

*Regulation of gene
expression, molecular
basis of signalisation*

Molecular Biology

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***REGULATION OF GENE
EXPRESSION IN
PROKARYOTES***

Levels of gene expression regulation

REPLICATION

TRANSCRIPTION



POSTTRANSCRIPTION PROCESSING



TRANSLATION



POSTTRANSLATION PROCESSING

Transcription regulation in prokaryotes

- **single type of RNA polymerase**
- **nevertheless genes are take-on and take-off according to needs**
- **prokaryotic cell must react to quick changes of environmental conditions**
- **signals for starting and stopping transcription are in the form of small molecules - substrates**
- **transport of signals to promoters are mediated by protein molecules**

Participants on the regulation of gene expression

Regulator

- any substance, which participate on regulation of molecular process

Regulatory protein

- innate entity, which participate on the regulation
- it usually binds to promoter

Allosteric effector

- substance of low molecular mass which by binding to regulatory protein change its conformation and by this its affinity to regulatory region

Types of regulators

Positive regulator

- **induces transcription, translation, ...**

Negative regulator

- **inhibite transcription, translation, ...**

Allosteric effect

= interaction of an effector with regulatory protein



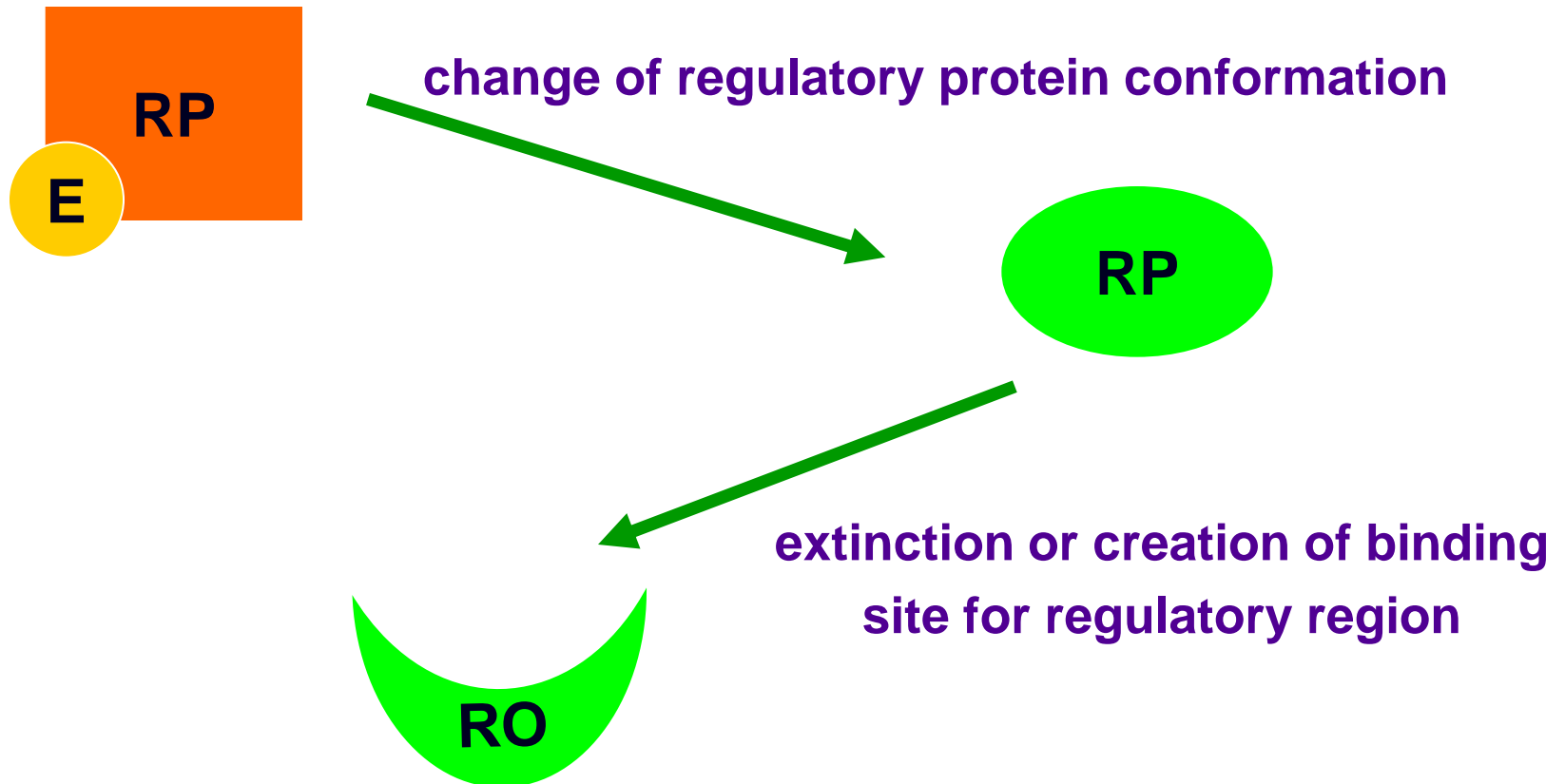
change of regulatory protein conformation



**extinction or creation of binding site for
regulatory region**

Alosteric effect

= interaction of an effector with regulatory protein



Types of allosteric effectors

Negative allosteric effector

= inhibites regulatory protein binding to regulatory region

Positive allosteric effector

= facilitates regulatory protein binding to regulatory region

Types of regulatory proteins

Negative regulatory proteins

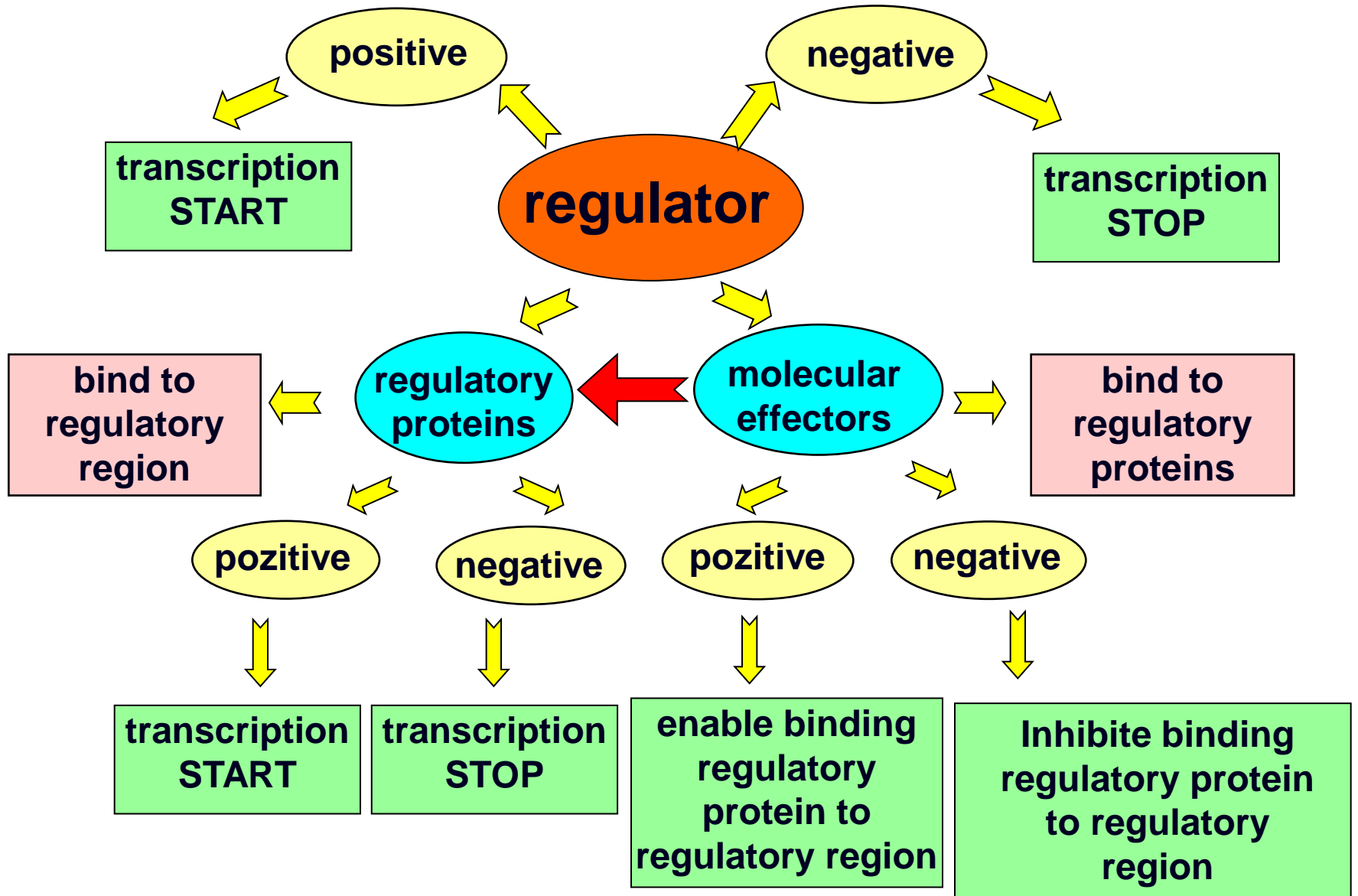
= their binding to regulatory region inhibits transcription of transcription unit by RNA polymerase

