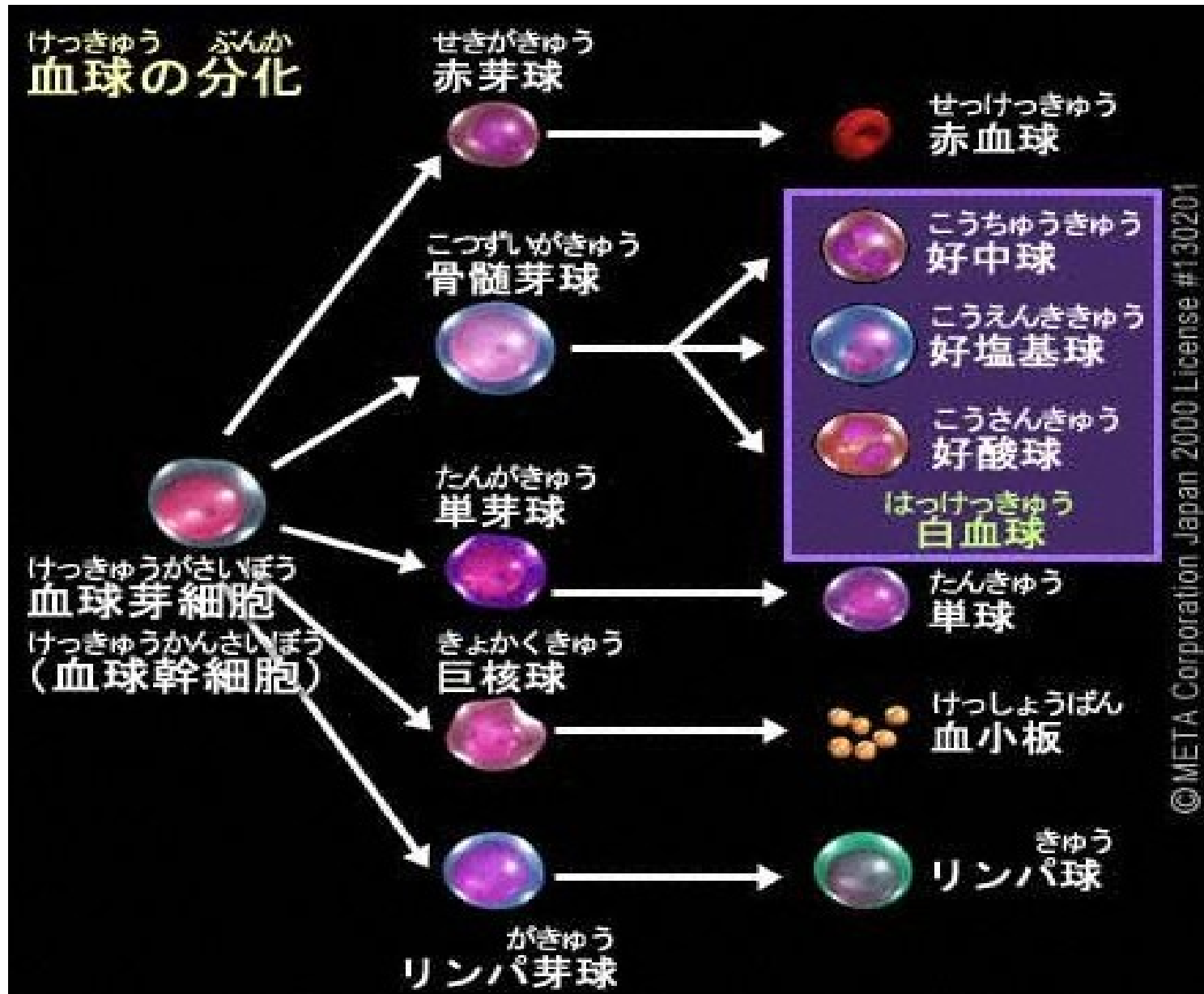
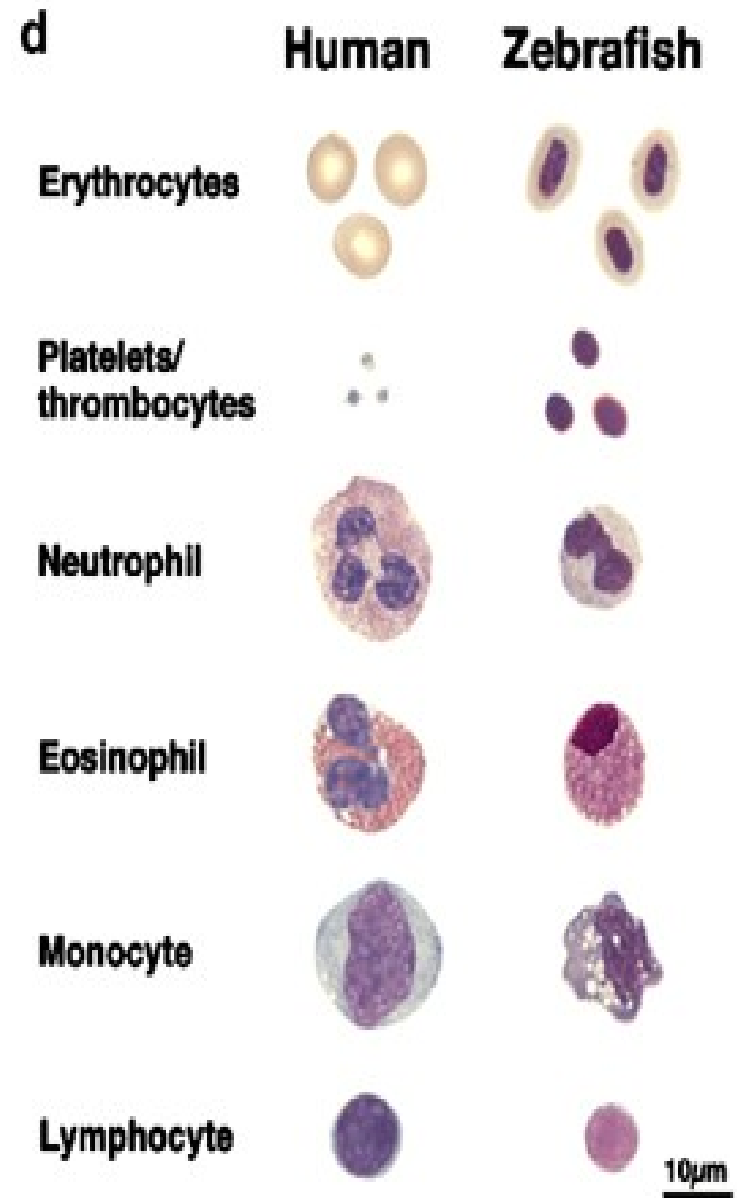
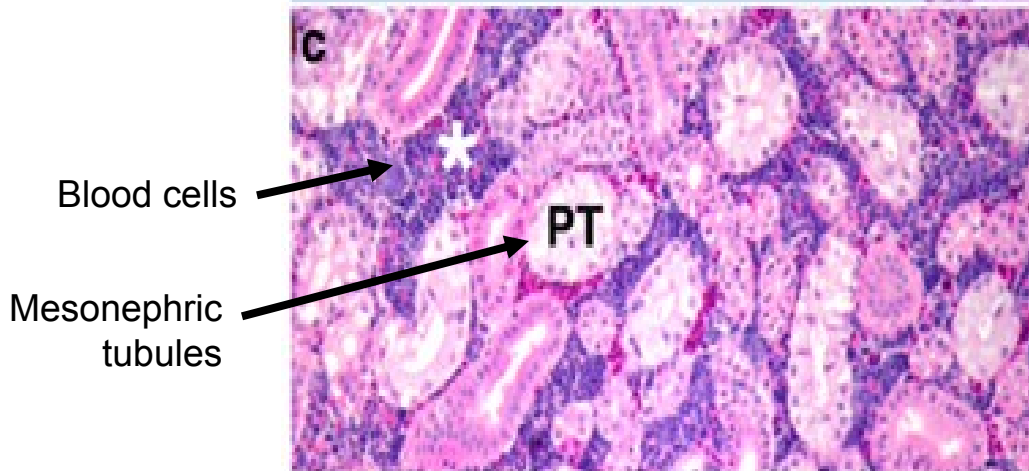
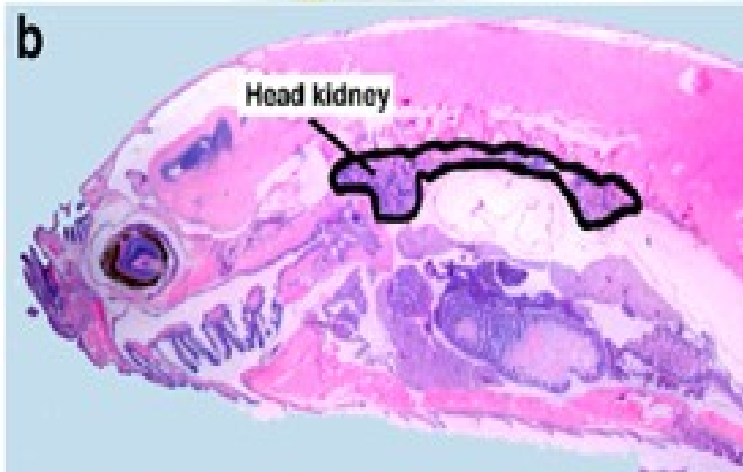
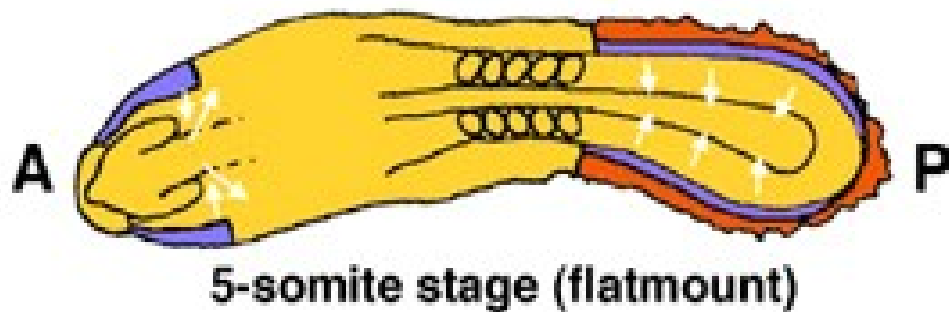
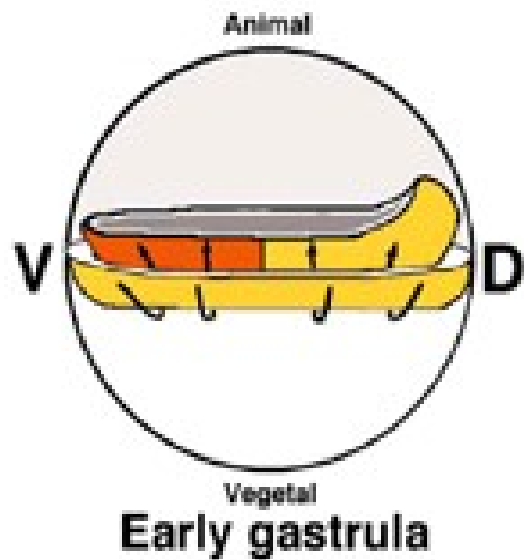


# 5. DEVELOPMENT OF BLOOD

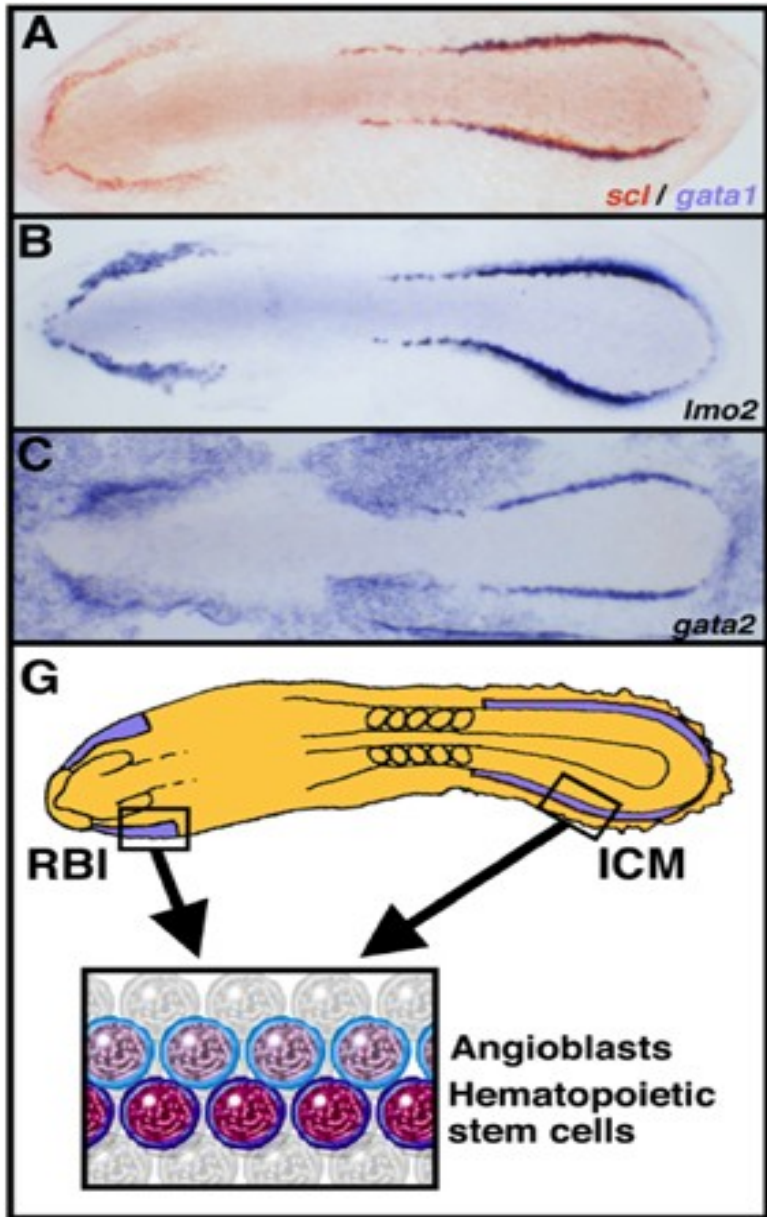






 primitive hematopoiesis  
 ventral mesoderm

**Fish gastrulation movie**

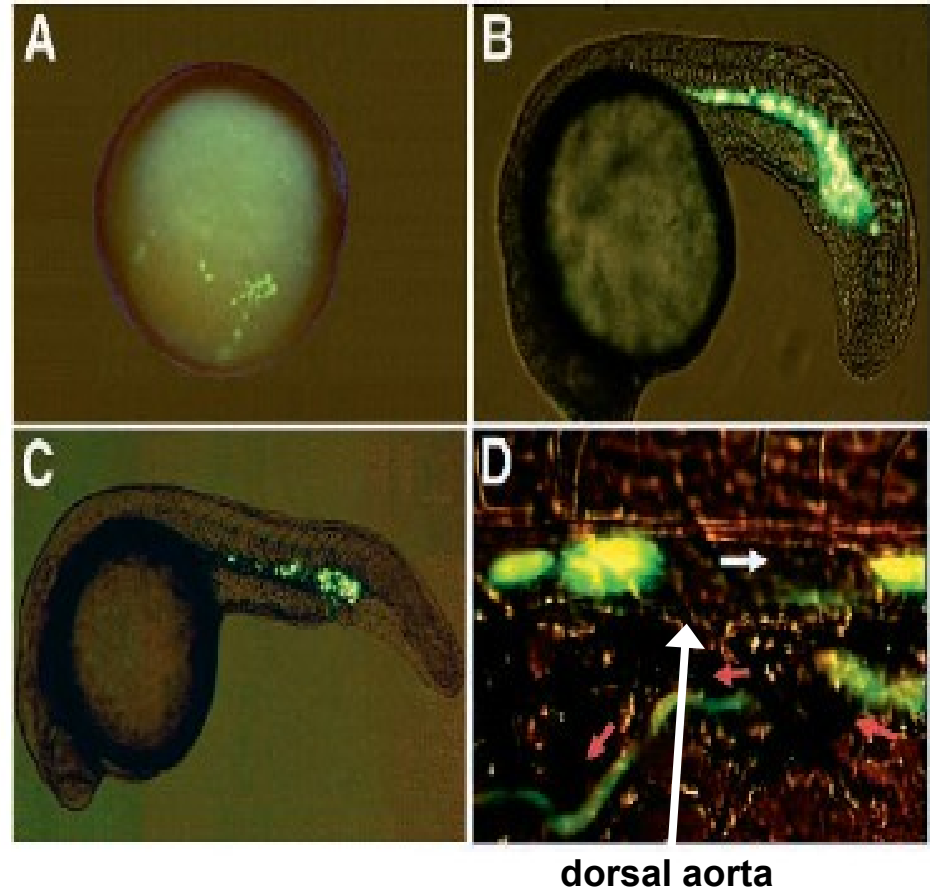


*scl, gata1/2, lmo2* - transcriptional factors that specify early hematopoietic tissue (*gata2* expressed also in ectoderm)

**RBI** – rostral blood islands

**ICM** – inner cell mass

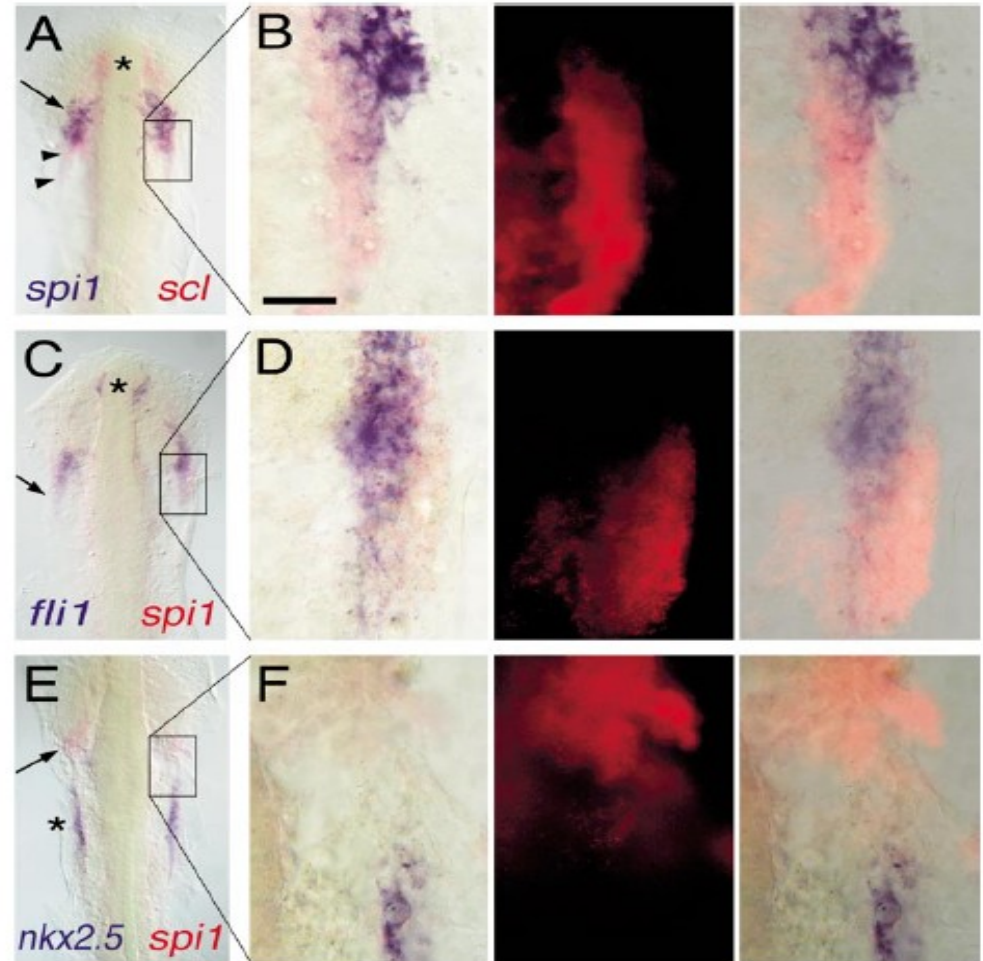
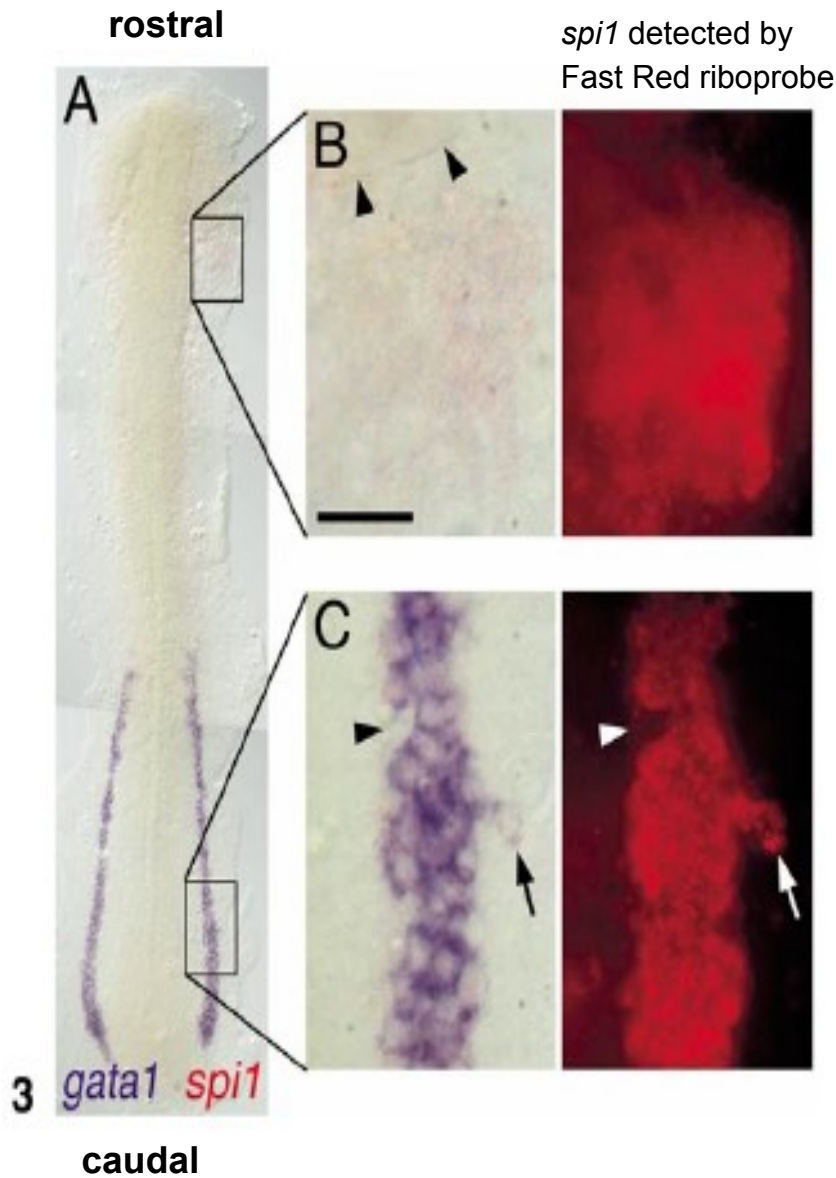
*gata1* promoter-driven GFP = early erythroid lineage



