

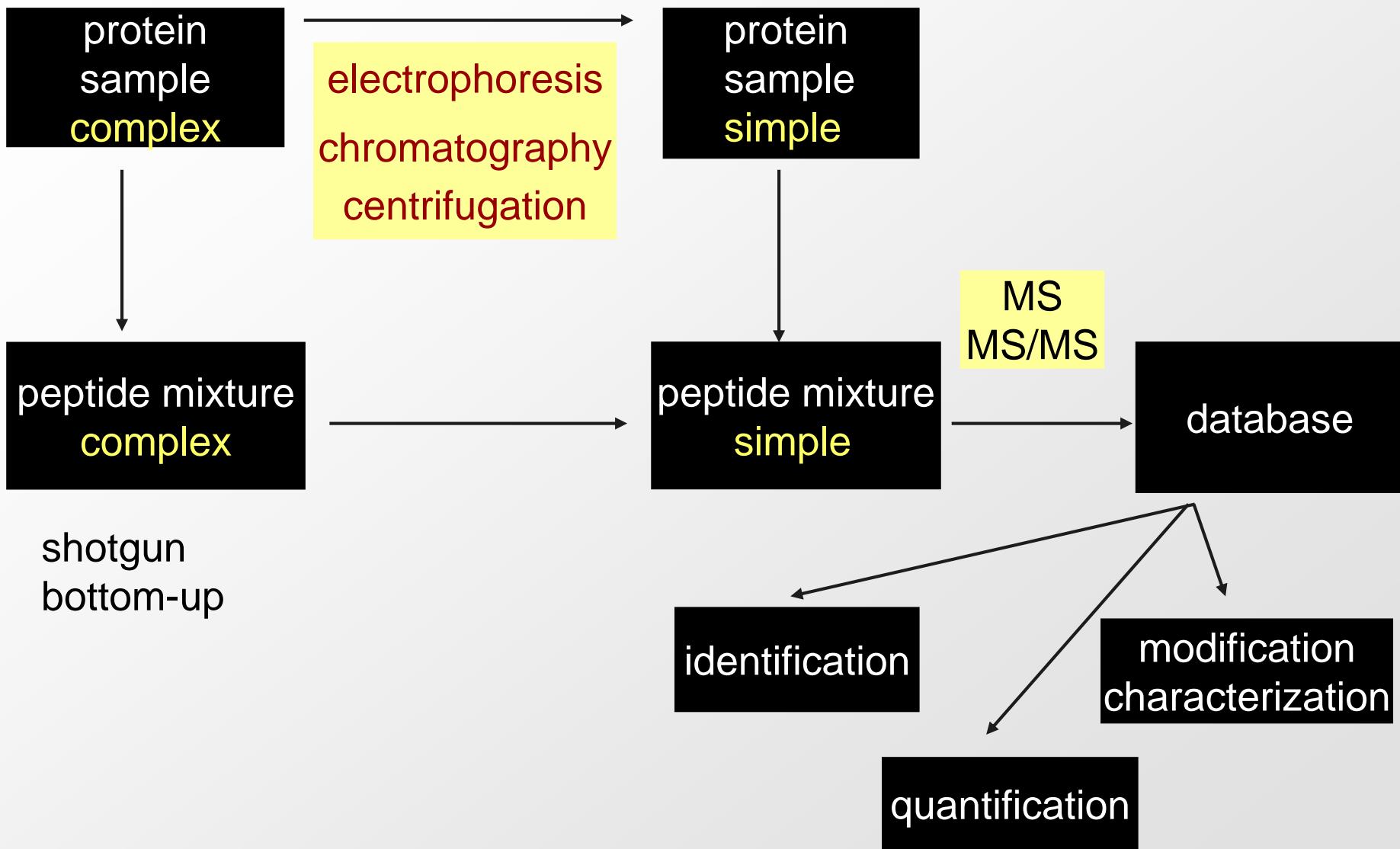
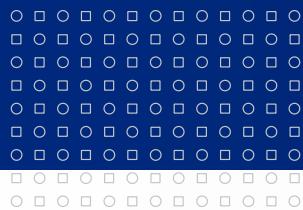
# PROTEOMIC SAMPLE PREPARATION

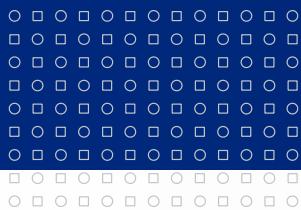
## Two-dimensional electrophoresis



Hana Konečná  
Proteomics Core Facility  
**CEITEC** Central European Institute of Technology  
**NCBR** National Centre for Biomolecular Research

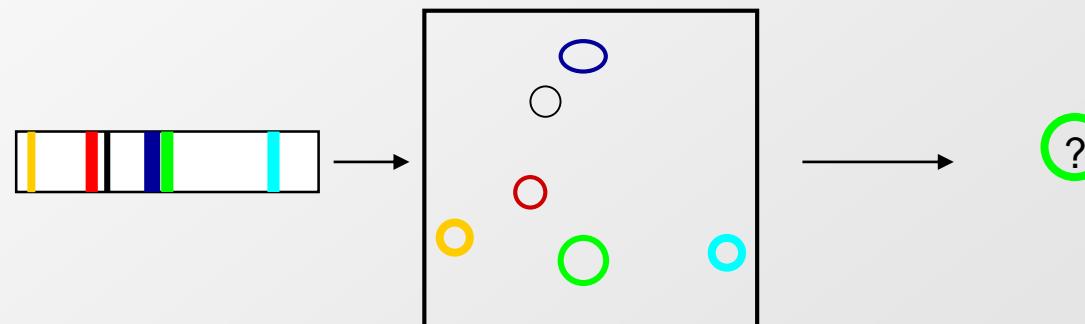


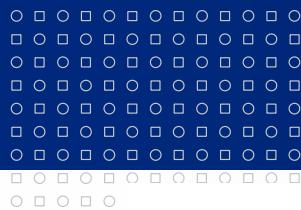




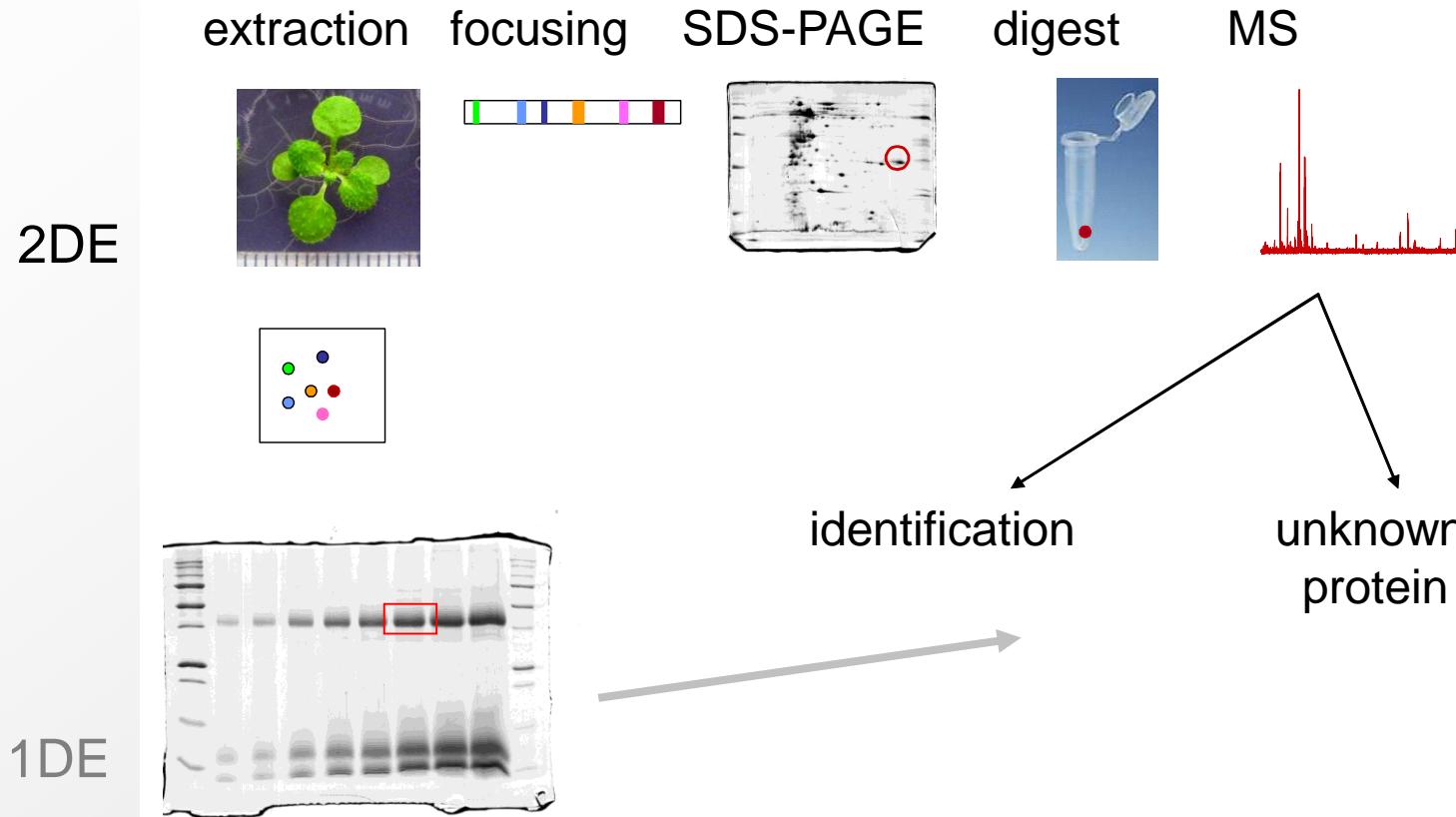
# I. SEPARATION II. PREFRACTIONATION

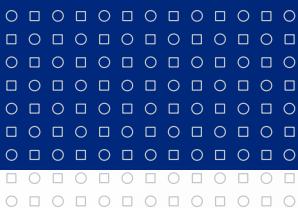
Two-dimensional electrophoresis 2-DE





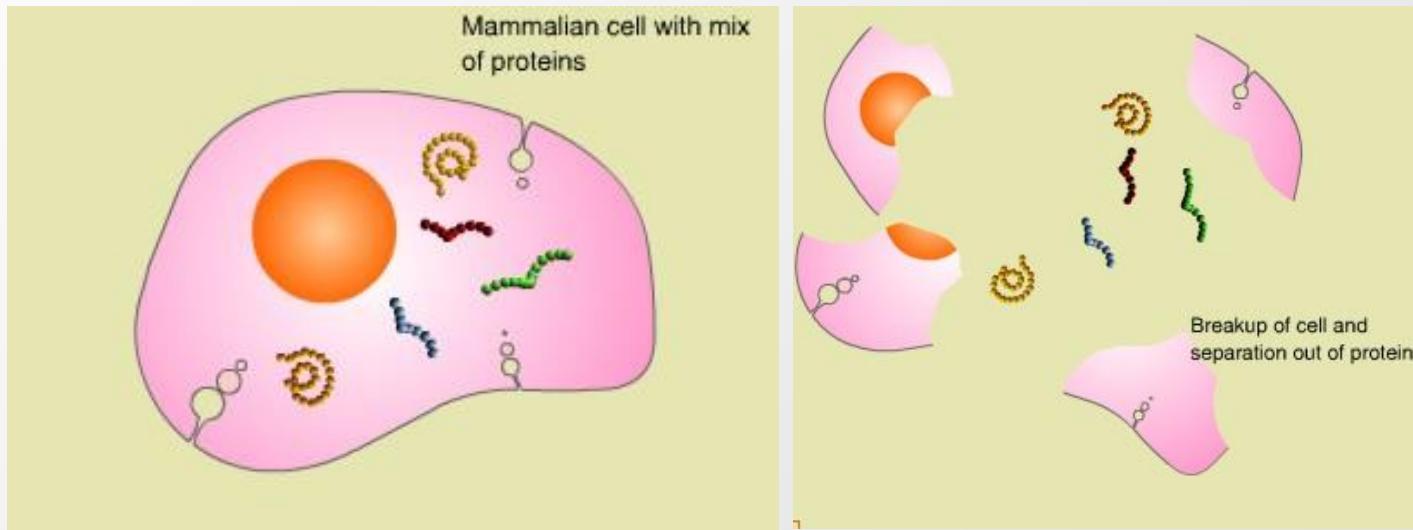
# Proteomic experiment

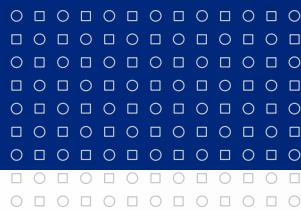




## HOMOGENIZATION

- mechanical
- ultrasound
- pressure
- freeze/thaw lysis
- detergent lysis





Watch for keratins!