Positive regulatory proteins

= their binding to regulatory region enables transcription of transcription unit by RNA polymerase

= activator of transcription

Net of regulatory internships



Negative and positive regulation of operon

NEGATIVE

- **it is a keystone of enzymatic induction and repression**
- **binding of active repressor to operator stops transcription**

POZITIVE

- **it is a keystone of catabolic repression**
- **binding of CAP to promoter in the presence of inductor stimulates transcription**

Enzymatic induction

- **Mostly for synthesis of enzymes which is stimulated by inducer**

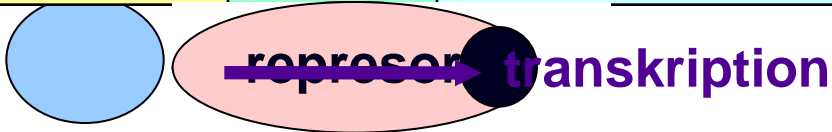
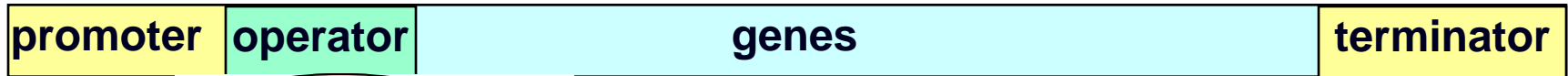
Inducible enzymes

Constitutive enzymes

- **their synthesis do not depend on the presence of inducer**
- **they are produced in cell in constant amount**

Enzymatic induction

induktor

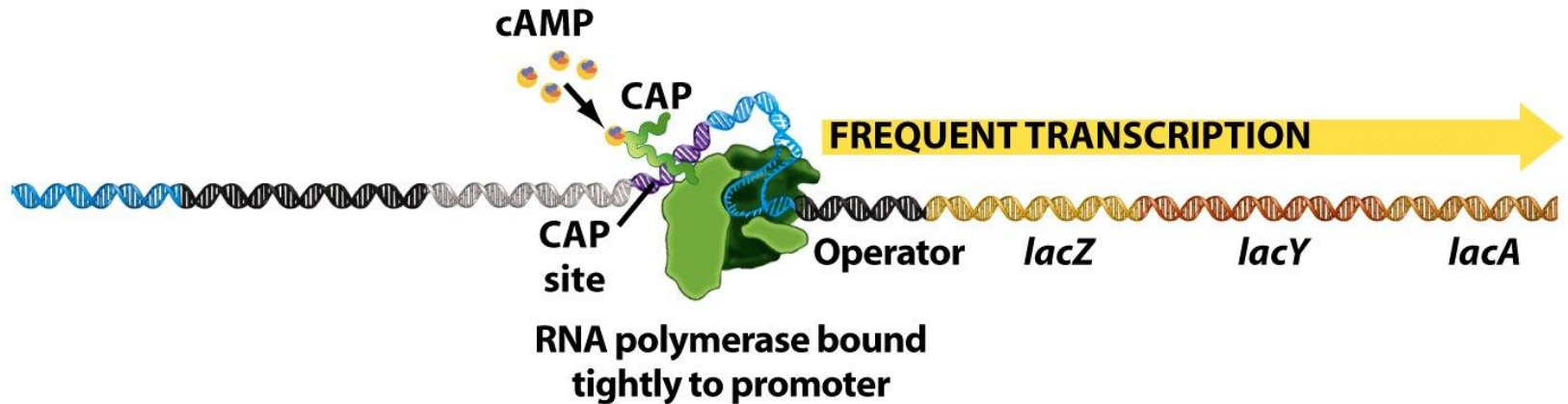


inactive complex

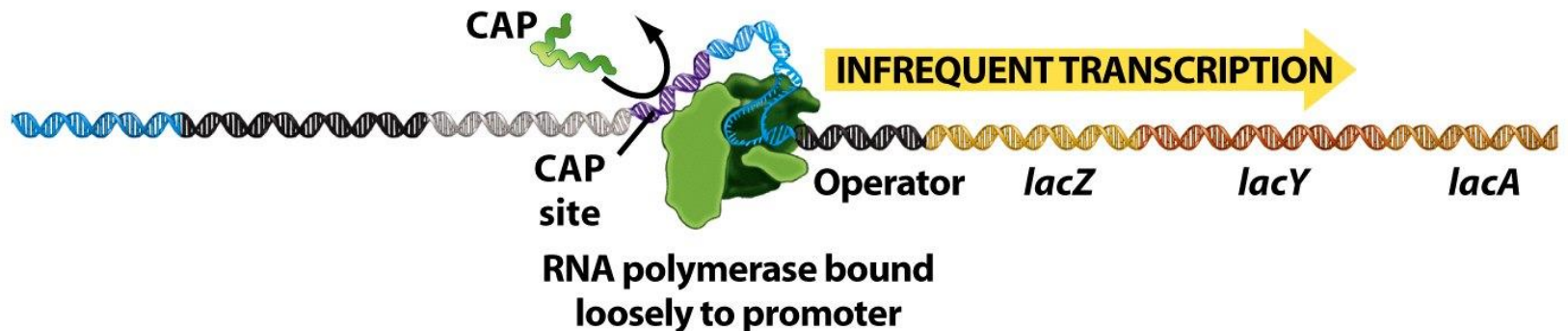
transcription of genes is ON – inducible enzymes are produced

Enzymatic induction

(a) When cAMP is present, it binds to CAP. The cAMP-CAP complex binds to DNA at the CAP site and increases binding of RNA polymerase to promoter. Transcription occurs frequently.



(b) When cAMP is absent, CAP does not bind to DNA. RNA polymerase does not bind the promoter efficiently, and transcription occurs rarely.

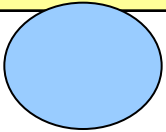
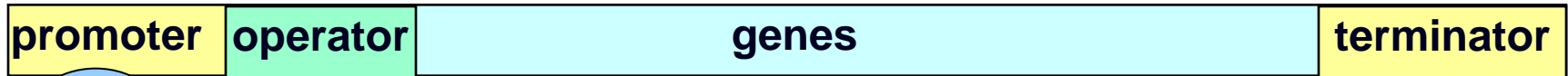


Enzymatic repression

- **Mostly for the enzymes of biosynthetic pathways**
- **The synthesis of these enzymes is inhibited by a specific metabolite of suitable metabolic pathway which is accumulated to critical amount and stops next synthesis**
- **The synthesis is renewed if concentration of the metabolite fall down under the critical amount**

