

# Molecular Biology II

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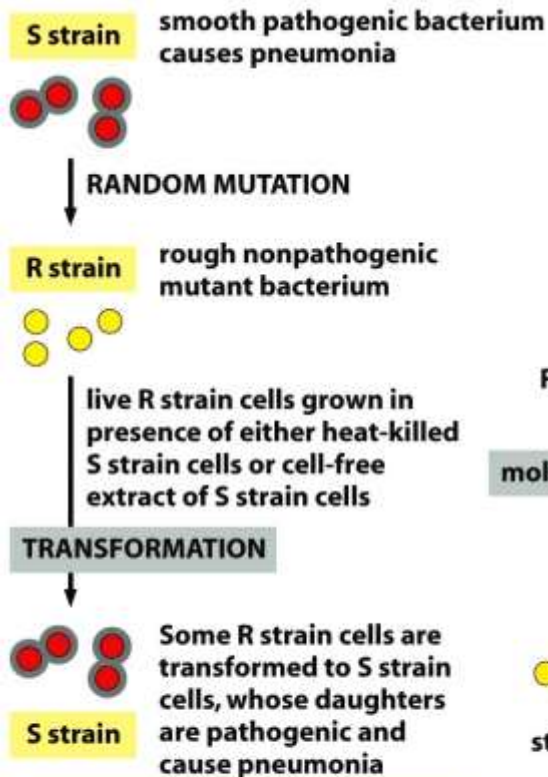
[petrmpharm.muni.cz,](mailto:petrmpharm.muni.cz)

# The structure and function of biopolymers during the transitions of genetic information

Daniel Renčiuk

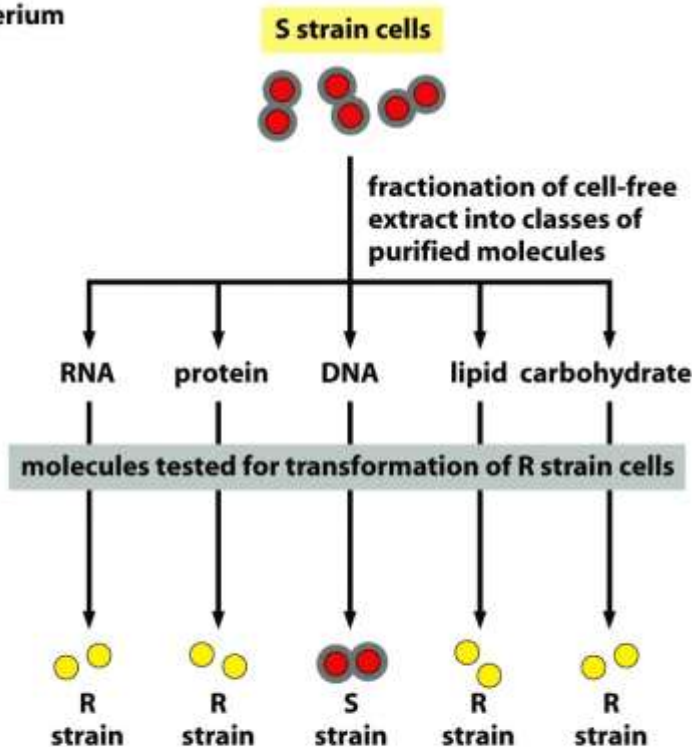
# Genetic information is coded by DNA

The experiment combining two strains of *Streptococcus pneumoniae* bacteria.



**CONCLUSION:** Molecules that can carry heritable information are present in S strain cells.

(A)



**CONCLUSION:** The molecule that carries the heritable information is DNA.

(B)

O. Avery  
C. MacLeod  
M. McCarty

STUDIES ON THE CHEMICAL NATURE OF THE SUBSTANCE INDUCING TRANSFORMATION OF PNEUMOCOCCAL TYPES

INDUCTION OF TRANSFORMATION BY A DEOXYRIBONUCLEIC ACID FRACTION ISOLATED FROM PNEUMOCOCCUS TYPE III

By OSWALD T. AVERY, M.D., COLIN M. MACLEOD, M.D., and MACLYN McCARTY,\* M.D.

(From the Hospital of The Rockefeller Institute for Medical Research)

PLATE I

(Received for publication, November 1, 1943)

Biologists have long attempted by chemical means to induce in higher organisms predictable and specific changes which thereafter could be transmitted in series as hereditary characters. Among microorganisms the most striking example of inheritable and specific alterations in cell structure and function that can be experimentally induced and are reproducible under well defined and adequately controlled conditions is the transformation of specific types of *Pneumococcus*. This phenomenon was first described by Griffith (1) who succeeded in transforming an attenuated and non-encapsulated (R) variant derived from one specific type into fully encapsulated and virulent (S) cells of a heterologous specific type. A typical instance will suffice to illustrate the techniques originally used and serve to indicate the wide variety of transformations that are possible within the limits of this bacterial species.



# Roles of genetic material

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**Genotype** – storage of genetic information and its transition to the offspring

**Phenotype** – expression of genetic information to particular properties of an individual

**Evolutionary role** – adaptation of an organism/species to the environment through the changes in genetic information

# Terminology

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**Gene** – several “definitions” depending on the point of view:

classic genetics (Mendel) – elementary unit of hereditary genetic information

molecular genetics – part of DNA coding for RNA (and as a consequence coding for some property of the individual)

structural genes coding for mRNA/protein (+ regulatory regions)

genes coding for functional RNA (miRNA, ...)

strict – structural gene – part of DNA that codes for protein sequence

**Allele** – particular variant of the respective gene

**Genome** – complete DNA of organism (molecular) x complete genetic information of organism x sum of genes (classic)

**Genotype** – the combination of particular alleles of all genes in individual

**Phenotype** – the sum of actual individual properties (as a result of expression of particular genotype in the respective environment)

**Genophore** – the carrier of genetic information, usually a molecule of DNA (often used for bacteria)

# Information biopolymers

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## **Deoxyribonucleic acid (DNA)**

- linear heteropolymer composed from 2-deoxyribonucleotides connected by phosphodiester bonds
- usually as a stable and resistant double helix
- serves as a storage of genetic information, as a template for its reproduction (replication) and as a template for the expression of genetic information to the phenotype (transcription)

## **Ribonucleic acid (RNA)**

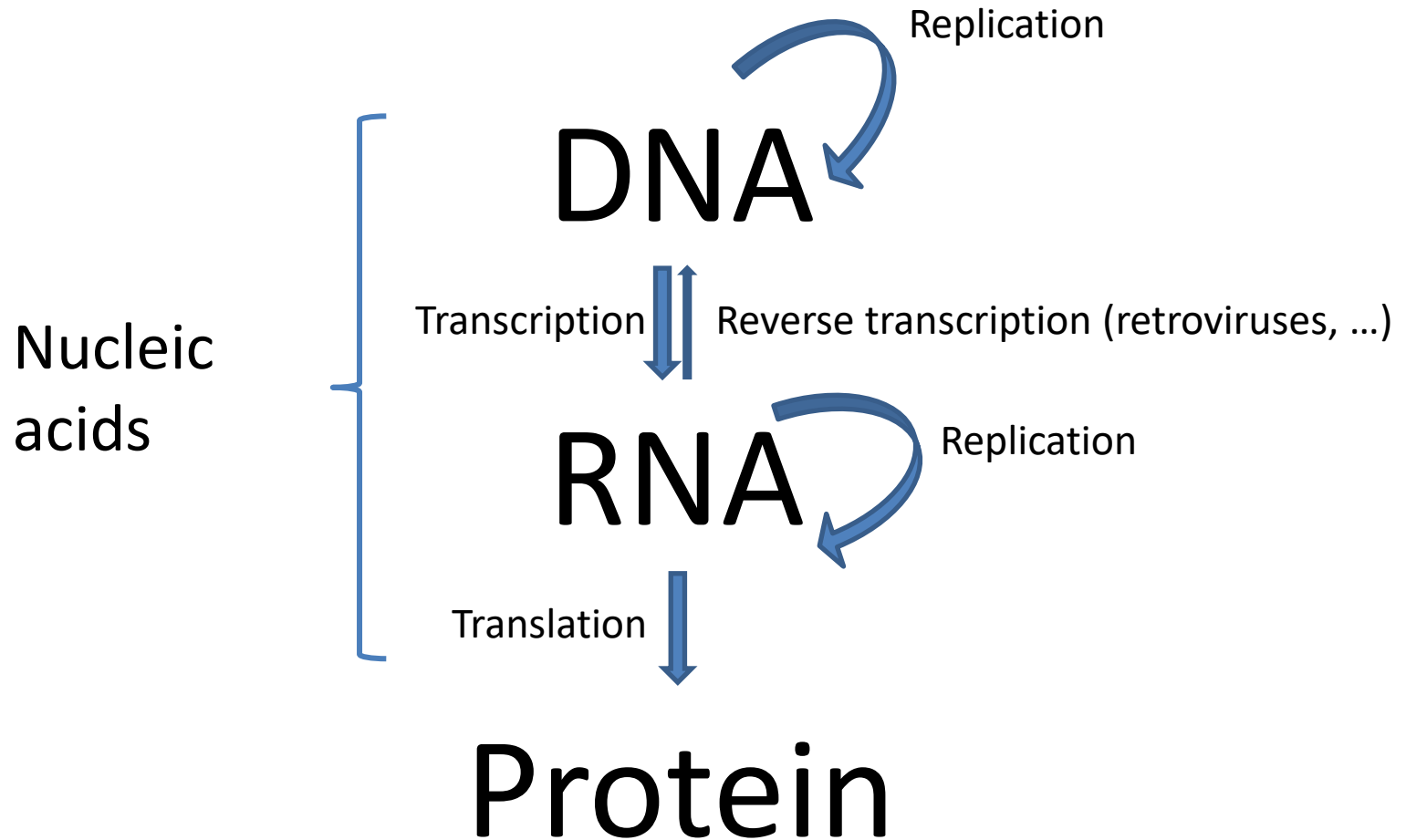
- linear heteropolymer composed from ribonucleotides connected by phosphodiester bonds
- usually as a single-stranded structure of variable length, structure and reactivity
- many functions depending on type of RNA

## **Protein**

- linear heteropolymer composed from 20 (21) amino acids connected by peptide bonds
- highly variable structures, properties and functions

# Central dogma of molecular biology

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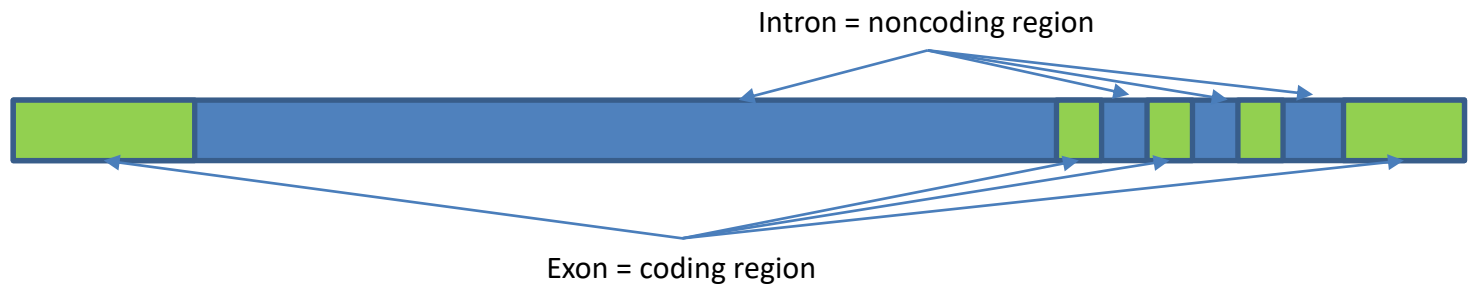




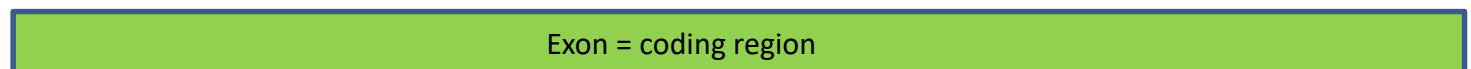
# Genes

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Most **eukaryotic** genes contain **introns**, that are transcribed into primary RNA transcript and introns are consecutively removed by **splicing** process on **spliceosome** to form final **mRNA**.



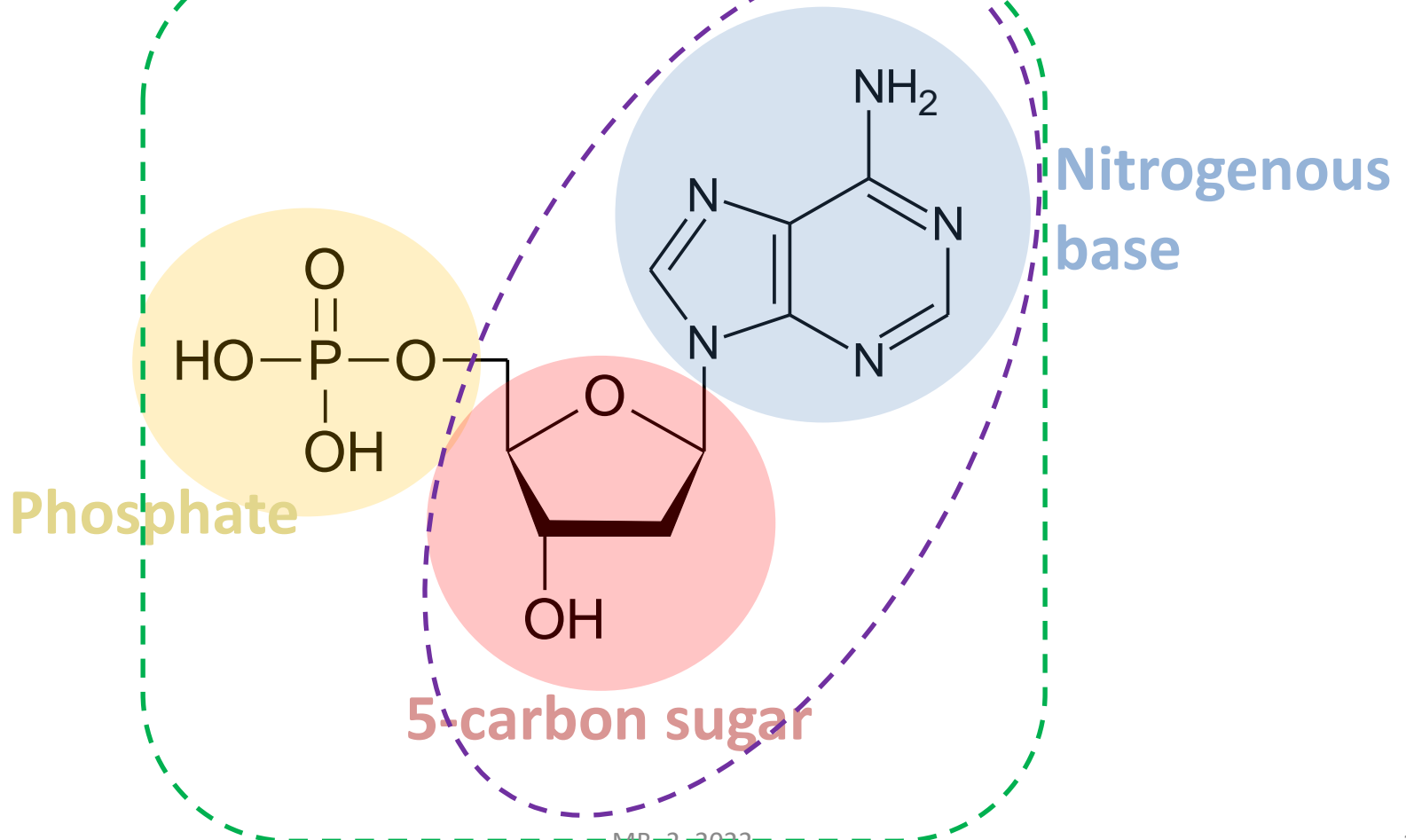
**Prokaryotic** genes do not contain **introns** and they are directly transcribed into mRNA that serves as a template for translation into protein.



