



CEITEC

Central European Institute of Technology
BRNO | CZECH REPUBLIC

Image analysis II

C9940 3-Dimensional Transmission Electron Microscopy
S1007 Doing structural biology with the electron microscope

April 4, 2016



EUROPEAN UNION
EUROPEAN REGIONAL DEVELOPMENT FUND
INVESTING IN YOUR FUTURE



OP Research and
Development for Innovation



Outline

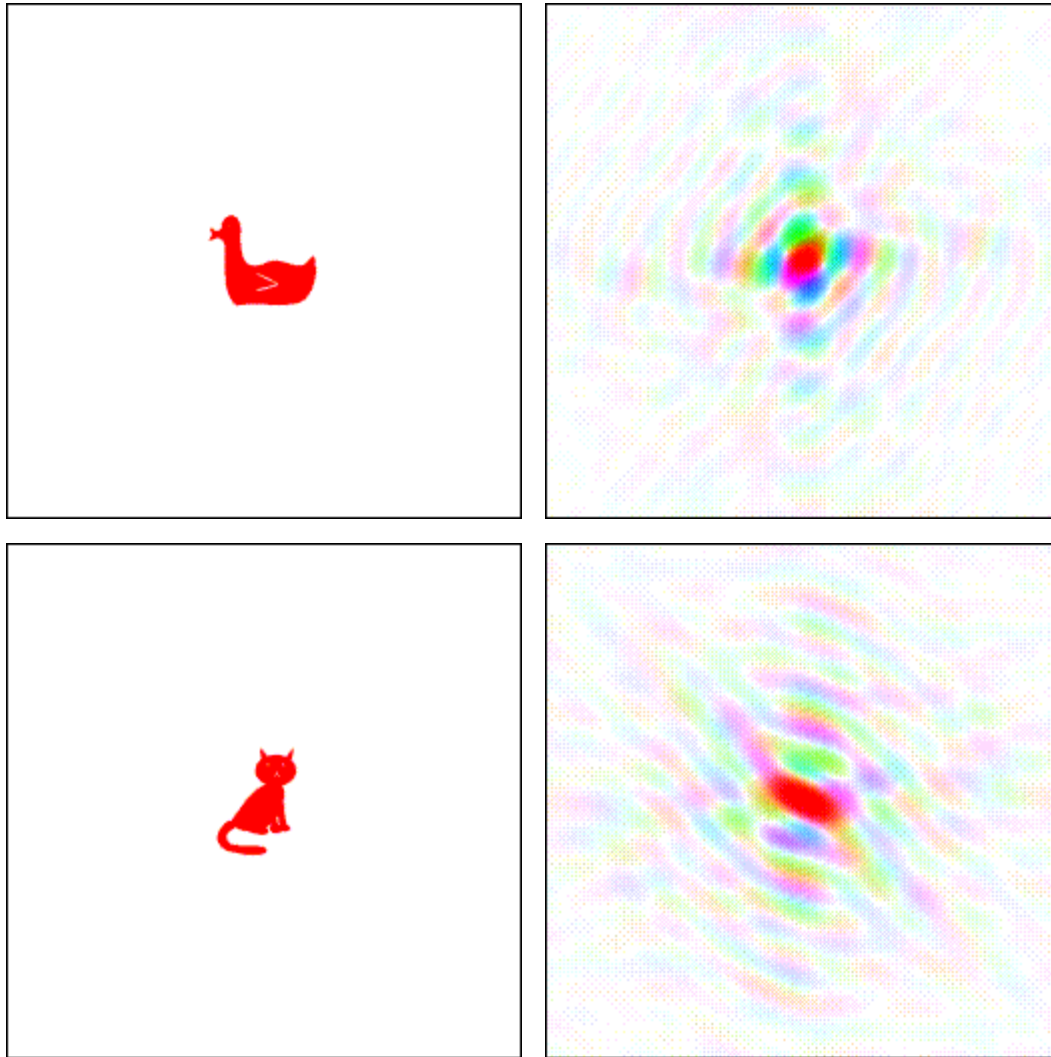
Image analysis II

- Fourier transforms revisited
 - Ducks and other animals
- Analogy to the Ewald sphere
- Aliasing
- Alignment
- Interpolation
- Multivariate data analysis

Outline

Image analysis II

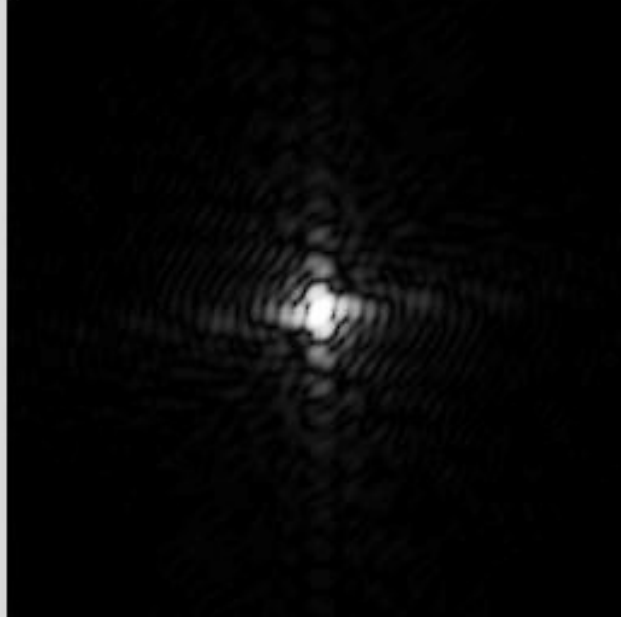
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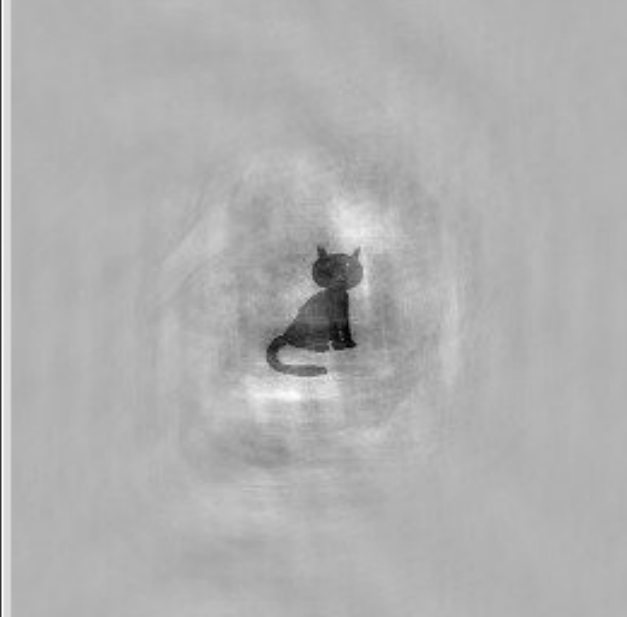
<http://www.yybl.york.ac.uk/~cowtan/fourier/magic.html>



picduck.spi



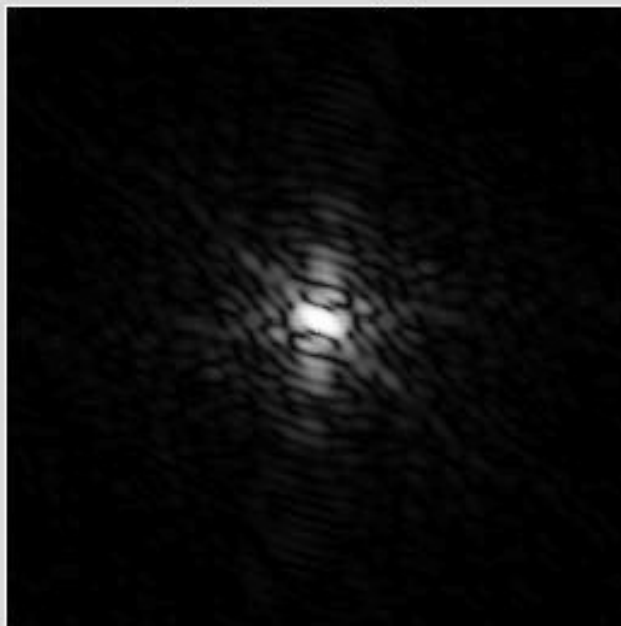
pws-picduck.spi



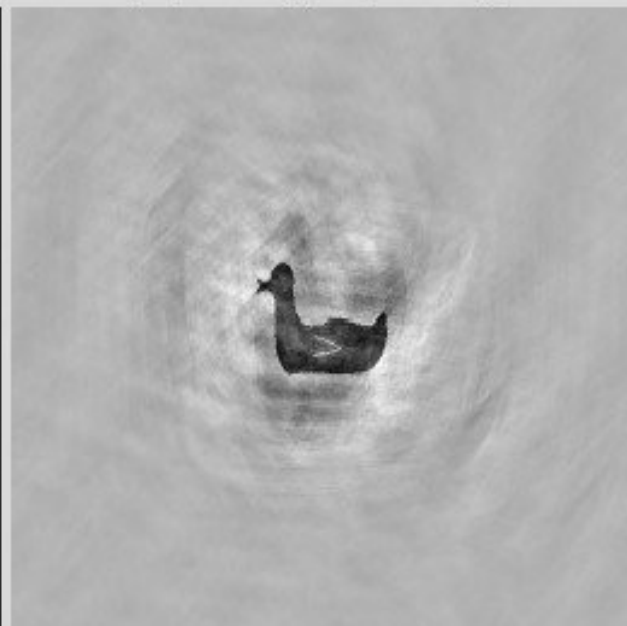
amp-picduck pha-piccat.spi



piccat.spi



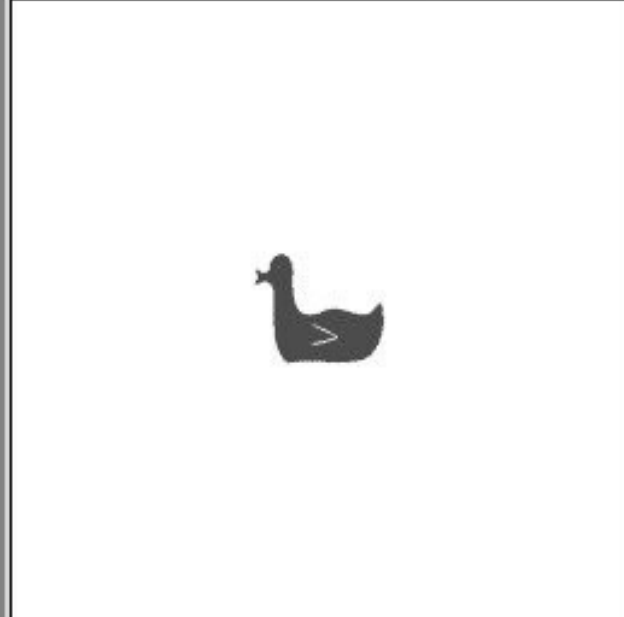
pws-piccat.spi



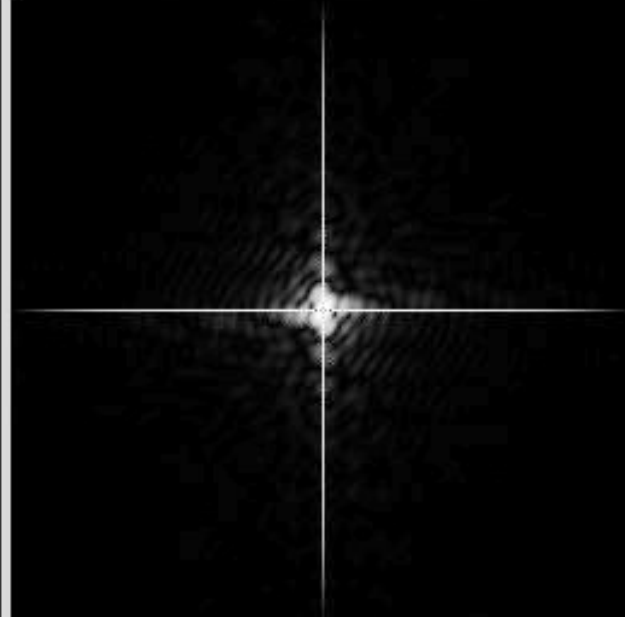
amp-piccat pha-picduck.spi

A quiz:

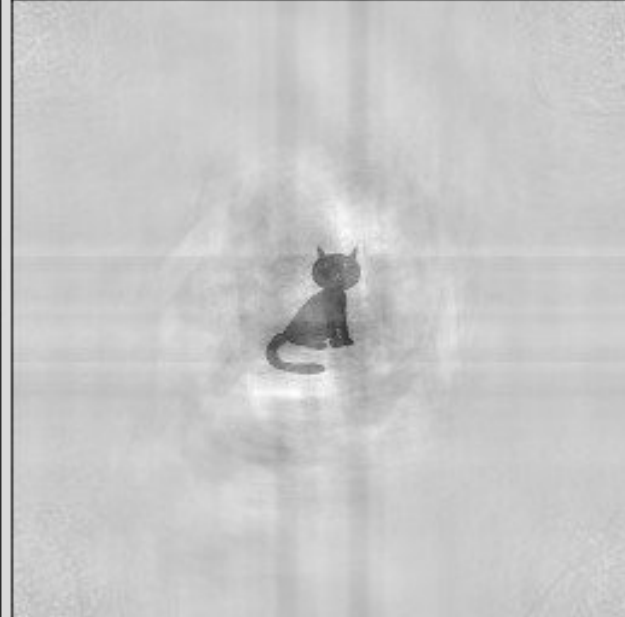
A problem I had with my first attempt



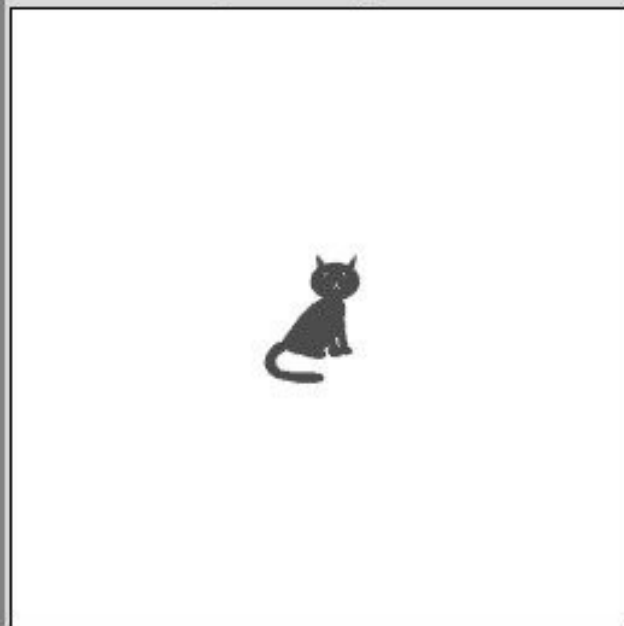
picduck.spi



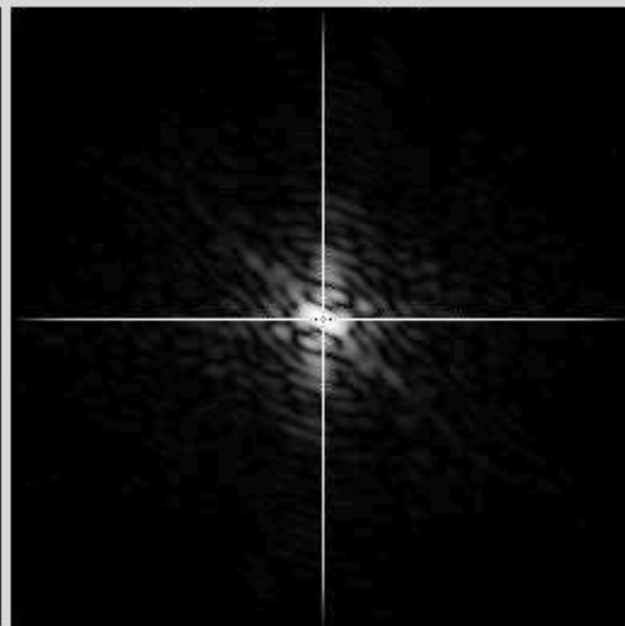
pws-picduck.spi



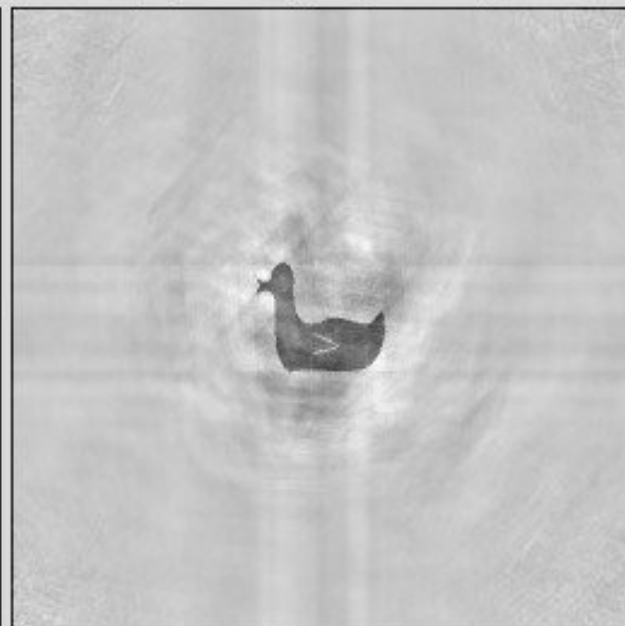
amp-picduck pha-piccat.spi



piccat.spi



pws-piccat.spi

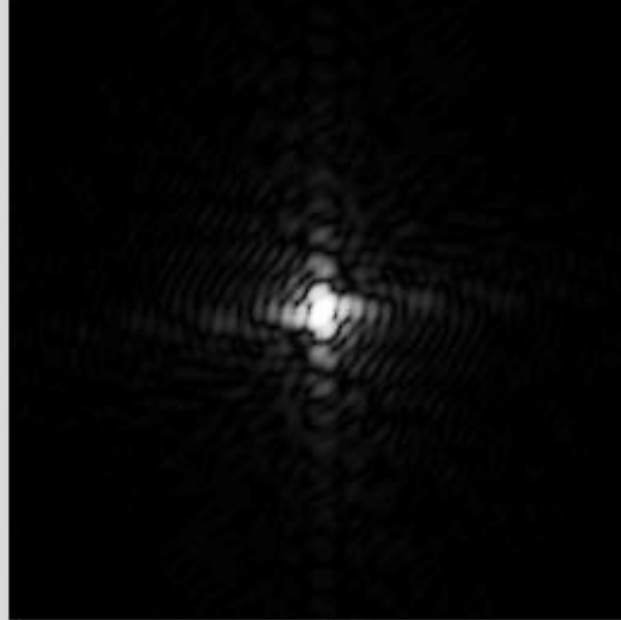


amp-piccat pha-picduck.spi

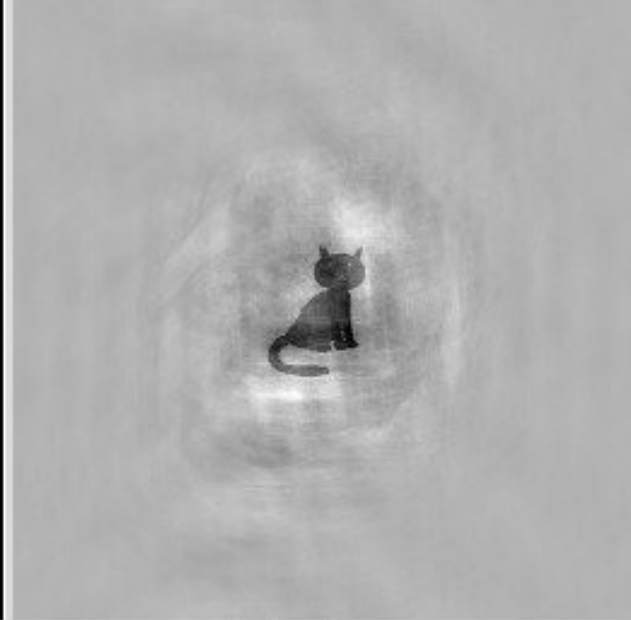
QUESTION: Where did that cross come from? 



picduck.spi



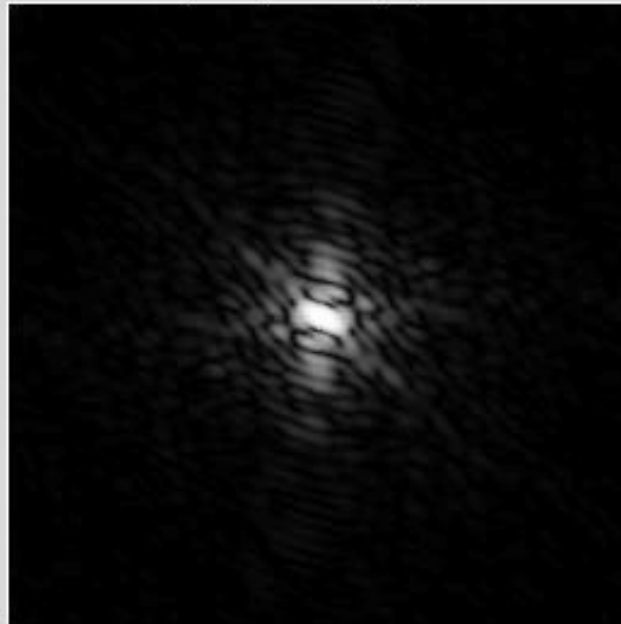
pws-picduck.spi



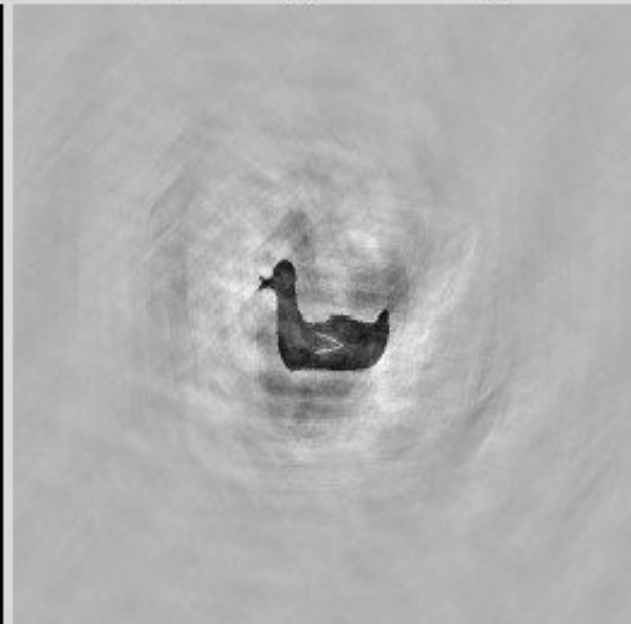
amp-picduck pha-piccat.spi



piccat.spi



pws-piccat.spi



amp-piccat pha-picduck.spi

Apodization: Smoothly bring the signal to zero near an edge

Outline

Image analysis II

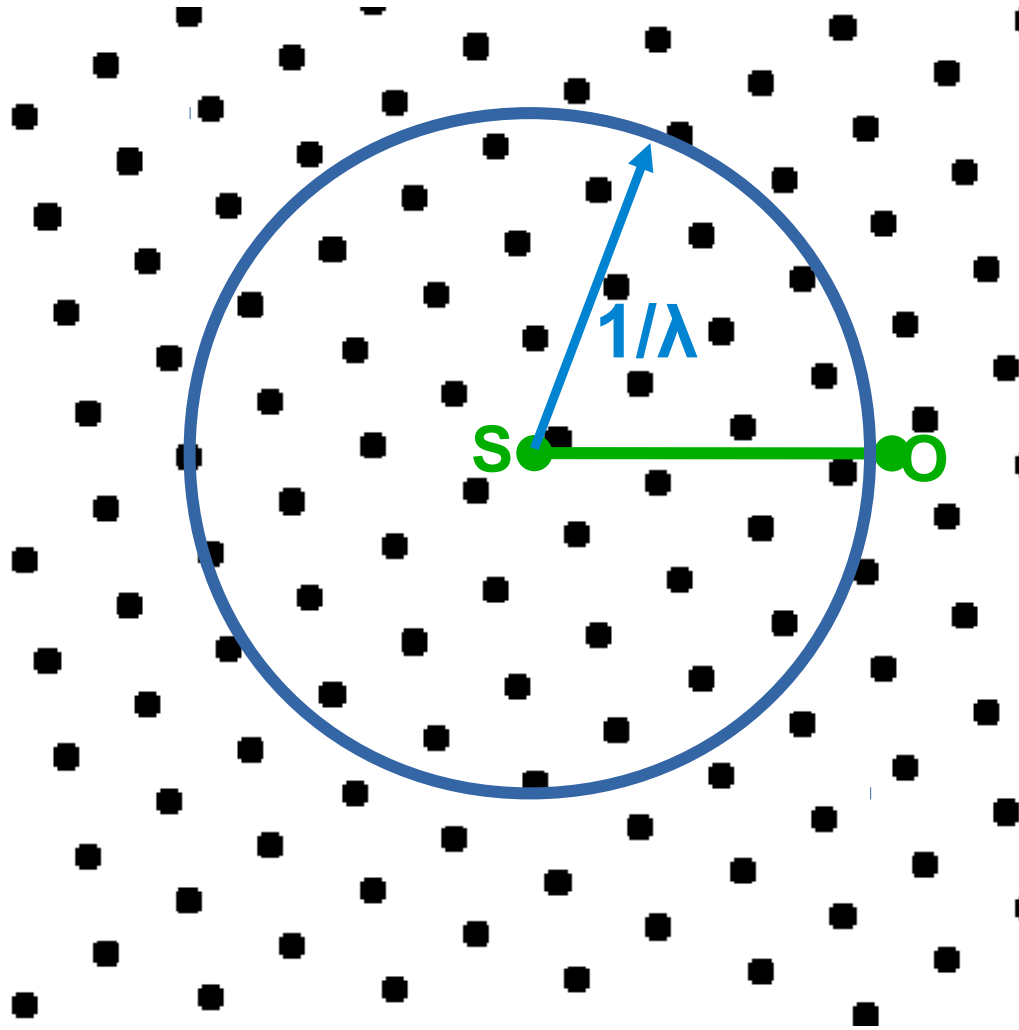
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Why are electrons useful?