Enzymatic repression

repressor

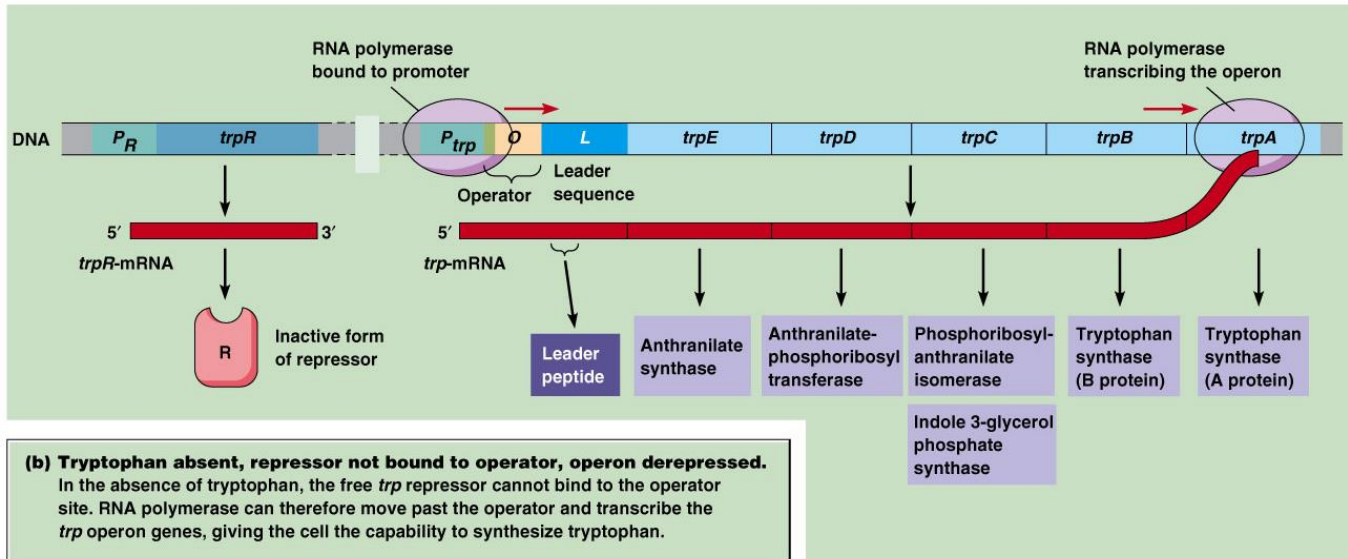


corepressor – activates
repressor

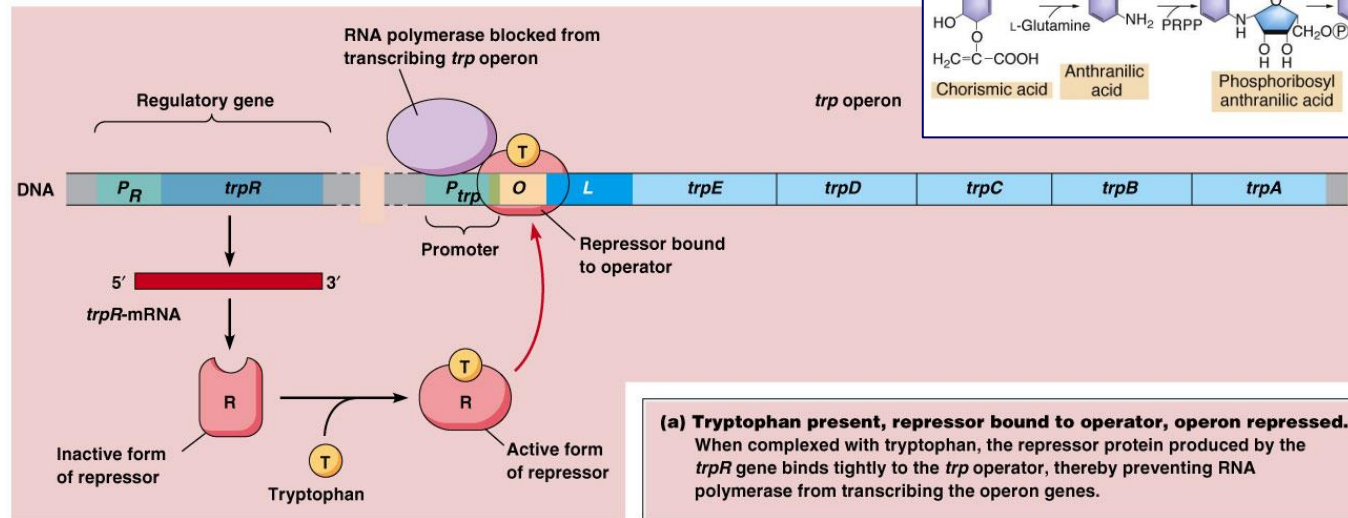
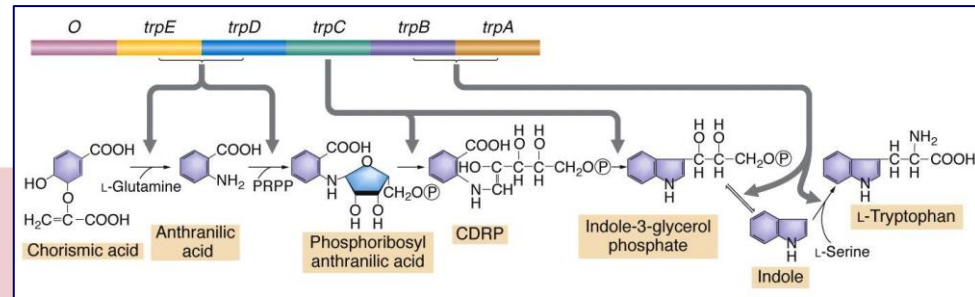


transcription of gene is OFF – synthesis of corepressor is stopped

Enzymatic repression



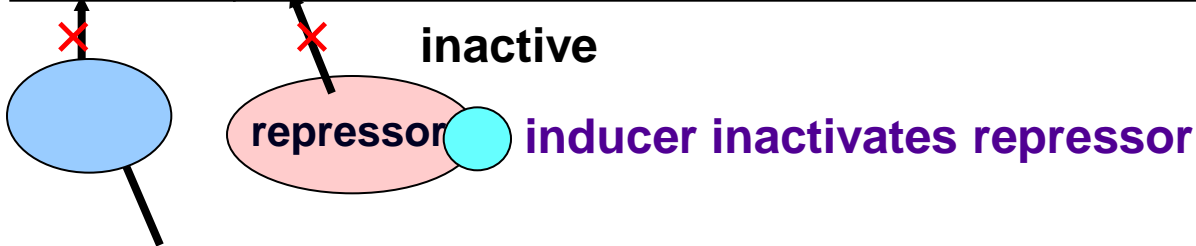
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Catabolic repression

- **Substrate inhibits synthesis of inducible enzymes although inducer is still present**
- **For example, glucose inhibits synthesis of β -galactosidase even in the presence of lactose as an inducer**

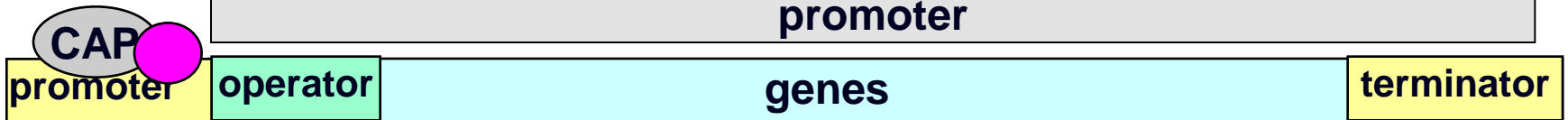
Catabolic repression



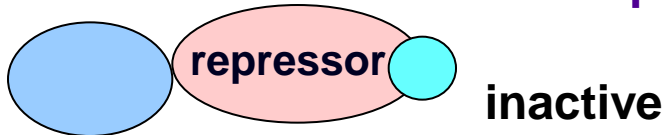
RNA polymerase has low affinity to the promoter, it is not binding, transcription is OFF or very slow



