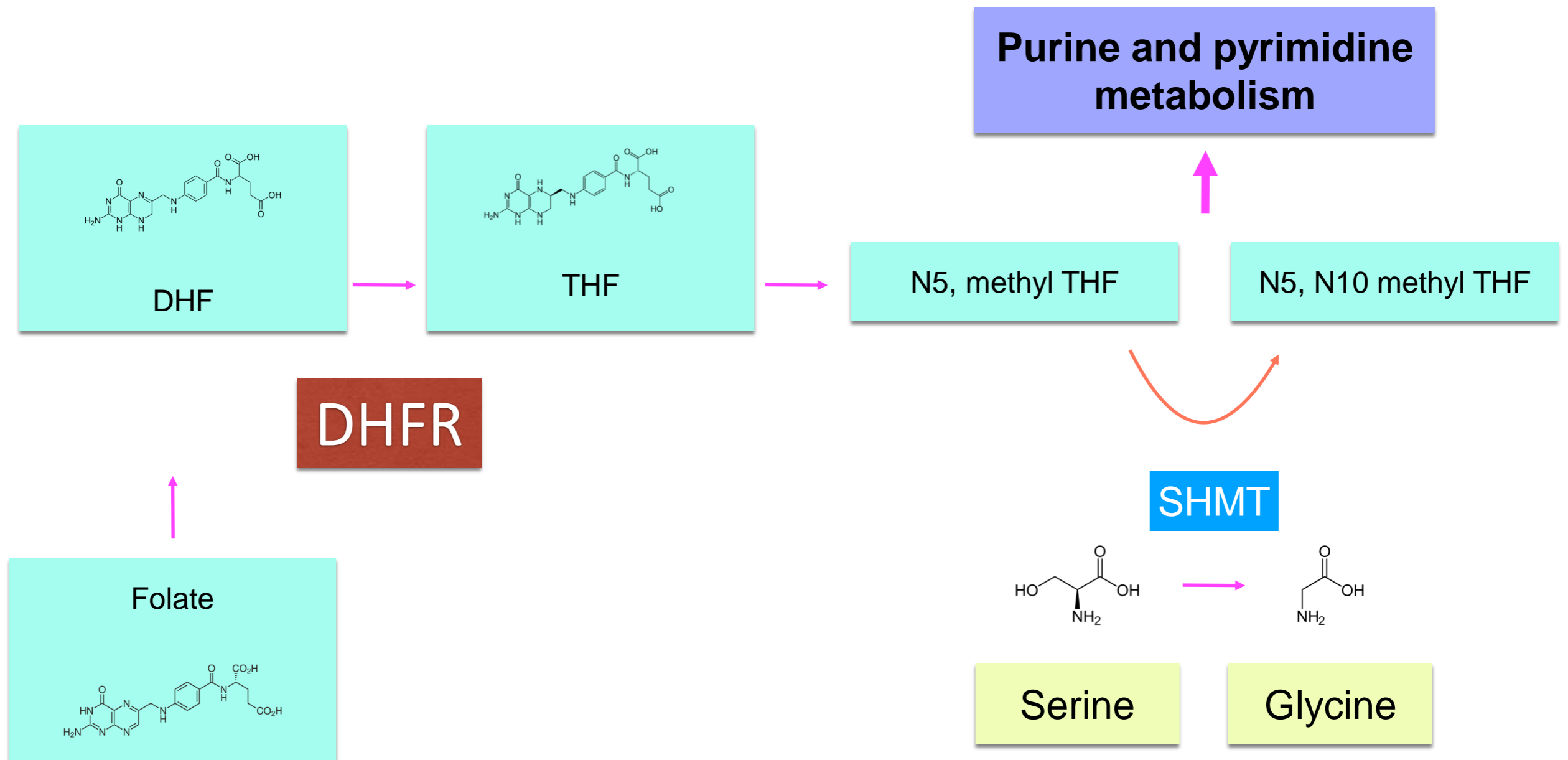
Glu     $\alpha$ -KG

H<sub>2</sub>O



2x Pyruvate

# Serine and one-carbon metabolism

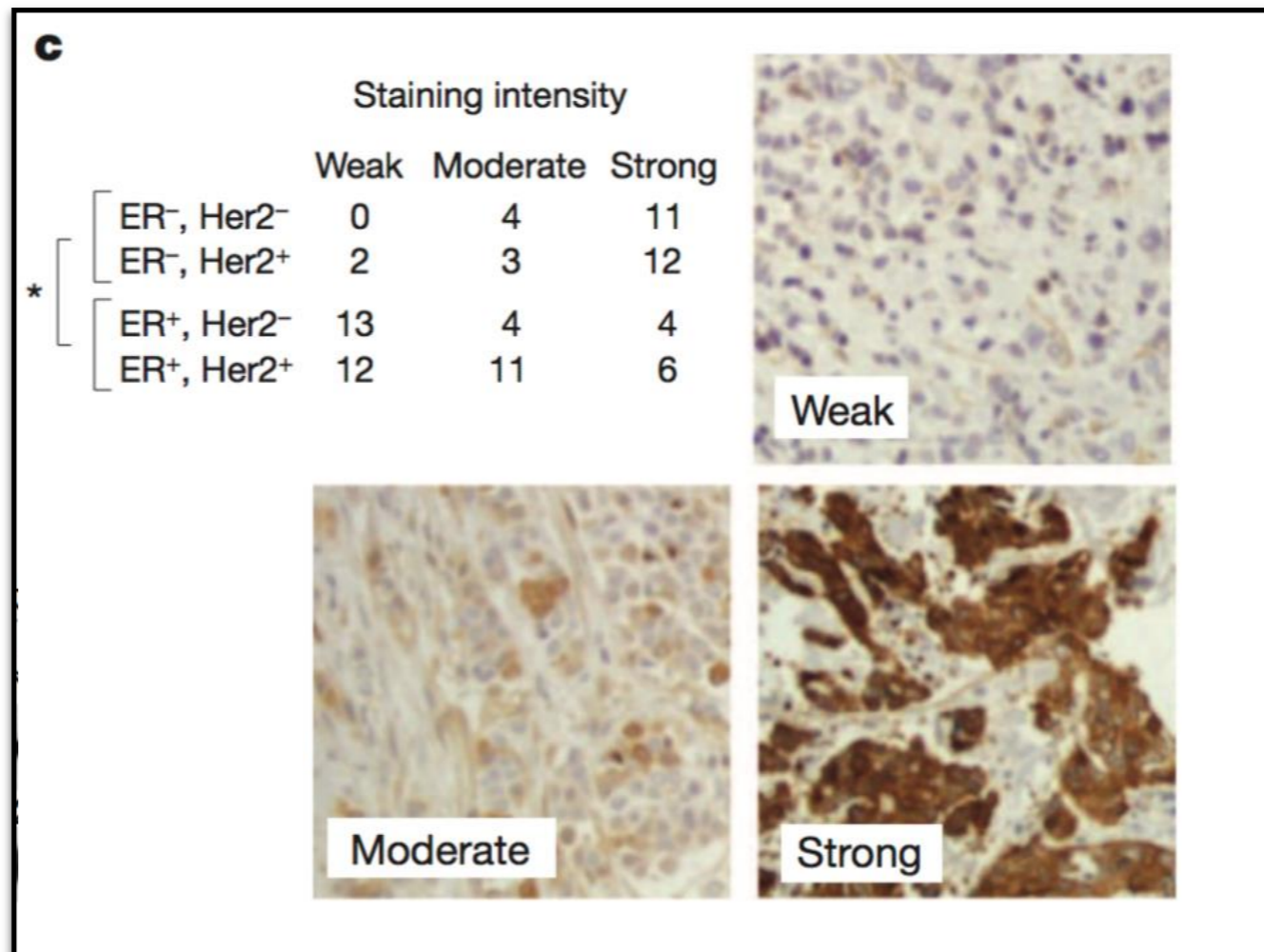


Serine conversion to glycine, transfer of one carbon to N5, methyl-THF

This carbon is essential for nucleotide metabolism

# PHGDH amplification in cancer

PHGDH expression is associated with aggressive breast cancer markers



Staining of PHGDH in different breast cancer samples

PHGDH is also overexpressed in gliomas

Possemato, R., Marks, K. M., Shaul, Y. D., Pacold, M. E., Kim, D., Birsoy, K., Sethumadhavan, S., Woo, H.