# Nucleic acids

**NUCLEOTIDE**

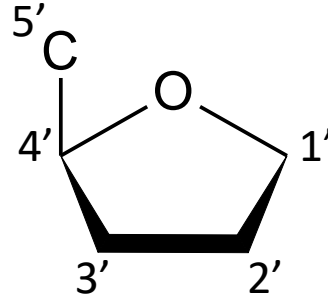
**NUCLEOSIDE**



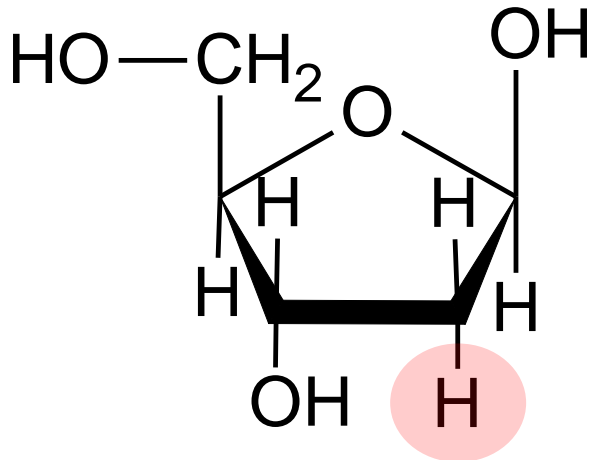
# 5-carbon sugar - pentose

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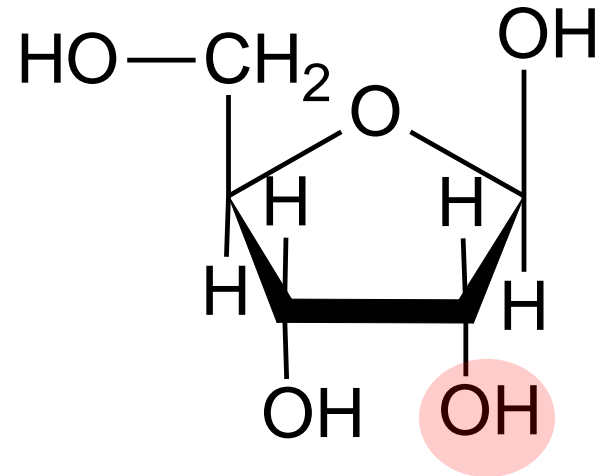
DNA



RNA



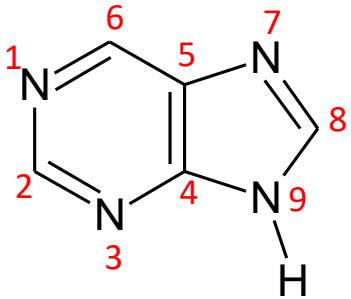
$\beta$ -D-2-deoxyribose



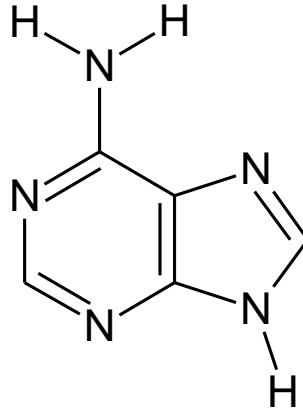
$\beta$ -D-ribose

# Base

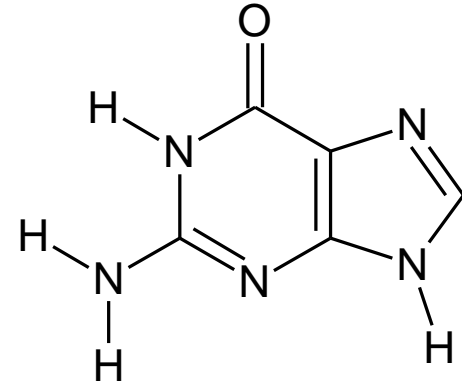
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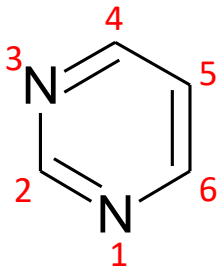
Purine



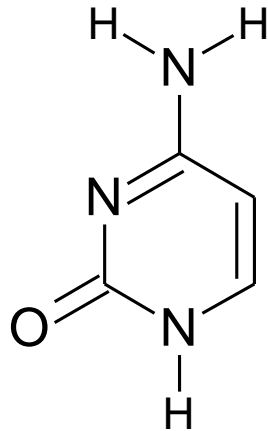
Adenine



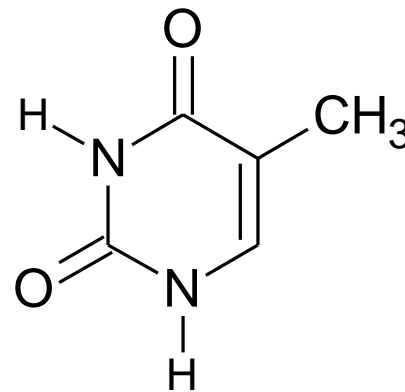
Guanine



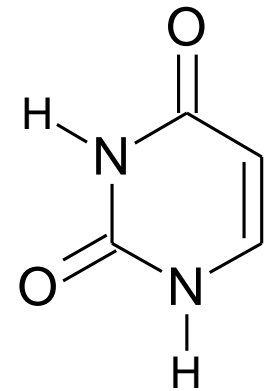
Pyrimidine



Cytosine



Thymine



Uracil

# Nucleic acid nomenclature

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	Base	Nucleoside	Nucleotide NTP
G	Guanine	Guanosine	Guanosine triphosphate
A	Adenine	Adenosine	Adenosine triphosphate
T	Thymine	Thymidine	Thymidine triphosphate
C	Cytosine	Cytidine	Cytidine triphosphate
U	Uracil	Uridine	Uridine triphosphate

DNA – prefix deoxy-

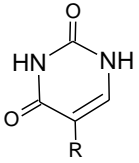
NMP = monophosphate

NDP = diphosphate

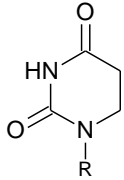
# Modified bases

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## tRNA

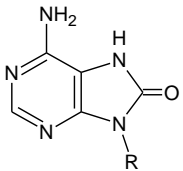


pseudouracil

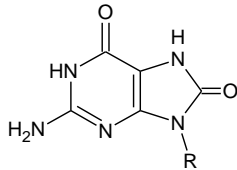


dihydrouracil

## Oxidative damage

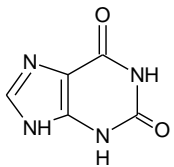


8-oxo adenine

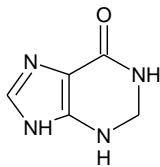


8-oxo guanine

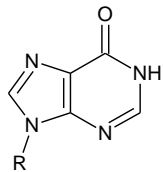
## Metabolism



xanthine

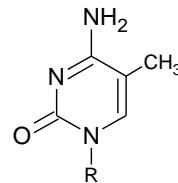


hypoxanthine

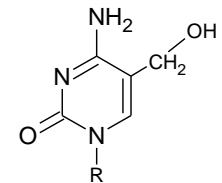


inosine

## Epigenetics



5-methyl cytosine



5-hydroxymethyl cytosine

# Conformation of N-glycosidic bond

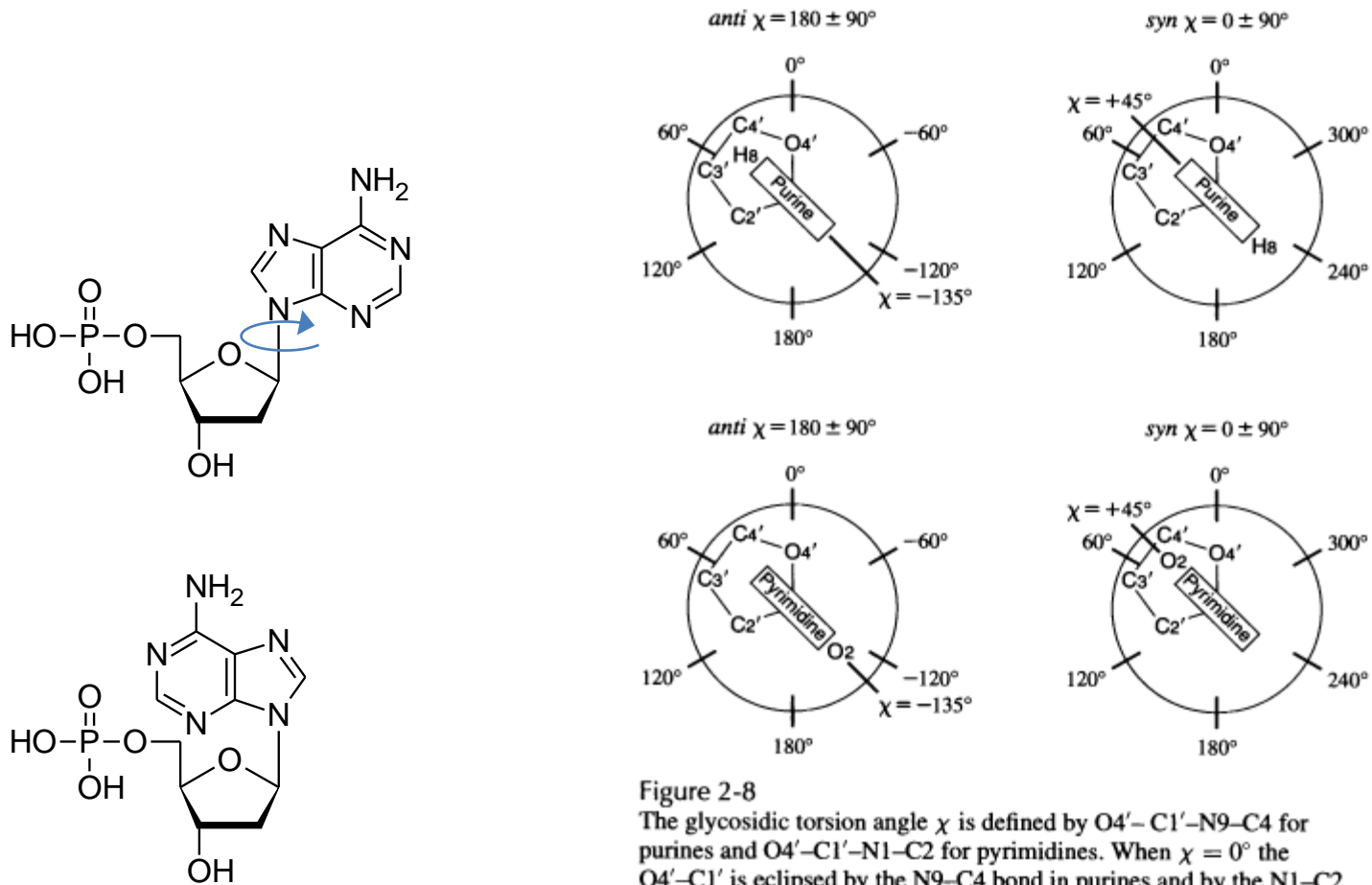
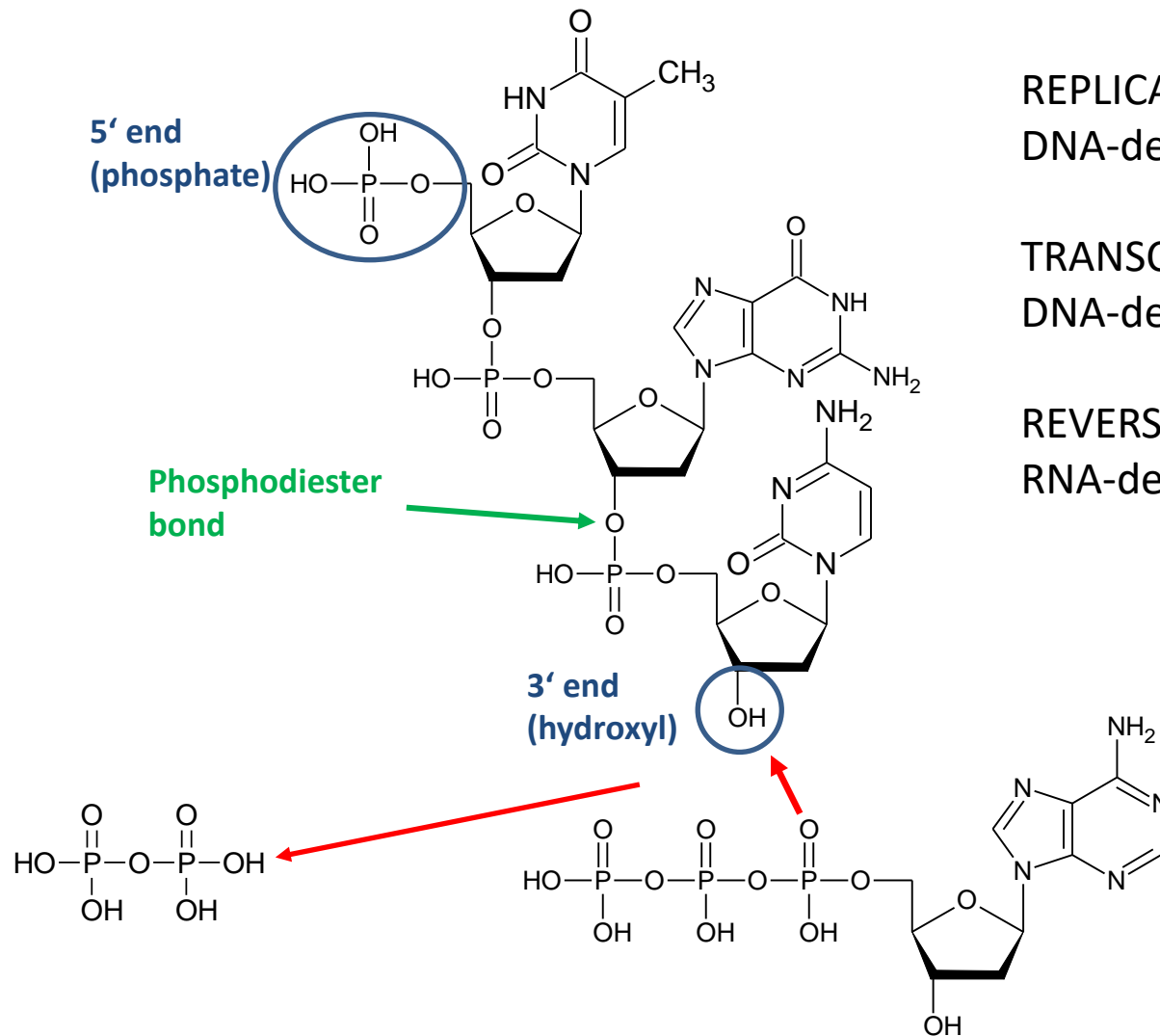


Figure 2-8

The glycosidic torsion angle  $\chi$  is defined by O4'–C1'–N9–C4 for purines and O4'–C1'–N1–C2 for pyrimidines. When  $\chi = 0^\circ$  the O4'–C1' is eclipsed by the N9–C4 bond in purines and by the N1–C2 bond in pyrimidines. The syn conformations correspond to  $0^\circ \pm 90^\circ$ ; anti conformations correspond to  $180^\circ \pm 90^\circ$ . In nucleotides steric hindrance limits the conformations actually found to a much narrower range of angles that depend on sugar pucker and base. The syn conformations are usually found with  $\chi = 45^\circ \pm 45^\circ$ ; anti conformations are usually found with  $\chi = -135^\circ \pm 45^\circ$ .