complex CAP~cAMP – escalate affinity of RNA polymerase to promoter

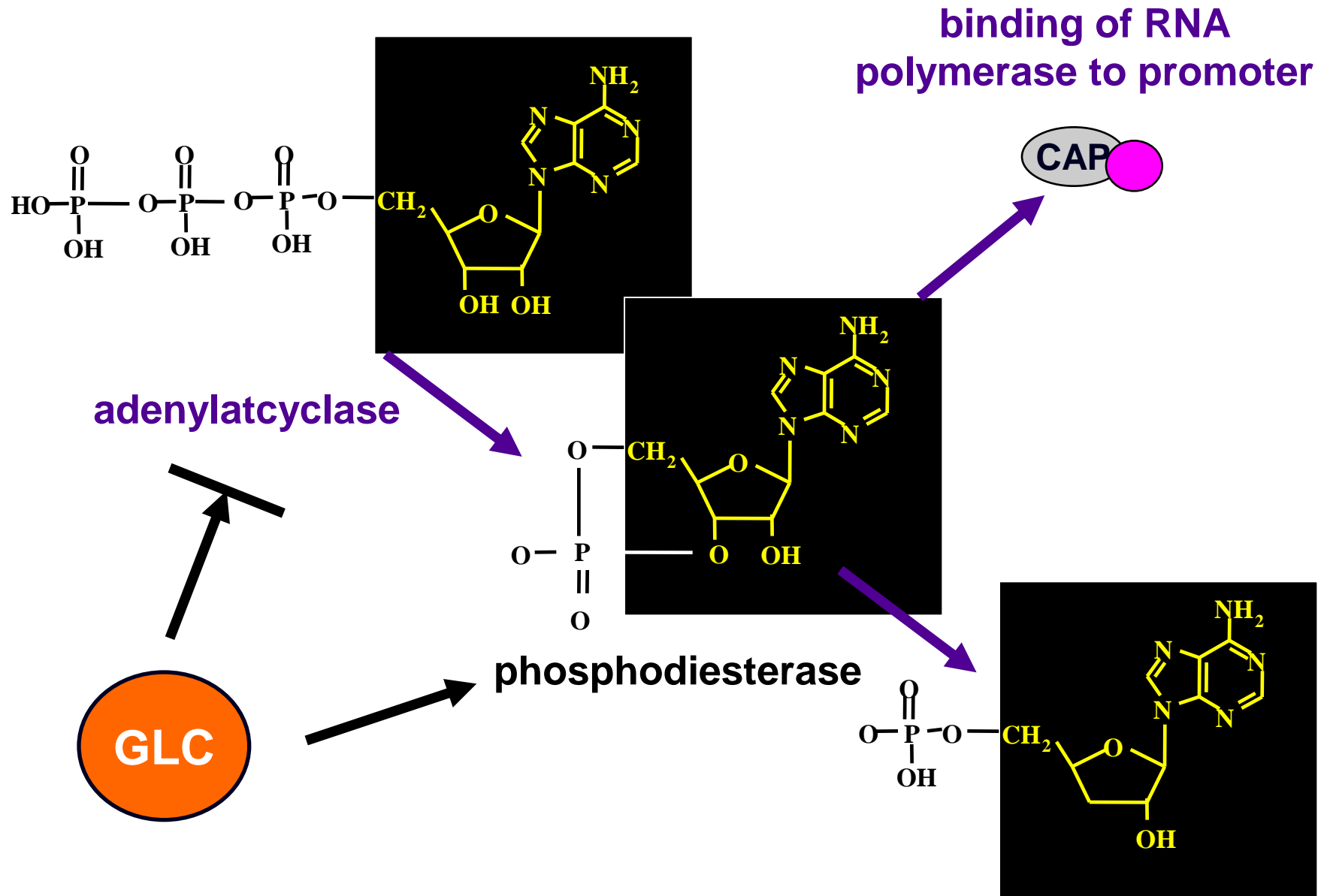


transcription



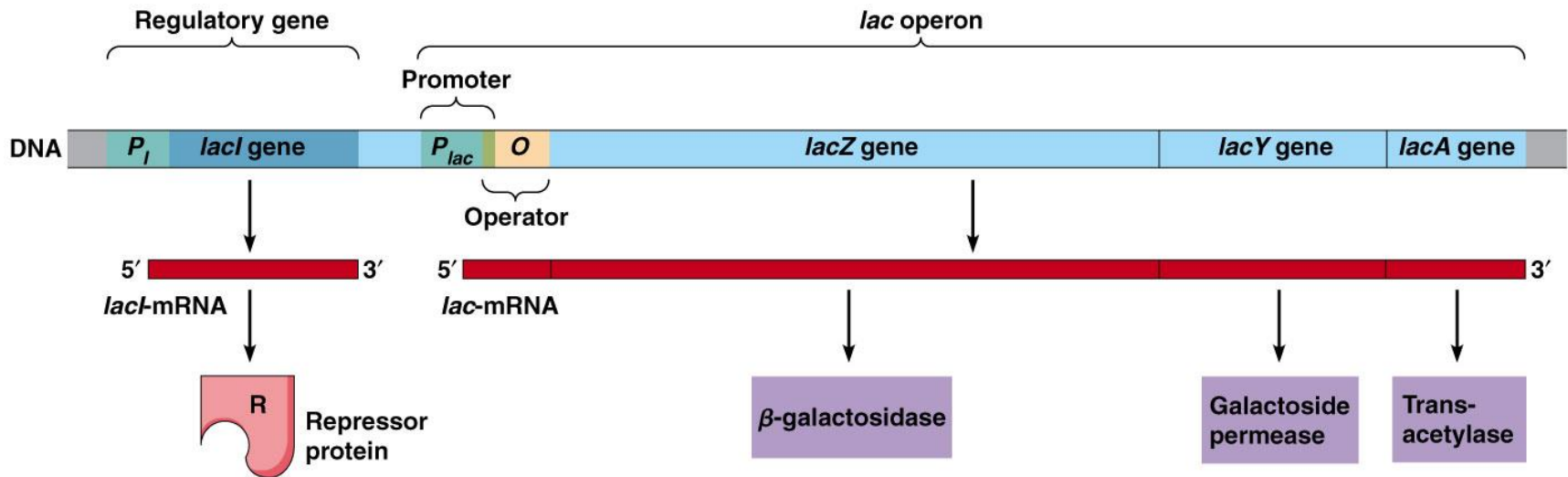
Affinity of RNA polymerase is going up, it binds to promoter, transcription is ON

Function of glc in catabolic repression



lac operon of Escherichia coli

the enzymes for metabolism of lactose



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I: codes repressor

Z: codes enzyme β -galaktosidase ($lac \rightarrow glu + gal$)

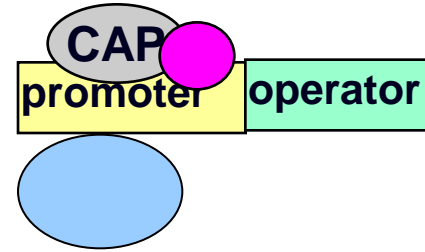
Y: codes enzyme permease

A: codes enzyme thiogalaktosidtransacetylase

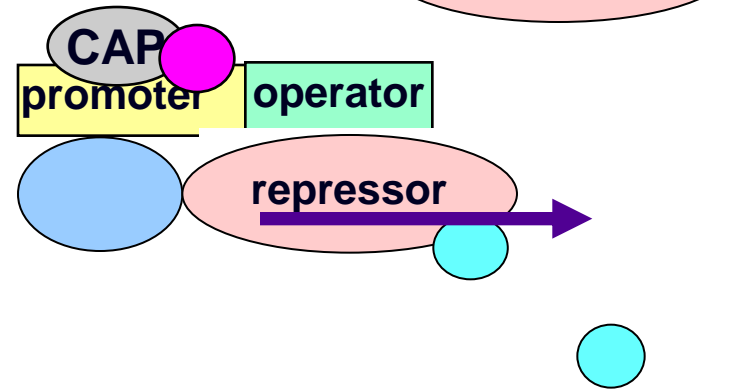
Inducer = lactose

Regulation of lac operone

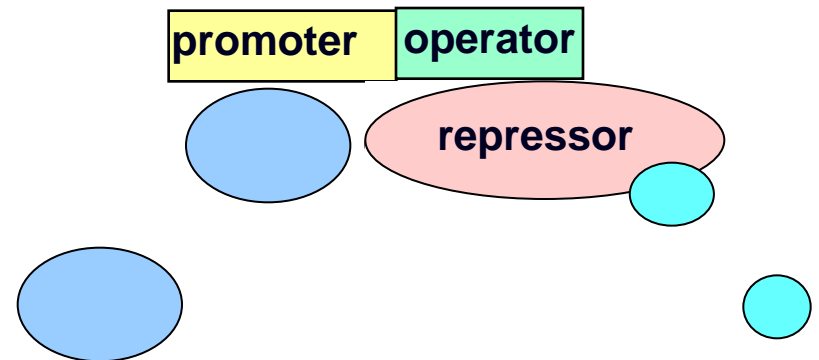
a) glc is not present, lac is not present
→ repressor binds to operator