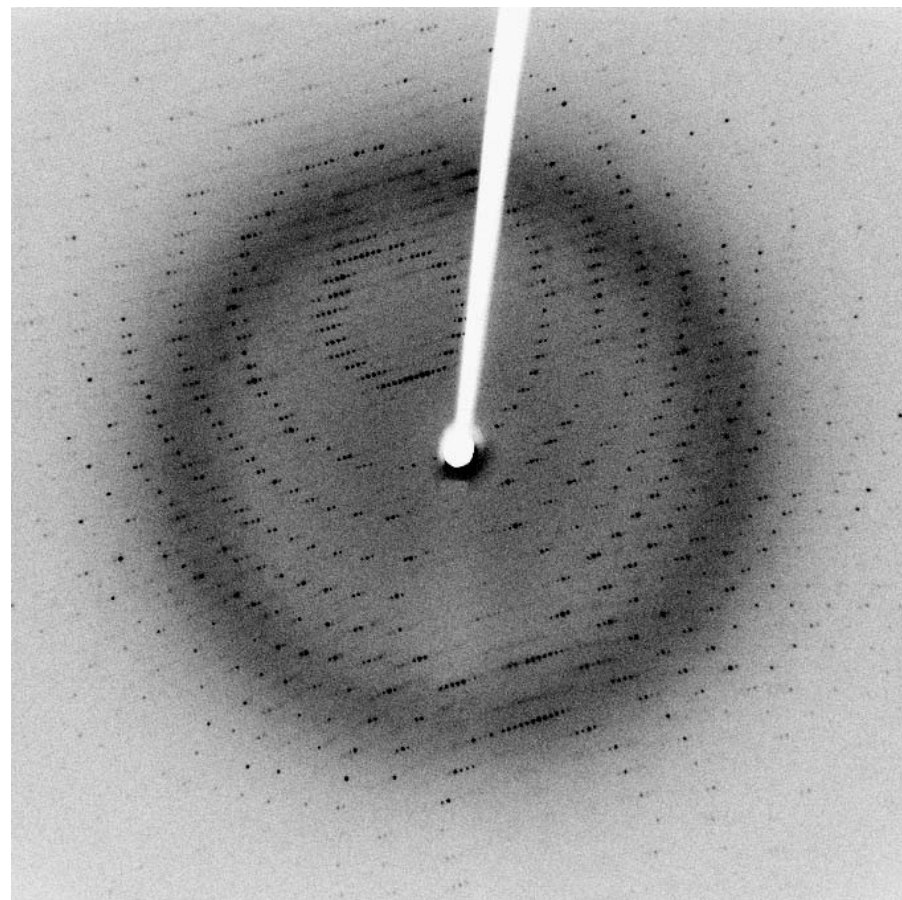
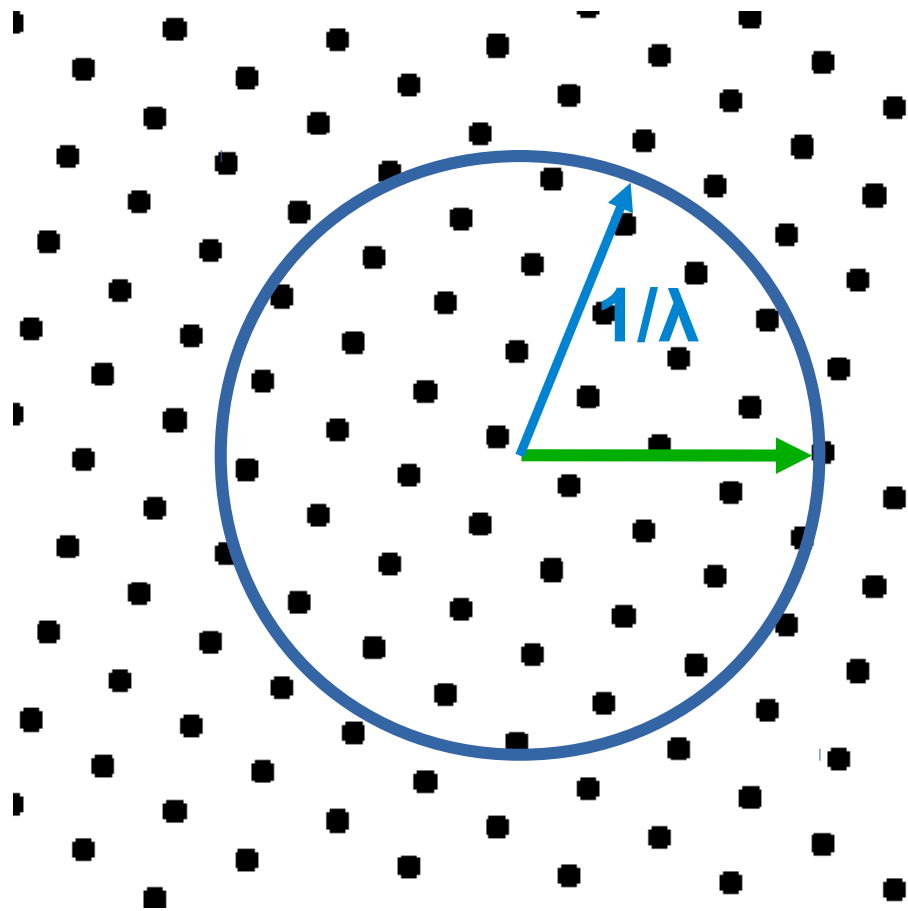
Wavelengths of various radiation types

- ◆ Visible light: $>380\text{nm}$
- ◆ X-rays (copper K): 0.154nm (1.54\AA)
- ◆ Electrons (300kV): 0.002nm (0.02\AA)

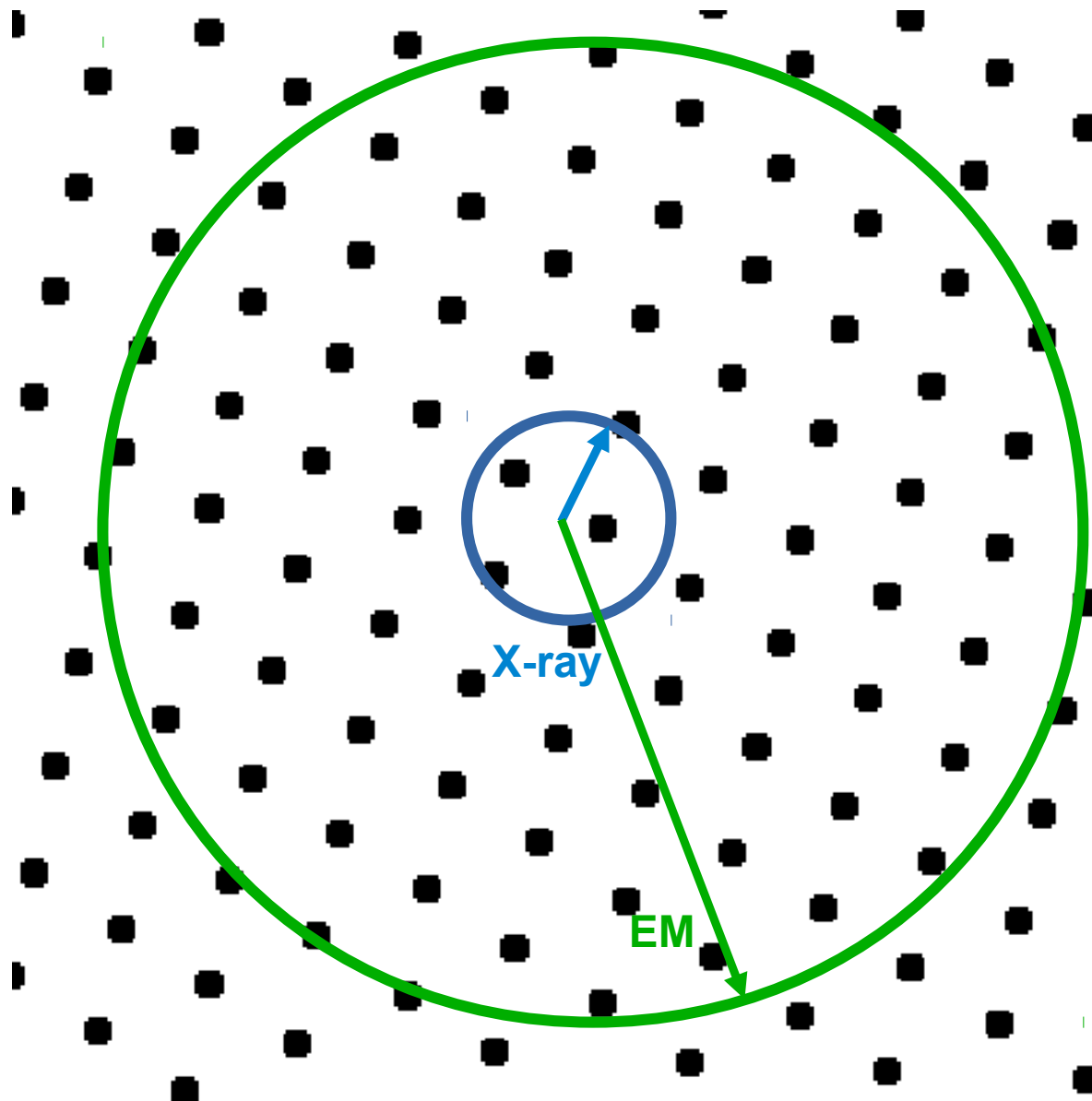
How wavelength limits resolution



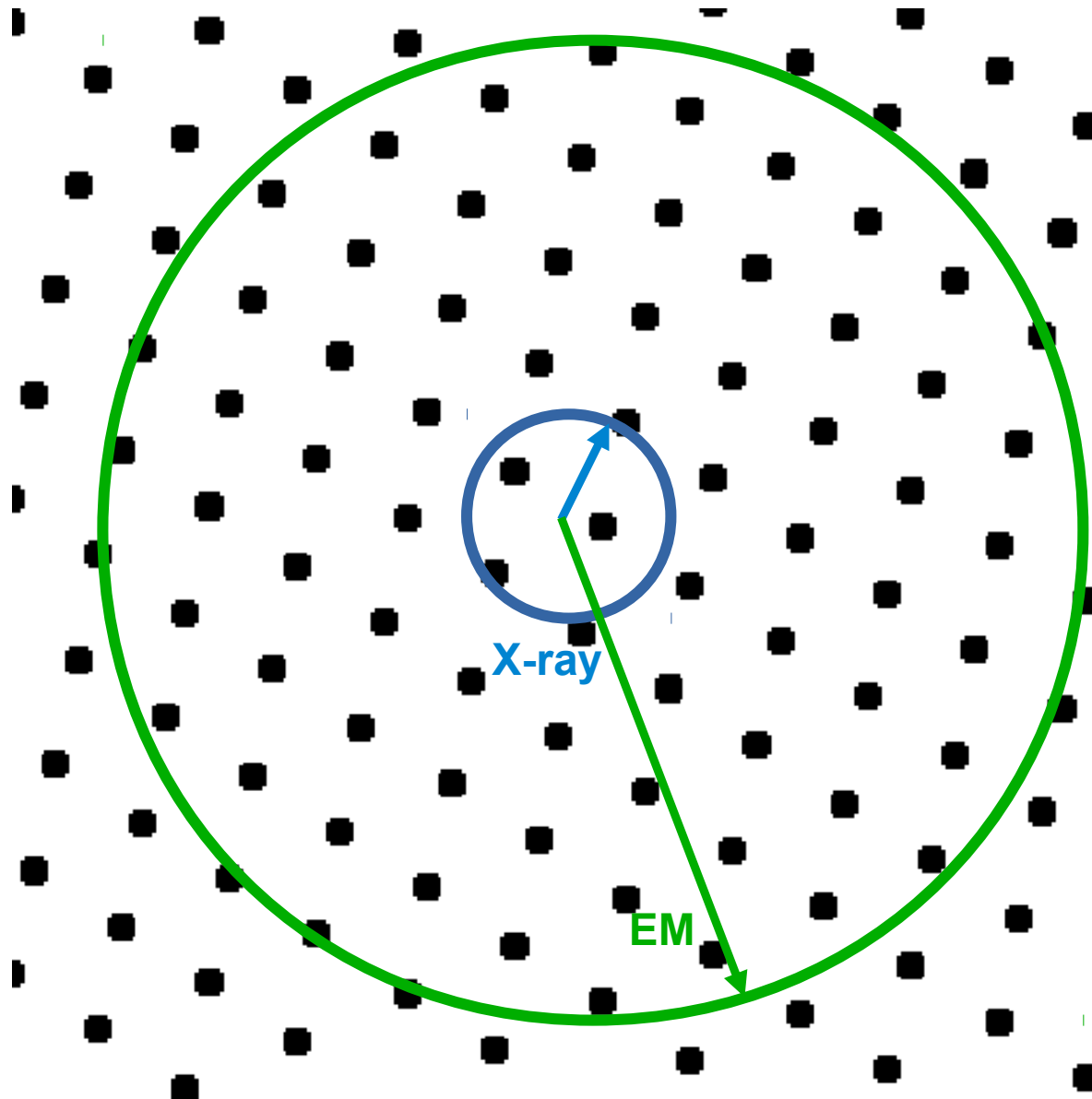
S: specimen origin
O: diffraction origin



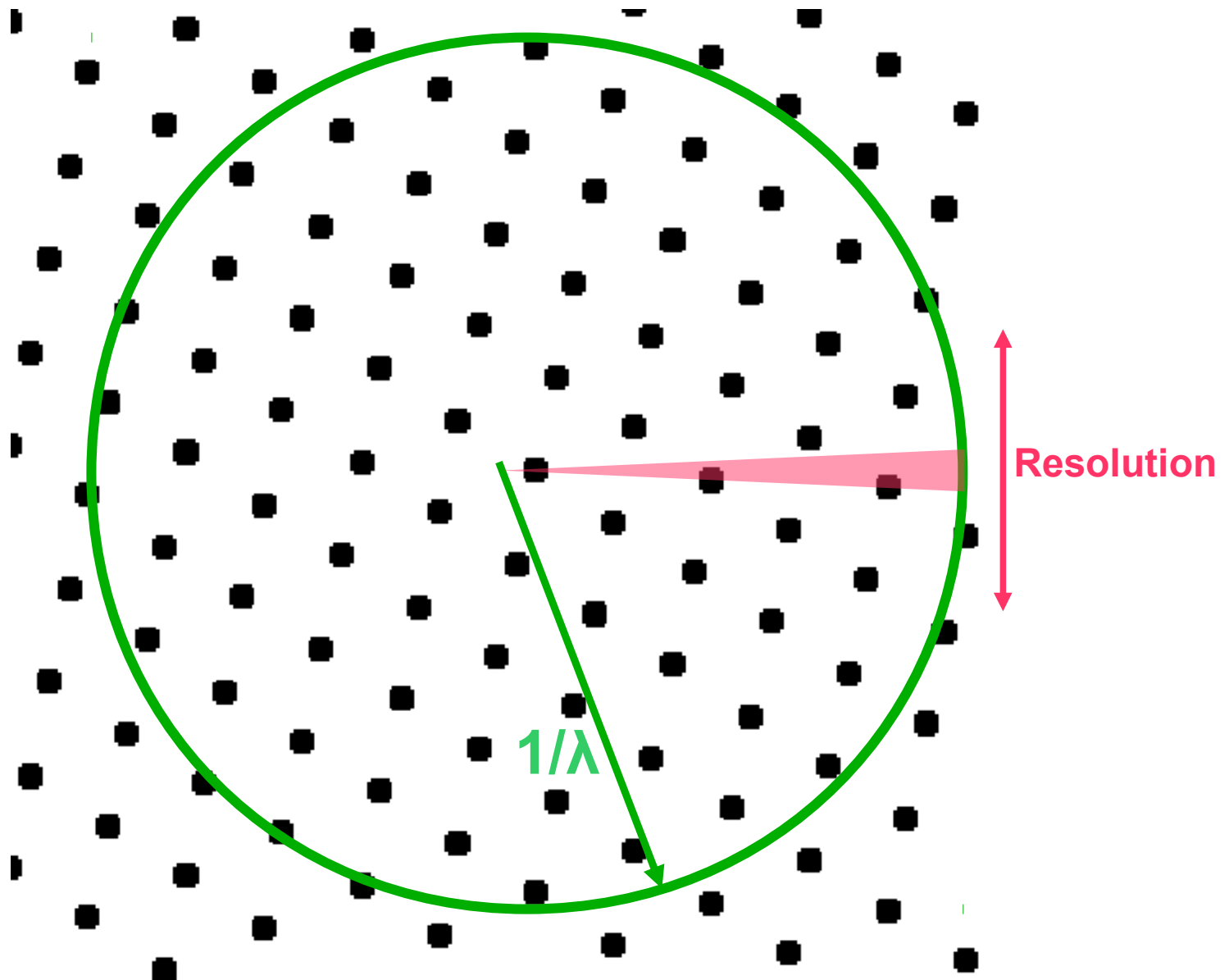
<http://en.wikipedia.org>



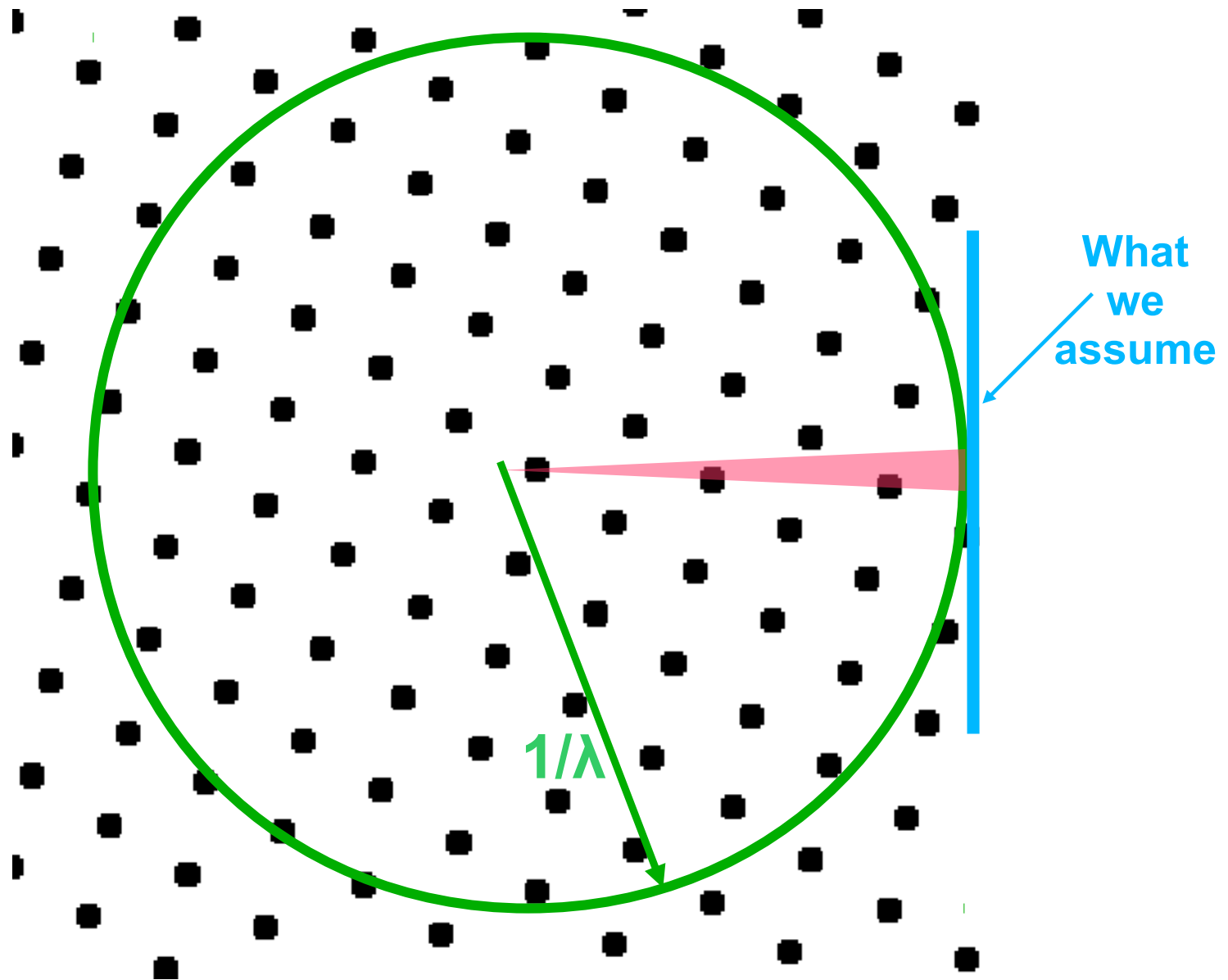
NOTE: Not to scale.
EM wavelength is ~ 80 smaller,
and therefore $1/\lambda$ would be 80X bigger.



NOTE 2: For practical purposes, the radius of the Ewald sphere is so large that we ignore its curvature.

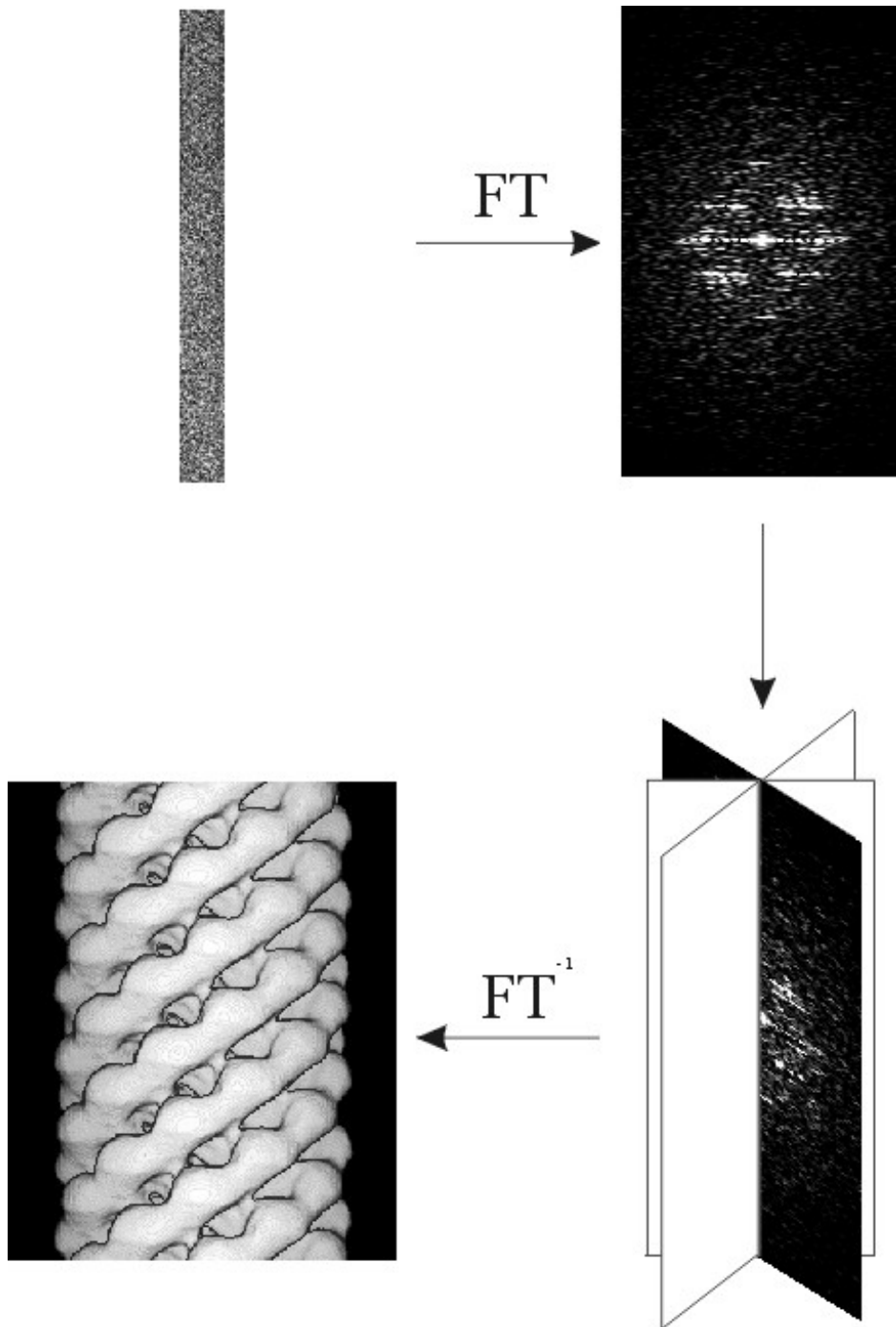


NOTE 3: Electron lenses are terrible, and biological samples are fragile, so in practice we'll see on a tiny fraction of the data we could theoretically get.