5-somite stage embryo – hematopoiesis starts officially

# MYELOPOIESIS STARTS IN RBI @ 10-SOMITE STAGE

*spi1* – marker of myeloid lineage

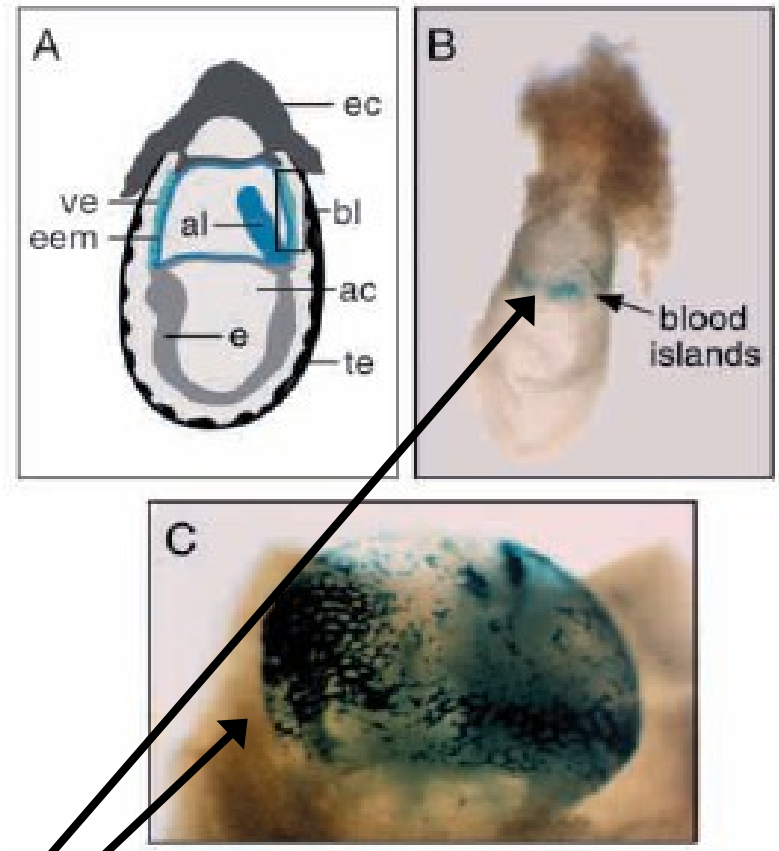
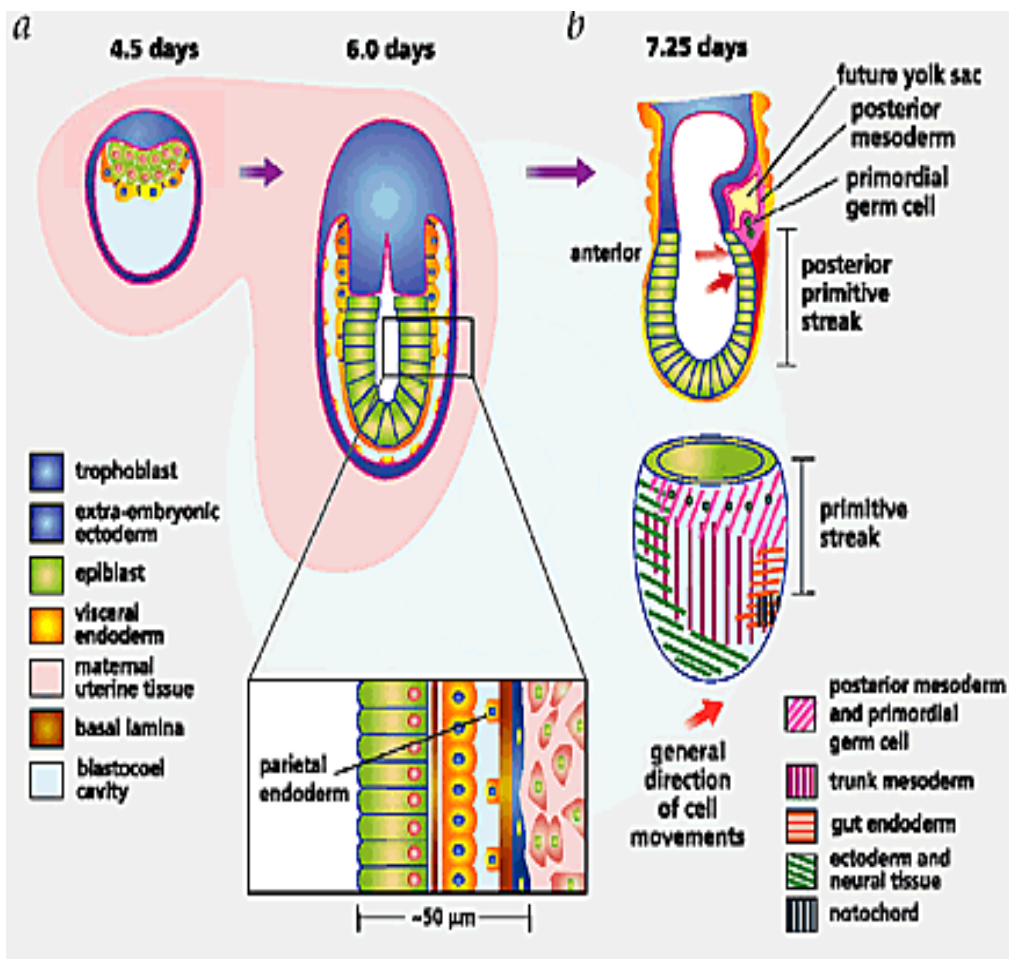


*scl* – early hematovascular cell fate

*fli1* – early vascular fate

*nkx2.5* – heart fate





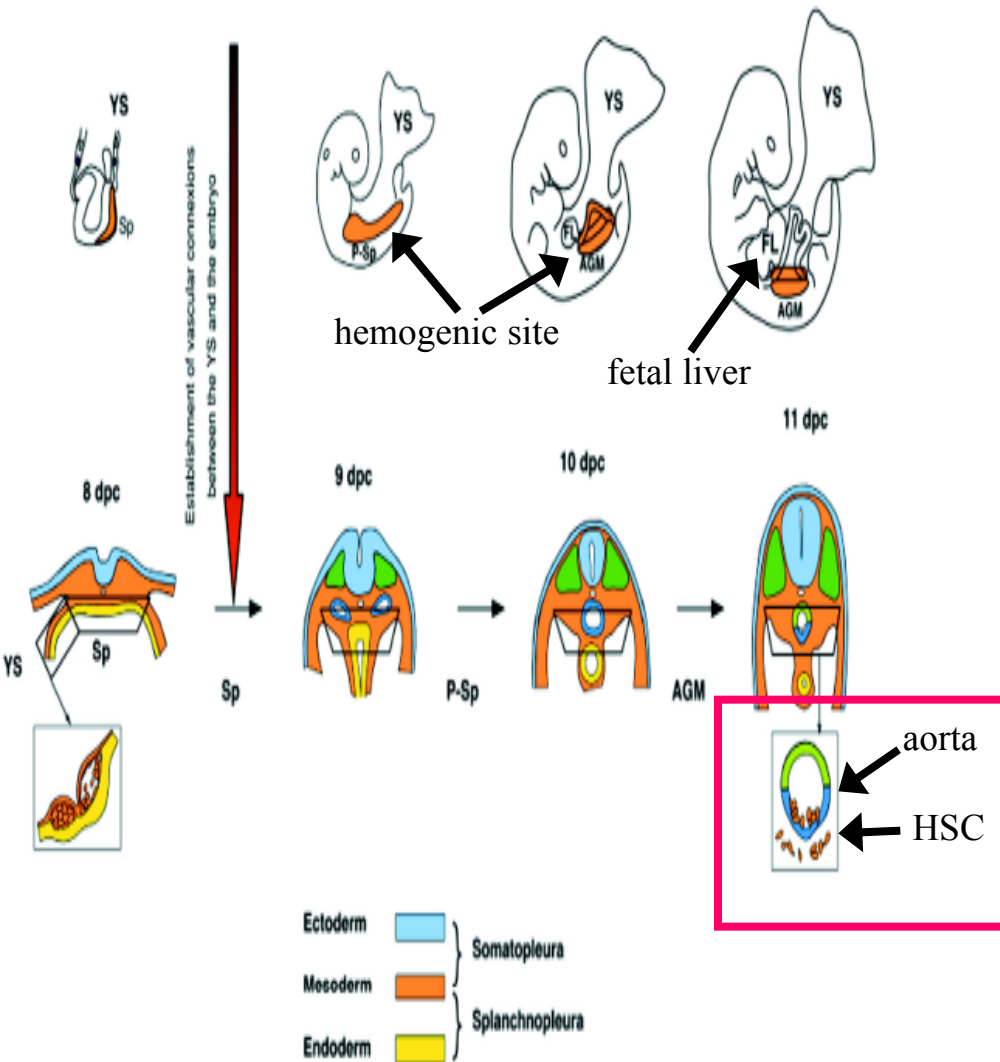
8.5 dpc – yolk sac encloses entire embryo  
blood islands merge to form vascular channels

LacZ driven by  $\beta$ -globin promoter  
(X-gal staining – primitive erythroblasts)

bl - blood islands  
ec - ectoplacental cone  
ac - amniotic cavity  
te - trophectoderm  
al - allantois  
eem - extraembryonic mesoderm (blue)  
ve - visceral endoderm  
e - embryonic ectoderm

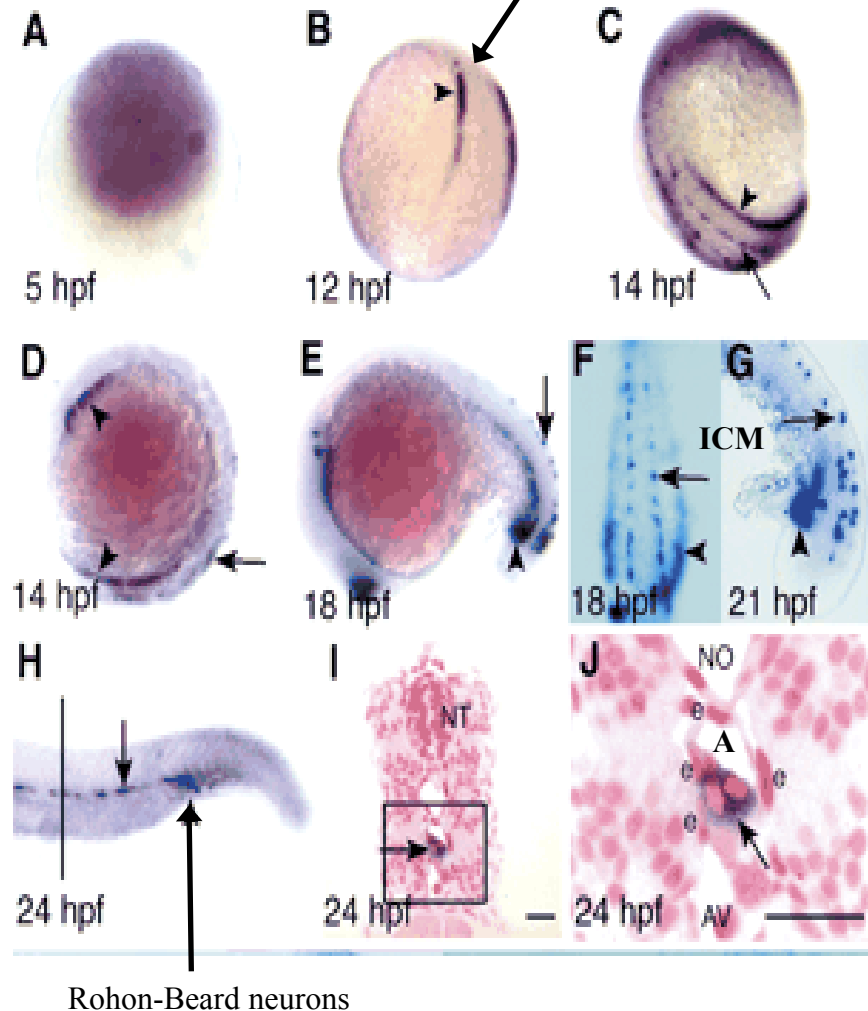
# AORTA-GONAD MESONEPHROS (AGM)

Development of intra-embryonic hemogenic site



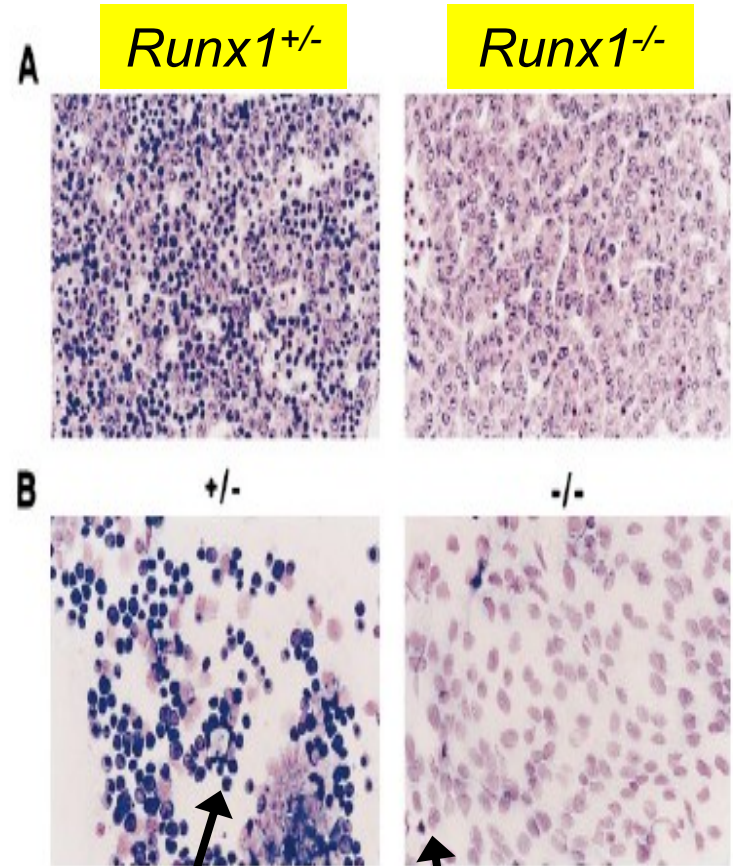
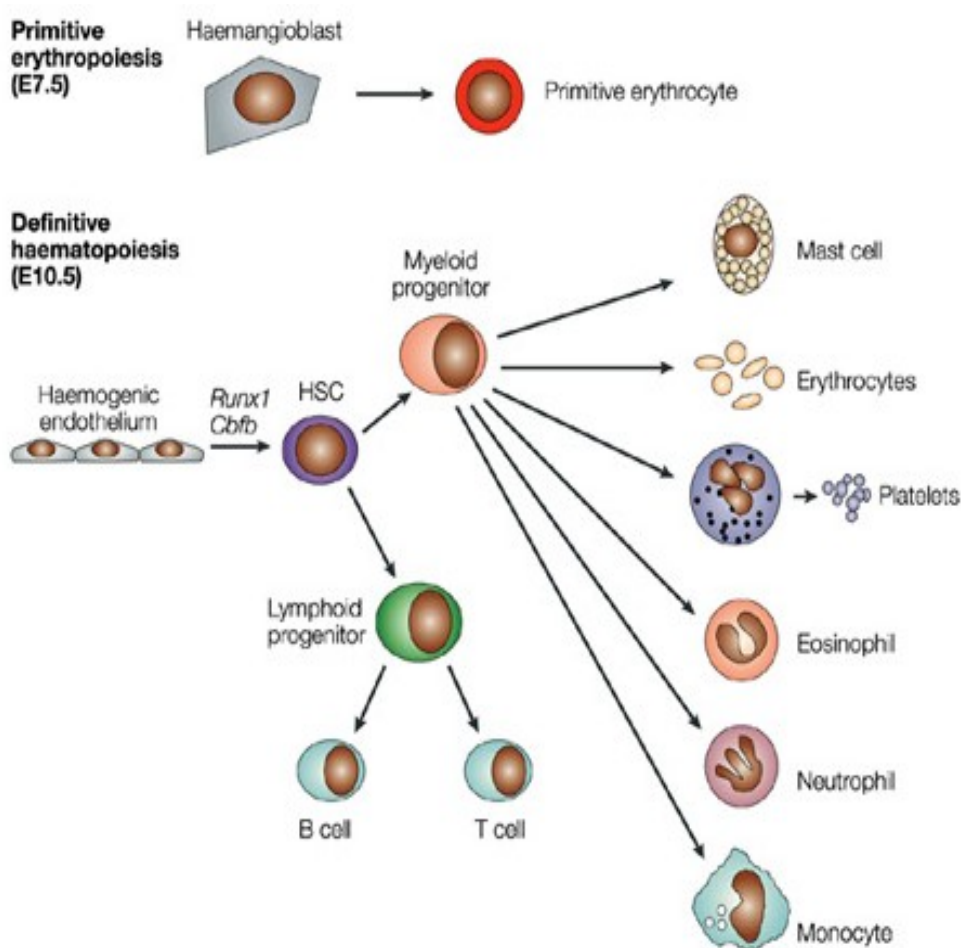
## RUNX1 riboprobe