b) glc is not present, lac is present
enzyme induction



c) glc is present, lac is present
catabolic repression



Regulate lac operonu

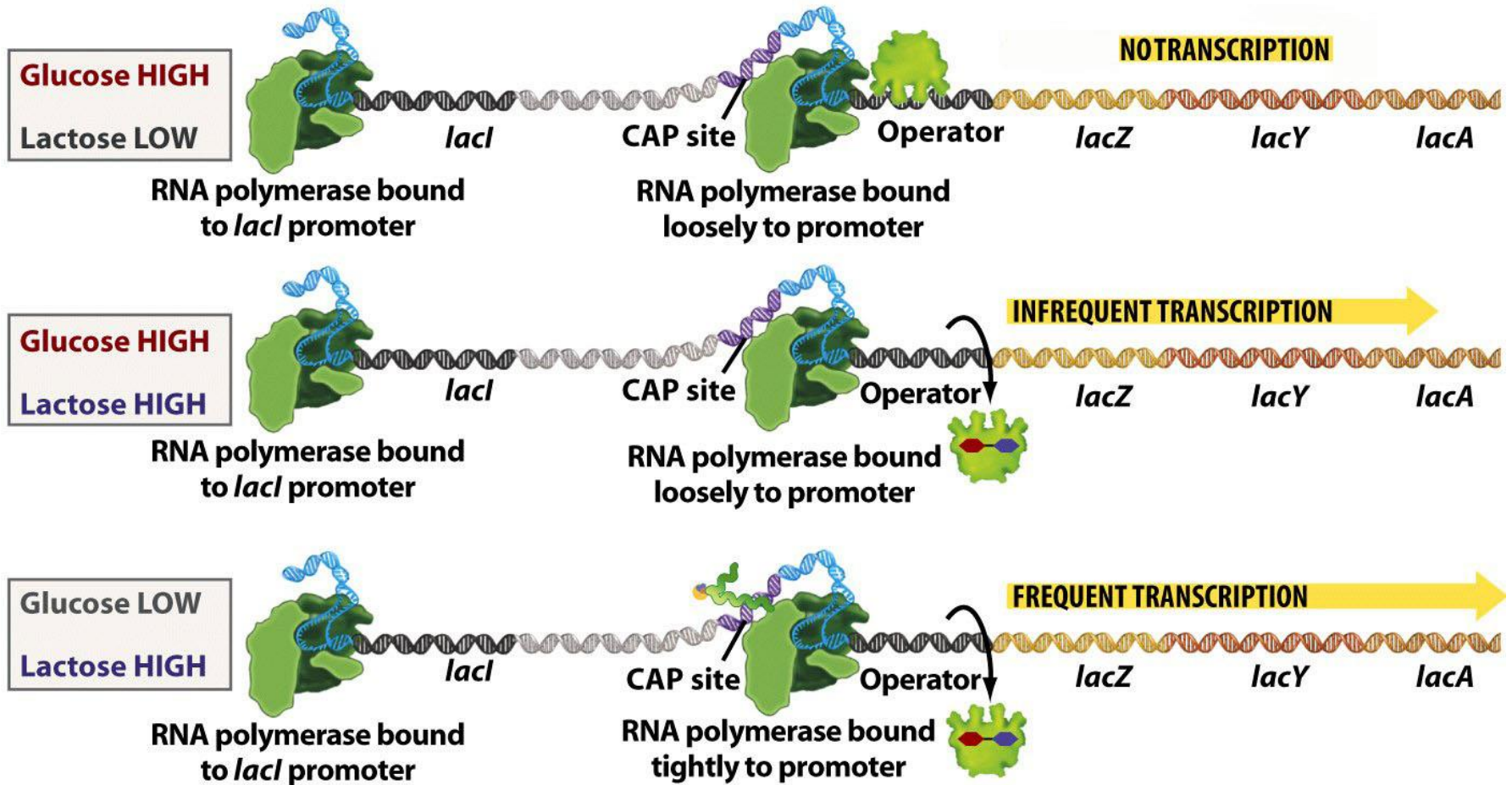


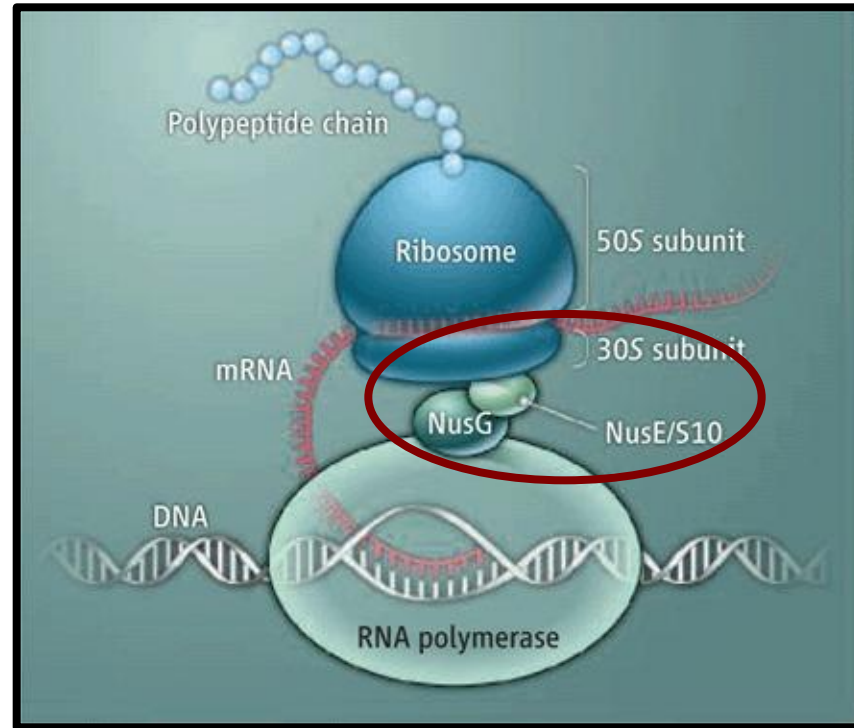
Figure 17-10 Biological Science, 2/e

Attenuation

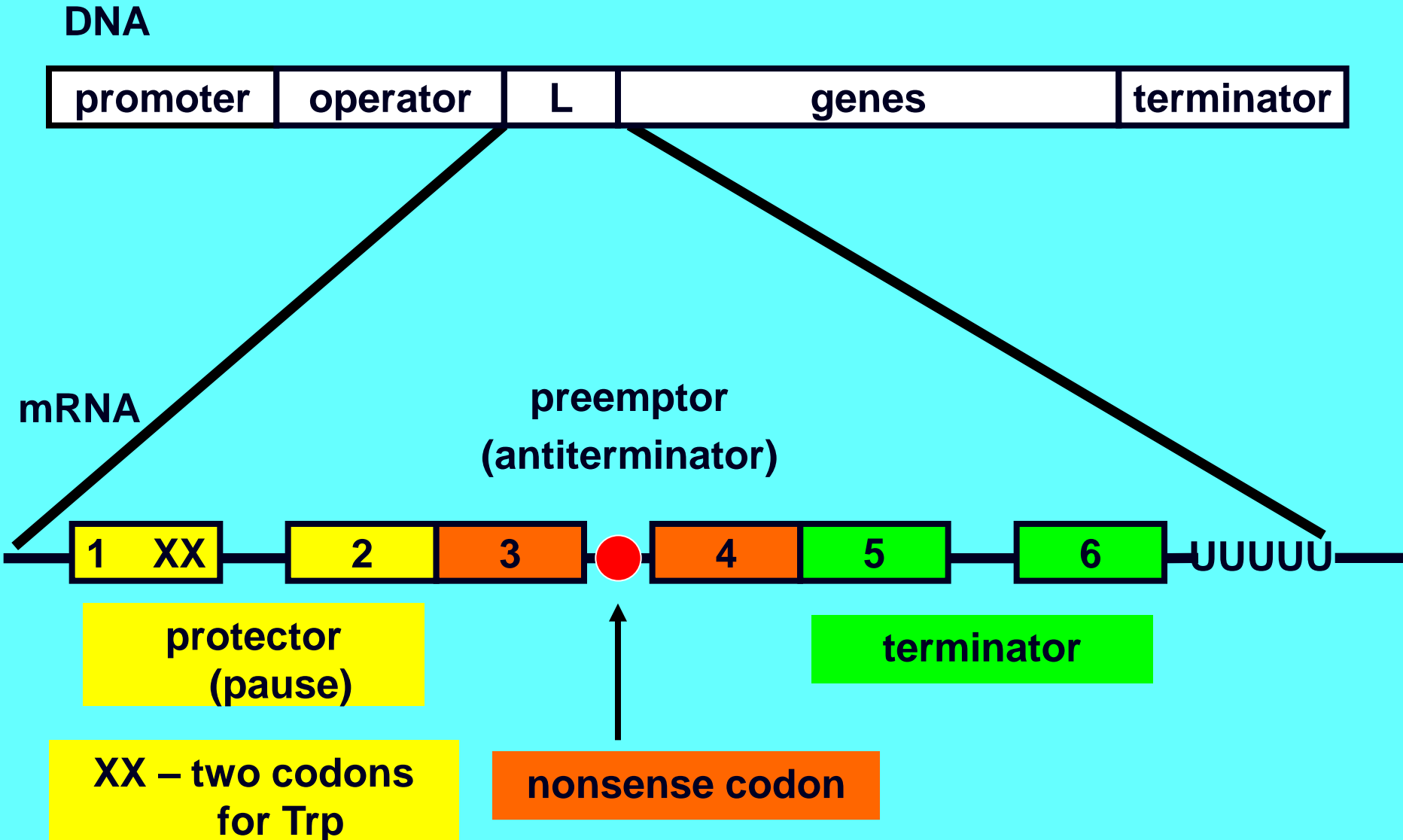
- **The process of regulation of transcription based on the attenuator**
- **Attenuator is a part of leader sequence of gene**
- **Attenuator acts as an early terminator of transcription**

**for example *trp* operon
contains genes necessary for tryptofan
synthesis**

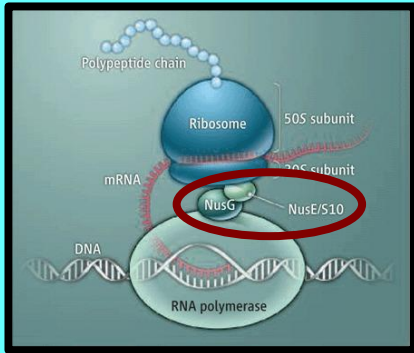
Do you still remember ?