## SAMPLE PREPARATION

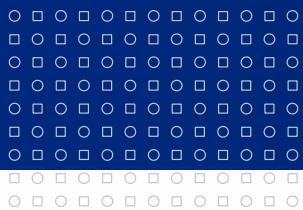
**Solubilization**      urea      thiourea      detergents

<b>Reduction</b>	DTT dithiotreitol	TBP tributylphosphine	TCEP Tris (2-carboxyethyl) phosphine hydrochloride
------------------	----------------------	--------------------------	--

**Inhibition**      of proteases, phosphatases glycosylases

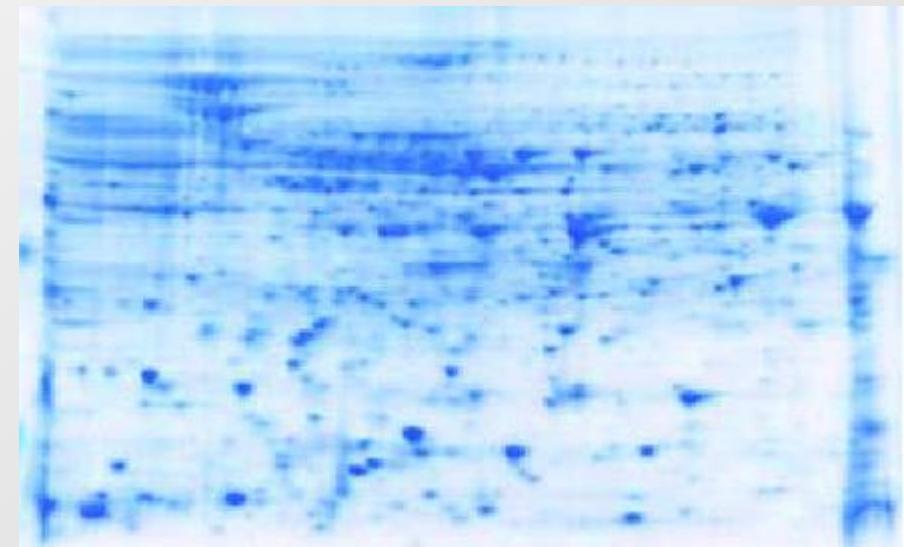
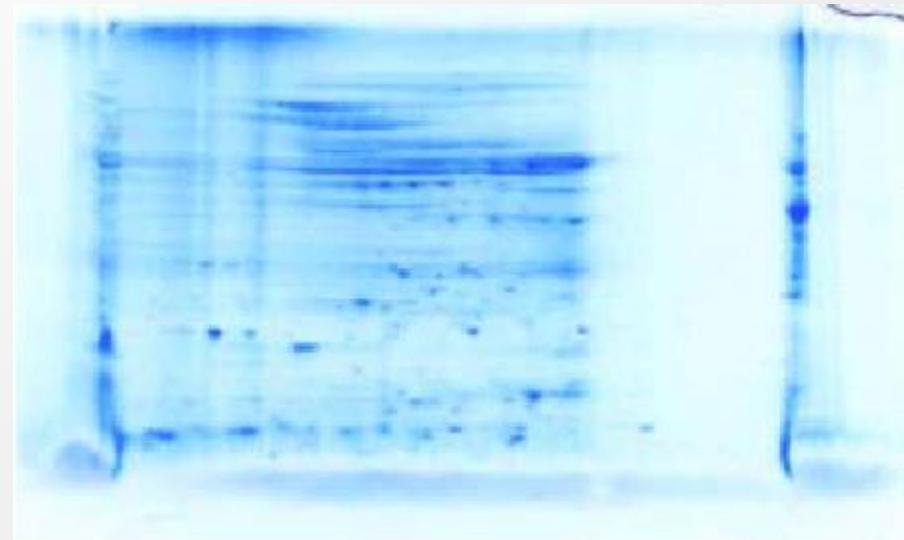
**Contaminants removal**

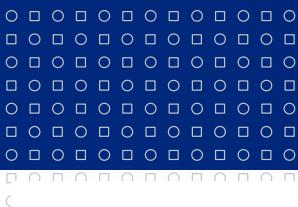
Protein assay



## DETERGENTS

- no net charge
  - 0.5 – 4%
  - working in high urea
- 
- non ionogenic
  - zwitterion
- 
- SDS only up to 0.25%





## C7BzO

3-(4-Heptyl)phenyl-3-hydroxypropyl)dimethylammoniopropanesulfonate

## CHAPS

3-[(3-Cholamidopropyl)dimethylammonio]-1-propanesulfonate hydrate

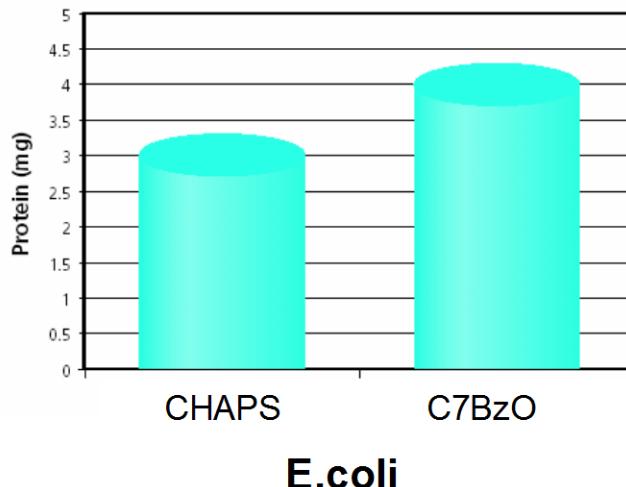
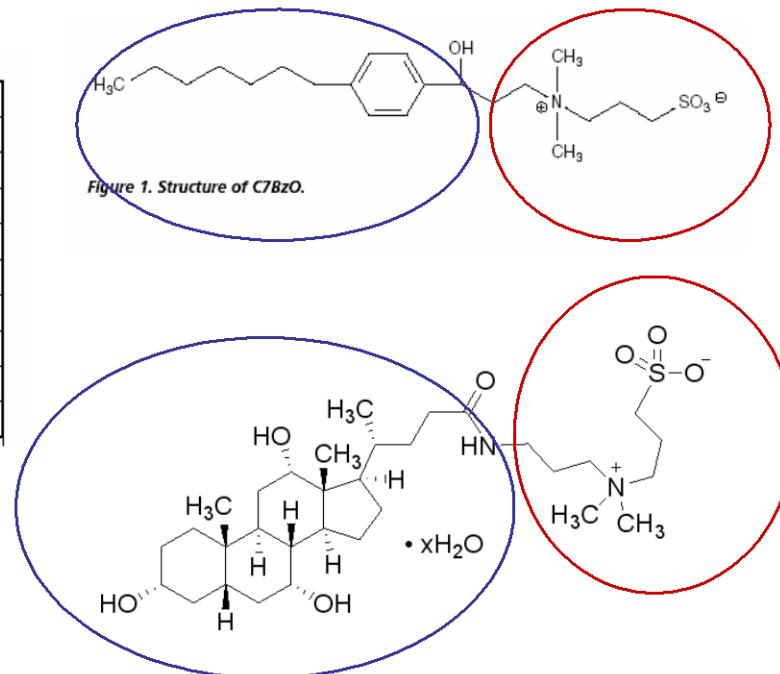
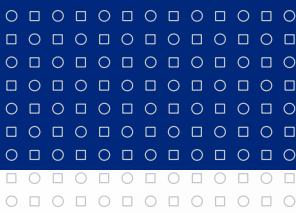


Figure 1. Structure of C7BzO.

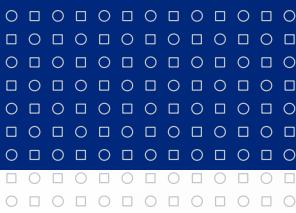




## RULE OF THUMB

- avoid proteolysis
- simple preparation
- fresh reagents
- fresh sample
- remove particles - spin
- remove contaminants

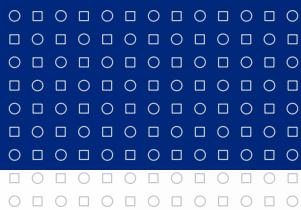




# CONTAMINANTS

- salts, buffers
- small molecules
- ionic detergents
- nucleic acids
- polysaccharides
- lipids
- phenols

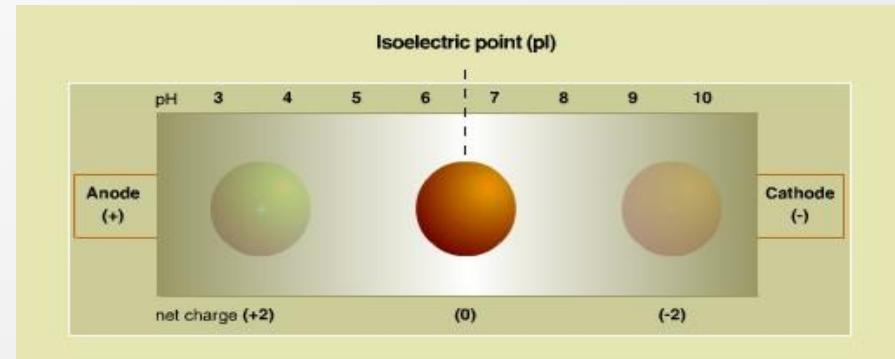




## 2-DE

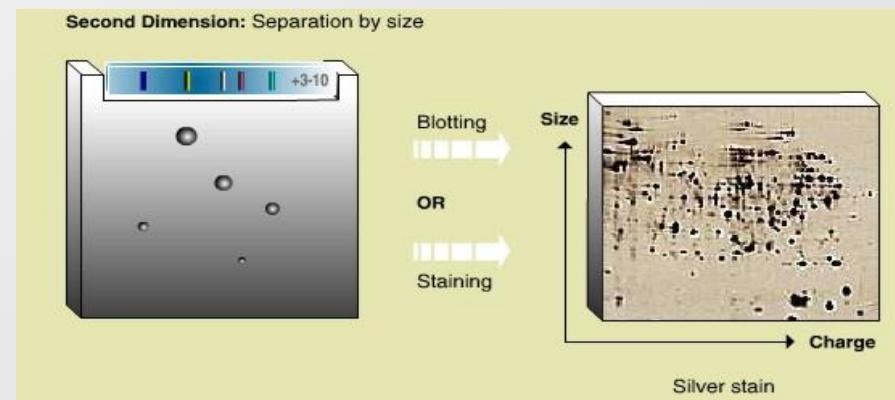
- first dimension

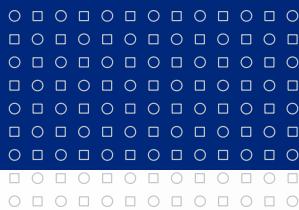
### IEF



- second dimension

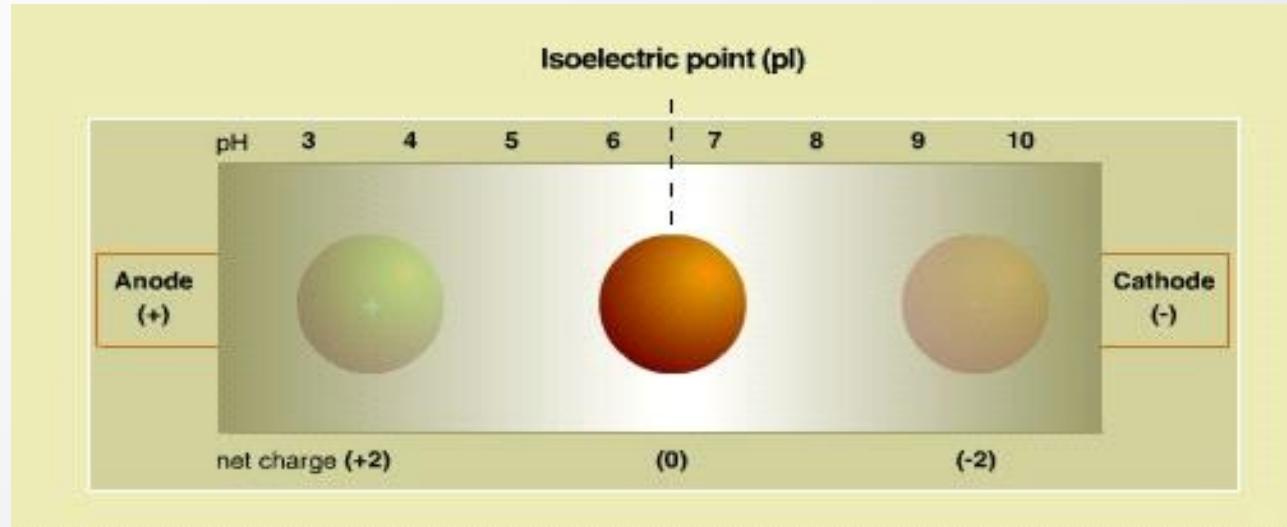
### SDS-PAGE

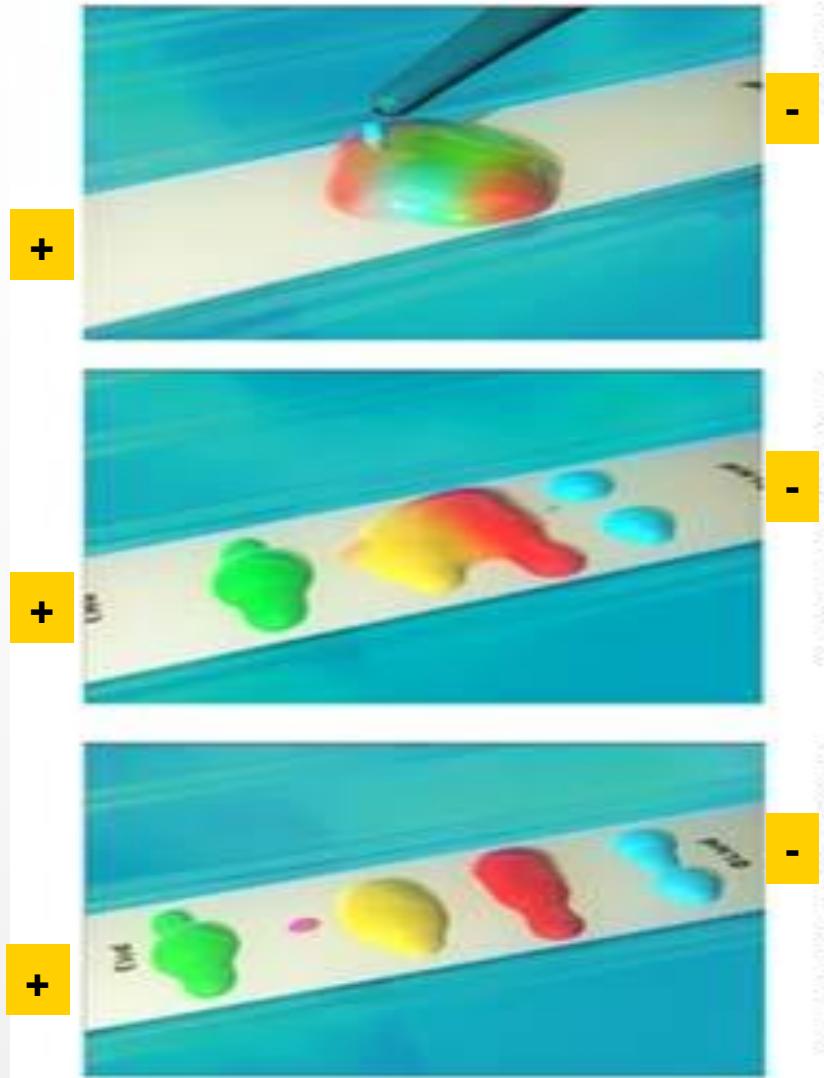
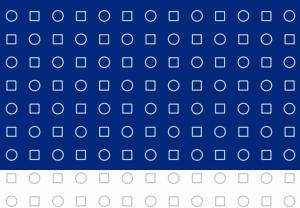




## 1<sup>st</sup> dimension ISOELECTRIC FOCUSING

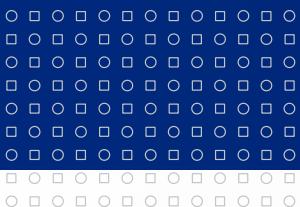
migration of charged molecules in pH gradient in electric field