lateral plate mesoderm



A- aorta  
 NO- notochord  
 AV- axial vein  
 e - endothelial cell

# RUNX1 (CBFA2, AML1) is necessary for 'definitive' hematopoiesis to start at AGM

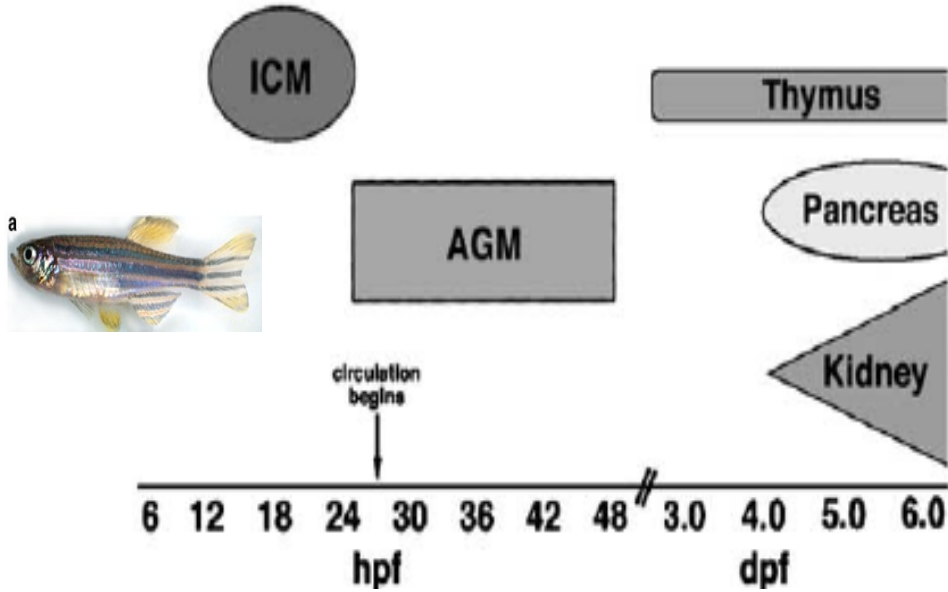


E11.5: **dark-** hematopoietic cells

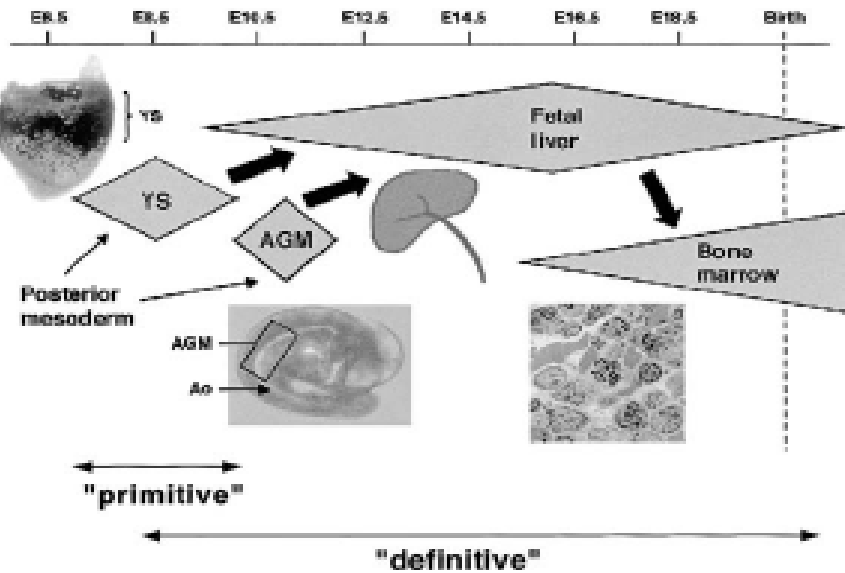
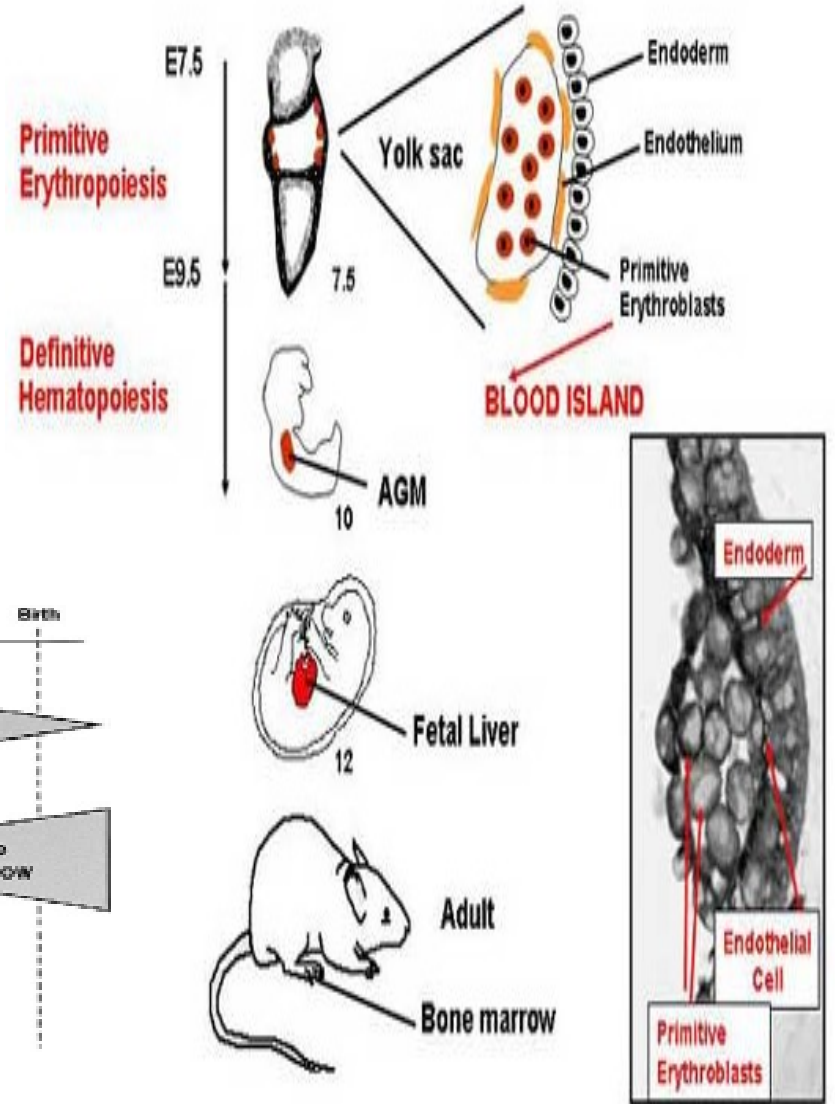
only a few primitive erythrocytes left – likely a carry-over from the 'primitive' hematopoiesis

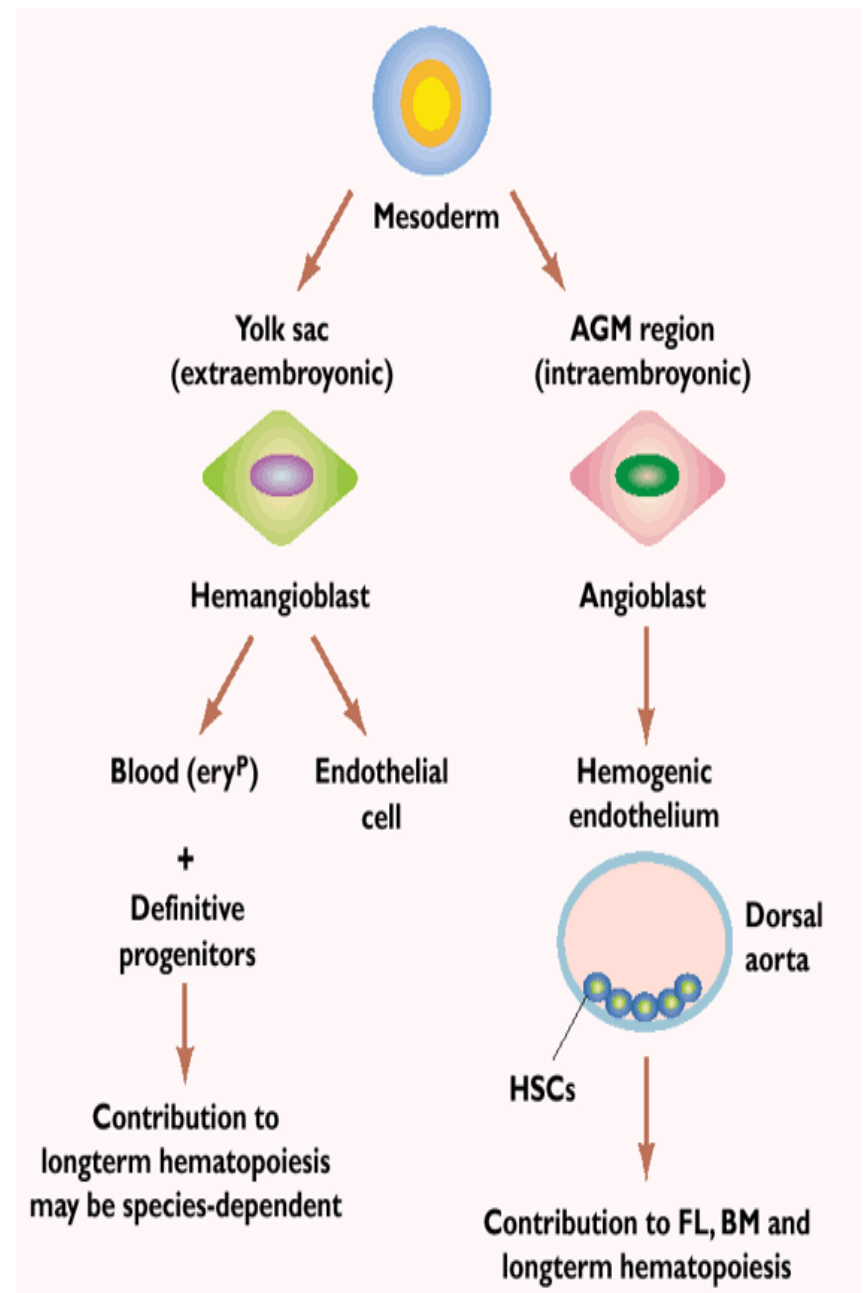
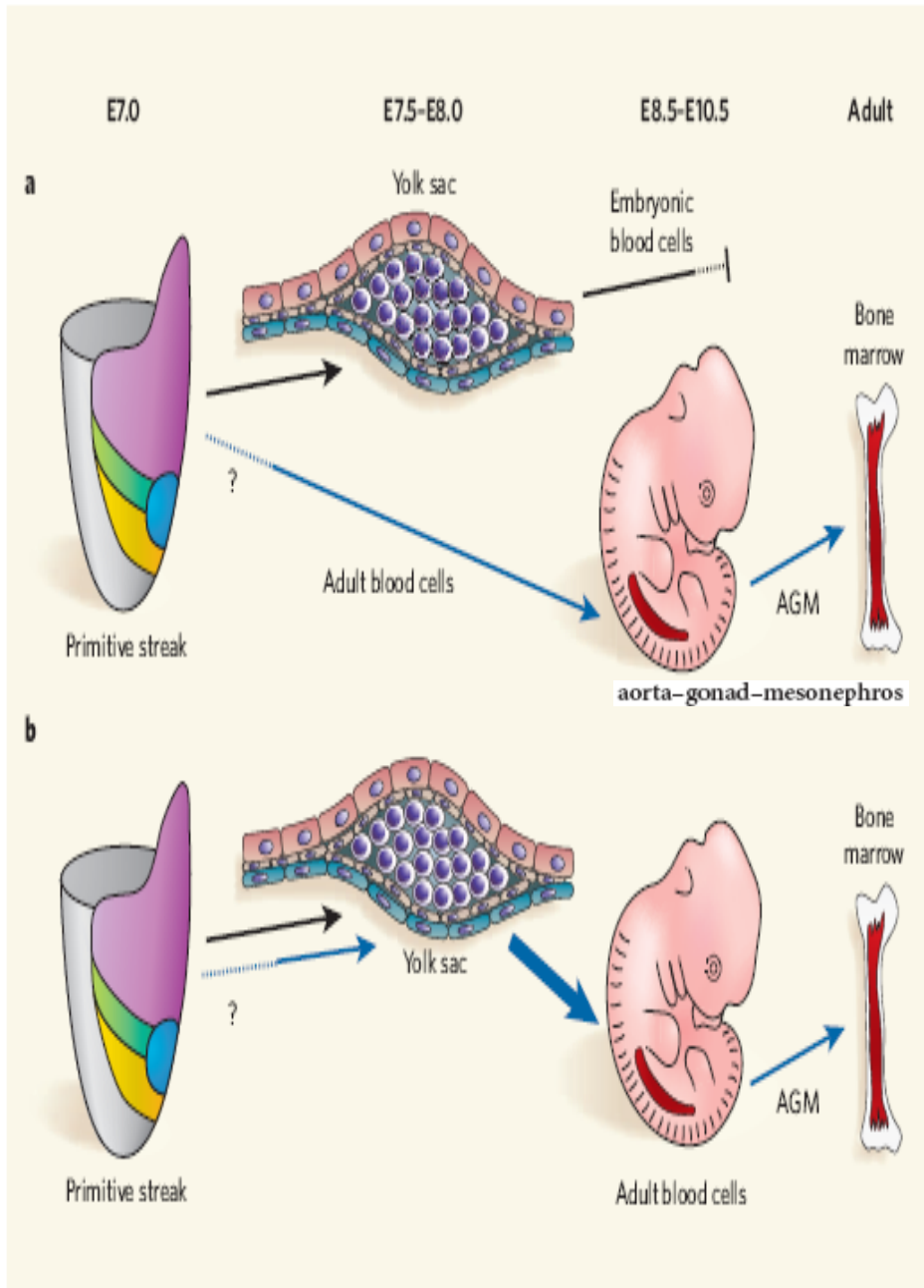


# EXTRAEMBRYONAL vs EMBRYONAL

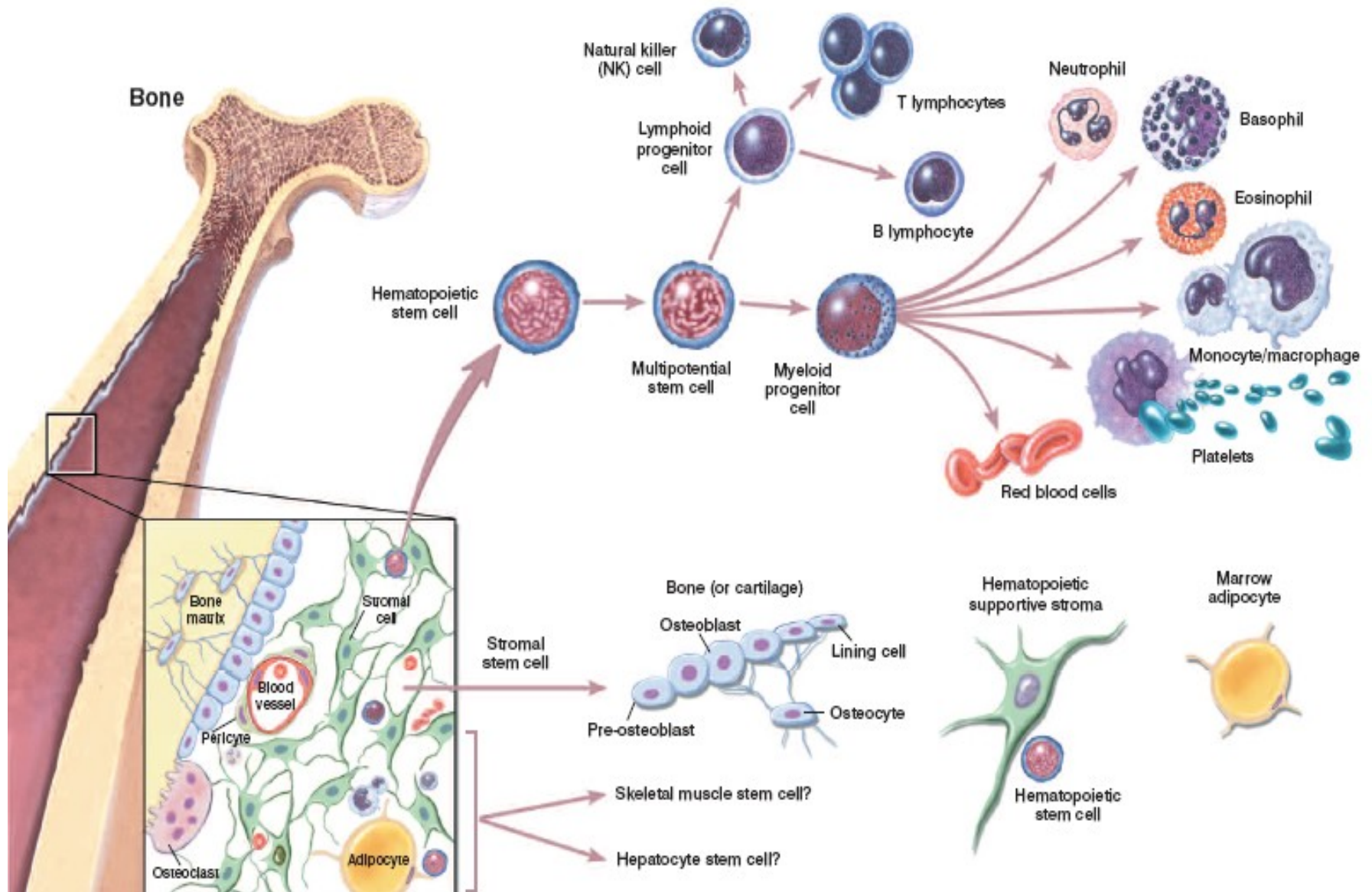


# PRIMITIVE vs. DEFINITIVE



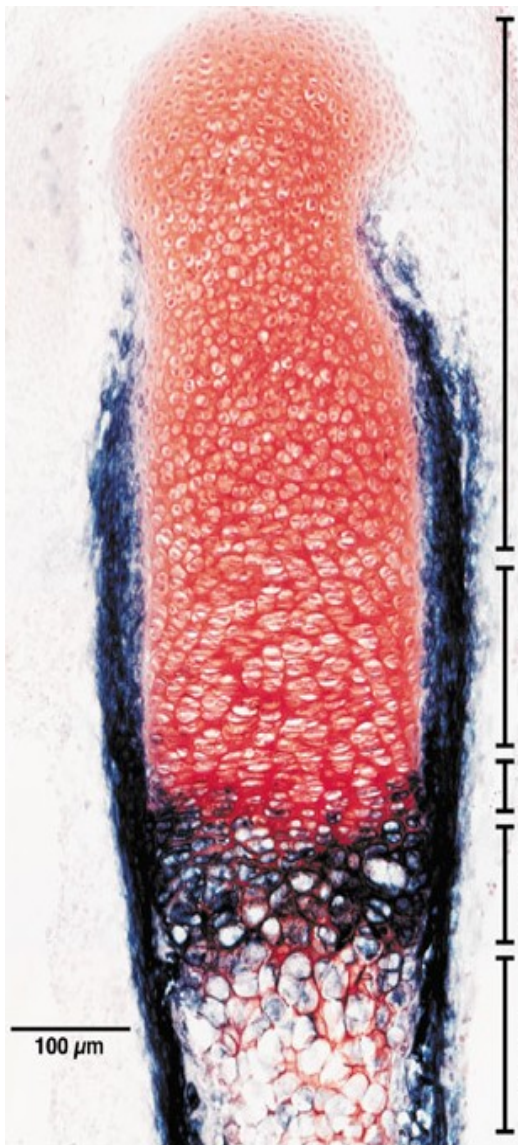


# BONE MARROW NICHE IS CRITICAL FOR HEMATOPOIESIS





# Collagen type X deletion



Resting chondrocytes  
Markers: type II collagen  
Moderate mitotic index

thymus

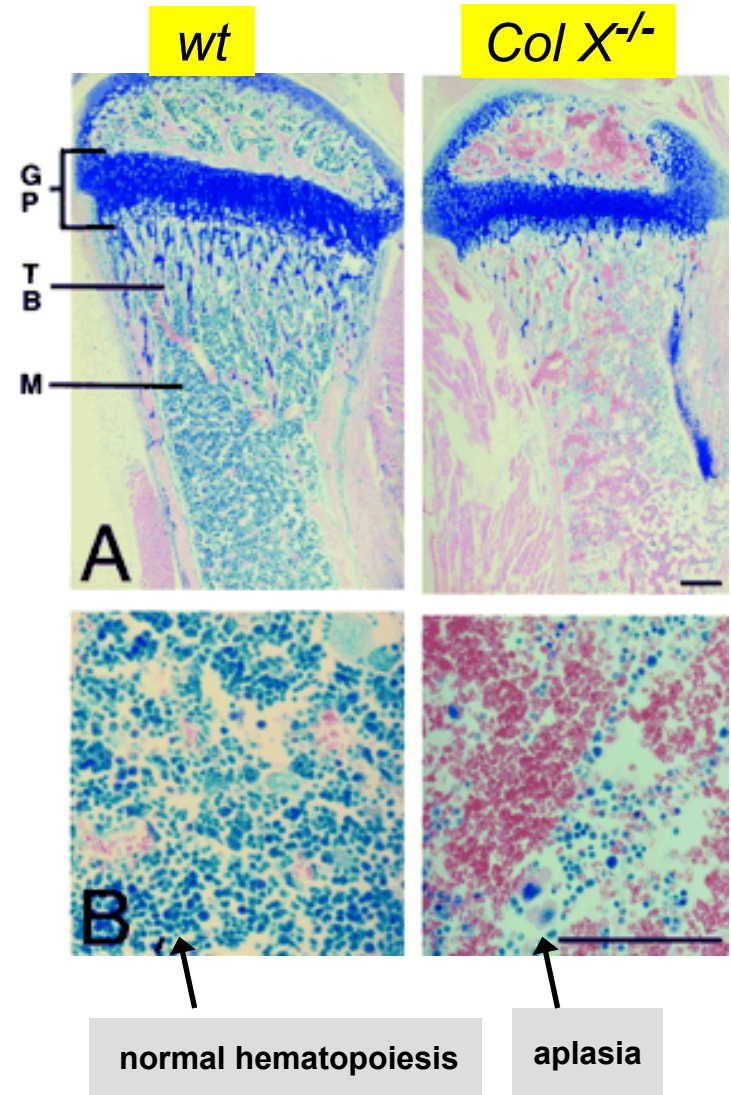
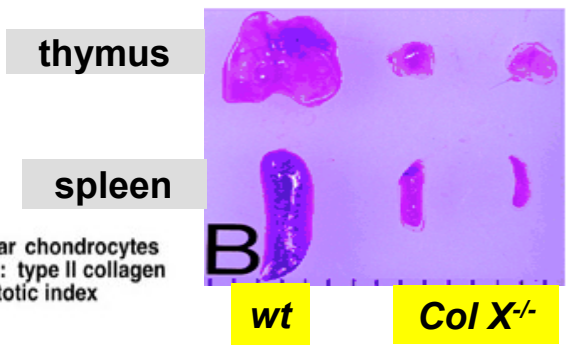
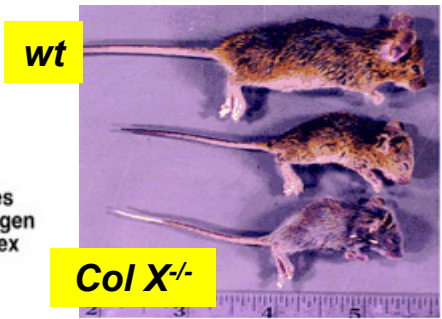
spleen