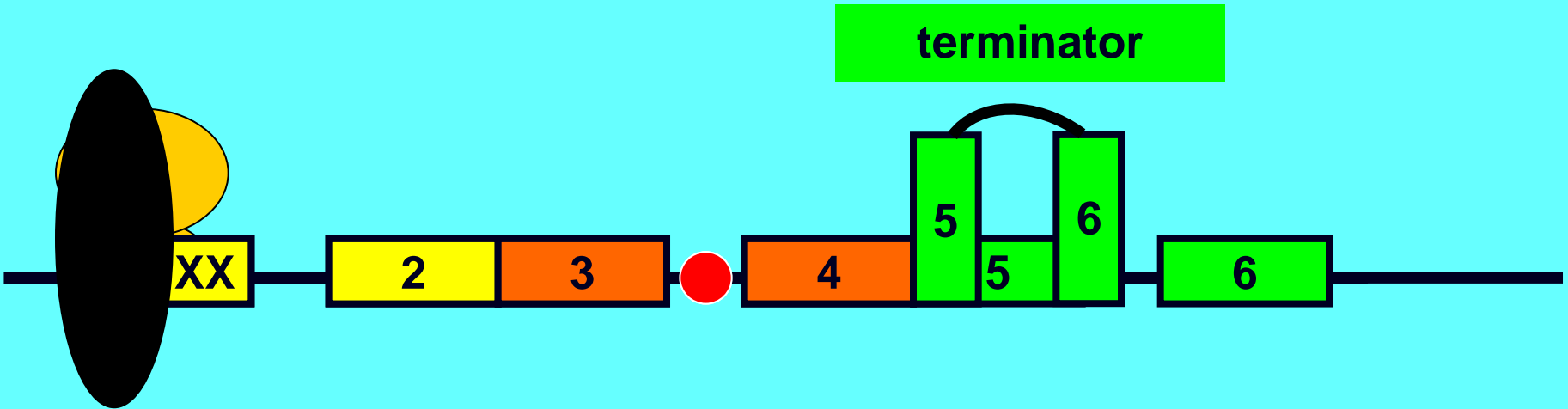
Structure of attenuator



Attenuation in redundancy of Trp



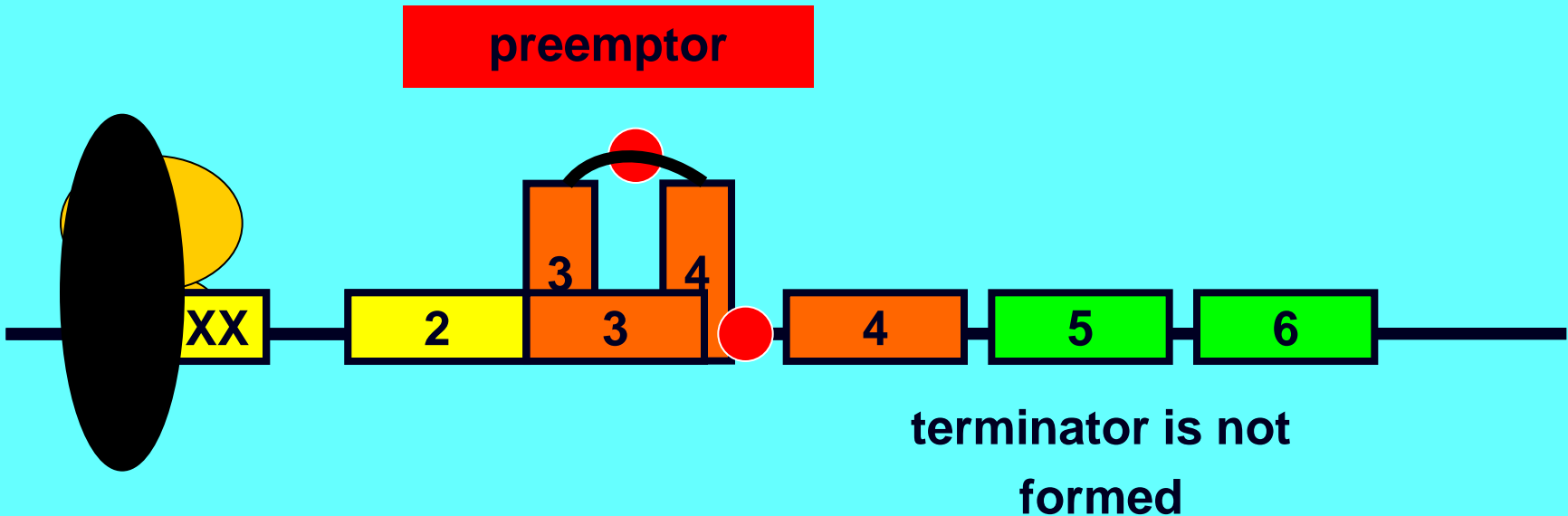
RNA polymerase
releases here



translation stops on the
nonsense codon

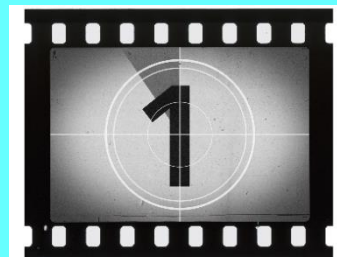
**TRANSCRIPTION IS
STOPPED**

Attenuation in deficiency of Trp



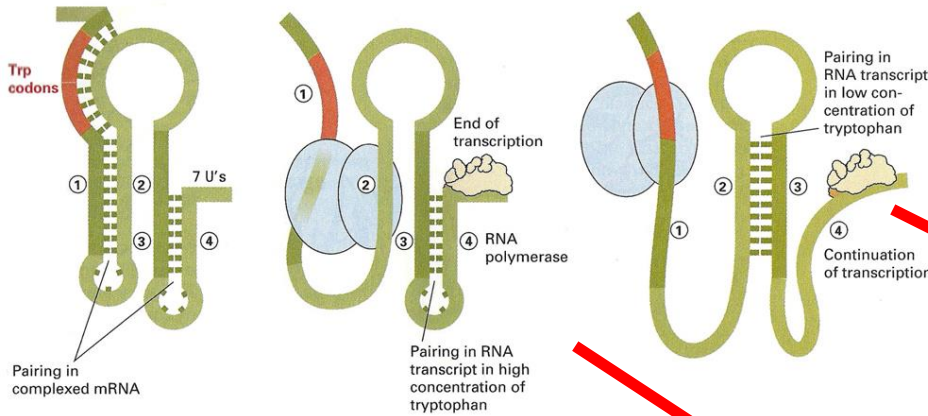
Translation stops on the codons for Trp because this amino acid is in low number of copies