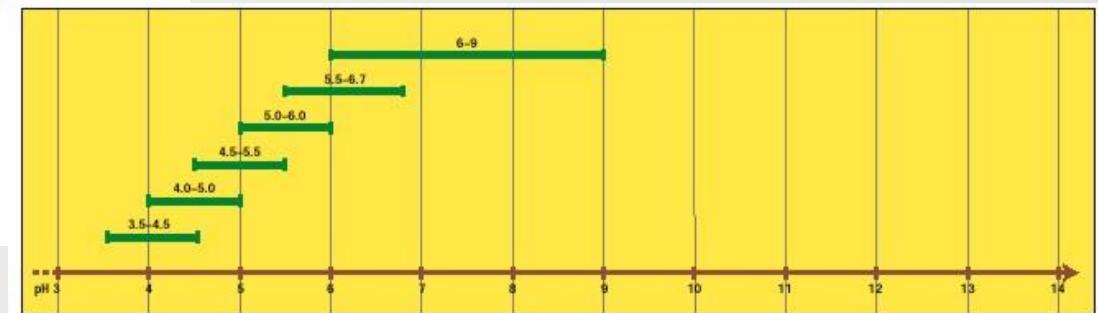


## ISOELECTRIC FOCUSING

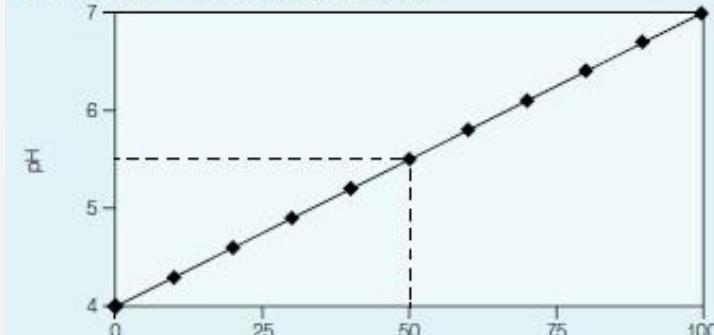
- immobilized pH gradient
- ampholytes



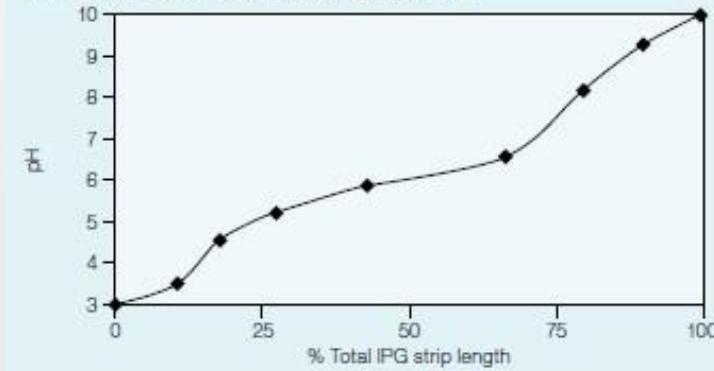
## RANGE OF STRIP SIZE OF STRIP



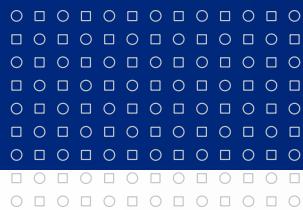
A. Linear pH 4-7 ReadyStrip IPG strip



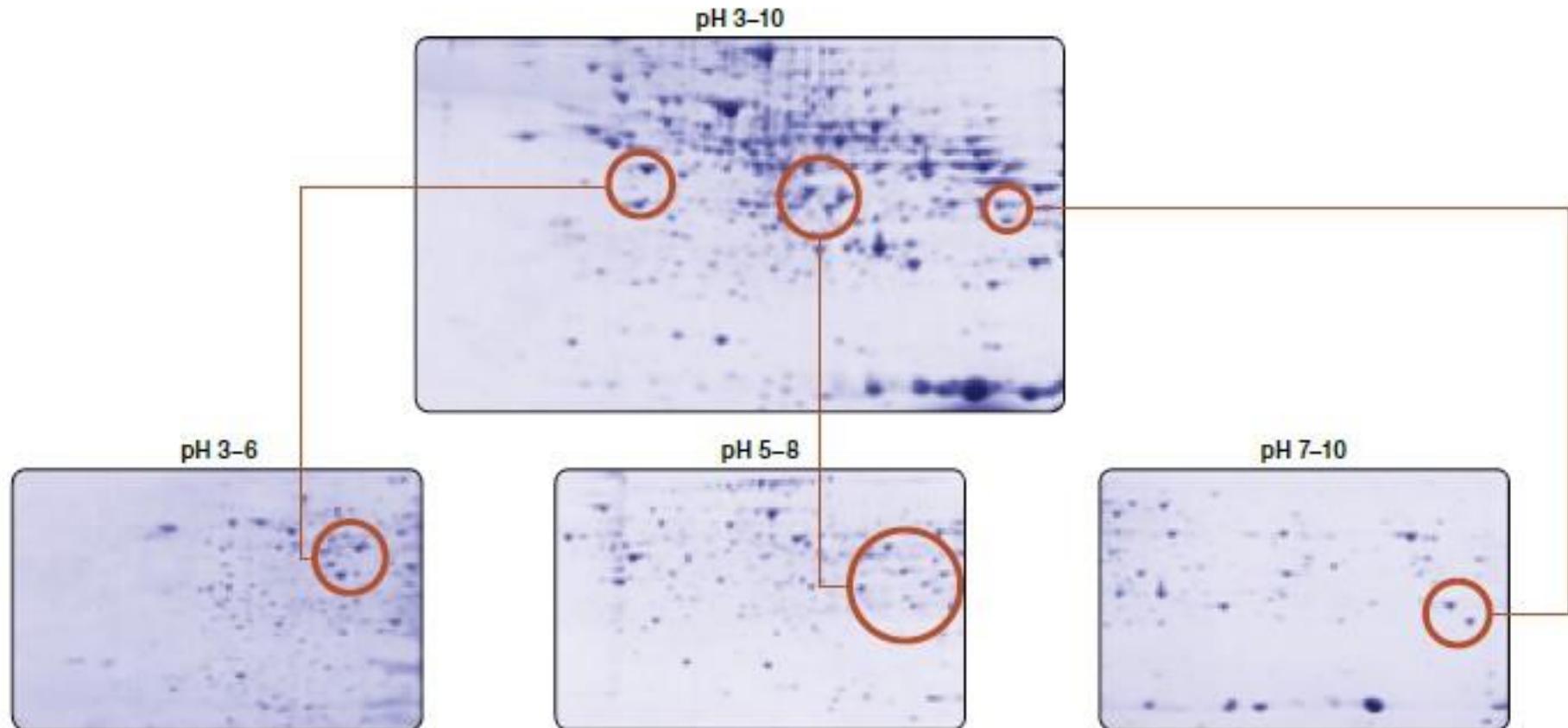
B. Nonlinear pH 3-10 ReadyStrip IPG strip

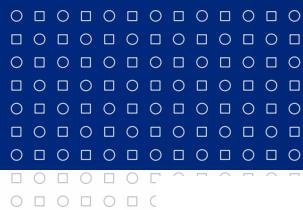


!



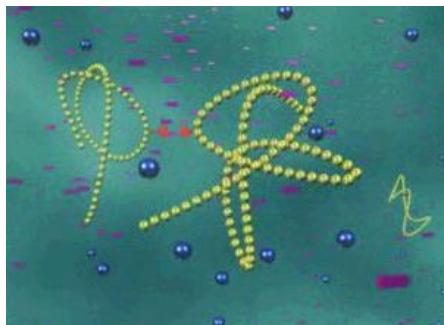
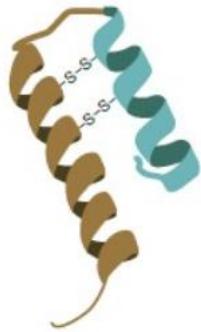
## RANGE OF STRIP



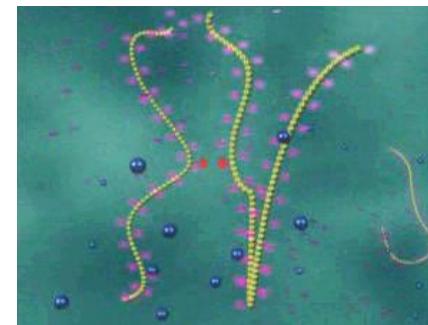


!

## EQUILIBRATION OF STRIP



denaturation SDS •

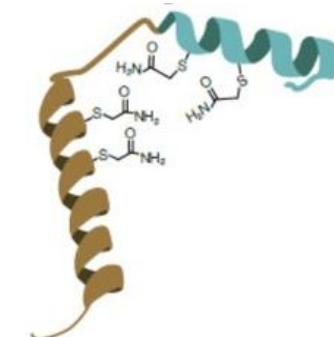


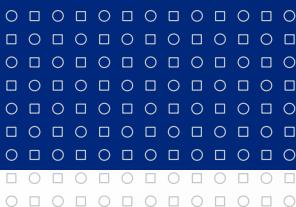
reduction

DTT •

alkylation

IAA •

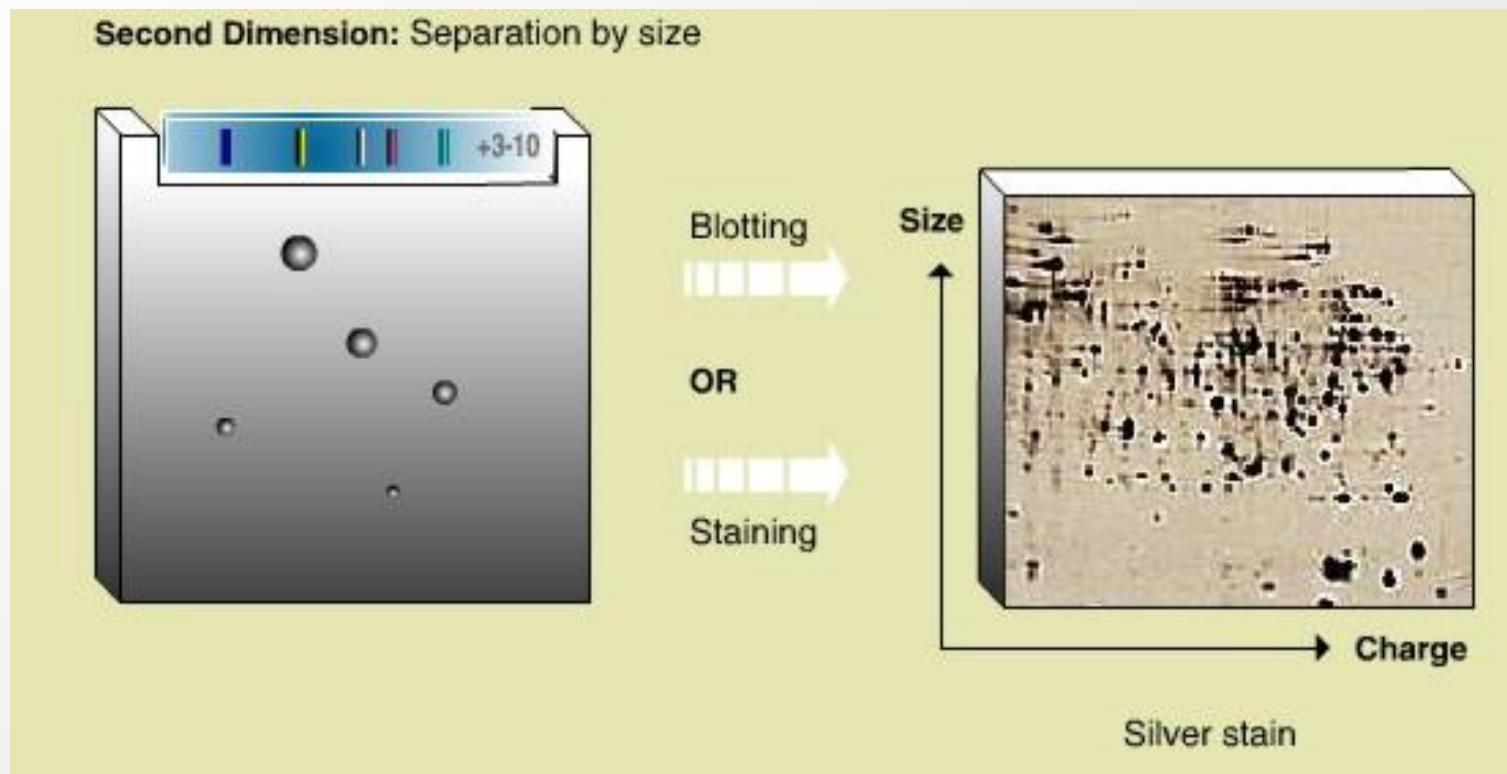


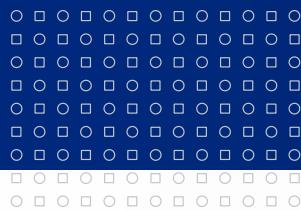


## 2<sup>nd</sup> dimension SDS-PAGE

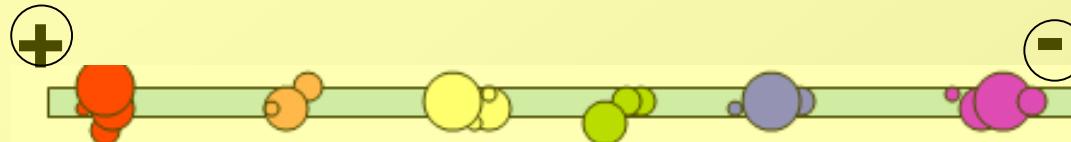
!

