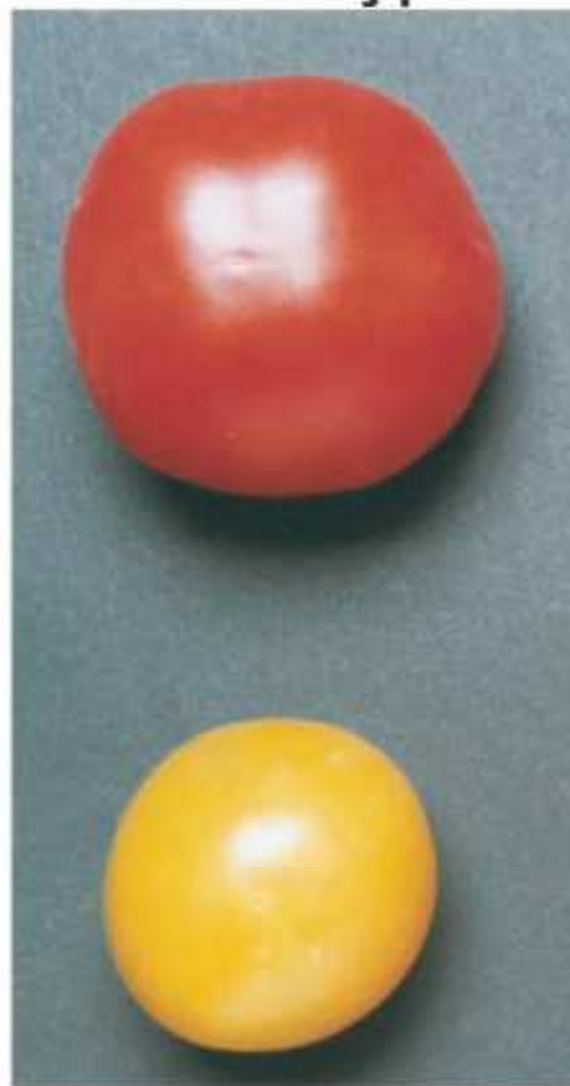


Regulation of plant development by ethylene

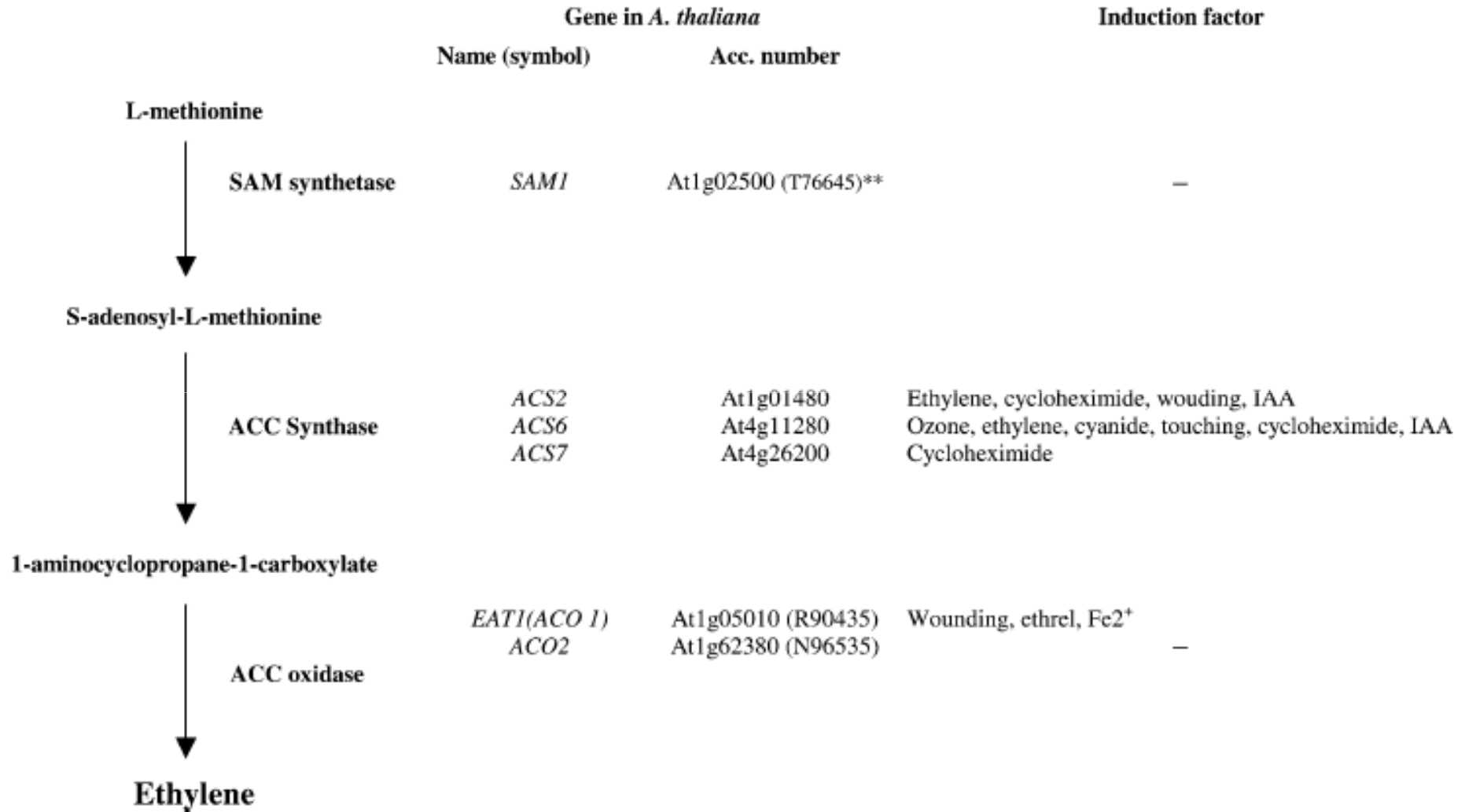


Wild Type

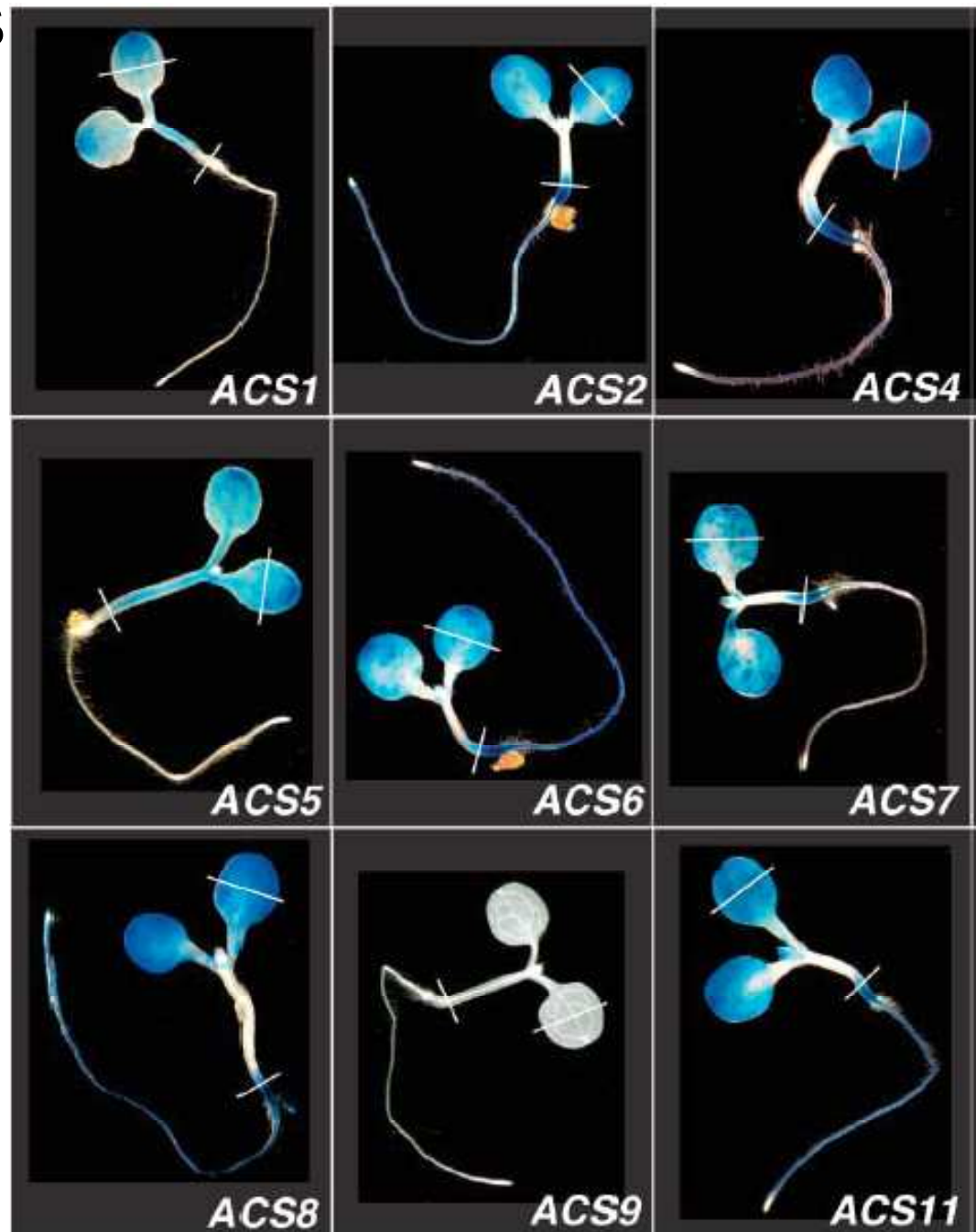


Never-ripe

Ethylene biosynthesis

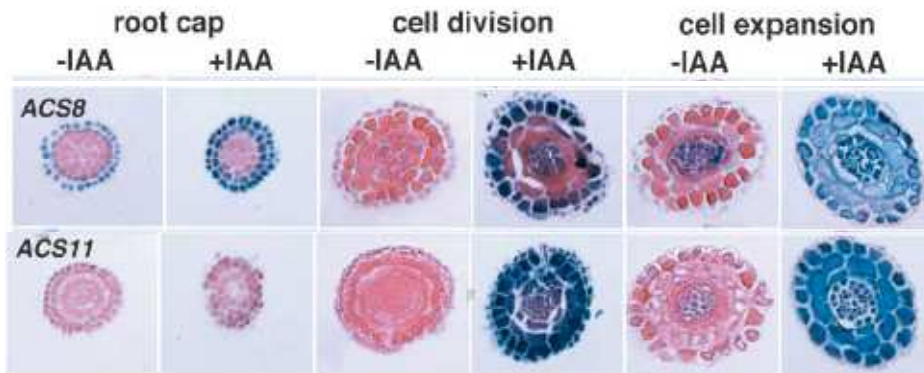
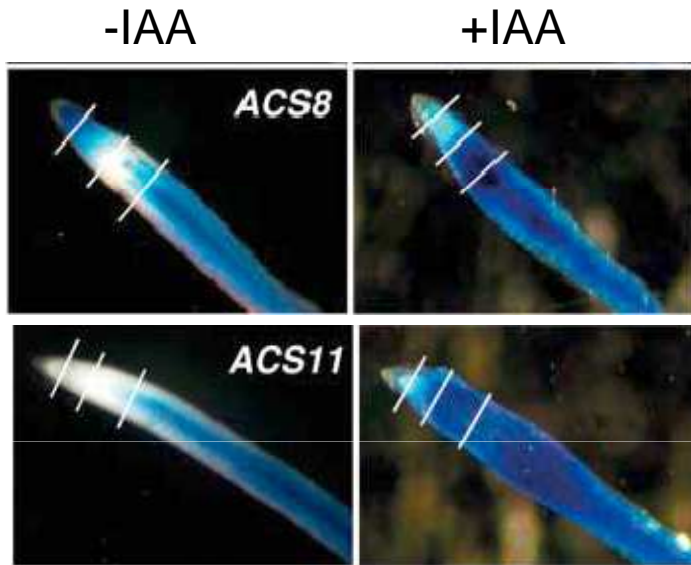