-K., Jang, H. G., Jha, A. K., Chen, W. W., Barrett, F. G., Stransky, N., Tsun, Z.-Y., Cowley, G. S., Barretina, J., Kalaany, N. Y., Hsu, P. P., Ottina, K., Chan, A. M., Yuan, B., Garraway, L. A., Root, D. E., Mino-Kenudson, M., Brachtel, E. F., Driggers, E. M., and Sabatini, D. M. (2011) Functional genomics reveal that the serine synthesis pathway is essential in breast cancer. *Nature*. 476, 346–350



# Metabolic enzymes as drug targets

All improve overall survival

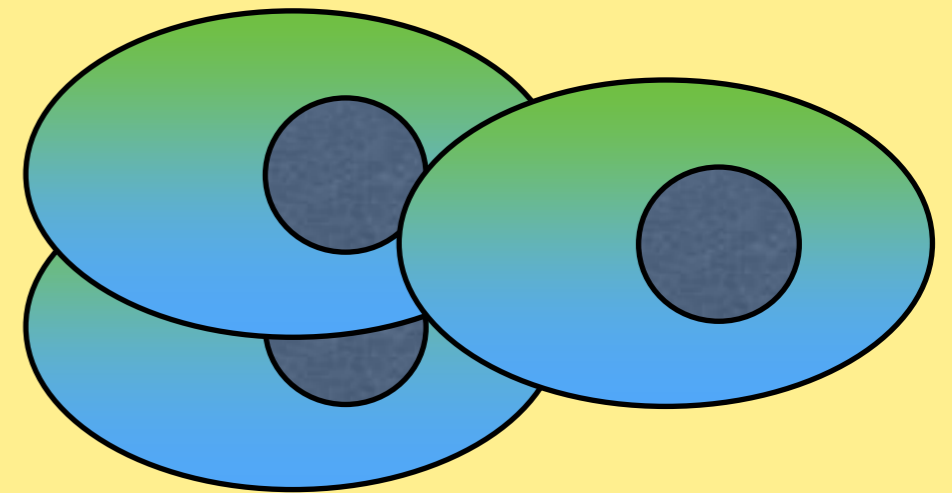
However, many of these drugs also target normal proliferating cells