### Migration of anions in electric field according to MW

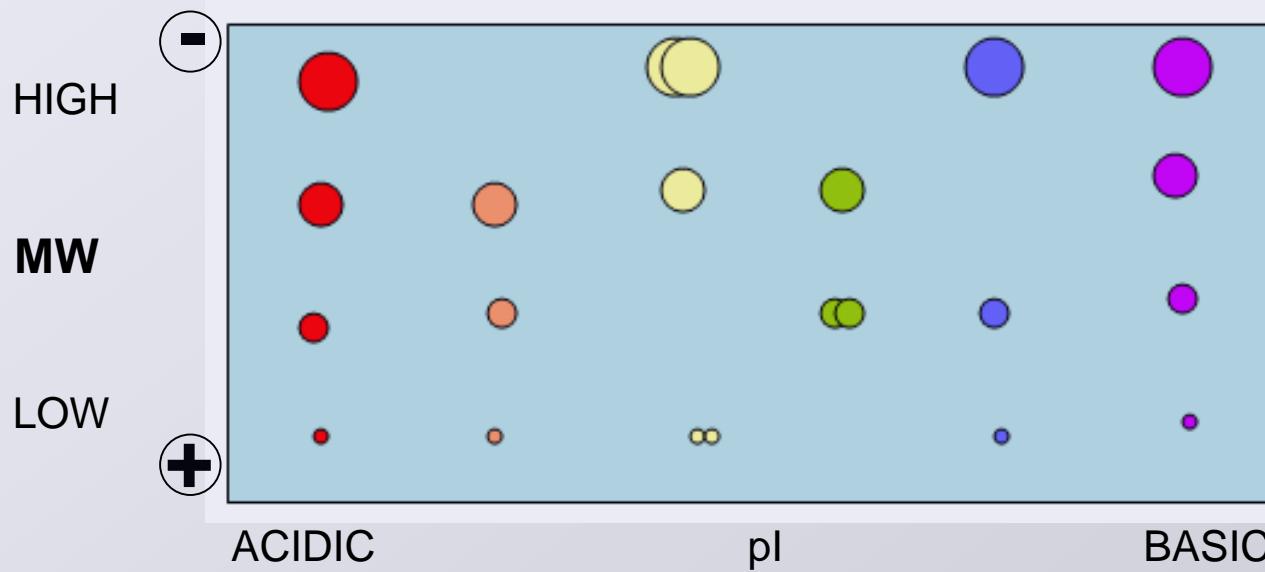


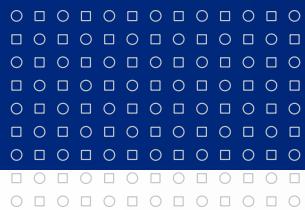


!

**FOCUSING****STRIP****SDS-PAGE**

equilibration

**GEL****Gel orientation**



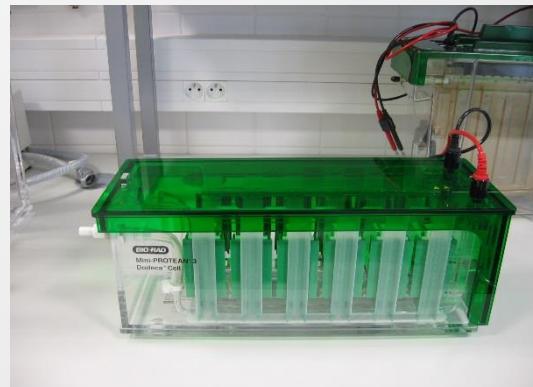
## 2-DE INSTRUMENTATION

- Protean IEF
- Protean Dodeca Cell
- Densitometer GS-800
- FLA-7000, STORM

*PDQuest, Quantity One*



Protean Plus Dodeca Cell

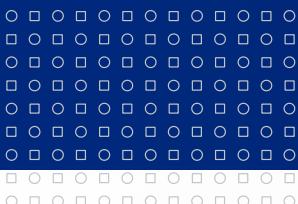


Mini-Protean 3 Dodeca Cell



Protean II xi Cell

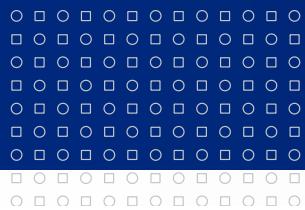




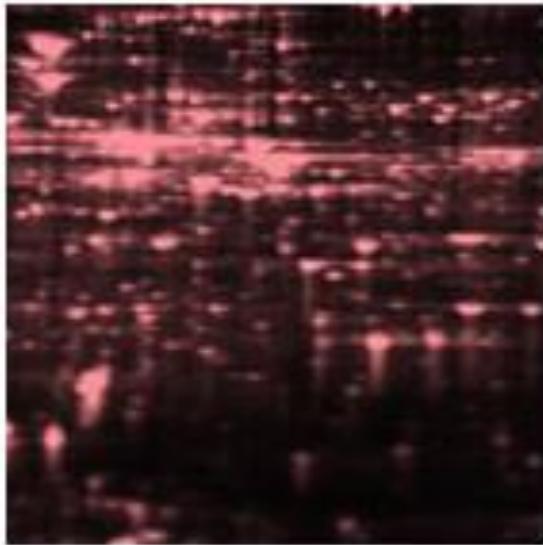
## PROTEIN DETECTION

- gel x blot
- visualisation → staining
  - radioactivity assay
  - immunodetection
- staining in gel
  - post-electrophoretic
  - pre-electrophoretic
- protein specific
- PTM specific
- visible spectrum
- fluorescence



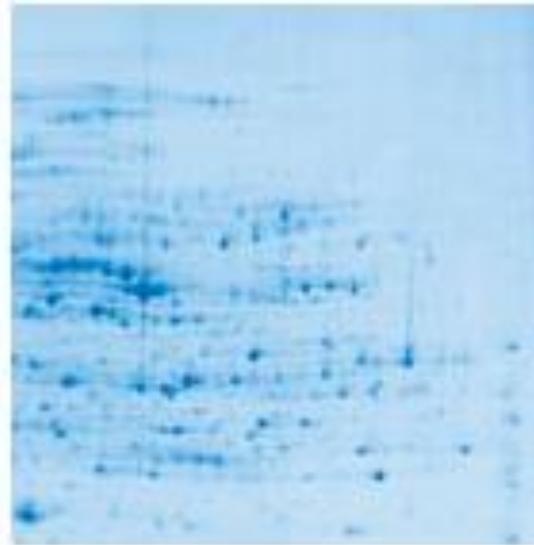


## PROTEIN DETECTION IN GEL



**Sypro Ruby**

**1.4 ng**



**Coomassie**

**36 ng**



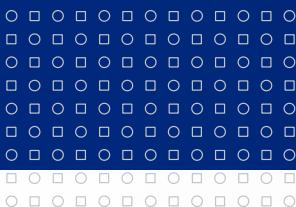
**silver**

**0.6 ng**

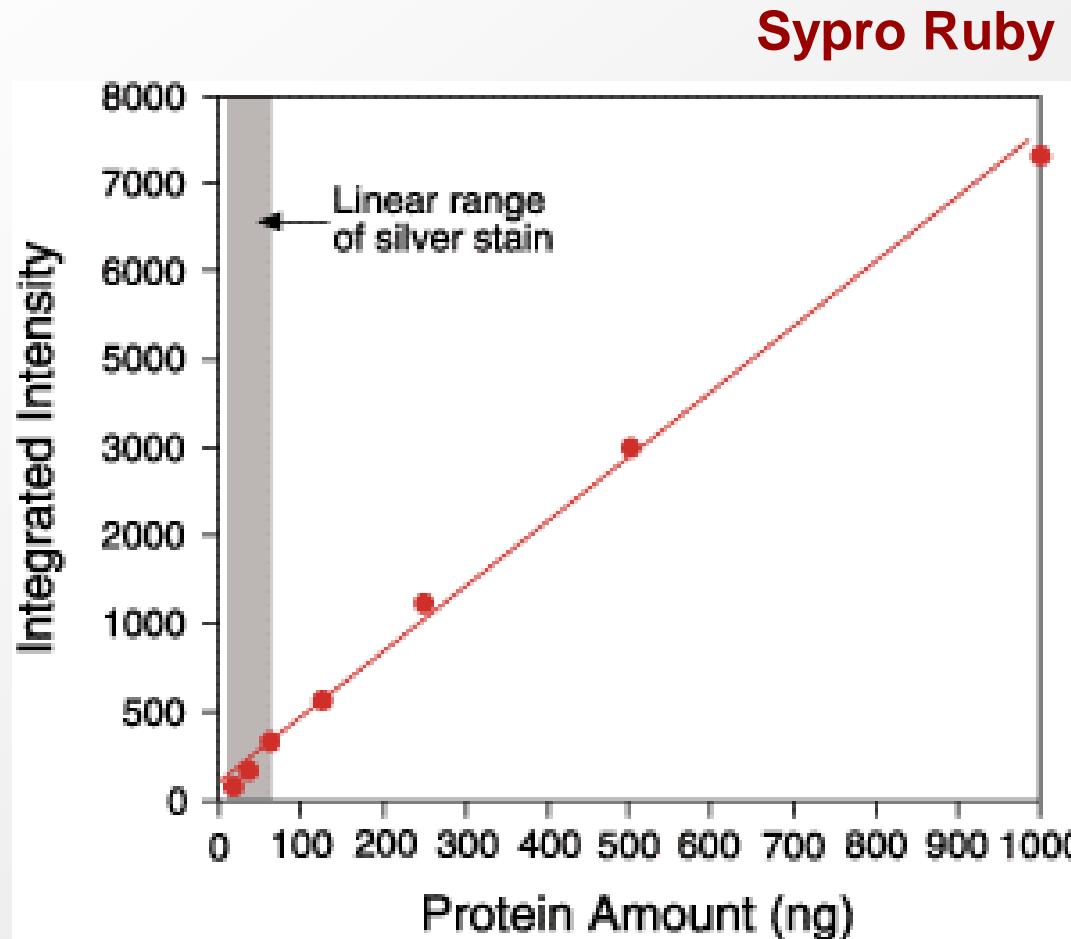
**PTM specific staining**

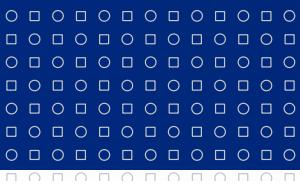
*Pro-Q Diamond*

*Pro-Q Emerald*

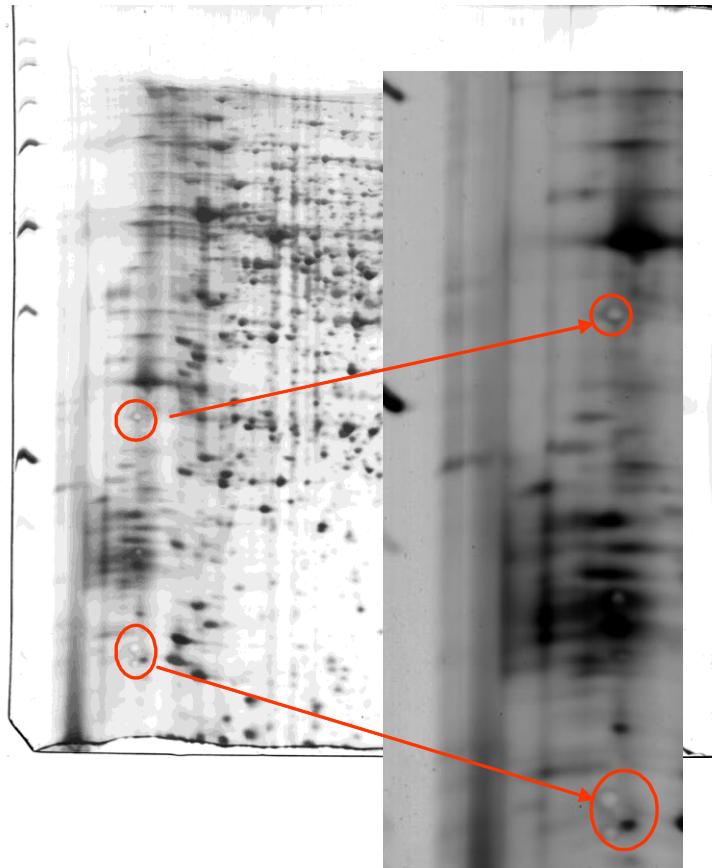


## PROTEIN STAINING – LINEARITY

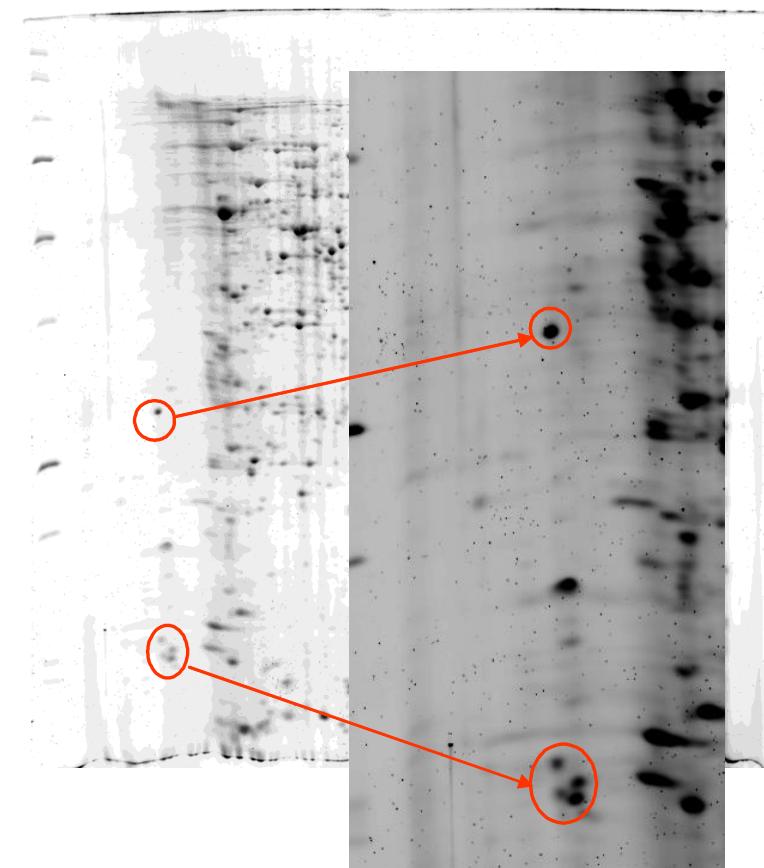


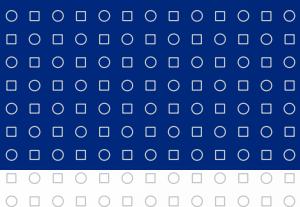


**Ag**



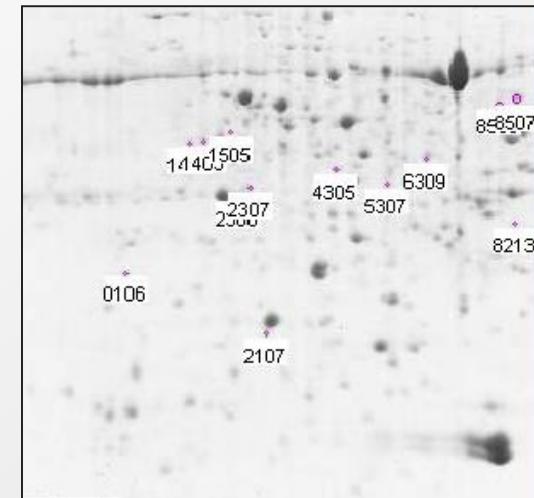
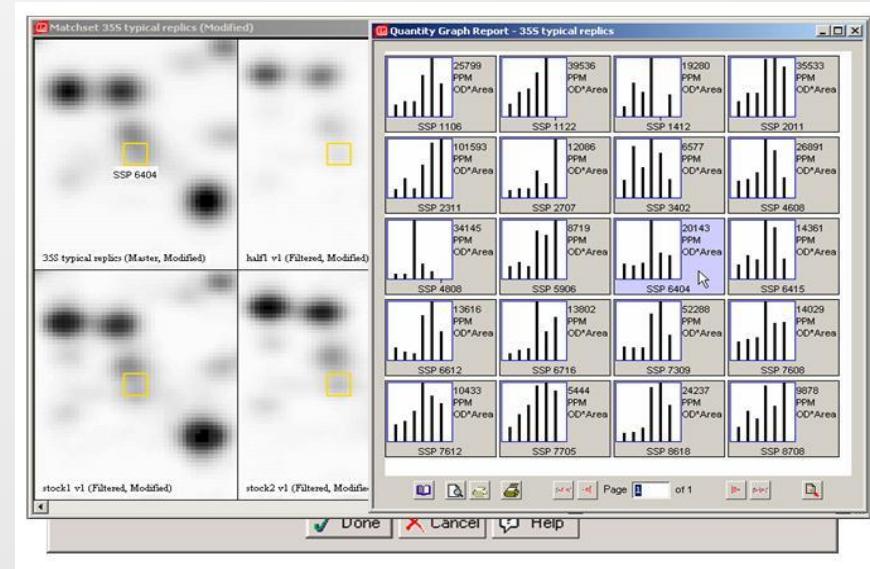
**Sypro Ruby**