Expression analysis of ACS

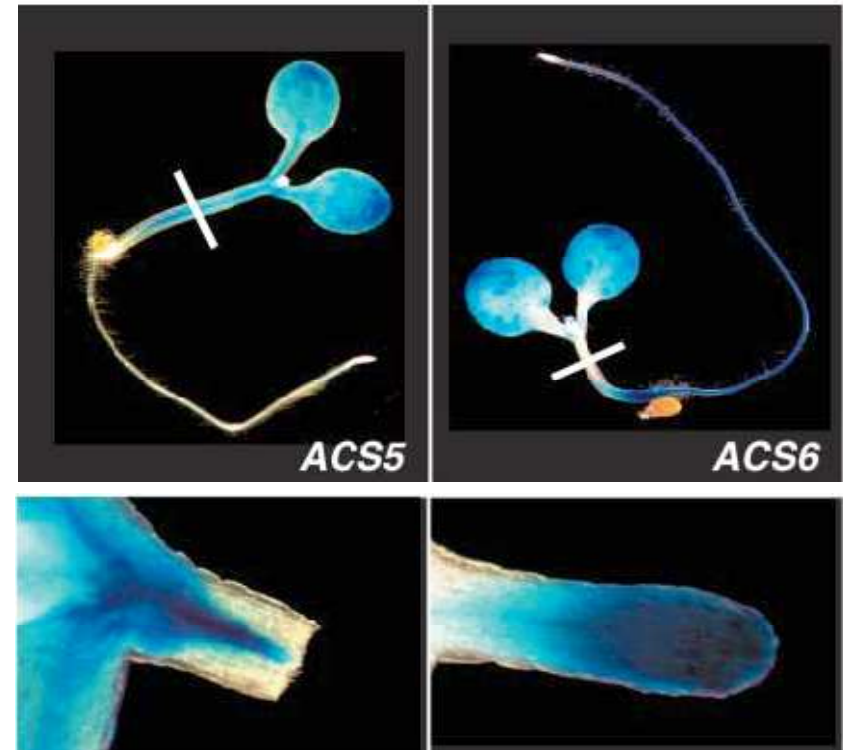


Transcriptional regulation of ACS expression

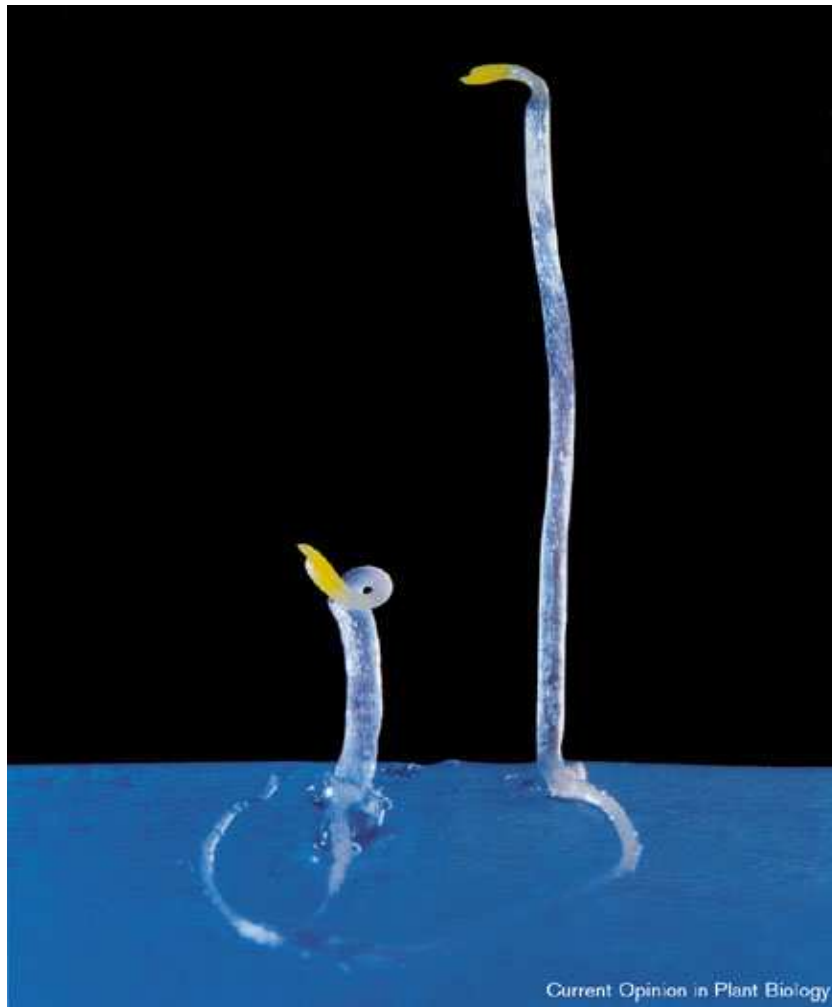
Auxin



Wounding

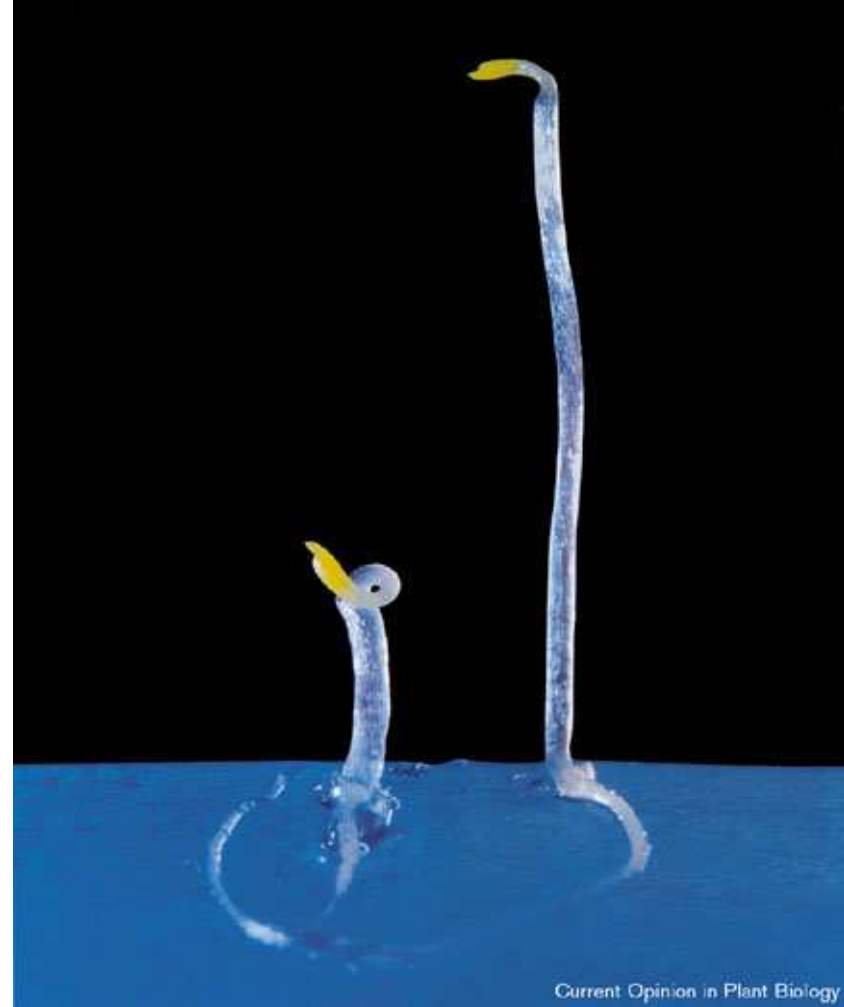


Mutant screens for ethylene pathway genes



C_2H_4

air

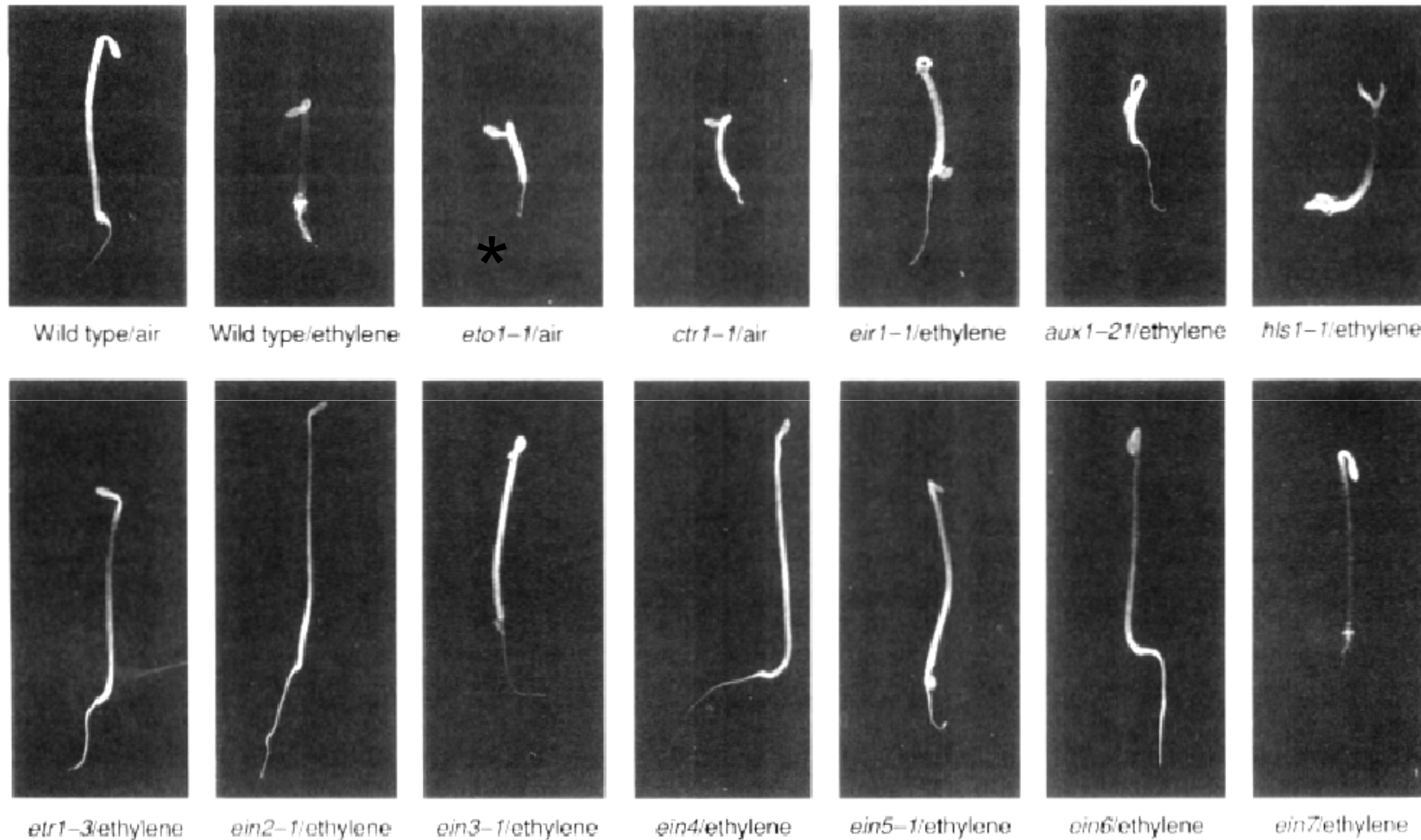


air

C_2H_4

Screen for ethylene mutants

I. Genes involved in regulation of biosynthesis

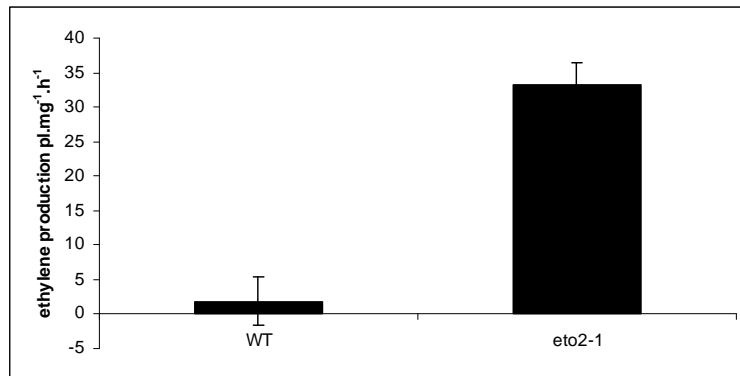


Roman et al., 1994

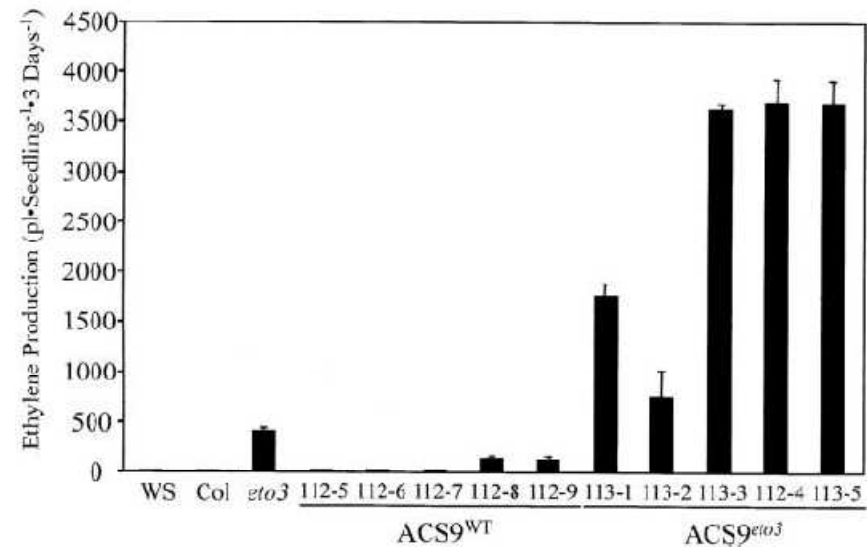
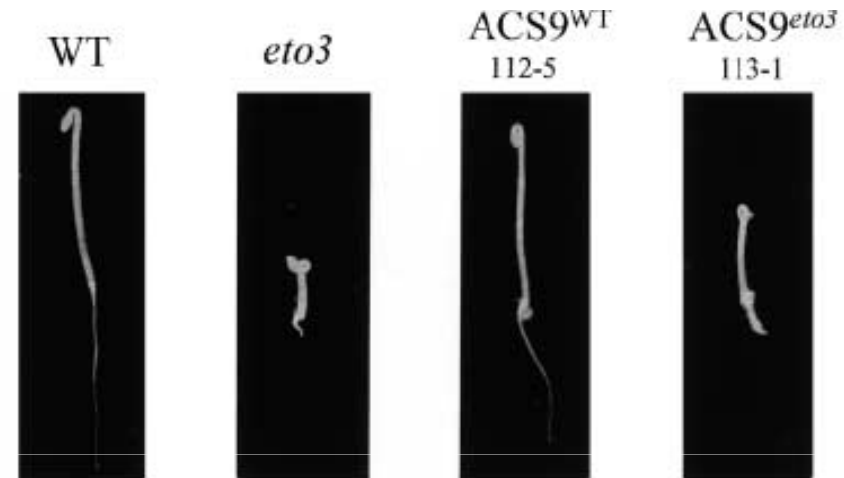
* phenotype rescued by inhibitor of ethylene biosynthesis

eto mutants – constitutive triple response

eto2

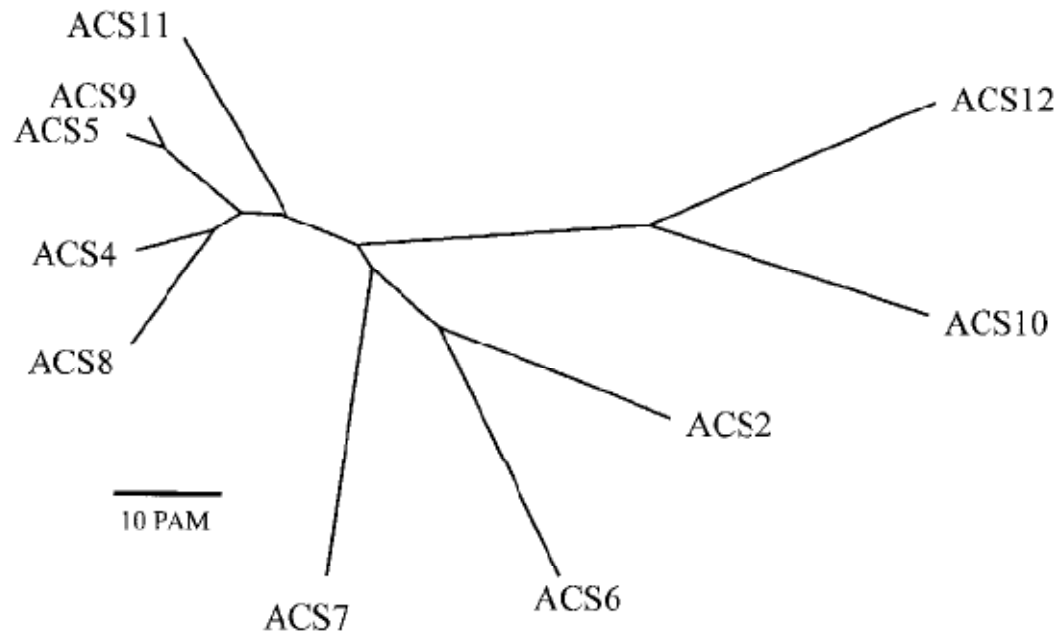


eto3

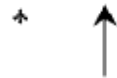


- overproduction of ethylene

eto2,eto3 dominant mutation results from single amino acid change in the C terminus ACS5,ACS9



ACS4	457	VSNWVFRLSFHDREAEEER
ACS8	452	VSNWVFRLSFHDREPEER
<i>eto2</i>	453	VSNWVFpgfmdrsct
ACS5	453	VSNWVFRVSWTDRVPDER
ACS9	453	VSNWVFRVSWTDRVPDER
<i>eto3</i>	453	VSNWDFRVSWTDRVPDER

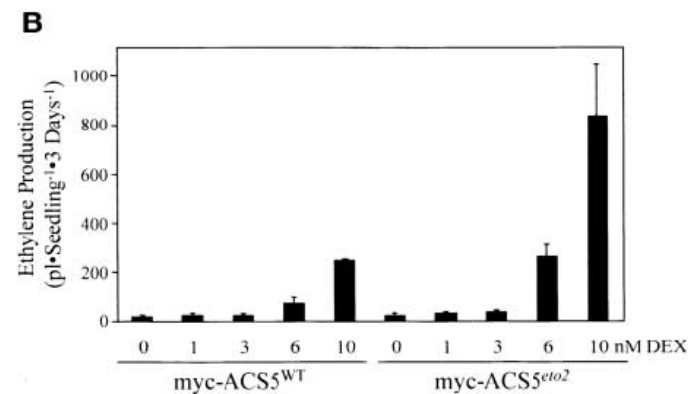
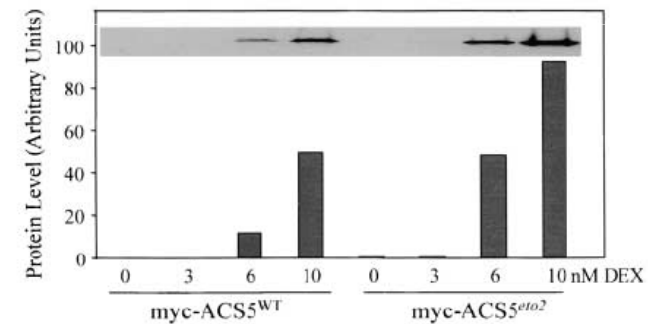
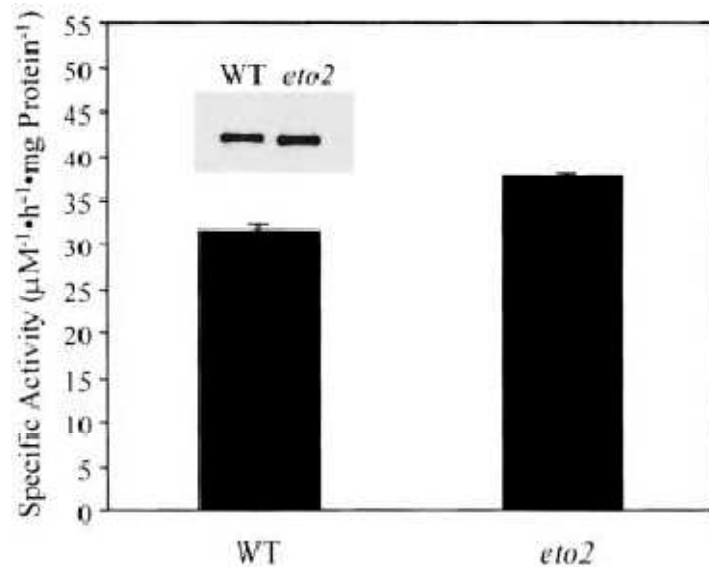


eto3 mutation does not affect level of ACS9 mRNA

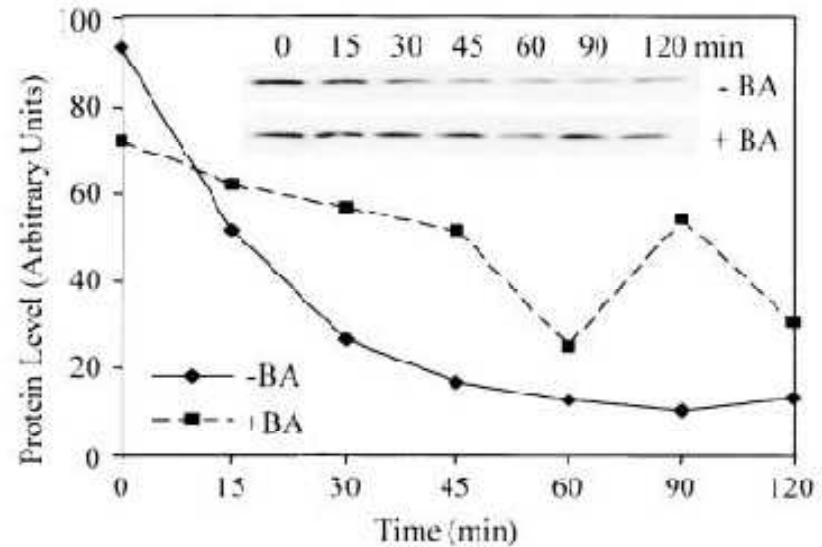
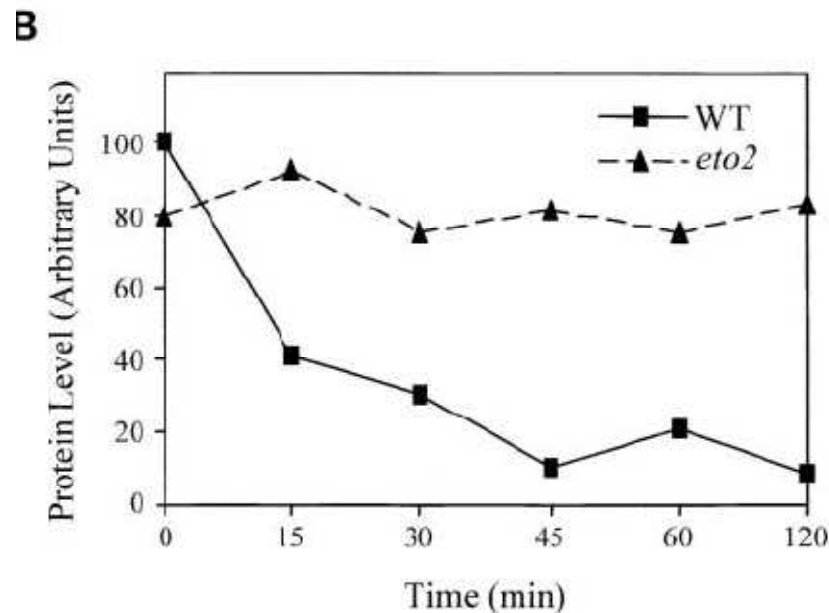
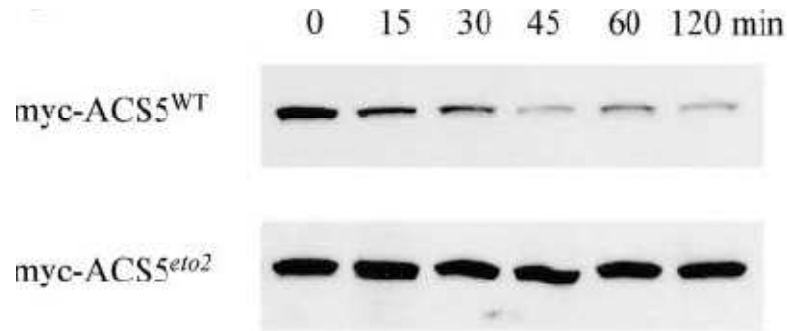
Table 1. Levels of ACS9 mRNA in Wild-Type and *eto3* Seedlings