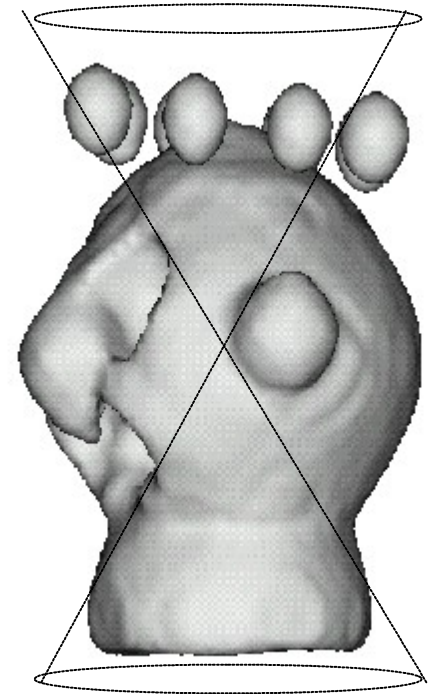
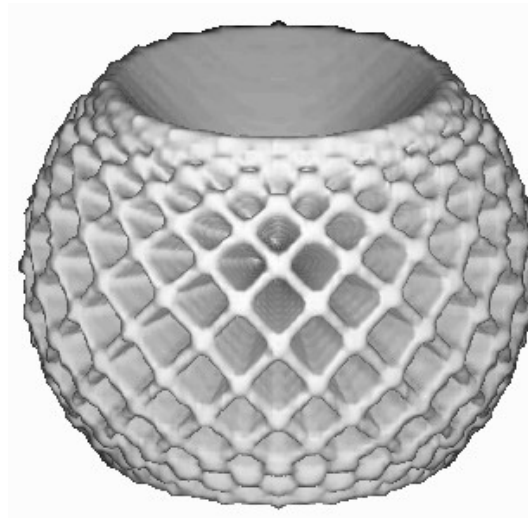
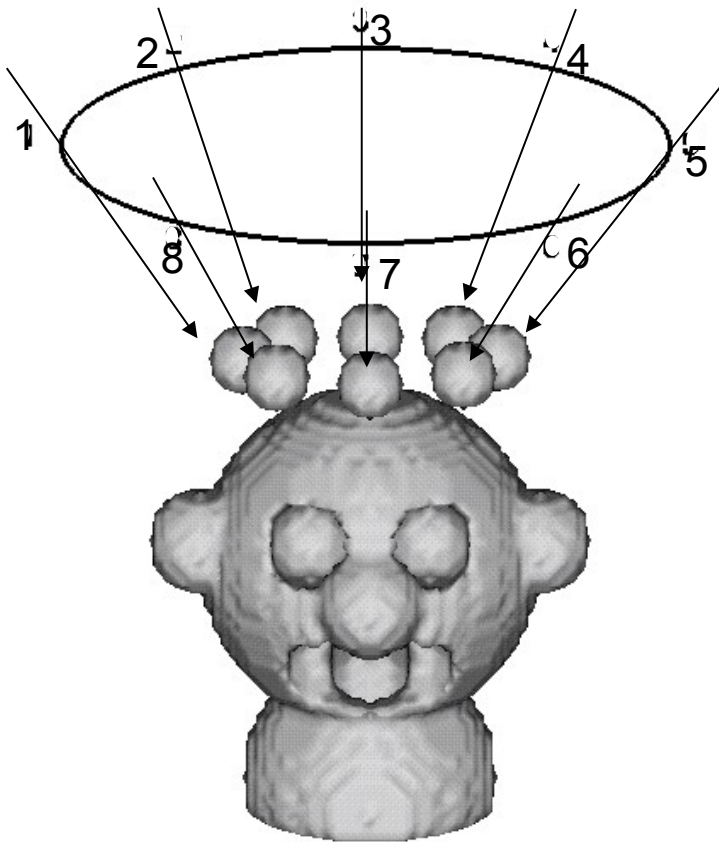


NOTE 4: For more information, see DeRosier (2000) "Correction of high-resolution data for curvature of the Ewald sphere."

Preview: 3D reconstruction



Preview: reconstruction

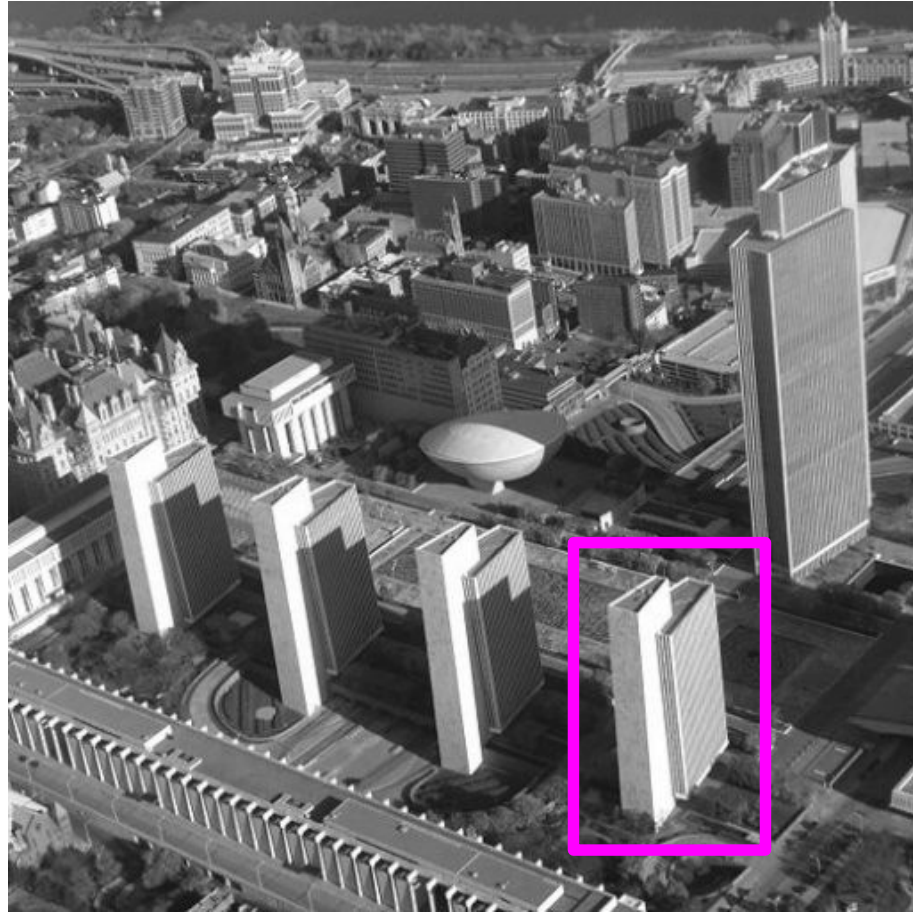


Outline

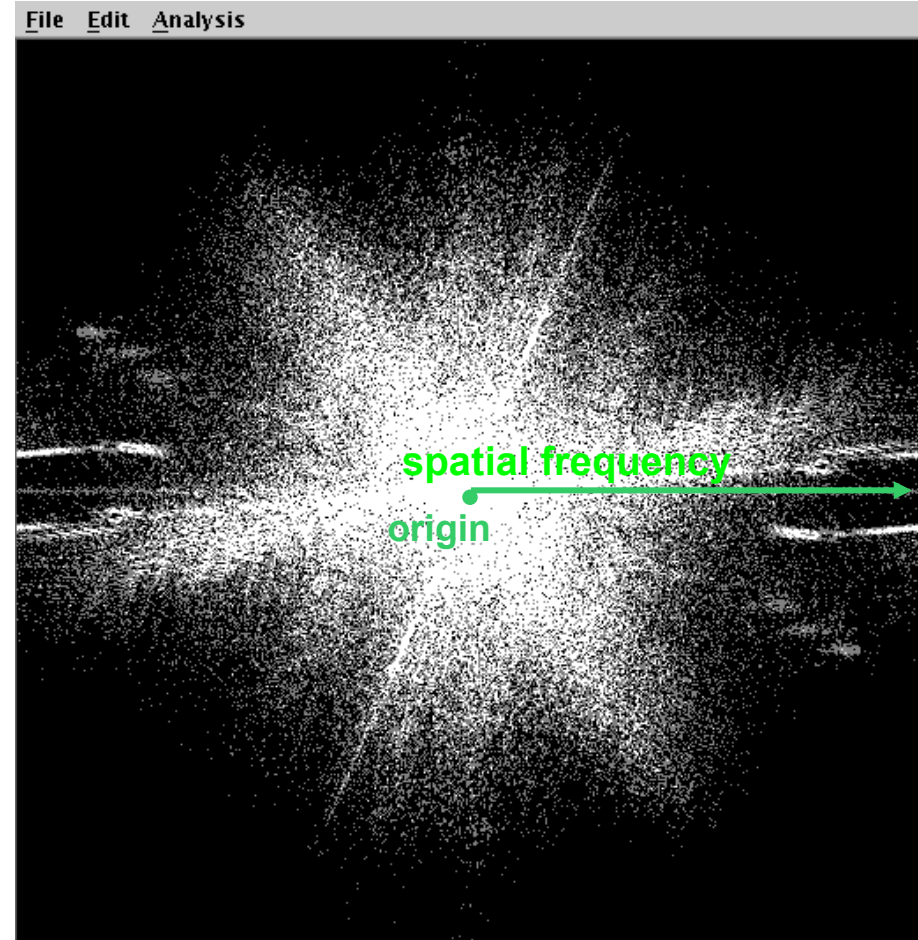
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From last week...



From Wikipedia



This is an example of aliasing.

An example using SPIDER

<https://youtu.be/6LzaPARy3uA?t=51>

Outline

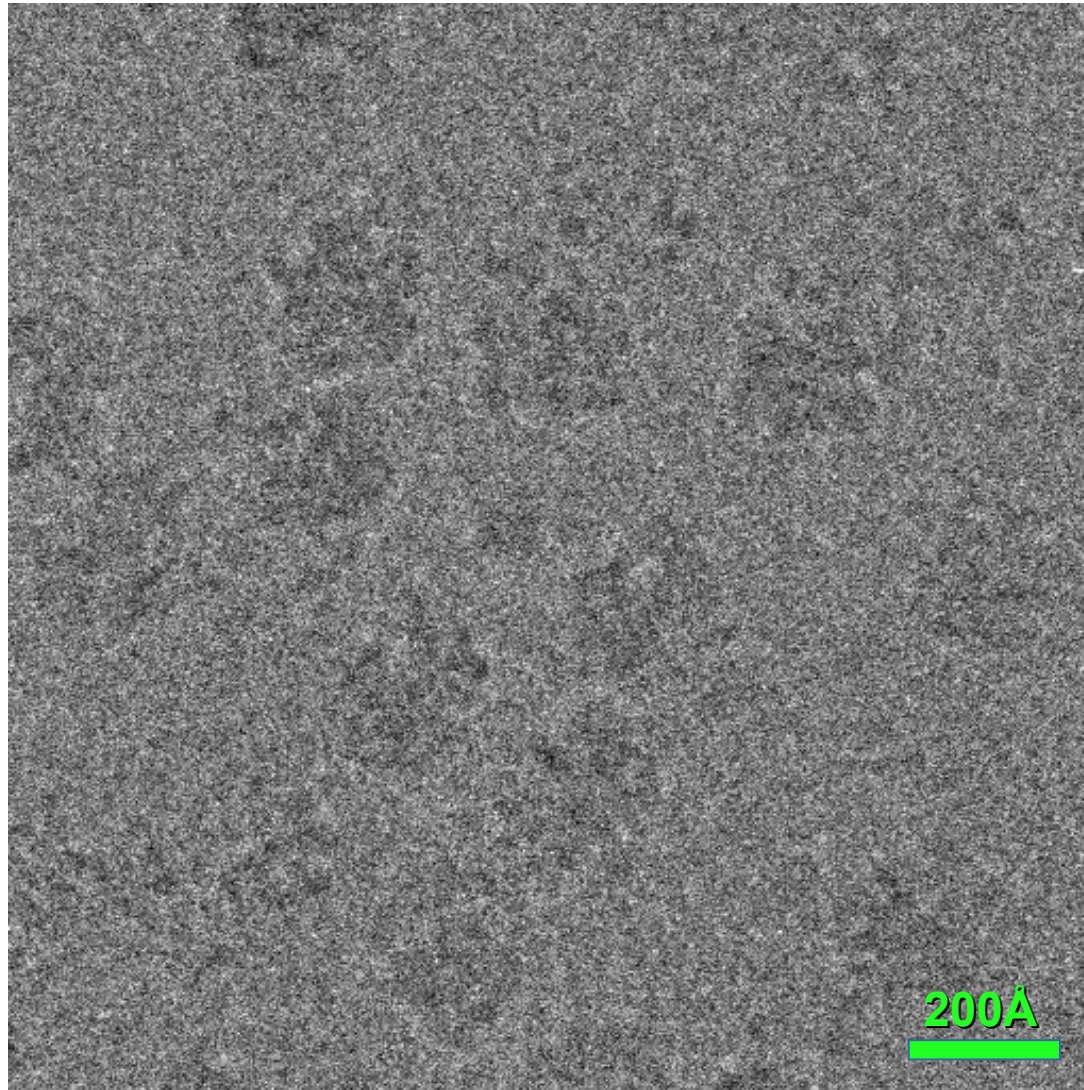
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- **Alignment**
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QUESTION:

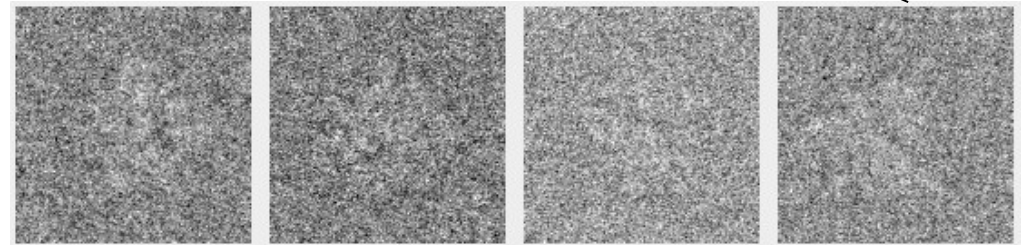
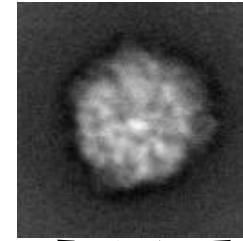
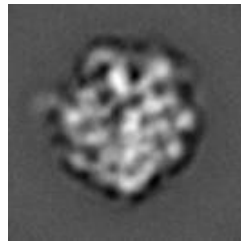
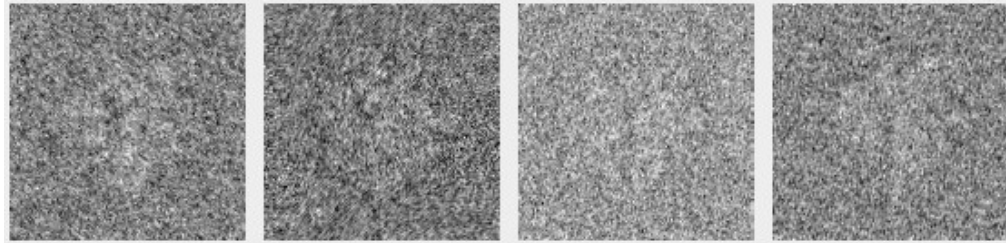
Why do we need to average the signal from many images?

ANSWER: Our signal-to-noise is poor



What happens if we don't align our images?

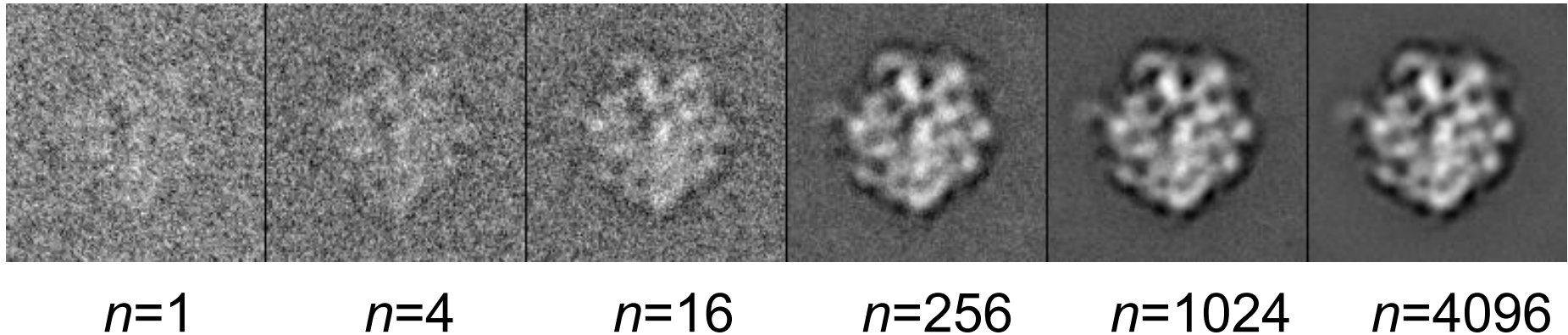
aligned images 1-4 of 4096 total



unaligned images 1-4 of 4096 total

This is a simple 2D case, but the effects are analogous in 3D.

What happens as we include more particles?



Signal-to-noise ratio increases with \sqrt{n}

(P)review of 3D reconstruction: The parameters required

Two translational:

✓ Δx

✓ Δy

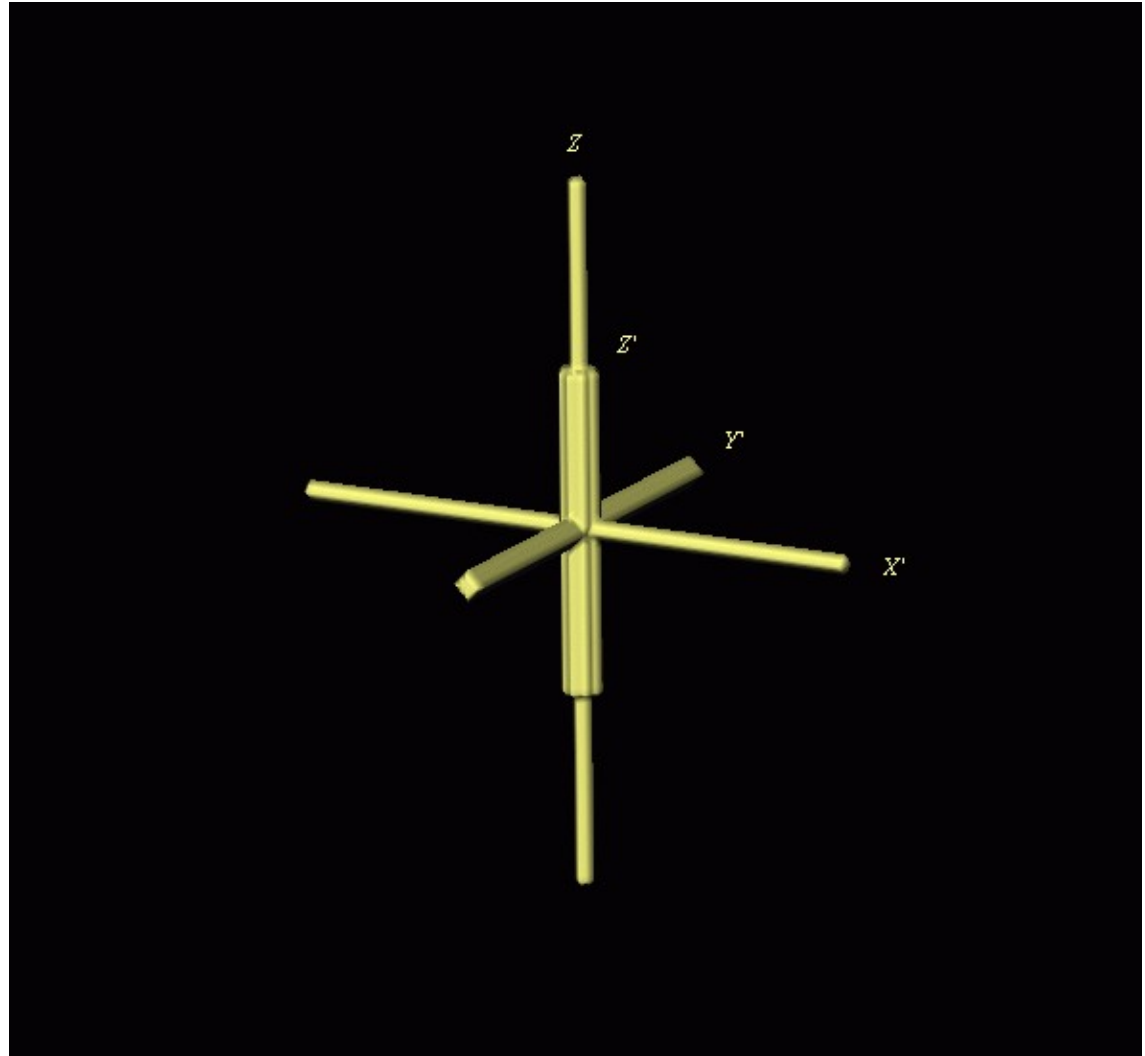
Three orientational
(Euler angles):

✓ phi (about z axis)

✓ theta (about y)

✓ ψ (about new z)

These are determined in 2D.
We'll concentrate on these 1st.



