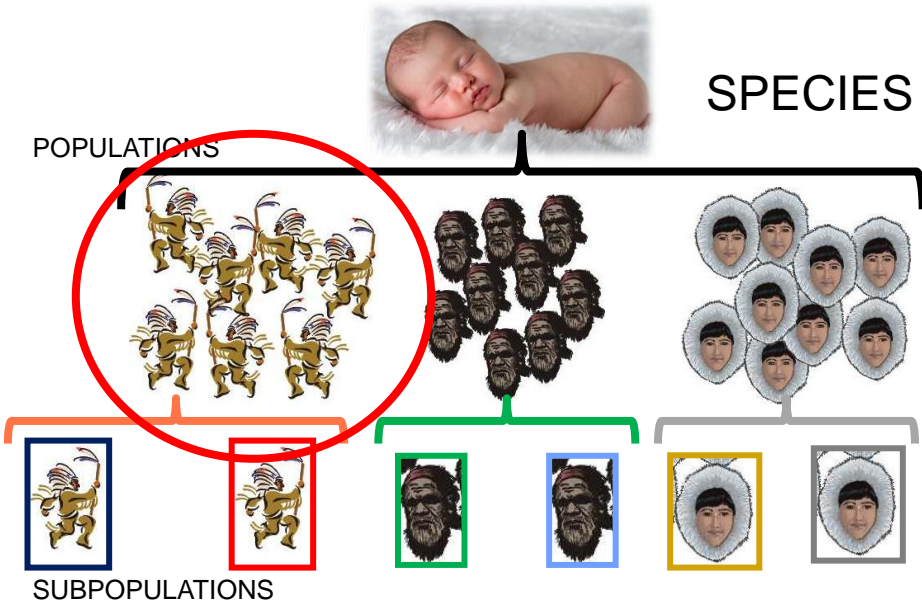
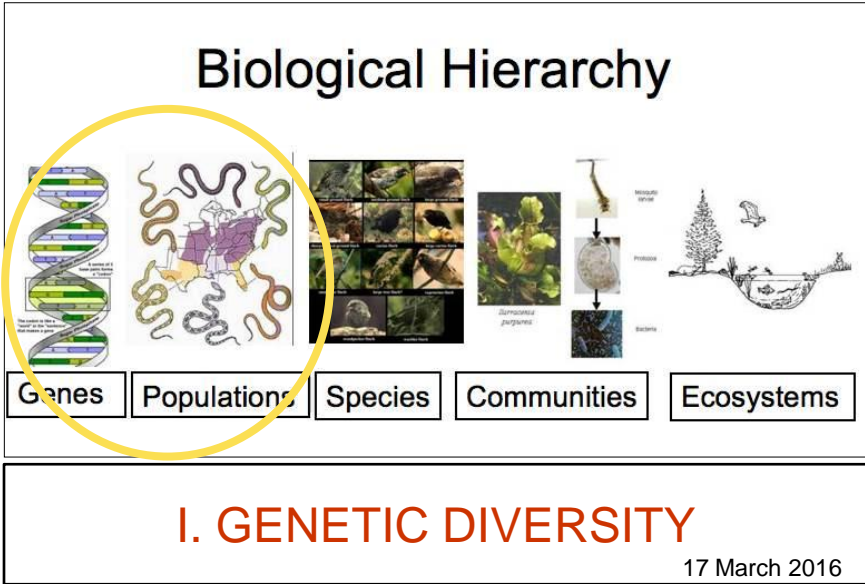


POPULATION GENETICS



I. GENETIC DIVERSITY – ANALYSIS OF SINGLE POPULATIONS

POPULATION

and problems of definition

- a population is a group of interbreeding individuals that exist together in time and space
- to develop the basic concepts of population genetics, we initially consider the **ideal population** = large, random-mating