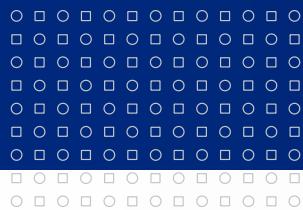




## IMAGE ANALYSIS

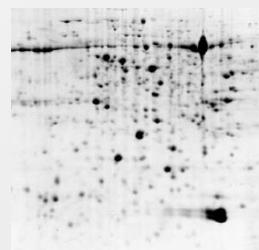
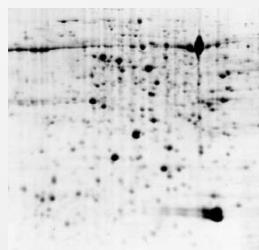
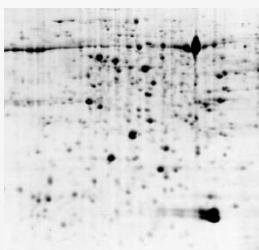
- quality
- quantity





## biological variability

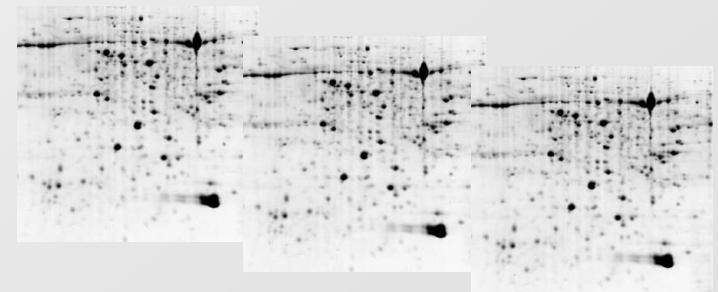
three plantelets analysed  
separately under same conditions



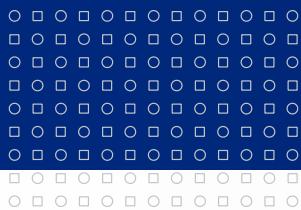
biological replicates

## technical variability

same plantelet analysed  
three times under same conditions



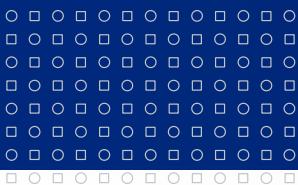
technical replicates



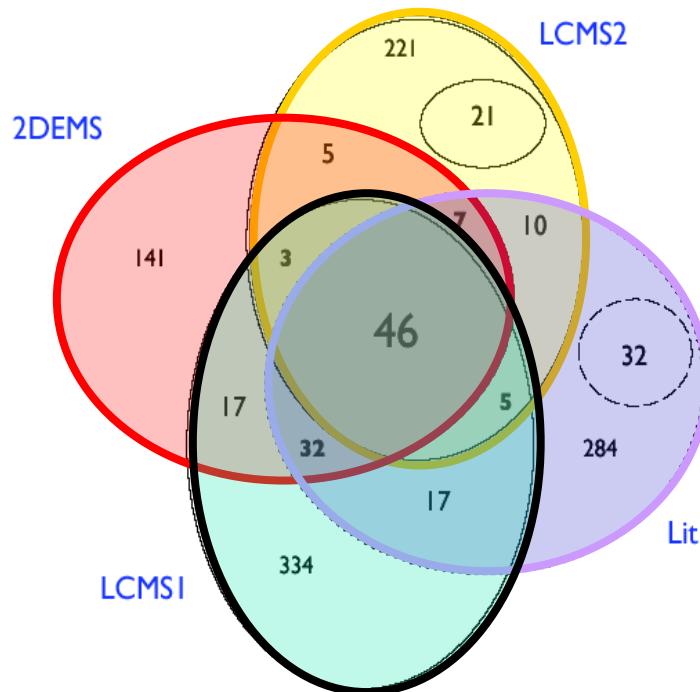
## 2D or not 2D ?

- visual aspects
- reproducibility
- dynamic range
- extreme proteins (membrane, basic...)
- difficult automatization
- postdigestion extraction



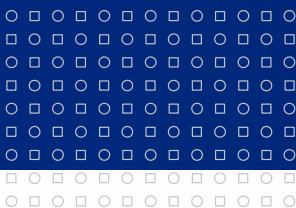


## Different Platforms See Different Plasma Proteomes: Small Overlap of Four Plasma Proteome Datasets (Number of NR proteins)



- 46 proteins in all four lists
- 195 proteins in 2 or more lists
- 1175 NR proteins total

From: The Human Plasma Proteome: A Non-Redundant List Developed by Combination of Four Separate Sources, N. L. Anderson et al, Molec. Cell Proteomics, 3: 311-326 (2004).



## MULTIDIMENSIONAL CHROMATOGRAPHY

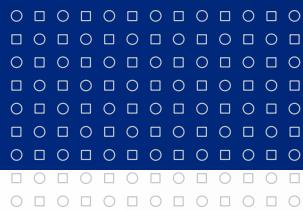
### FOR

- large sample volumes
- on-column concentration
- membrane proteins, basic proteins
- no staining
- peptides – going directly to MS
- automatization

### AGAINST

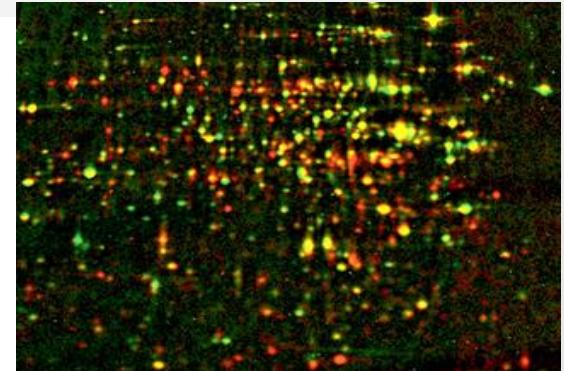
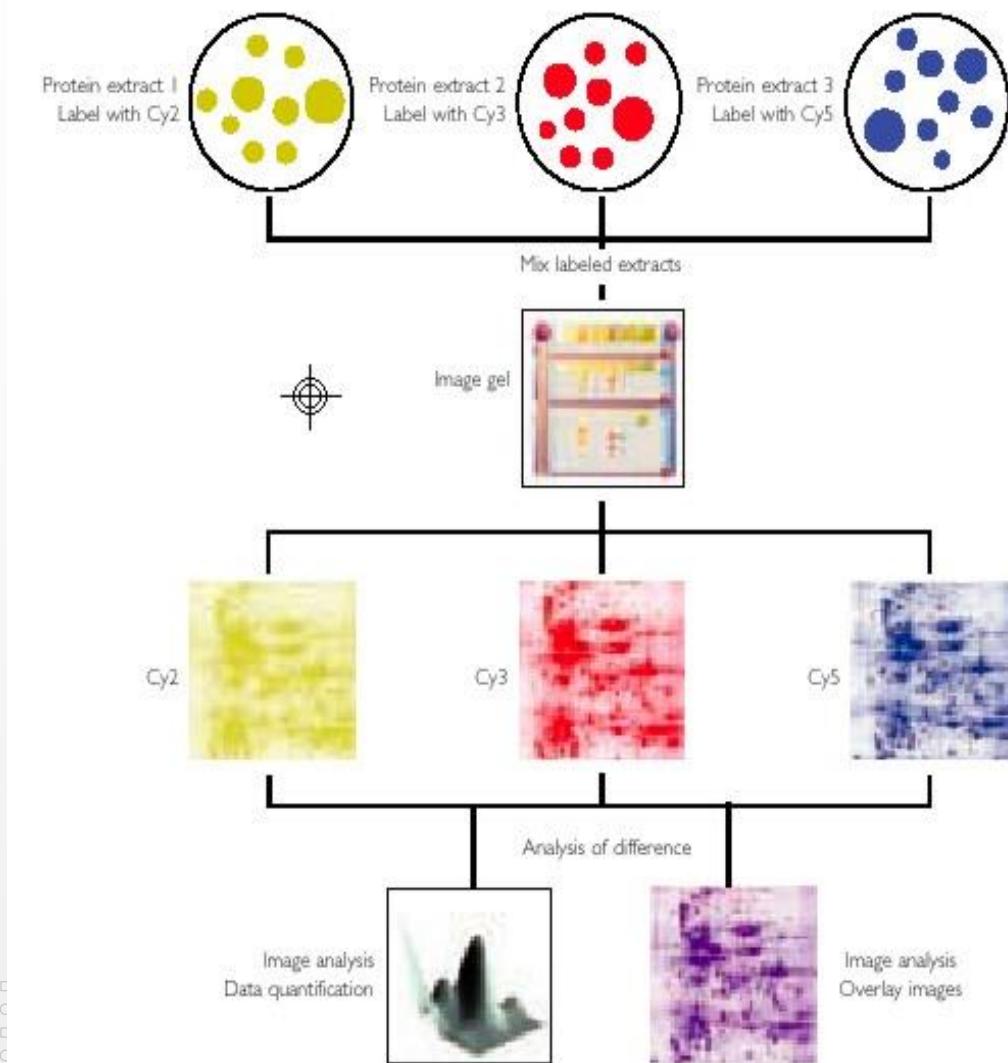
- vizual aspects lost: pl a Mr
- LC - serial analysis
- GE - more samples in parallel

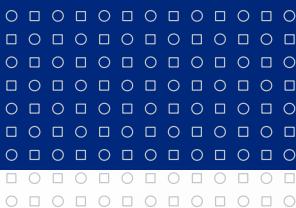




# Difference Gel Electrophoresis

# DIGE





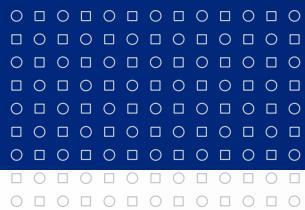
# BIOMARKERS

## ... NEEDLE IN HAYSTACKS

prefractionation ▪ separation ▪ identification ▪ control vs. sample

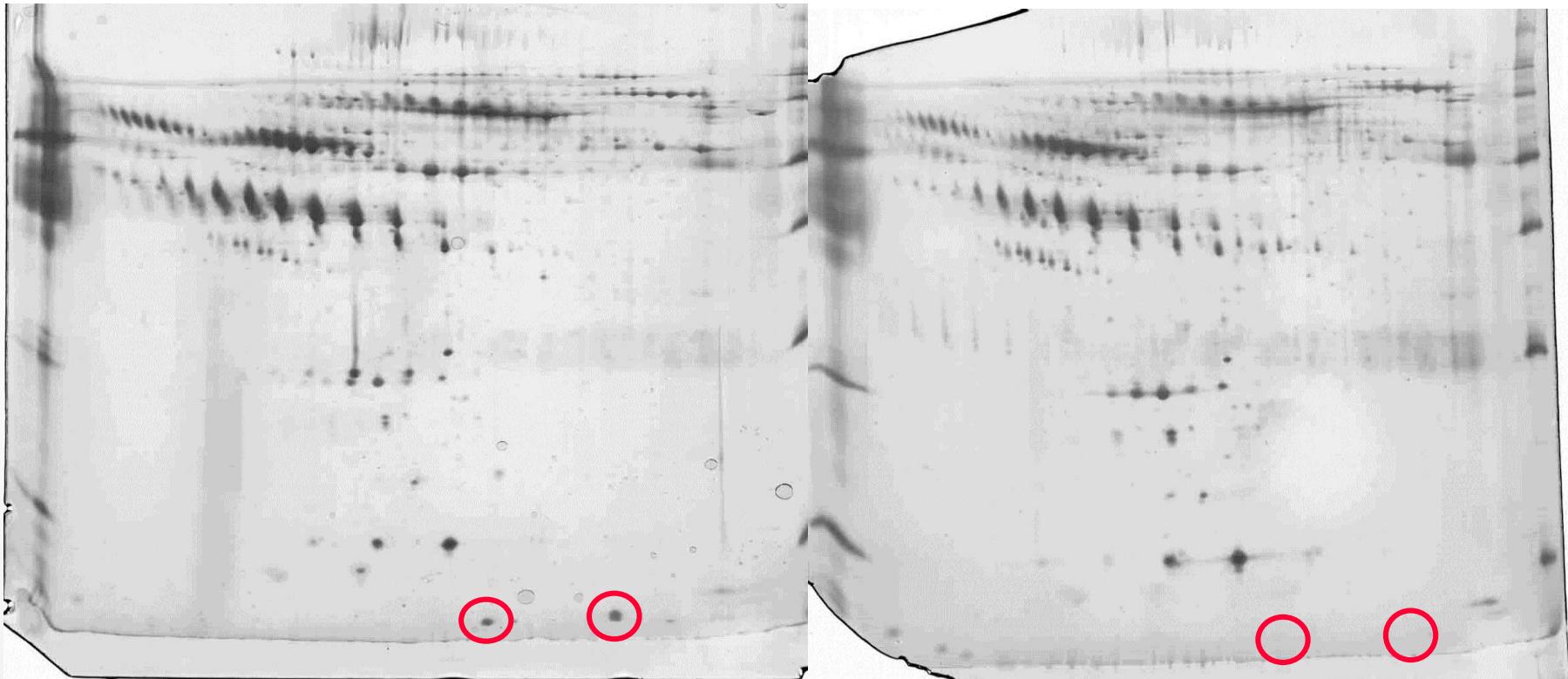
- **haystack** - proteins without relation to disease
- **needles** – disease specific proteins
- potential needles **difficult to validate** biological variability!
- are needles worth further examination?
- often contain **PTM**, difficult to be identified by MS