Columnar chondrocytes  
Markers: type II collagen  
High mitotic index

Prehypertrophic chondrocytes  
Markers: type II collagen, Ihh  
Post-mitotic

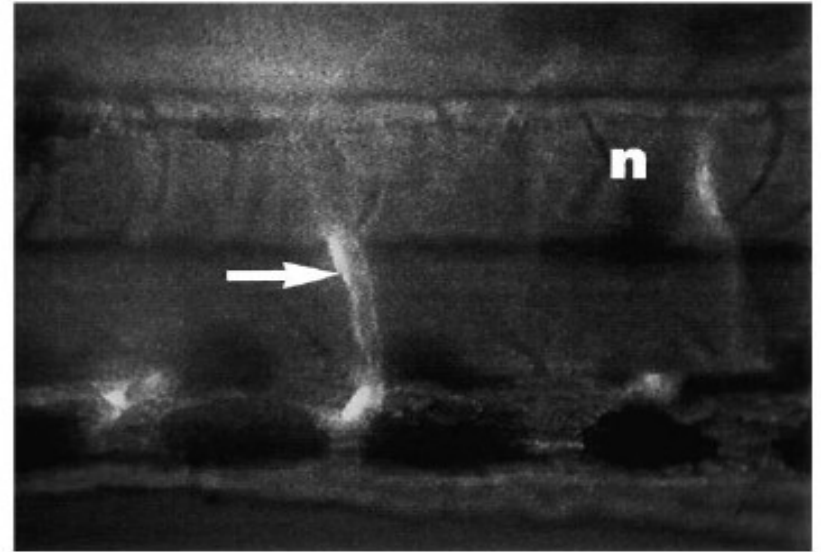
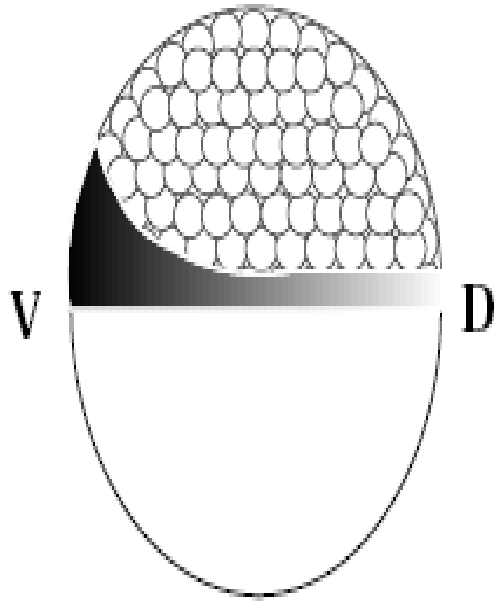
Hypertrophic chondrocytes  
Markers: alkaline phosphatase,  
type X collagen, VEGF  
Post-mitotic

Primary spongiosum (bone)  
Markers: osteocalcin, alkaline  
phosphatase, osteopontin  
High mitotic index



# HEMANGIOBLAST

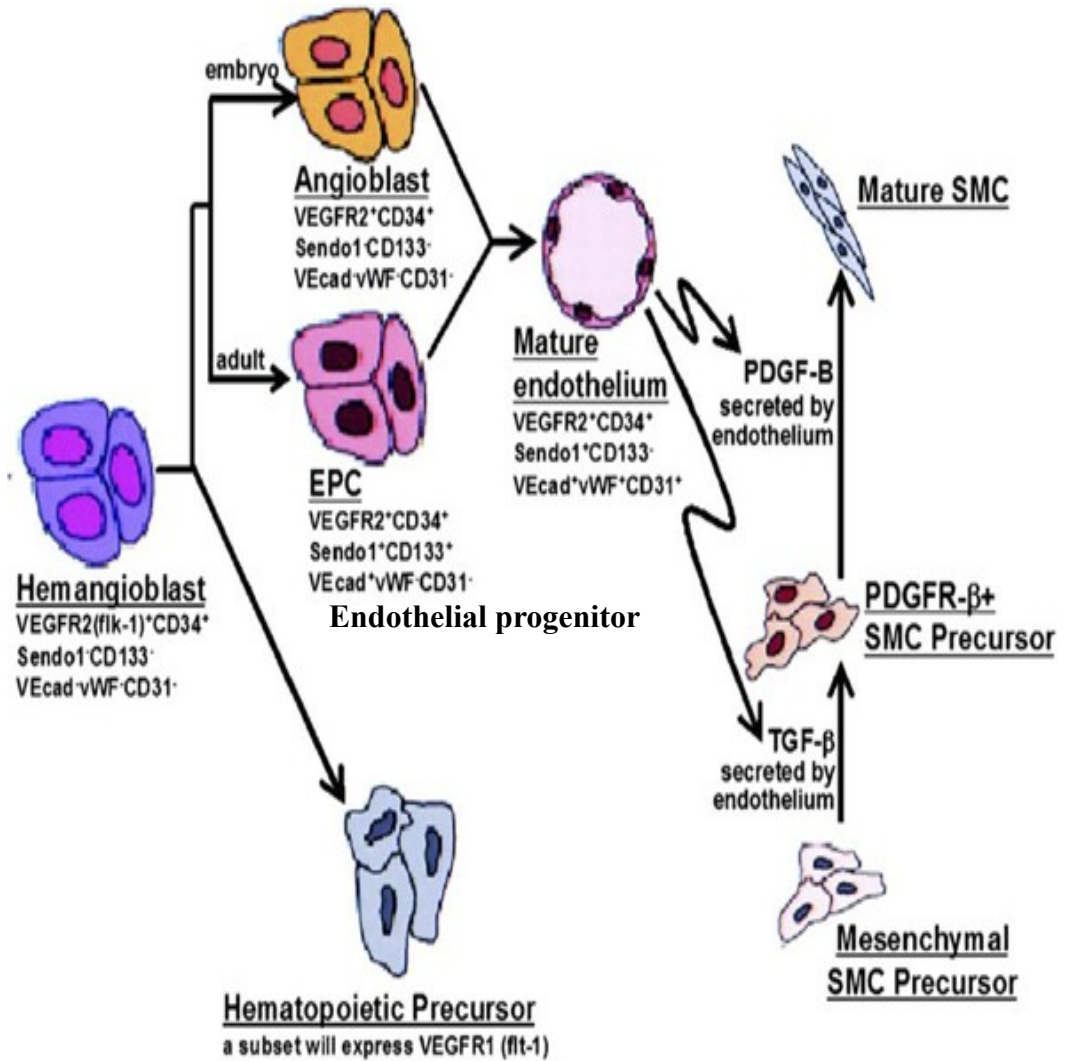
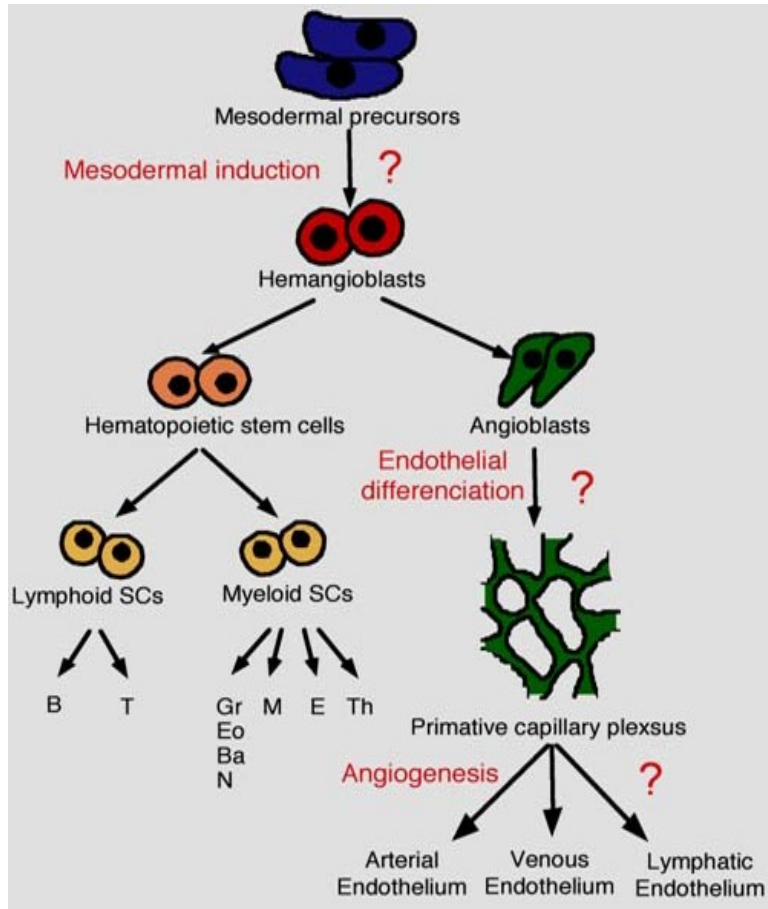
scattered among endothelial and HSC in heart field of the gastrula



**ventral marginal cells of the heart field, n- notochord**

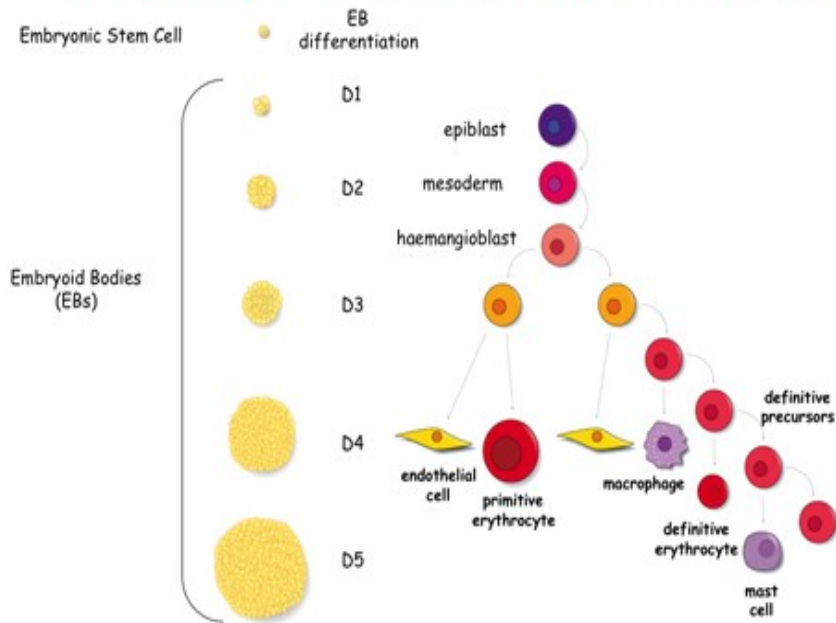
Fig. 5. The heart field. Diagrammatic representation of the heart field in the early blastula. The intensity of grey represents the propensity to form heart.



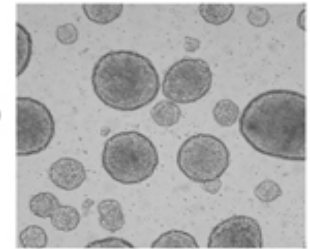
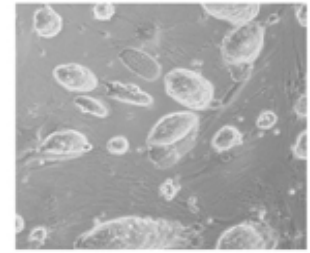
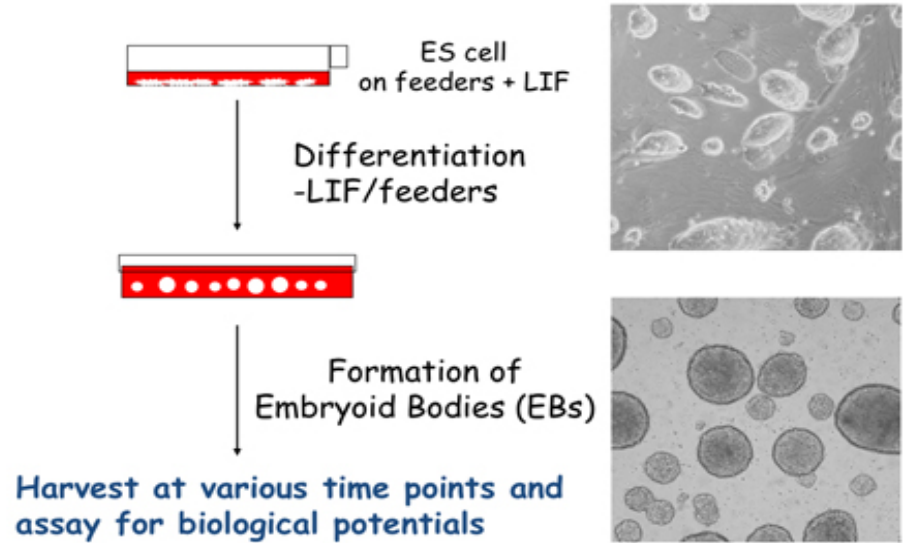


HEMANGIOBLAST or HAEMOGENIC EPITHELIA, or BOTH?

# ES/EB as a Model of Yolk Sac Haematopoiesis

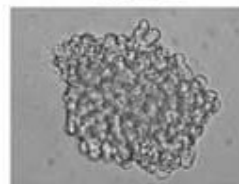
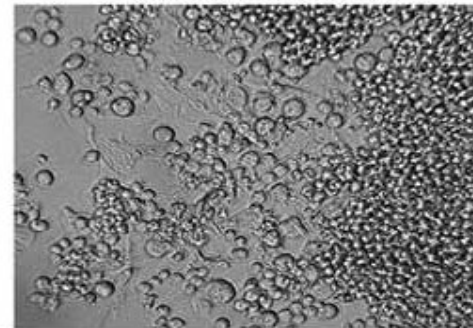
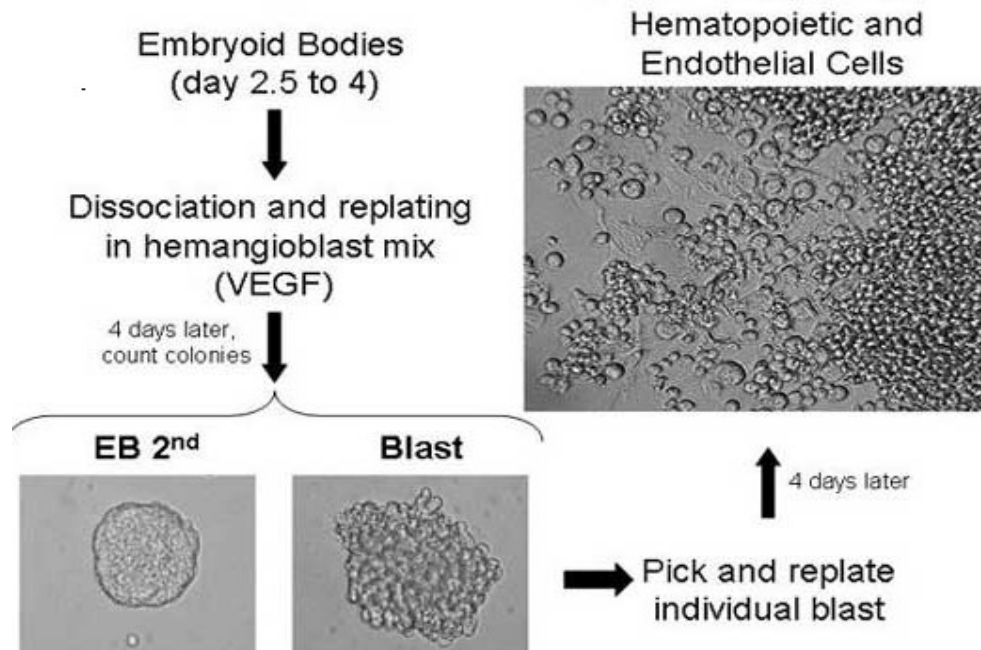


# In vitro differentiation of ES cells



blast colony-forming cell

## Hemangioblast FLK-1<sup>+</sup> (BL-CFC) Assay



# The haemangioblast generates haematopoietic cells through a haemogenic endothelium stage

NATURE | Vol 457 | 12 February 2009

Christophe Lancrin<sup>1</sup>, Patrycja Sroczynska<sup>1</sup>, Catherine Stephenson<sup>1</sup>, Terry Allen<sup>2</sup>, Valerie Kouskoff<sup>3</sup> & Georges Lacaud<sup>1</sup>

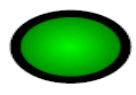
**Embryonic Stem (ES) cell**

**Haemangioblast Fik1<sup>+</sup>**

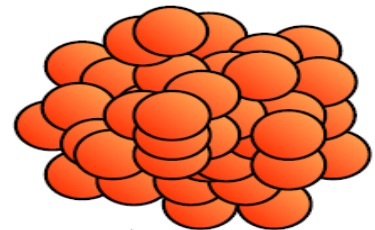
**Blast Colony**



3 to 3.5 Days  
ES differentiation

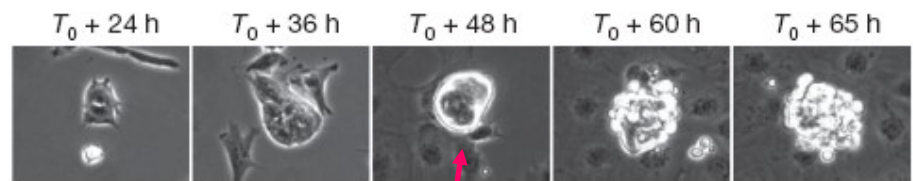
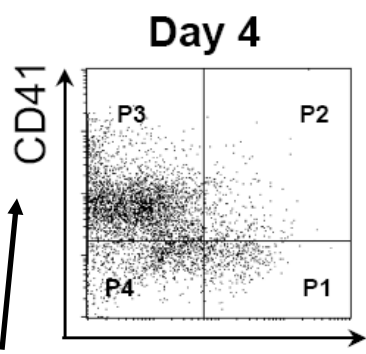