*How do find the relative translations
between two images?*

Cross-correlation

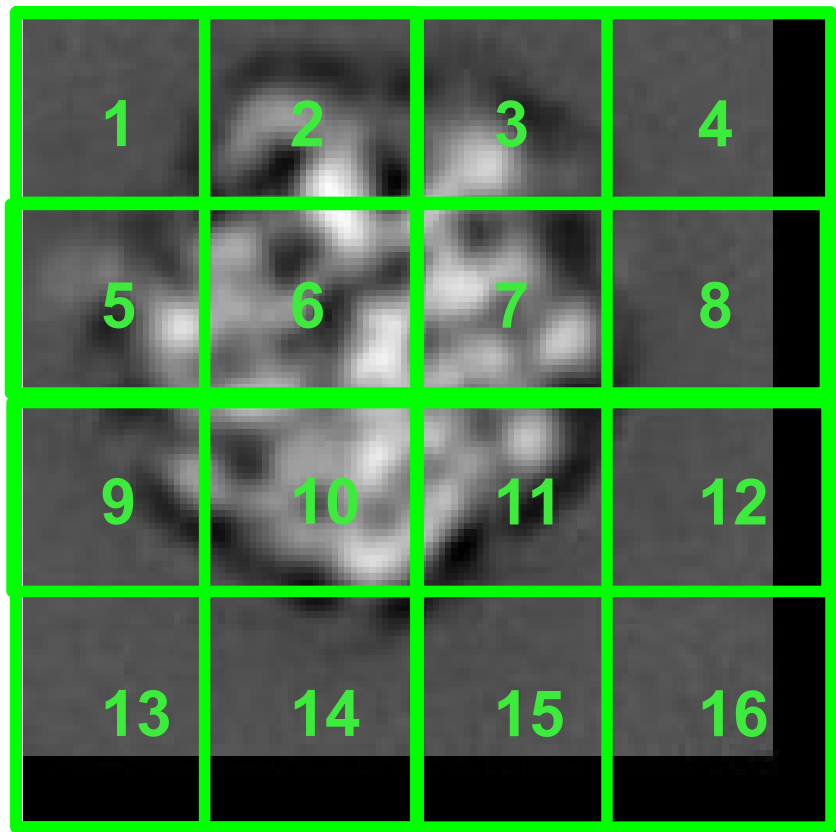


Image f

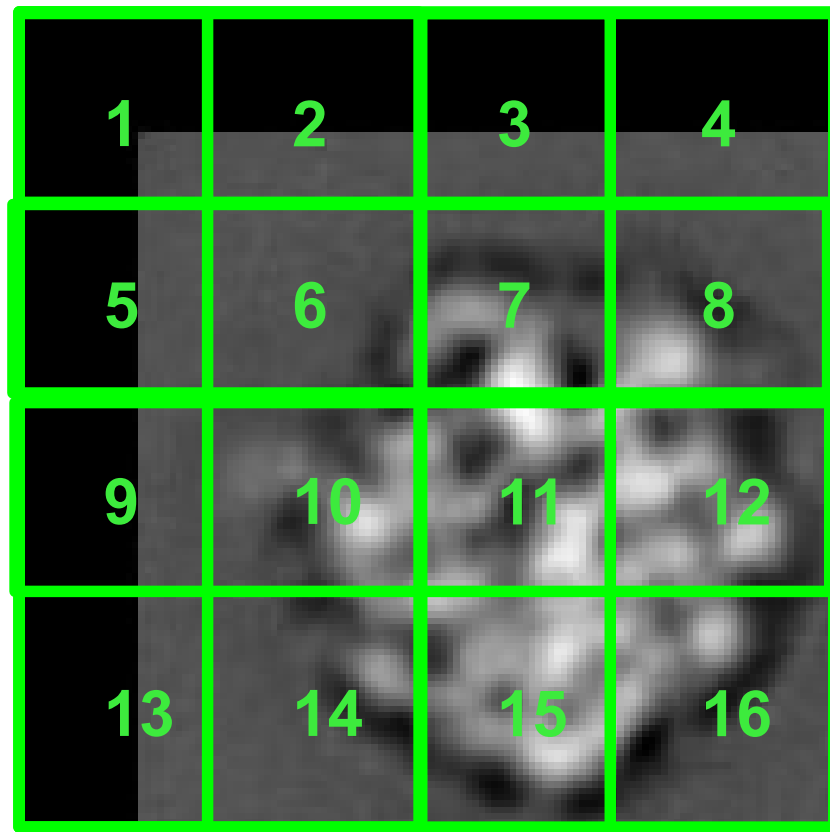


Image g

Cross-correlation coefficient:

$$\frac{\sum_{N=1}^{16} f(\vec{x}) g(\vec{x})}{\sigma_f \sigma_g}$$

constant
"normalization"

Cross-correlation

1	2	3	4
5	6	7	8
9	10	11	12
13	14	15	16

Image f

1	2	3	4
5	6	7	8
9	10	11	12
13	14	15	16

Image g

$$\begin{aligned} \text{Unnormalized CCC} = & f_1g_1 + f_2g_2 + f_3g_3 + f_4g_4 + f_5g_5 + f_6g_6 + f_7g_7 + f_8g_8 \\ & + f_9g_9 + f_{10}g_{10} + f_{11}g_{11} + f_{12}g_{12} + f_{13}g_{13} + f_{14}g_{14} + f_{15}g_{15} + f_{16}g_{16} \end{aligned}$$

Cross-correlation

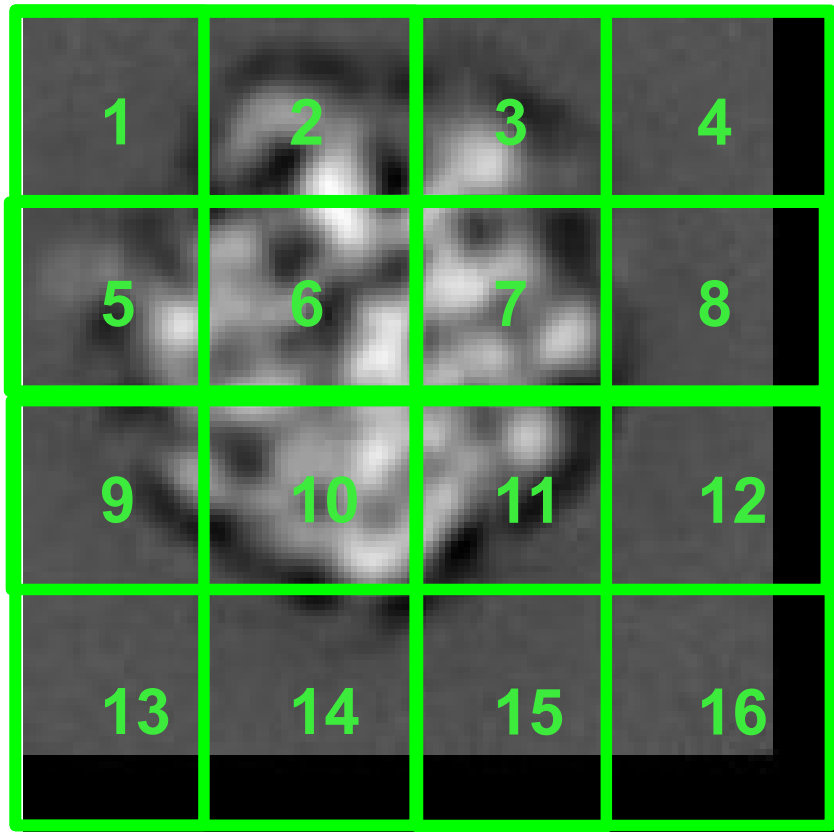


Image f

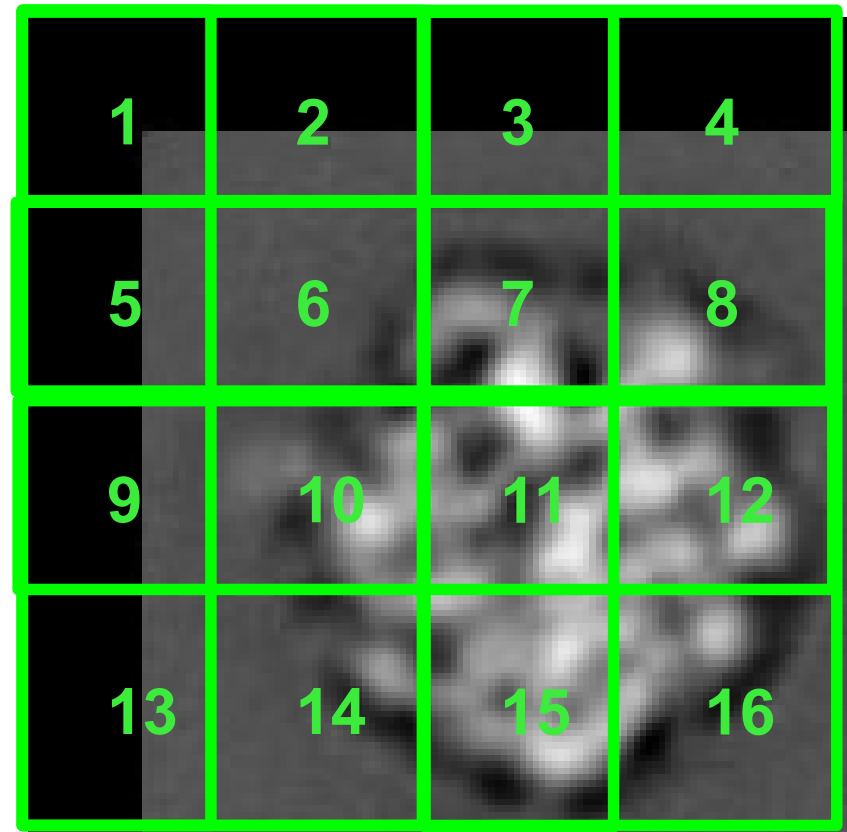


Image g

$$\begin{aligned} \text{Unnormalized CCC} = & f_1g_1 + f_2g_2 + f_3g_3 + f_4g_4 + f_5g_5 + f_6g_6 + f_7g_7 + f_8g_8 \\ & + f_9g_9 + f_{10}g_{10} + f_{11}g_{11} + f_{12}g_{12} + f_{13}g_{13} + f_{14}g_{14} + f_{15}g_{15} + f_{16}g_{16} \end{aligned}$$

Cross-correlation coefficient

Cross-correlation coefficient:
$$\frac{\sum_{N=1}^{16} f(\vec{x}) g(\vec{x})}{\sigma_f \sigma_g}$$

If the alignment is perfect, the correlation value will be 1.

What if the correlation isn't perfect?

Cross-correlation

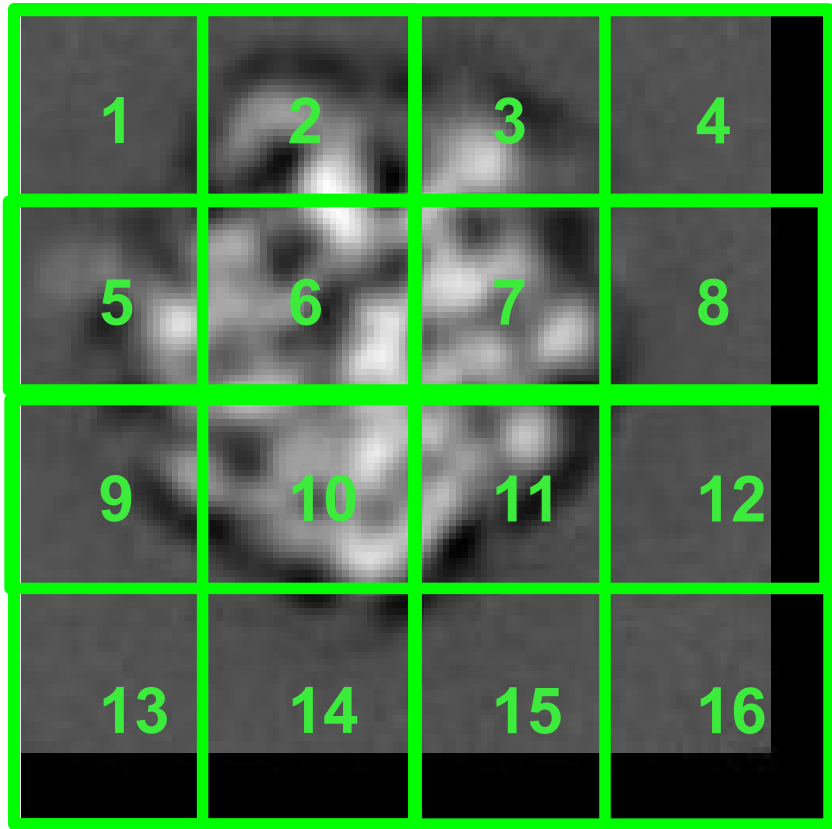


Image f

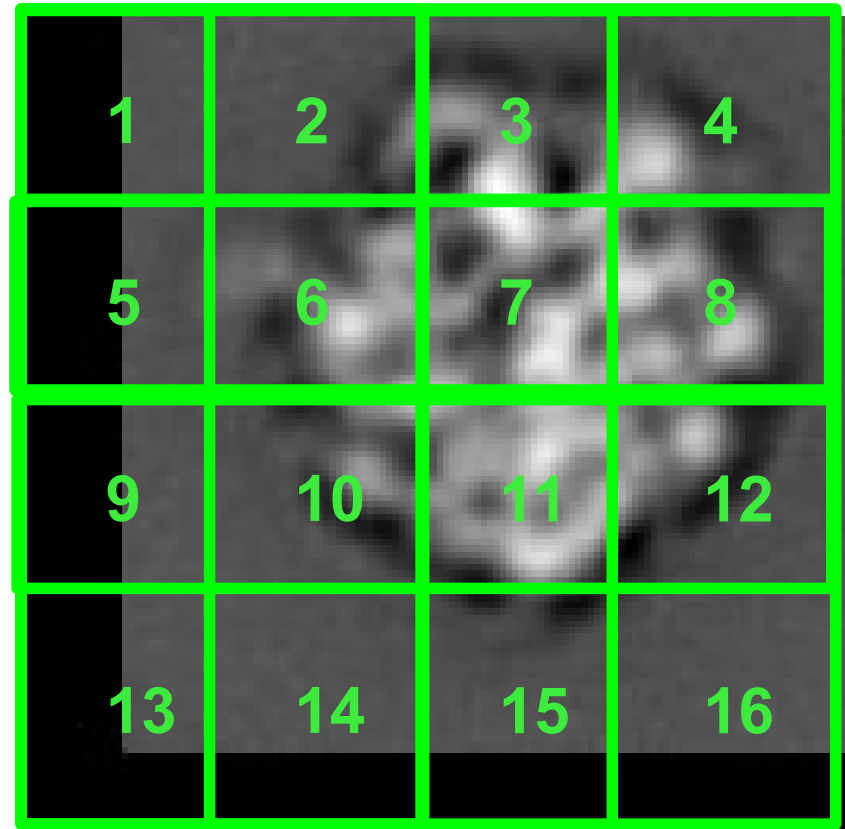
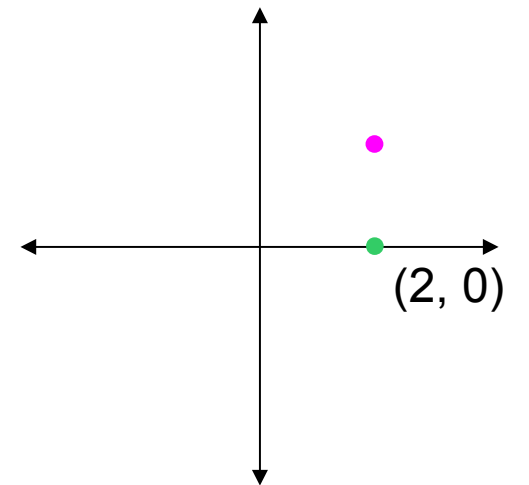
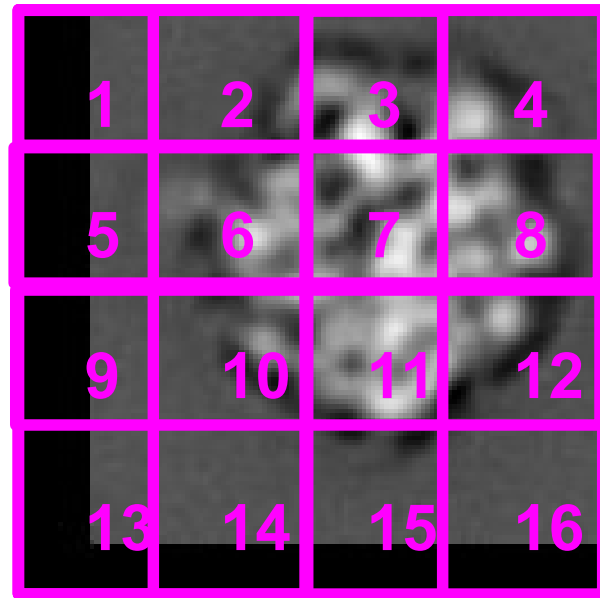
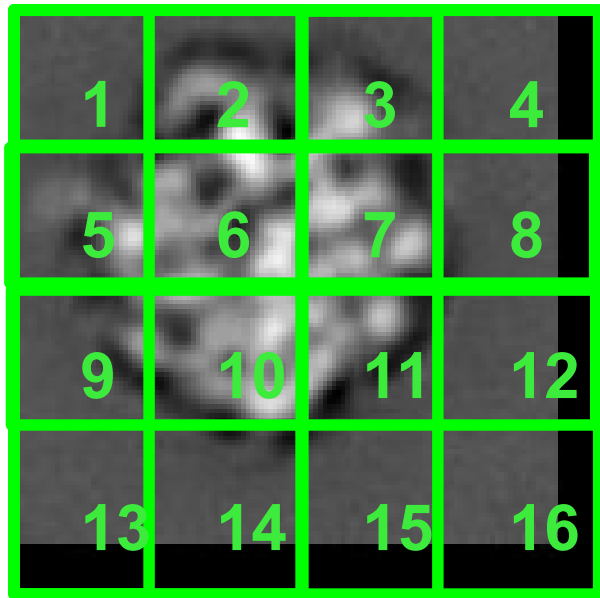
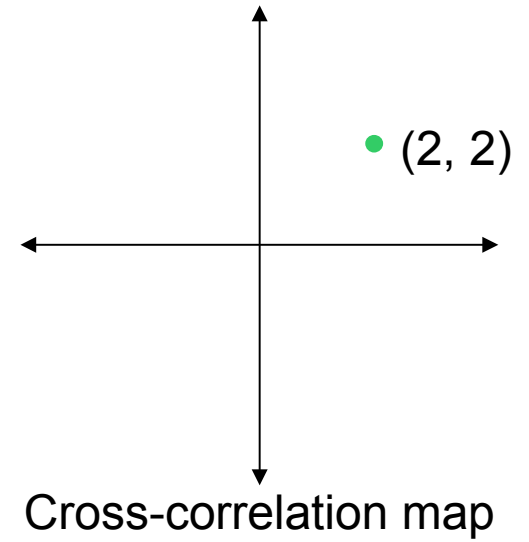
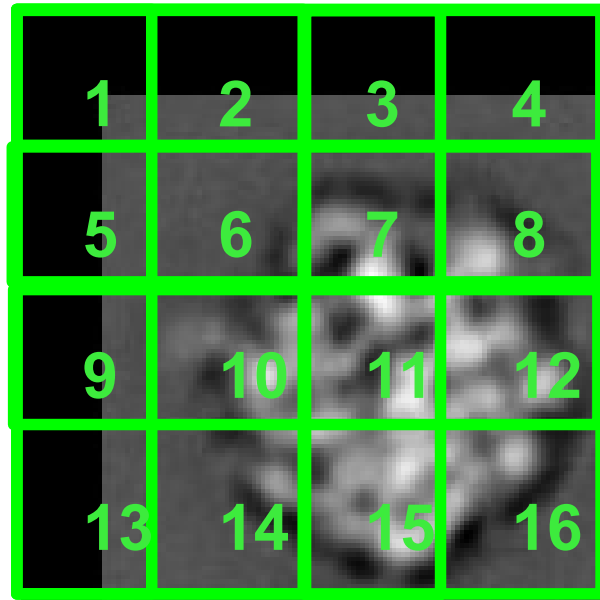
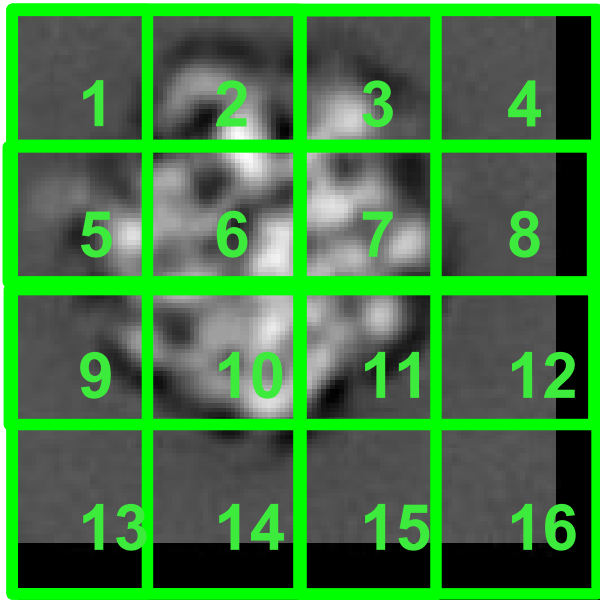


Image g

What if the correlation isn't perfect?

ANSWER: You try other shifts (perhaps all).



We would need to repeat this for all combinations of shifts.

Cross-correlation function (CCF)

Brute-force translational search is CPU-intensive

BUT

Fourier transforms can help us.

	Real space	$f(x)$	$g(x)$
Some notation:			
	Fourier space	$F(X)$	$G(X)$

Cross-correlation function (CCF)

Brute-force translational search is CPU-intensive

BUT

Fourier transforms can help us.

Complex conjugate:

If a Fourier coefficient $F(X)$ has the form: $a + bi$

The complex conjugate $F^*(X)$ has the form: $a - bi$

$$F^*(X) G(X) = \text{F.T.}(\text{CCF})$$

This gives us a map of all possible shifts.

Cross-correlation function (CCF)

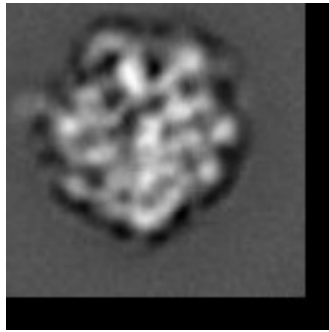


Image $f(x)$

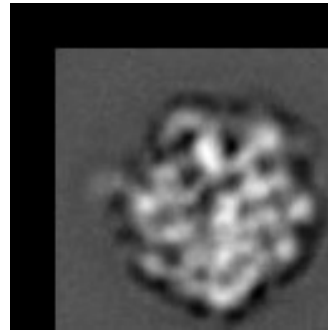
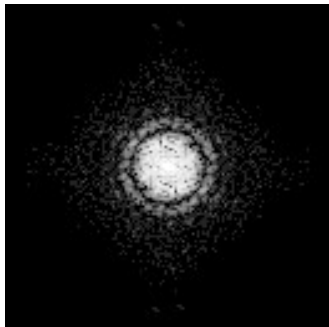
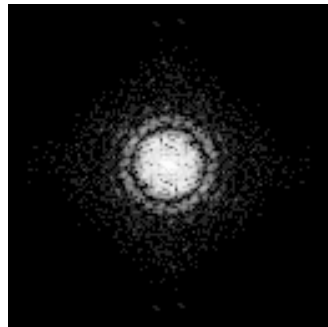


Image $g(x)$



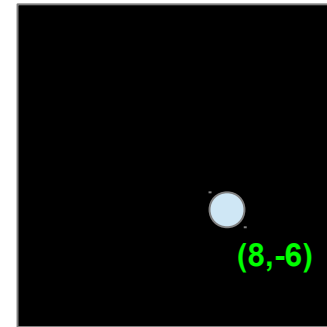
F.T. $F^*(X)$
(complex conjugate)

x



F.T. $G(X)$

=



F.T. (CCF)

The position of the peak gives us the shifts that give the best match, e.g., (8,-6).