## Antimetabolites

5-fluorouracil  
Methotrexate  
Hydroxyurea



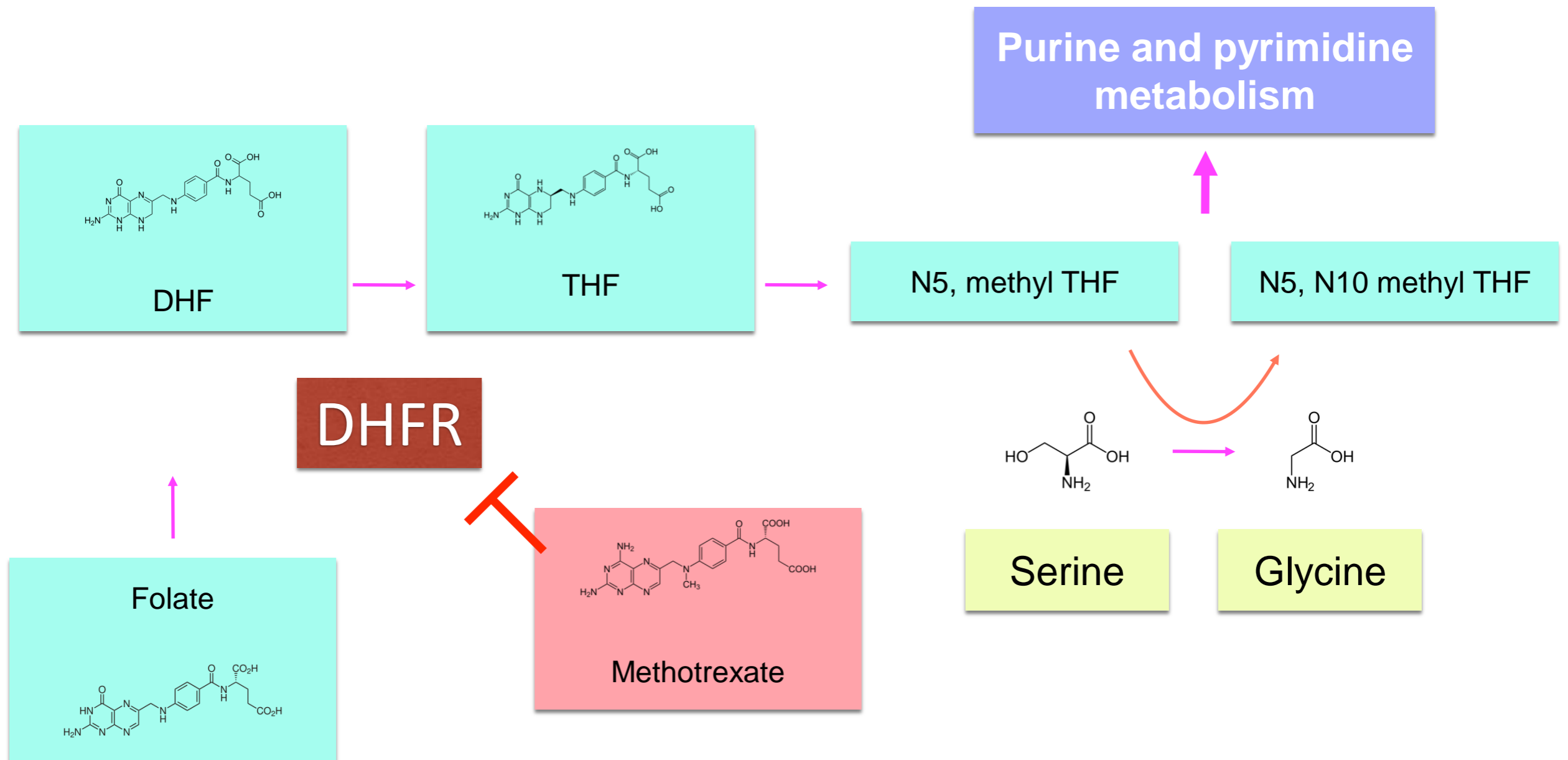
Proliferating cells



Building blocks

Support rapid growth

# Methotrexate

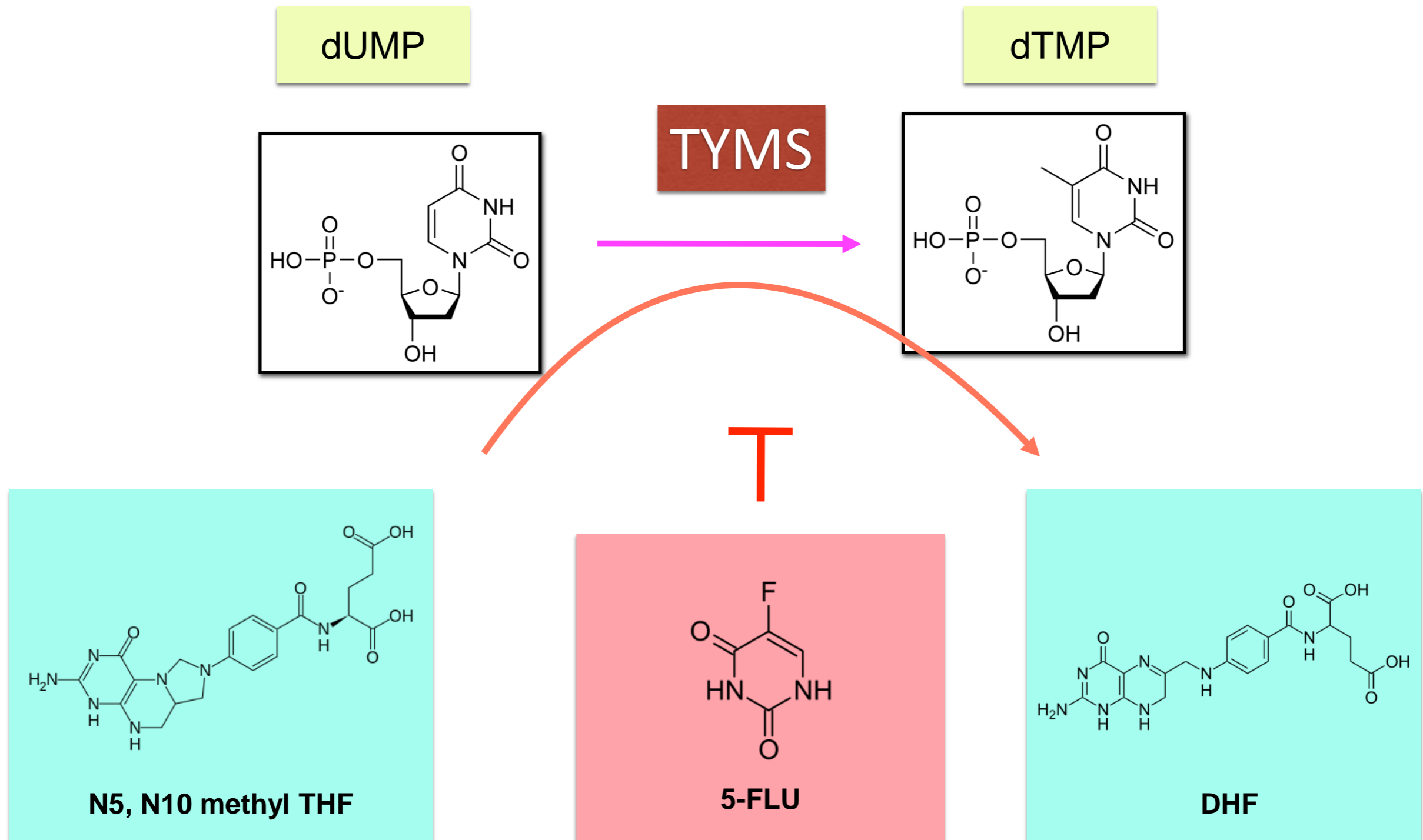


Serine conversion to glycine, transfer of one carbon to N5, methyl-THF

This carbon is essential for nucleotide metabolism

Methotrexate inhibits the ability of cells to synthesise nucleotides

# 5-fluorouracil



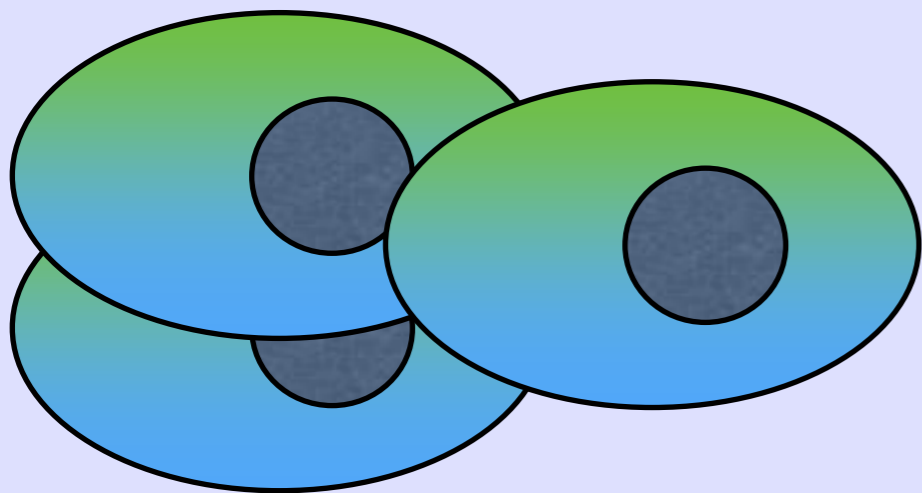
- 5-FLU inhibits TYMS, an important enzyme in pyrimidine synthesis

# Metabolic liabilities

Several tumor types are auxotrophic for 1 or more amino acids owing to deficiencies in a corresponding endogenous biosynthesis or salvage pathway.