Sample	C _T nor (Experiment 1) ^a	C _T nor (Experiment 2)	C _T nor (Experiment 3)	C _T nor (mean ± SD)	ACS9 mRNA ^b
Wild type	6.95	6.36	6.53	6.61 ± 0.30	1.0
<i>eto2</i>	7.23	6.99	6.07	6.76 ± 0.61	0.9

eto2 mutation does not affect specific activity of ACS5

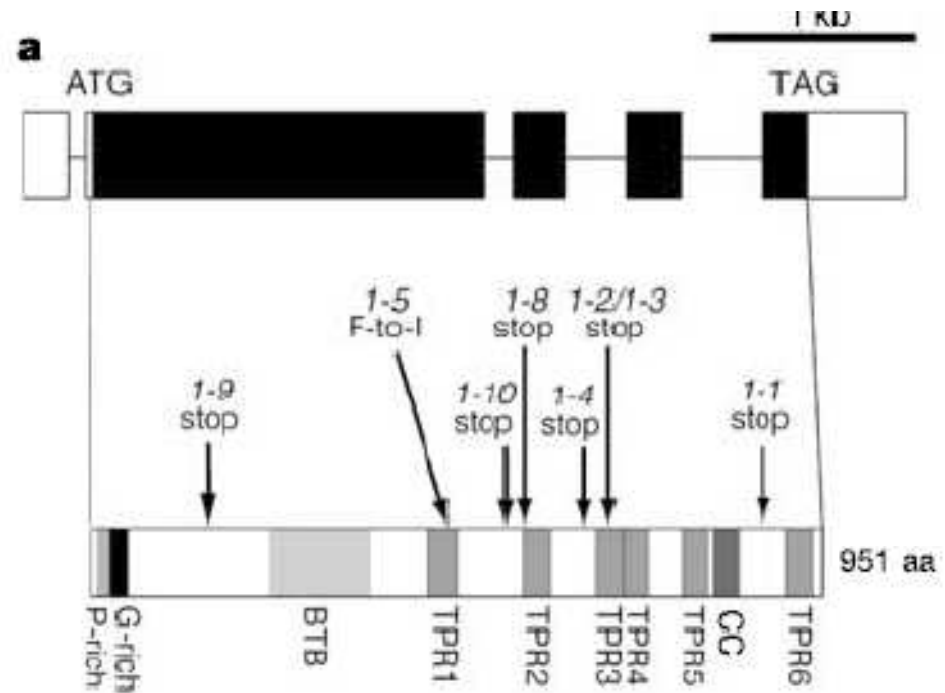
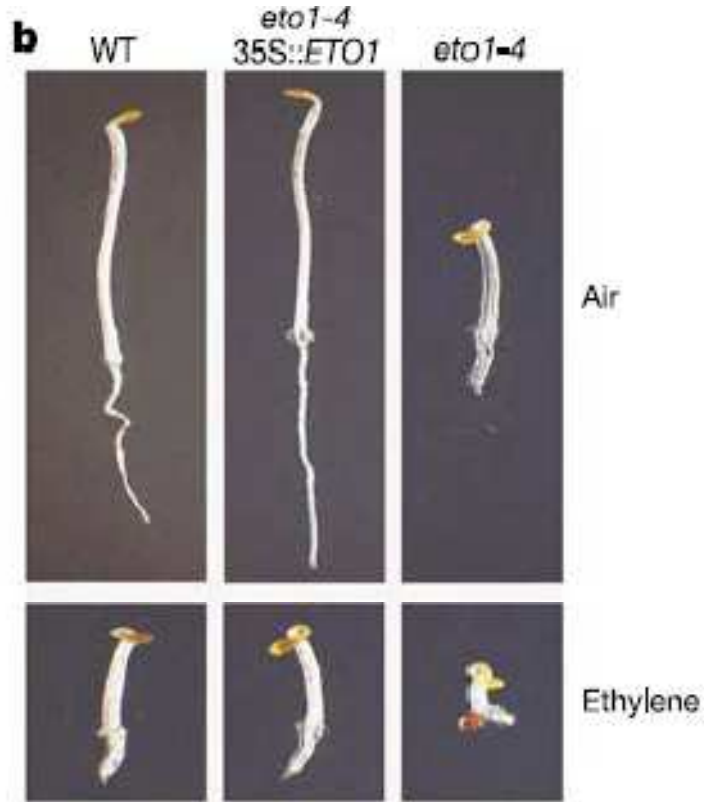


Effect of *eto2* mutation on ACS5 protein stability

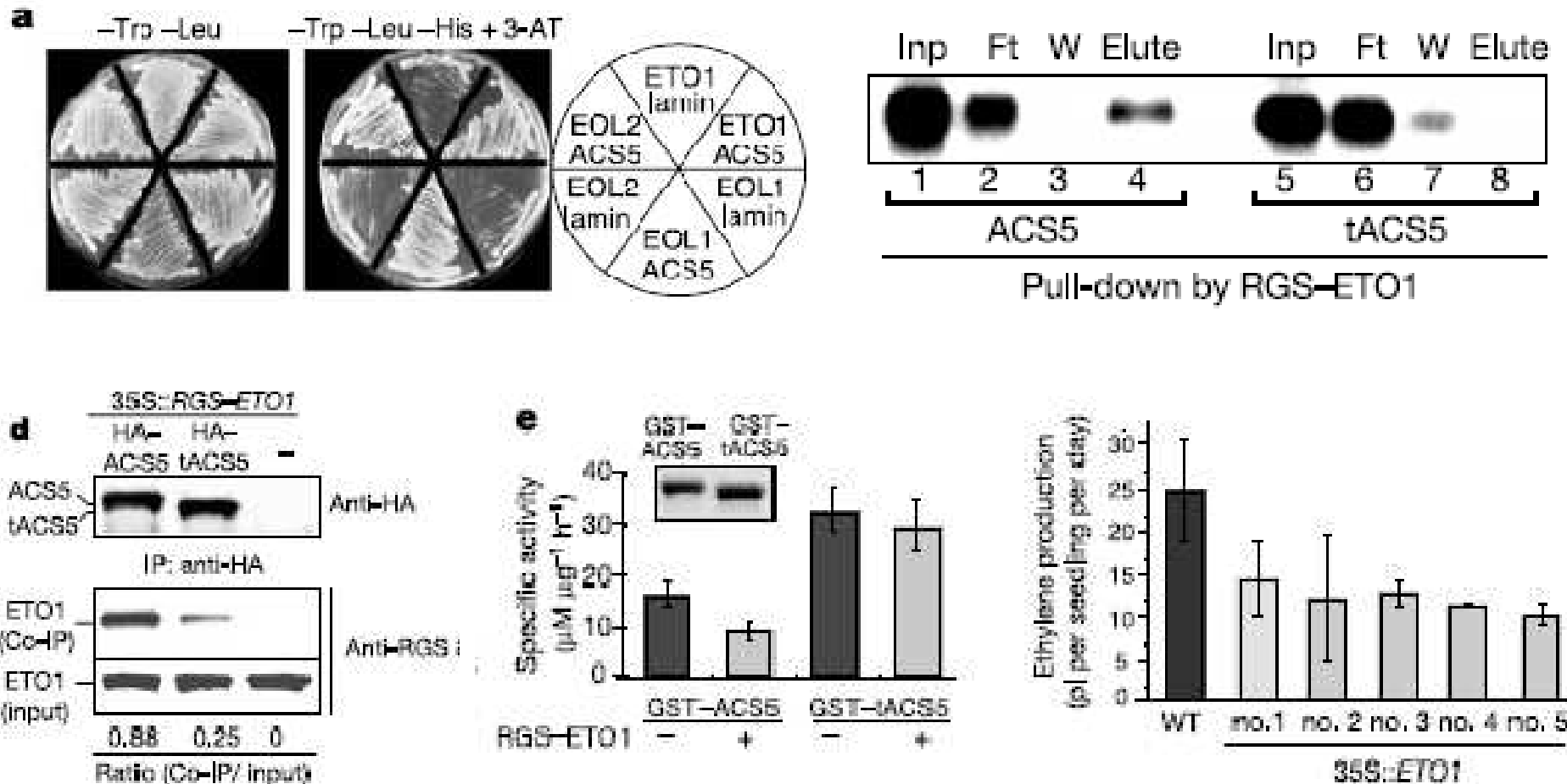


Posttranscriptional regulation of ACS

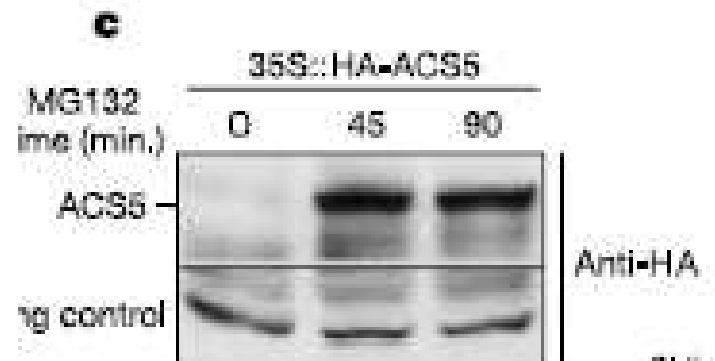
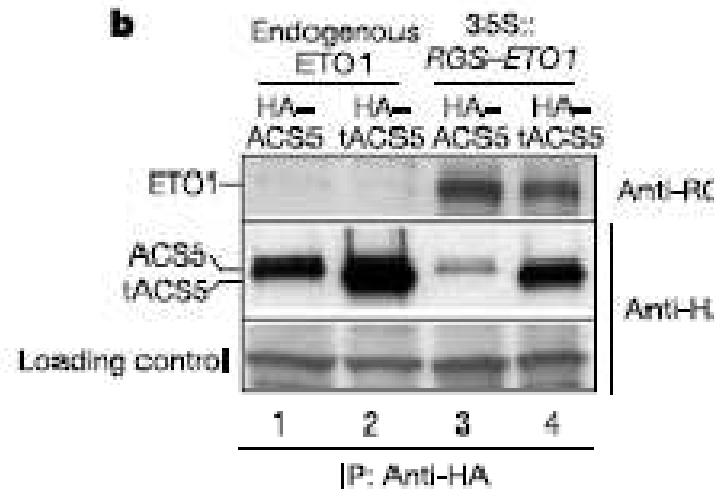
Eto1-recessive mutant with constitutive triple response



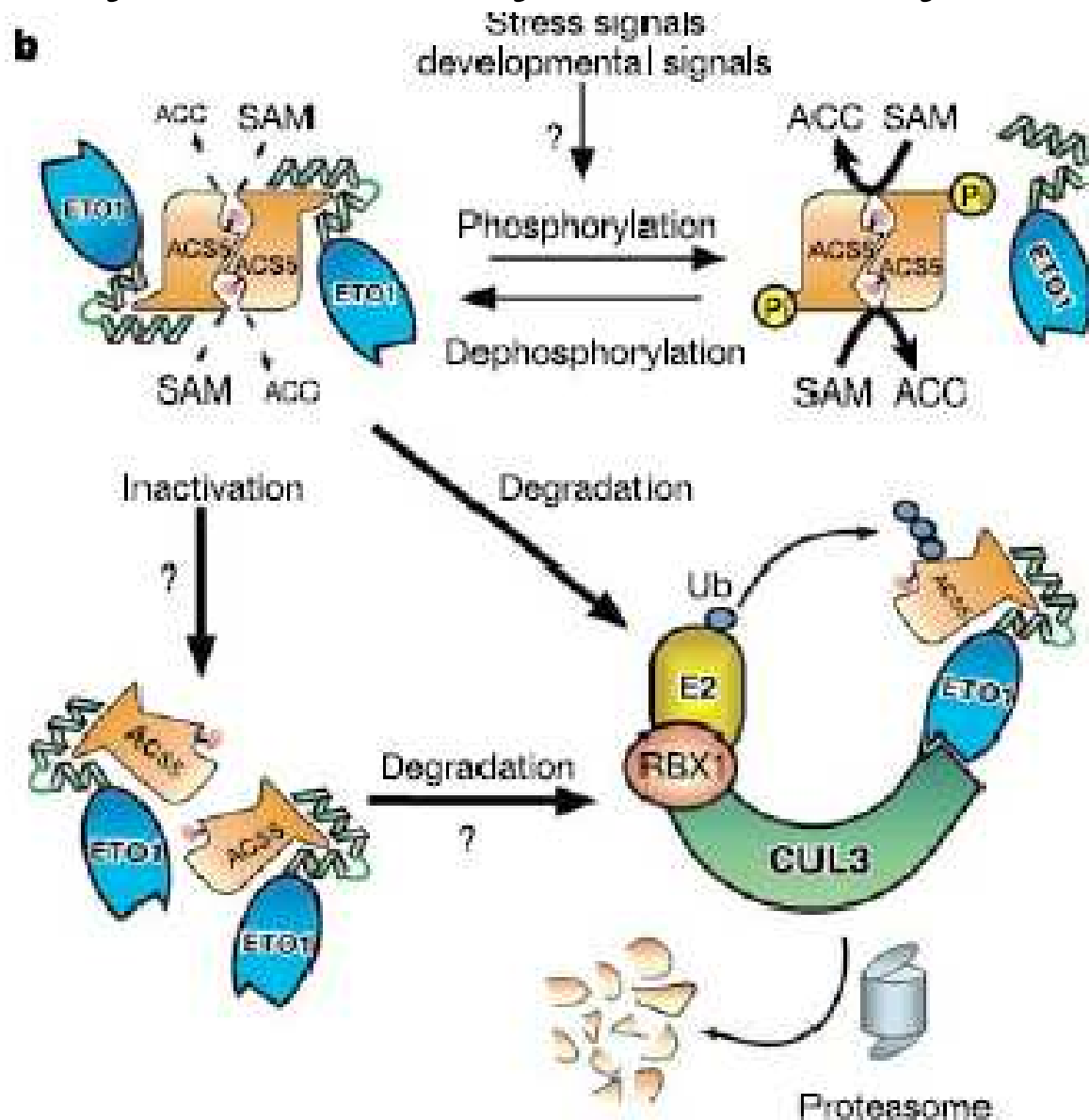
ETO1 interacts with ACS5 and regulates its activity



ETO1 promotes ACS5 degradation through proteasome dependent pathway

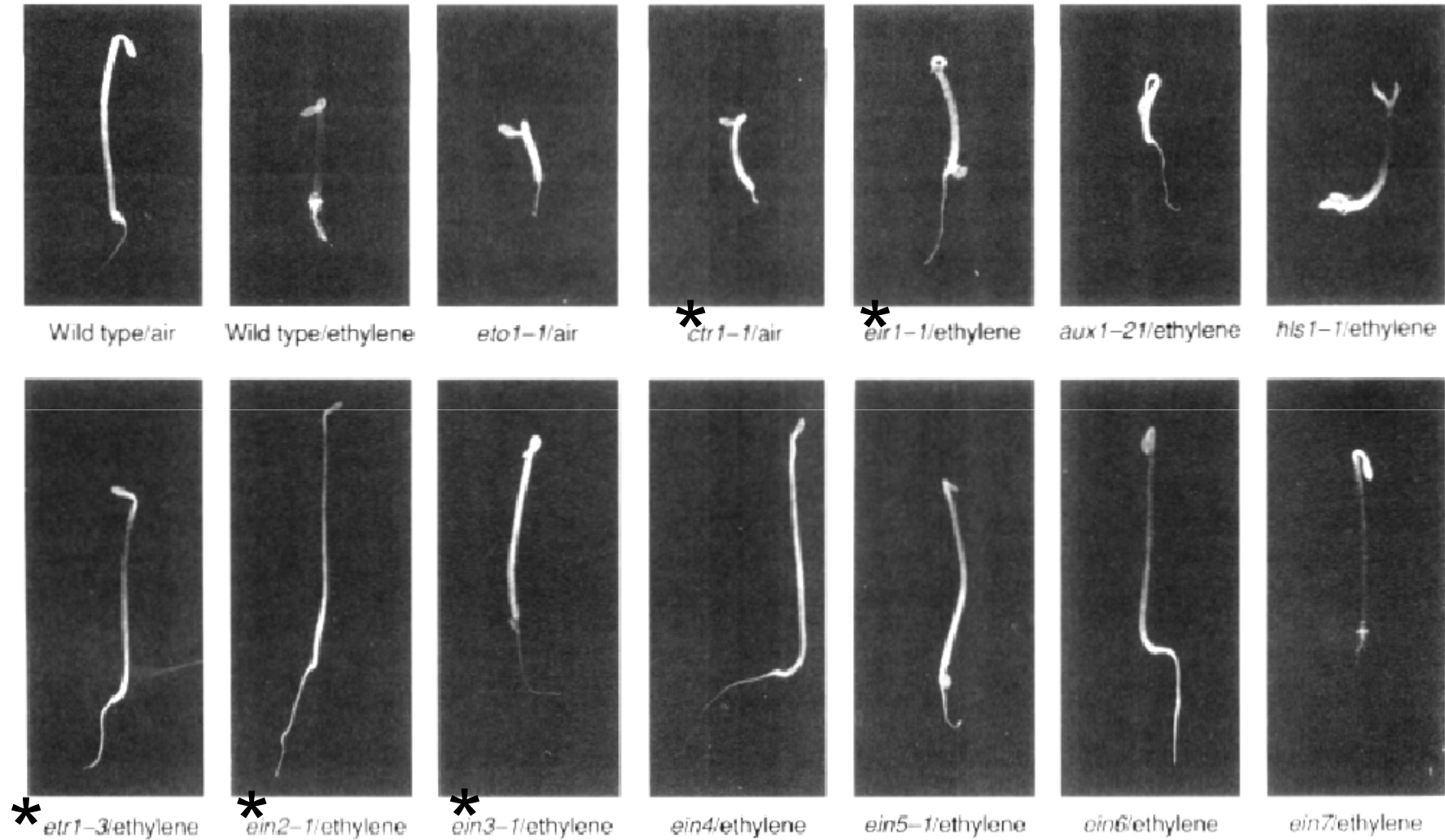


Model for regulation of ethylene biosynthesis by ETO1



Screen for ethylene mutants

II. Genes involved in signalling pathway



Roman et al., 1994

	Ecotype	Phenotype ^b
A. Strains ^a		
<i>aux1-7</i>	Columbia	Aux ⁻
<i>aux1-21</i>	Columbia	Aux ⁻
<i>aux1-22</i>	Columbia	Aux ⁻
* <i>ctr1-1</i>	Columbia	Ctr ⁻
<i>ctr1-5</i>	Wassilewskija	Ctr ⁻ , kan ^r
* <i>ein2-1</i>	Columbia	Ein ⁻
<i>ein2-6</i>	Wassilewskija	Ein ⁻
* <i>ein3-1</i>	Columbia	Ein ⁻
<i>ein3-2</i>	Wassilewskija	Ein ⁻ , kan ^r
* <i>ein4</i>	Columbia	Ein ⁻
<i>ein5-1</i>	Columbia	Ein ⁻
<i>ein5-2</i>	Columbia	Ein ⁻
<i>ein6</i>	Landsberg	Ein ⁻
<i>ein7</i>	Columbia	Ein ⁻
<i>eir1-1</i>	Columbia	Eir ⁻
<i>eir1-2</i>	Columbia	Eir ⁻
<i>eto1-1</i>	Columbia	Eto ⁻
* <i>etr1-3</i>	Columbia	Ein ⁻
<i>hls1-1</i>	Columbia	Hls ⁻
<i>ein2-1 tt4</i>	Mixed	Ein ⁻ , Tt ⁻
<i>eir1-1 ap1</i>	Mixed	Eir ⁻ , Ap ⁻
DP28	Landsberg	Dis ⁻ , Clv ⁻ , Tt ⁻
W2	Landsberg	Dis ⁻ , An ⁻
W100	Landsberg	Tt ⁻ , and more
M10	Landsberg	Ap ⁻ , Clv ⁻