# Formation of sugar-phosphate backbone



REPLICATION

DNA-dependent DNA polymerase

TRANSCRIPTION

DNA-dependent RNA polymerase

REVERSE TRANSCRIPTION

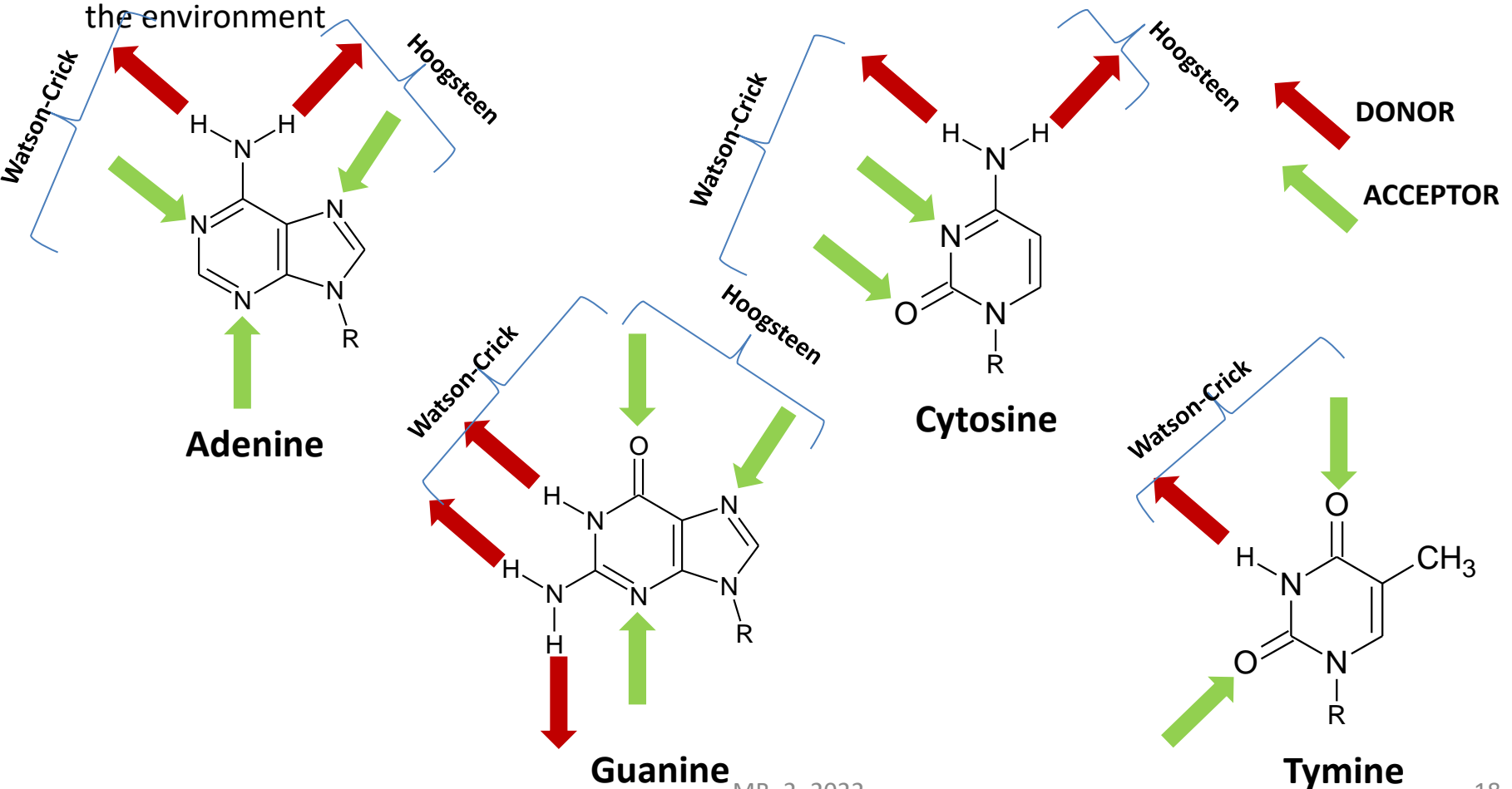
RNA-dependent DNA polymerase

# Base reactivity – hydrogen bonds

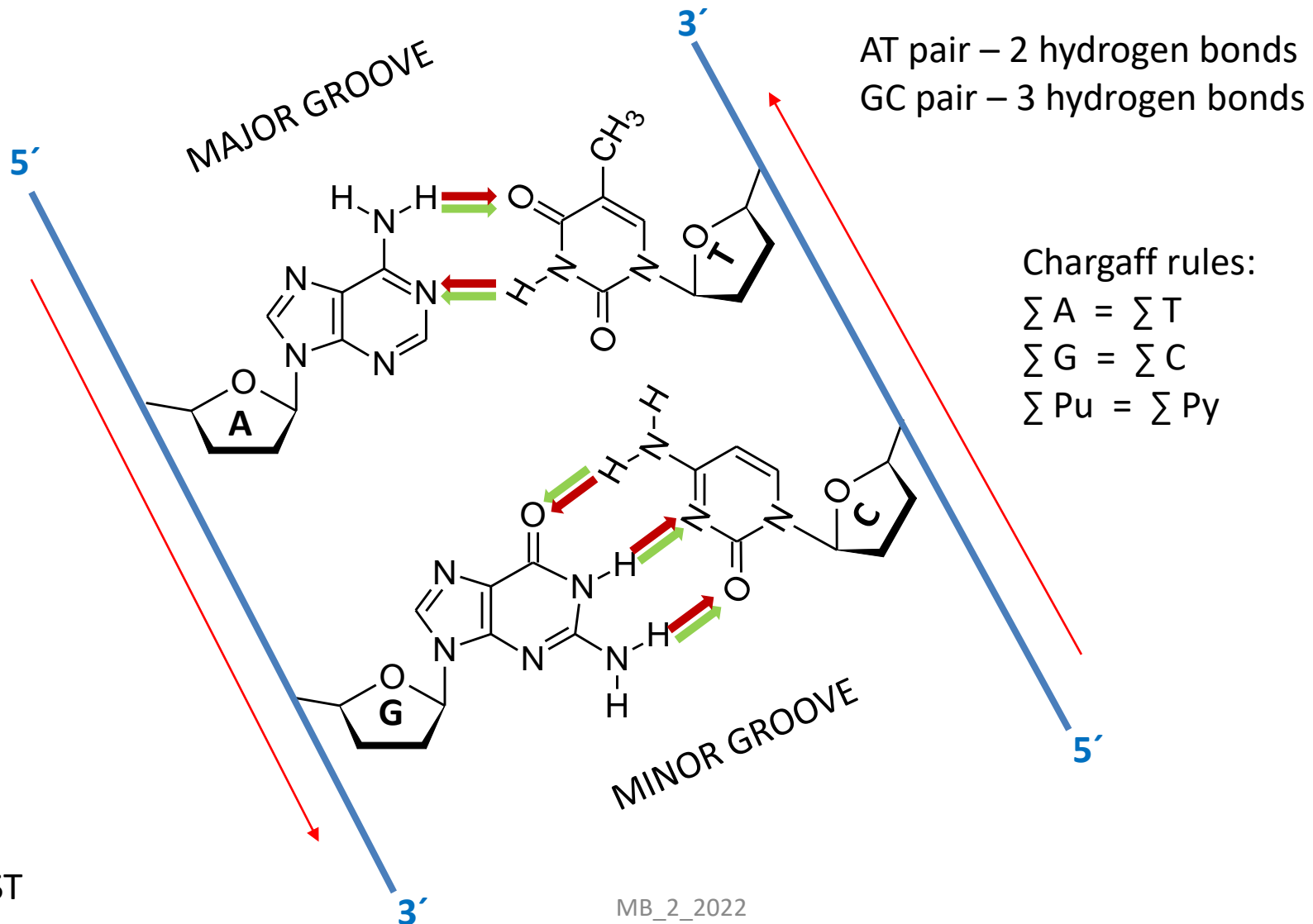
**Hydrogen bond** – weak electrostatic interaction of two polar groups – one covalently binds hydrogen (DONOR – usually -N-H or -O-H); the second (ACCEPTOR) is usually N or O

Length:  $\sim 2.8 \text{ \AA}$  (2 - 3,4  $\text{ \AA}$ )

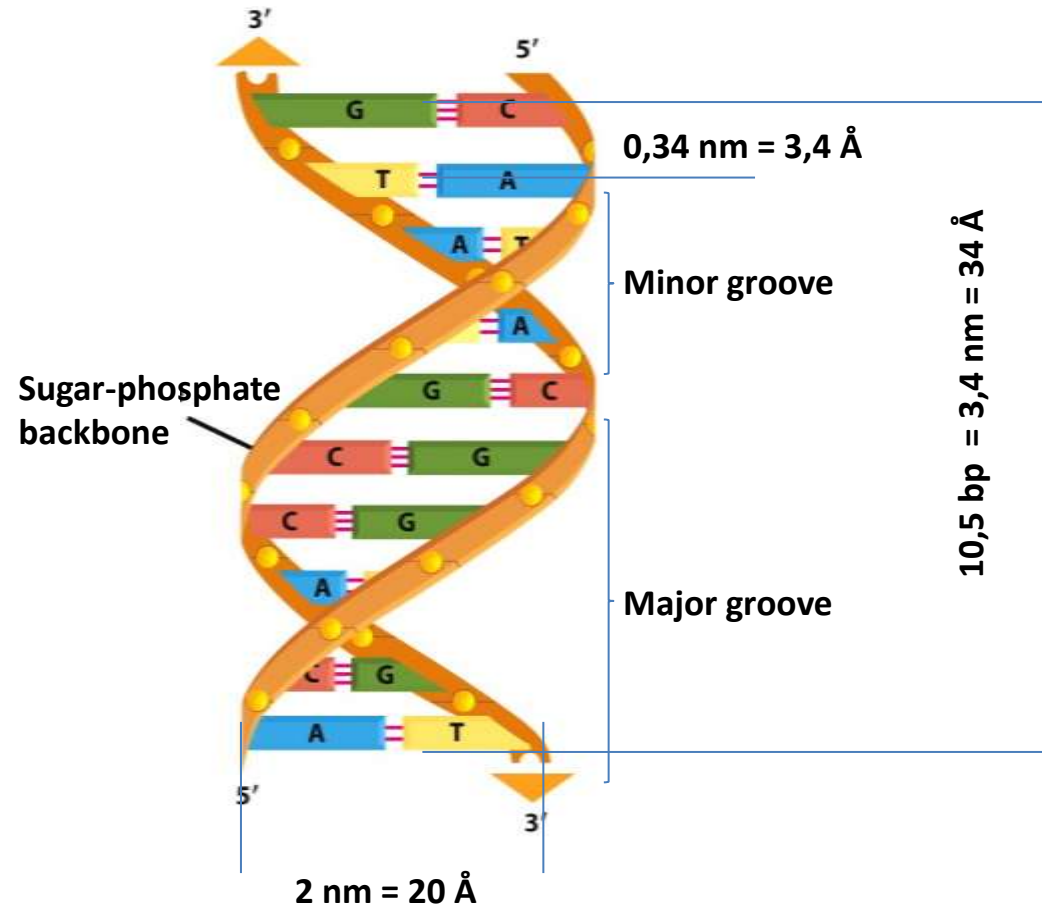
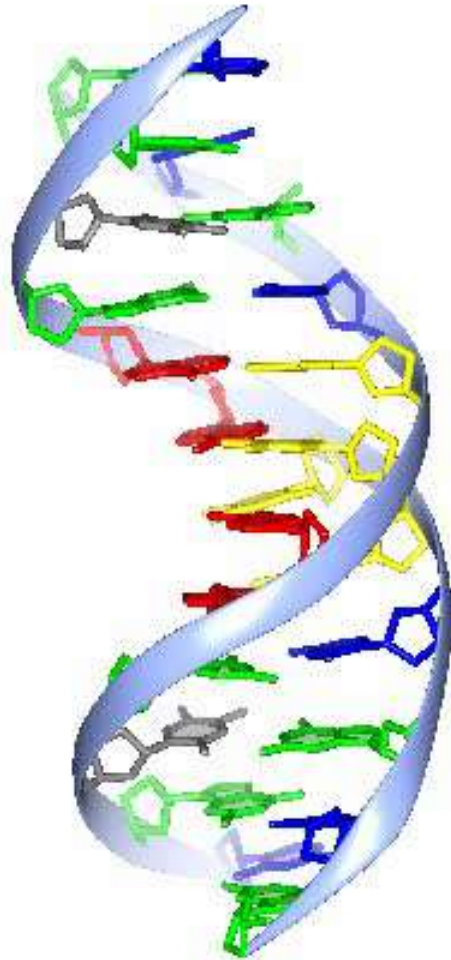
Energy:  $< 1 \text{ kcal/mol}$  both depend on particular atoms and on the environment



# Watson-Crick base pairing



# DNA double helix



# DNA double helix

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- **two** molecules (**strands**) of DNA
- the helix is **right-handed**
- the strands are **antiparallel** – their 5′-3′ direction is opposite in the context of double helix direction
- similar content of purines and pyrimidines; content of A = T, G = C (Chargaff rules)
- result – the strands are **complementary** – i.e. according to the Watson-Crick base pairing rules we can predict the sequence of one strand according to the sequence of the other
- on average the double helix contains **10,5 base pairs** per turn of the helix, which is about **3,4 nm** in length

# DNA structure – Watson and Crick model



F. Crick

J.D. Watson



M. Wilkins



R. Franklin



E. Chargaff

April 25, 1953 NATURE 737

equipment, and to Dr. O. E. J. Drenth and the captain and officers of H.M.S. *Discovery II* for their part in making the observations.

\*Dunn, P. W., *Invent.*, 11, and *Science*, 70, 143 (1951).

\*Langer, August, M. S., *Ann. N.Y. Acad. Sci.*, London, 1952, 4, 146 (1952).

\*Yon, J. W. S., *Weeks 50th Papers in Paper Science*, Dover, 11 (1952).

\*Langer, A. W., *19th. Nat. Jour.*, *Proc. (London)*, 1111 (1953).

## MOLECULAR STRUCTURE OF NUCLEIC ACIDS

### A Structure for Deoxyribonucleic Acid

WE wish to suggest a structure for the salt of deoxyribonucleic acid (D.N.A.). This structure has novel features which are of considerable biological interest.

A structure for nucleic acid has already been proposed by Pauling and Corey<sup>1</sup>. They kindly made their manuscript available to us in advance of publication. Their model consists of three intertwined chains, with the phosphates near the fibre axis, and the bases on the outside. In our opinion, this structure is unsatisfactory for two reasons: (1) We believe that the material which gives the X-ray diagram is the acid, not the free acid. Without the acidic hydrogen atoms it is not clear what forces would hold the structure together, especially as the negatively charged phosphates near the axis will repel each other. (2) Some of the van der Waals distances appear to be too small.

Another three-chain structure has also been suggested by Foweraker (in the press). In his model the phosphates are on the outside and the bases on the inside, linked together by hydrogen bonds. This structure as described is rather ill-defined, and for this reason we shall not comment on it.

We wish to put forward a radically different structure for the salt of deoxyribonucleic acid. This structure has two helical chains each wound round the same axis (see diagram). We have made the usual chemical assumptions, namely, that each chain consists of phosphate diester groups joining β-D-deoxythiobarbituric residues with 3',5' linkages. The two chains (but not their bases) are related by a dyad perpendicular to the fibre axis. Both chains follow right-handed helices, but owing to the dyad the sequence of the atoms in the two chains run in opposite directions. Each chain loosely resembles Purberg's model No. 1; that is, the bases are on the inside of the helix and the phosphates on the outside. The configuration of the sugar and the atoms near it is close to Purberg's 'standard configuration', the sugar being roughly perpendicular to the attached base. There is a residue on each chain every 3.4 Å. In the 2-direction. We have assumed an angle of 36° between adjacent residues in the same chain, so that the structure repeats after 10 residues on each chain, that is, after 34 Å. The distance of a phosphate atom from the fibre axis is 10 Å. As the ribonucleotides are on the outside, esters have

The structure is in no way a rather high. At low angles the bases will become more compact.

The novel features of it in which the two chains are perpendicular to the together in pairs, a single hydrogen-bonded to a chain, so that the two 2-co-ordinates. One of the other a pyrimidine hydrogen, bonds are made 1 to pyrimidine position 4.

If it is assumed that structure in the model (that is, with the later figures) it is found bases can bond together (purine) with thymine (purine) with cytosine (In other words, if so a pair, on either chain, then on these assumptions the other member must be thymine; similarly for guanine and cytosine. The sequence of bases on a single chain does not appear to be restricted in any way. However, if only specific pairs of bases can be formed, it follows that if the sequence of bases on one chain is given, then the sequence on the other chain is automatically determined.

It has been found experimentally<sup>2</sup> that the ratio of guanine to cytosine, are always very close to unity for deoxyribonucleic acid.

It is probably impossible to build this structure with a ribose sugar in place of the deoxyribose, as the extra oxygen atom would make too close a van der Waals contact.

The previously published X-ray data<sup>3,4</sup> on deoxyribonucleic acid are insufficient for a rigorous test of our structure. So far as we can tell, it is roughly compatible with the experimental data, but it must be regarded as unproved until it has been checked against more exact results. Some of these are given in the following communications. We were not aware of the details of the results presented there when we devised our structure, which rests mainly though not entirely on published experimental data and stereochemical arguments.

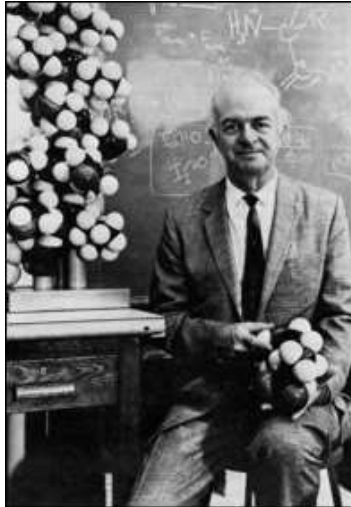
It has not escaped our notice that the specific pairing we have postulated immediately suggests a possible copying mechanism for the genetic material.

Full details of the structure, including the conditions assumed in building it, together with a set of co-ordinates for the atoms, will be published elsewhere.