ALLELE FREQUENCY

- proportion of an allele in comparison to all the other alleles of the same locus (gene) in a population sample
- basic characteristics for genetic diversity (variation) of a population
- population genetics studies genetic diversity and processes that have created it and influence it – i.e. the dynamics of distribution and frequency of alleles (genotypes → phenotypes), i.e. processes shaping **evolution**:

increase of gen. diversity: **mutation** and **migration**

decrease of gen. diversity: **genetic drift** (and **natural selection**)

MUTATIONS

increase genetic diversity
responsible for variation/heterogeneity in
populations – essential to **evolution**

1. substitutions (transitions, transversions)

non-coding regions

synonymous } = silent substitutions
GTC → GTA
Val → Val

nonsynonymous

missense

GTC → TTC

Val → Phe

nonsense

AAG → TAG

Lys → ochre (stop)

2.

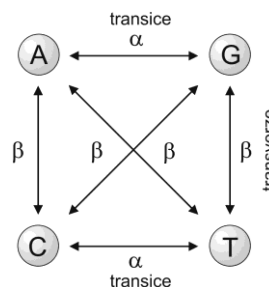
insertion

ACGGT → ACAGGT

deletion

ACGGT → AGGT

} = indels
→ frameshift mutations



Mutation rate – rate at which number of various types of
mutations occur in a given position over time

OBSERVATION

*Callimorpha
dominula*

přástevník
hluchavkový

Scarlet tiger moth



OBSERVATION

Callimorpha dominula

přástevník
hluchavkový

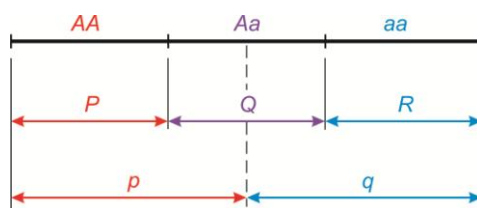
Scarlet tiger mo

Table 3.1. Data from a collection of 1612 scarlet tiger moths.

| Phenotype | No. of individuals |
|-----------------|--------------------|
| White spotting | 1469 |
| Intermediate | 138 |
| Little spotting | 5 |



Genotype and allele frequency



Relative numbers = frequencies: genotype f.: $P (G_{AA})$, $Q (G_{Aa})$, $R (G_{aa})$

allele (gene) f.: $p (A)$, $q (a)$

$$P + Q + R = 1$$

$$p + q = 1$$

| | | | | |
|-----------|-----------------------|-------------|-----------------------|-------|
| Genotype | A_1A_1 | A_1A_2 | A_2A_2 | Total |
| Number | n_1 | n_2 | n_3 | N |
| Frequency | $P = n_1/N$ | $Q = n_2/N$ | $R = n_3/N$ | |
| | $p = (2n_1 + n_2)/2N$ | | $q = (n_2 + 2n_3)/2N$ | |

Hardy-Weinberg Equilibrium (HWE)

Ex. Single locus with 2 alleles

| Allele | Allele frequency |
|--------|------------------|
| A | p |
| a | q |

$p + q = 1$
 p, q - Allele frequencies known from our samples

| Genotype | Expected genotype frequency |
|----------|-----------------------------|
| AA | p^2 |
| Aa | $2pq$ |
| aa | q^2 |

= Hardy-Weinberg equilibrium

➤ Observed genotype frequencies (H_o) are known from our samples

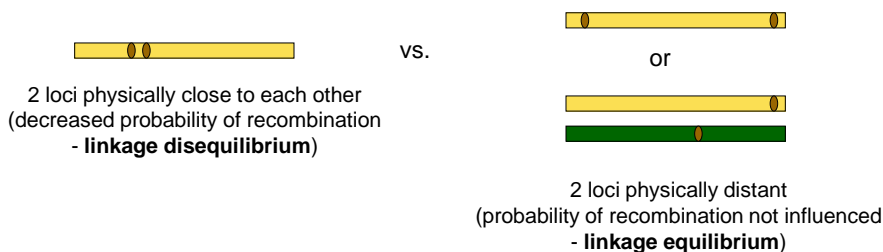
➤ deviation of H_o from HWE conditions \Rightarrow for example χ^2 test