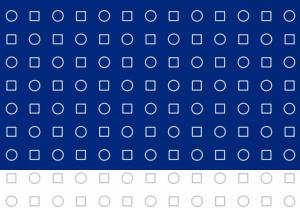


# Biomarkers in human plasma

Day 21 – before clinical manifestation



separation → identification

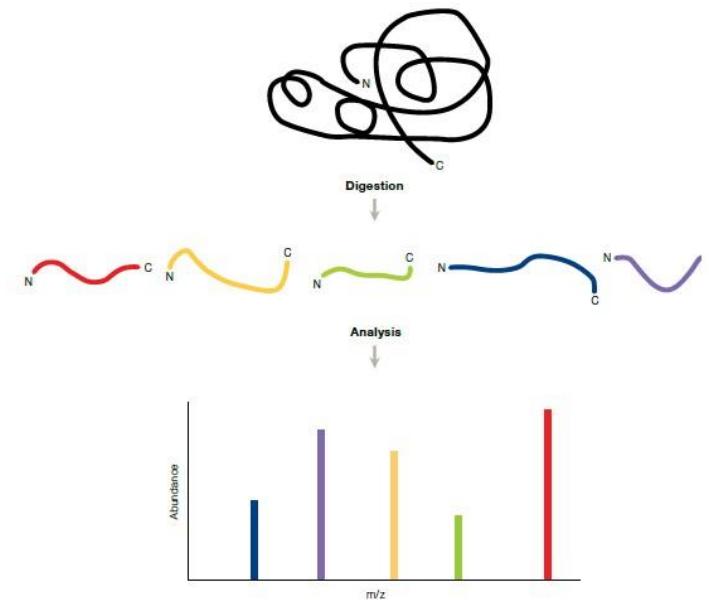


# ↓ DIGEST

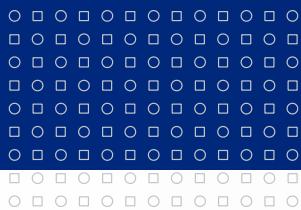
trypsin      Glu-C      Asp-N      thermolysin

MAVEPFPRRPITRPHASIEVDTSGTGGSAGSSEK**VF**  
**CLIGQAEGGEPTVYELRNYAQAKRLFRSGELLDAI**  
ELAWGSNPNTAGRILAMRIEDAKPASAEIGGLKIT  
SKIYGNVANNIQVGLEKNTLSDSLRLRVIFQDDRFN  
**EYVDNIGNIFTIKYKGEEANATFSVEHDEETQKASR**  
LVLKVGDQEVKSYDLTGGAYDYTNAlITDINQLPDF  
EAKLSPFGDKNLESSKLDKIENANIKDKAVYVKAVF  
GDLEKQTAYNGIVSFQLNAEGEVPSNVEVEAGEE  
SATVTATSPIKTIEPFELTKLKGGTNGEPPATWADKL  
**DKFAHEGGYYIVPLSSKQSVHAEVASFKERSDAGE**  
PMRAIVGGGFNESKEQLFGRQASLSNPRVSLVANS  
GTFVMDDGRK**NHVPAYMVAVALGGLASGLEIGES**  
ITFKPLRVSSLDQIYESIDLDELNENGIIISIEVRNR**TN**  
**TFFRIVDDVTTFNDKSDPVKAEMAVGEANDFLVSE**  
LKVQLEDQFIGTRTINTSASIICKDFIQSYLGR**KKRDN**  
EIQDFPAEDVQVIVEGNEARISM**TVYPIRSFKKISVS**  
**LVYKQQTLQA**

- IN-GEL
- IN-SOLUTION



MS

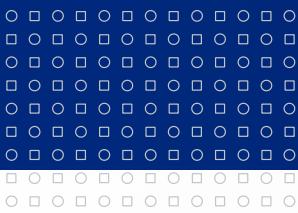


MASARYK UNIVERSITY

[www.muni.cz](http://www.muni.cz)

G I G O

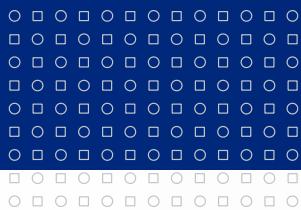




G I G O

GARBAGE IN - GARBAGE OUT

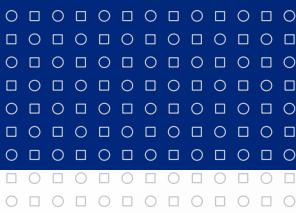




# LITERATURE

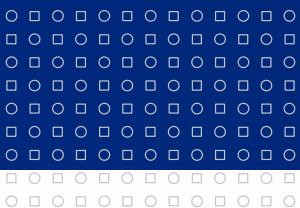
- R.M. Twyman: Principles of Proteomics
- R.Westermeier, T.Naven, H-R Höpker: Proteomics in Practice
- A.J.Link: 2D Proteome Analysis Protocols
- Current Protocols in Protein Science
- R.J.Simpson: Proteins and Proteomics
- T.Rabilloud: Proteome Research: Two-dimensional Gel Electrophoresis and Identification Methods
- A. Görg, W. Weiss, M.J.Dunn: Proteomics 2004, 4, 3665, rev.
- I. Miller, J. Crawford, E. Gianazza: Proteomics 2006, 6, rev.
- F.Chevalier: Proteome Science 2010, 8:23, review
- R. Burgess, M. Deutscher: Guide to Protein Purification





I. SEPARATION  
**II. PREFRACTIONATION**

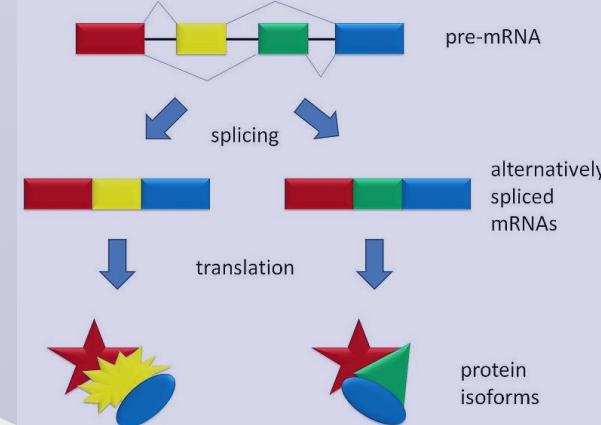




GENOME



PROTEOME

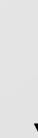


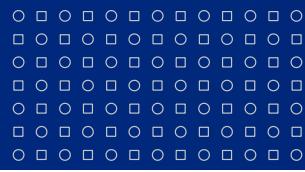
ISOFORMS

PTM ~200 variants (fosforylation, glykosylation, acylation, methylation...)

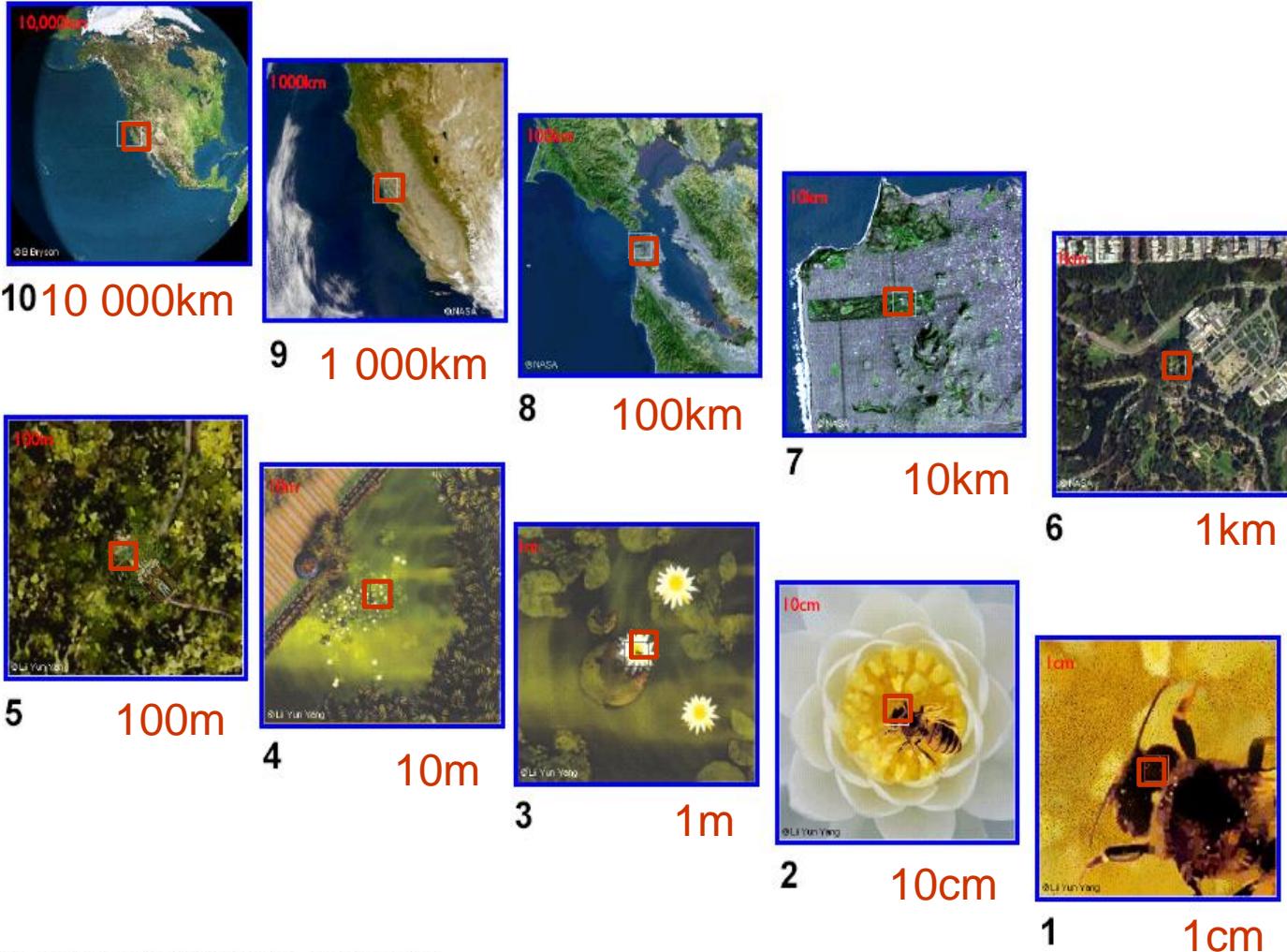
CONCENTRATION RANGE ~ 10 orders of magnitude

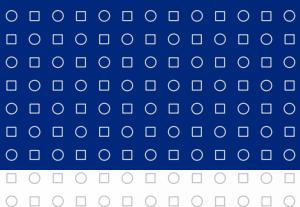
PREFRACTIONATION → MS



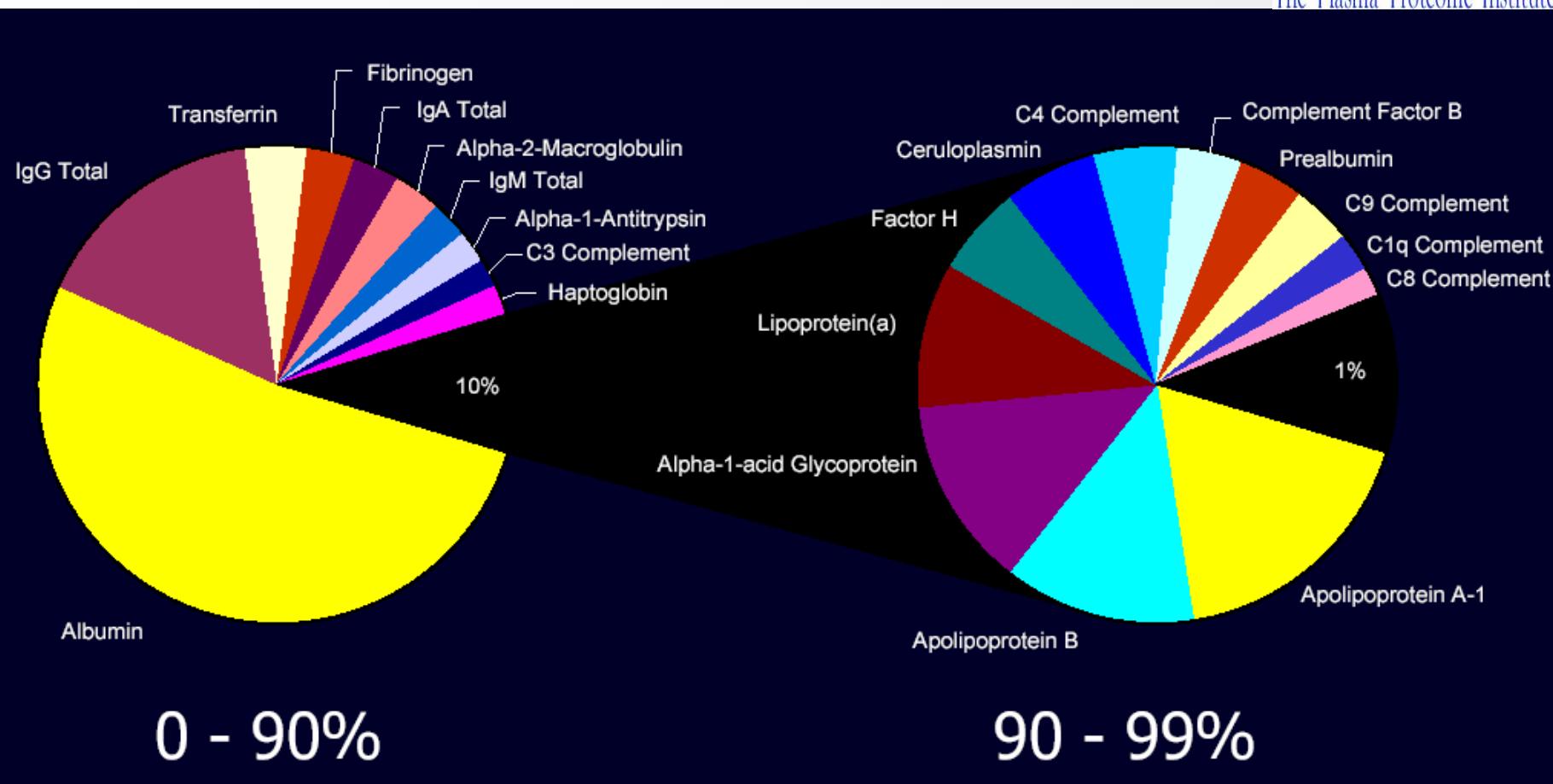


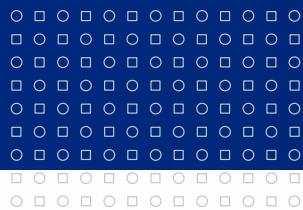
# $10^{10}$ Really Is Wide Dynamic Range