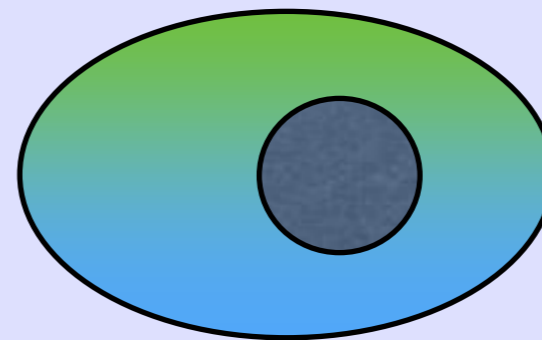
Thus,

Cancer cells



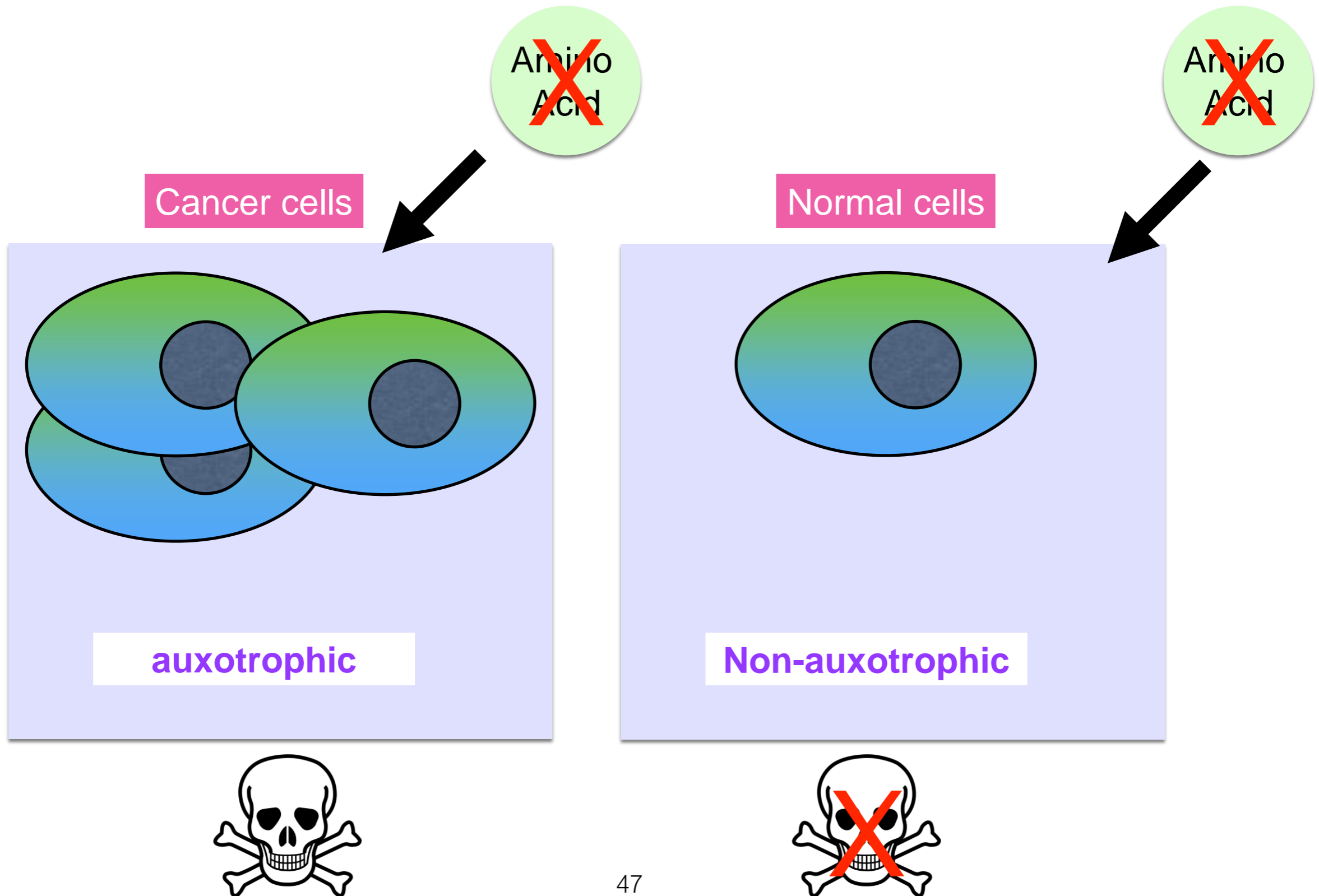
auxotrophic

Normal cells



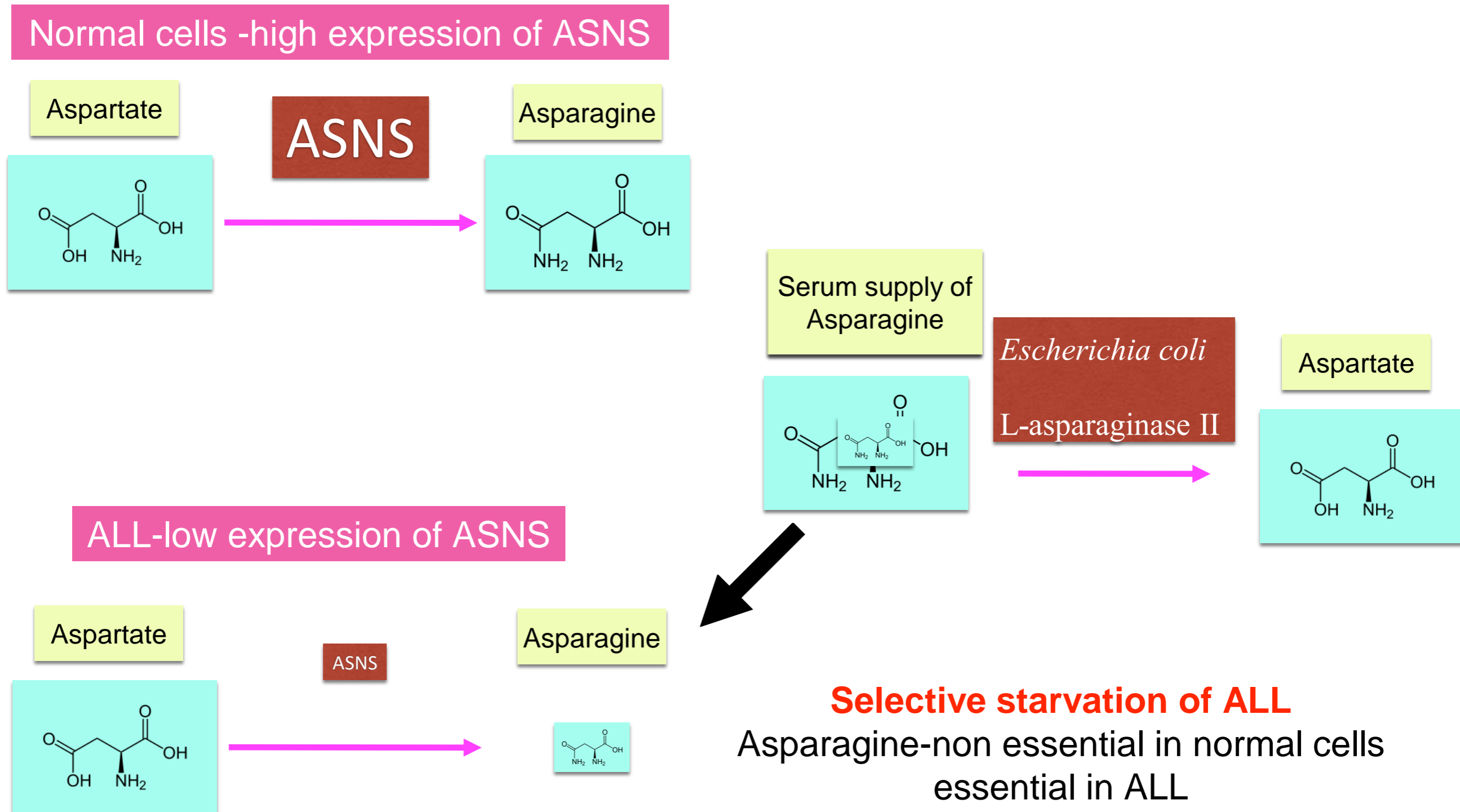
auxotrophic

# Metabolic liabilities

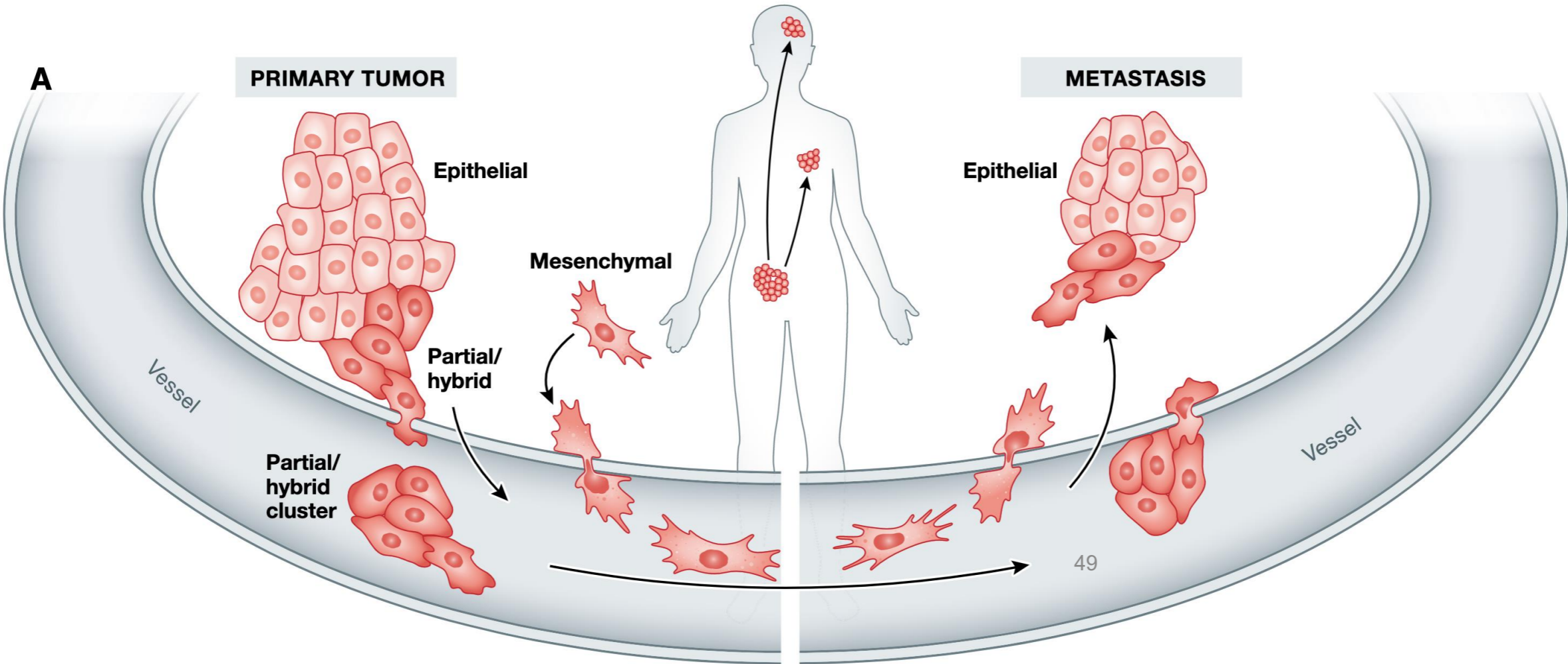




# L-asparaginase in the treatment of acute lymphoblastic leukemia (ALL)



# Metastatic Cascade



1. S. Brabletz, H. Schuhwerk, T. Brabletz, M. P. Stemmler, Dynamic EMT: a multi-tool for tumor progression. *Embo J*, e108647 (2021).

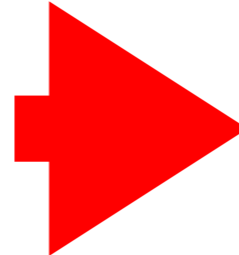