Expected heterozygosity, (H_e) under HWE

$H_e = 1 - (p^2 + q^2)$ for 1 locus with the allele frequencies p and q

Assumptions for ideal population in HWE

- random-mating
- negligible effect of mutations and migration („closed populations“)
- infinitely large population (negligible effect of random fluctuations in allele frequencies in time – genetic drift) – **in HWE population the allele frequencies are stable = do not change between generations**
- Mendelian inheritance of the analysed loci
- neutral loci – not under selection
- diploid, sexually reproducing organisms with discrete generations
- loci are independent from each other – test for „linkage disequilibrium“



LINKAGE DISEQUILIBRIUM (LD)

loci in LINKAGE EQUILIBRIUM – segregate independently of each other during meiosis

the most common reason for non-random association among loci (LD) is the **proximity of two loci on a chromosome** (others e.g. small pop. size – gen. drift, immigration, overlapping generations, admixture, etc.)

haplotype diversity – $p(AB) \neq p(A) \times p(B)$

in presence of LD:

we have **fewer** independent loci for our genetic analysis than anticipated

neutral loci (alleles) linked to selected ones will appear non-neutral

presence of LD **needs to be tested** when analysing data from multiple loci

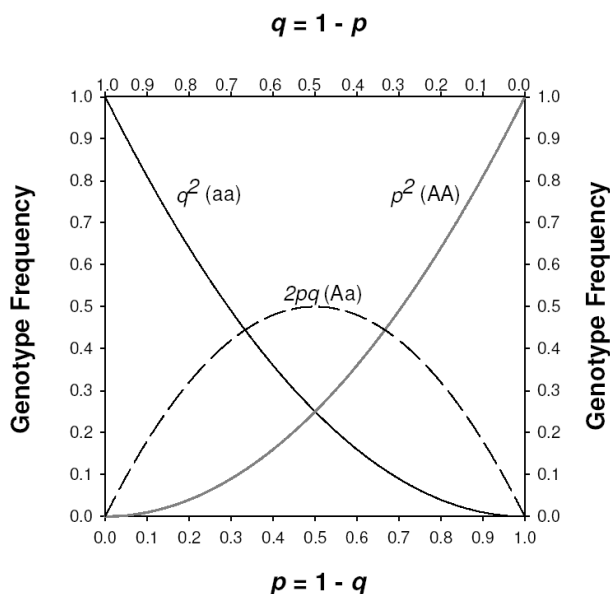


Figure 3.4 The combinations of homozygote and heterozygote frequencies that can be found in populations that are in HWE. Note that the frequency of heterozygotes is at its maximum when $p = q = 0.5$. When the allele frequencies are between $1/3$ and $2/3$, the genotype with the highest frequency will be the heterozygote.

Example of genetic diversity estimation in a sample of 4 individuals (on 4 loci)

| Individual | Locus 1 | Locus 2 | Locus 3 | Locus 4 | Average |
|------------|---------|-------------------|---------|---------|--------------|
| Ind 1 | 170/170 | 223/227 | 116/116 | 316/316 | |
| Ind 2 | 170/172 | 223/225 | 112/112 | 316/316 | |
| Ind 3 | 172/172 | 223/225 | 112/112 | 316/316 | |
| Ind 4 | 170/172 | 223/227 | 112/112 | 316/316 | |
| Počet alel | 2 | 3 | 2 | 1 | 2 |
| H_o | 0,5 | 1,00 | 0 | 0 | 0,375 |
| p | 0,5 | p = 0,5 | 0,75 | 1,00 | |
| q | 0,5 | q = 0,25 r = 0,25 | 0,25 | 0 | |
| H_e | 0,5 | 0,625 | 0,375 | 0 | 0,375 |

$$H_e = 1 - (p^2 + q^2)$$

Proportion of polymorphic loci (polymorphism) = 0,75

$$H_e = 1 - (p^2 + q^2 + r^2)$$

Is our population in HWE?

Callimorpha dominula

přástevník
hluchavkový

Scarlet tiger moth





Is our population in HWE?