3-4 Days



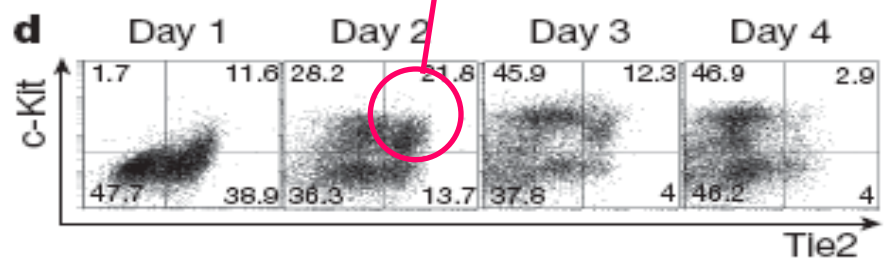
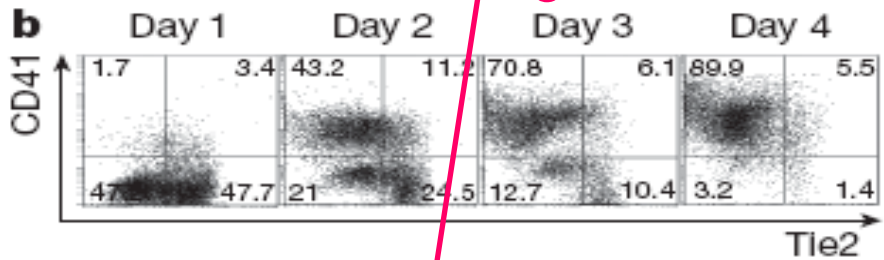
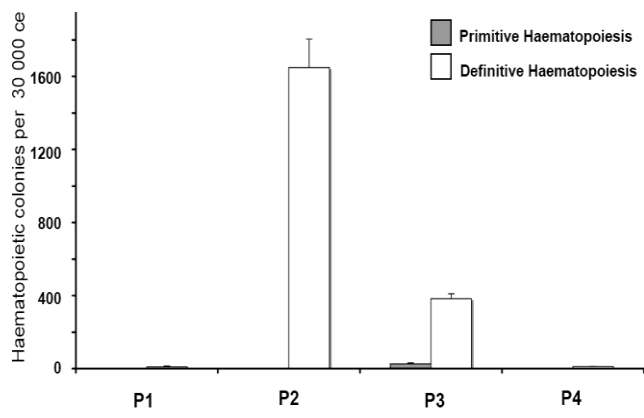
Sort of Fik1<sup>+</sup> cells

Hemangioblast culture



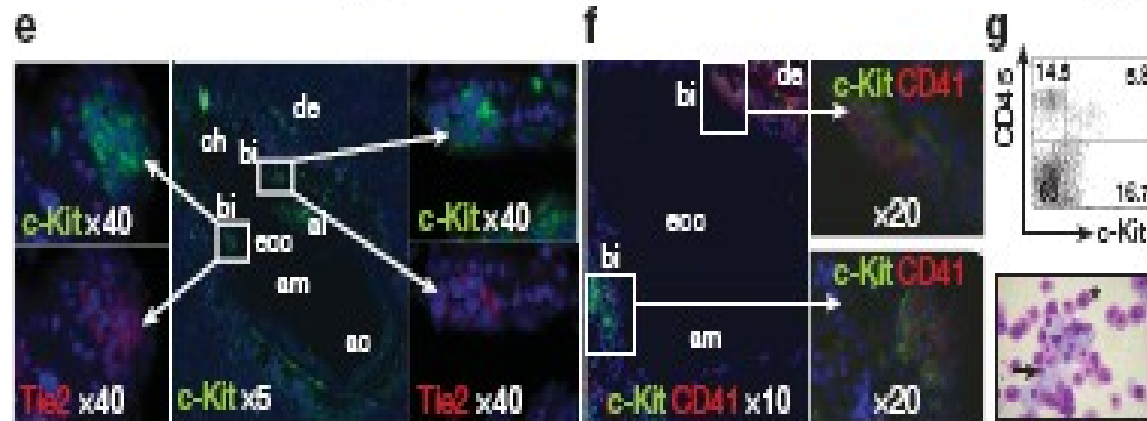
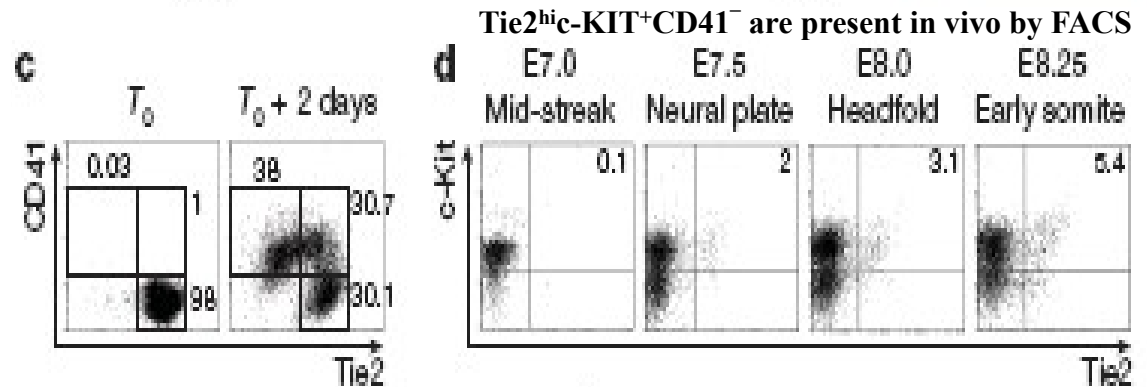
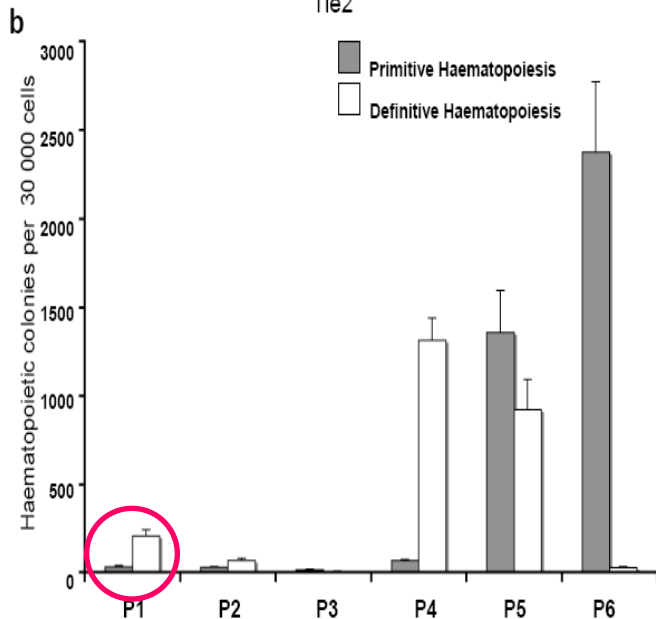
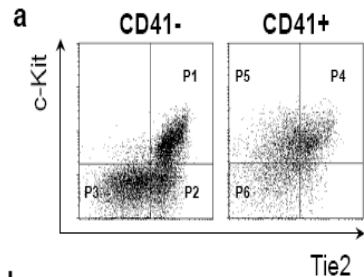
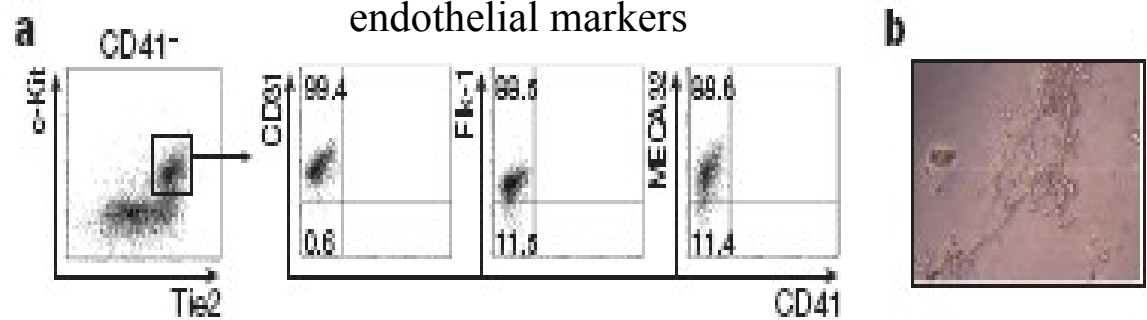
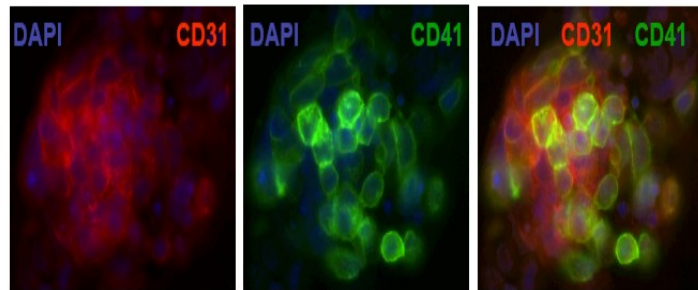
tight adherent structure

Integrin alpha 2B Tie2 Angiopoietin 1 rec.



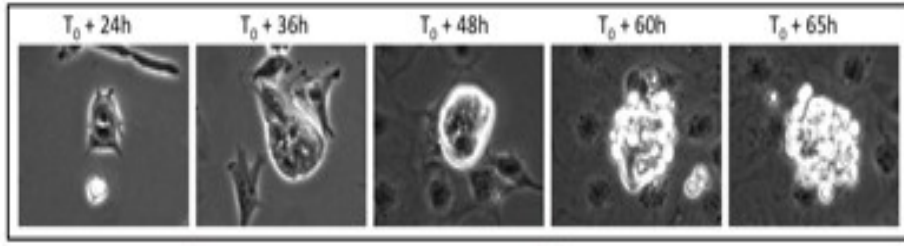
# Tie2<sup>hi</sup>c-KIT<sup>+</sup>CD41<sup>-</sup> can generate hematopoietic precursors

endothelia in matrigel



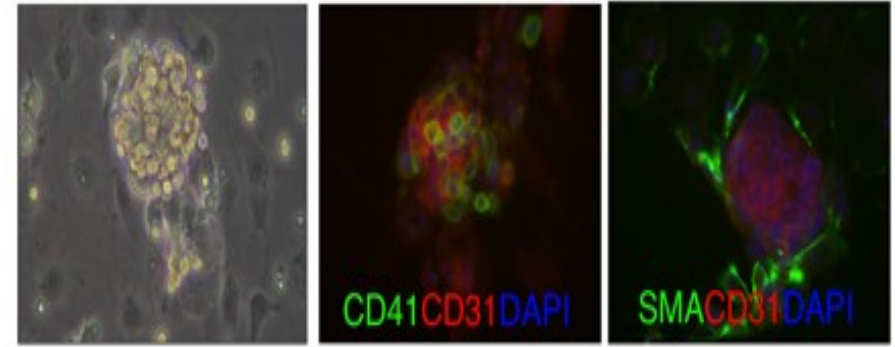
Tie2<sup>hi</sup>c-KIT<sup>+</sup>CD41<sup>-</sup> are present in vivo by IHC





Generation of structure of tightly associated cells

Generation of round cells



Haemangioblast

Haemogenic endothelium

Definitive Haematopoiet progenitors



SCL/TAL-1

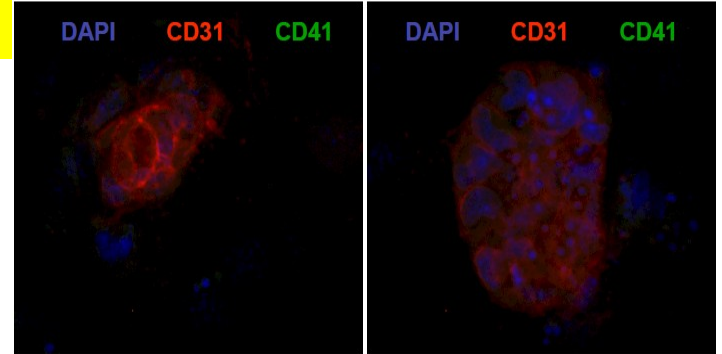
RUNX1/AML-1

Flk1<sup>+</sup>  
Bry-GFP<sup>+</sup>

Tie2<sup>Hi</sup>  
c-Kit<sup>+</sup>

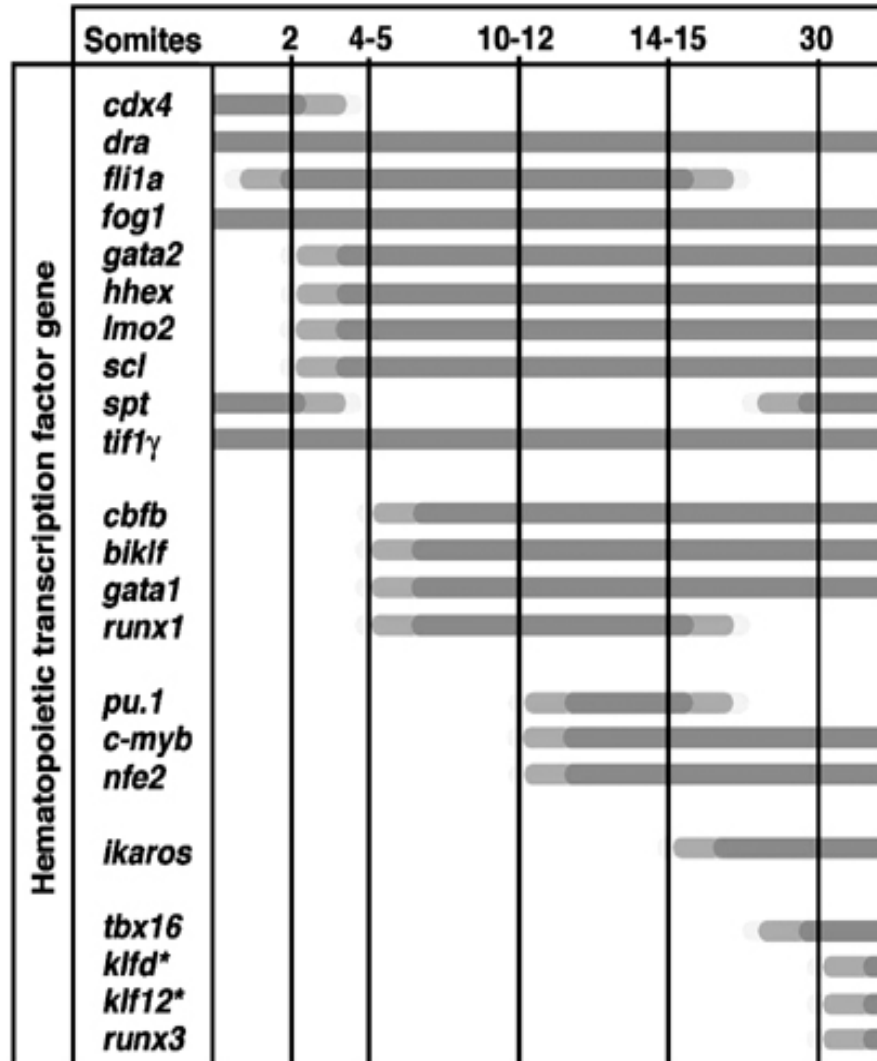
CD41<sup>+</sup>  
CD45<sup>+</sup>

*Runx1*<sup>-/-</sup>

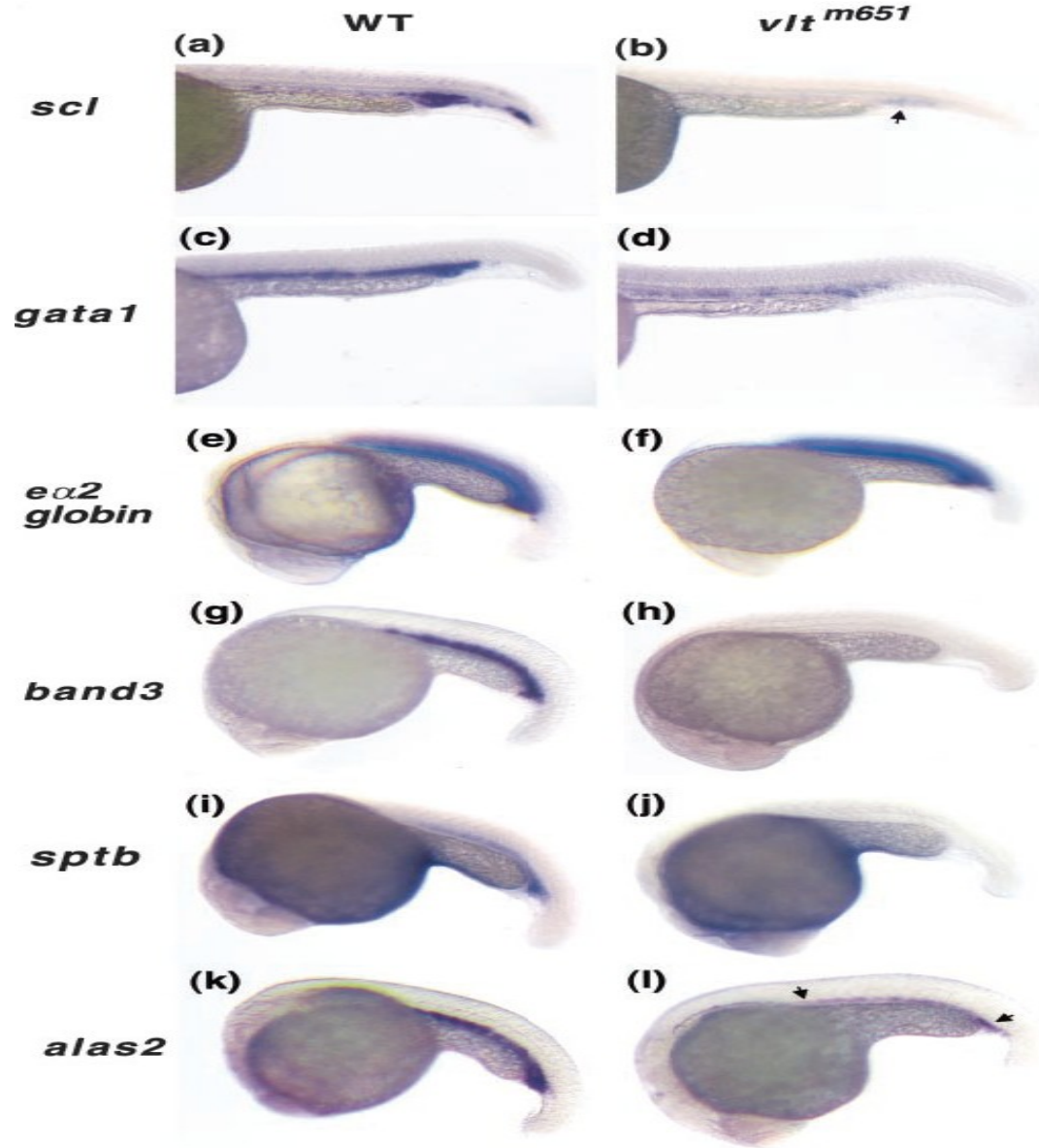




# TRANSCRIPTIONAL FACTORS IN BLOOD DEVELOPMENT



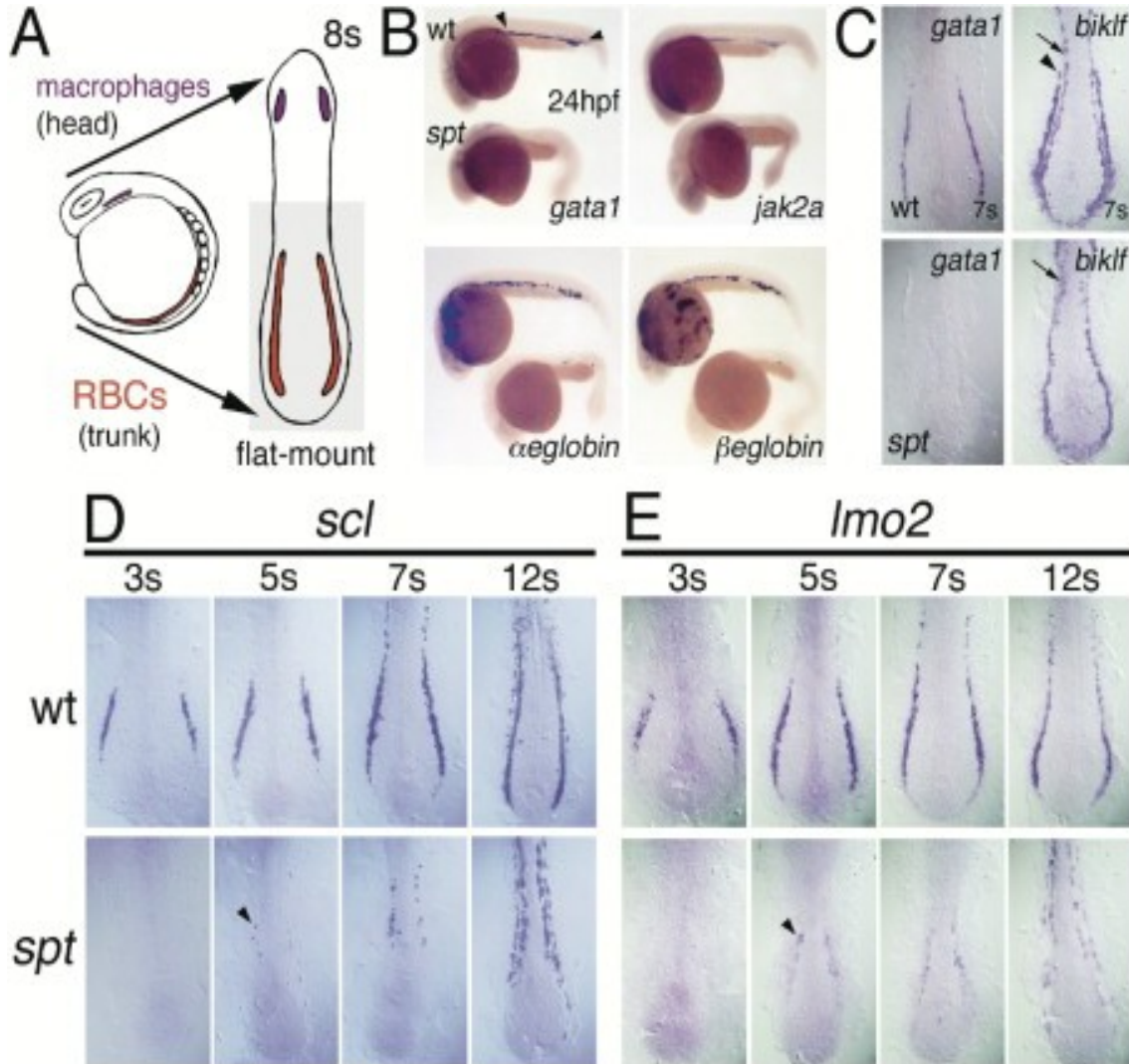
*GATA-1* - binds DNA via a zinc finger motif