What makes the cell leave the tumor?



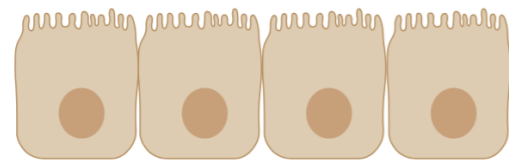


In order to execute the journey the cells needs to change





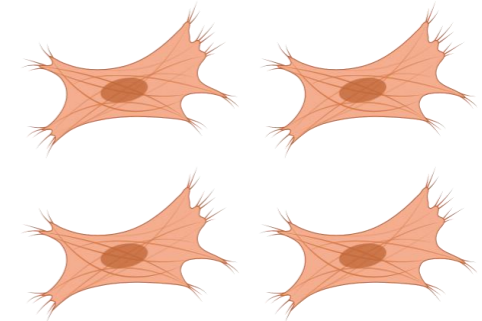
# Epithelial-Mesenchymal Transition (EMT)



**Epithelial**



**EMT**



**Partially Mesenchymal**

**Drug Response**



shutterstock · 1174498804

**Sensitive**

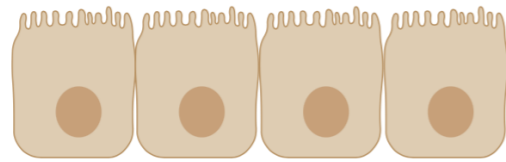


shutterstock · 1174498804

52

**Resistant**

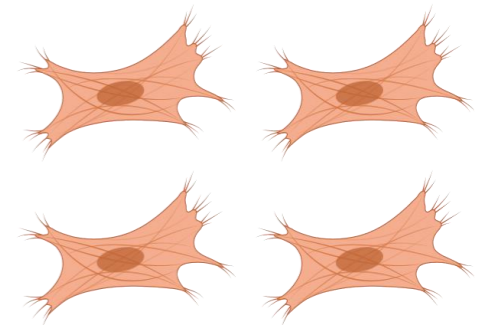
# Epithelial-Mesenchymal Transition (EMT)