We are much indebted to Dr. Jerry Donohue for constant advice and criticism, especially on inter-atomic distances. We have also been stimulated by a knowledge of the general nature of the unpublished experimental results and ideas of Dr. M. H. F. Wilkins, Dr. R. B. Franklin and their co-workers at

Franklin's X-ray photograph shows DNA's 'B'-form (1952)

# DNA structure – Pauling model



Linus Pauling

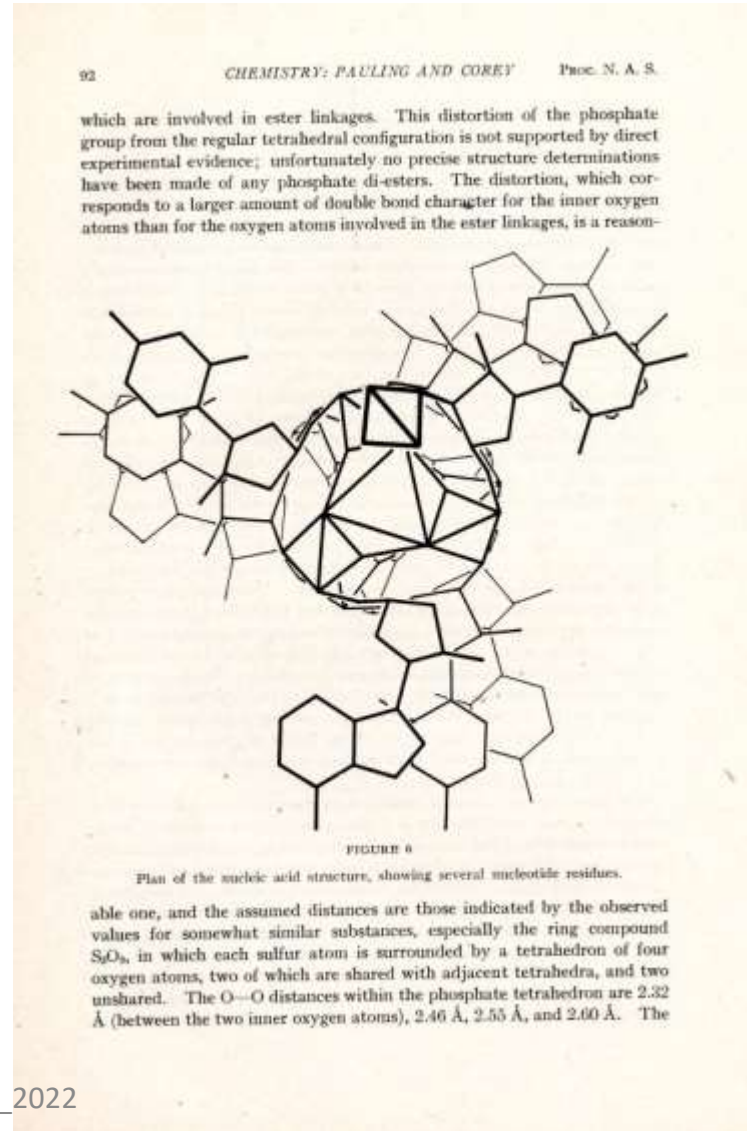
## *A PROPOSED STRUCTURE FOR THE NUCLEIC ACIDS*

BY LINUS PAULING AND ROBERT B. COREY

GATES AND CRELLIN LABORATORIES OF CHEMISTRY,\* CALIFORNIA INSTITUTE OF TECHNOLOGY

Communicated December 31, 1952

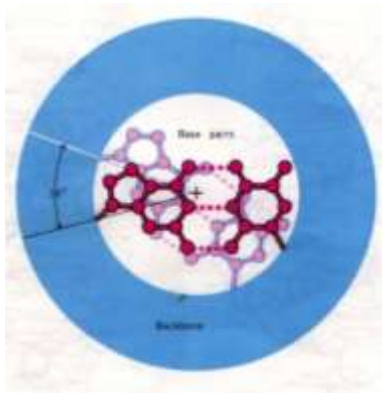
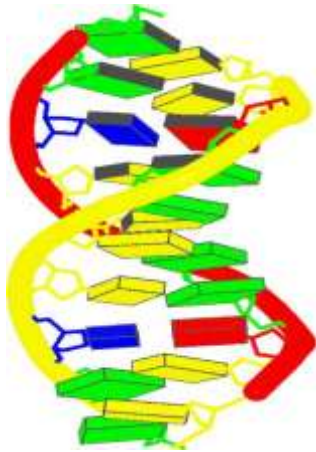
The nucleic acids, as constituents of living organisms, are comparable in importance to the proteins. There is evidence that they are involved in the processes of cell division and growth, that they participate in the transmission of hereditary characters, and that they are important constituents of viruses. An understanding of the molecular structure of the nucleic acids should be of value in the effort to understand the fundamental phenomena of life.



# Various types of double helix

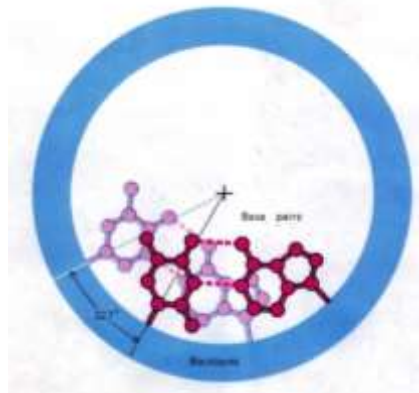
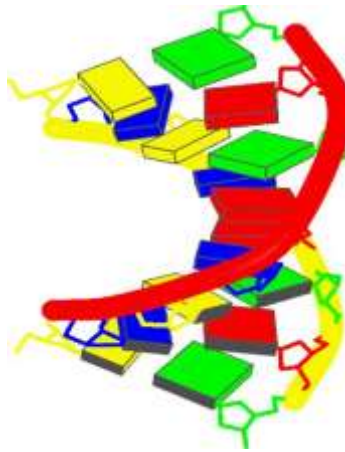
## B-DNA

- DNA in water/salt solutions



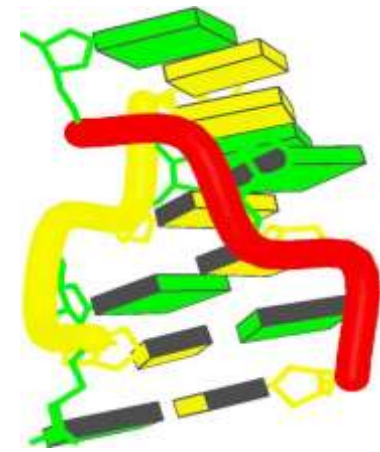
## A-DNA

- DNA in crowding solutions
- RNA



## Z-DNA

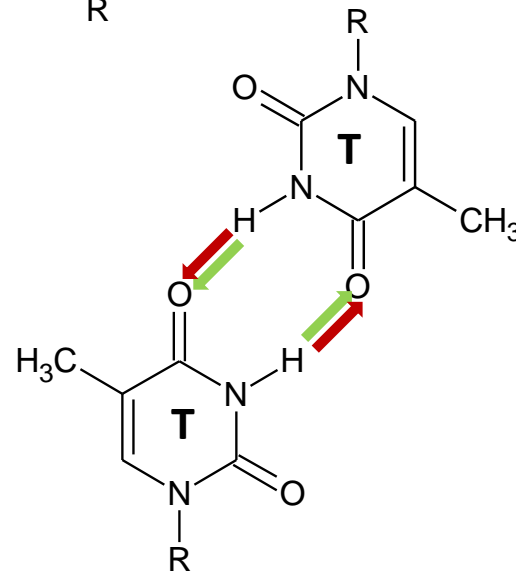
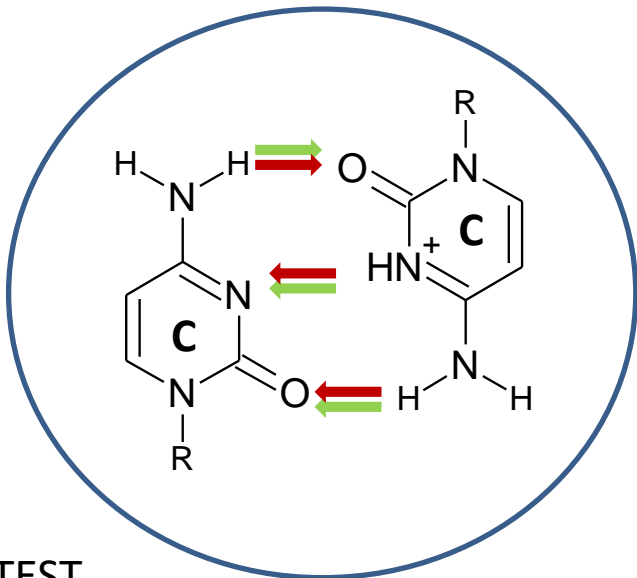
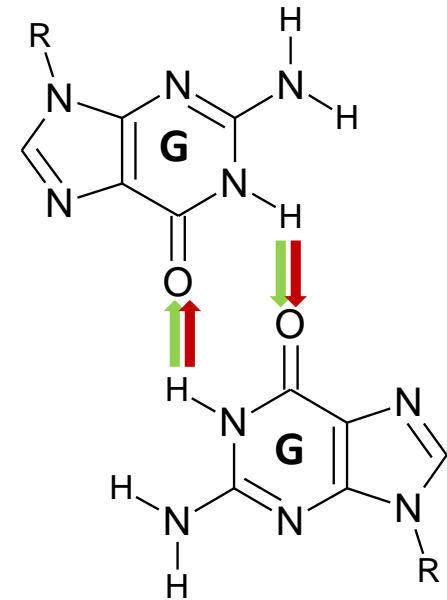
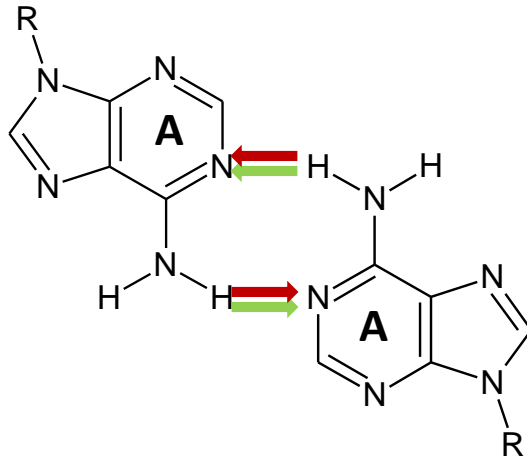
- CpG sequences in crowding conditions



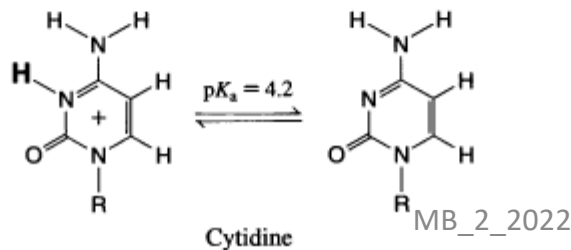
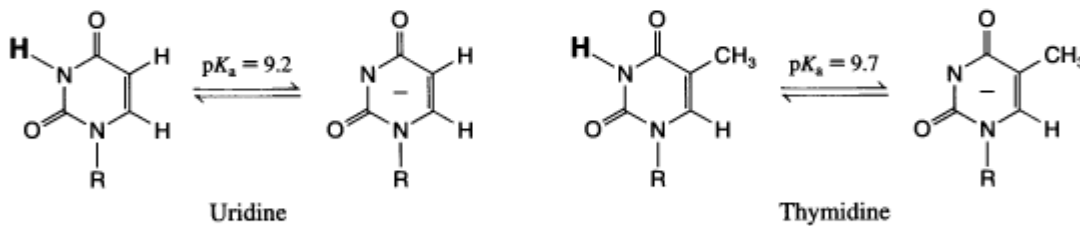
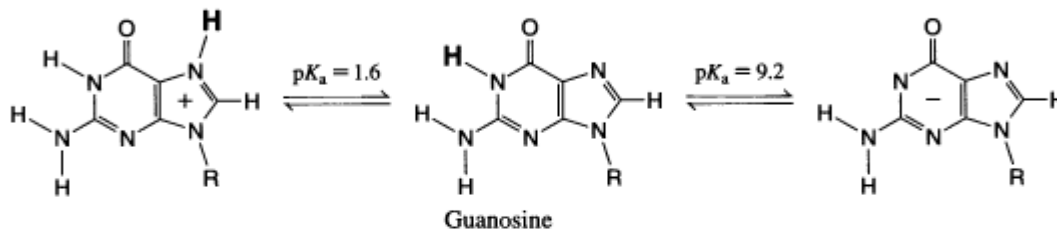
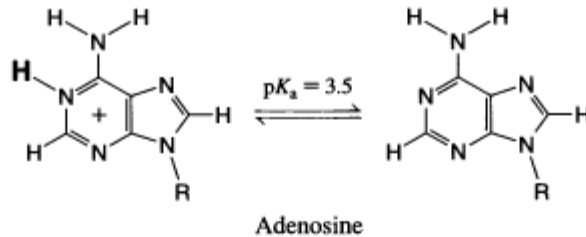
Left handed  
Zig-zag step



# Reversed Watson-Crick pairing



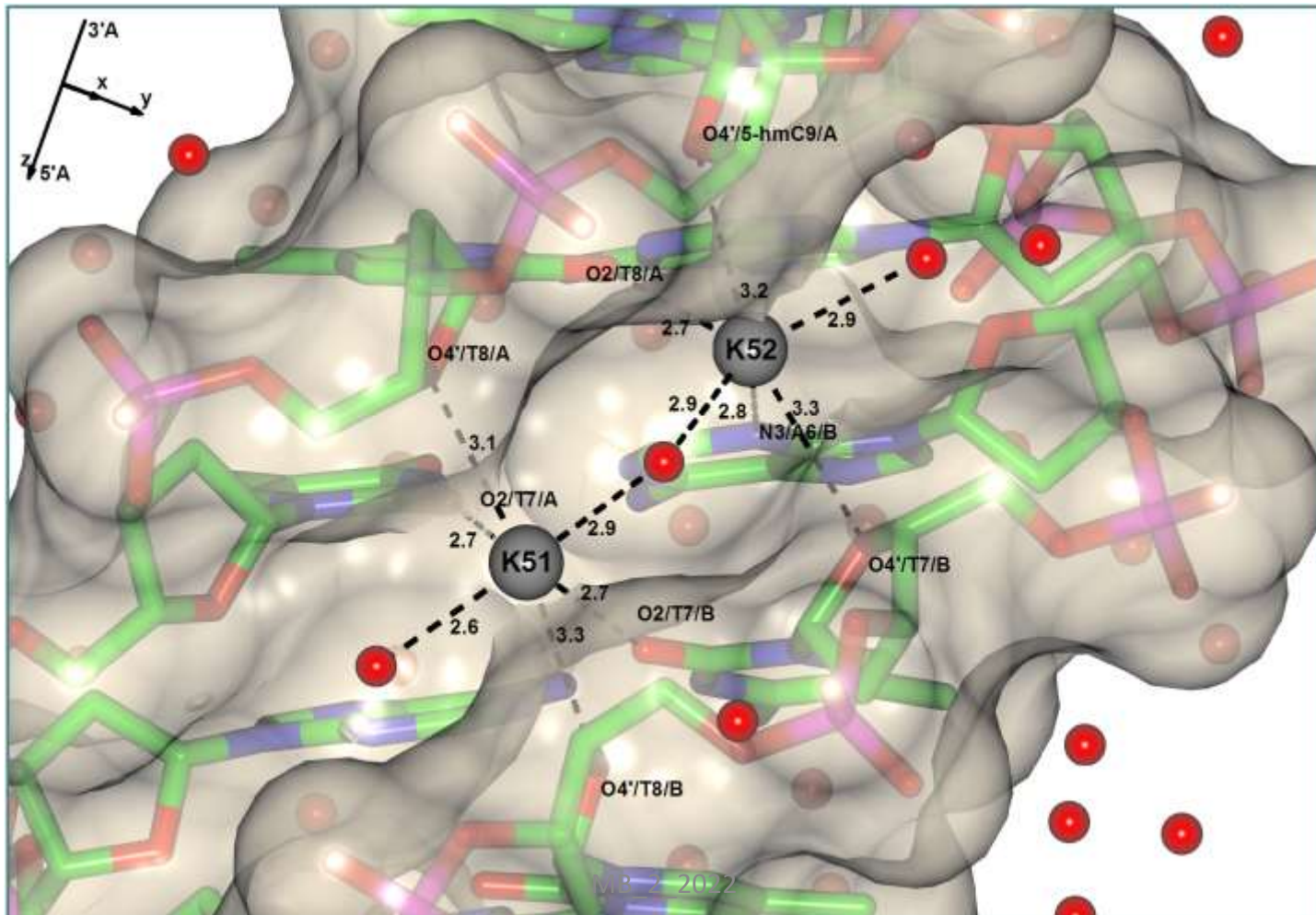
# Base protonation



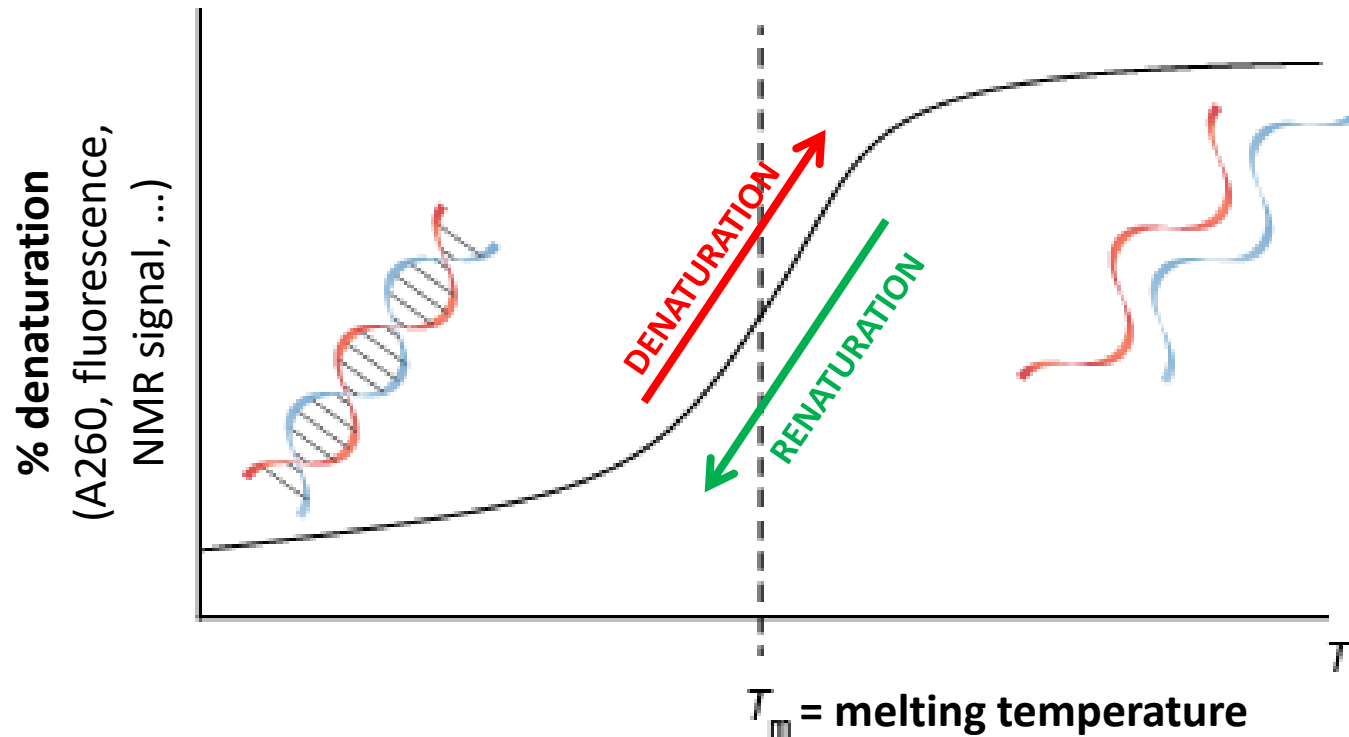
- base protonation might alter the base reactivity
- free bases have pK far from physiological
- pK of bases in DNA might be closer to pH 7.4
- cytosine in C<sub>n</sub> sequences has pK~7 – cytosine i-motif