*Well, that was an easy case.
We only needed to do translational alignment.
What about orientation alignment?*

Orientation alignment

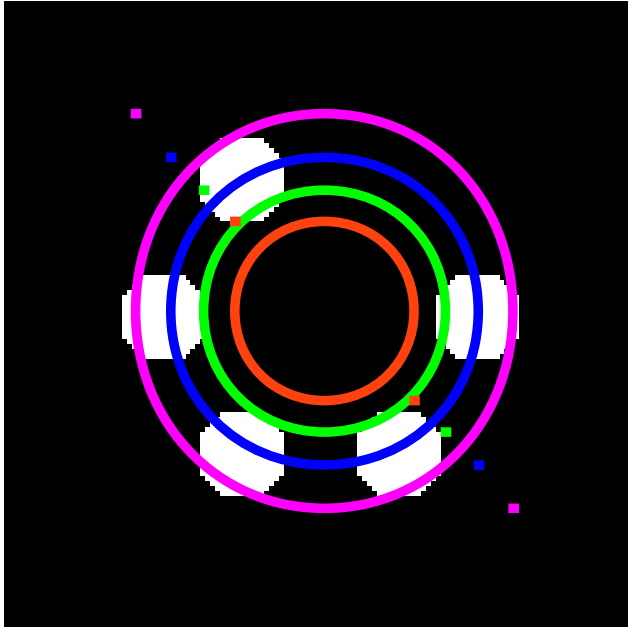


Image 1

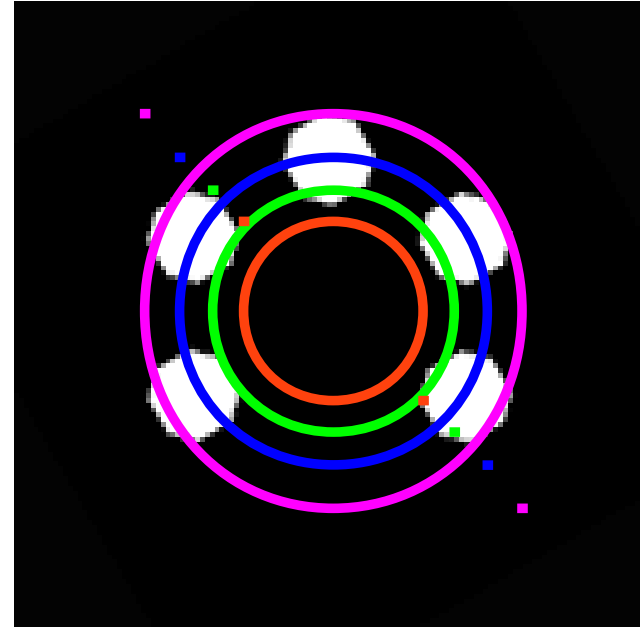
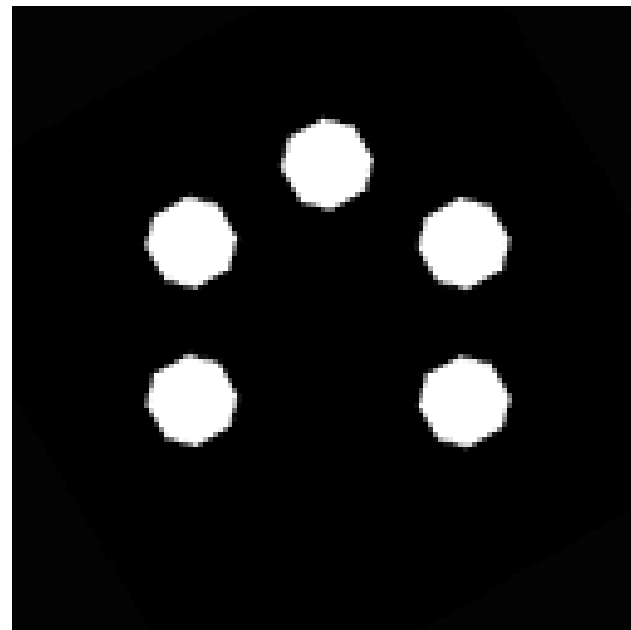
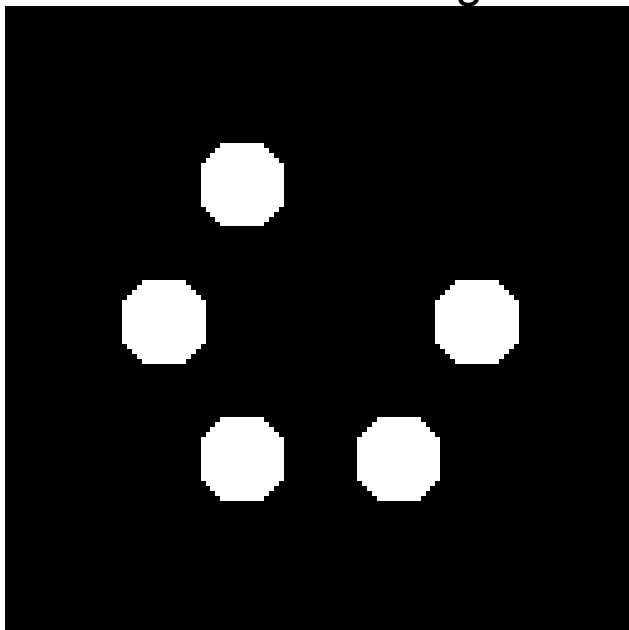


Image 2

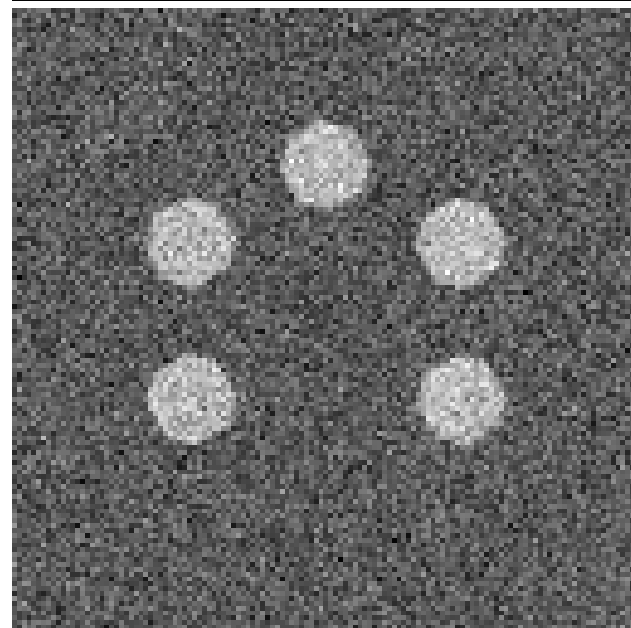
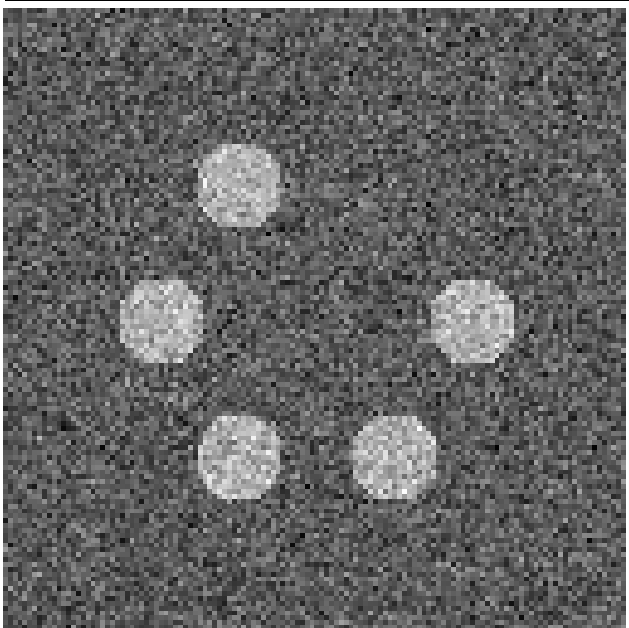
We take a series of rings from each image, unravel them, and compute a series of 1D cross-correlation functions.

Shifts along these unraveled CCFs is equivalent to a rotation in Cartesian space.

Reference image



Noise added



Orientation alignment

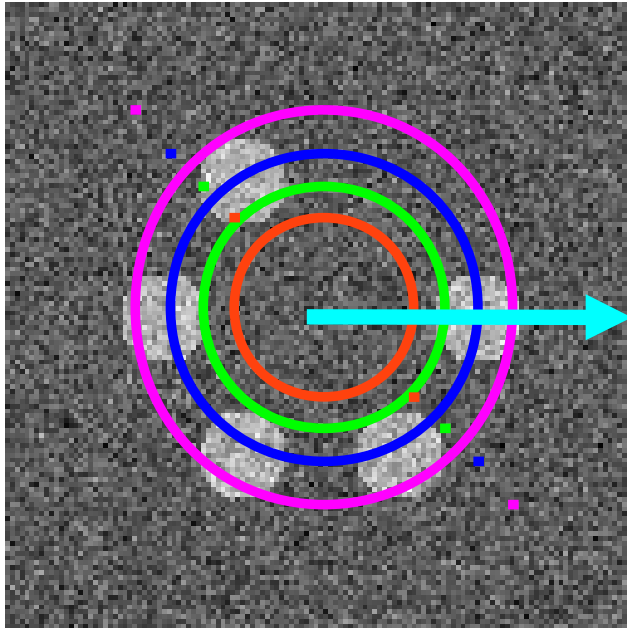


Image 1

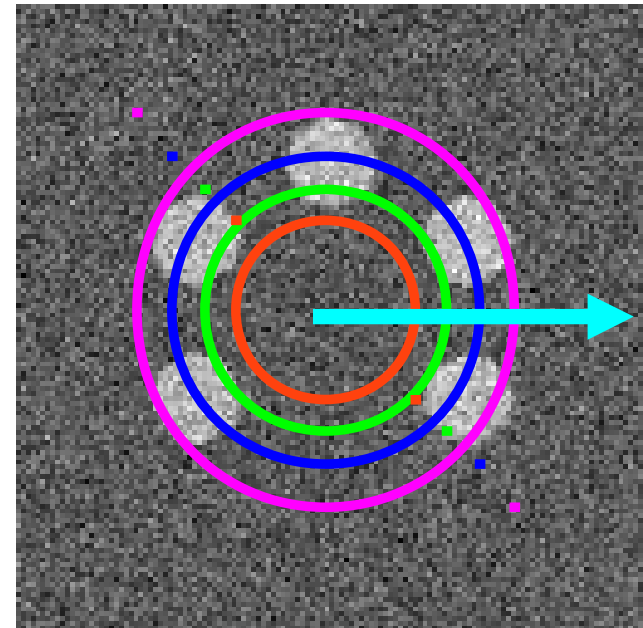
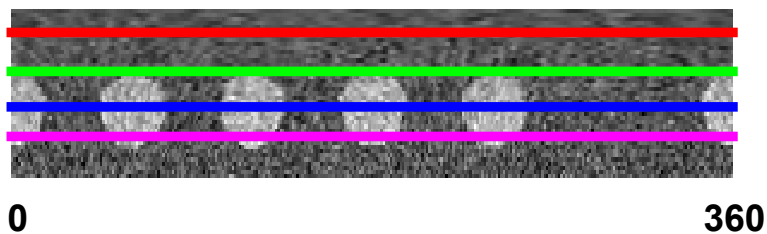
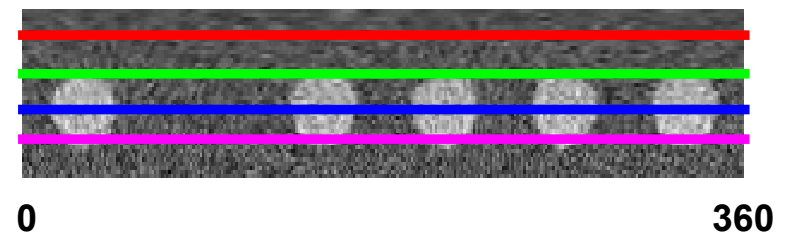


Image 2

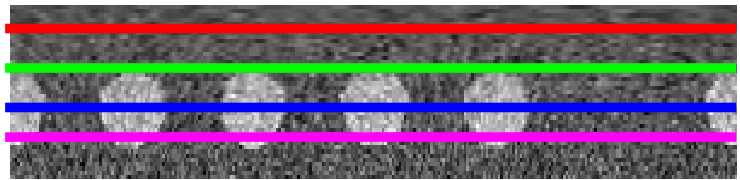


radius 1
radius 2
radius 3
radius 4



Polar representation

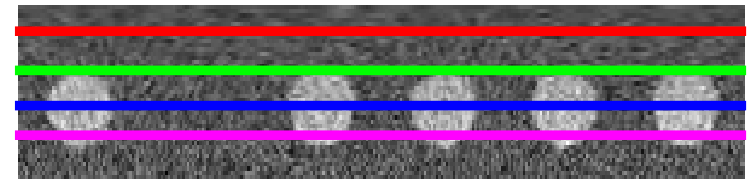
Orientation alignment



radius 1
radius 2
radius 3
radius 4

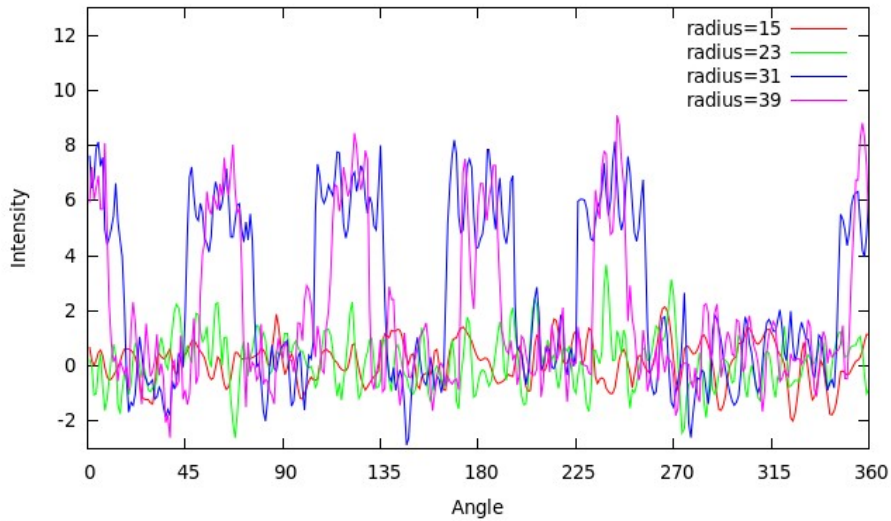
0

360

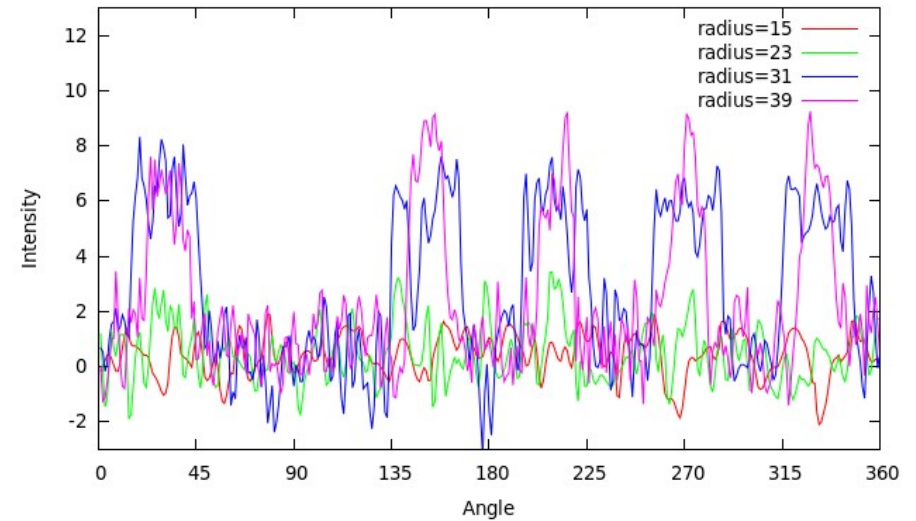


0

360

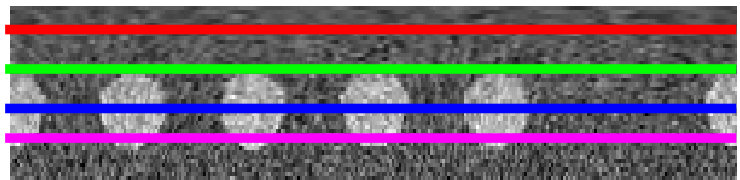


374.951, 4.53721



356.141, -2.50024

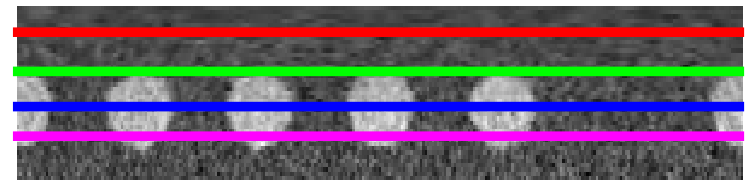
Orientation alignment: After rotation



radius 1
radius 2
radius 3
radius 4

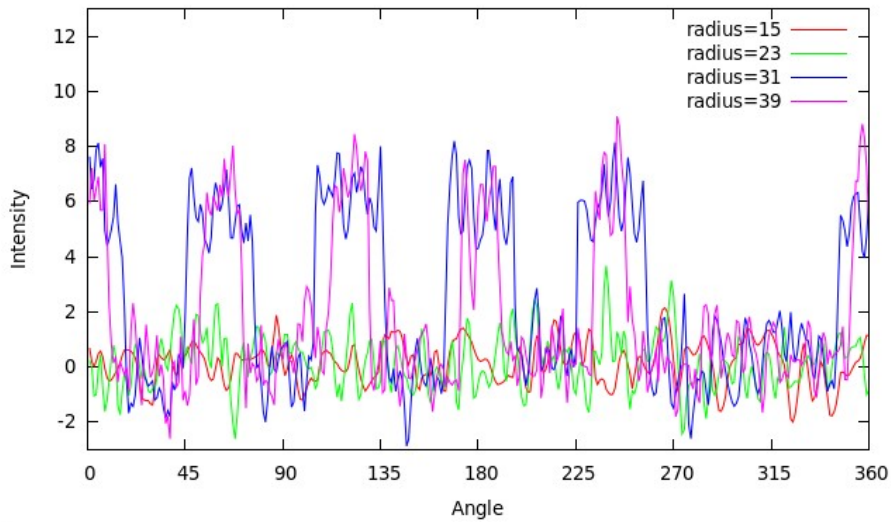
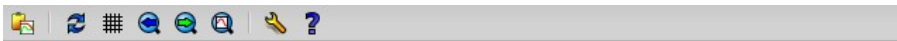
0

360

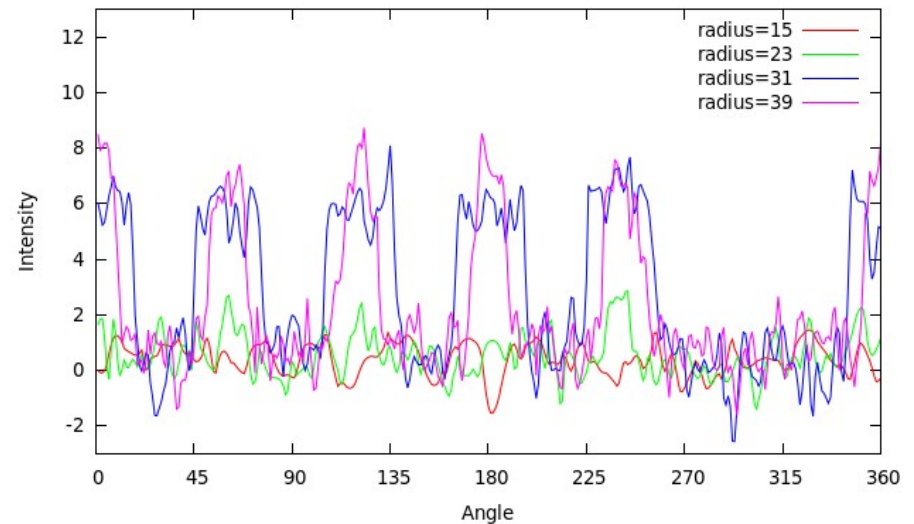
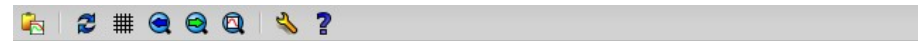


0

360



374.951, 4.53721



372.357, -3.21418

*Which do you perform first?
Translational or orientation alignment?*

Translational and orientation alignment are interdependent

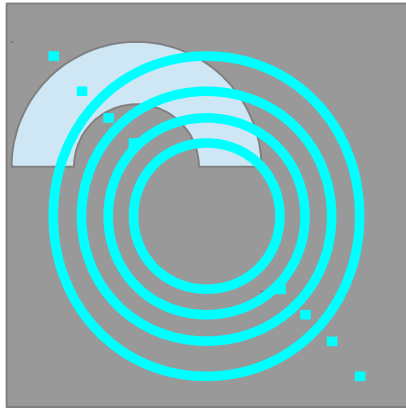


Image 1

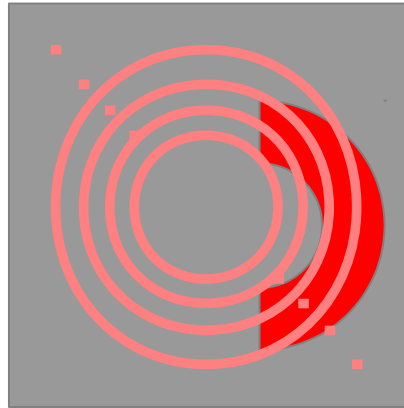
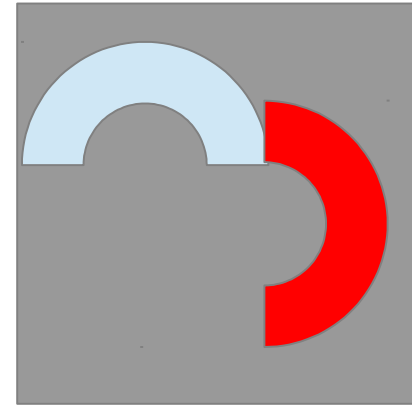


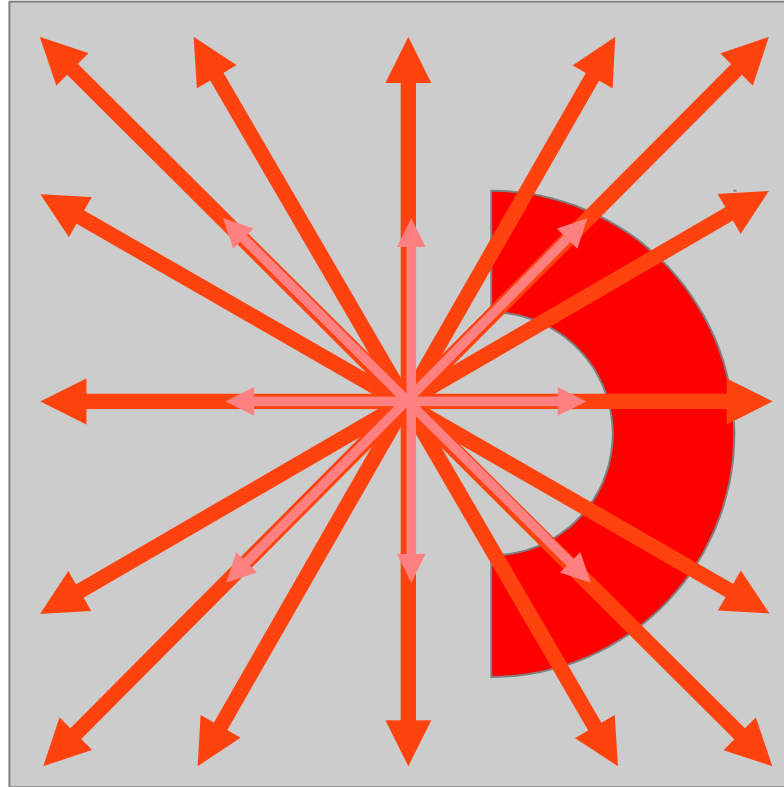
Image 2



Superimposed

SOLUTION: You try a set of reasonable shifts,
and perform separate orientation alignments for each.

Translational and orientation alignment are interdependent



Set of all shifts of up to 1 pixel

Set of all new shifts of up to 2 pixels

Shifts of (0, +/-1, +/-2) pixels results in 25 orientation searches.

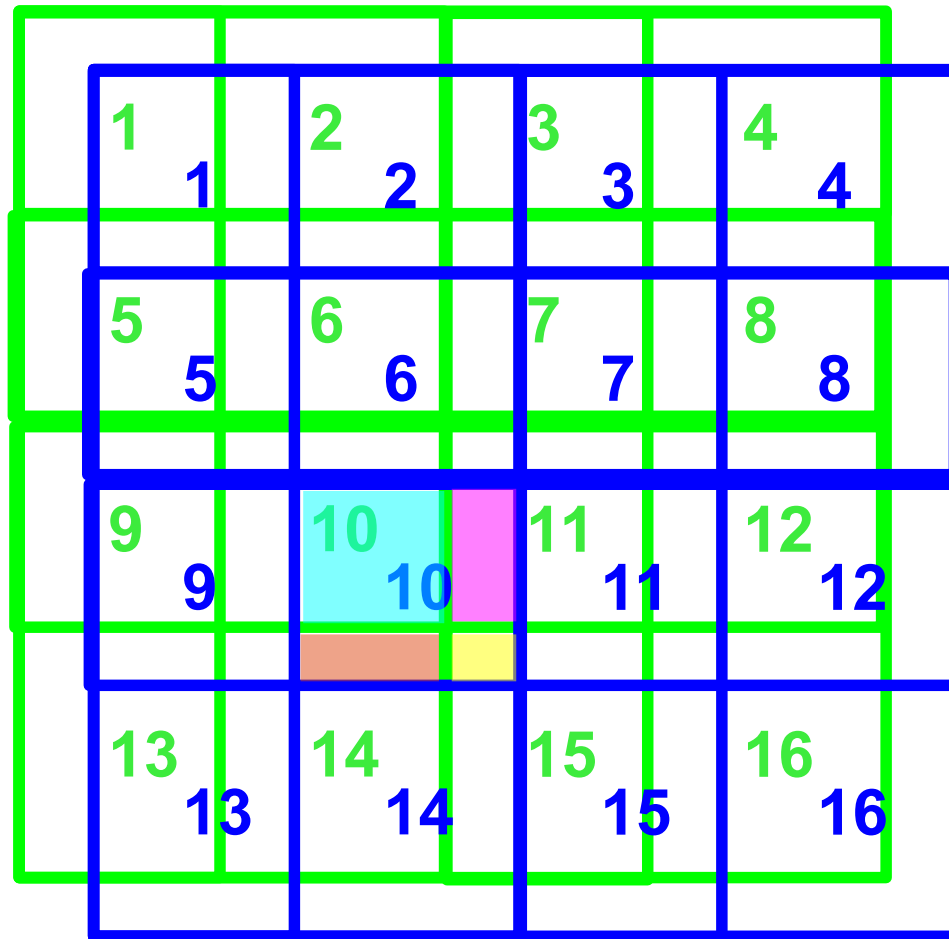
Outline

Image analysis II

- Fourier transforms revisited
 - Ducks and other animals
- Analogy to the Ewald sphere
- Aliasing
- Alignment
- **Interpolation**
- Multivariate data analysis

How to apply the best shift and rotation?