**Epithelial**



**EMT**



**Partially Mesenchymal**

**Cell  
Migration**



**Slow**

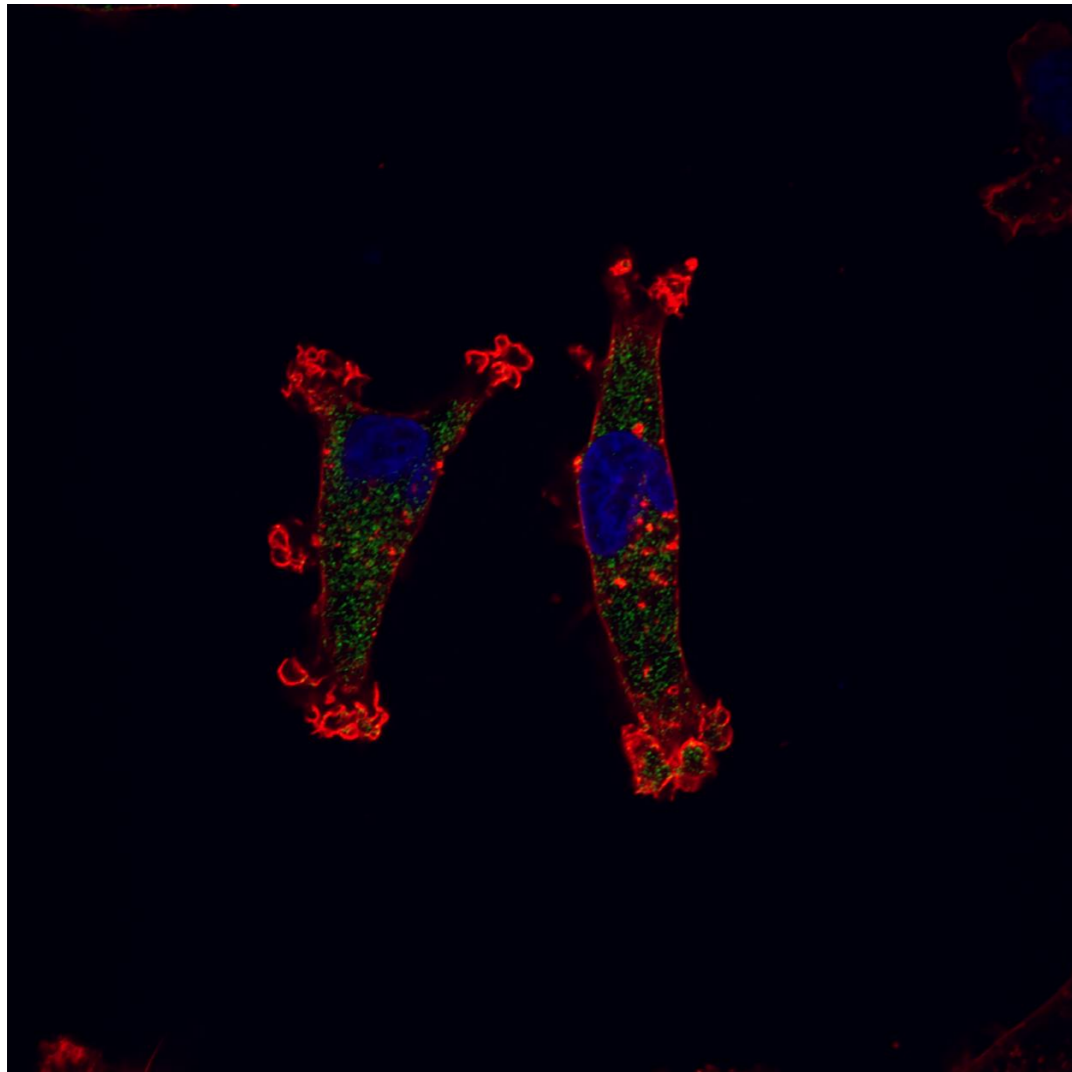
53



**Fast**



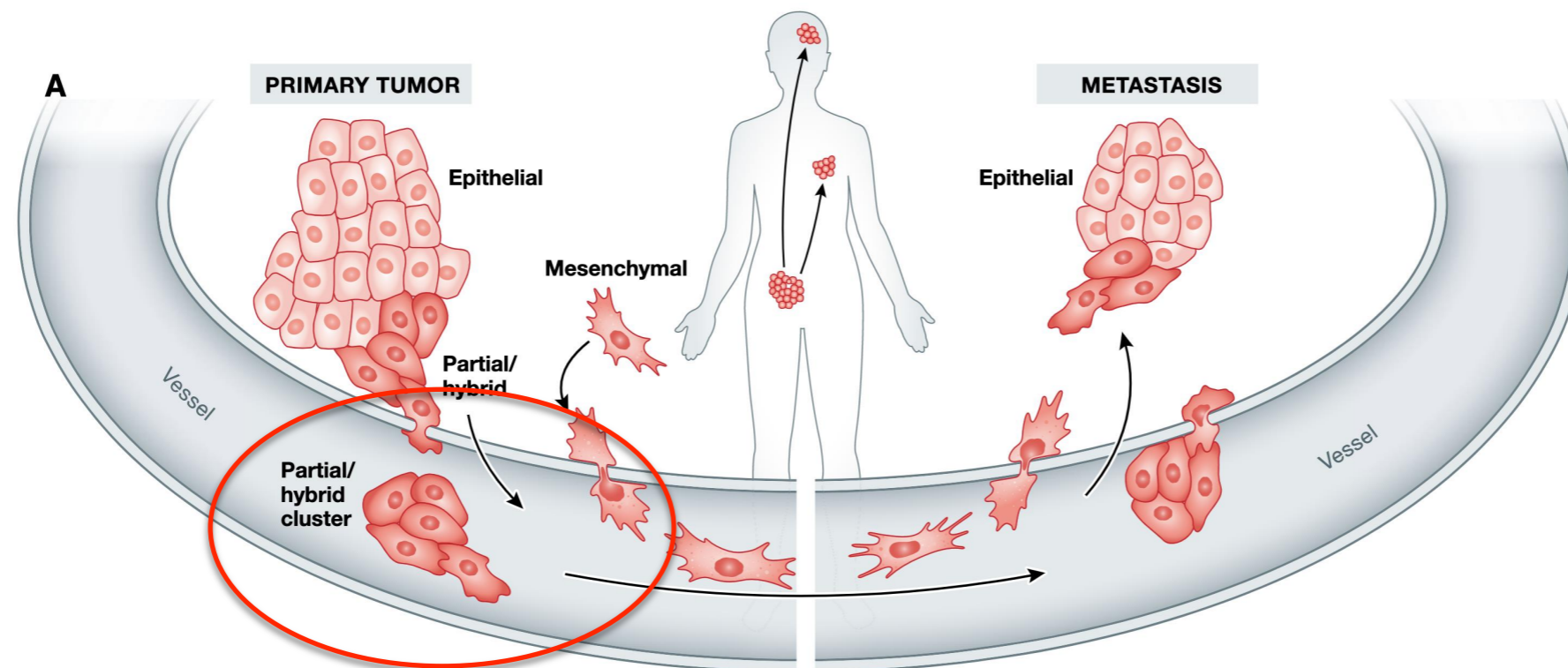
The migrating cells contains “hands” that help them migrate