# DNA double helix x ions / water

- phosphates in DNA backbone are negatively charged – repulsion
- this is compensated by interaction with ions ( $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ , ...) or water (H-H bonds)



# Stability of DNA double helix

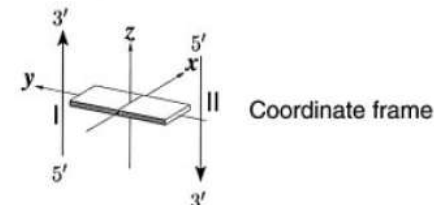
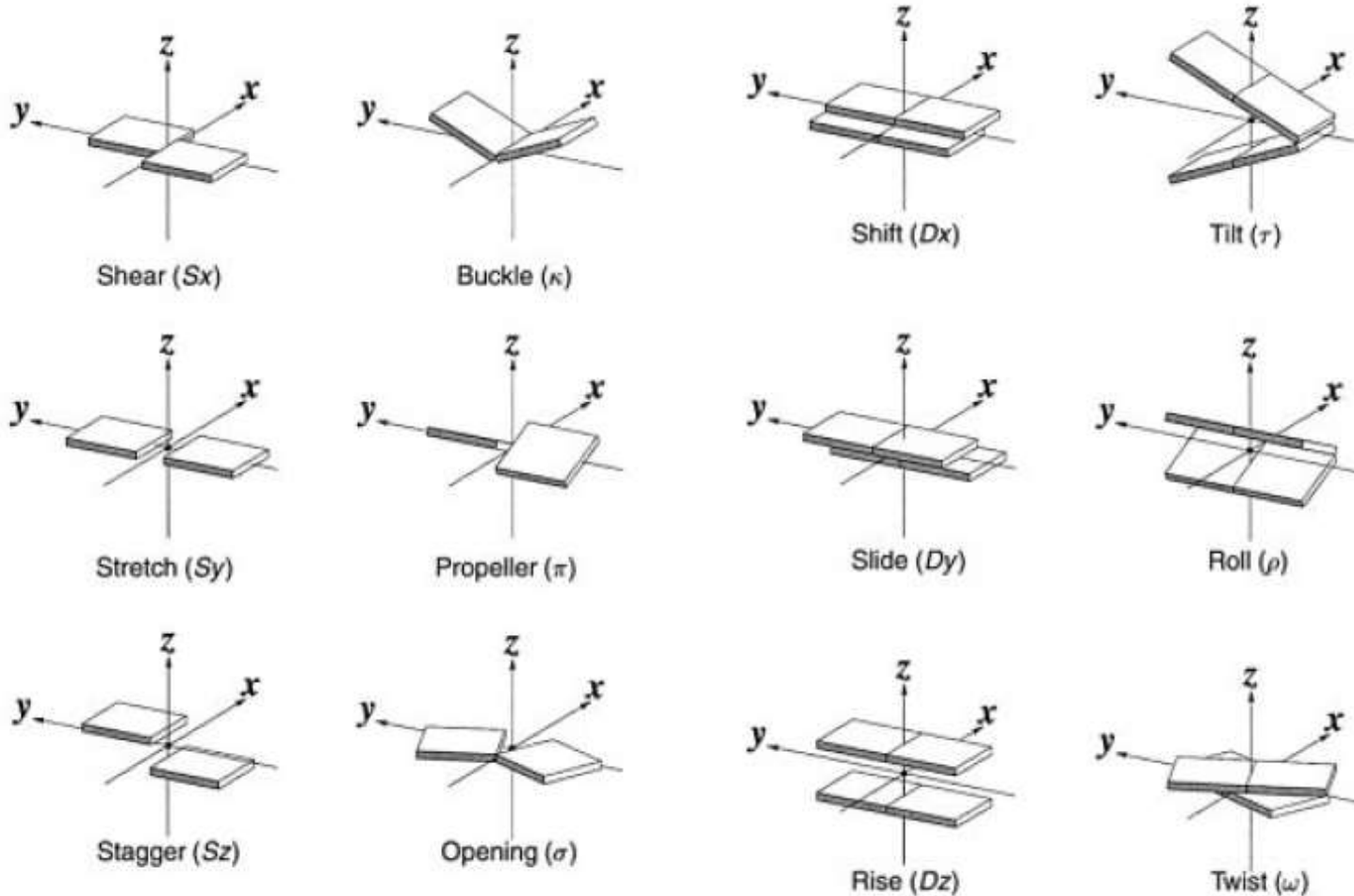


- hydrogen bonds
- base stacking
- repulsion of backbone phosphates

AT = 2 x GC = 3  
various  
 $Mg^{2+} > Na^+$

$T_m$  increases with GC and length  
 $T_m$  increases with length and ions  
 $T_m$  increases with ions

# Base-pair parameters in double helix

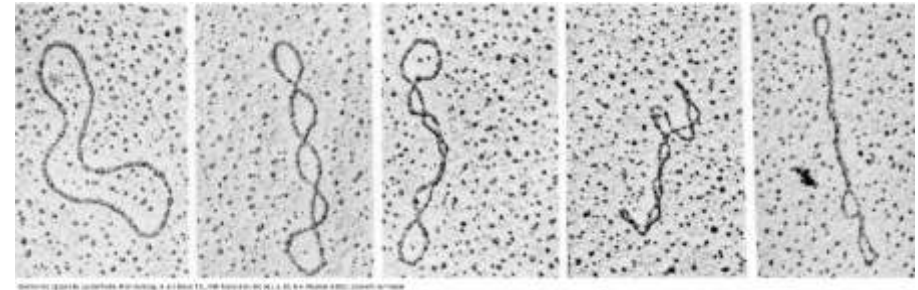
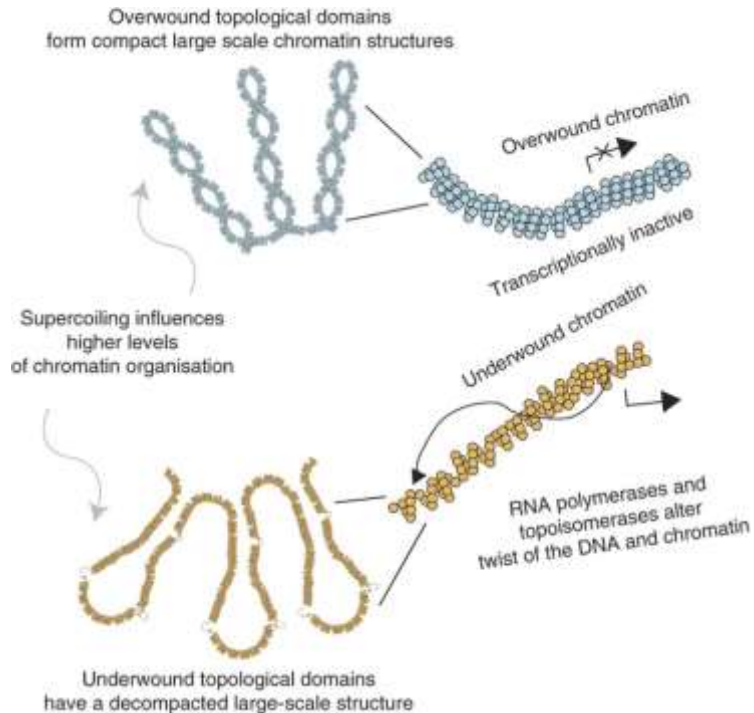


# Types of nucleic acids

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- **linear** (human chromosome) x **circular** (bacterial genome)
- **single-stranded** (most RNAs) x **double-stranded** (human DNA)

# Superhelicity



Superhelicity happen mostly as a result of transition of polymerase complex and unwinding of DNA (helicase, ...) during replication and transcription.

## Topoisomerases

- Enzymes that relax the superhelicity
- Topo I – works on 1 DNA strand
- Topo II – works on 2-strand DNA

# Reactivity of bases with amino acids

Double-stranded NA:  
Interaction of Hoogsteen side  
with amino acid in major groove.

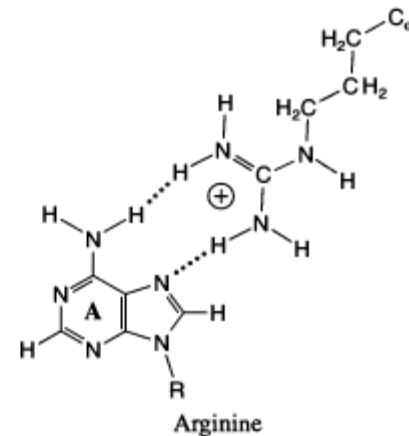
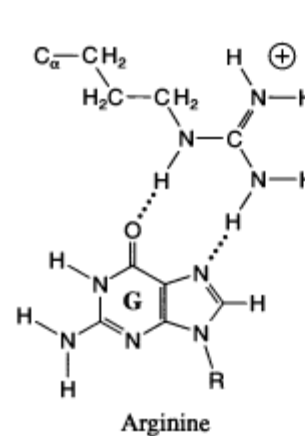
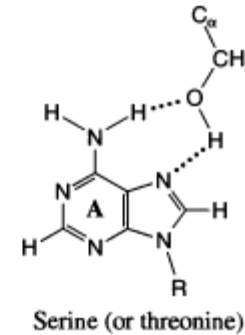
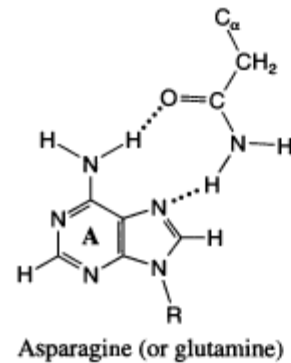


Figure 2-16  
Interactions involving two hydrogen bonds between amino acids and bases that can occur through the major groove of a double helix.



# Reactivity of bases with amino acids

Single-stranded NA:  
Interaction of Watson-Crick side  
with amino acid.

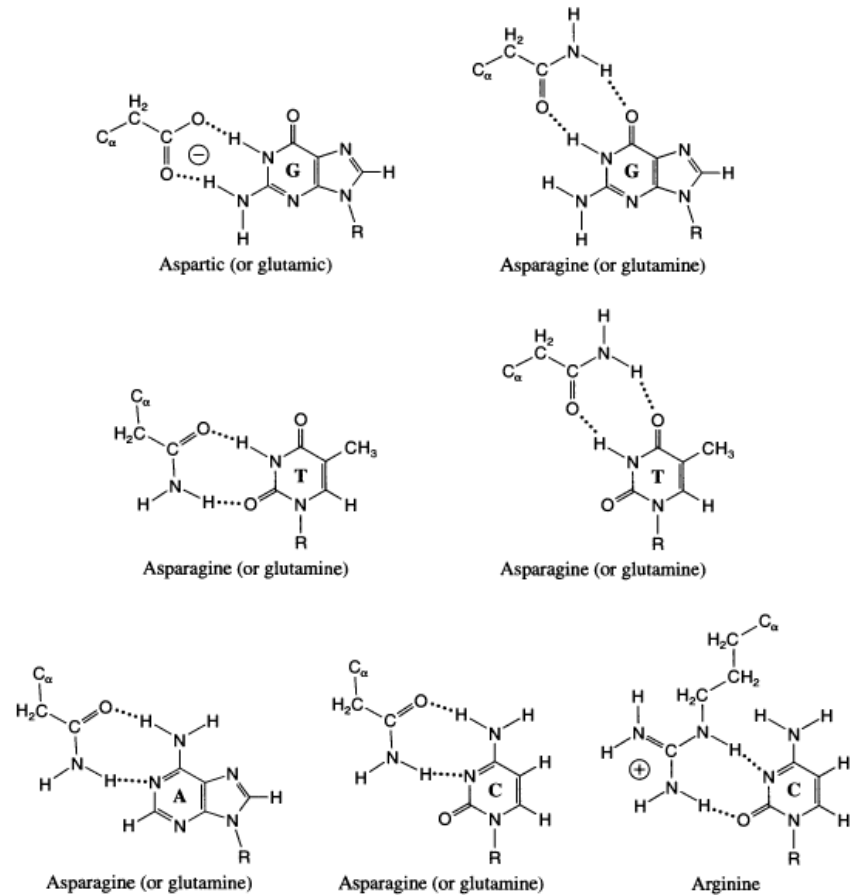
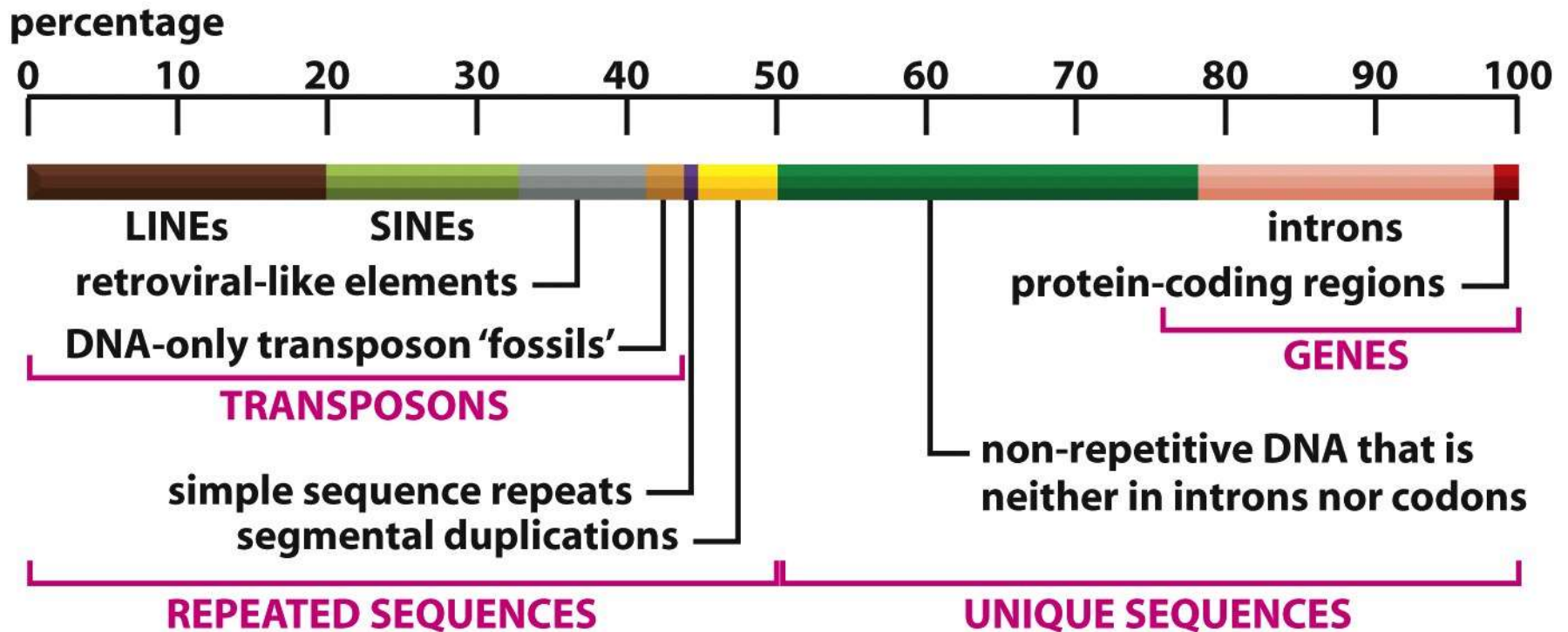


Figure 2-17  
Interactions involving two hydrogen bonds between amino acids and bases that take the place  
of Watson-Crick base pairing.

# Genome composition



# Repetitive sequences - repeats

Some sequences in genome are **unique**, usually the genomic sequences (both coding and non-coding). In contrast, other sequences exist in many copies – **repetitive sequences (repeats)**. The length of repeat (microsatellites 2-6 bp x LINE 6-7000 bp), as well as the number of copies (several – 1.5M SINE in human) is highly variable.