Shifts

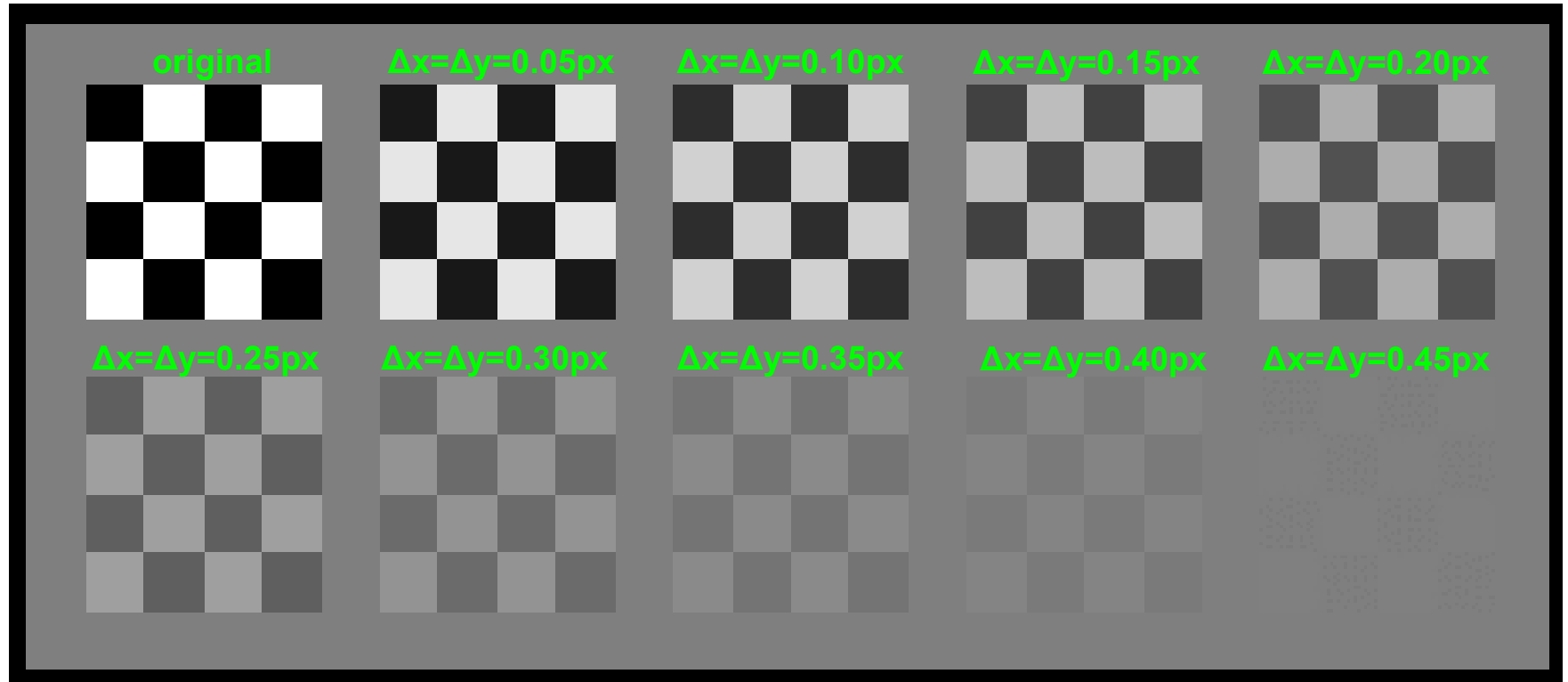


Suppose we shift the image in x & y .

The new pixels will be weighted averages of the old pixels.

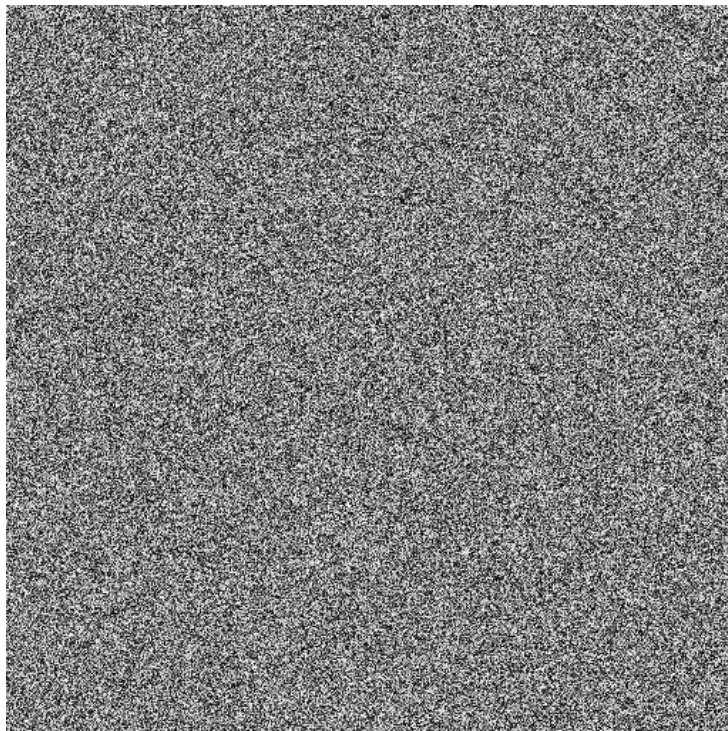
The more the mix the pixels, the worse the result will be. 

Effect of shifts

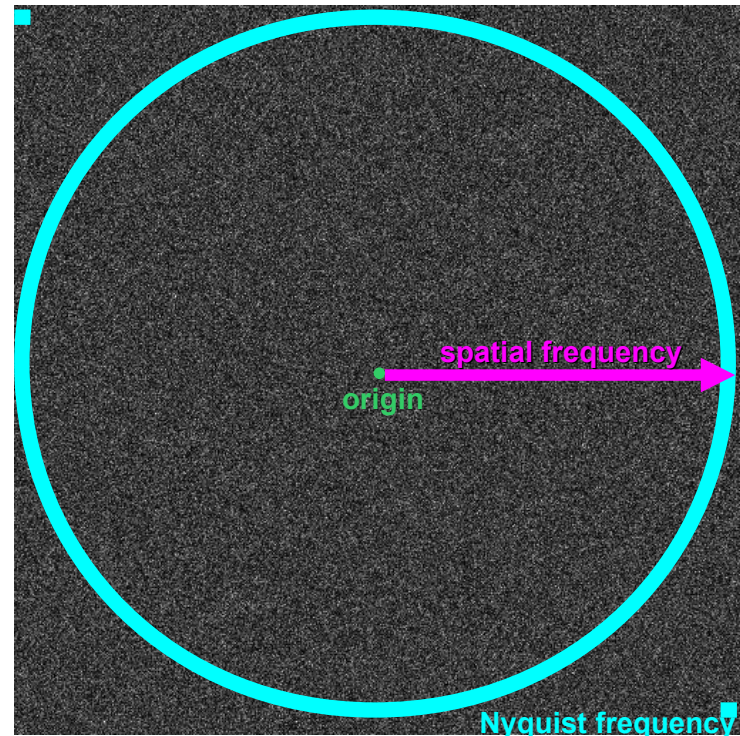


Two more properties of Fourier transforms: Noise

- ◆ The Fourier transform of noise is noise
- ◆ “White” noise is evenly distributed in Fourier space
 - “White” means that each pixel is independent



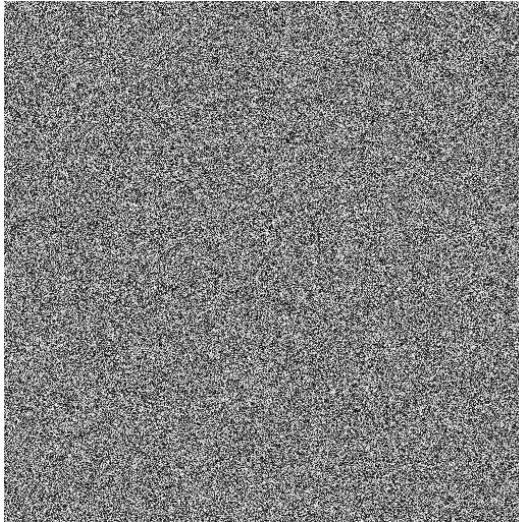
White noise



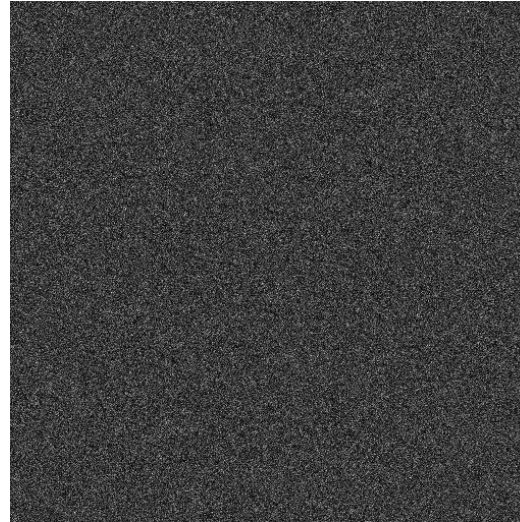
Power spectrum

Effects of interpolation are resolution-dependent

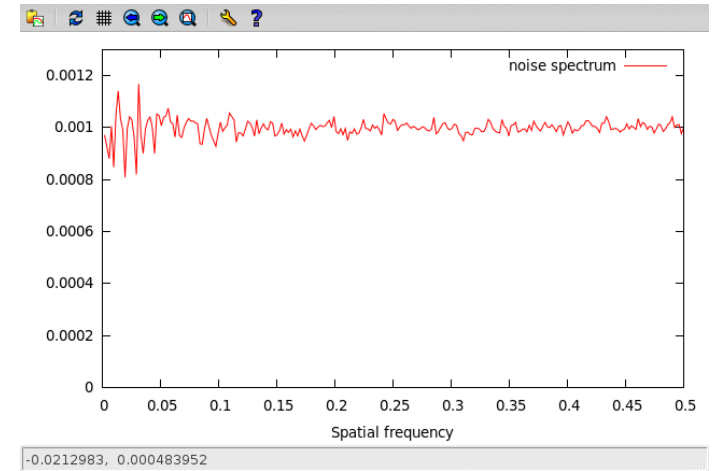
Original



Image

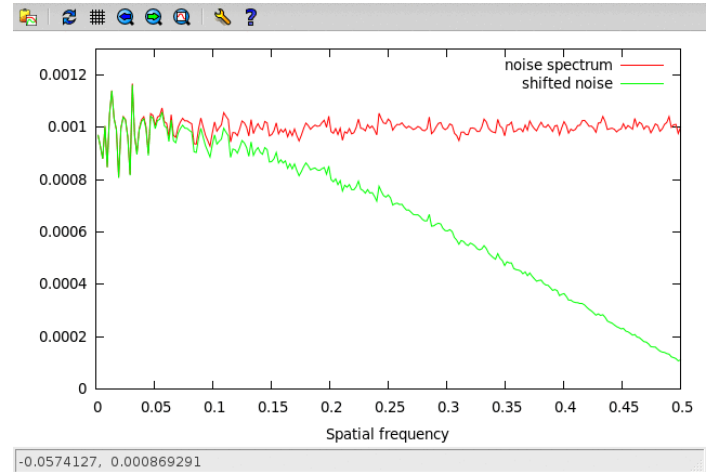
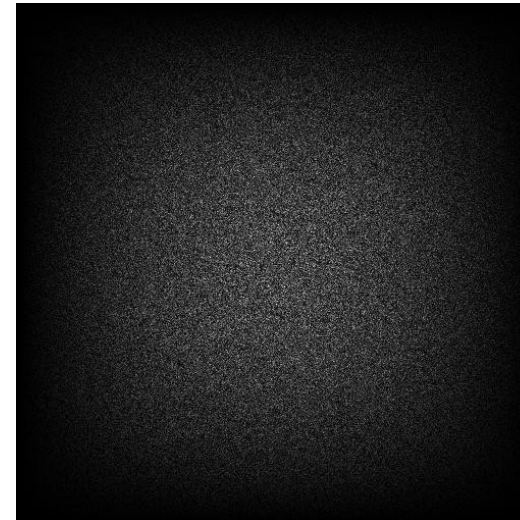
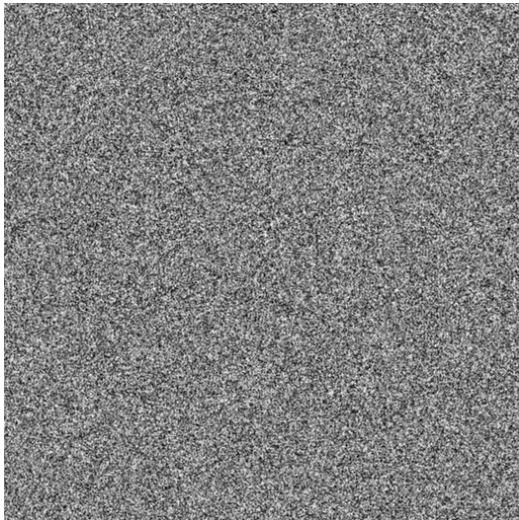


Power spectrum

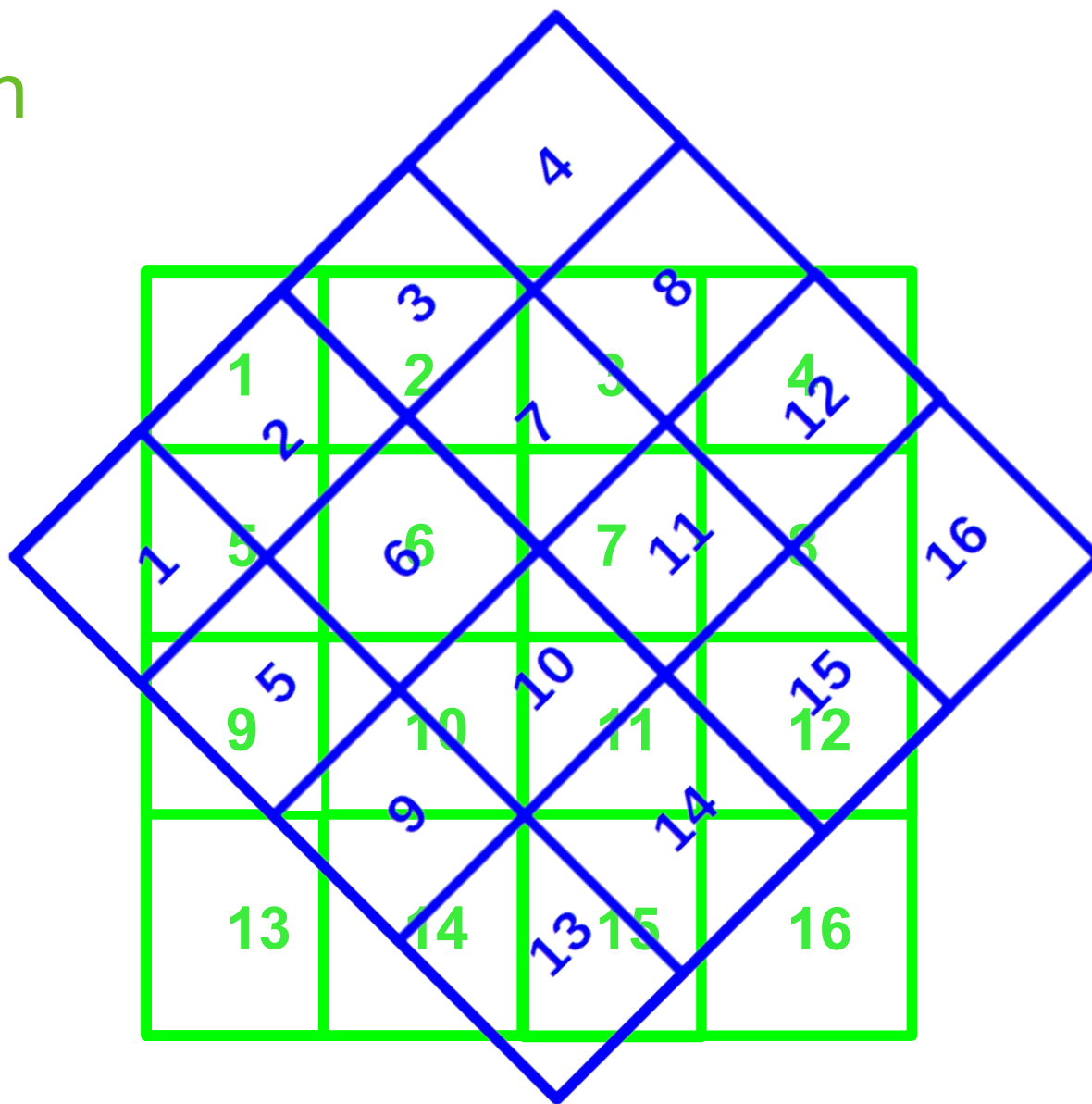


Profile

Shifted by (0.5,0.5) px



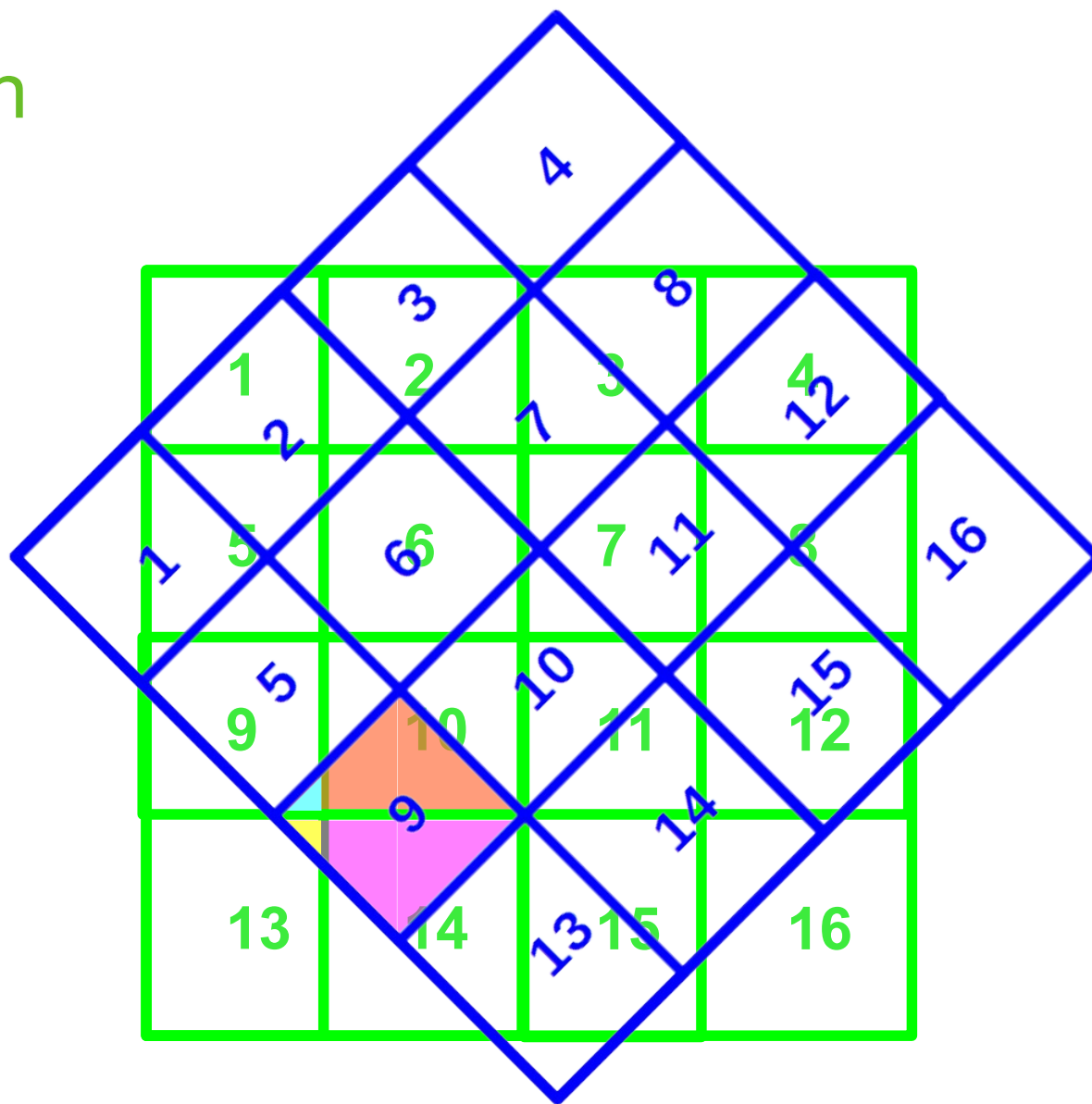
Rotation



Suppose we rotate the image.

The new pixels will be weighted averages of the old pixels.

Rotation



Suppose we rotate the image.

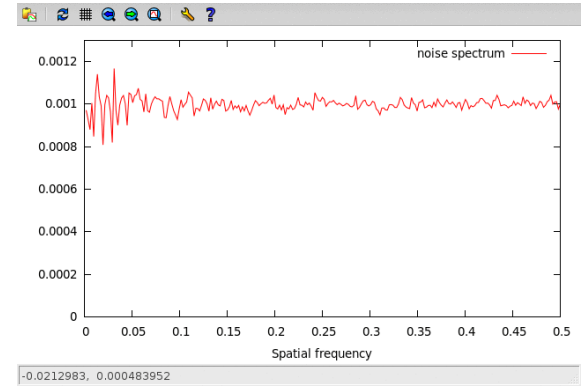
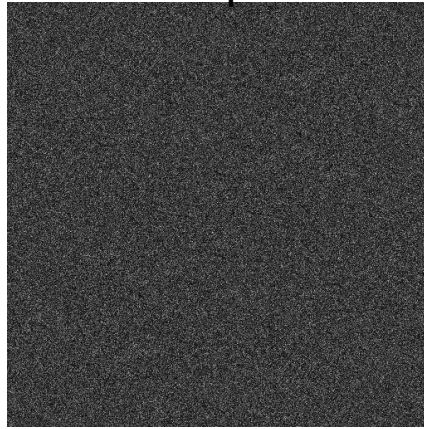
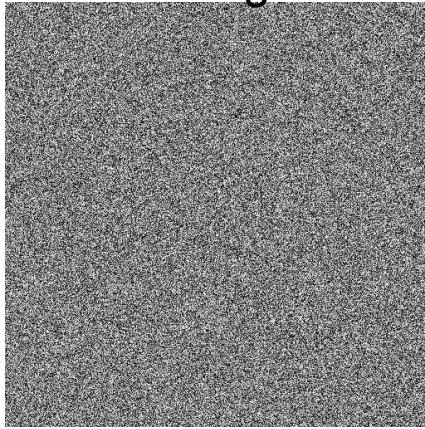
New pixel #9 will be a weighted sum of old pixels 9, 10, 13, and 14.