# Abundant proteins in human plasma



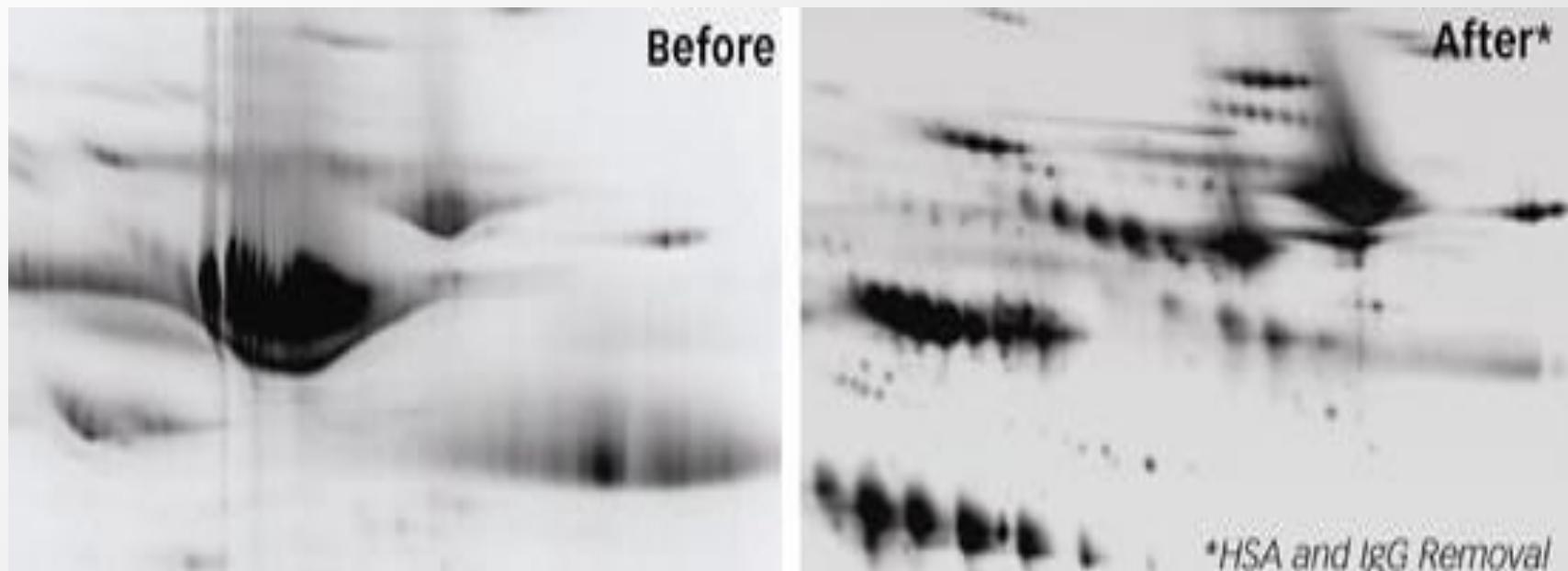


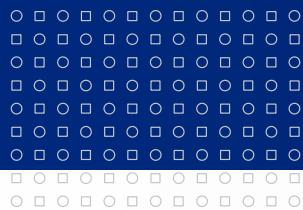
## AFFINITY DEPLETION

Removal of abundant proteins by affinity chromatography

**HSA**

**IgG**





## Human plasma – bound fractions after affinity depletion

ALBUMIN

IgG

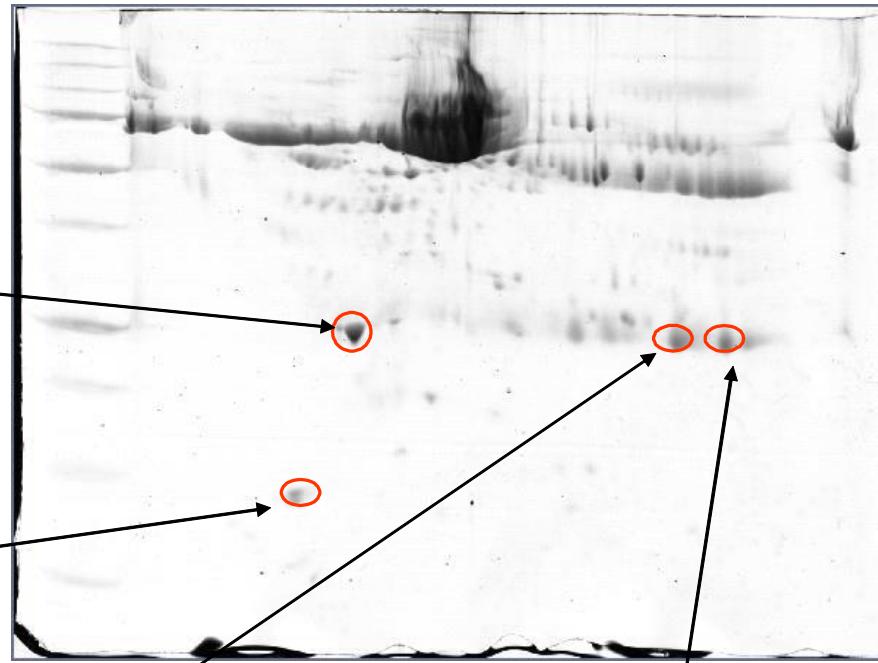
Staining CBB G-250

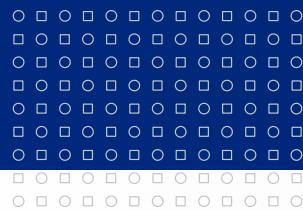
Apolipoprotein

albumin

Immunoglobulin kappa light chain

Immunoglobulin light chain



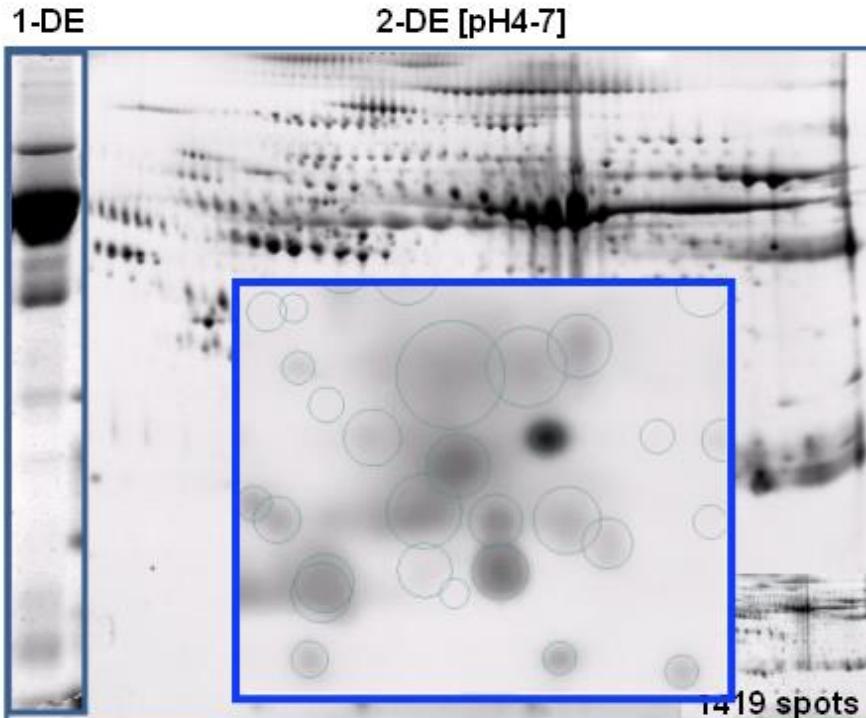


# CPPL

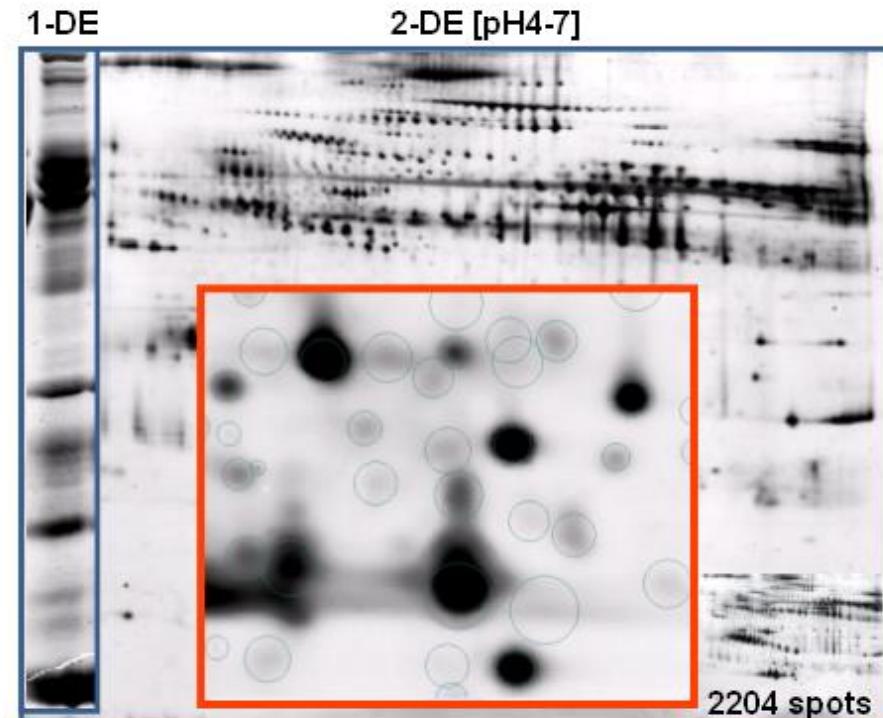
## Combinatorial Peptide Ligand Library

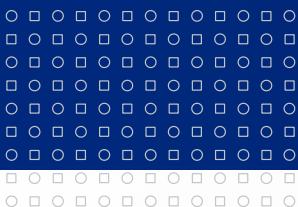


Native Human Serum



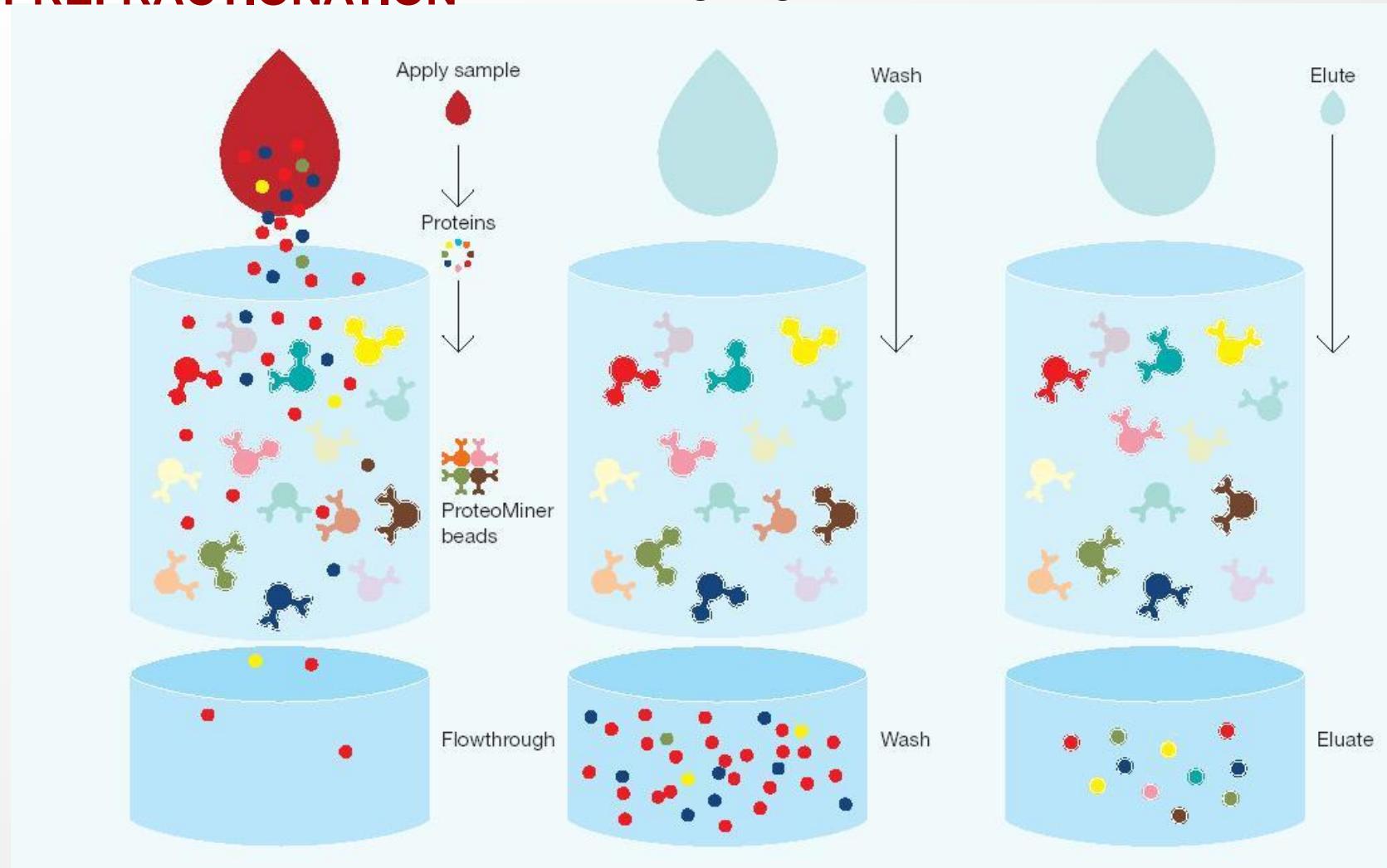
Human Serum Fractionated by ProteoMiner

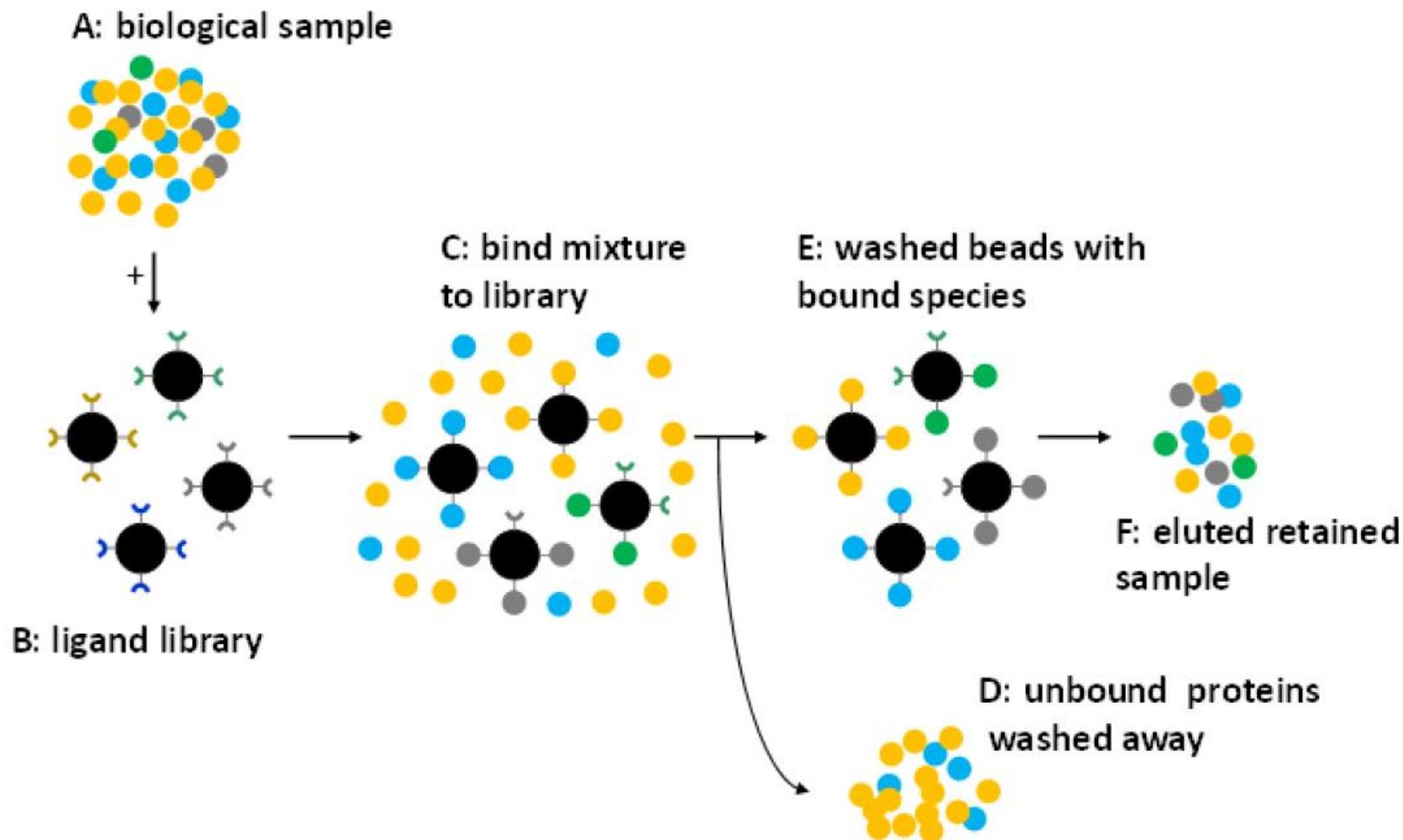
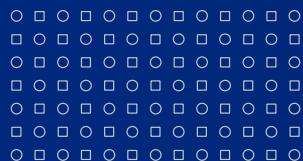


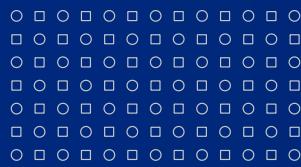


## PREFRACTIONATION

## PROTEOMINER







IEF