## Structure:

### • **direct** repeats

5' . . . **AGTC** . . . **AGTC** . . . 3'  
3' . . . **TCAG** . . . **TCAG** . . . 5'

5' . . . **AGTC** . . . **CTGA** . . . 3'  
3' . . . **TCAG** . . . **GACT** . . . 5'

### • **inverted** repeats + **palindromes**

5' . . . **AGTC** . . . **GACT** . . . 3'  
3' . . . **TCAG** . . . **CTGA** . . . 5'

## Position:

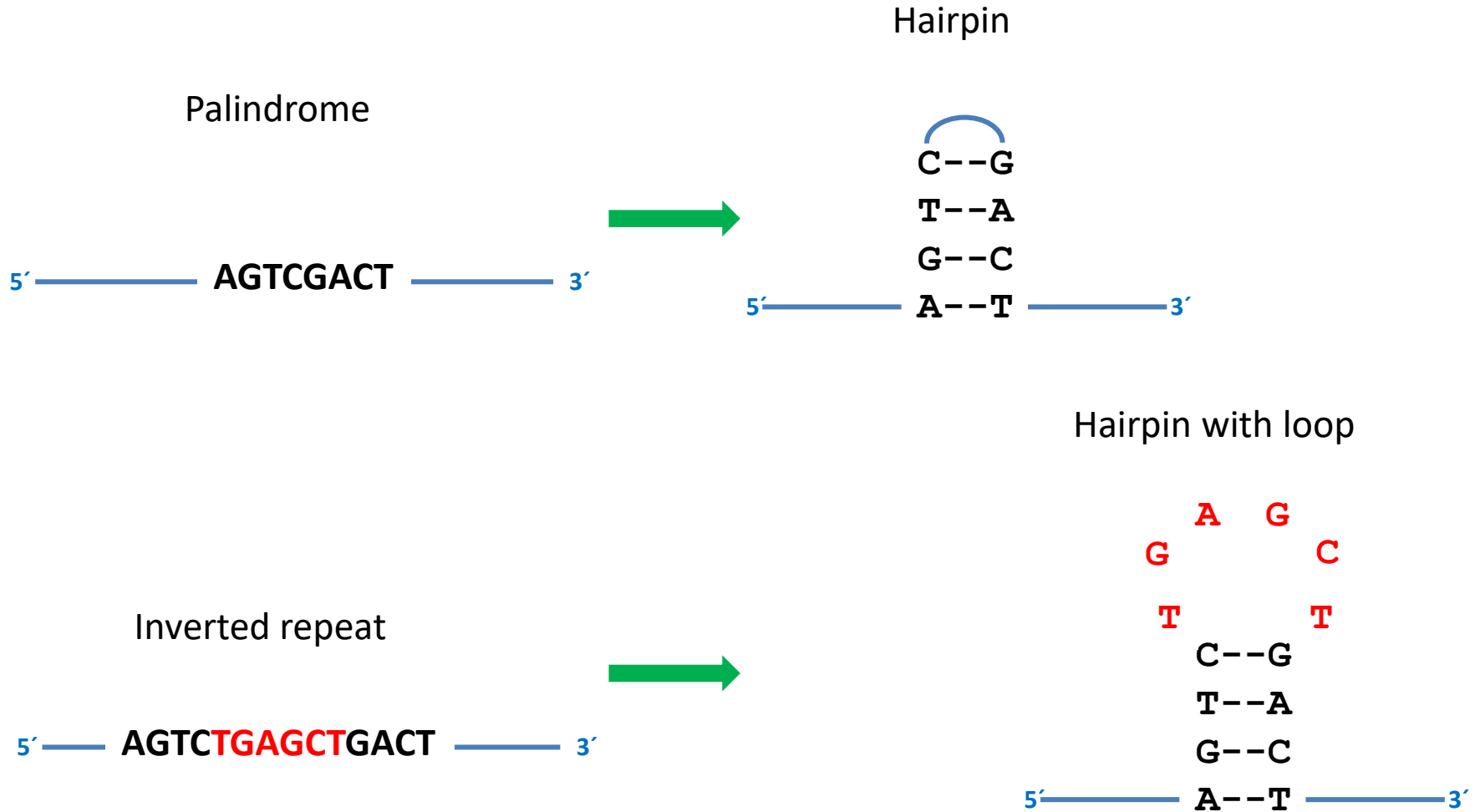
### • **Tandem** repeats



### • **Interspersed** repeats



# Inverted repeats

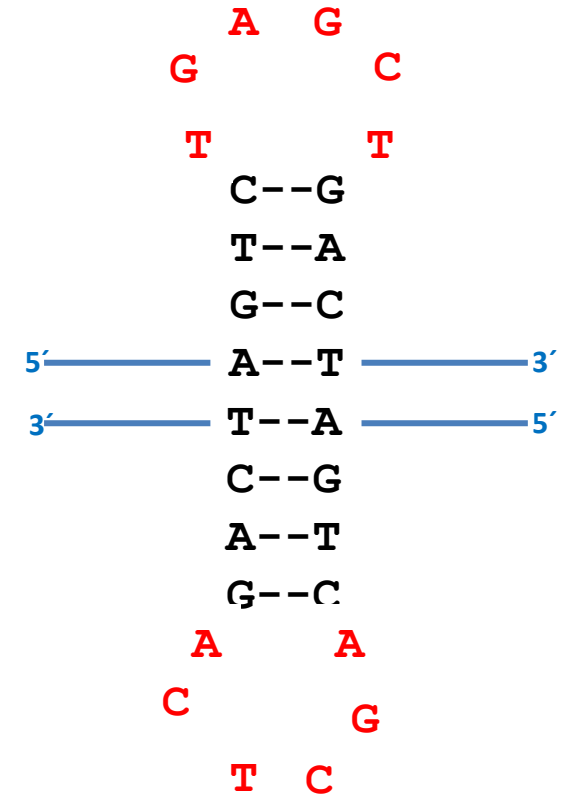


# Inverted repeats

Inverted repeat



Cruciform



# Special types of repetitions - transposons

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Interspersed repetitions with various lengths and number of copies.

LTR – long terminal repeat - 100 bp – 5 kbp – variant of retrotransposons

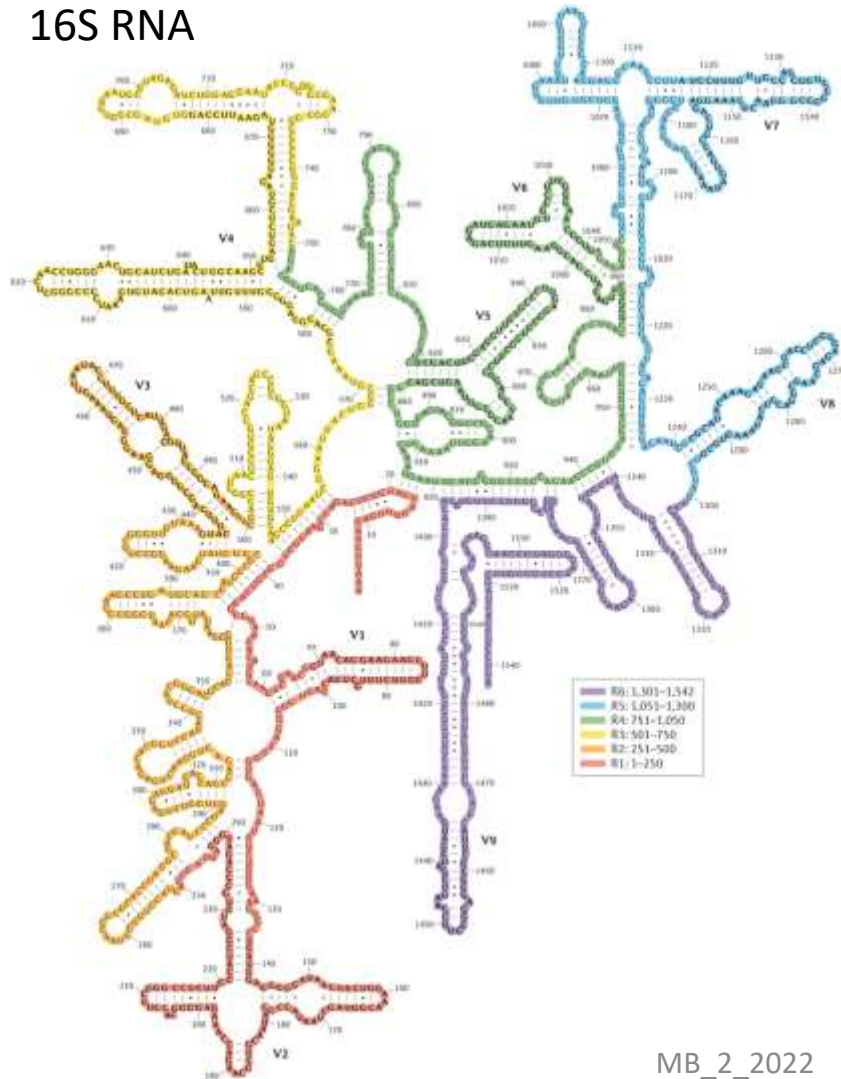
LINE – long interspersed nuclear elements – up to 6 kbp – human > 500k copies

- 3 types (L1, L2, L3) – only some L1 are able to transpose

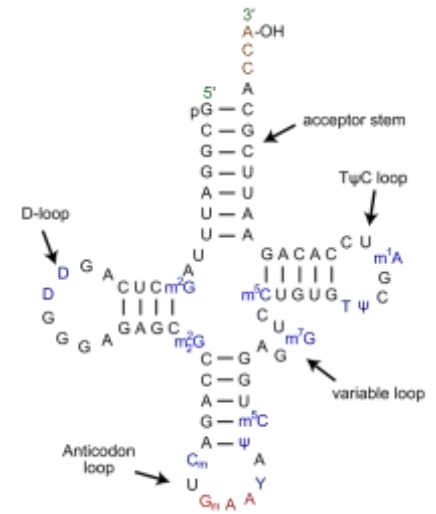
SINE – (Alu, ...) short interspersed nuclear elements – up to 500 bp – human ~ 1,5M copies

# Loops and hairpins in RNA

16S RNA

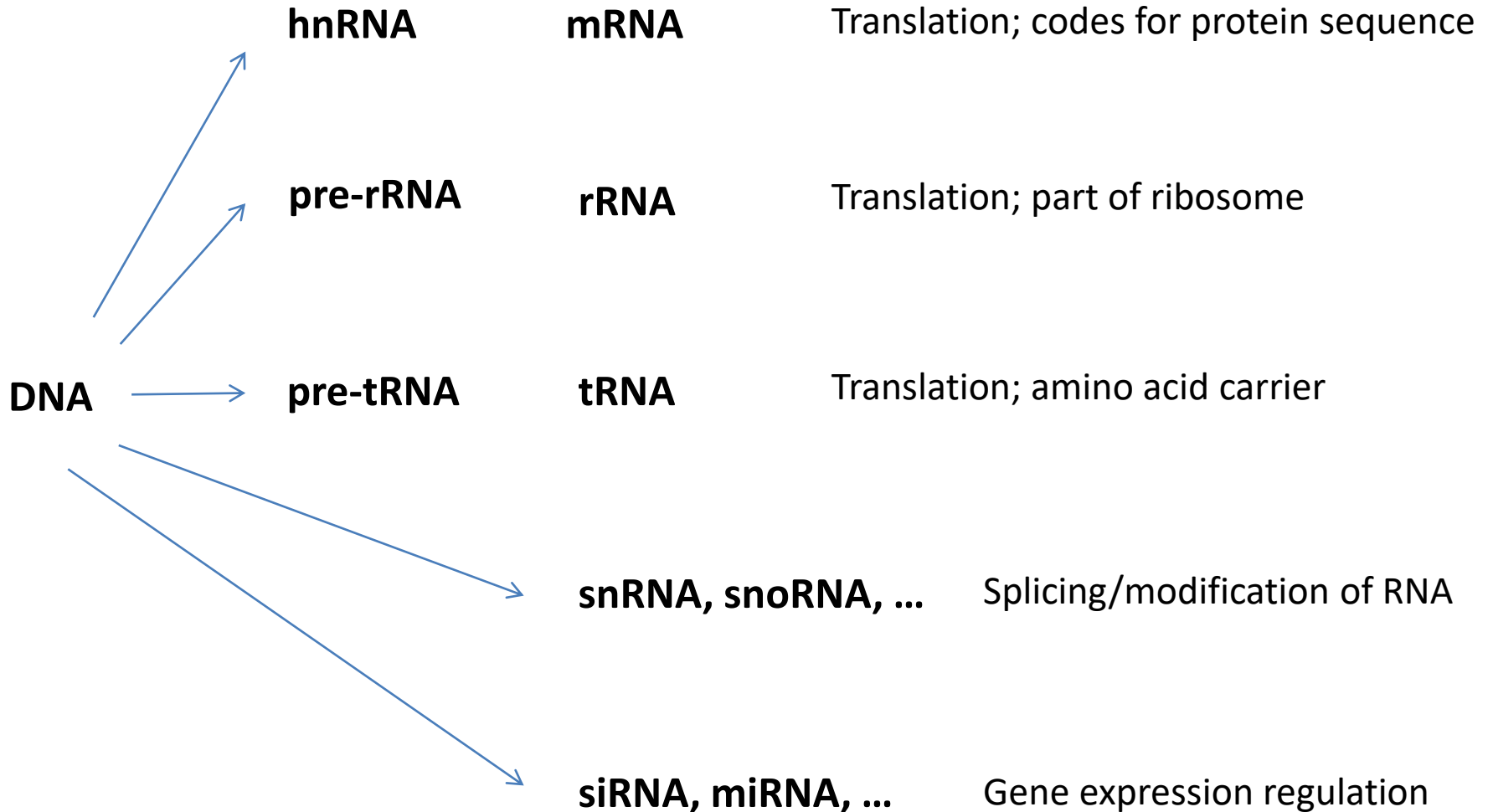


tRNA (Lys)



# Functional types of RNA

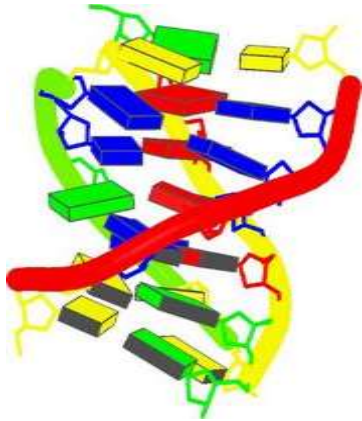
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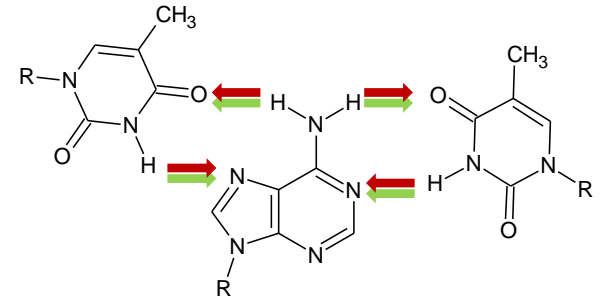
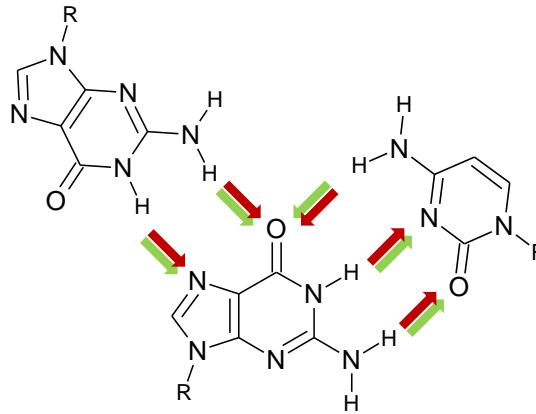
# Hoogsteen pairing - triplexes

- gene expression regulation



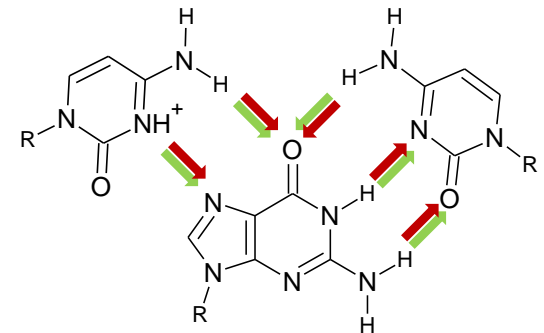
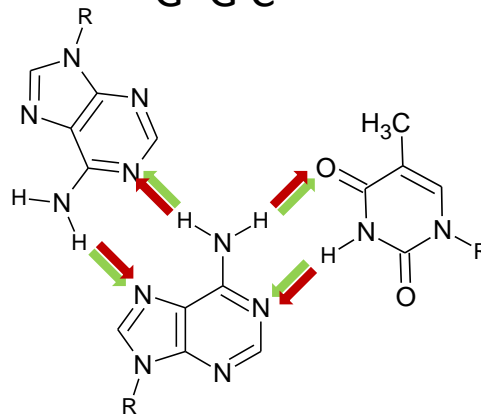
Pu\*Pu Py