Image

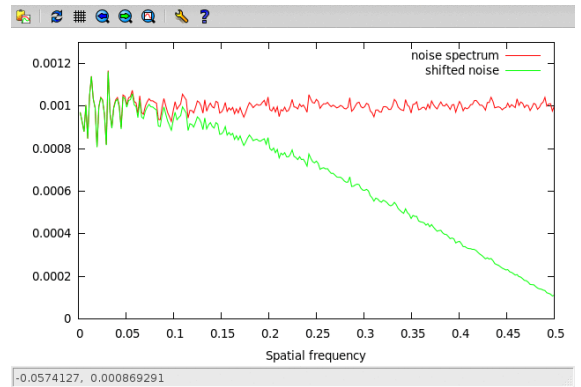
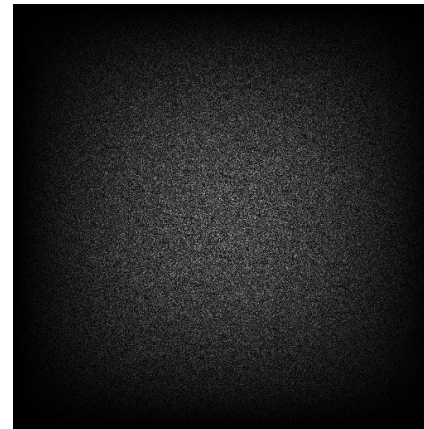
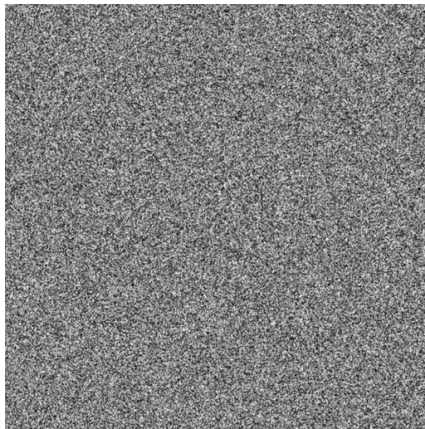
Power spectrum

Power spectrum profile

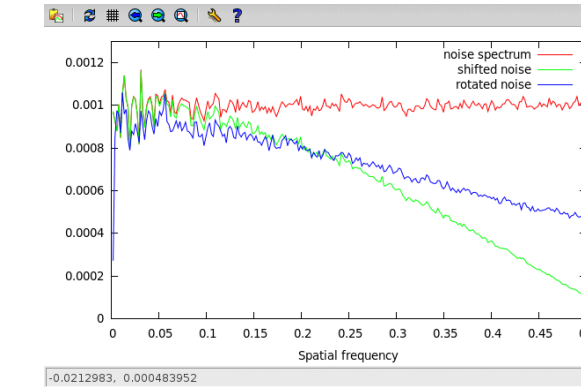
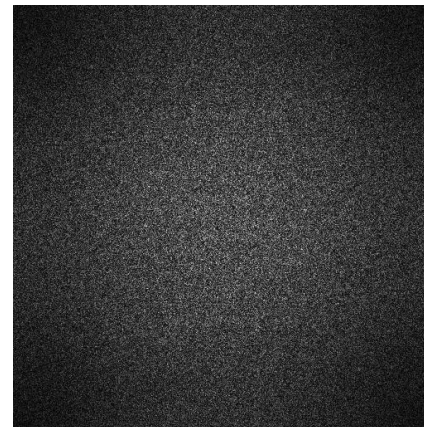
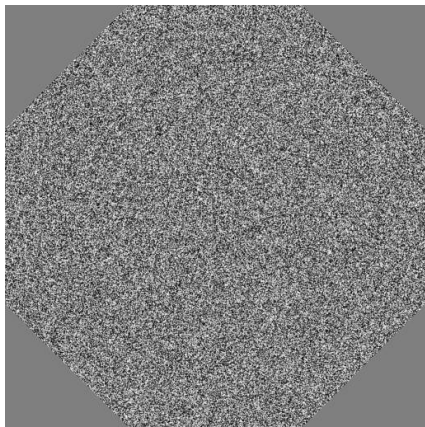
Original



Shifted by (0.5,0.5) px



Rotated by 45°



The degradation of the images means that we should minimize the number of interpolations.

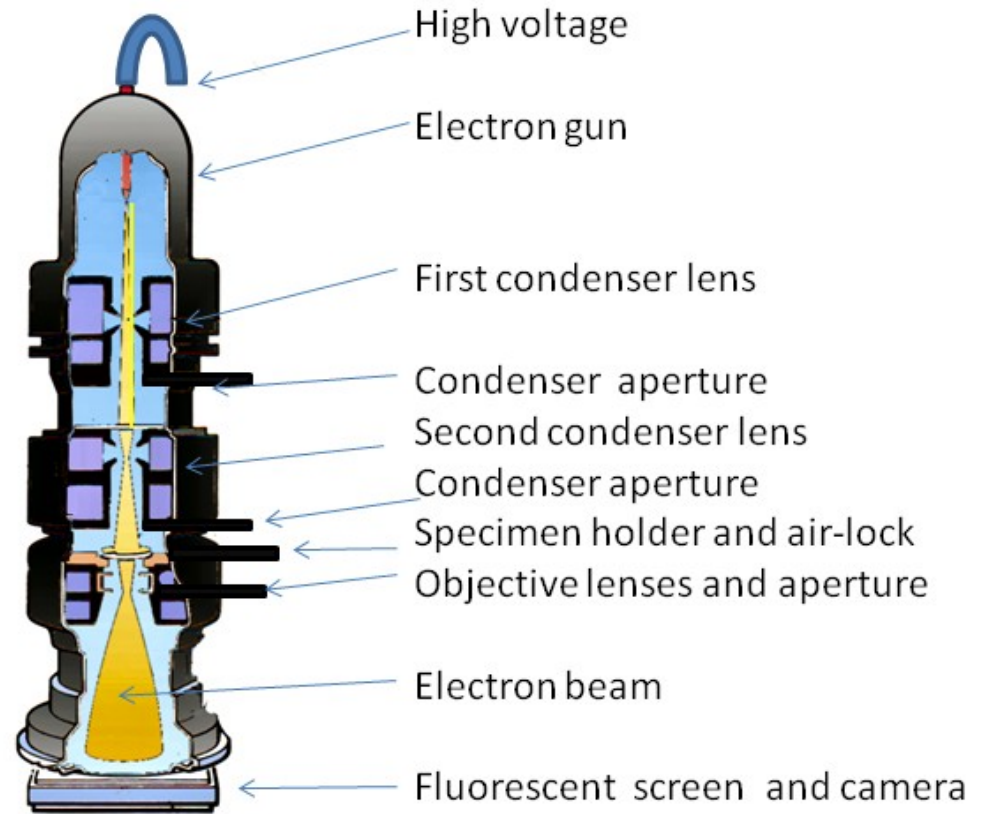
From two weeks ago...

Typical magnification: 50,000X
Typical detector element: 15 μ m
(pixel size on the camera scale)

Pixel size on the specimen scale:
 $15 \times 10^{-6} \text{ m/px} / 50000 =$
 $3.0 \times 10^{-10} \text{ m/px} = \mathbf{3.0 \text{ \AA/px}}$

In other words,
the best resolution we
can achieve (or, the
finest oscillation we
can detect) at 3.0 \AA/px
is **6.0 \AA** .

It will be worse due to interpolation,
so to be safe, a pixel should be 3X
smaller than your target resolution.

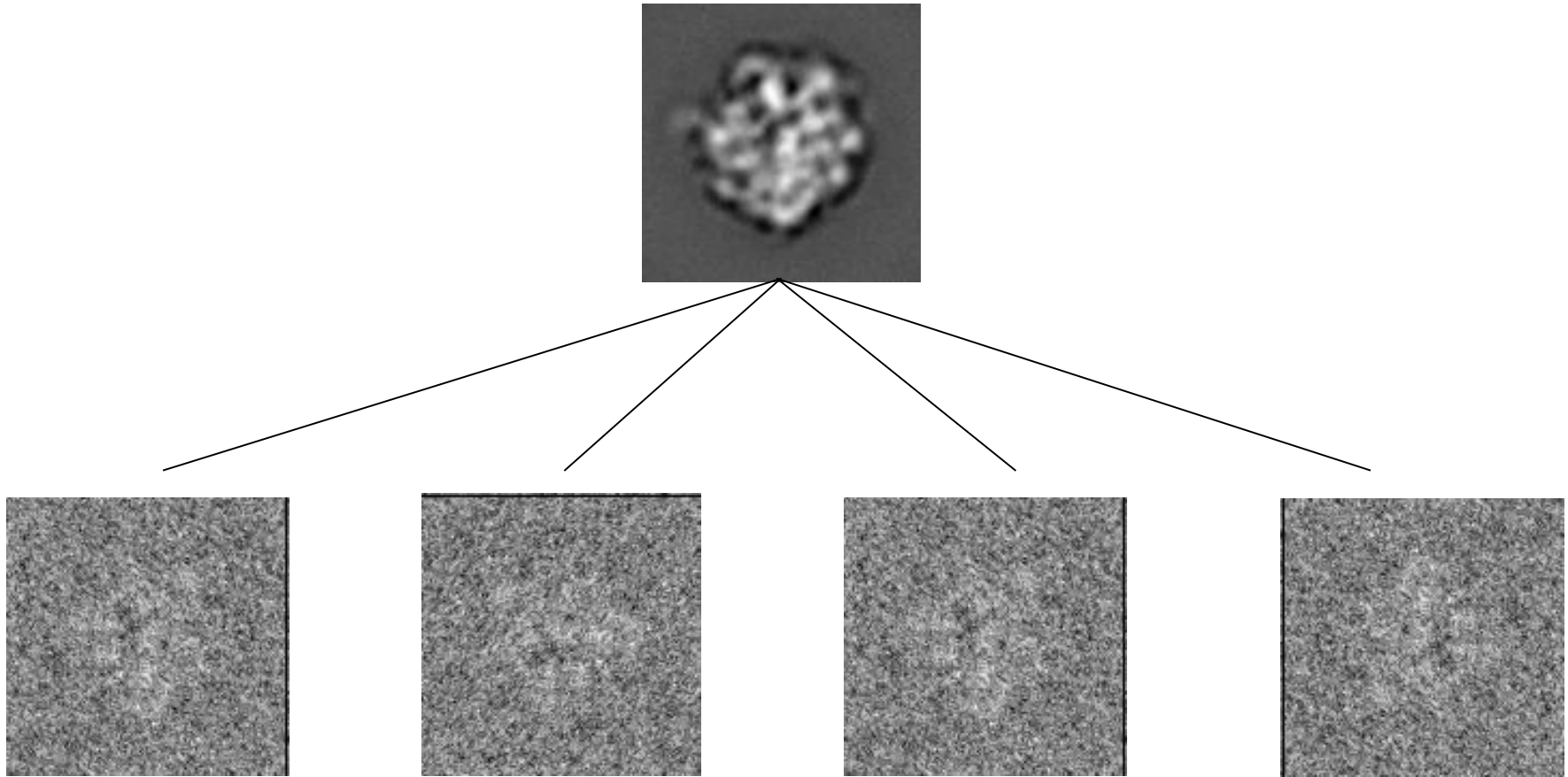


Transmission Electron Microscope

<http://www.en.wikipedia.org>

Different alignment strategies

Reference-based alignment



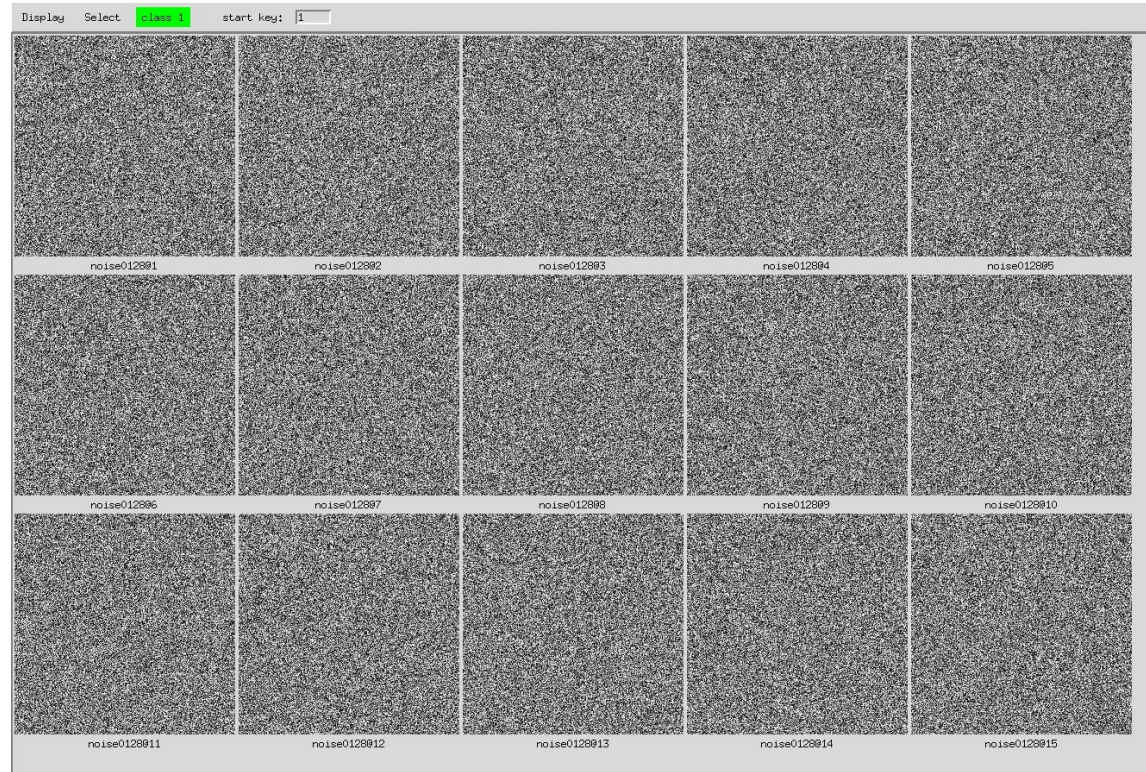
There's a problem with reference-based alignment:

Model bias

Model bias

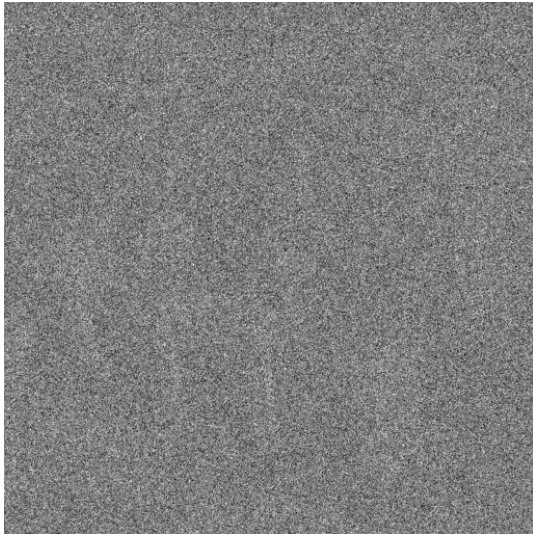


Reference

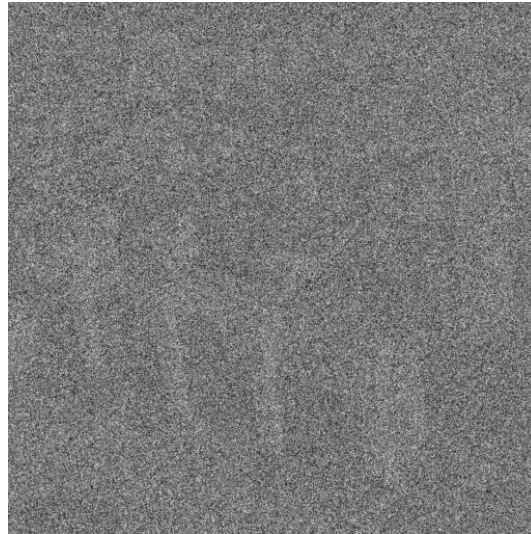


Images of pure noise

Averages of images of pure noise



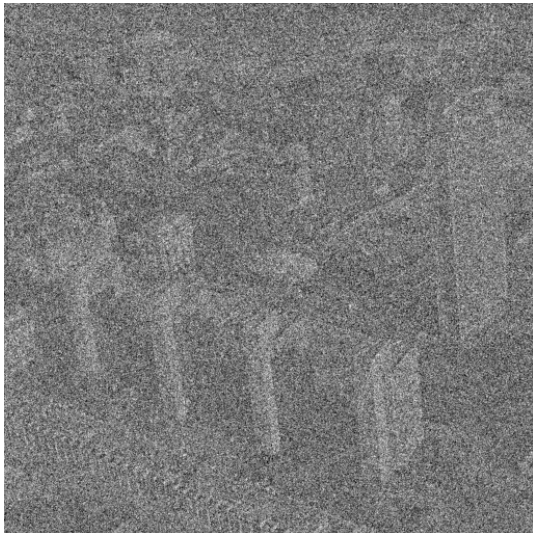
N = 128



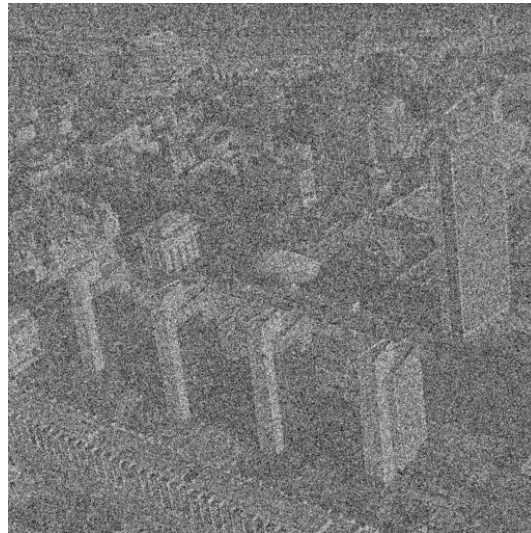
N = 256



N = 512



N = 1024



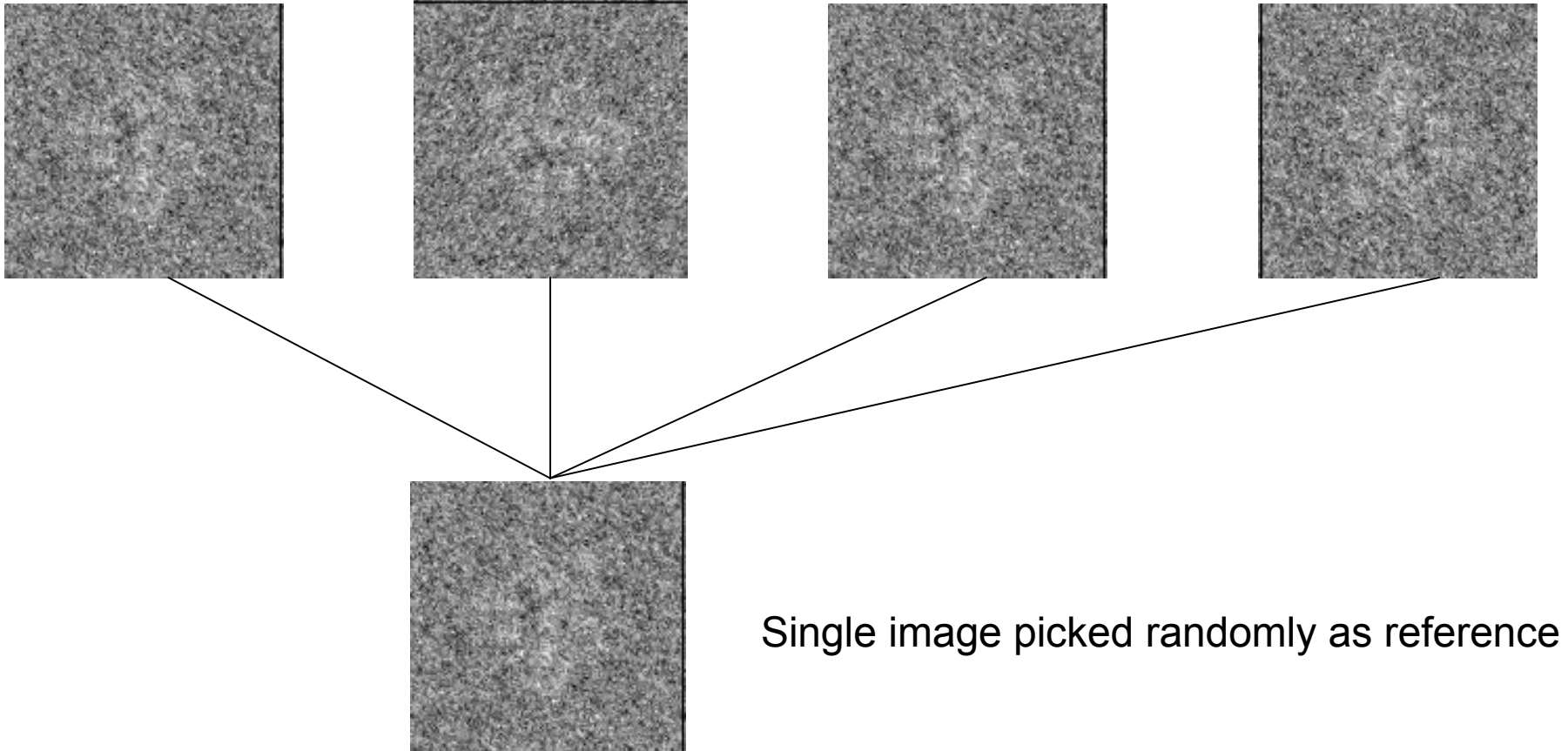
N = 2048



original

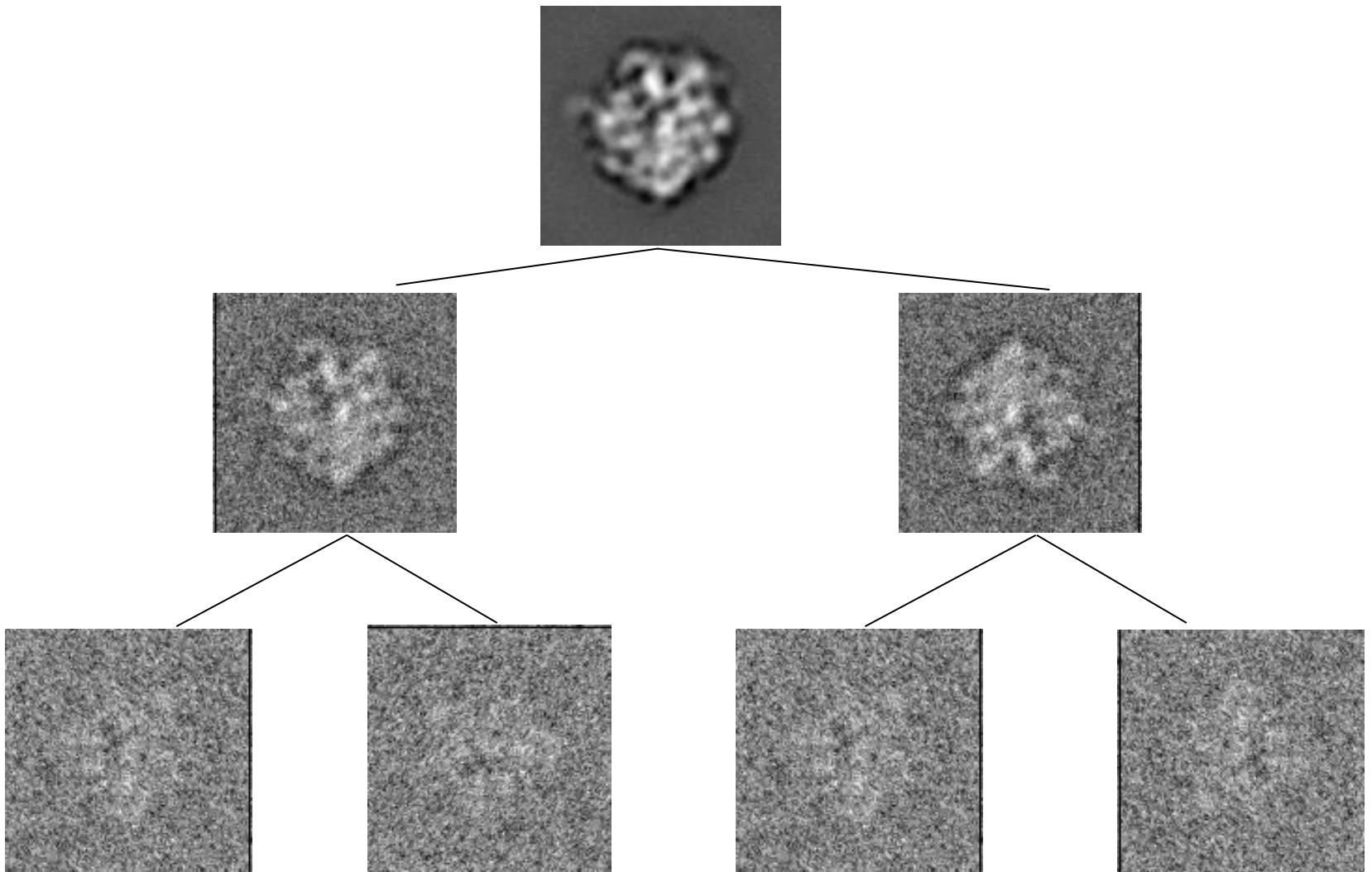
There are reference-free alignment schemes

Reference-free alignment (SPIDER command AP SR)



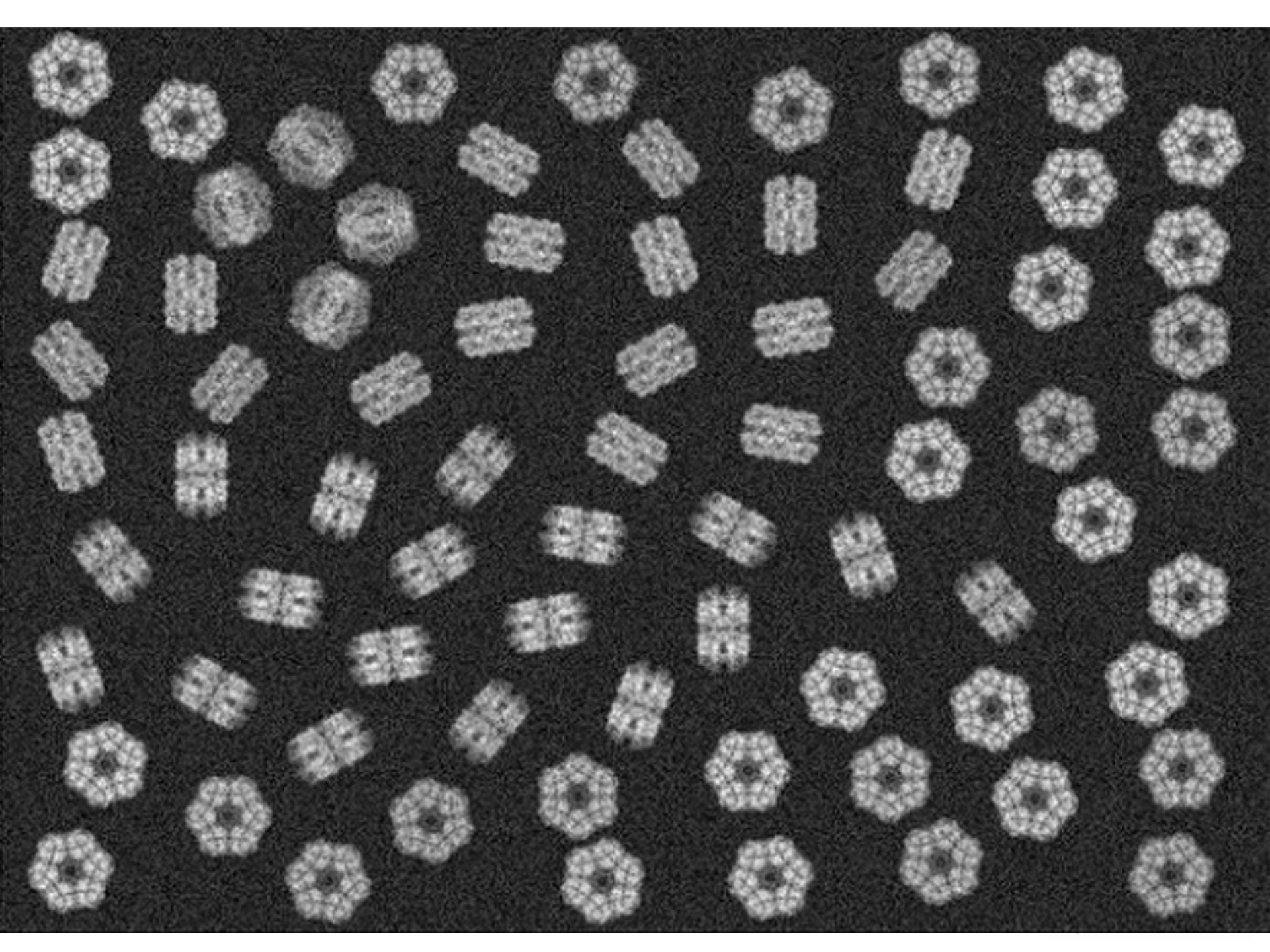
Disadvantage: Alignment depends on the choice of random seed.