Table 3.1. Data from a collection of 1612 scarlet tiger moths.

| Phenotype | No. of individuals | Assumed genotype | No. of <i>A</i> alleles | No. of <i>a</i> alleles |
|-----------------|--------------------|------------------|-------------------------|-------------------------|
| White spotting | 1469 | <i>AA</i> | 1469x2=2938 | - |
| Intermediate | 138 | <i>Aa</i> | 138 | 138 |
| Little spotting | 5 | <i>aa</i> | - | 5x2=10 |

d the scarlet tiger moth, *Panaxia agnita* in the scoring of the *omiga*

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 SP, U.K.
 Oxford Polytechnic, Headington, Oxford OX3 9BP, U.K.

In gene frequency and usually among selection against
 melanism (summarized by Jones (1989)).
 In the present paper we suggest that one fact
 contributing to the Colwell findings may have been
 variability in the scoring of melanisms by different
 observers over many years. The evidence for this comes
 first from two Colwell reports, and second from our
 own studies using Winstan Way material.

omiga

the same

all have been

collected over

the course of the

collected black spot is sometimes replaced by a yellow

one (see figure 2) and larvae which also have a

larval spot are extremely similar to the typical for

one found (1975).

I. H. Cook (personal communication, 1980) has

noted that as far as he knows Colwell's entries were valid

inclusions as melanisms all made with the yellow

larval spot and no black whatever or was the correct

larval spot was retained. This view was also held by

Kitchinson (see the Reddish-Greenish-Reddish

Collection in the Natural History Museum, London

Levy (1970) confirms Cook's comments but also

notes larvae which produced all melanisms and 1



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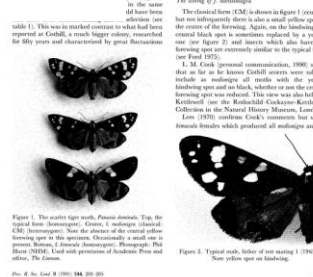


Figure 1. The scarlet tiger moth, *Panaxia agnita*. Top, the typical form (homotypic); Centre, *f. melanica* (homocoll.); Bottom, melanism; Note the absence of the central yellow larval spot in this specimen. Occasionally a small one is present (hetero-*f. melanica* homocoll.). Photographed by Denis F. OUVEN. Cook with permission of Academic Press and others, The Canon.

Figure 2. Typical moth larvae of one mating (1976). Note yellow spot on hindwing.

Proc. R. Soc. Lond. B (1981) 184, 205-209

20

Deviation from HWE

- **HWE test** – e.g. Genepop software („exact probability tests“) – any case of **significant deviations from HWE** indicates that some of HWE **assumptions were not fulfilled** → detailed inspection required:
- **heterozygote excess**
 - negative **assortative mating** (i.e. intentional mating of distinct individuals)
 - used loci are advantageous in heterozygote situation (= balancing **selection** favouring heterozygotes, e.g. MHC genes)
 - **mutation**
 - **migration**
- **heterozygote deficit**
 - **inbreeding** (all loci are equally affected), assortative mating
 - genetic **structure** in populations
 - **null alleles** (only some loci affected by heterozygote deficit)

Quantifying genetic diversity

Polymorphism (proportion of polymorphic loci) - P

- **polymorphic locus** = with at least two alleles with having frequency of more numerous allele being **less or equal 0.95** (or 0.99)
- e.g. a population sample with four polymorphic loci out of five → $P = 0.8$

Number of alleles - N_a

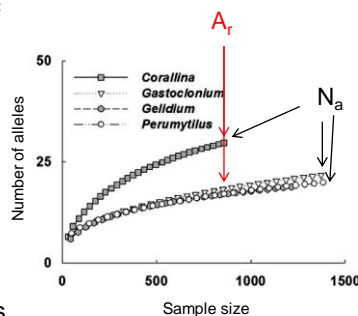
- number of alleles per locus (mean over loci)

Allelic richness - A_r

- number of alleles corrected for sample size (rarefaction method e.g. in FSTAT software)

Observed heterozygosity - H_o

- observed frequency of heterozygote genotypes (mean over loci)