**Dracula fish – loss-of-function mutation in GATA1 – impaired erythroid differentiation**

# Spadetail/TBX16

DNA binding domain derived from the prototype gene called transcription factor T



**impaired erythroid but not myelopoietic differentiation**

# Moonshine

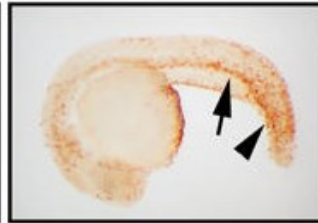
TIM-family of transcriptional factors

**B**

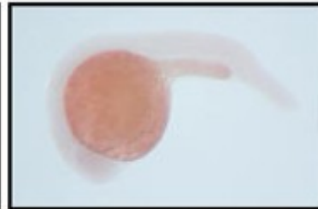
wild-type

*montg234*<sup>-/-</sup>

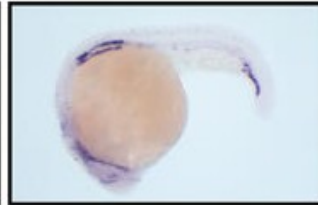
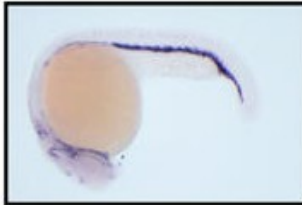
TUNEL



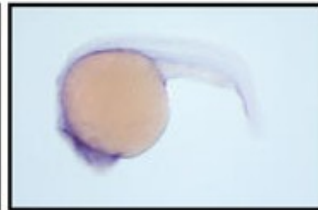
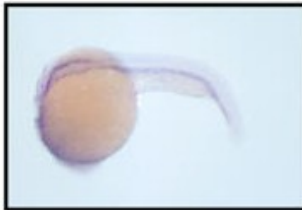
*gata1*



*scl*

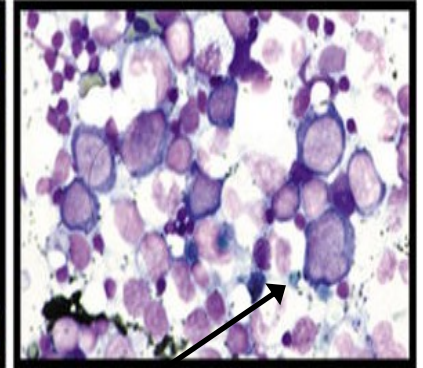
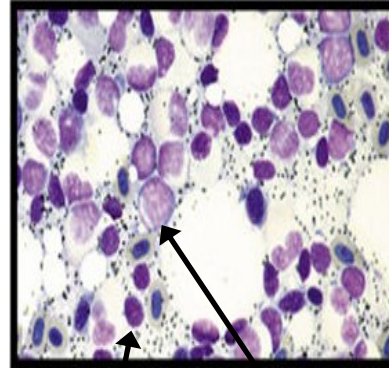


*gata2*



wild type

*mon*<sup>tb222</sup><sup>-/-</sup>



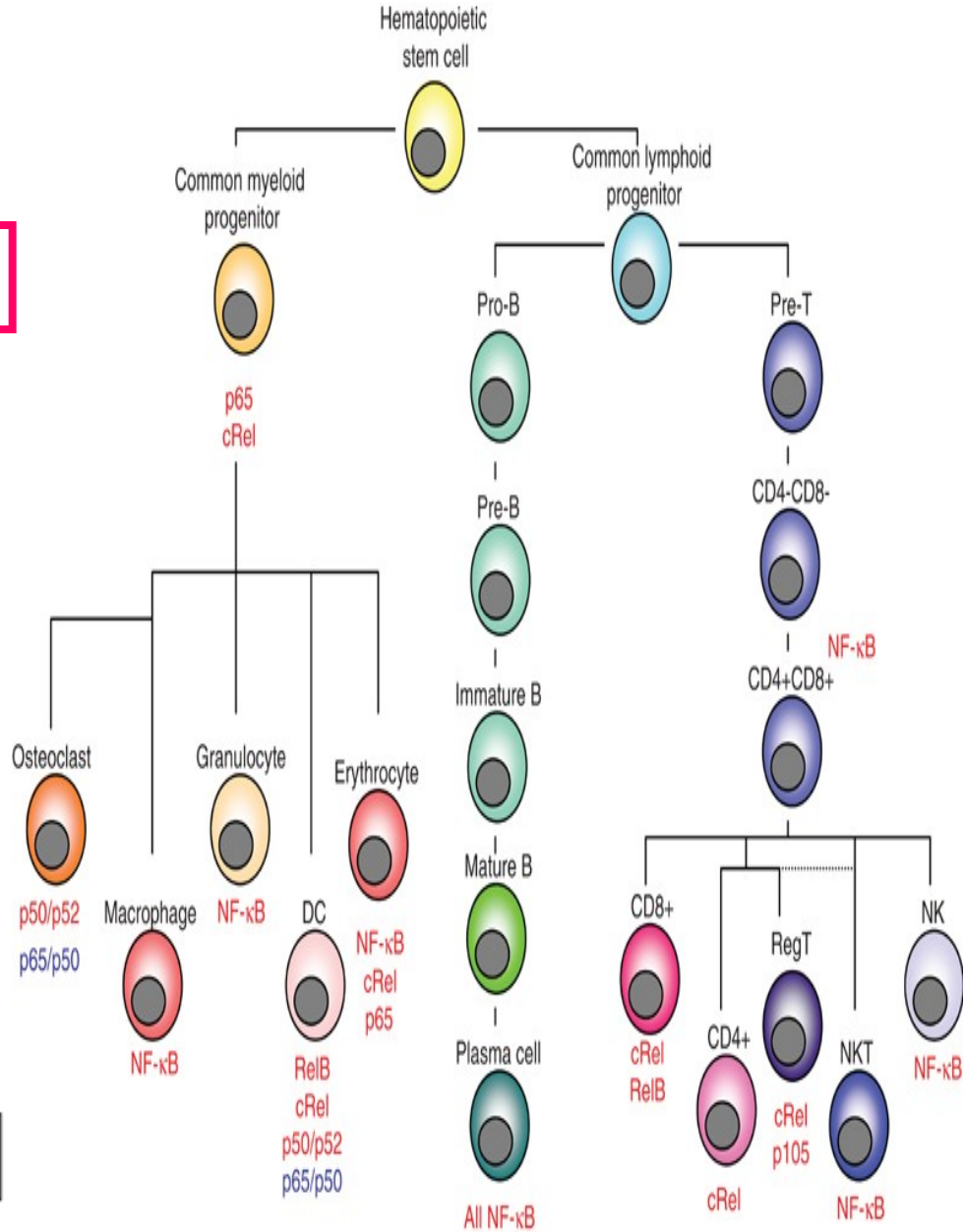
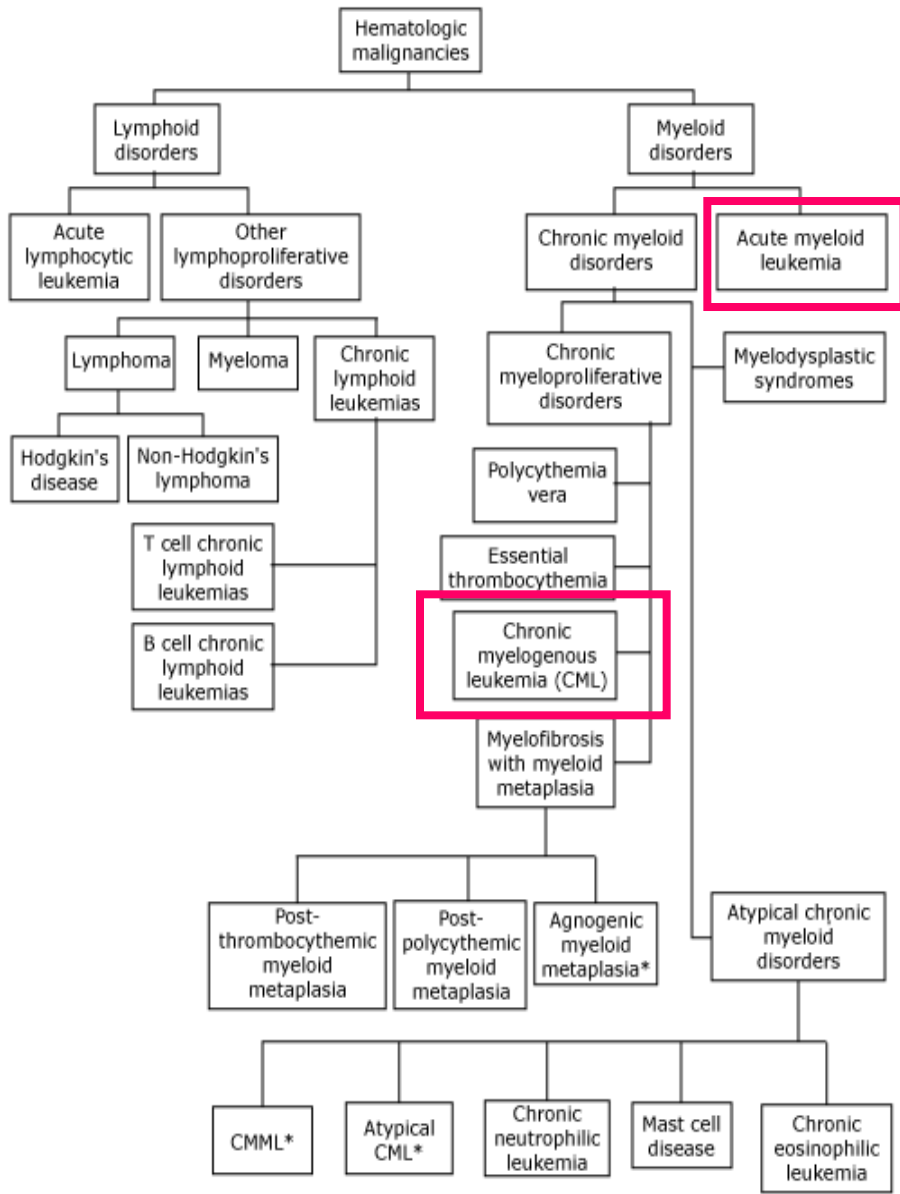
erythrocyte

proerythroblast

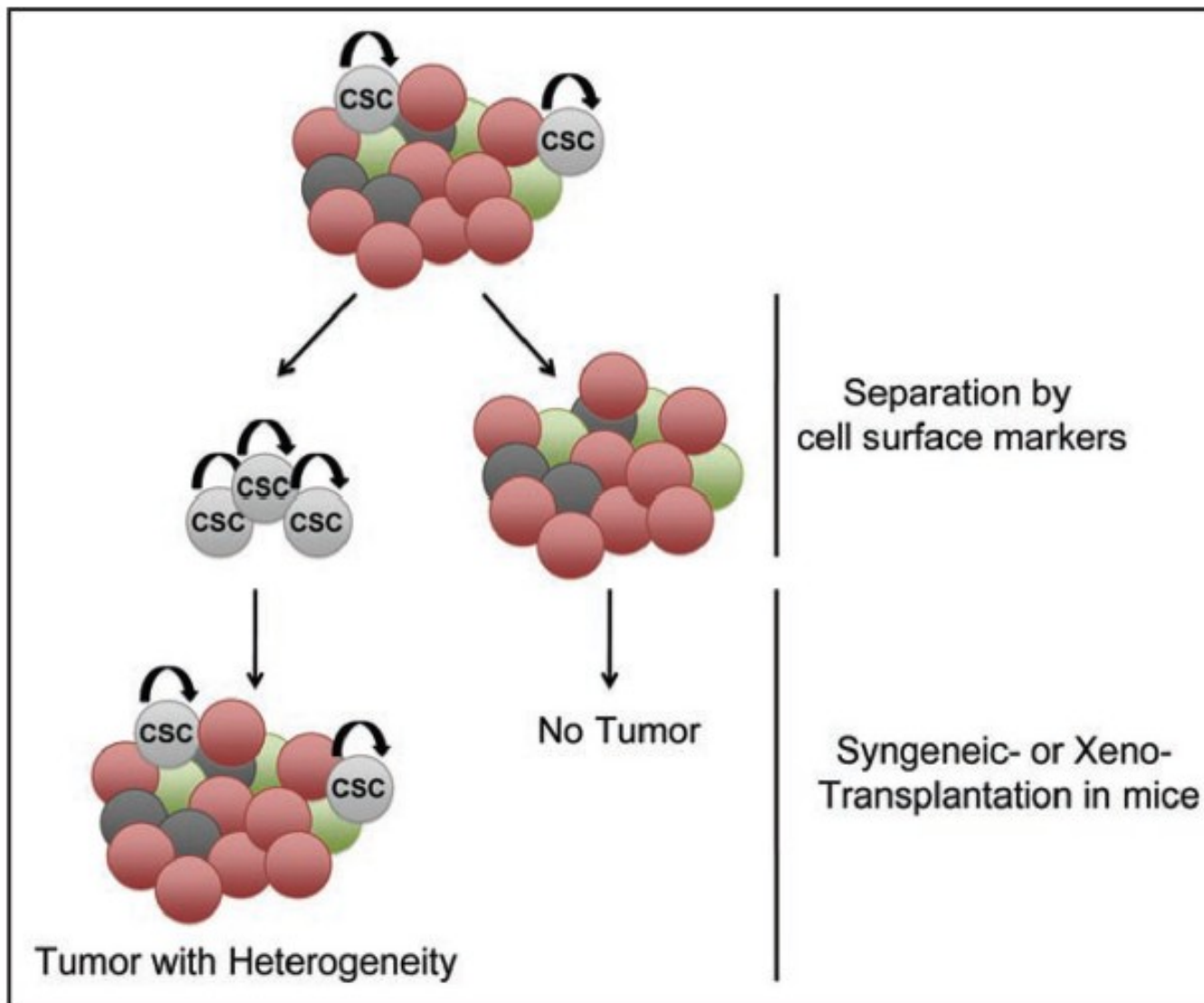
'primitive' – survival of HSC

'definitive' – cardiomegaly and impaired red cell differentiation

# WHEN SOMETHING GOES WRONG WITH BLOOD







**Figure 1.**

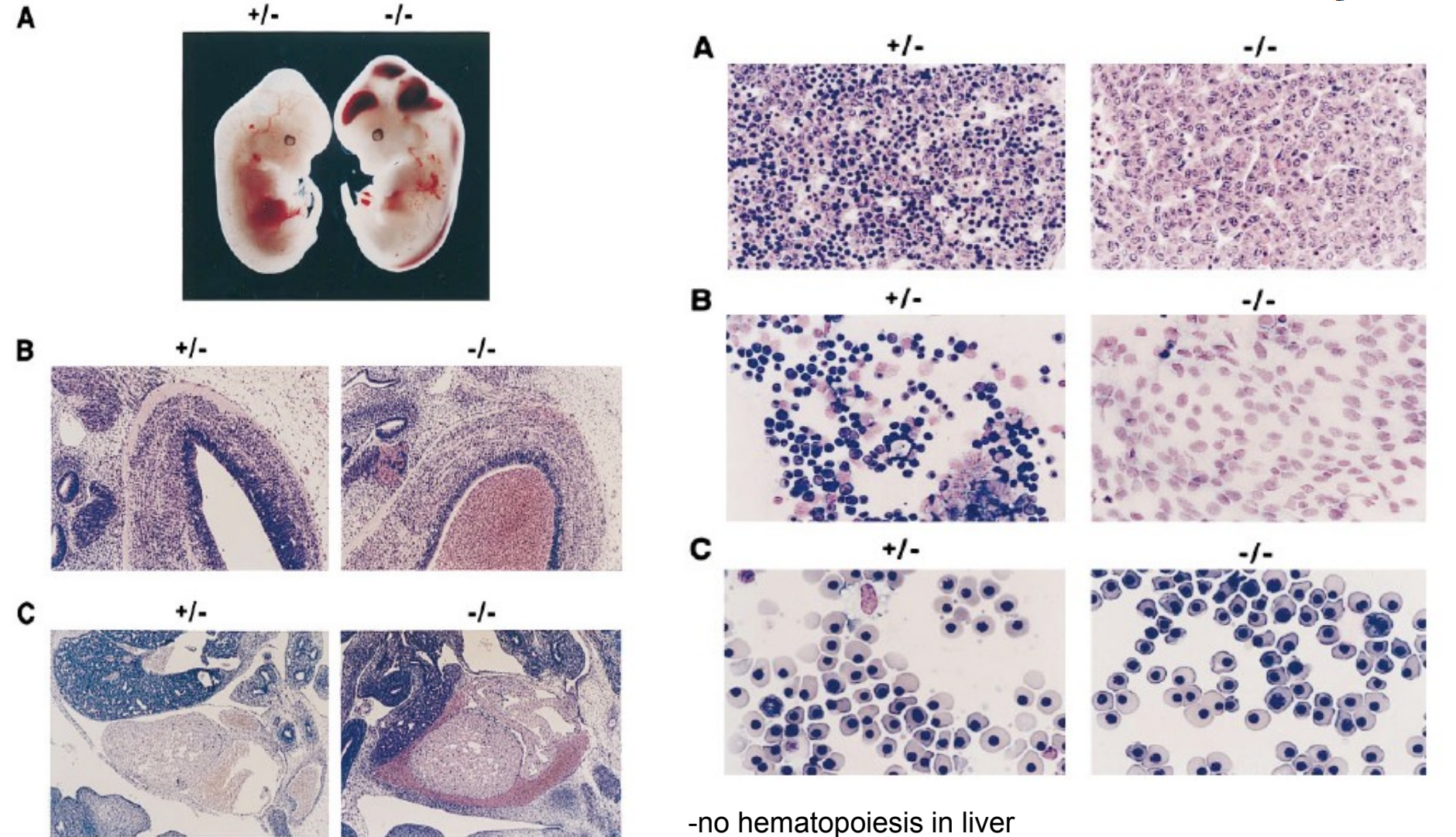
The Cancer Stem Cell Hypothesis. Only a specific subset of tumor cells, i.e., the cancer stem cells (CSC), is capable of forming tumors and generating the heterogeneous population of cells in a tumor. Arrows indicate self-renewal potential that is unique to CSC.