Pyramidal/pairwise alignment



Marco... Carrascosa (1996) Ultramicroscopy

*You have aligned images,
but they don't all look the same.*

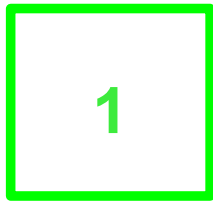


Outline

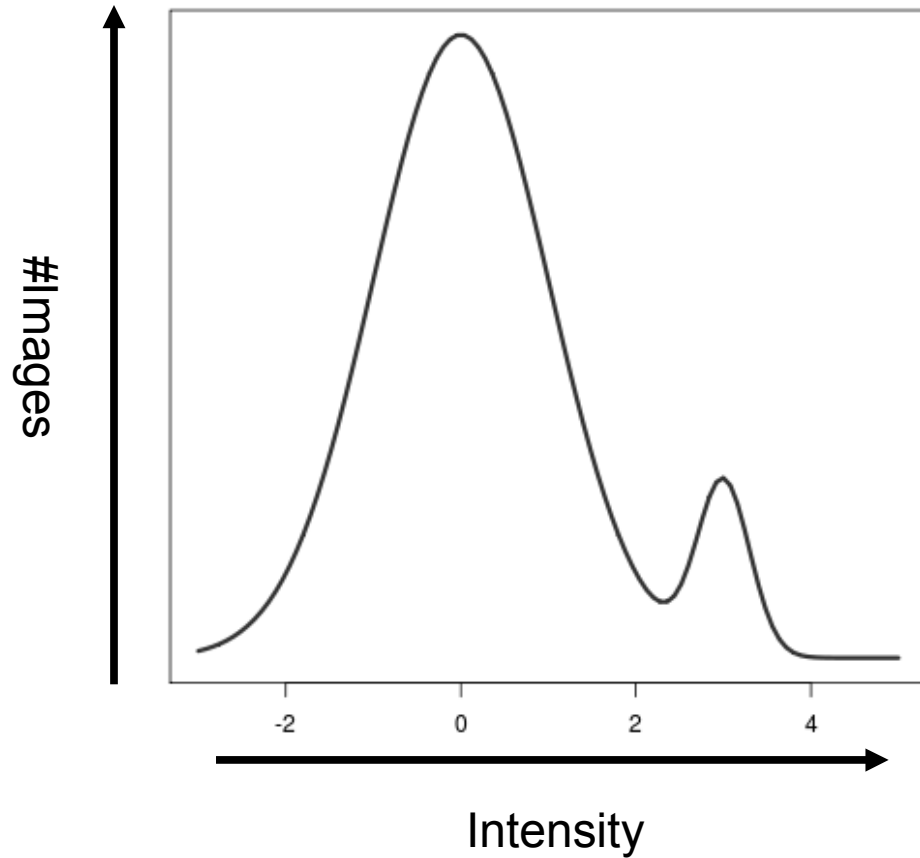
Image analysis II

- Fourier transforms revisited
 - Ducks and other animals
- Analogy to the Ewald sphere
- Aliasing
- Alignment
- Interpolation
- Multivariate data analysis

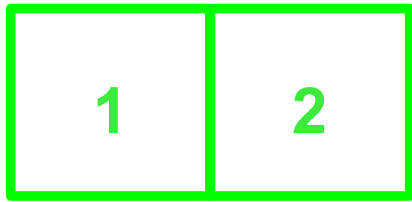
A one-pixel image



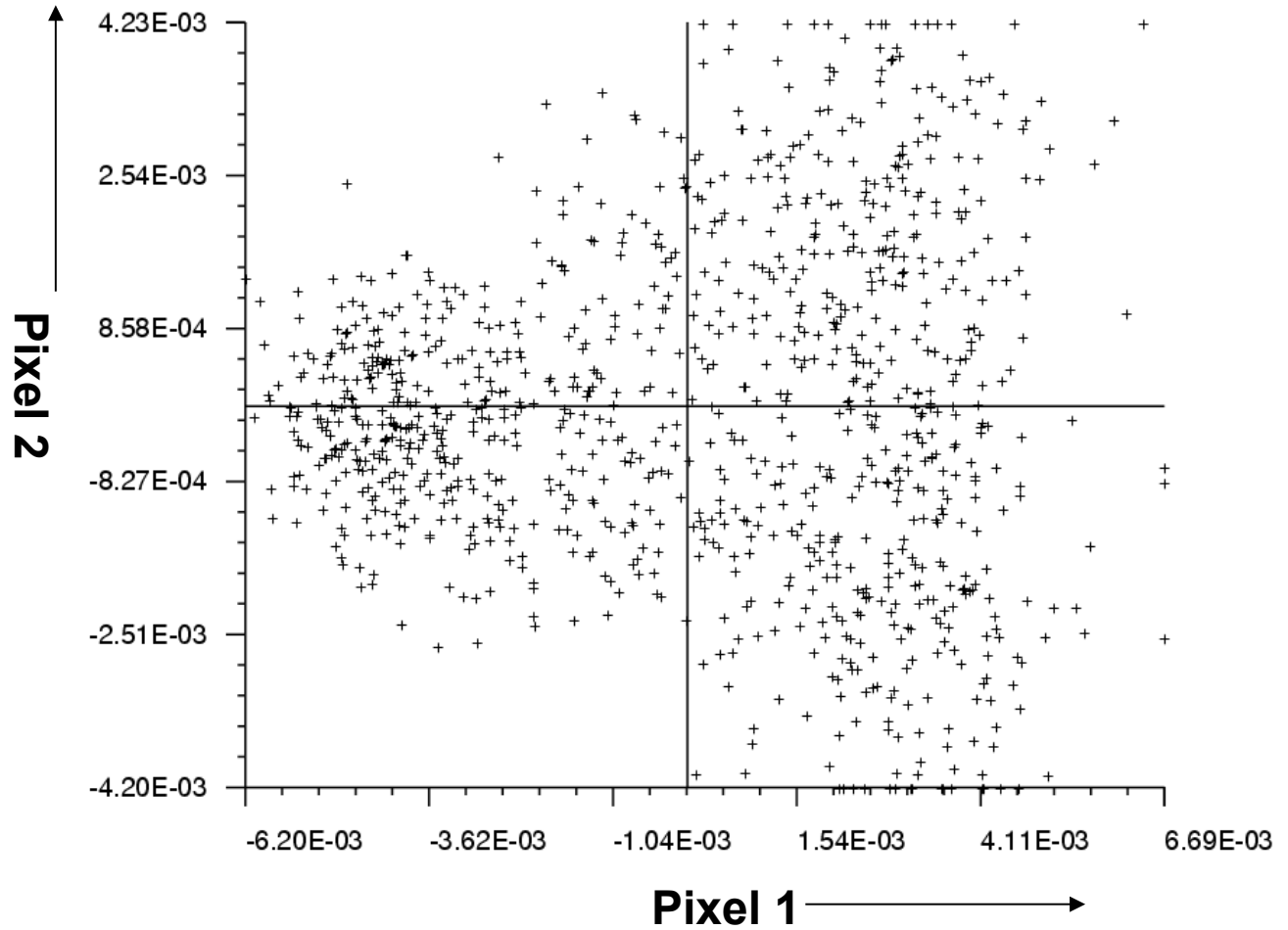
1-pixel image



A two-pixel image



2-pixel image



A 16-pixel image

1	2	3	4
5	6	7	8
9	10	11	12
13	14	15	16

Now, we have a 16-dimensional problem.

Multivariate data analysis (MDA), or Multivariate statistical analysis (MSA)

1	2	3	4
5	6	7	8
9	10	11	12
13	14	15	16

1	2	3	4
5	6	7	8
9	10	11	12
13	14	15	16

Suppose pixel 6 coincided with pixel 11,
And pixel 7 coincided with pixel 10.
Then, we're back to two variables, and a 2D problem.

Multivariate data analysis (MDA), or Multivariate statistical analysis (MSA)



Our 16-pixel image can be reorganized into a 16-coordinate vector.

Covariance of measurements x and y :

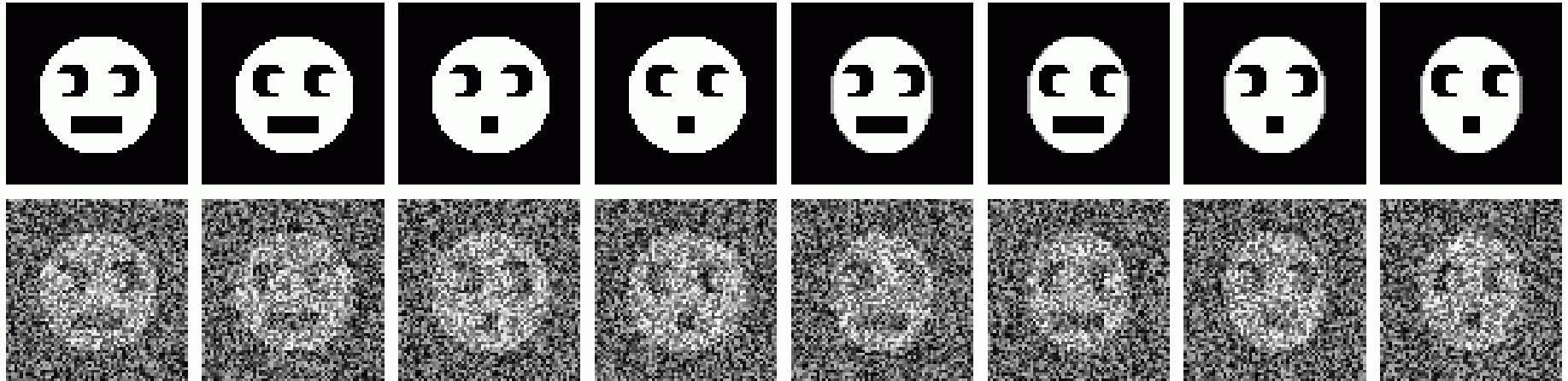
$$\langle xy \rangle - \langle x \rangle \langle y \rangle,$$

where $\langle x \rangle$ is the mean of x .

A high covariance is a measure of the correlation between two variables.

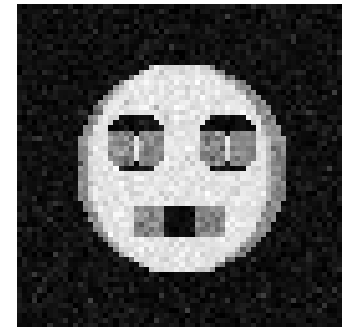
MDA: An example

8 classes of faces, 64x64 pixels



With noise added

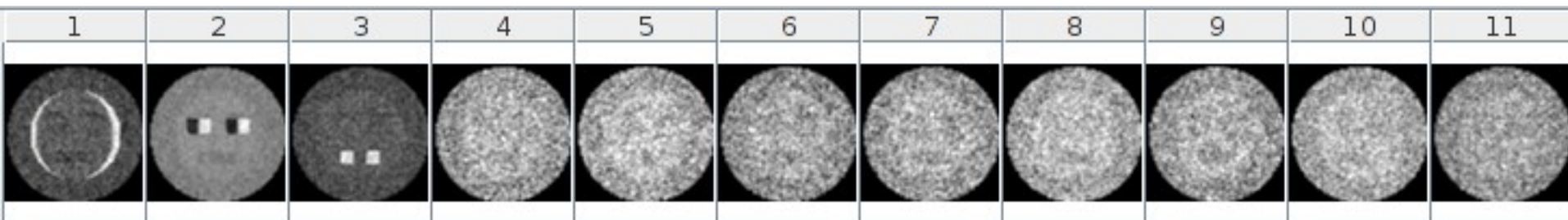
Average:



From http://spider.wadsworth.org/spider_doc/spider/docs/techs/classification/tutorial.html

Principal component analysis (PCA) or Correspondence analysis (CA)

- ◆ For a 4096-pixel image, we will have a 4096x4096 covariance matrix.
- ◆ Row-reduction of the covariance matrix gives us “eigenvectors.”
 - The eigenvectors describe correlated variations in the data.
 - The eigenvectors have 4096 elements and can be converted back into images, called “eigenimages.”
 - The first eigenvectors will account for the most variation. The later eigenvectors may only describe noise.
 - Linear combinations of these images will give us approximations of the classes that make up the data.

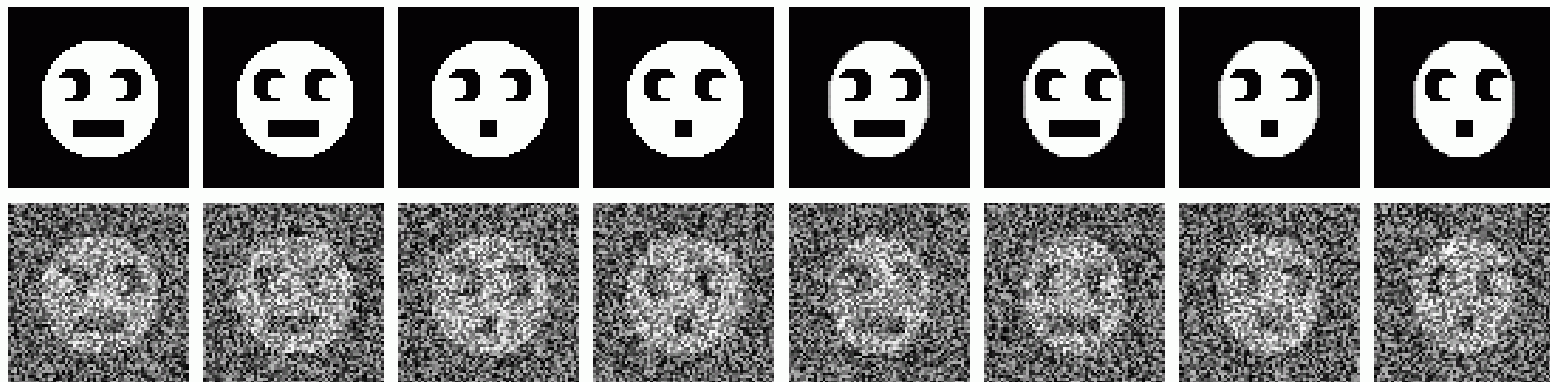


Eigenimages

Reconstituted images

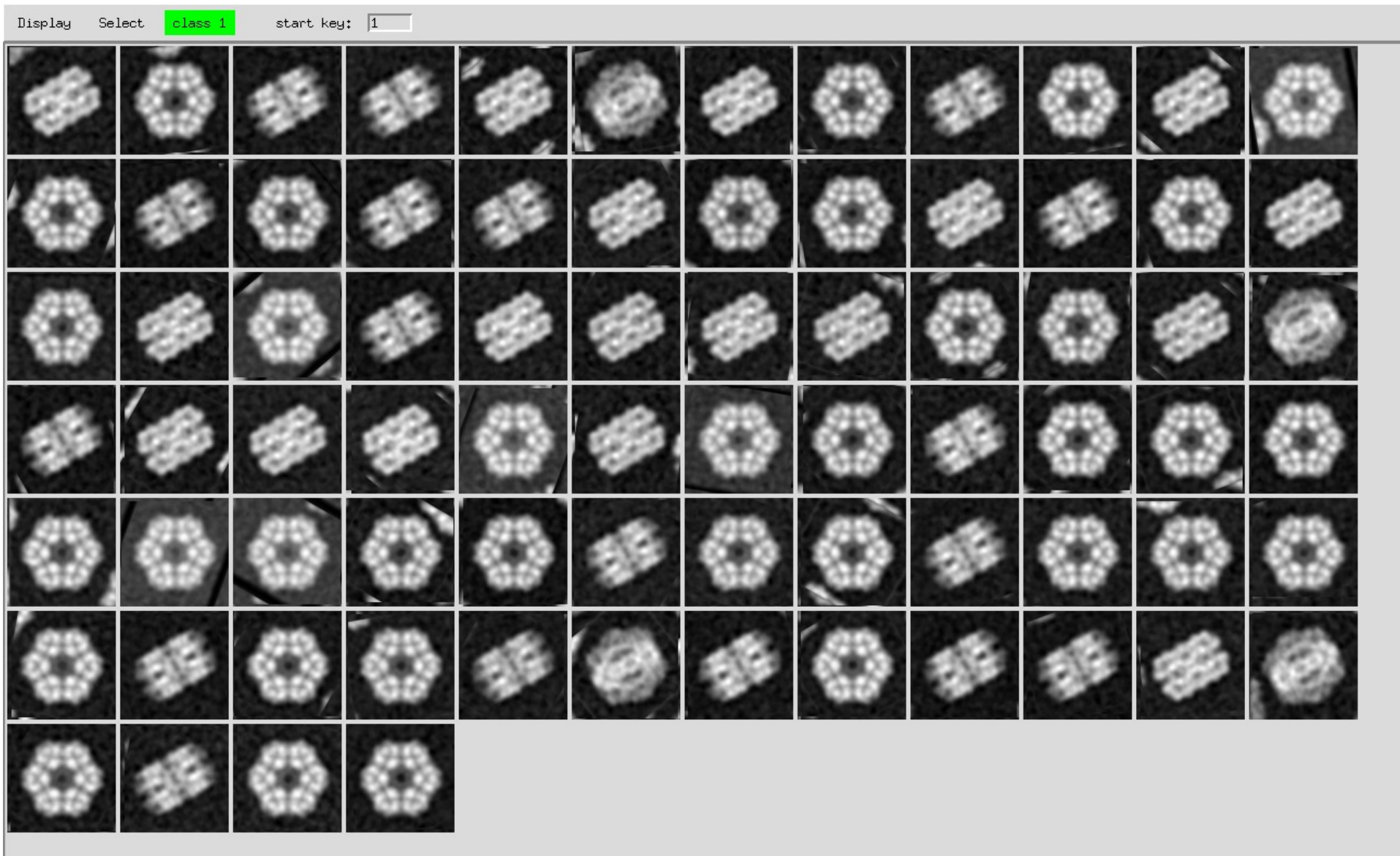
Linear combinations of these images will give us approximations of the classes that make up the data.

$$c_0 \begin{array}{c} \text{Average} \\ \text{Eigenimage \#1} \\ \text{Eigenimage \#2} \\ \text{Eigenimage \#3} \end{array} + c_1 \begin{array}{c} \text{Eigenimage \#1} \\ \text{Eigenimage \#2} \\ \text{Eigenimage \#3} \end{array} + c_2 \begin{array}{c} \text{Eigenimage \#2} \\ \text{Eigenimage \#3} \end{array} + c_3 \begin{array}{c} \text{Eigenimage \#3} \end{array} + \dots$$



A reminder of what our original images looked like

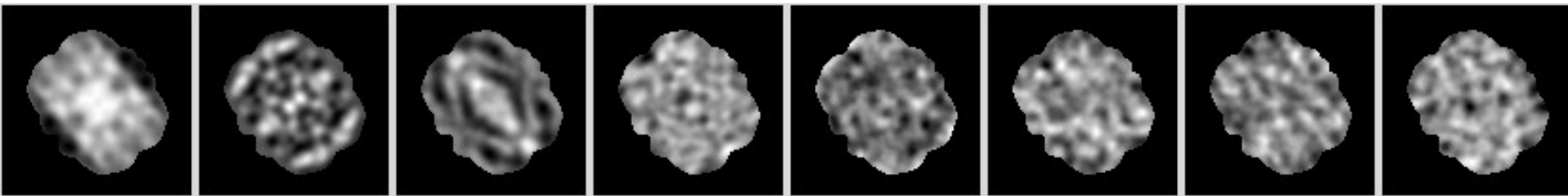
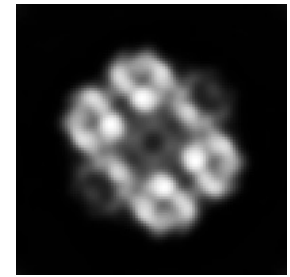
Another example: worm hemoglobin



Phantom images of worm hemoglobin

PCA of worm hemoglobin

Average:



stkeigenimg@1 stkeigenimg@2 stkeigenimg@3 stkeigenimg@4 stkeigenimg@5 stkeigenimg@6 stkeigenimg@7 stkeigenimg@8

$+c_0$

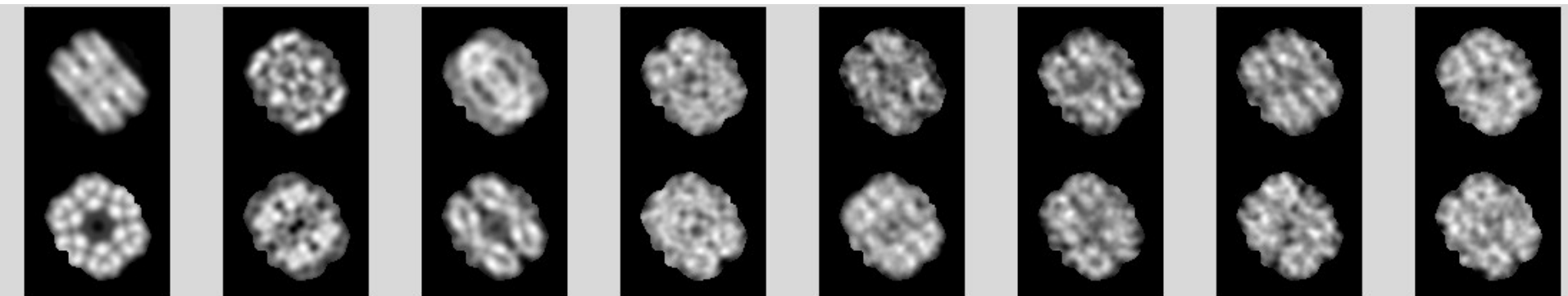
$+c_1$

$+c_2$

$+c_3$

$+c_4$

$+c_5$



stkreconstituted@1 stkreconstituted@2 stkreconstituted@3 stkreconstituted@4 stkreconstituted@5 stkreconstituted@6 stkreconstituted@7 stkreconstituted@8

$-c_0$

$-c_1$

$-c_2$

$-c_3$

$-c_4$

$-c_5$

*Next week:
Classification & 3D Reconstruction*

Thank you for your attention



Central European Institute of Technology
Masaryk University
Kamenice 753/5
625 00 Brno, Czech Republic

www.ceitec.muni.cz | info@ceitec.muni.cz



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