HAPLOID DIVERSITY

- genetic diversity for haploid data

HAPLOTYPE DIVERSITY (h ; Nei et Tajima 1981) – frequency of different haplotypes

$$H = \frac{N}{N-1} \left(1 - \sum_i x_i^2 \right)$$

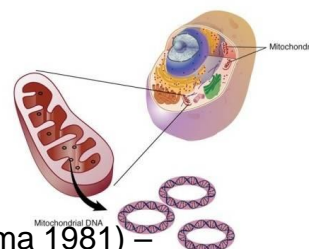
x_i – haplotype frequency of each haplotype in the sample
 N – sample size

NUCLEOTIDE DIVERSITY (π ; Nei 1987)

- quantifies the mean nucleotide divergence between sequences
- probability that two randomly chosen homologous nucleotides will be identical

$$\pi = \sum_{ij} x_i x_j \pi_{ij}$$

x_i and x_j – respective frequencies of the i th and j th sequences
 π_{ij} – number of nucleotide differences per nucleotide site between the i th and j th sequences



WHAT INFLUENCES GENETIC DIVERSITY?

- influenced by a multitude of factors
- varies considerably between populations


MOST IMPORTANT DETERMINANTS OF GENETIC DIVERSITY:

- genetic drift
- population bottlenecks
- natural selection
- methods of reproduction

GENETIC DRIFT

population not infinitely large → population not in HWE → increase of influence of CHANCE → allele frequencies vary between generations

in absence of selection, each allele goes to:

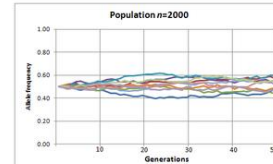
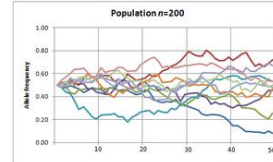
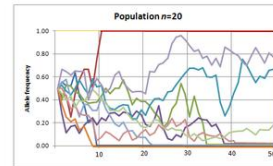
1. fixation
 2. extinction
- more quickly in smaller populations
-  **DECREASE of genetic diversity**

genetic drift – process causing a population's allele frequencies to change from one generation to the next as a result of **CHANCE**

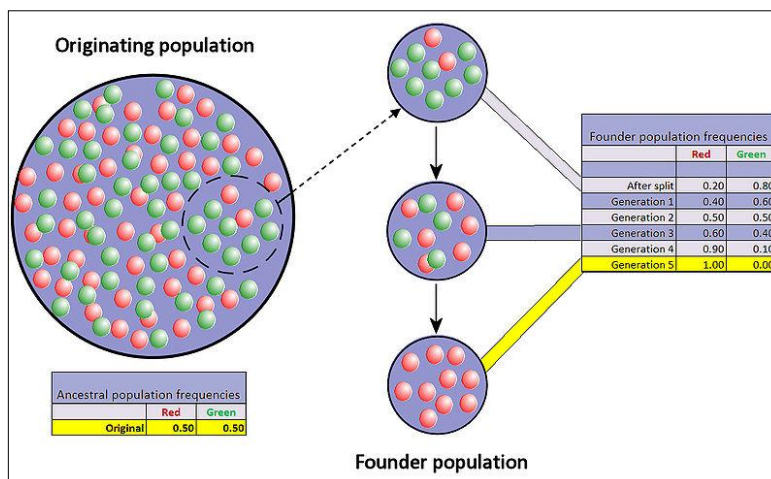
GENETIC DRIFT



very profound effect of genetic drift in small populations – **founder effect**, **bottleneck**
 inextricable link between genetic drift and population size – **the effective population size**