**TRANSCRIPTION
FOLLOWS**

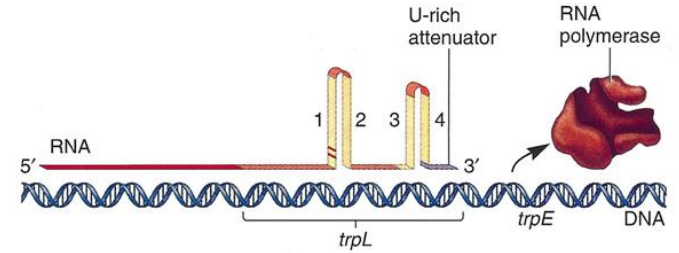


Trp attenuation

Translation of leader peptide and transcription of *trp* operon

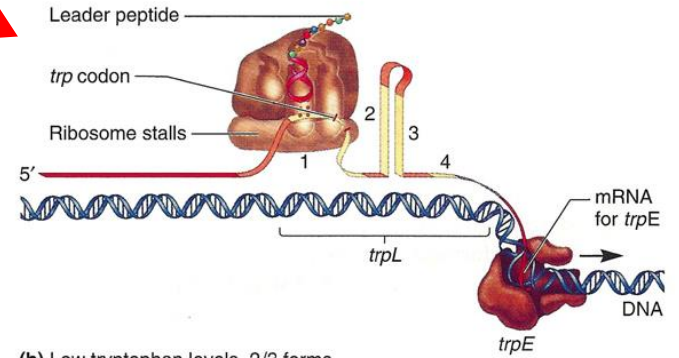


trp derepressed without cotranslation



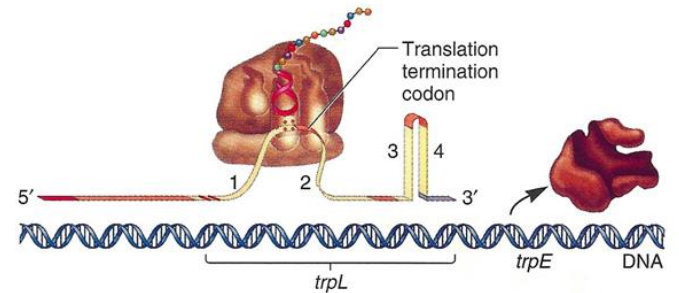
(a) No translation

trp derepressed at low tryptophan levels

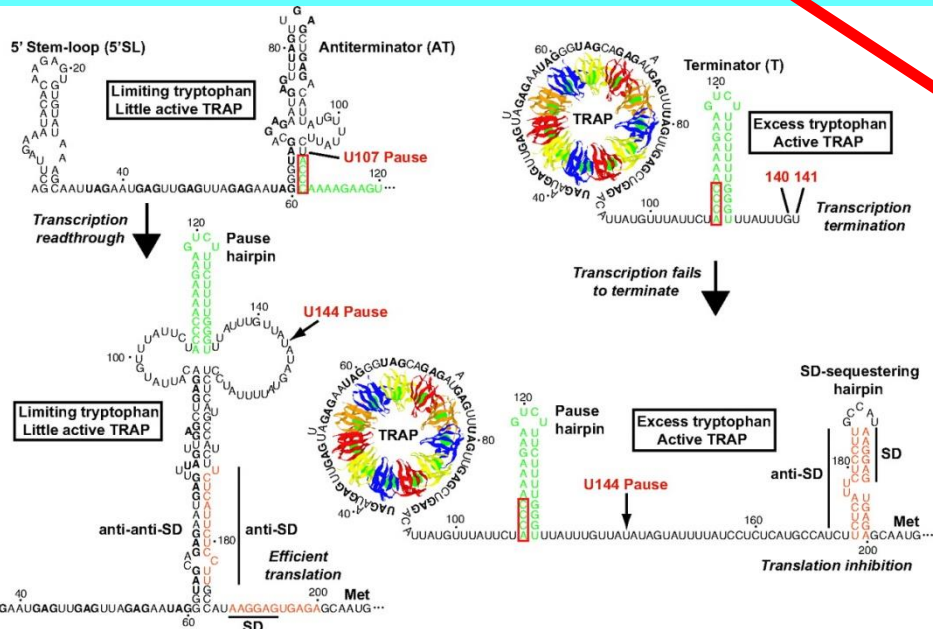


(b) Low tryptophan levels, 2/3 forms

trp derepressed at high tryptophan levels



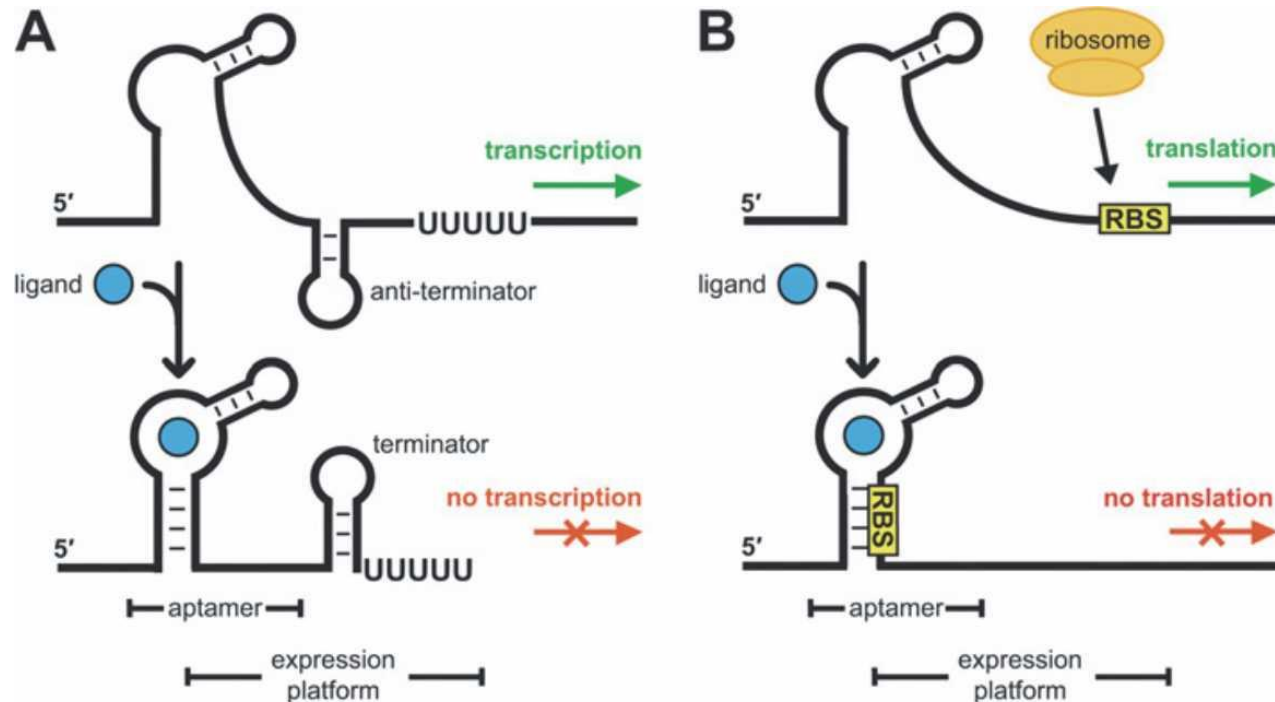
(c) High tryptophan levels, 3/4 forms



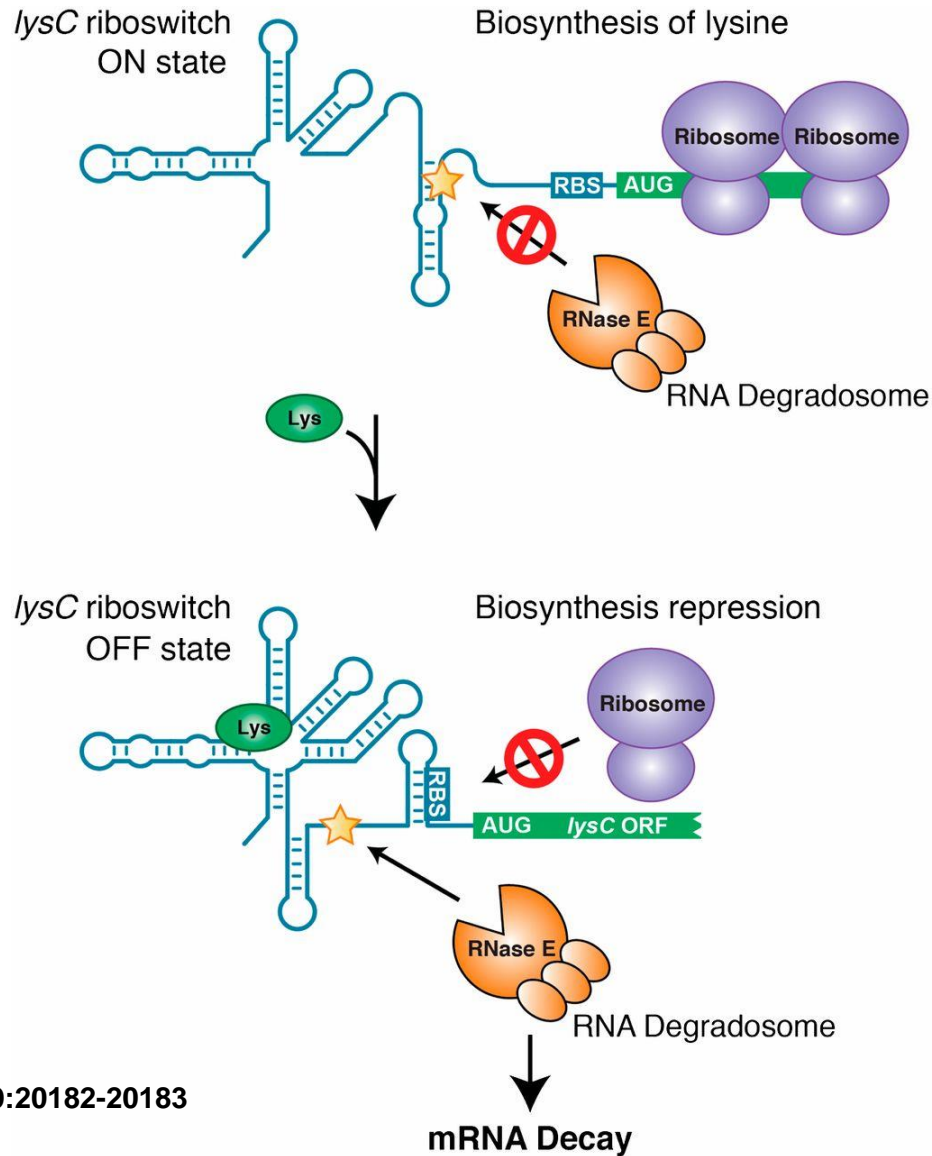
Alexander V. Yakhnin et al.
PNAS 2008;105:16131-16136