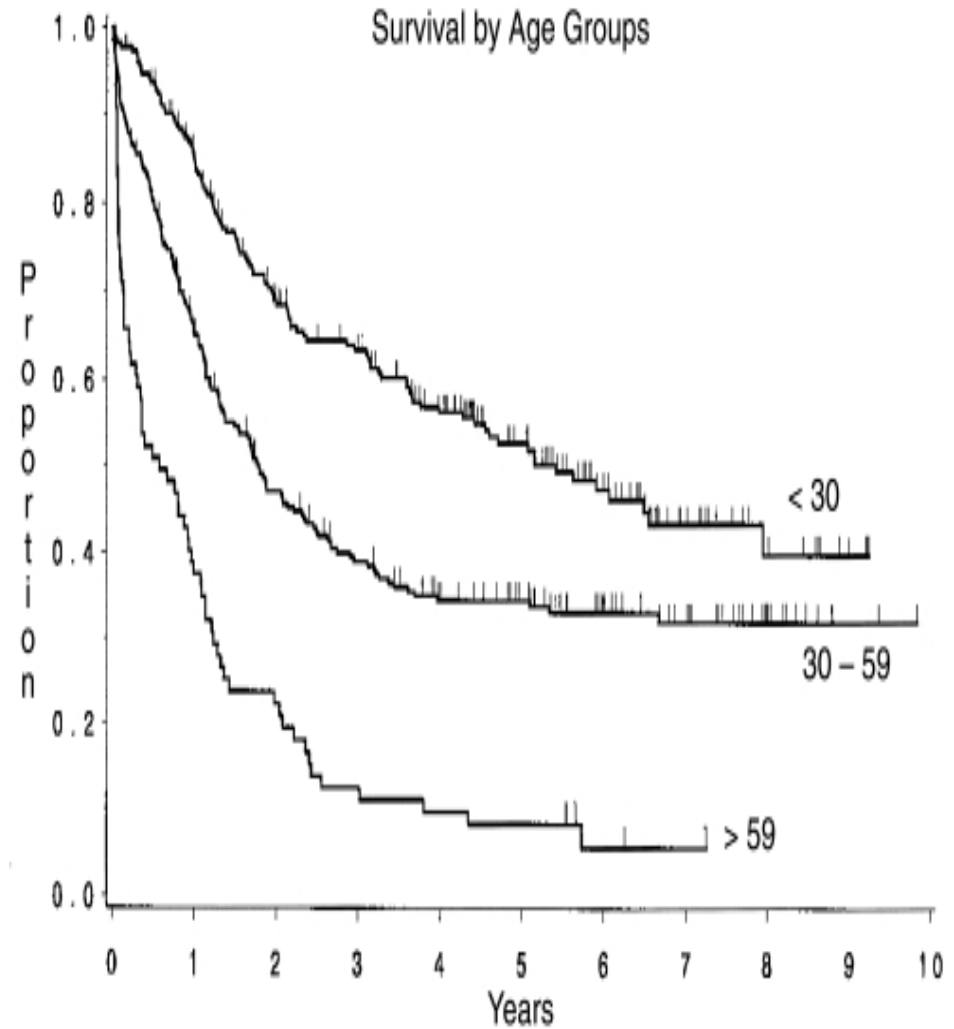
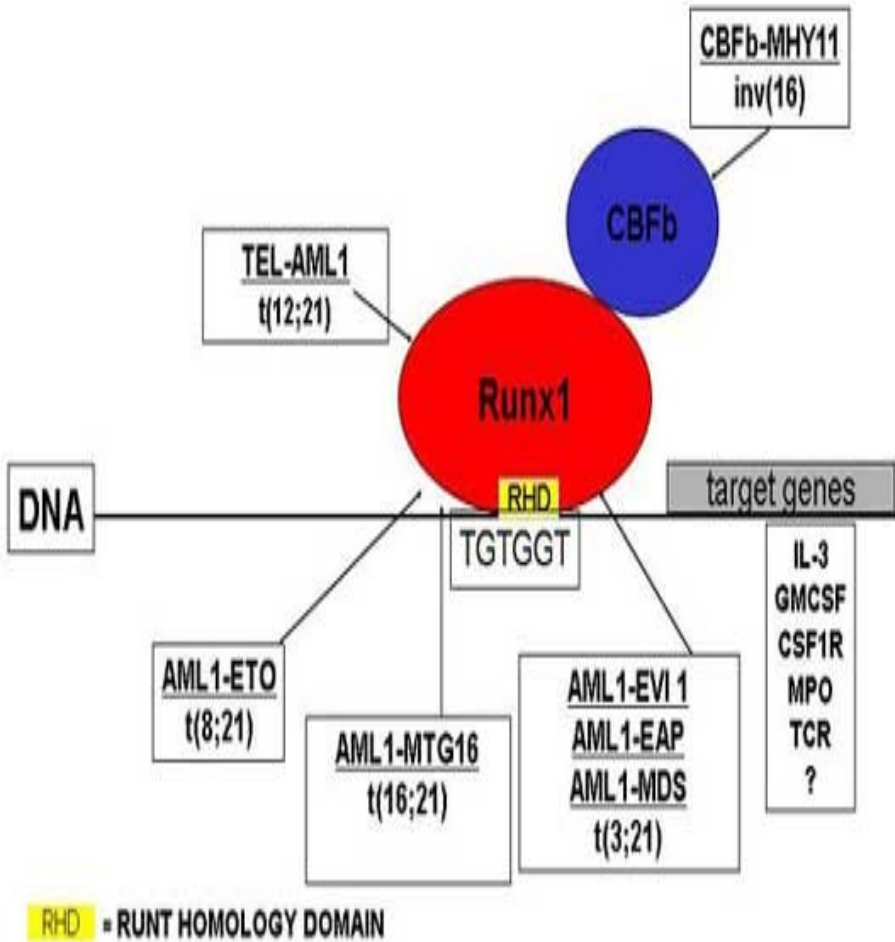
# AML1, the Target of Multiple Chromosomal Translocations in Human Leukemia, Is Essential for Normal Fetal Liver Hematopoiesis

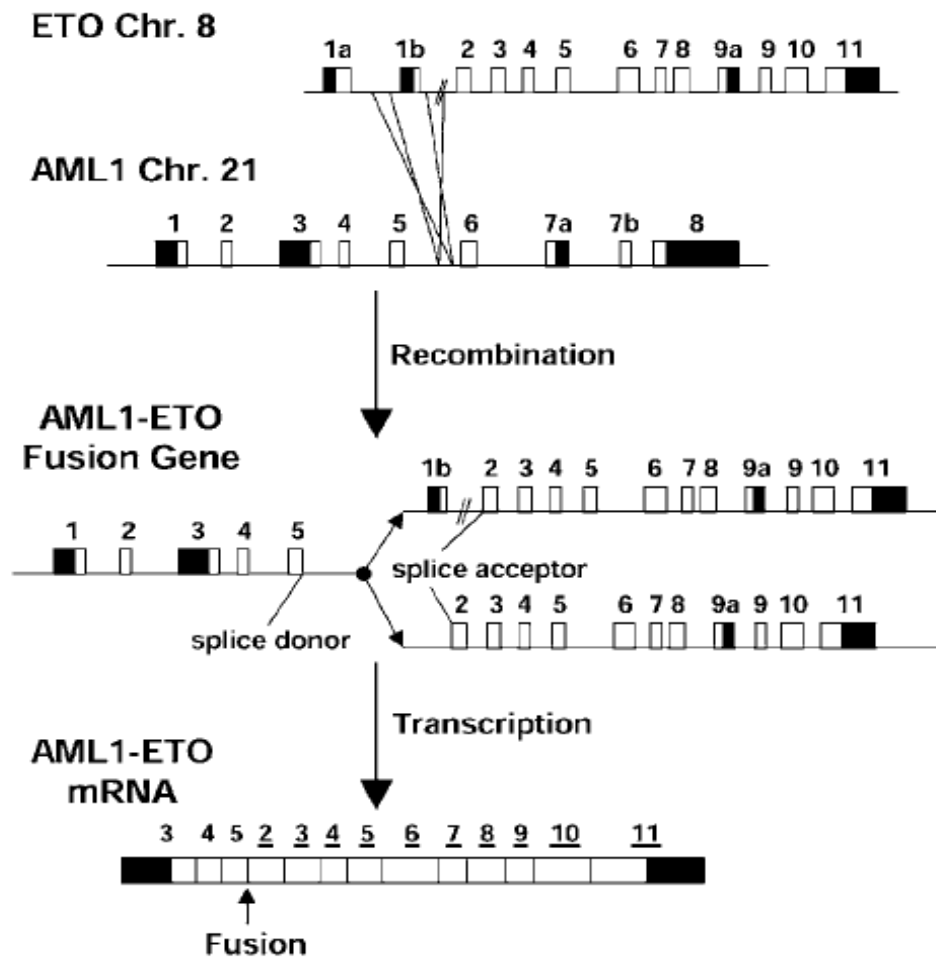
Cell, Vol. 84, 321-330, January 26, 1996,



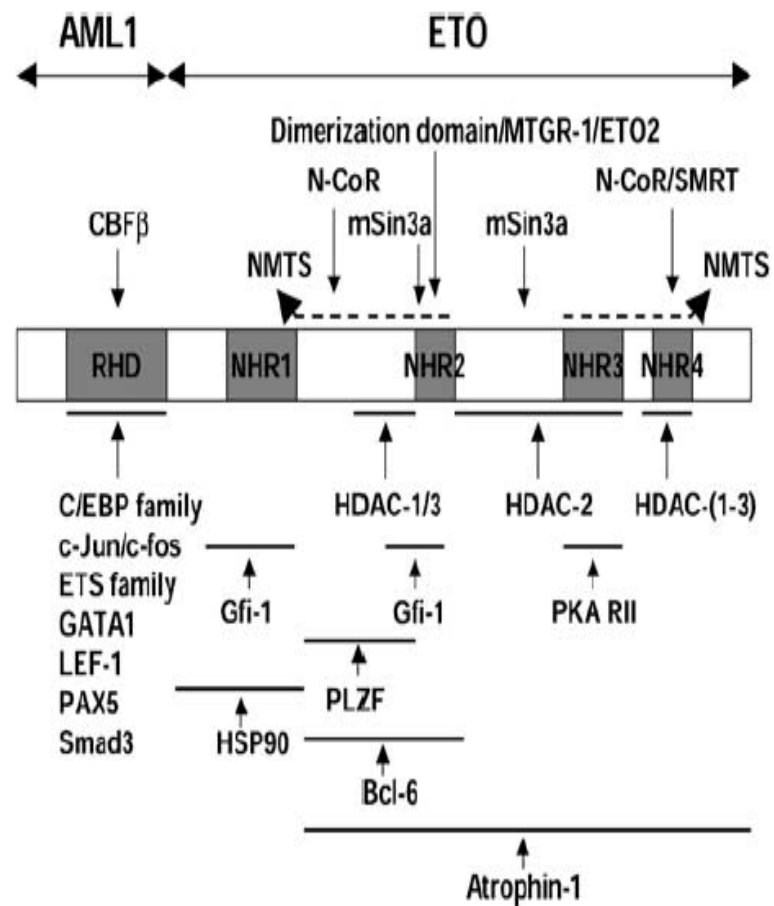
-no hematopoiesis in liver  
-only primitive erythrocytes in peripheral blood, no platelets

# ACUTE MYELOID LEUKEMIA





**Figure 1** Genomic structure of t(8;21). Chromosome 8 containing the *ETO* gene is made up of 13 exons spanning approximately 87kb, which can give four alternative splice forms and is regulated by two promoters. Chromosome 21 contains the *AML1* gene with nine exons that give various alternative splice forms and is regulated by two promoters and spans 260kb. The breakpoint cluster areas are denoted by the crossing lines between *ETO* and *AML1*. Owing to the absence of a splice acceptor in exon 1b of *ETO*, the mRNA of the fusion transcripts does not include this exon. White boxes and black boxes indicate translated and untranslated exon sequences, respectively. Underlined numbers in the *AML1-ETO* mRNA denote exons contributed by the *ETO* gene

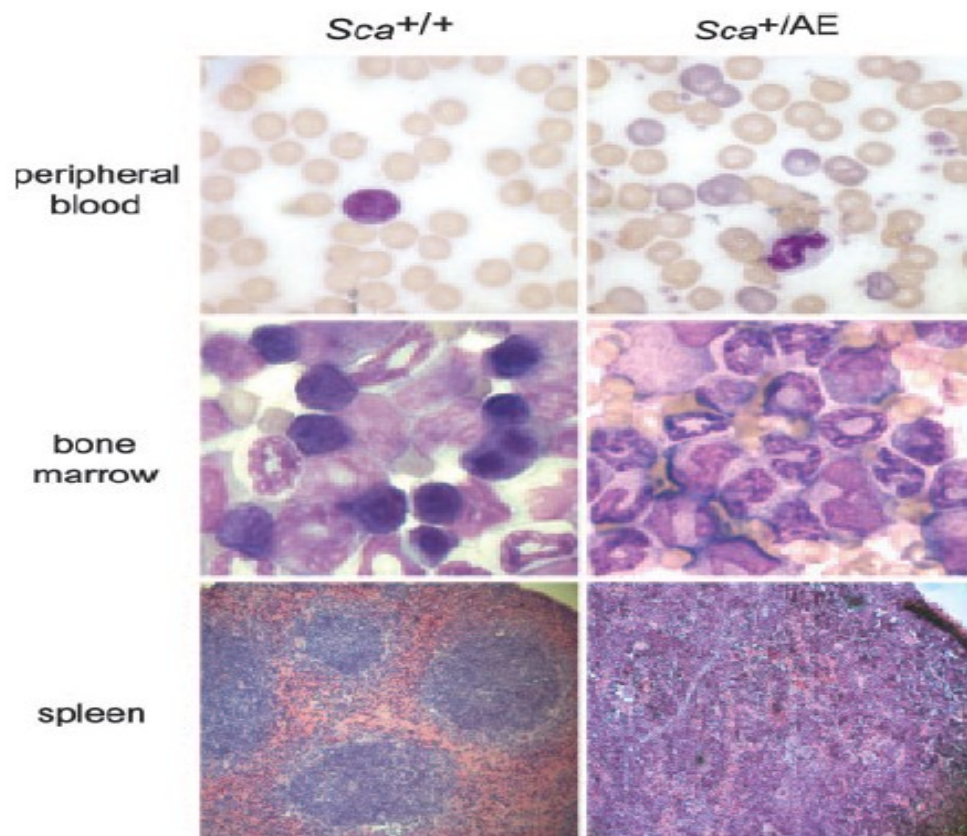


**Figure 2** AML1-ETO-interacting proteins. AML1-ETO contains the N-terminal sequences of AML1, including the RHD. Subsequent a.a.'s are of ETO, containing the four NHR1-4. Indicated as well are the regions containing the NMTS of ETO (broken line arrows). Known RHD and ETO/AML1-ETO-interacting proteins (or family of proteins) are shown. The dimerization domain of AML1-ETO and ETO family members is located in NHR2

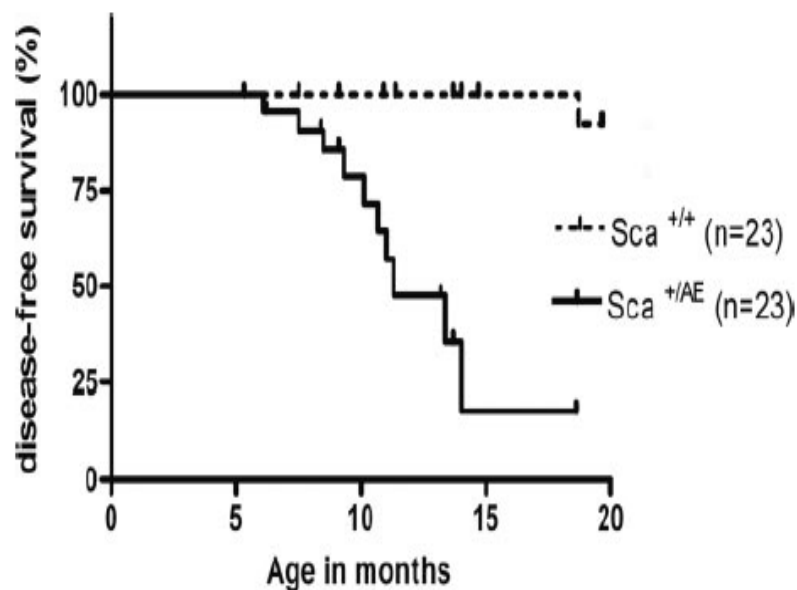


# Stem cell expression of the AML1/ETO fusion protein induces a myeloproliferative disorder in mice

*Sca1* – locus active in hematopoietic stem cells in adult mice



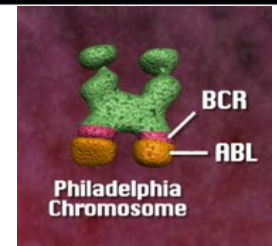
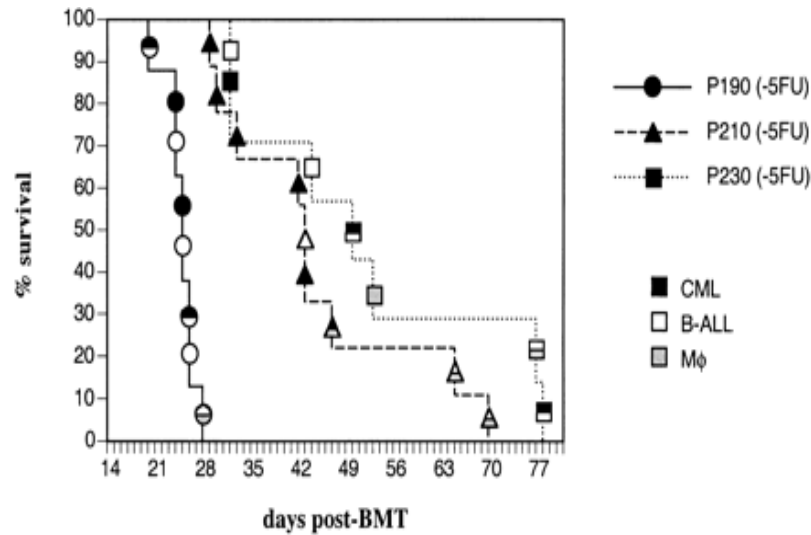
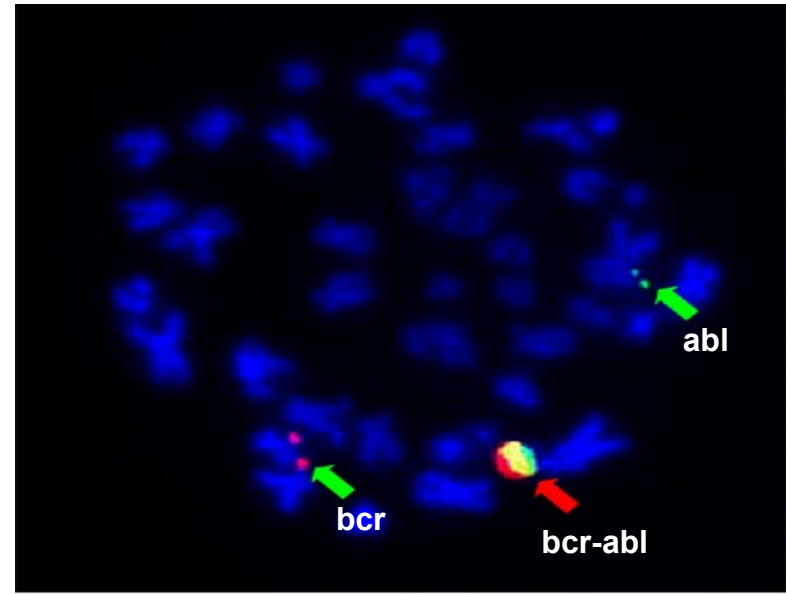
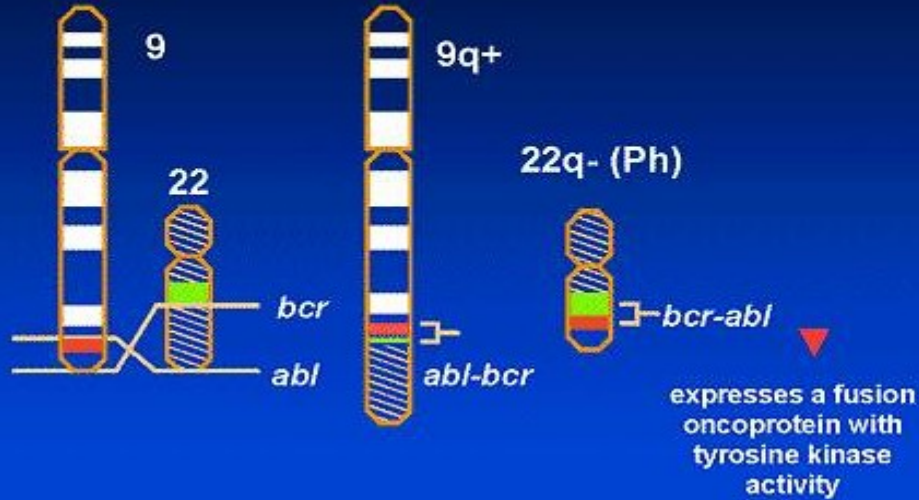
**Fig. 4.** Morphologic analysis of MPD. Peripheral blood and bone marrow slides show evidence of myeloid hyperplasia with loss of erythroid and lymphoid precursors in the bone marrow and polychromasia in peripheral blood of *Sca*<sup>+/AE</sup> mice compared with WT littermate mice. Increased extramedullary hematopoiesis with disruption of follicular architecture is evident in the spleens of *Sca*<sup>+/AE</sup> mice.