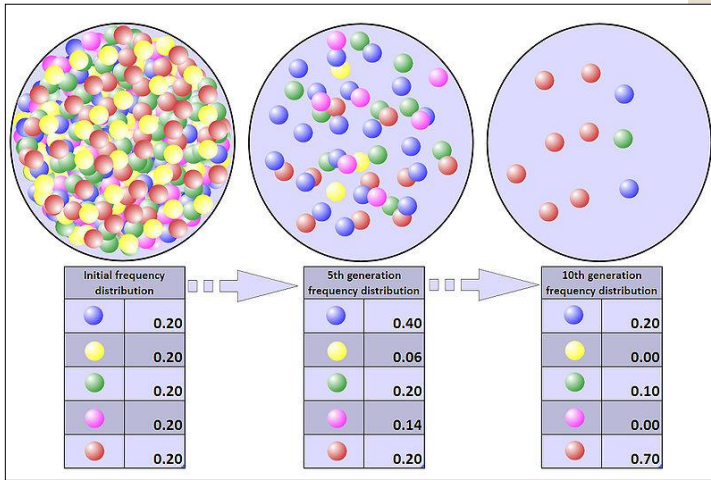
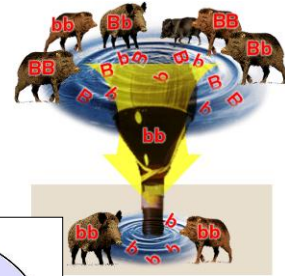


Founder effect

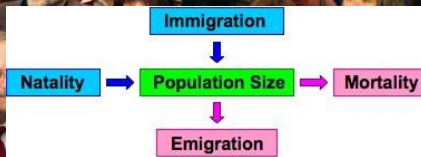




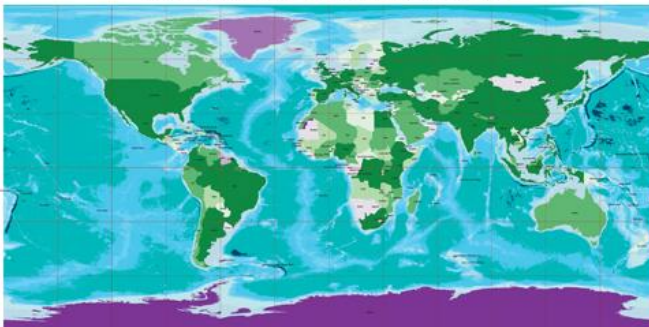
Bottleneck



N_e – effective population size



WORLD POPULATION MAP



N_e – effective population size

vs. N_c – census population size (may be estimated from N_e)
 – see Luikart *et al.* 2010 *Conserv Genet*)

all else being equal, LARGE pops are MORE LIKELY to survive than small pops

N_e – reflects the rate at which genetic diversity will be lost following genetic drift (this rate is inversely proportional to a population's N_e)

single-sample estimators of N_e – level of LD due to drift
 double sample estimators of N_e – temporal changes in allele frequencies due to genetic drift

OVERVIEW

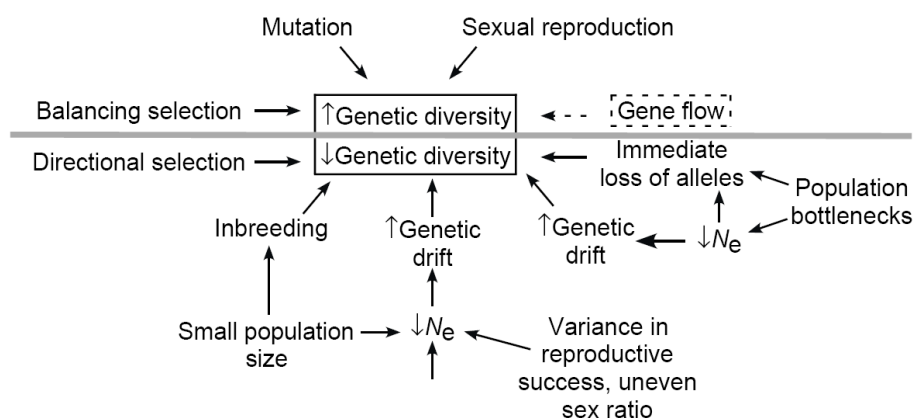


Figure 3.16 An overview of some of the main factors that influence levels of genetic diversity within populations.

Freeland *et al.* 2011