Roman et al., 1994

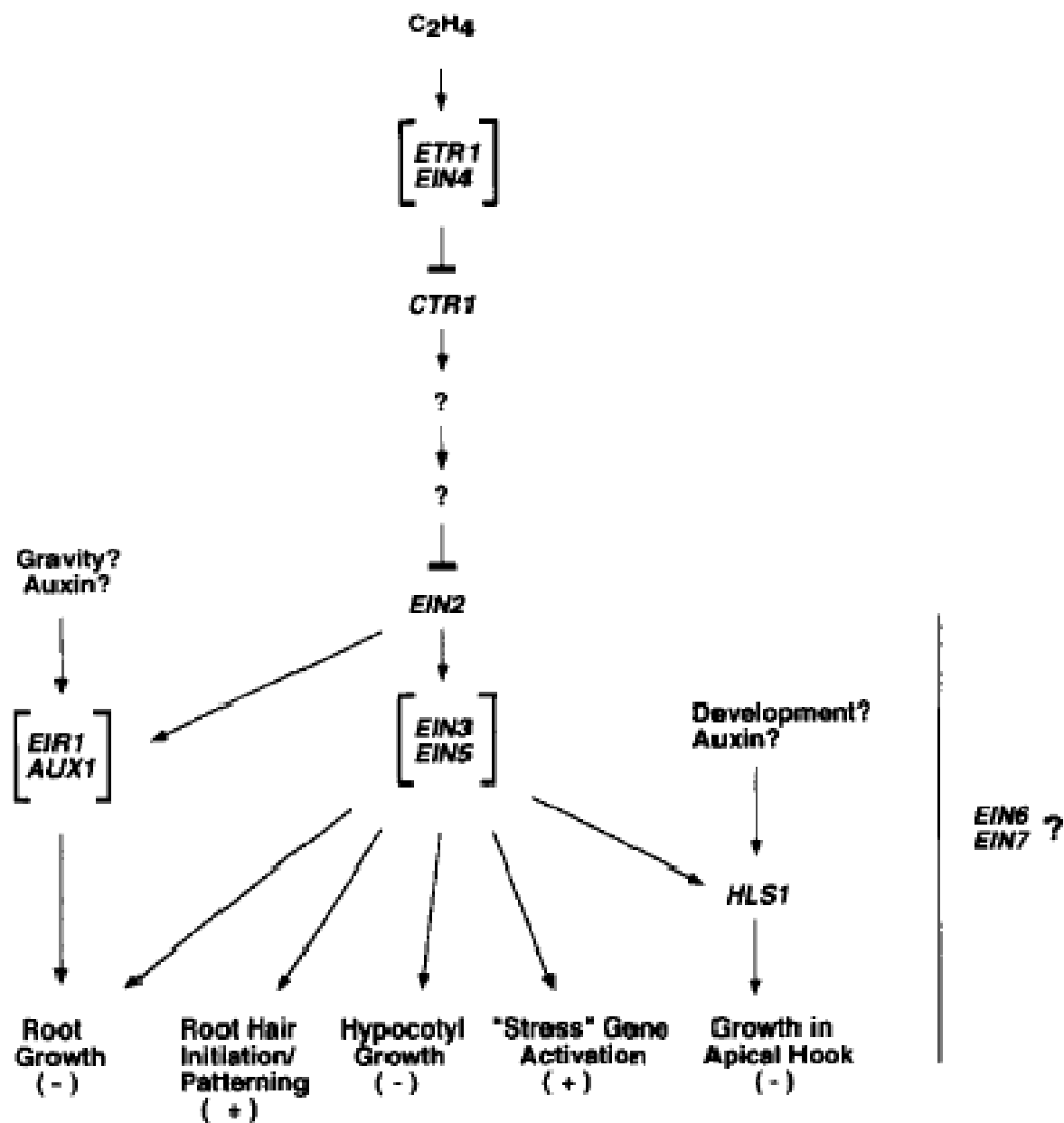
Quantifying the ethylene response phenotype

Strain	Ethylene				Air			
	Root length	Hypocotyl length	Total seedling	Hook angle	Root length	Hypocotyl length	Total seedling	Hook angle
Columbia	1.5 ± 0.1	3.0 ± 0.1	4.5 ± 0.1	250 ± 8	3.9 ± 0.2	4.8 ± 0.1	8.7 ± 0.2	114 ± 9
Landsberg	2.1 ± 0.1	2.7 ± 0.1	4.8 ± 0.1	233 ± 18	nd ^b	nd	nd	nd
Wasslewskija	1.0 ± 0.1	3.1 ± 0.1	4.1 ± 0.1	270 ± 6	4.0 ± 0.2	6.1 ± 0.1	10.1 ± 0.2	166 ± 7
<i>aux1-21</i>	4.7 ± 0.2	3.4 ± 0.1	8.1 ± 0.3	197 ± 8	6.3 ± 0.3	6.0 ± 0.1	12.3 ± 0.3	126 ± 6
<i>ctr1-1</i>	0.8 ± 0.0	2.8 ± 0.1	3.6 ± 0.1	247 ± 5	0.9 ± 0.0	3.1 ± 0.1	4.0 ± 0.1	246 ± 10
<i>ctr1-5</i>	0.4 ± 0.0	1.9 ± 0.1	2.3 ± 0.1	252 ± 5	0.6 ± 0.0	2.4 ± 0.1	3.0 ± 0.1	237 ± 8
<i>ein2-1</i>	6.1 ± 0.2	6.5 ± 0.2	12.6 ± 0.2	39 ± 4	5.8 ± 0.2	6.9 ± 0.2	12.7 ± 0.3	43 ± 7
<i>ein3-1</i>	3.6 ± 0.1	5.2 ± 0.1	8.8 ± 0.2	118 ± 7	5.4 ± 0.3	5.9 ± 0.1	11.3 ± 0.4	77 ± 7
<i>ein3-2</i>	3.1 ± 0.1	5.5 ± 0.1	8.5 ± 0.2	176 ± 6	5.2 ± 0.3	6.3 ± 0.2	11.4 ± 0.3	152 ± 8
<i>ein4</i>	7.1 ± 0.2	7.3 ± 0.3	14.4 ± 0.3	64 ± 9	6.8 ± 0.3	6.9 ± 0.3	13.7 ± 0.4	45 ± 5
<i>ein5-1</i>	2.5 ± 0.1	4.8 ± 0.1	7.3 ± 0.1	144 ± 10	5.6 ± 0.2	5.3 ± 0.2	11.0 ± 0.3	89 ± 8
<i>ein5-2</i>	2.6 ± 0.1	4.6 ± 0.2	7.2 ± 0.2	156 ± 10	4.3 ± 0.2	5.6 ± 0.2	9.9 ± 0.3	113 ± 10
<i>ein6</i>	3.5 ± 0.1	6.2 ± 0.2	9.7 ± 0.2	95 ± 6	7.0 ± 0.2	6.0 ± 0.2	13.0 ± 0.2	47 ± 4
<i>ein7</i>	2.9 ± 0.1	5.2 ± 0.1	8.1 ± 0.2	176 ± 4	5.2 ± 0.2	6.7 ± 0.2	11.9 ± 0.2	137 ± 8
<i>eir1-1</i>	3.4 ± 0.1	3.1 ± 0.1	6.5 ± 0.1	282 ± 7	5.2 ± 0.9	6.2 ± 0.1	11.4 ± 0.1	106 ± 7
<i>eir1-2</i>	3.0 ± 0.1	3.1 ± 0.1	6.1 ± 0.1	261 ± 7	4.8 ± 0.3	5.7 ± 0.2	10.5 ± 0.4	109 ± 10
<i>eto1-1</i>	1.4 ± 0.1	3.3 ± 0.1	4.7 ± 0.2	244 ± 10	1.9 ± 0.1	3.3 ± 0.1	5.3 ± 0.1	239 ± 8
<i>etr1-3</i>	4.6 ± 0.2	6.1 ± 0.1	10.7 ± 0.3	89 ± 7	4.9 ± 0.3	5.5 ± 0.2	10.4 ± 0.3	96 ± 5
<i>hls1-1</i>	0.9 ± 0.1	3.4 ± 0.1	4.3 ± 0.2	5 ± 1	3.3 ± 0.1	5.1 ± 0.1	8.4 ± 0.2	4 ± 1

Roman et al., 1994

B. Double mutants^a

<i>aux1-21 ctr1-1</i>	Aus ⁻ , Ctr ^{-c}
<i>aux1-21 eir1-1</i>	Aux ⁻
<i>ctr1-5 ein 2-1</i>	Ein ⁻
<i>ctr1-1 ein3-2</i>	Ein ⁻
<i>ctr1-1 ein5-1</i>	Ein ⁻
<i>ctr1-1 ein7</i>	Ein ⁻
<i>ctr1-1 eir1-1</i>	Eir ⁻ , Ctr ^{-c}
<i>ctr1-1 etr1-3</i>	Ctr ⁻
<i>ctr1-1 hls1-1</i>	Hls ⁻ , Ctr ^{-c}
<i>ein2-1 eir1-1</i>	Ein ⁻ , Eir ⁻
<i>ein2-6 eir1-1</i>	Ein ⁻ , Eir ⁻
<i>ein2-1 eto1-1</i>	Ein ⁻
<i>ein2-6 eto 1-1</i>	Ein ⁻
<i>ein2-1 etr1-3</i>	Ein ⁻
<i>ein2-1 hls1-1</i>	Ein ⁻ , Hls ⁻
<i>ein3-1 eir1-1</i>	Ein ⁻ , Eir ⁻
<i>ein5-1 eir1-2</i>	Ein ⁻ , Eir ⁻
<i>eir1-1 hls 1-1</i>	Eir ⁻ , Hls ⁻



Roman et al., 1994

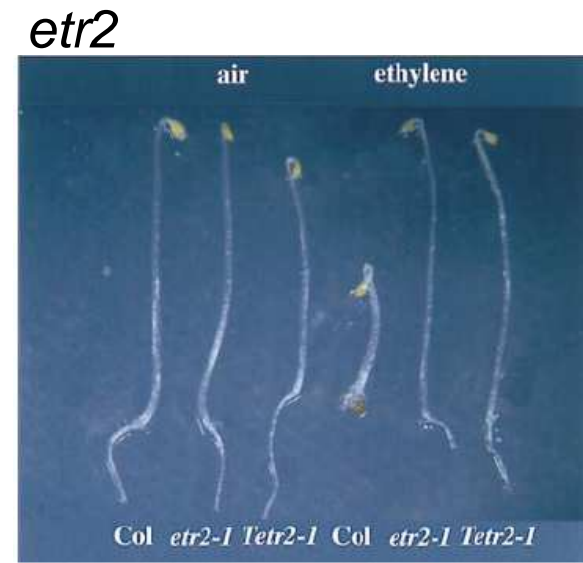
ETR1 codes for histidine kinase



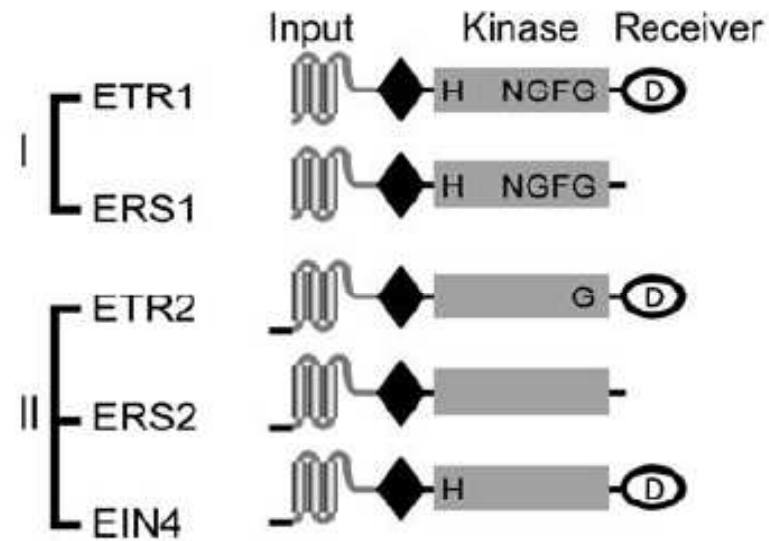
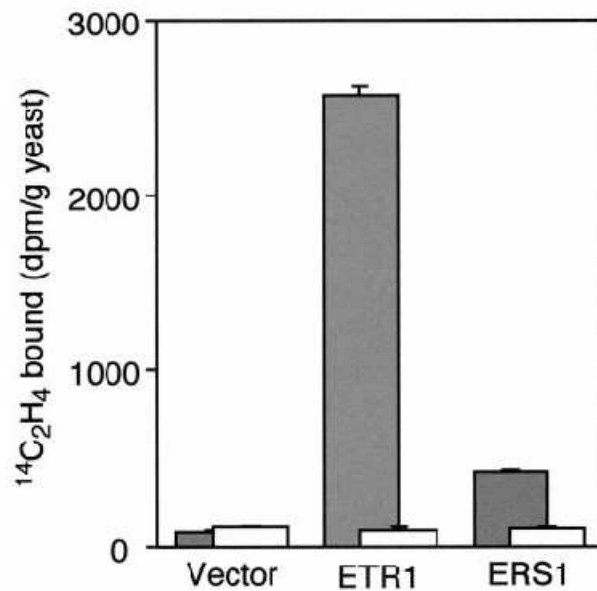
Bleecker et al., 1988



Hua et al., 1998

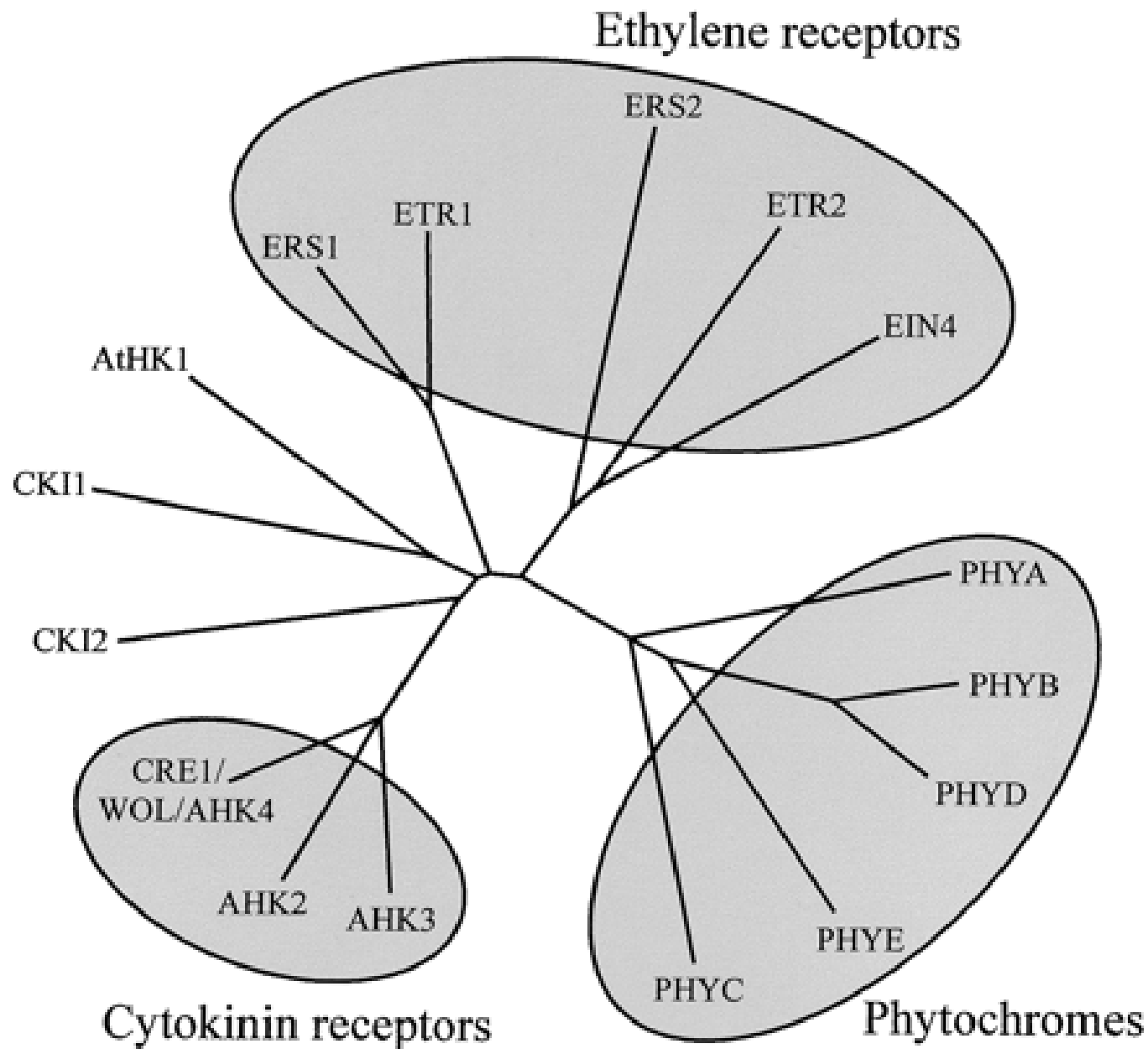


Sakai et al., 1998



... Ethylene signal transduction. In the absence of ethylene...