**Fig. 5.** Survival analysis. Kaplan-Meier plot demonstrates high penetrance and long latency of a nonlethal MPD in *Sca*<sup>+/AE</sup> mice compared with WT littermates ( $P < 0.0001$ ).

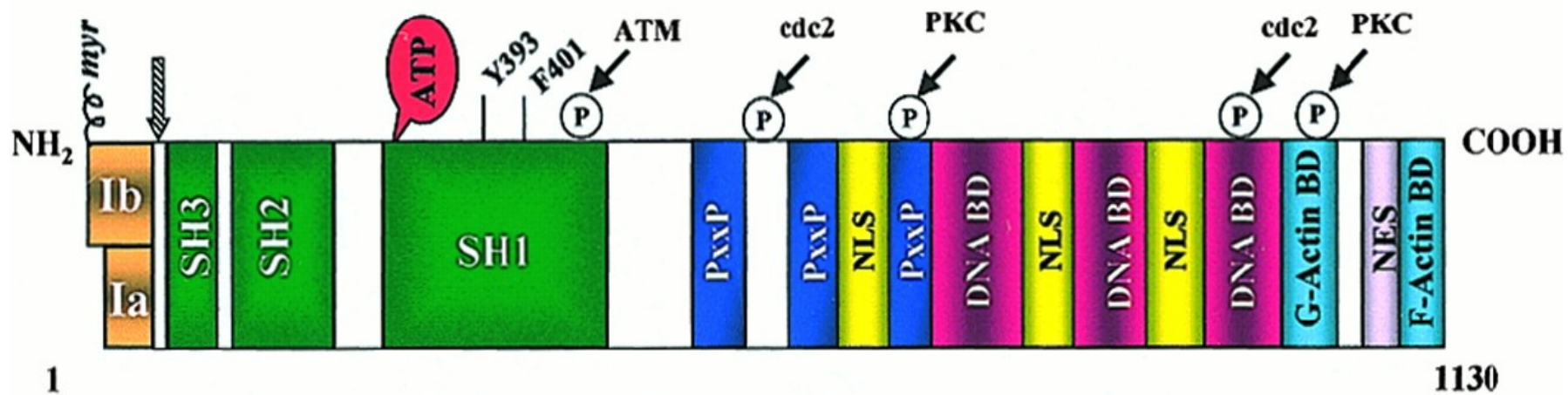
# CHRONIC MYELOID LEUKEMIA

The t(9;22) translocation produces the Philadelphia (Ph) chromosome

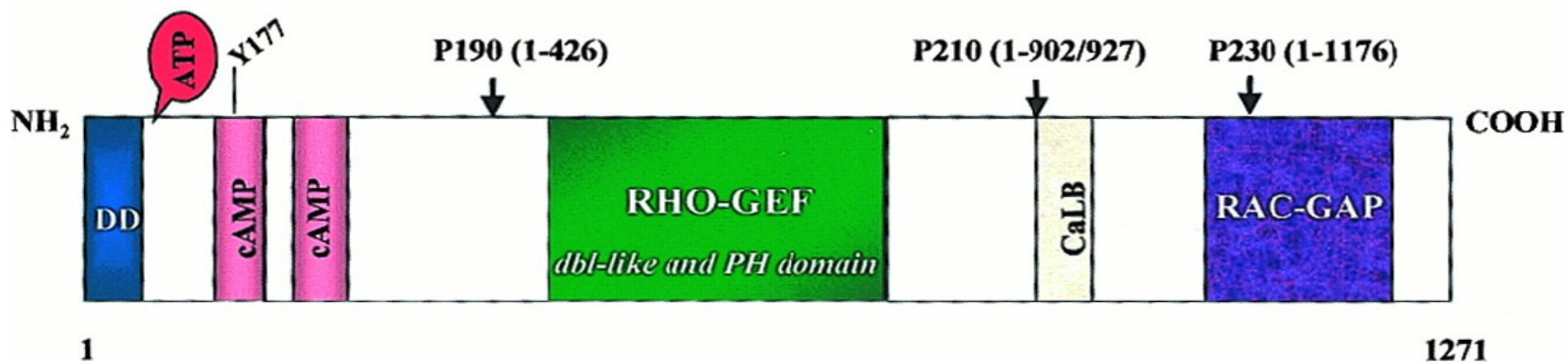


mice transplanted with patient bone marrow

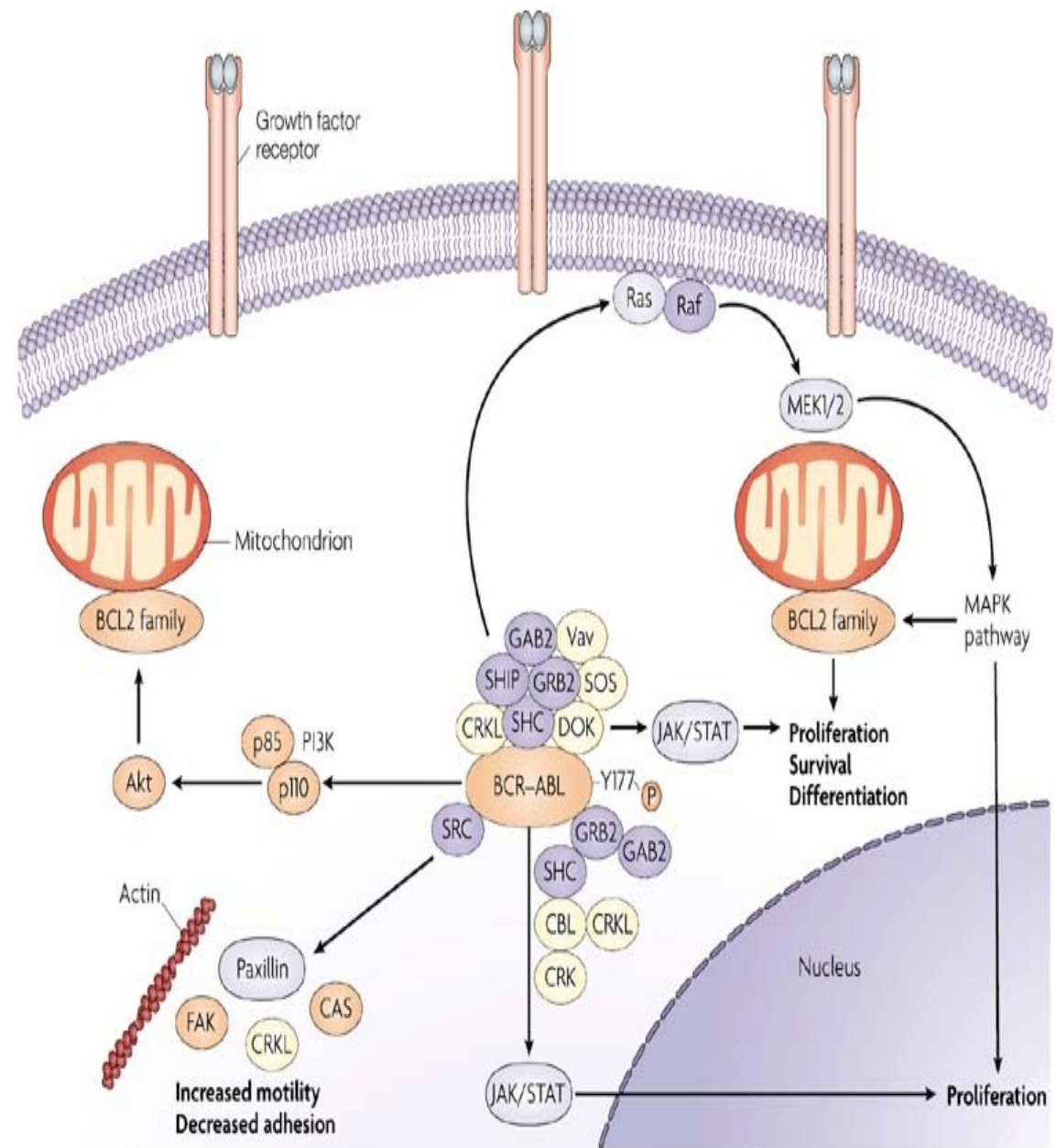
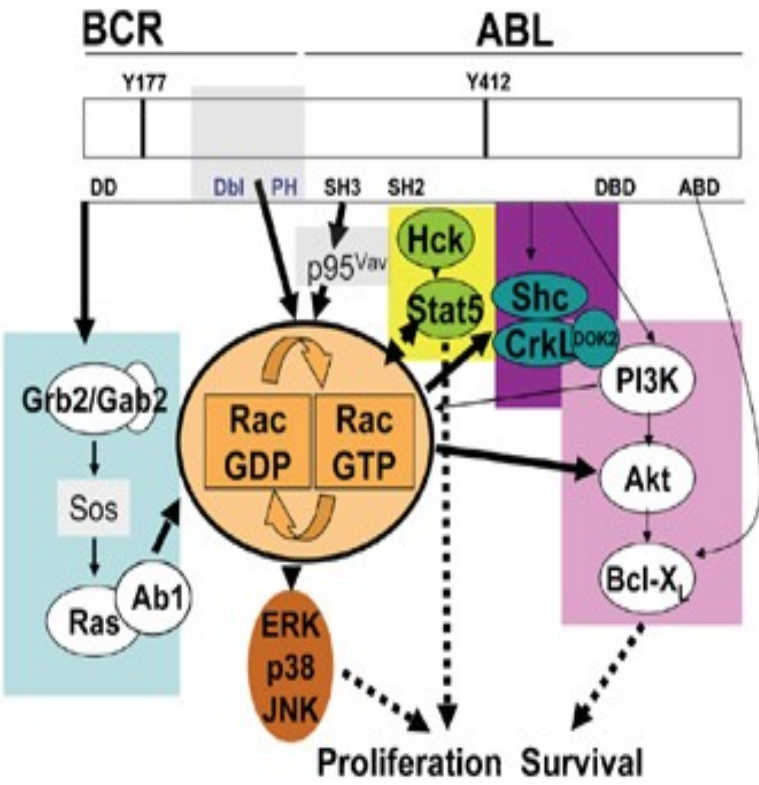
# p145 ABL



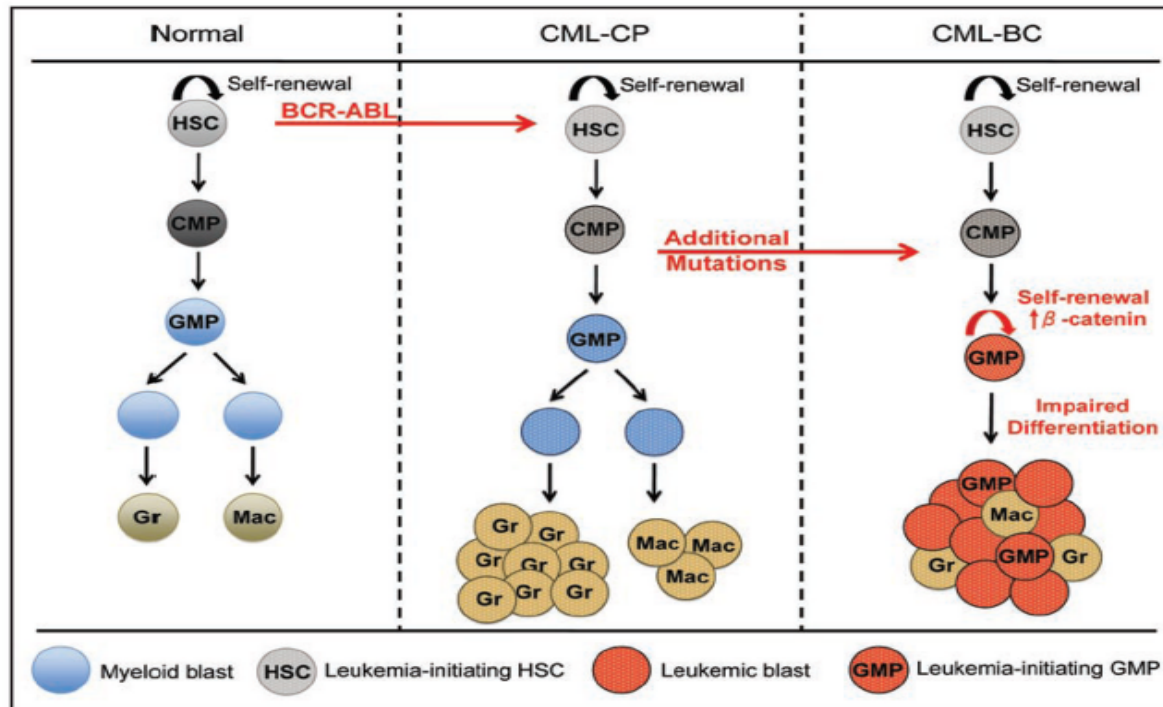
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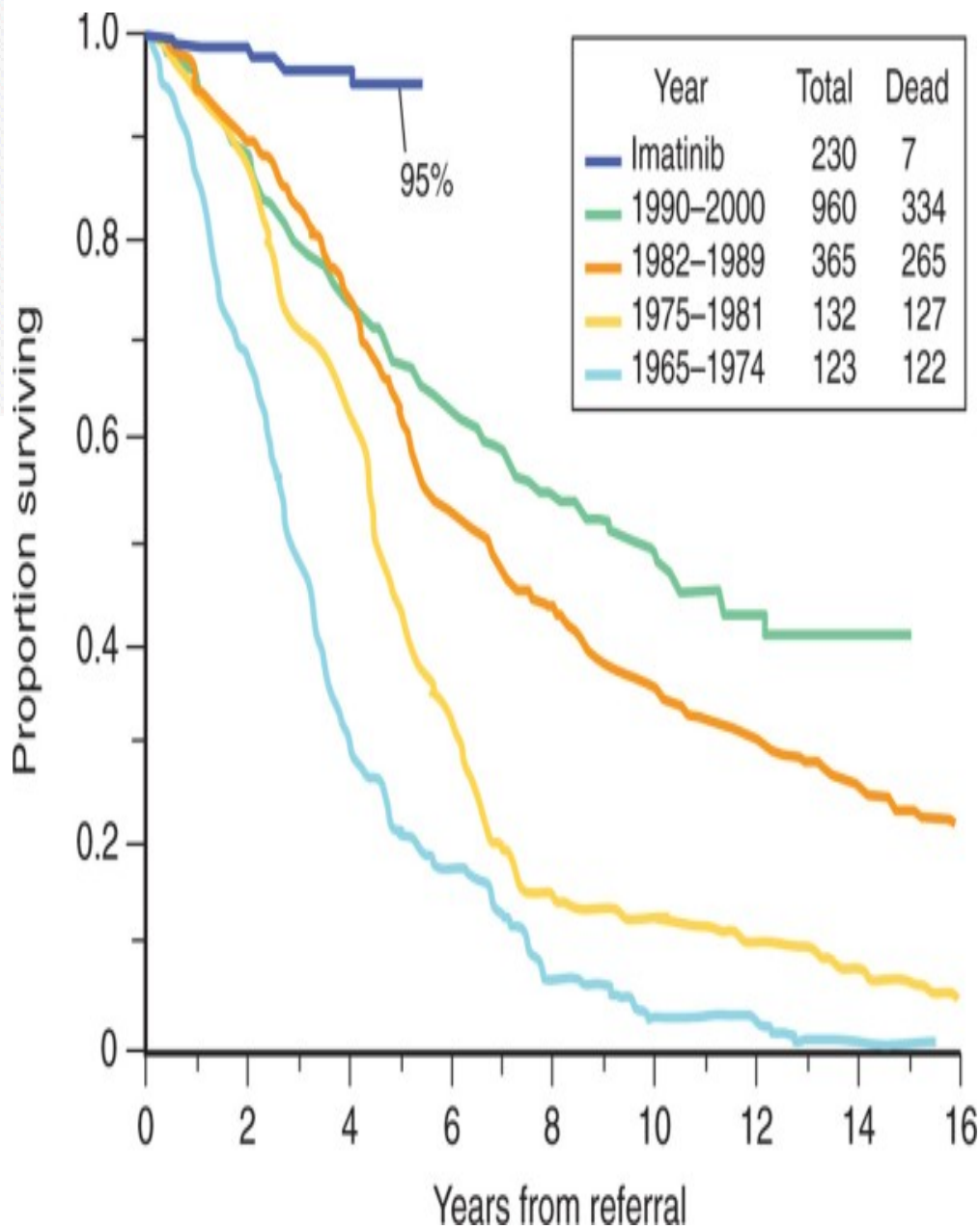
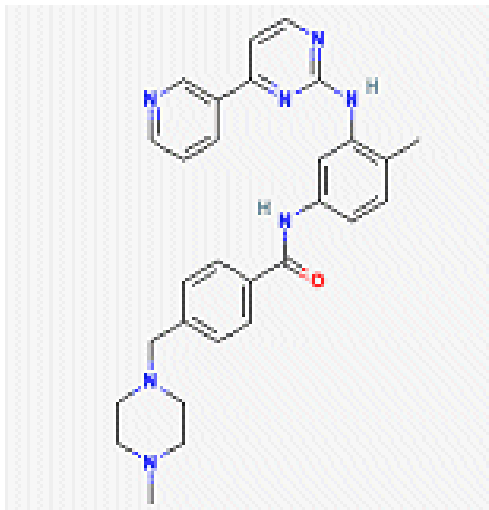
Am J Med. 1977 Jul;63(1):125-30. **Chronic myelocytic leukemia: clonal origin in a stem cell common to the granulocyte, erythrocyte, platelet and monocyte/macrophage.** [Fialkow PJ](#), [Jacobson RJ](#), [Papayannopoulou T](#).



**Figure 2.**

Evolution of Leukemic Stem Cells in CML Disease Progression. In normal hematopoiesis, the hematopoietic stem cell (HSCs) gives rise to all other lineages of hematopoietic cells. In CML chronic phase (CP) patients, expression of BCR-ABL in the HSC compartment leads to an expansion of the myeloid lineage resulting in an abnormal number of mature granulocytes. The HSC likely functions as the leukemia-initiating cell during CML-CP. Transition to blast crisis (BC) involves additional genetic and epigenetic alterations leading to the accumulation of immature blasts. Progression to blast crisis also results in the acquisition of self-renewal potential by a GMP population with elevated  $\beta$ -catenin activity. In a mouse model, as few as fifty cells from the BCR-ABL-transformed GMP compartment can initiate a CML-like disease. GMP from CML-BC patients also initiate leukemia when transplanted into immunocompromised mice. Arrows mark critical events in disease progression and self-renewal potential. Dotted circles indicate BCR-ABL-positive cells. (HSC, hematopoietic stem cell; CMP, common myeloid progenitor; GMP, granulocyte-macrophage progenitor; Gr, granulocyte; Mac, macrophage).





### Gleevec: HOW IT WORKS