A picture taken in our lab



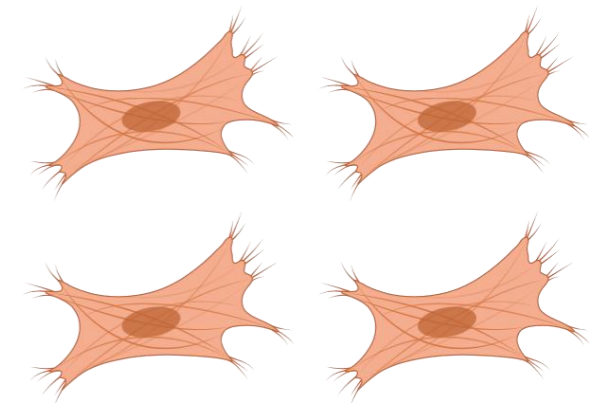
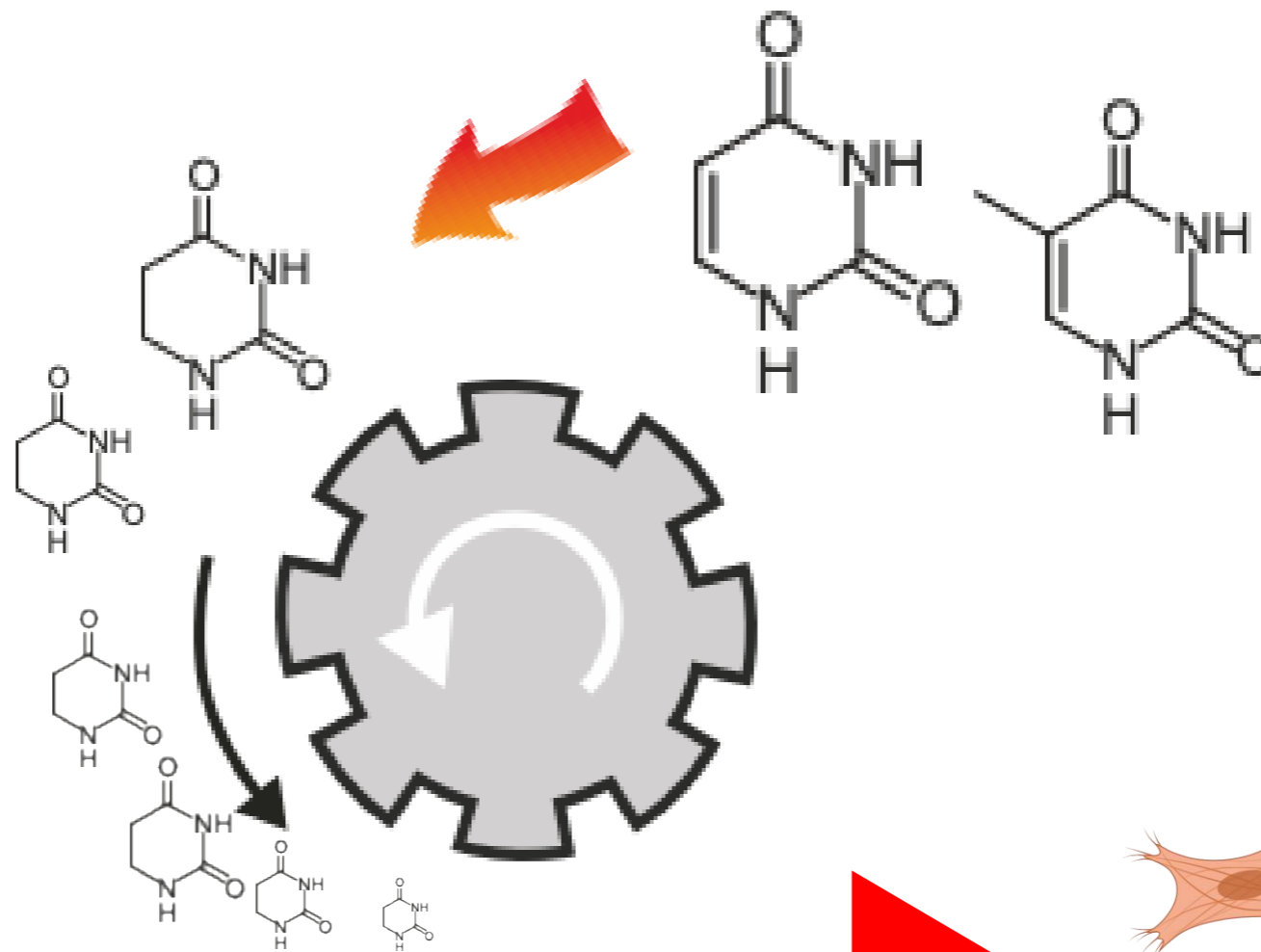
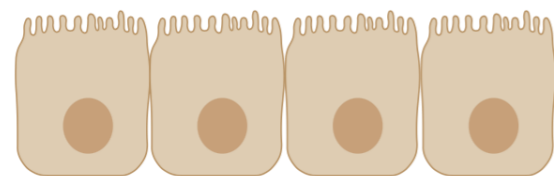
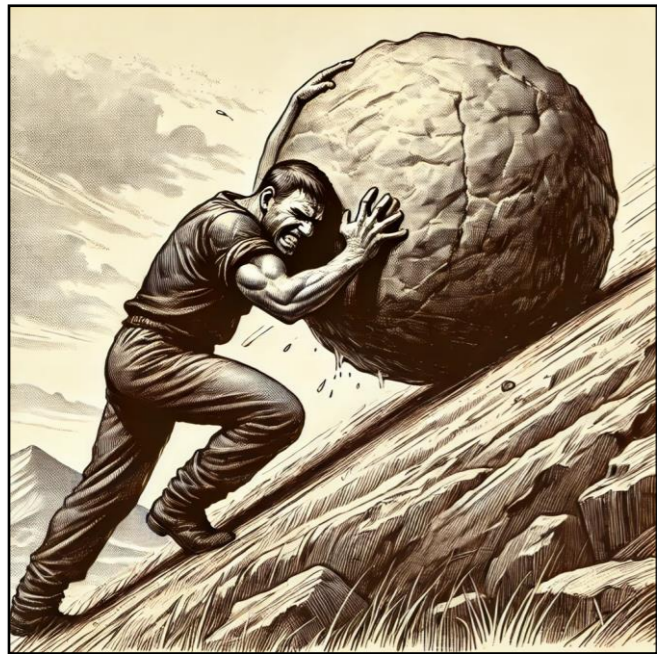
# Metastatic Cascade



55

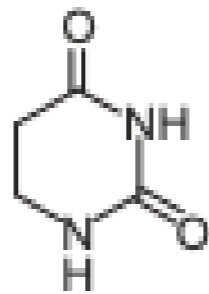
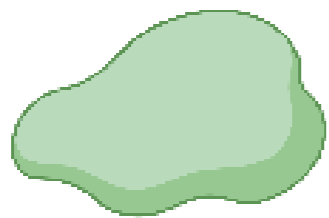
Currently, as far as we know, **there are no available anti-EMT drugs.**