His-kinases in Arabidopsis

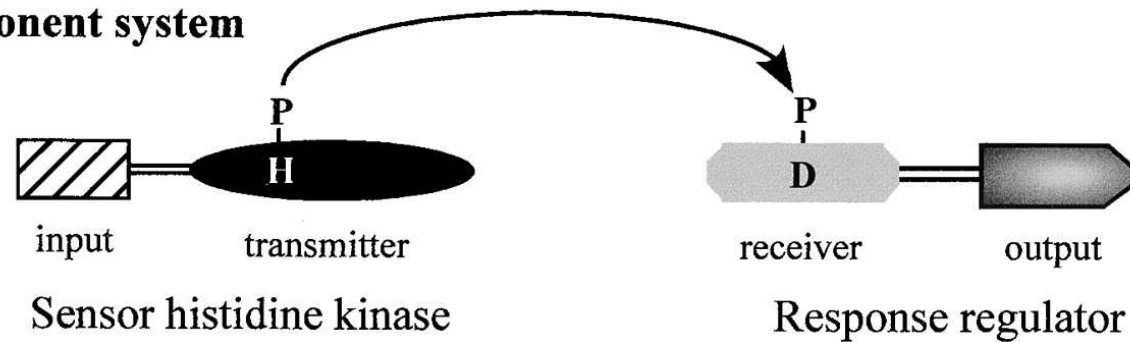


Ethylene signalling

– homology to two component system ?

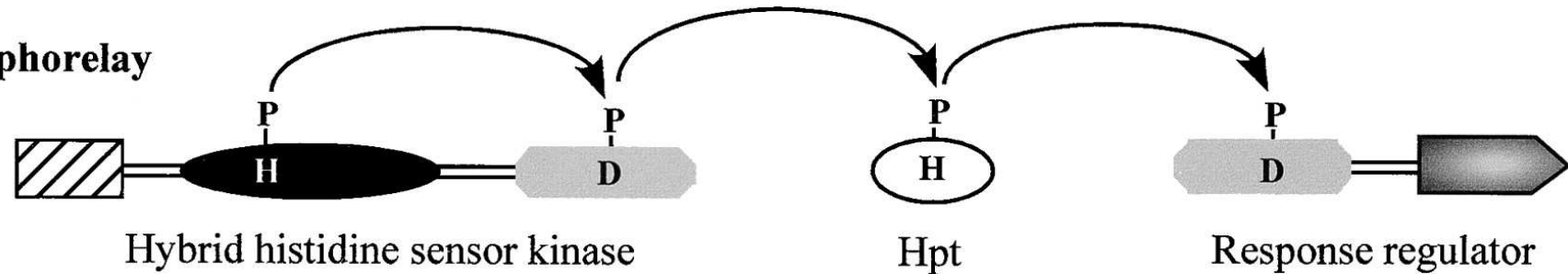
A

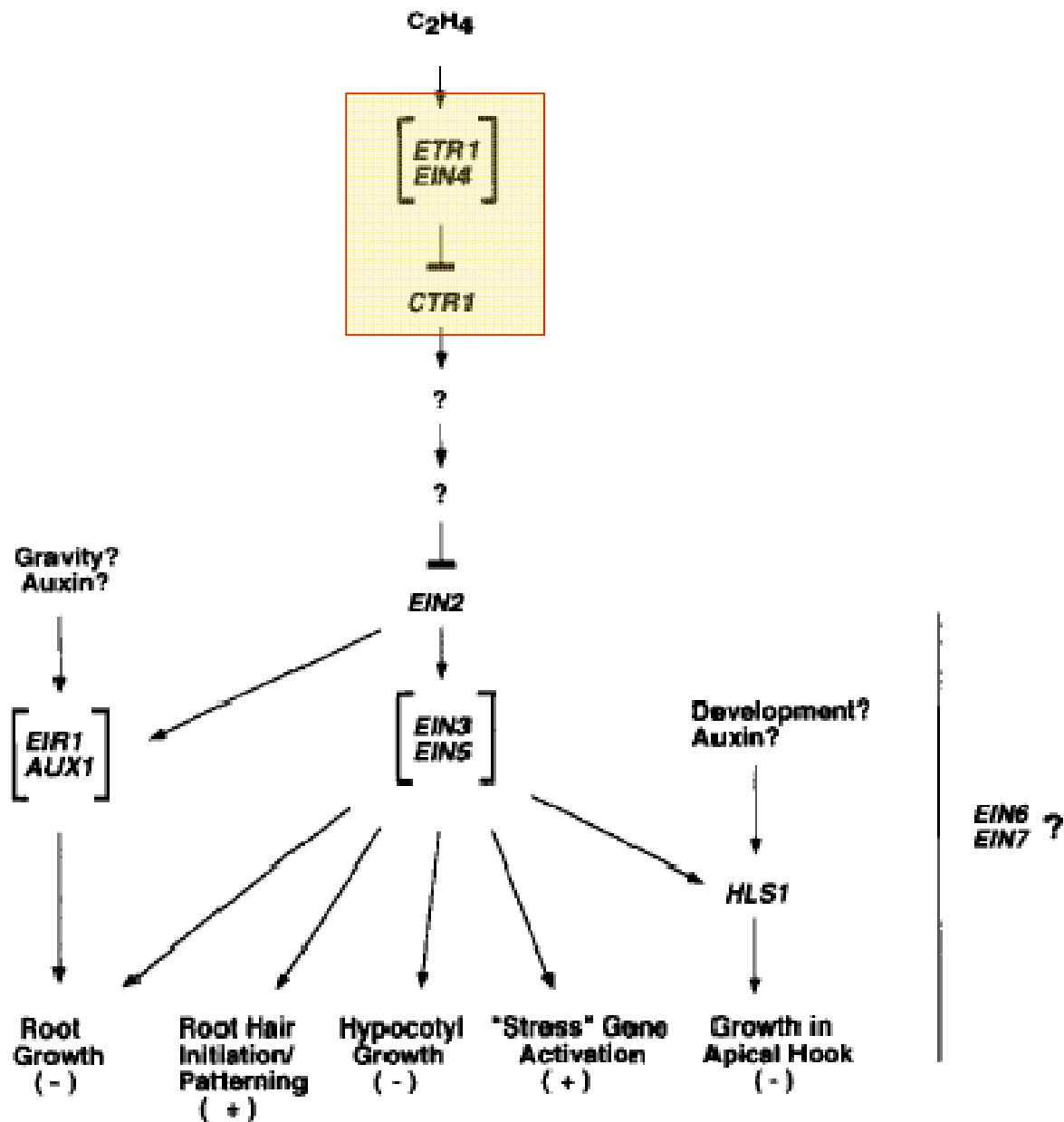
Simple two-component system



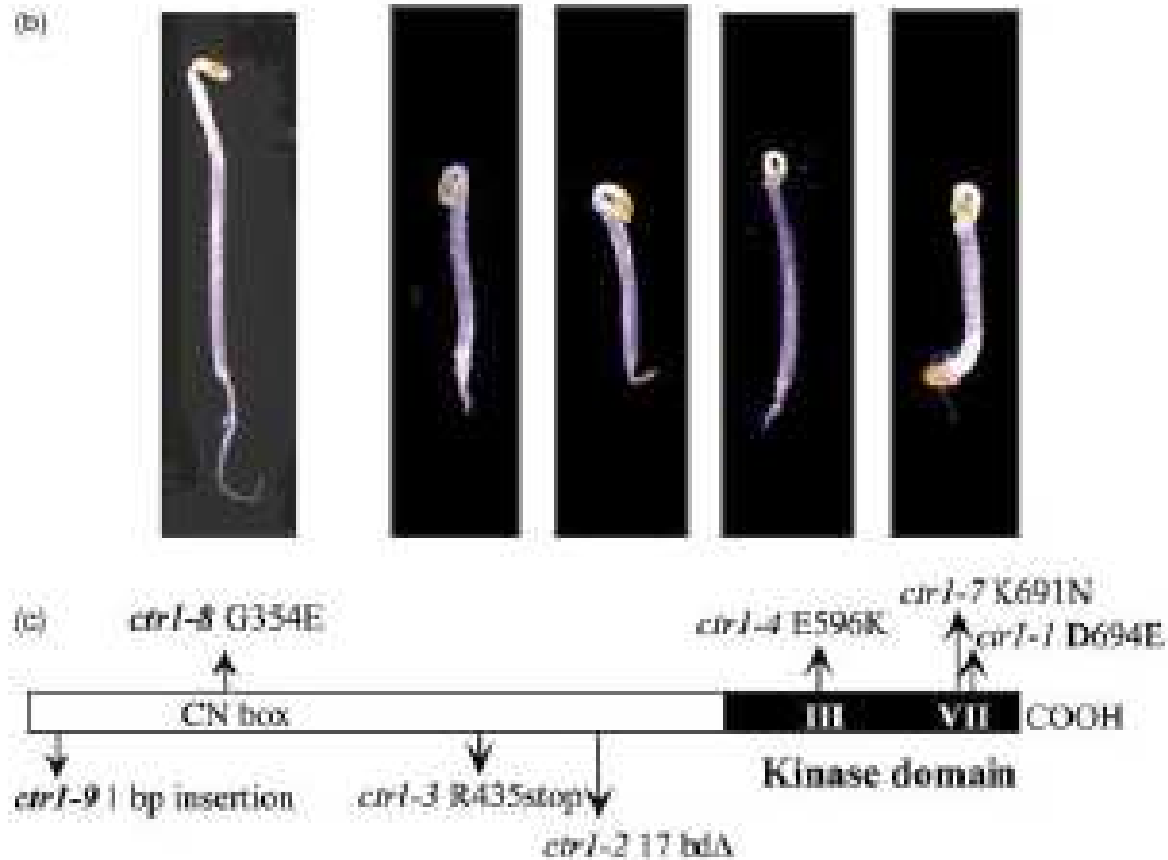
B

Phosphorelay

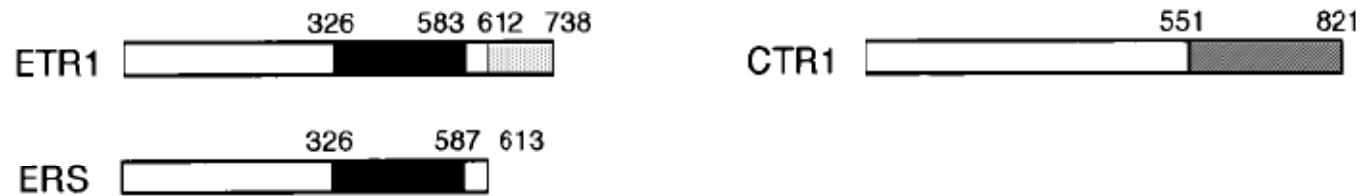




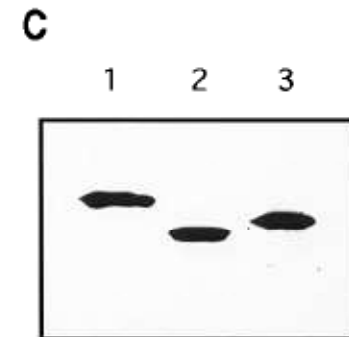
Ctr1 – codes for protein kinase of Raf family



ETR1 interacts with CTR1

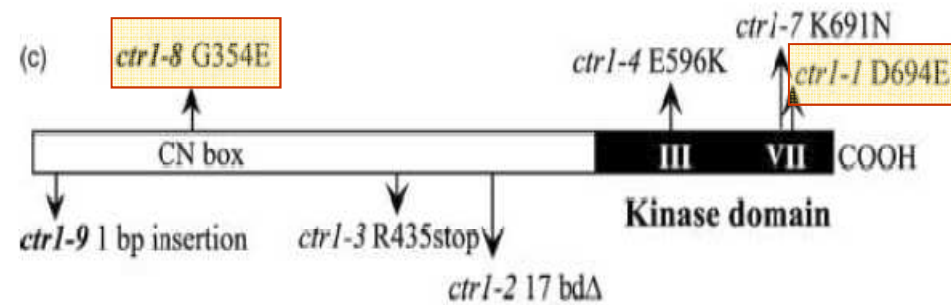
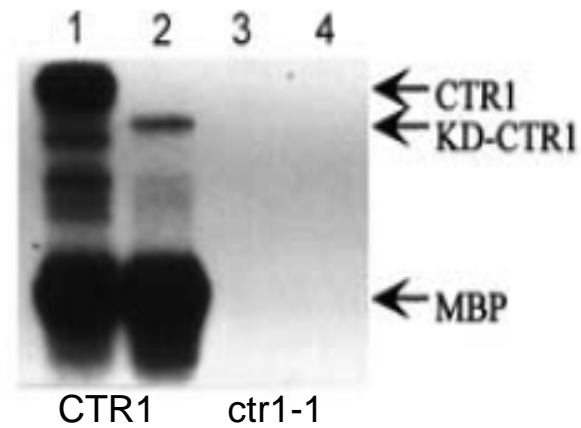


	DB FUSION	AD FUSION	HIS	lacZ	β -gal units
1	ETR1 293-729	CTR1 53-568			71 \pm 5.0
	"	CTR1 538-821			0.10 \pm 0.02
	"	vector			0.07 \pm 0.02
2	ETR1 293-610	CTR1 53-568			0.10 \pm 0.02
	"	vector			0.04 \pm 0.00
3	ERS 261-613	CTR1 53-568			4.4 \pm 0.20
	"	vector			0.05 \pm 0.01
	lamin	CTR1 53-568			0.05 \pm 0.01