Riboswitches

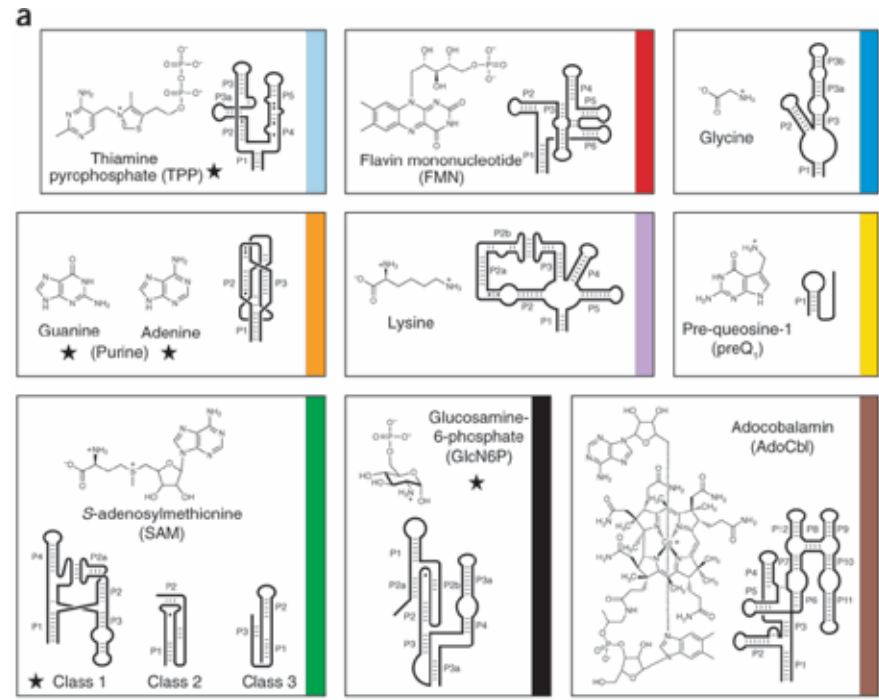
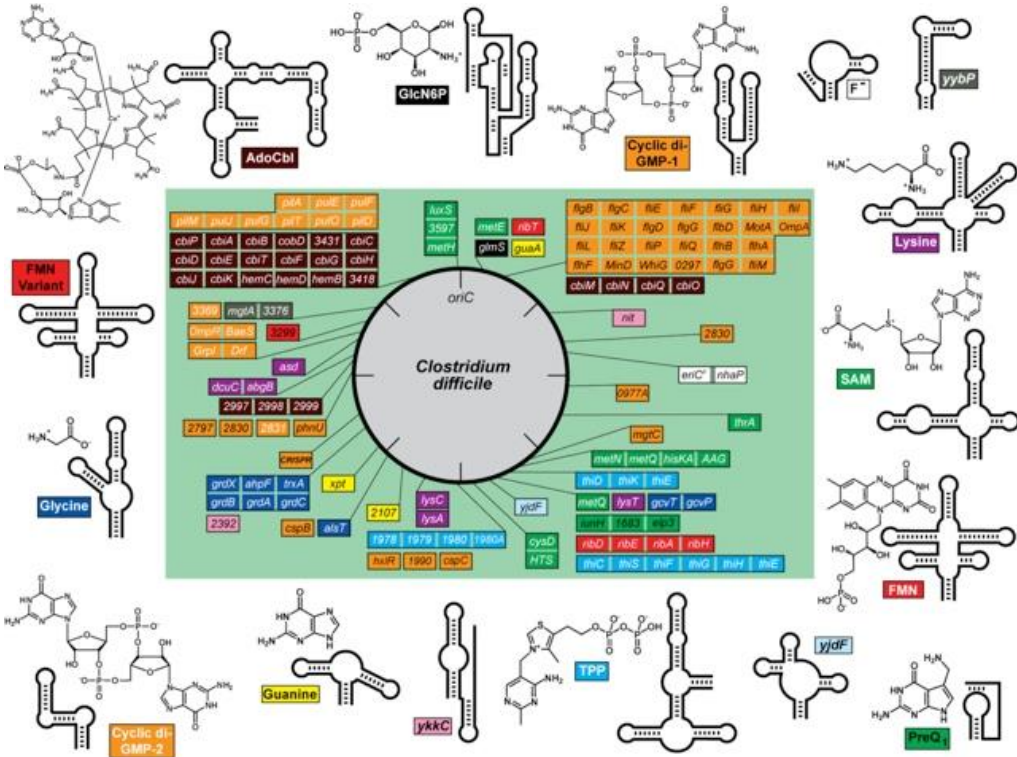
- Secondary mRNA structure capable to bind molecular effector (ligand)
- Binding of ligand on mRNA changes its secondary structure \Rightarrow the gene expression is „ON“ or „OFF“



Regulatory mechanisms of the *lysC* riboswitch controlling translation initiation and mRNA decay



Other types of riboswitches



Panel b: Distribution of riboswitch targets in various bacterial species

| Riboswitch target metabolite: | TPP | FMN | AdoCbl | Purine | SAM1 | SAM2 | SAM3 | Lysine | GlcN6P | Glycine | PreQ ₁ |
|-----------------------------------|-------|------|--------|--------|--------|------|------|--------|--------|---------|-------------------|
| <i>Acinetobacter baumannii</i> | 1(3) | 1(1) | 1(1) | - | - | - | - | - | - | 1*(1) | - |
| <i>Bacillus anthracis</i> | 6(19) | 2(5) | 1(1) | 6(9) | 17(36) | - | - | 4(4) | 1(2) | 1(1) | 2(5) |
| <i>Brucella melitensis</i> | 2(11) | 1(1) | 2(5) | - | 1(1) | - | - | - | - | 1*(3) | - |
| <i>Enterococcus faecalis</i> | 2(5) | - | 2(4) | 1(2) | - | 1(1) | - | 1(1) | 1(1) | - | 2(3) |
| <i>Escherichia coli</i> | 3(11) | 1(1) | 1(2) | - | - | - | - | 1(1) | - | - | - |
| <i>Francisella tularensis</i> | 1(1) | 1(5) | - | - | 1(1) | - | - | - | - | - | 1(1) |
| <i>Haemophilus influenzae</i> | 3(11) | 1(1) | - | - | - | - | - | 1(1) | - | 1*(1) | 1(1) |
| <i>Helicobacter pylori</i> | 1(2) | - | - | - | - | - | - | - | - | - | - |
| <i>Listeria monocytogenes</i> | 2(5) | 1(1) | 2(20) | 2(3) | 7(14) | - | - | 1(1) | 1(1) | 1(3) | 1(1) |
| <i>Mycobacterium tuberculosis</i> | 2(6) | - | 2(4) | - | - | - | - | - | - | 2*(3) | - |
| <i>Pseudomonas aeruginosa</i> | 1(1) | 1(2) | 5(24) | - | - | - | - | - | - | - | - |
| <i>Salmonella enterica</i> | 3(11) | 1(1) | 2(22) | - | - | - | - | - | - | - | - |
| <i>Staphylococcus aureus</i> | 2(7) | 2(5) | - | 1(2) | 4(6) | - | - | 2(2) | 1(1) | 1(3) | 2(4) |
| <i>Streptococcus pneumoniae</i> | 4(11) | 1(4) | - | 1(2) | - | 1(1) | - | - | - | 1(1) | - |
| <i>Vibrio cholerae</i> | 2(7) | 1(1) | 1(1) | - | - | - | - | 3(3) | - | 1(1) | - |
| <i>Yersinia pestis</i> | 3(8) | 1(1) | 1(2) | - | - | - | - | - | - | - | - |

Heat shock proteins

- **Proteins which are induced by higher temperature**
- **They are expressed by higher speed in the temperatures 42-45°C**
- **Regulated by sigma factor sigma-32, which binds to other promoters than the standard sigma factor**
- **Auto regulation by feedback loop = after accumulation the induced protein binds to sigma factor and releases it from RNA polymerase**

Regulation the genes for rRNA and tRNA

- **Speedy of cell division is connected to speedy of ribosome formation**
- **Synthesis of rRNA and tRNA is TAKE OFF if level of amino acid pool is low, synthesis of mRNA continues at least minimum amount**