My lab interest is to identify metabolic processes that push cancer cells to become aggressive

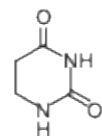
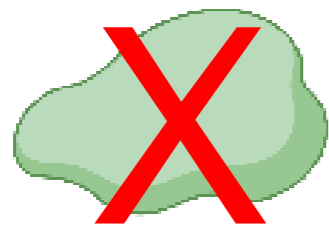




# How do we study?

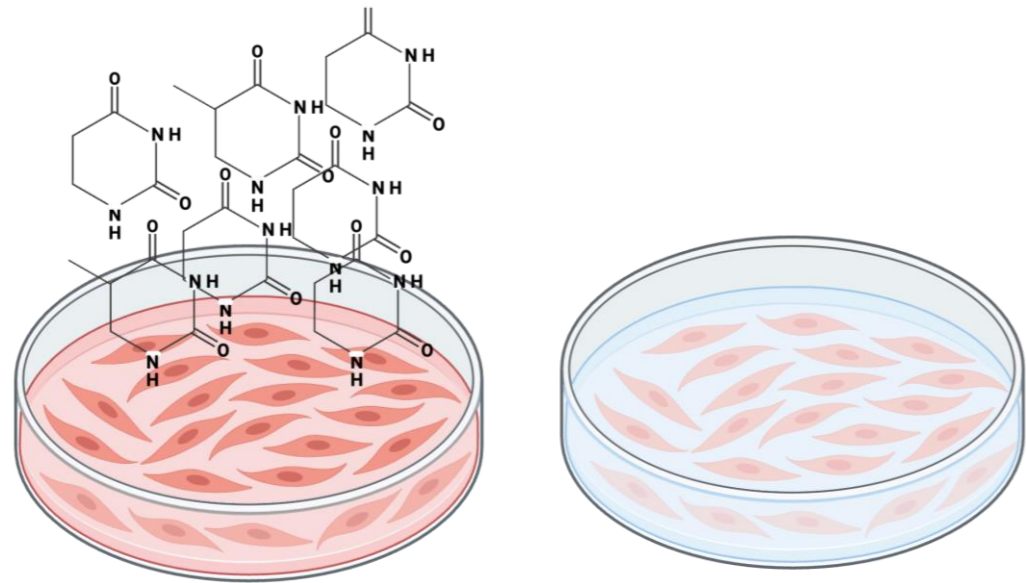


Active Enzyme

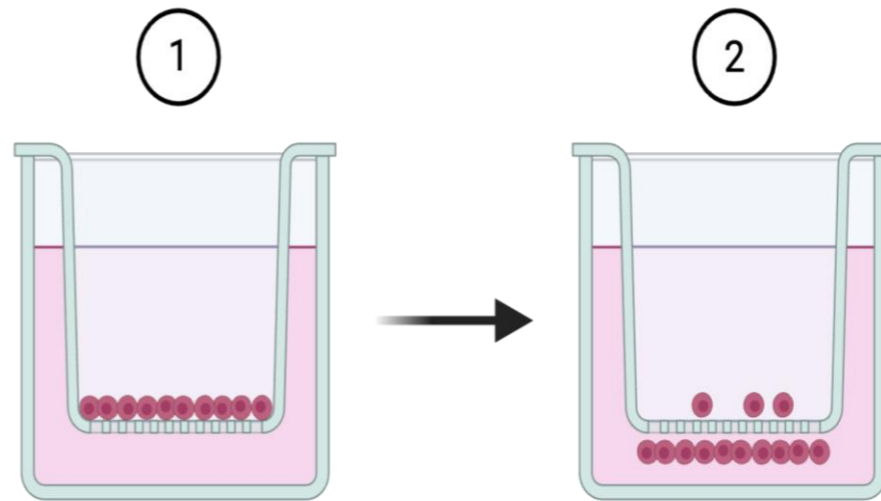


Non-active Enzyme

# Cell migration competition

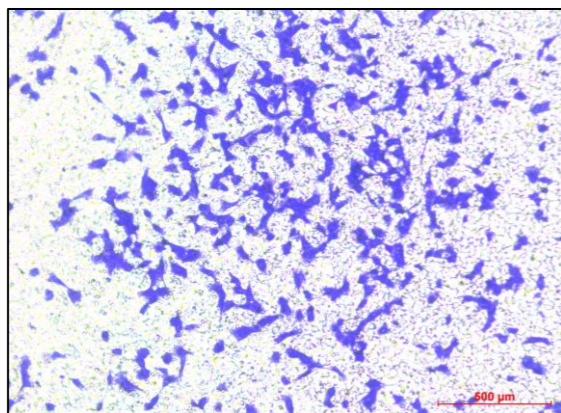


Active Enzyme Non-active Enzyme

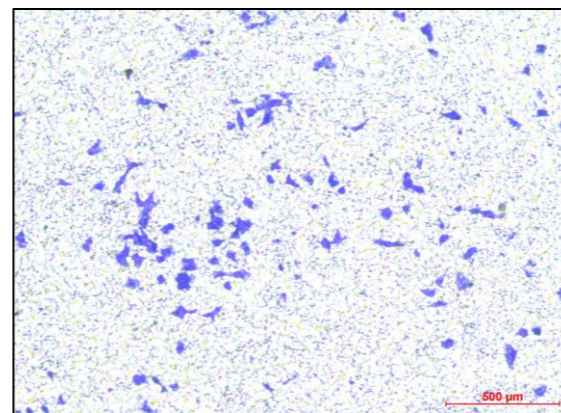


Transwell migration assay

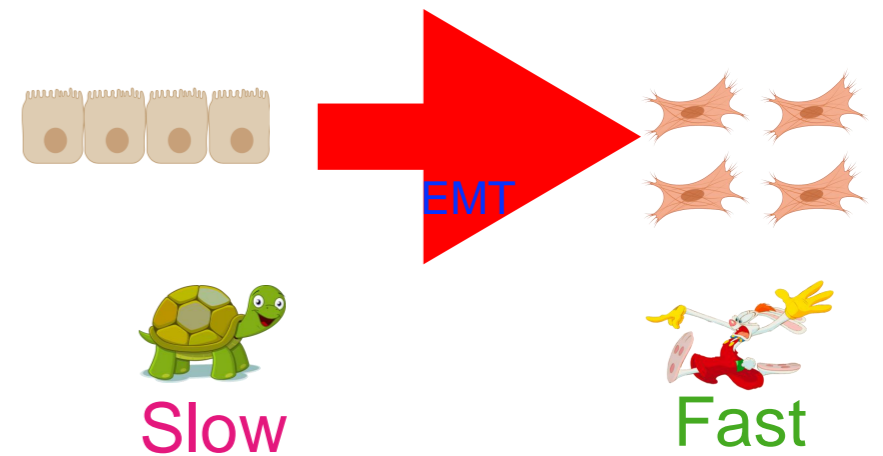
## Results



Active Enzyme



Non-active Enzyme





# My Team

Email: [yoavsh@ekmd.huji.ac.il](mailto:yoavsh@ekmd.huji.ac.il)