Clark et al., 1998

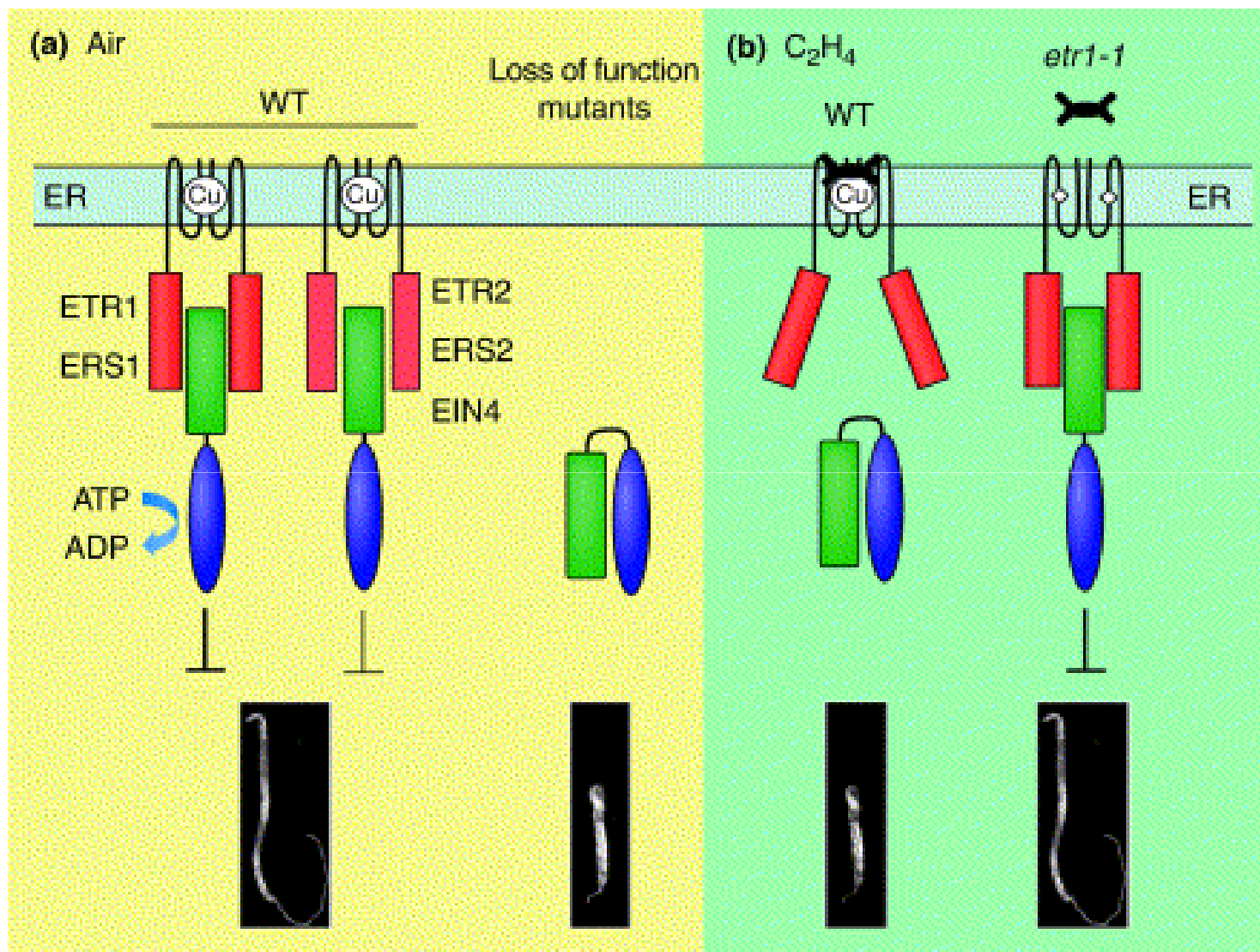
CTR1 has protein kinase activity

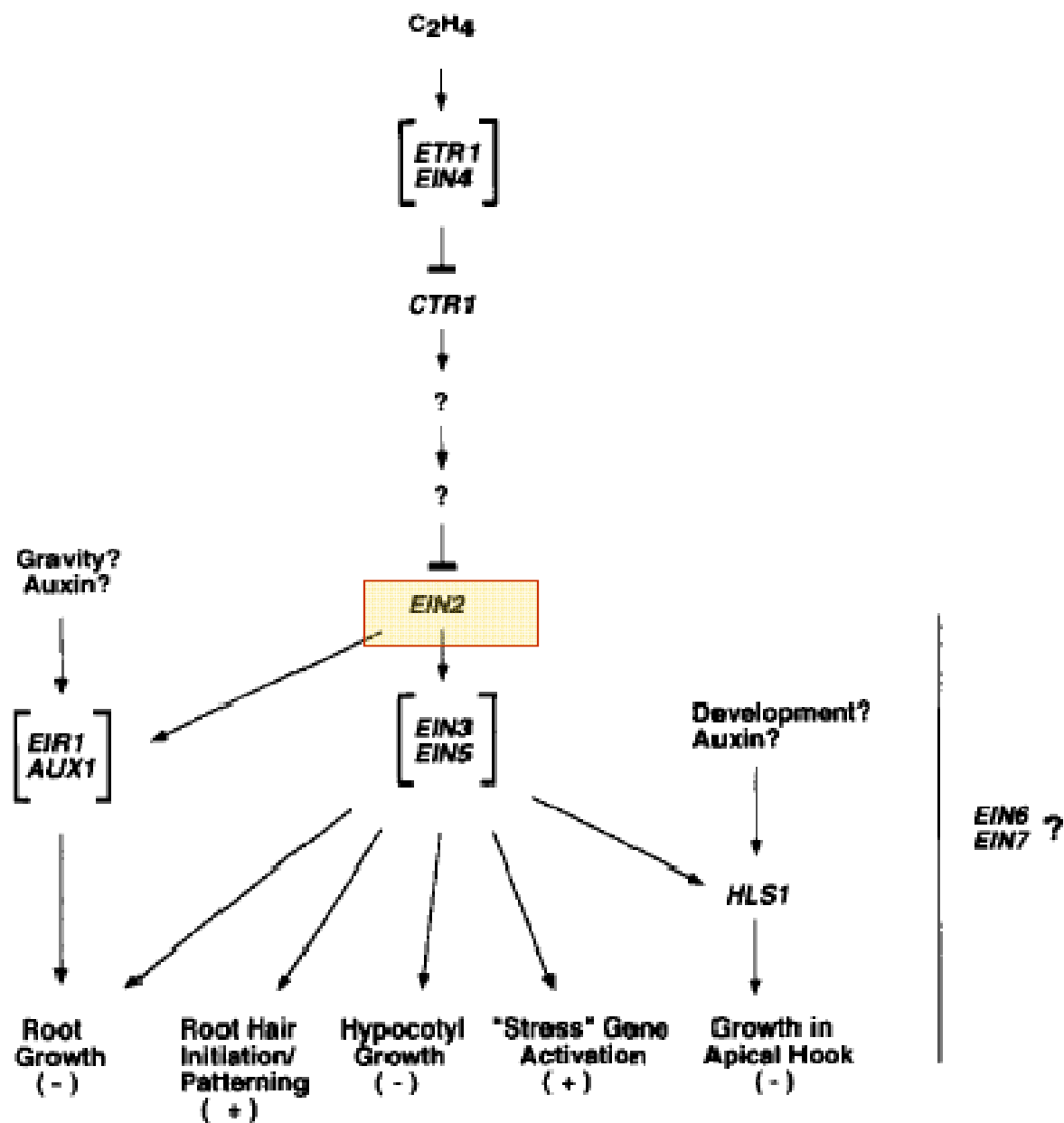




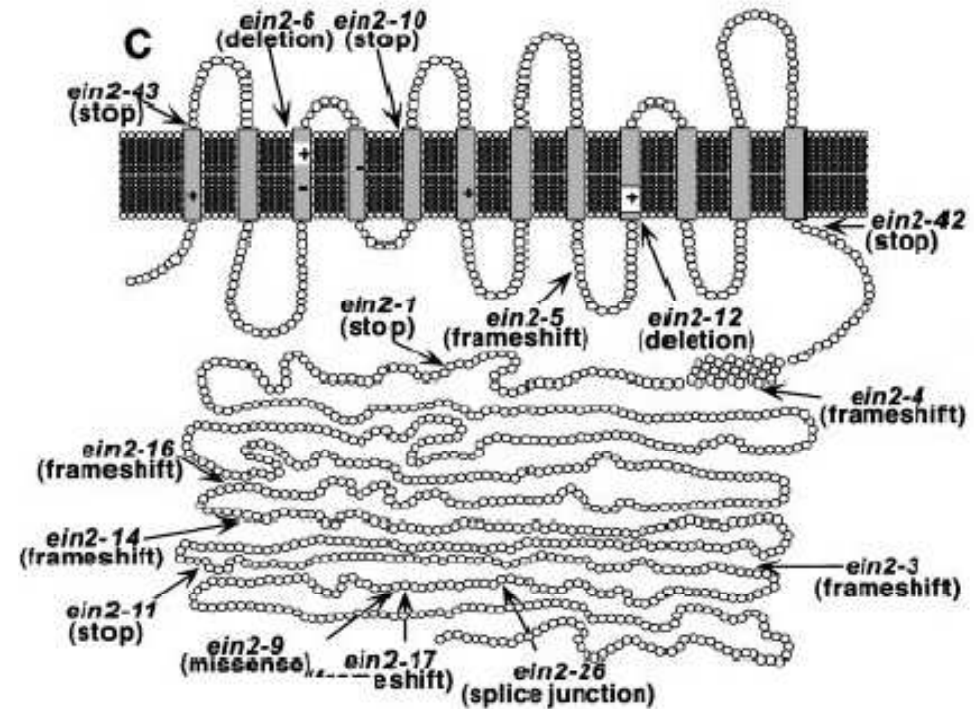
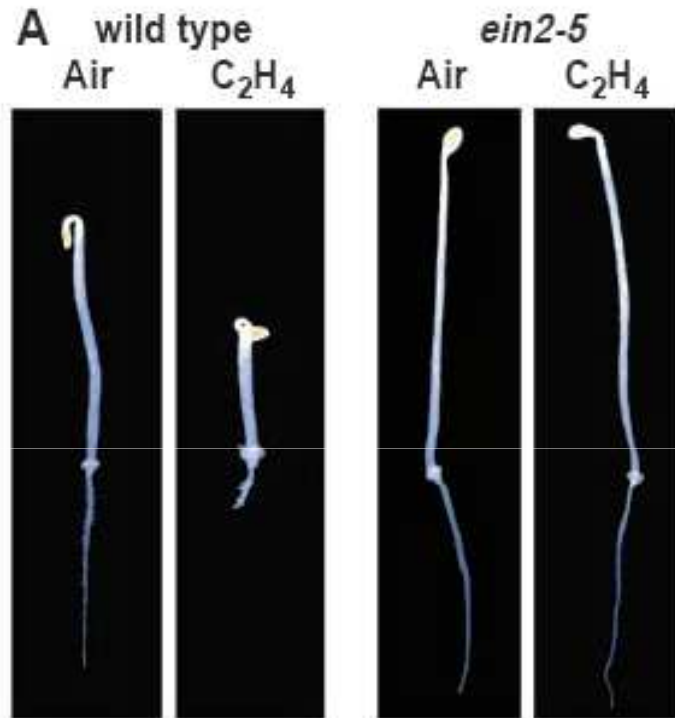
Current Biology

- ethylene receptor dominant mutation > ethylene insensitivity
- ethylene receptor loss of function mutation > constitutive ethylene response
- ctr1* loss of function mutation > constitutive ethylene response

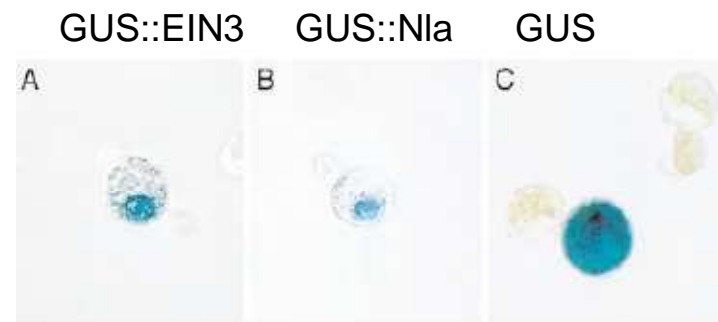
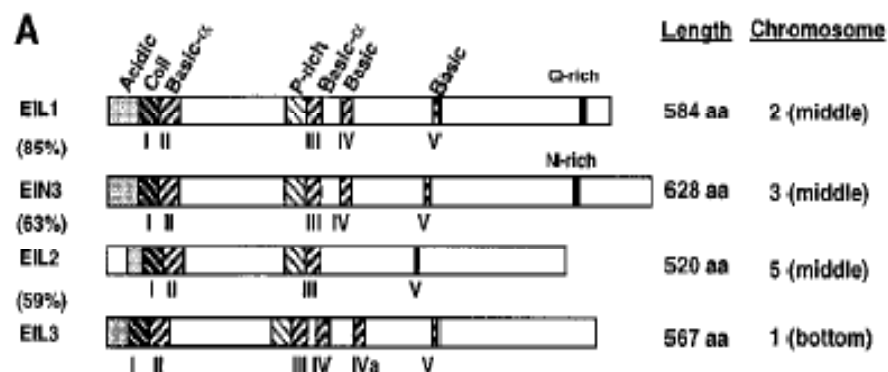
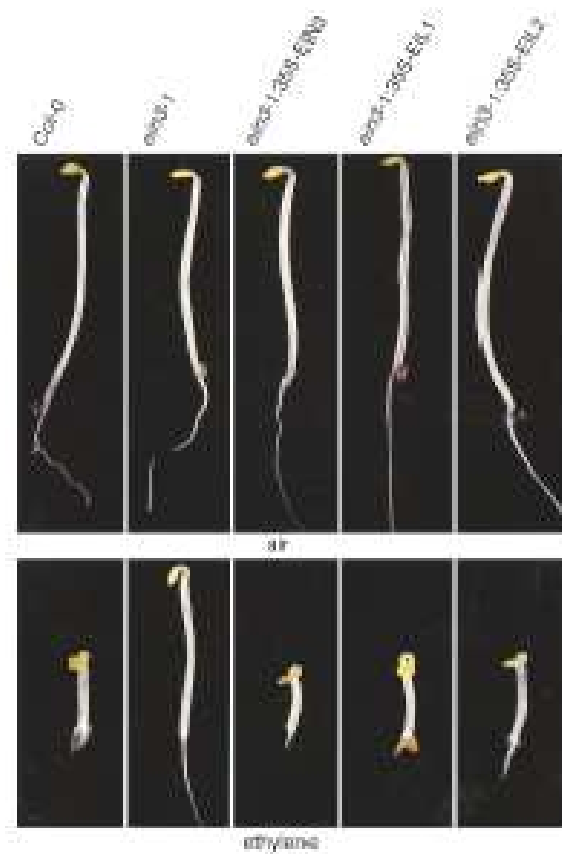
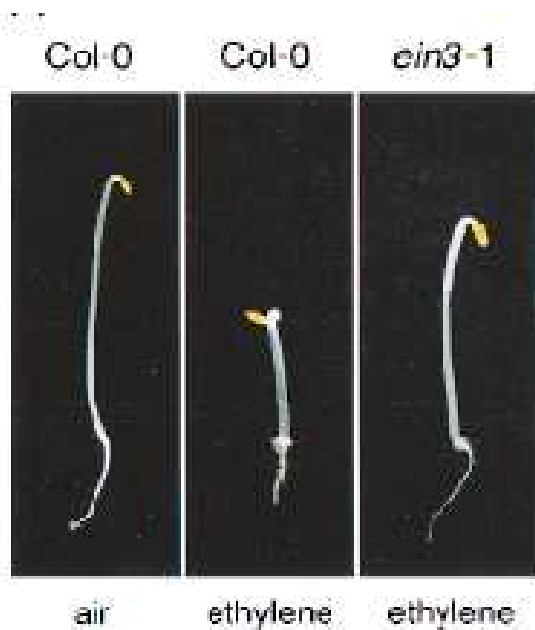




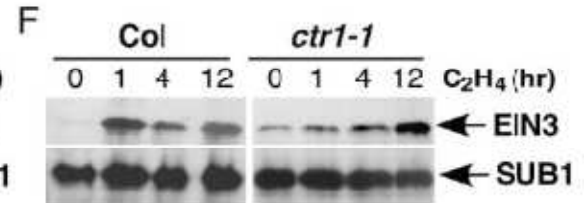
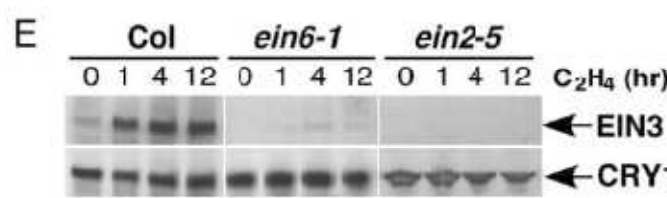
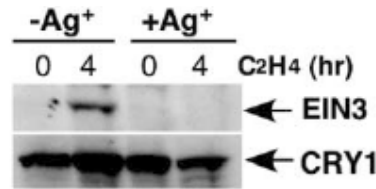
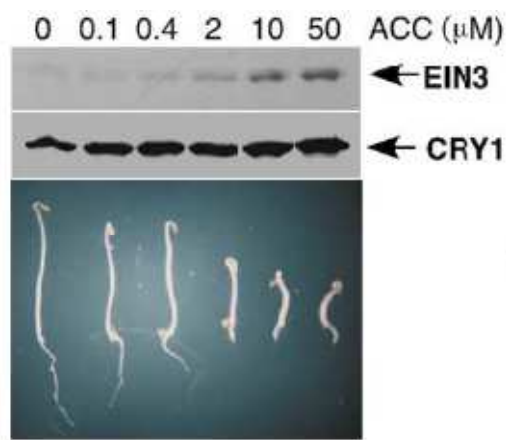
EIN2 - contains domain similar to Nramp metal transporters protein.



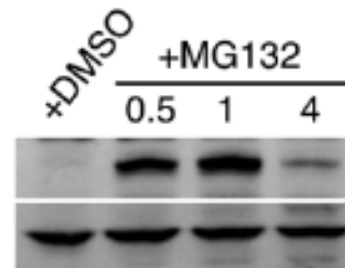
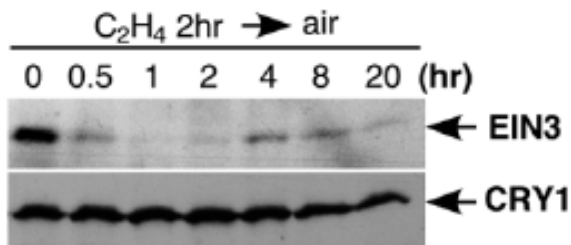
EIN3 codes for transcription factor



EIN3 protein level is controlled by ethylene

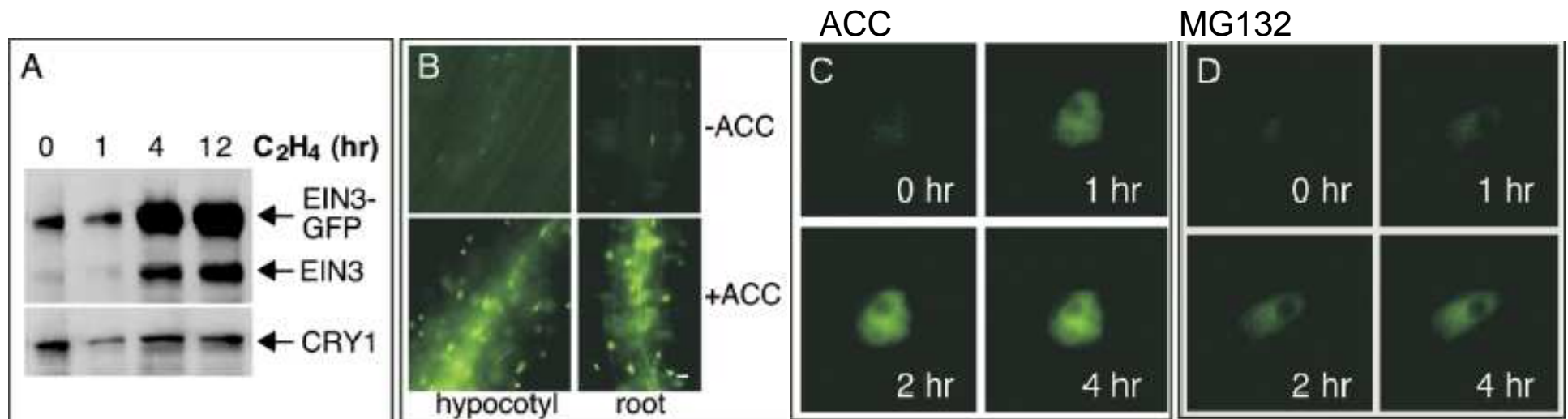


EIN3 is rapidly degraded by a proteasome-mediated pathway

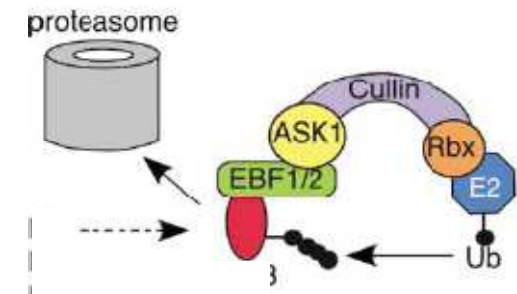
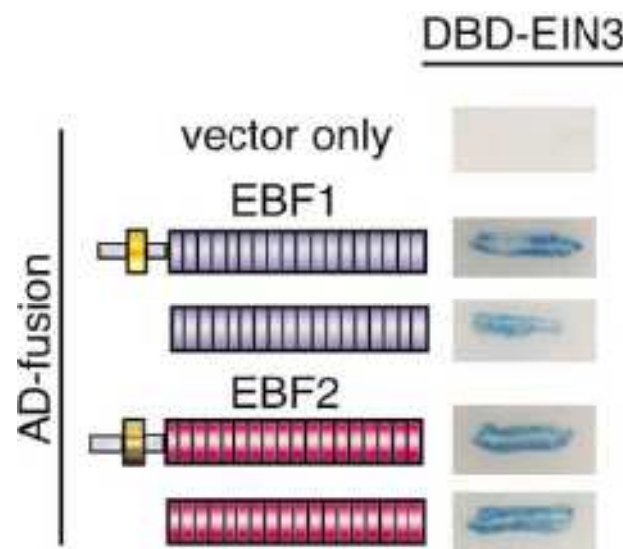
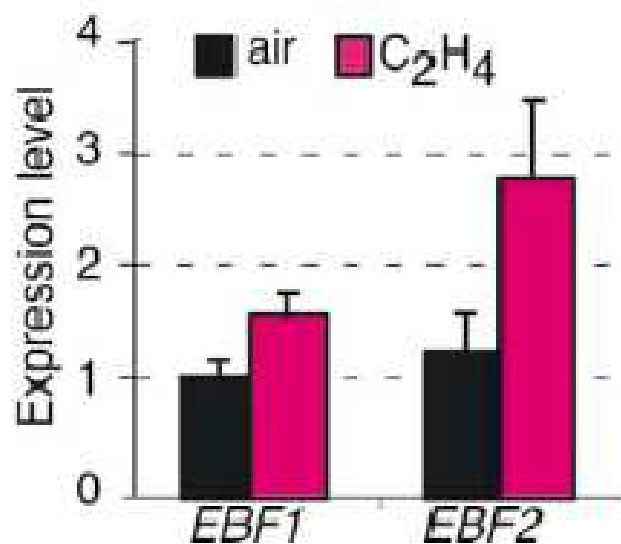