Py\*Pu Py



G\*G C

T\*AT



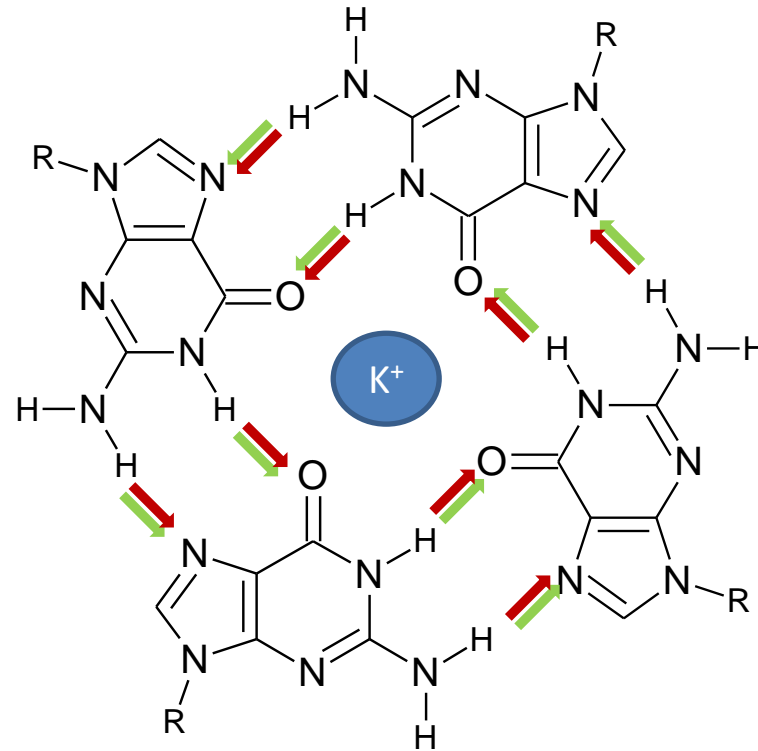
A\*AT

C\*G C



# Hoogsteen pairing – G-quadruplexes

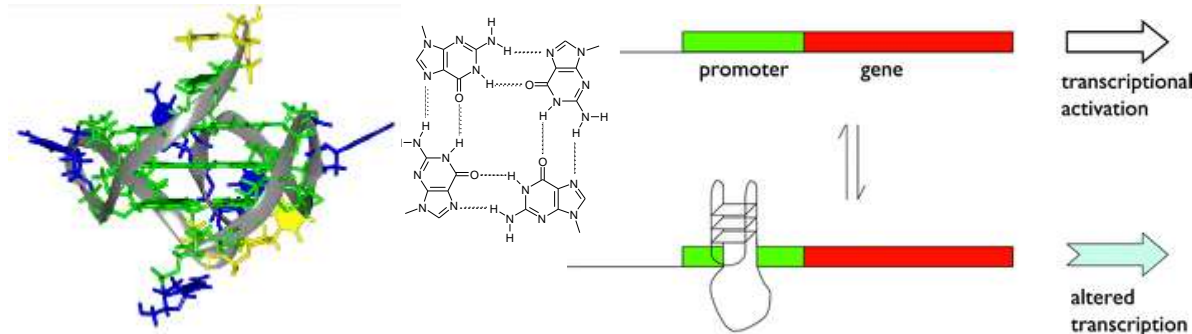
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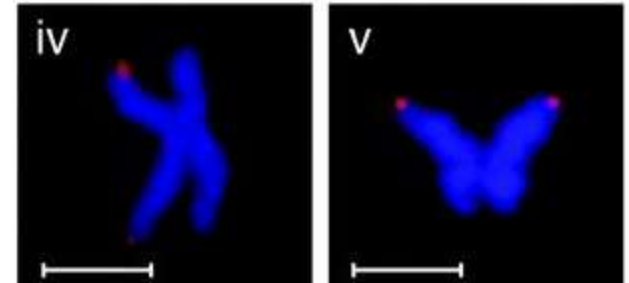
# Guanine quadruplexes



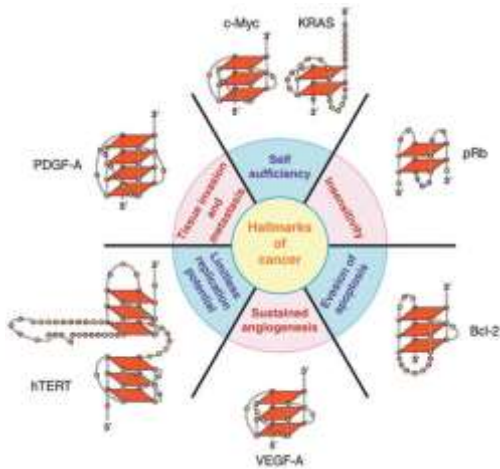
- gene expression regulation
- telomere structure



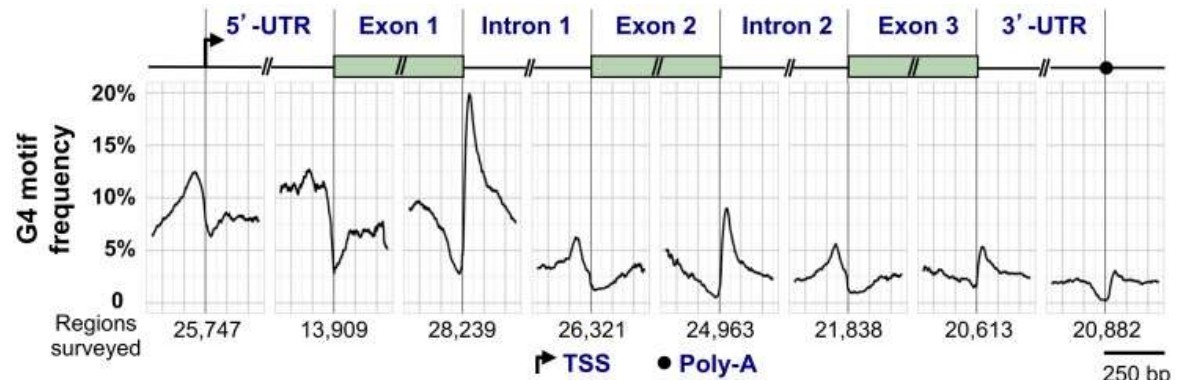
(Huppert J.L., Chem Soc Rev, 2008)



(Biffi G., et al., Nat Chem, 2013)



(Brooks T. A., et al., FEBS J, 2010)



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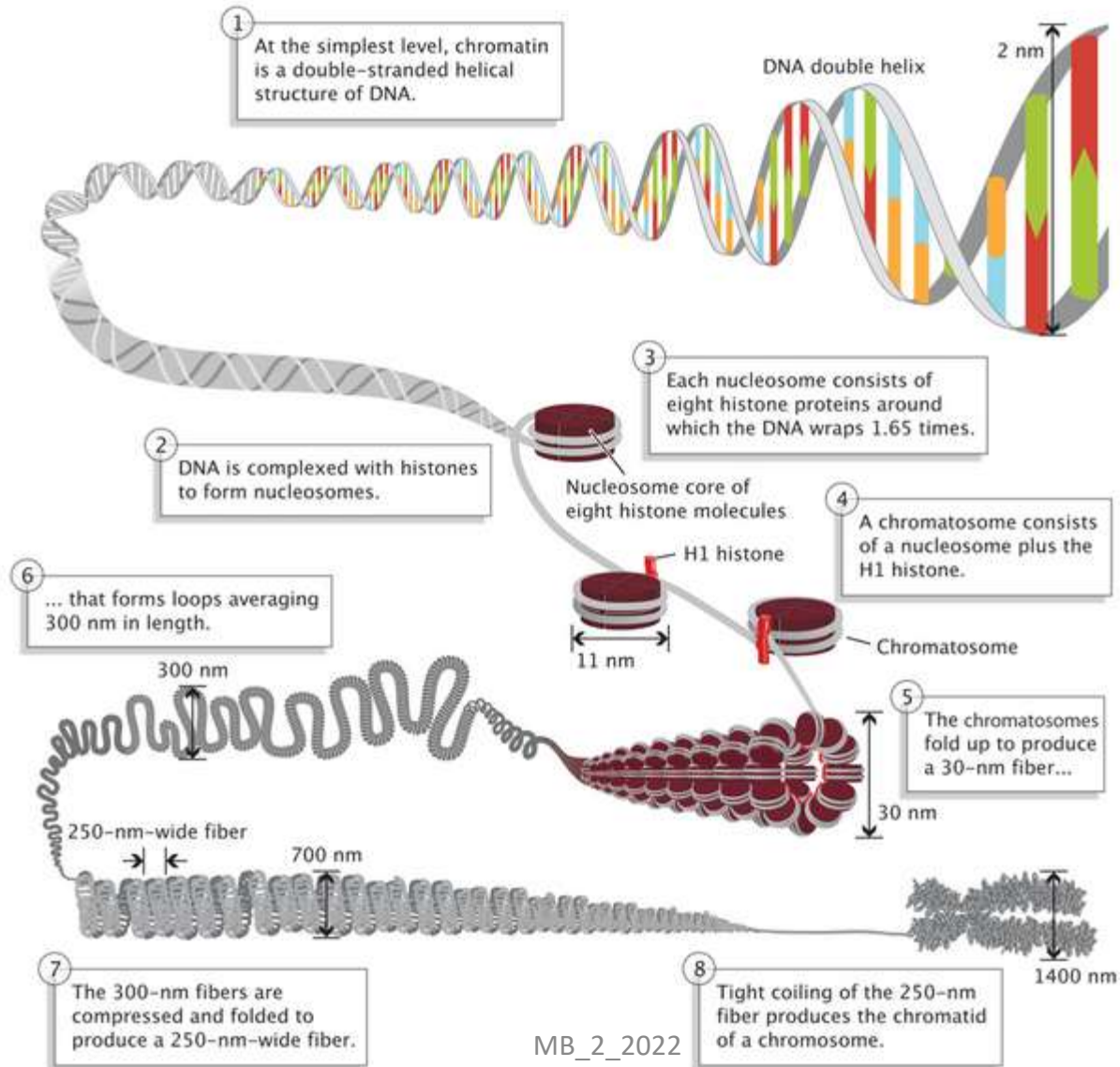
(Maizels N. and Gray L.T., PloS Genet., 2013)

# Base reactivity

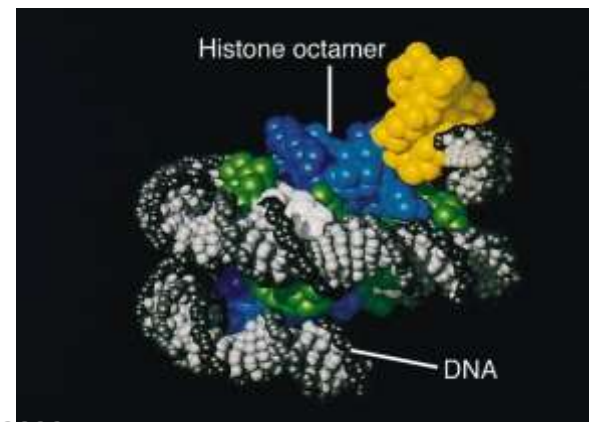
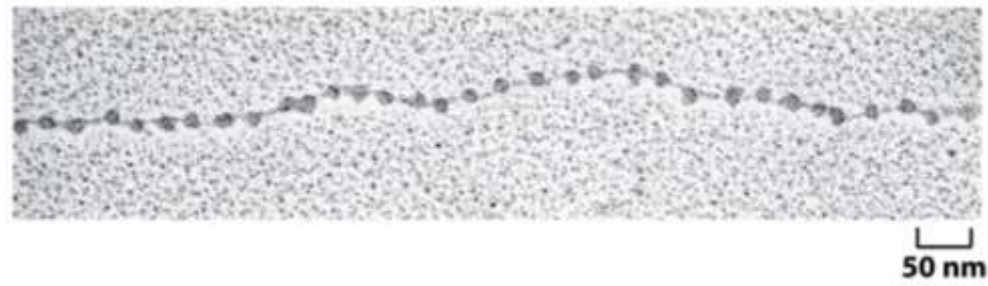
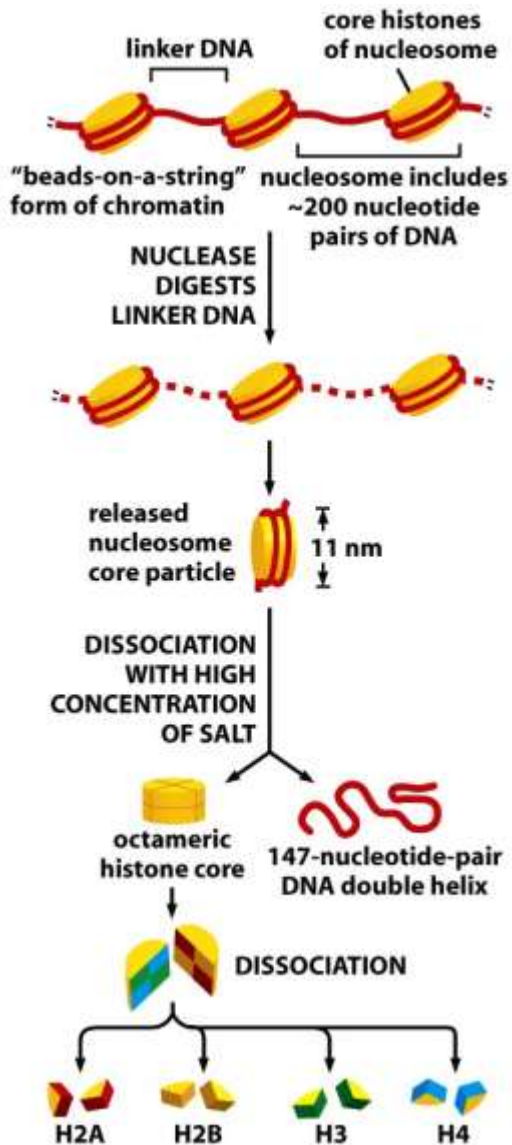
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Hydrophobic bases with high ability to form hydrogen bonds are reluctant to be freely expressed into water environment around – if there is any chance to avoid this and lower the base exposition to the environment by any type of base pairing or base stacking, the bases tend to form a structure. Even the “single-stranded” RNA or DNA forms, in fact, compact structure with number of base pairs.

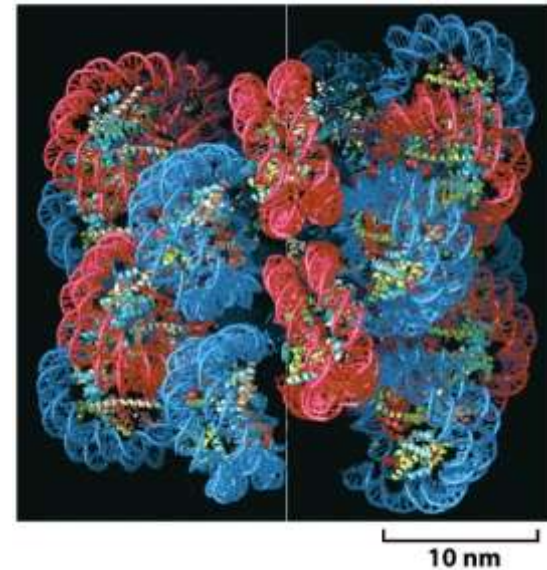
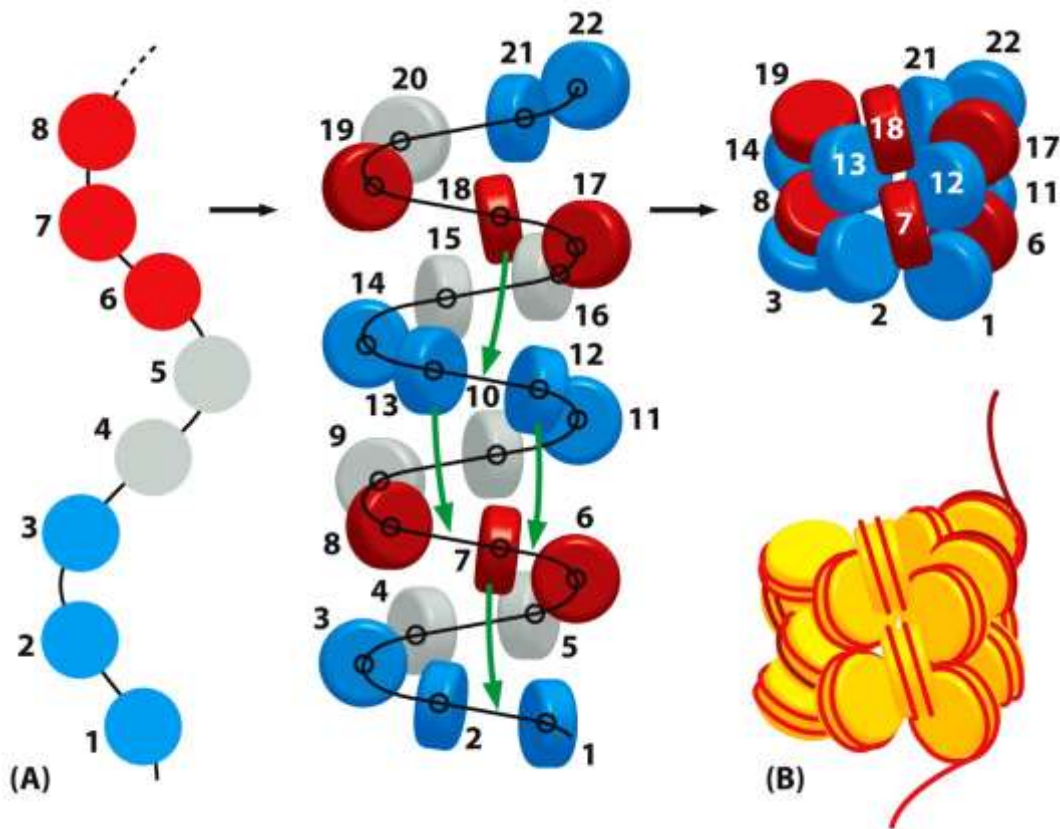
# Packing of DNA into chromosome