Guo and Ecker, 2003

EIN3 accumulates in nucleus upon ethylene or MG132 treatment

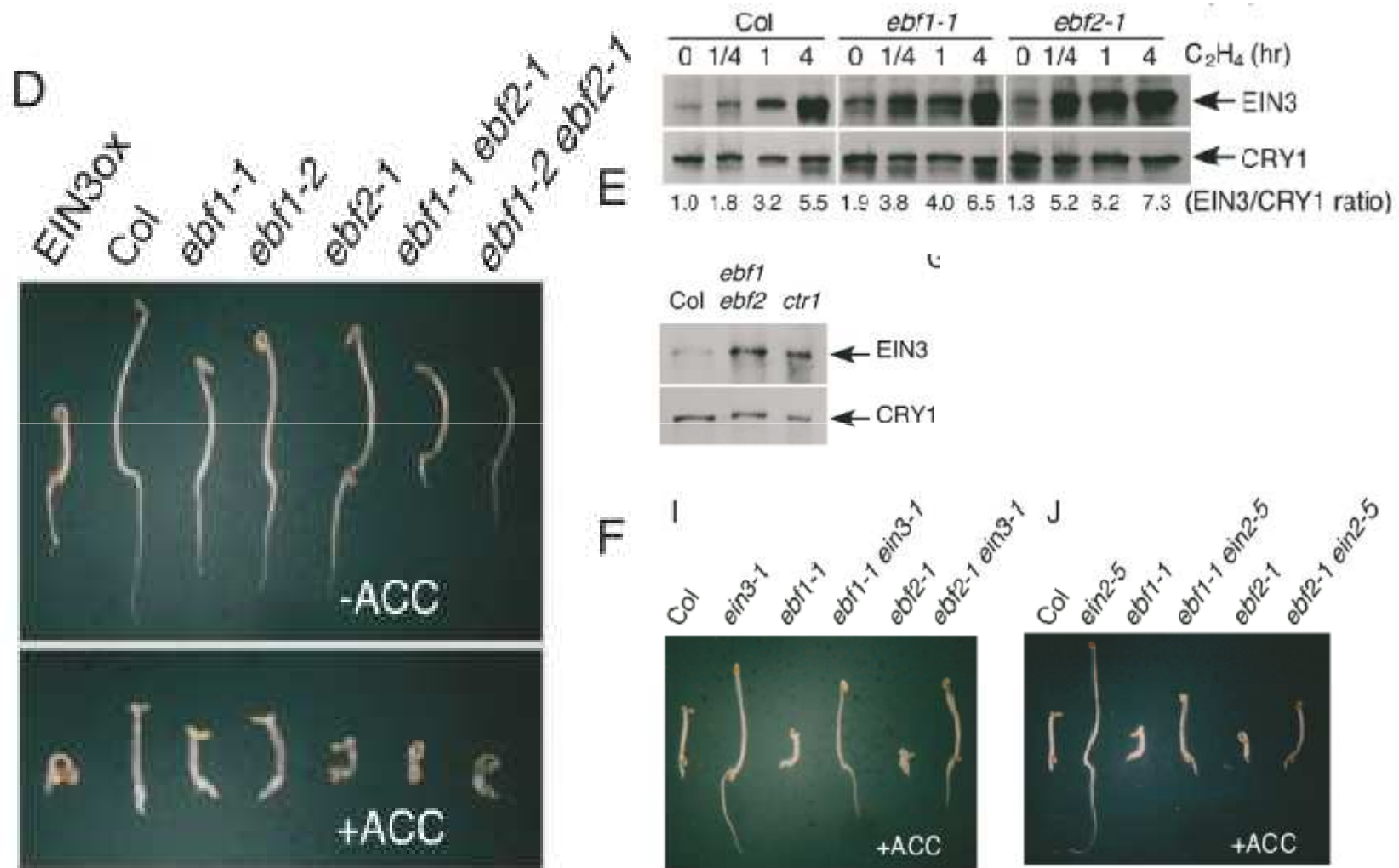


EIN3 interacts with F-box proteins

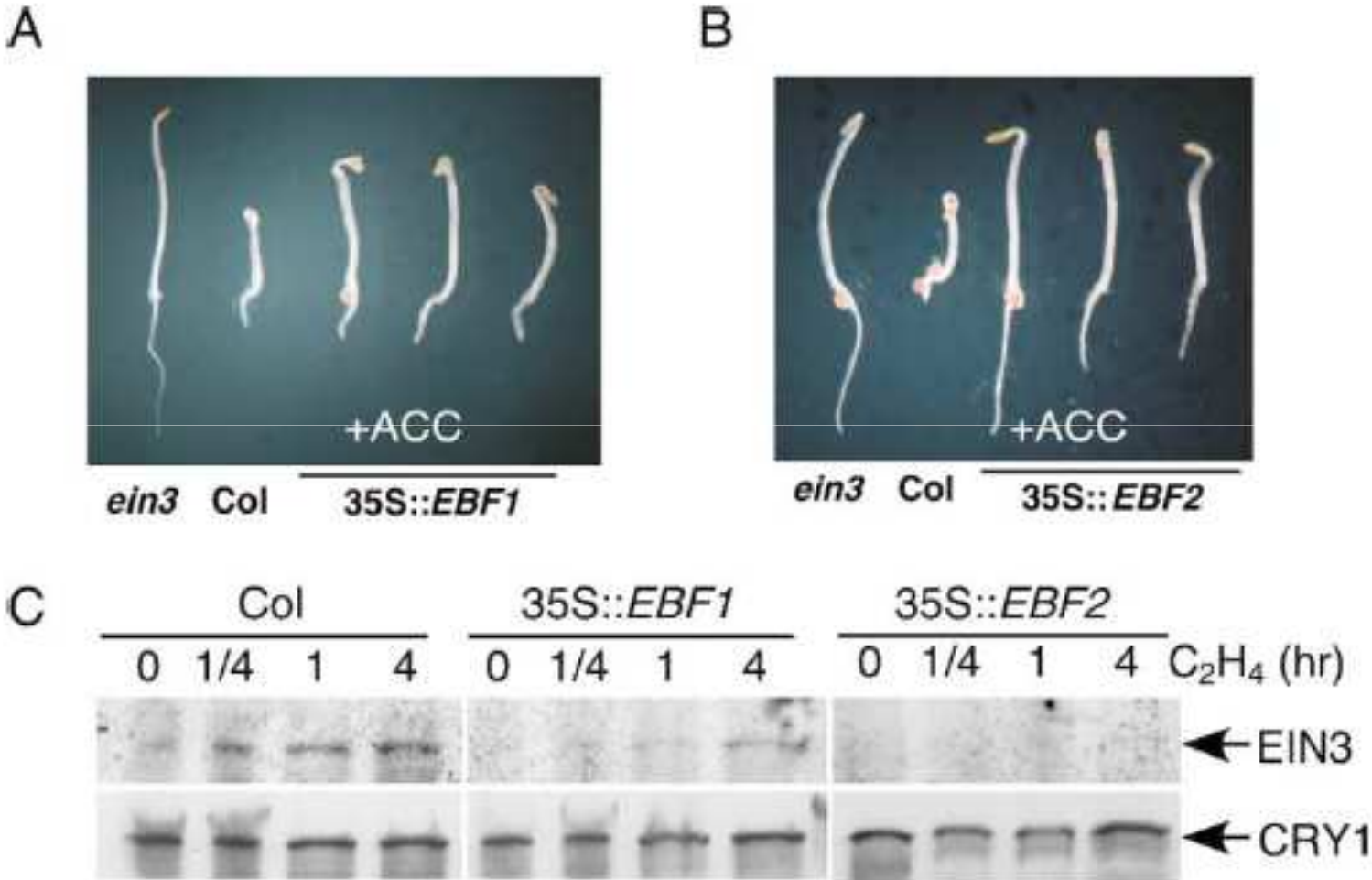


Guo and Ecker, 2003

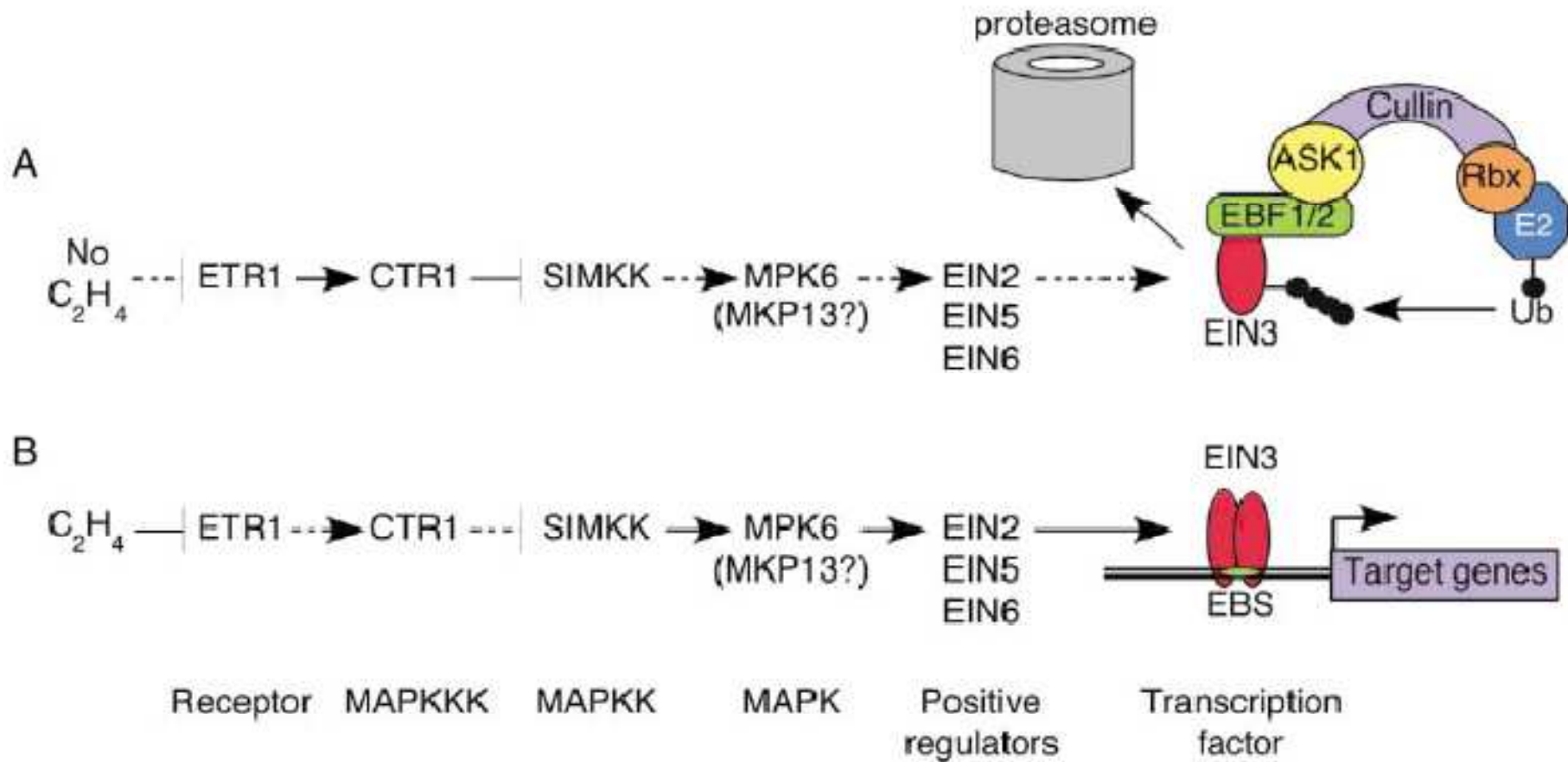
Mutation in EBF1 and EBF2 results in hypersensitivity to ethylene and accumulation of EIN3 protein



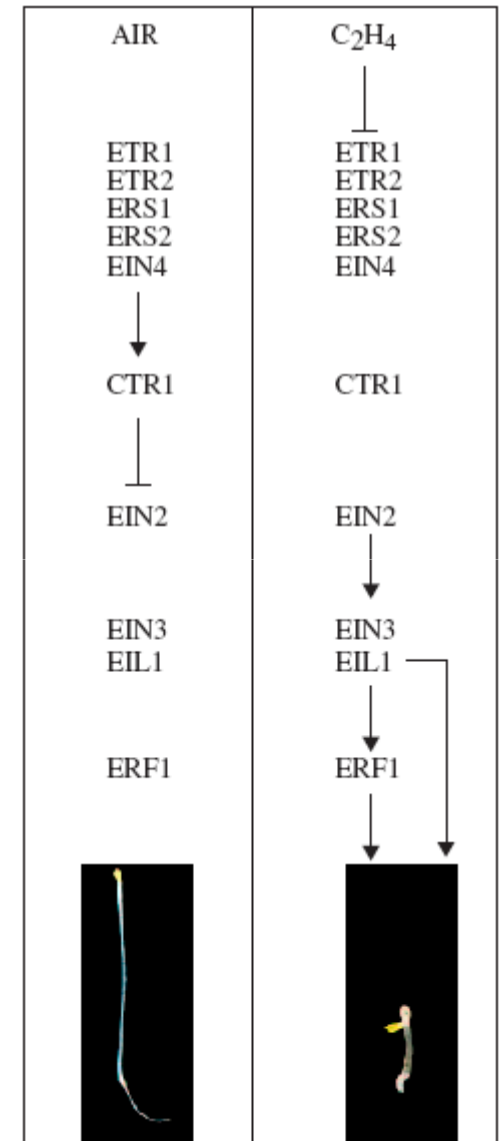
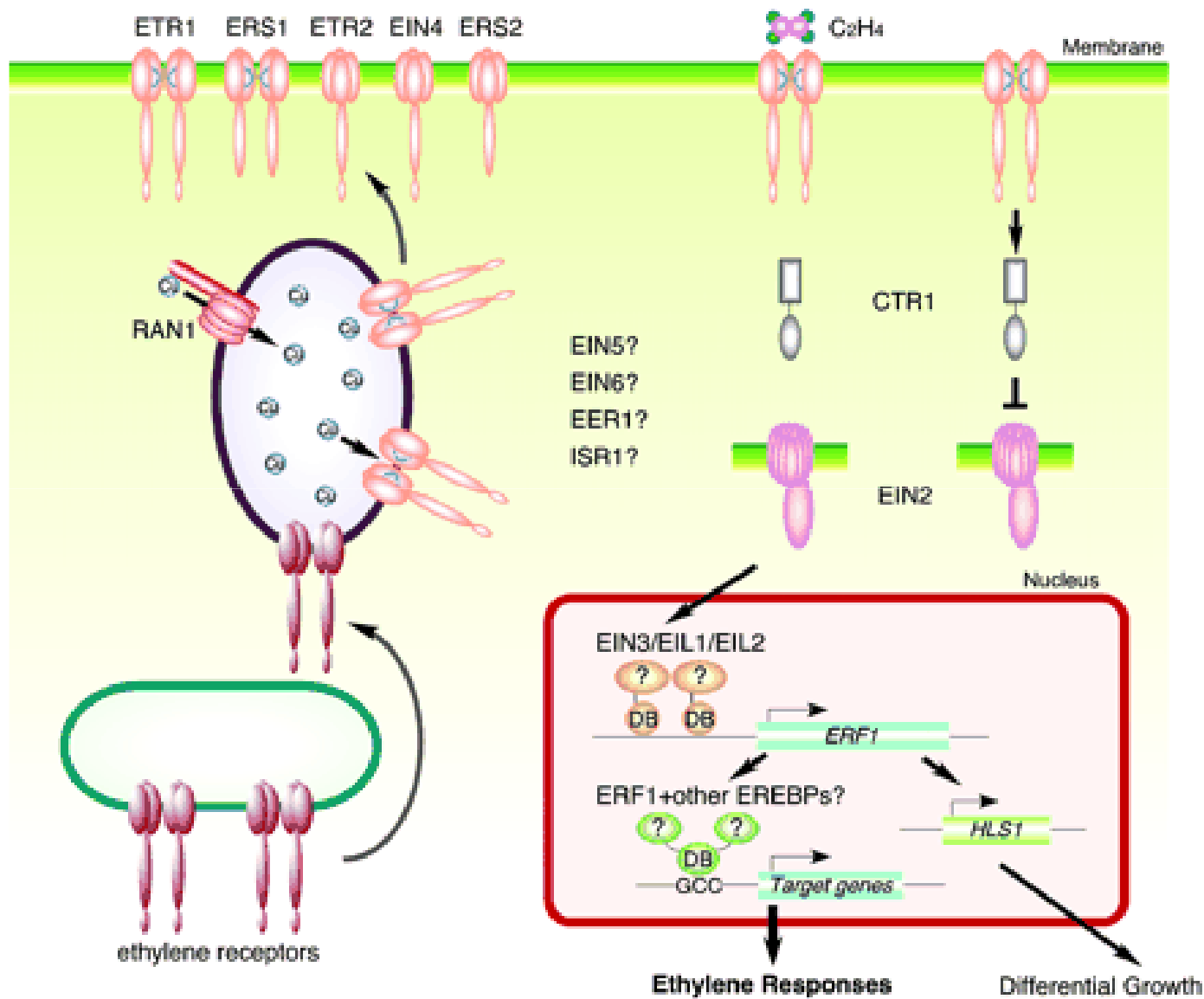
Overexpression of EBF1 and EBF2 results in ethylene insensitivity and reduced accumulation of EIN3 protein



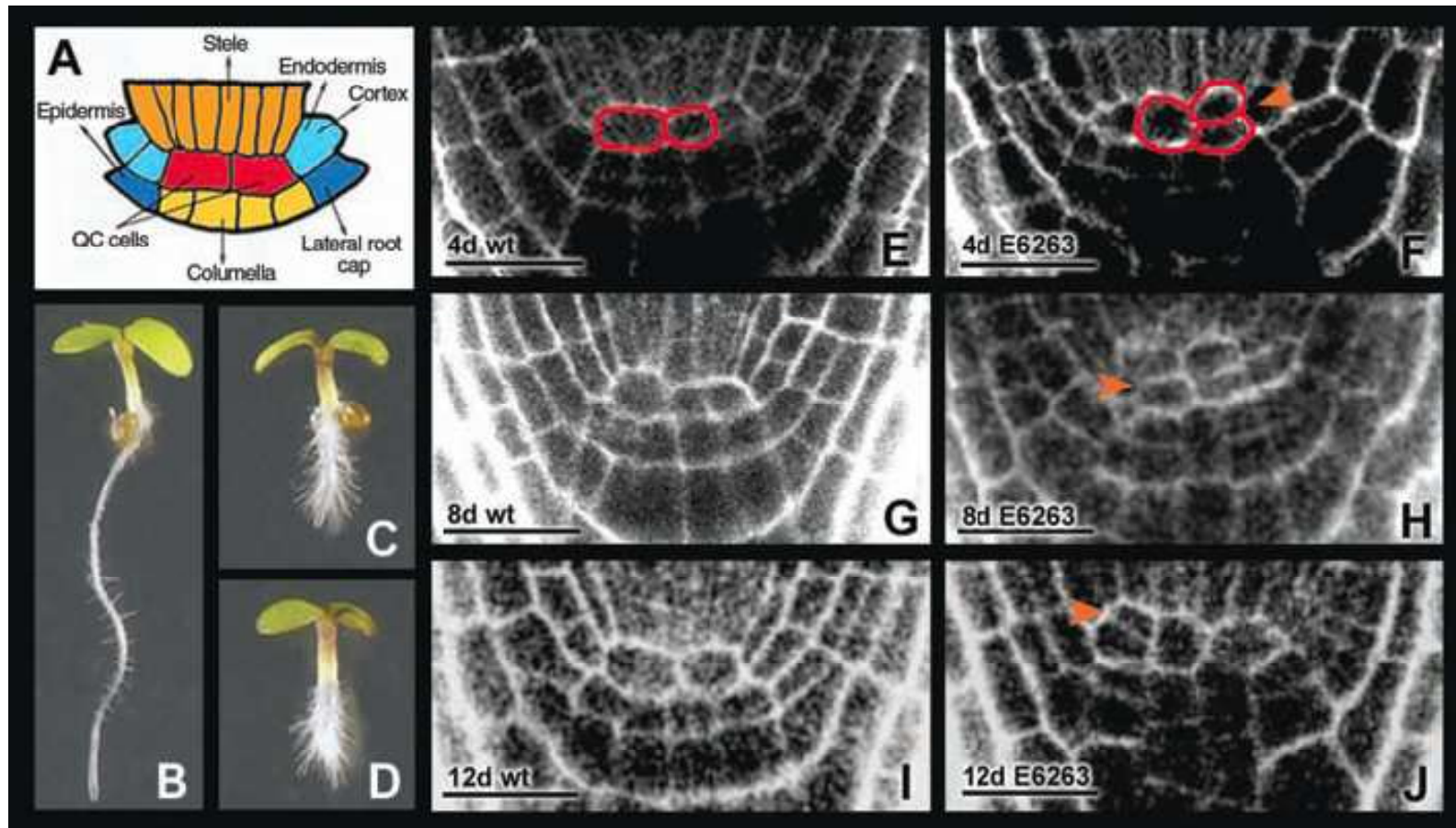
MODELS



Ethylene signal transduction pathway

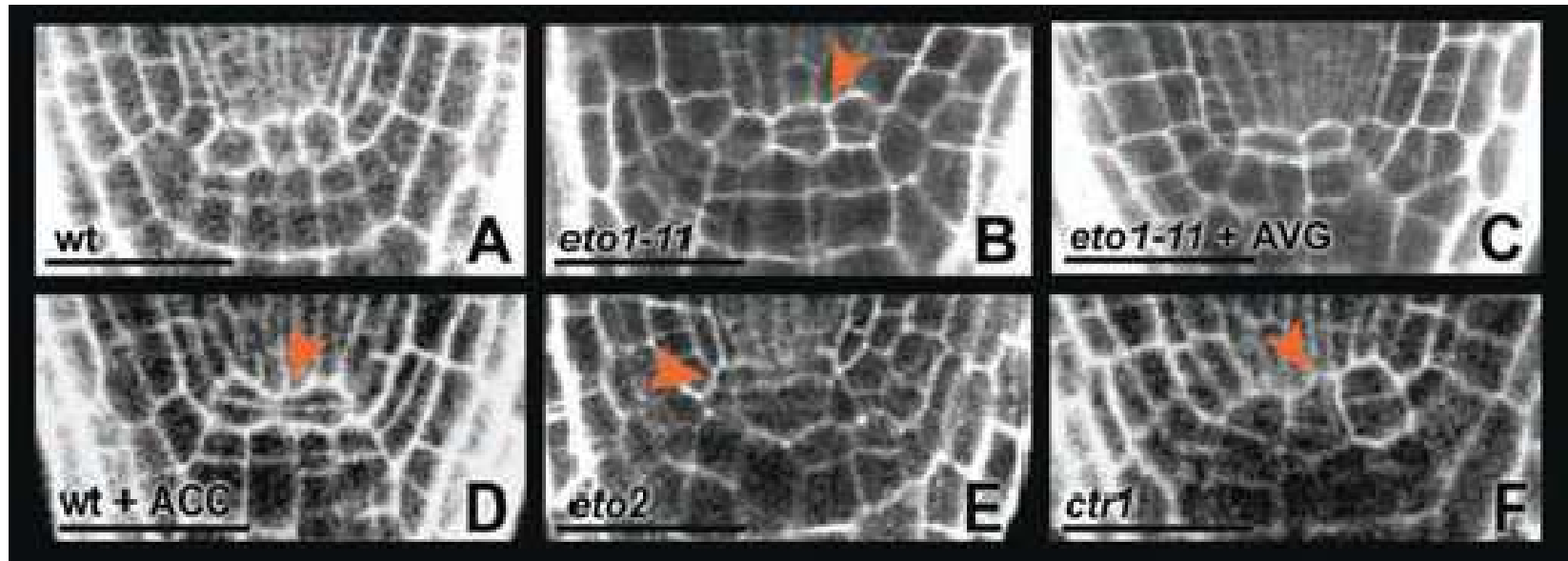


Ethylene Modulates Stem Cell Division in roots



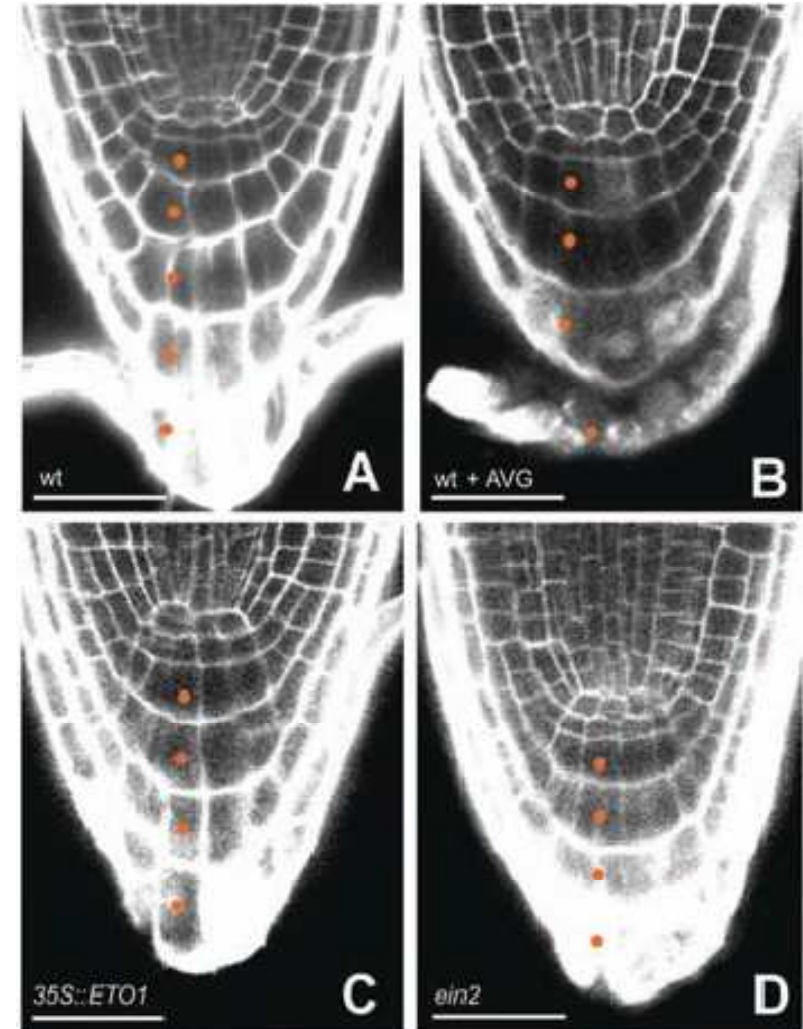
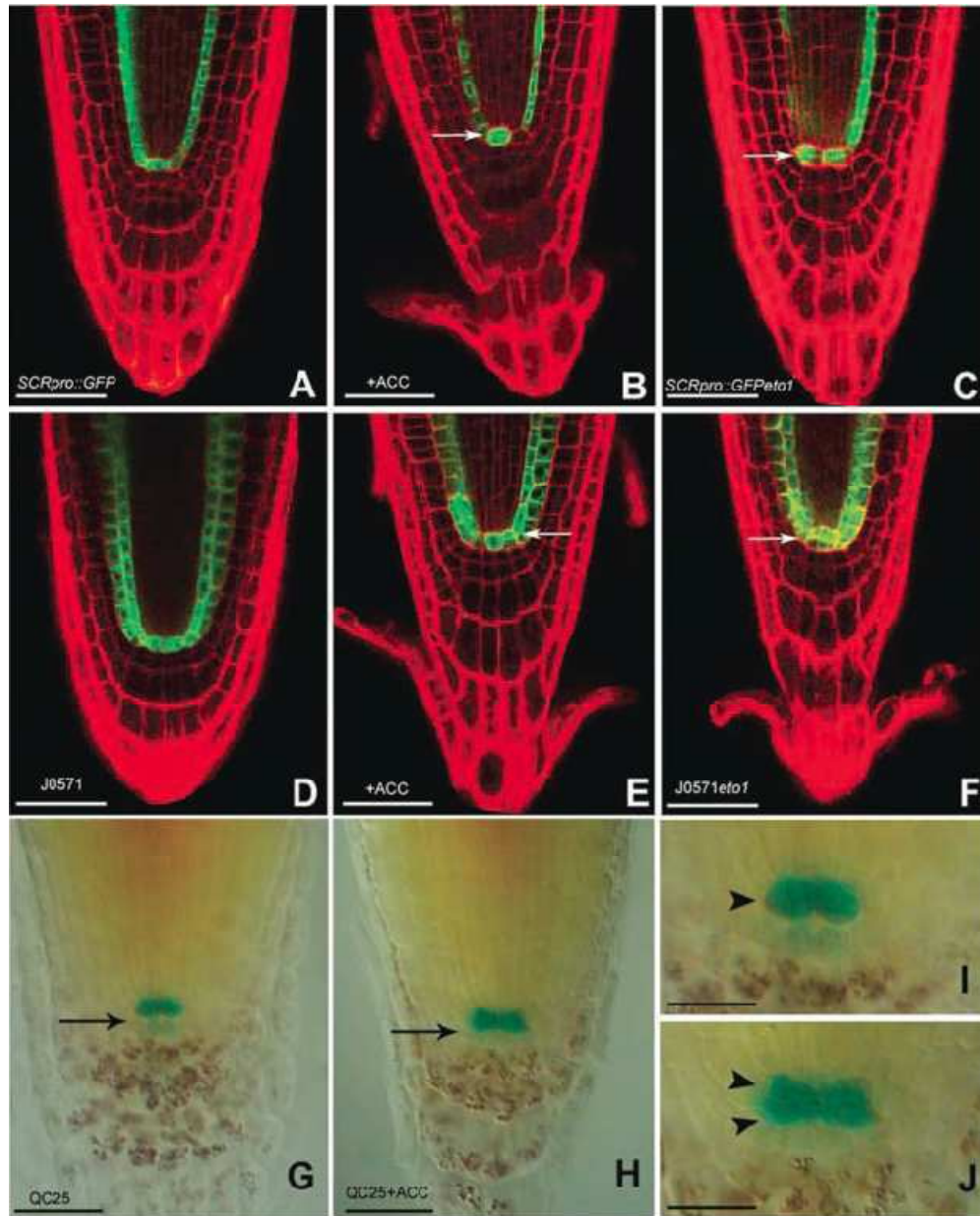
Olga Ortega-Martínez et al., 2007

Ethylene promotes QC cell division.

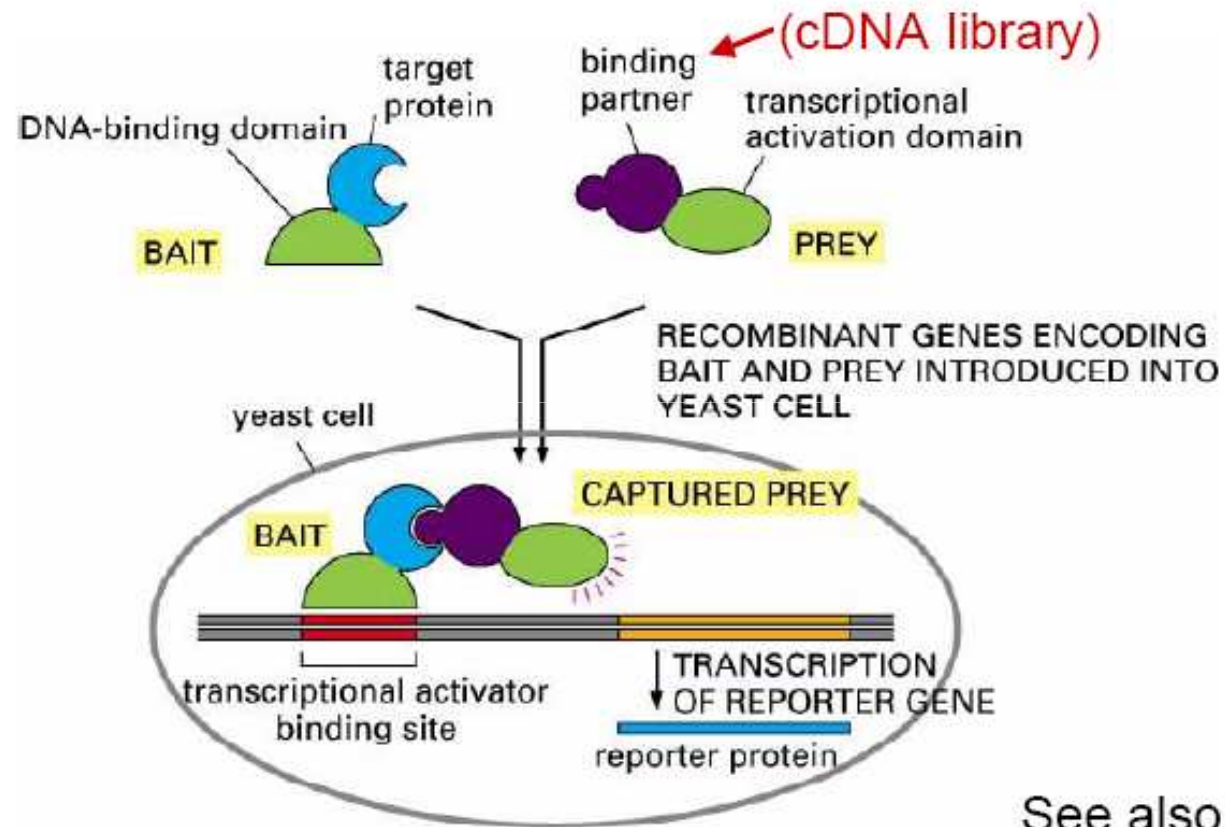


QC cell identity and function are maintained in *eto1* mutants

Ethylene stimulates formation of additional columella cell layers



Yeast two hybrid screen



See also MCB
Fig. 11-39

Figure 8-51. Molecular Biology of the Cell, 4th Edition.

recombinant DNA techniques are used to make fusion between protein X and glutathione S-transferase (GST)

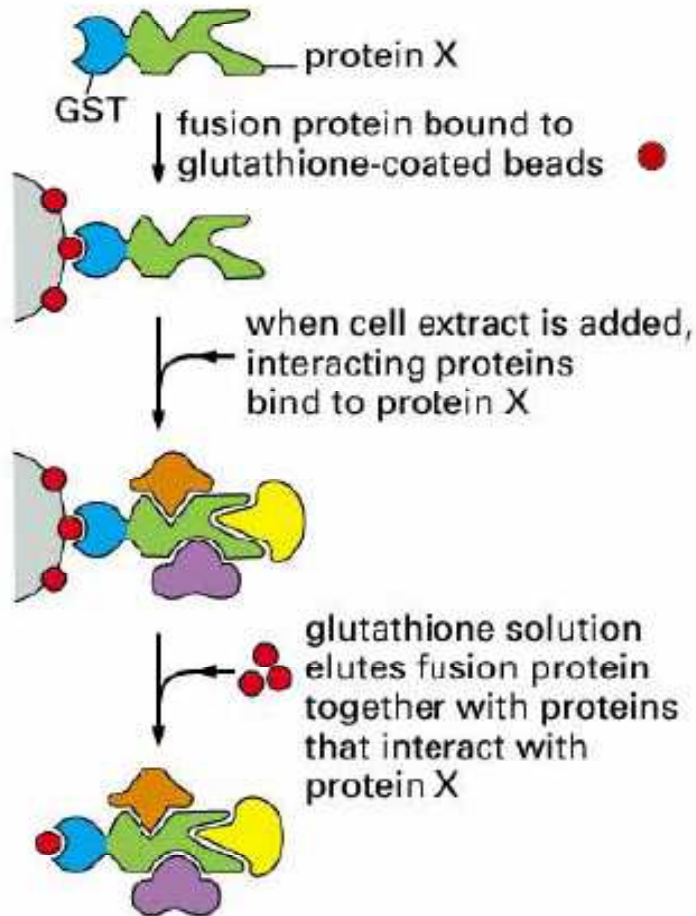


Figure 8-50. Molecular Biology of the Cell, 4th Edition.

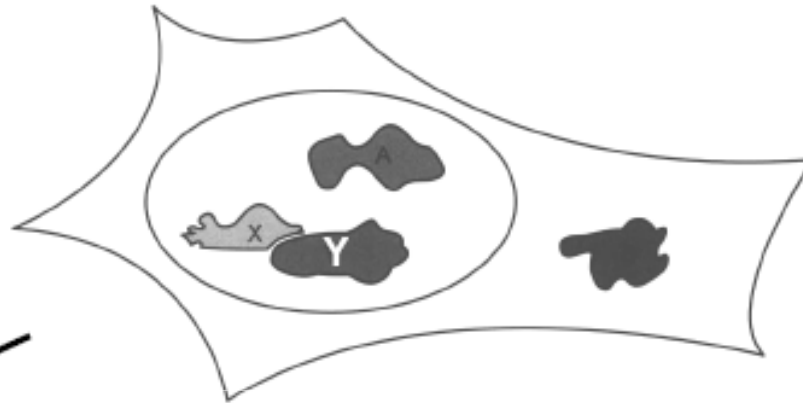
GST “pull downs”

- GST protein is usually expressed in *E. coli* as microgram quantities are used in typical assays
- Detection of bound proteins are usually by western blotting, using antibody to the putative interactor
- Used extensively with GST-domain fusions in structure function studies
- New proteins can be identified if metabolically labeled cells are used

Co-IP's

To be implicated in signal transduction, the interaction should depend on activation of the signaling pathway (compare co-IP before and after stimulation/ligand binding)

For regular IP, lyse cells using harsh conditions



Lyse cells and IP protein X using co-IP conditions (1% triton X-100, 1% Brij96, 2% digitonin, or 2% CHAPS, etc.)