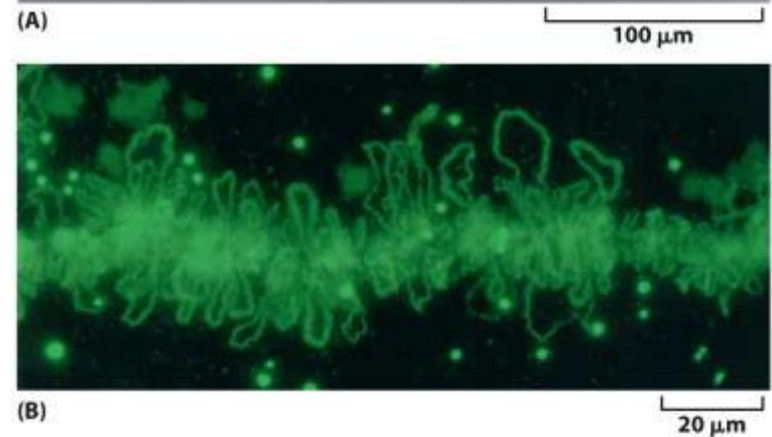
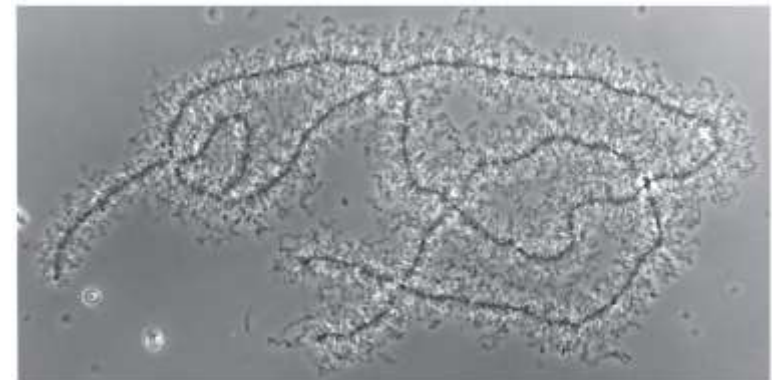
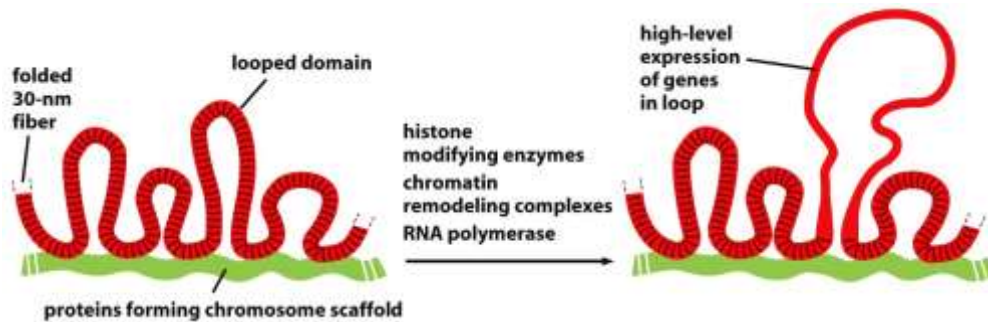
# Binding of DNA to a histone octamer



# Folding of nucleosomes into 30 nm fiber

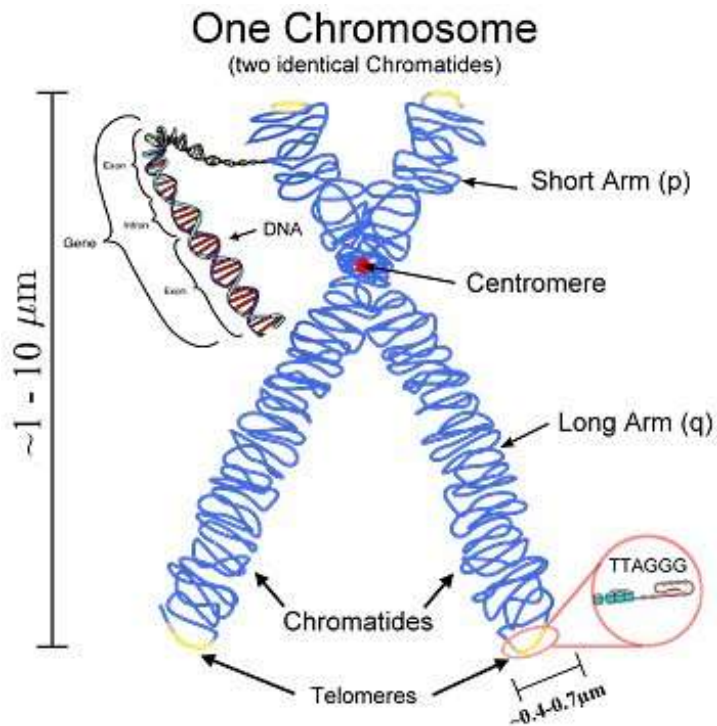


# 30 nm fiber binds to protein scaffold



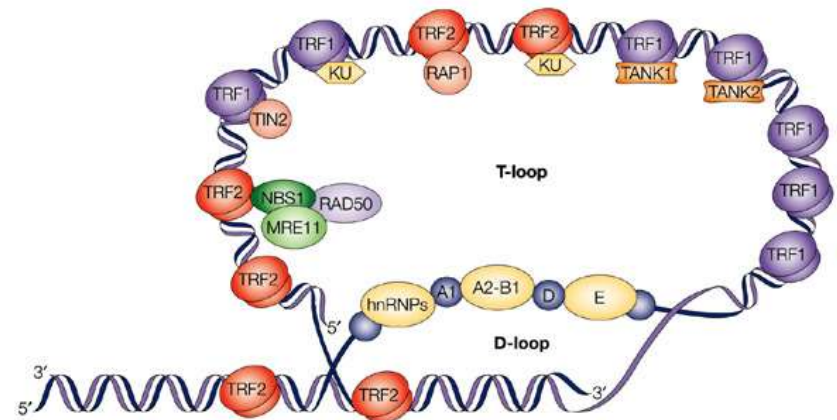


# Chromosome



**Centromere** – here are the chromosomes connected to the system of cellular microtubules – important for chromosome segregation during cell division

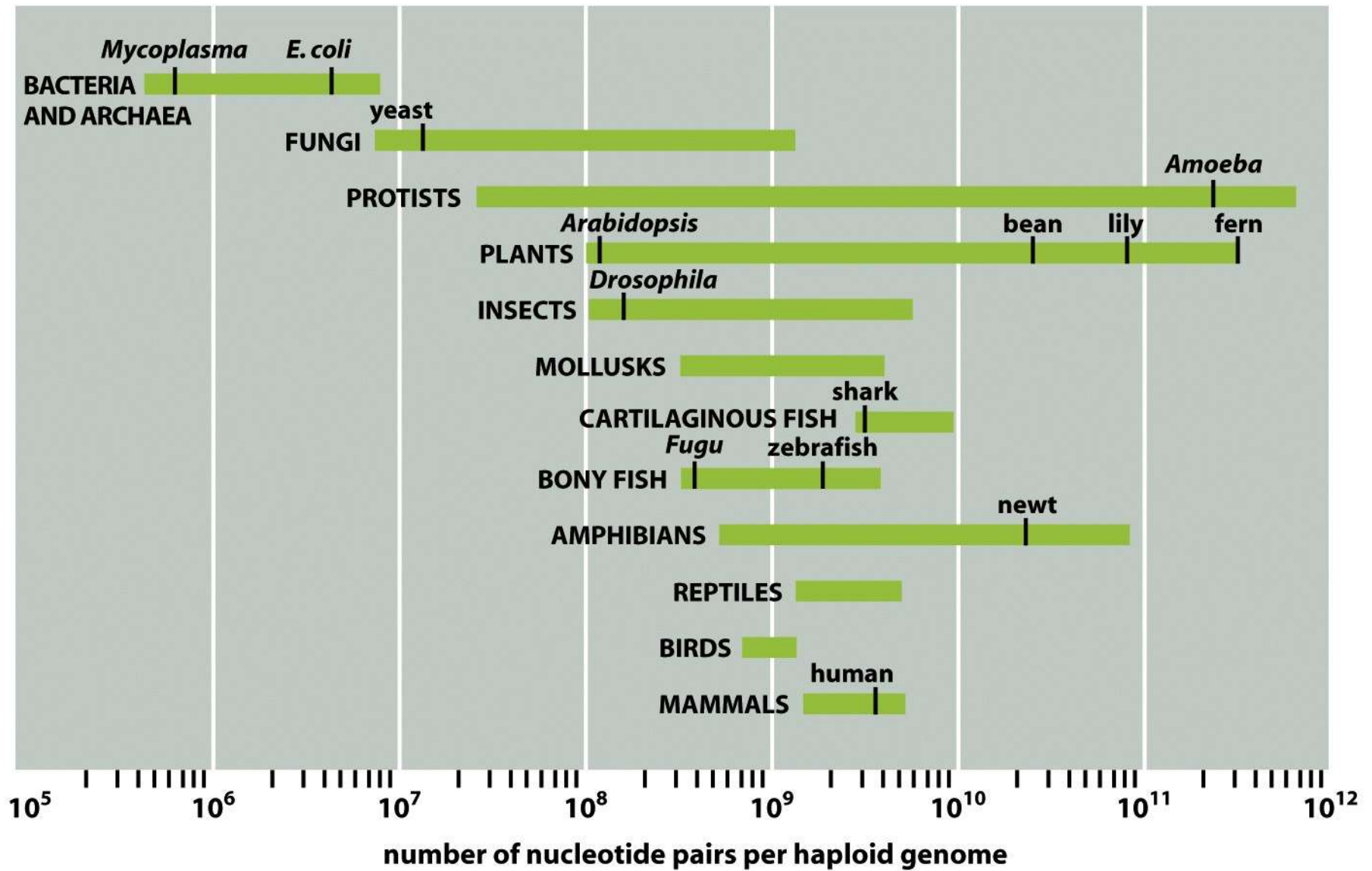
**Telomere** – terminal part of chromatides that protect the end from being recognised as a double-strand break by a DNA repair machinery



# Chromosome

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Fully condensed chromosomes are present only during the cell division, otherwise they are more or less decondensed to a lower levels of structure, especially in transcriptionally active sites (**euchromatin**). Transcriptionally inactive parts of DNA, as well as repetitive regions or telomeres are much more condensed (**heterochromatin**). Various types of chromatin differ in **epigenetic** markers of both DNA (5-methyl cytosine) and histones (methylation a acetylation).


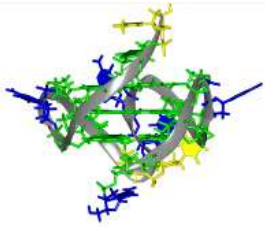
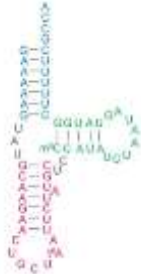
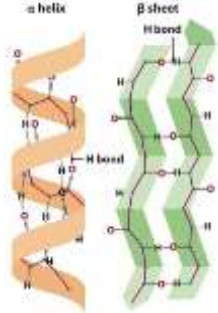

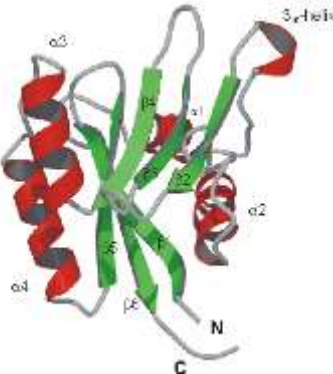


**Table 1–1 Some Genomes That Have Been Completely Sequenced**

SPECIES	SPECIAL FEATURES	HABITAT	GENOME SIZE (1000s OF NUCLEOTIDE PAIRS PER HAPLOID GENOME)	ESTIMATED NUMBER OF GENES CODING FOR PROTEINS
<b>ARCHAEA</b>				
<i>Methanococcus jannaschii</i>	lithotrophic, anaerobic, methane-producing	hydrothermal vents	1664	1750
<i>Archaeoglobus fulgidus</i>	lithotrophic or organotrophic, anaerobic, sulfate-reducing	hydrothermal vents	2178	2493
<i>Nanoarchaeum equitans</i>	smallest known archaean; anaerobic; parasitic on another, larger archaean	hydrothermal and volcanic hot vents	491	552
<b>EUCARYOTES</b>				
<i>Saccharomyces cerevisiae</i> (budding yeast)	minimal model eucaryote	grape skins, beer	12,069	~6300
<i>Arabidopsis thaliana</i> (Thale cress)	model organism for flowering plants	soil and air	~142,000	~26,000
<i>Caenorhabditis elegans</i> (nematode worm)	simple animal with perfectly predictable development	soil	~97,000	~20,000
<i>Drosophila melanogaster</i> (fruit fly)	key to the genetics of animal development	rotting fruit	~137,000	~14,000
<i>Homo sapiens</i> (human)	most intensively studied mammal	houses	~3,200,000	~24,000

Genome size and gene number vary between strains of a single species, especially for bacteria and archaea. The table shows data for particular strains that have been sequenced. For eucaryotes, many genes can give rise to several alternative variant proteins, so that the total number of proteins specified by the genome is substantially greater than the number of genes.

# Levels of structure of biopolymers

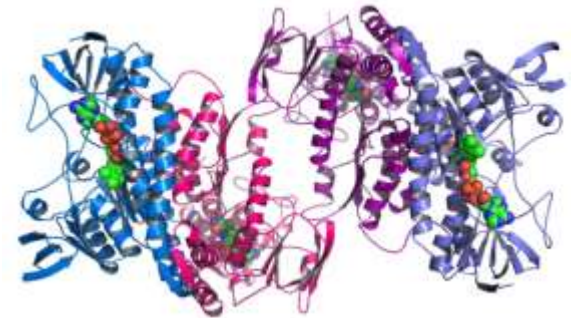
	DNA	RNA	Protein
Primary	AGGCTGCAAGTCGAT	UCCGACGUUCAGCUA	Ser-Asp-Val-Gln-Leu
Secondary		 	
Tertiary			

# Genetic code

Set of rules that assign a sequence of amino acids in the protein to the sequence of nucleotides in DNA or RNA.



Transcription  
+  
Translation



M.W.Nirenberg

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*RNA CODEWORDS AND PROTEIN SYNTHESIS, III.  
ON THE NUCLEOTIDE SEQUENCE OF A CYSTEINE  
AND A LEUCINE RNA CODEWORD*

BY PHILIP LEDER AND MARSHALL W. NIRENBERG

NATIONAL HEART INSTITUTE, NATIONAL INSTITUTES OF HEALTH

Communicated by Richard E. Roberts, October 1, 1964

Previous studies utilizing randomly ordered synthetic polynucleotides to direct amino acid incorporation into protein in *E. coli* extracts indicated that RNA codewords corresponding to valine, leucine, and cysteine contain the bases (UUG).<sup>1-4</sup> The activity of chemically defined trinucleotides in stimulating the binding of a specific C<sup>14</sup>-aminoacyl-sRNA to ribosomes, prior to peptide bond formation,<sup>2</sup> provided a means of investigating base sequence of RNA codewords and showed that the sequence of a valine RNA codeword is GpUpU.<sup>5</sup>

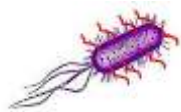
# Properties of genetic code

- genetic code is based on **triplets** – one amino acid in protein is coded by a sequence of three nucleotides in DNA (RNA)

mRNA    CGUGGUACGAUUGGAUGU  
Protein    **Arg Gly Thr Ile Gly Cys**

Triplet = Codon x anticodon = complementary sequence on particular tRNA that carries the respective amino acid

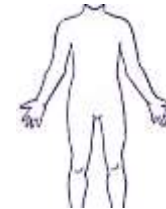
- genetic code is **universal** – individual triplets code for the same amino acid in almost all organisms (x mitochondria)



CGU = **Arginine**



CGU = **Arginine**



CGU = **Arginine**

- genetic code is **degenerated** – one amino acid might be coded by several different triplets (but the opposite is not true)



# Genetic code

	Second nt				
First nt	U	C	A	G	Third nt
U	Phe	Ser	Tyr	Cys	U
	Phe	Ser	Tyr	Cys	C
	Leu	Ser	STOP	STOP/Sel	A
	Leu	Ser	STOP	Trp	G
C	Leu	Pro	His	Arg	U
	Leu	Pro	His	Arg	C
	Leu	Pro	Gln	Arg	A
	Leu	Pro	Gln	Arg	G
A	Ile	Thr	Asn	Ser	U
	Ile	Thr	Asn	Ser	C
	Ile	Thr	Lys	Arg	A
	Met/START	Thr	Lys	Arg	G
G	Val	Ala	Asp	Gly	U
	Val	Ala	Asp	Gly	C
	Val	Ala	Glu	Gly	A
	Val	Ala	Glu	Gly	G



# Reading frames

---

Genetic code is based on triplets – three possible ways of reading (**reading frames**), but only one is correct.

mRNA    CGUGGUACGAUUGGAUGU  
Protein1    **Arg Gly Thr Ile Gly Cys**

mRNA    CGUGGUACGAUUGGAUGU  
Protein2    **Val Val Arg Leu Asp**

mRNA    CGUGGUACGAUUGGAUGU  
Protein3    **Trp Tyr Asp Trp Met**

# Genetic code

---

Although the genetic code is universal, the usage of particular codons, as well as the amount of particular tRNAs and aminoacyl transferases differ

Modification of synthetic genes for recombinant protein production according to the expression system used (Bacteria, human, ...) might be highly beneficial.