prefractionation



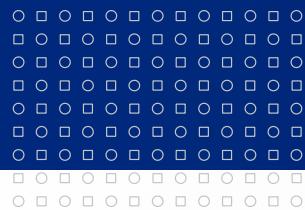
MicroRotofor

- prefractionation in solution

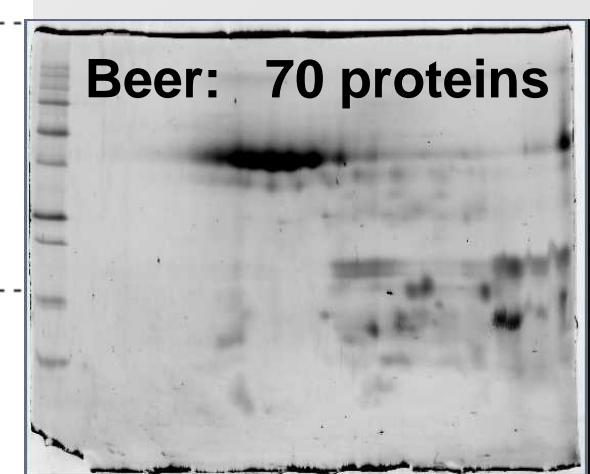
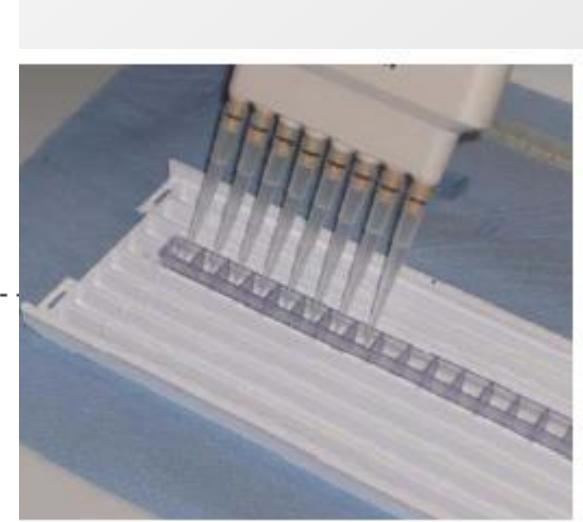
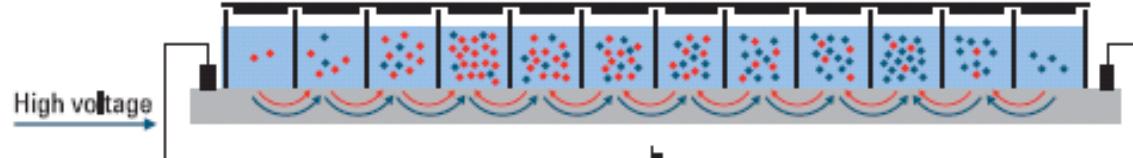
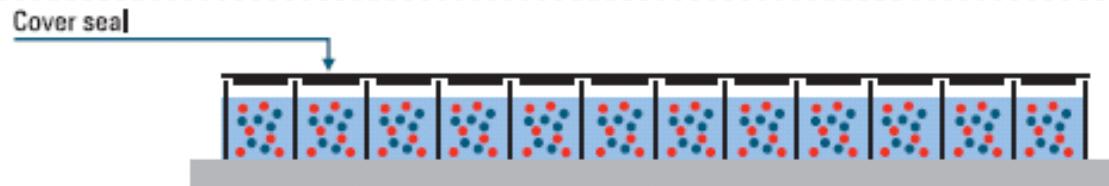
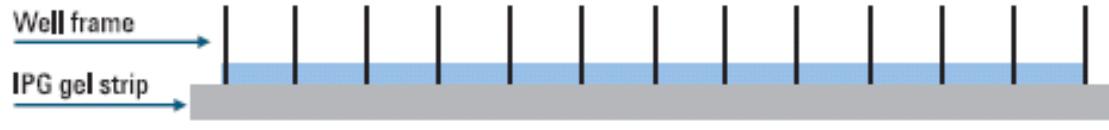


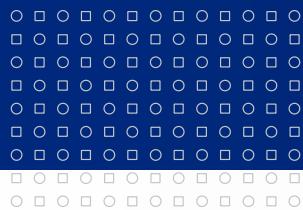
OffGel Fractionator

- prefractionation in solution using IPG strip

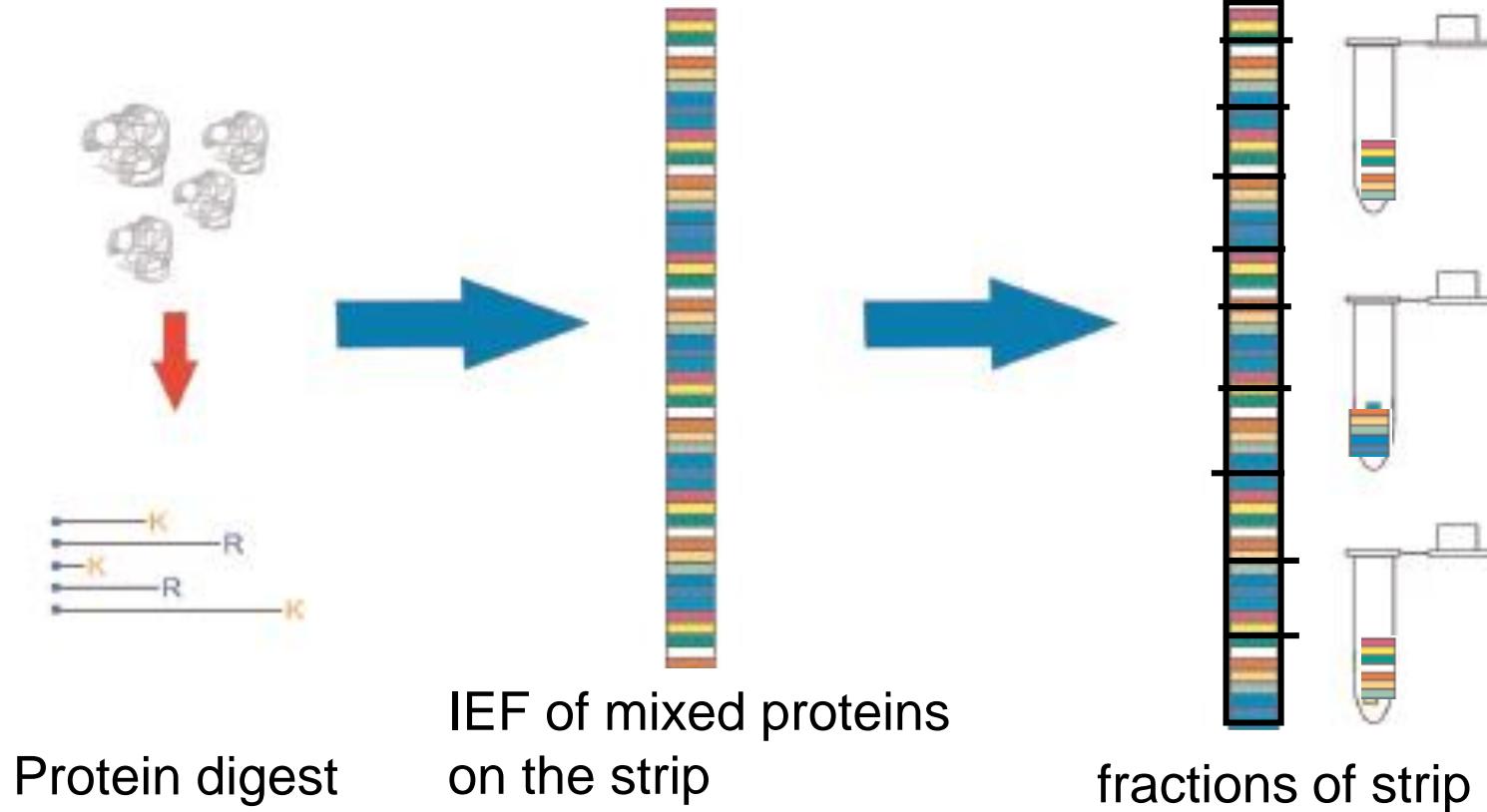


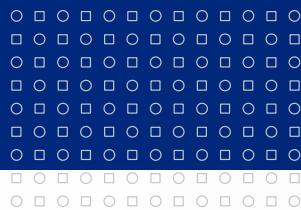
## OFFGEL IEF prefractionation of proteins or peptides





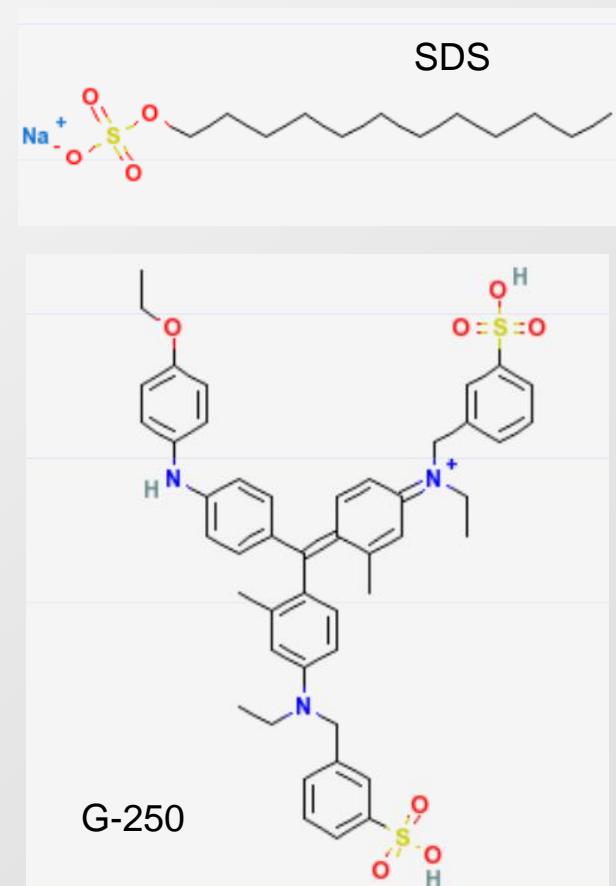
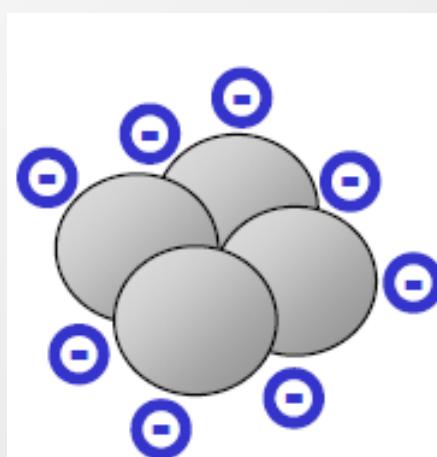
## IPG-IEF

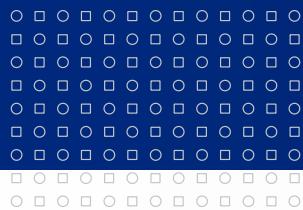




## Blue Native Electrophoresis BNE

- Separation of native proteins
- Separation of membrane complexes
- Solubilization by non-ionic detergents
- Charged by Coomassie G-250
- BN PAGE gel (strip/band) as 2D

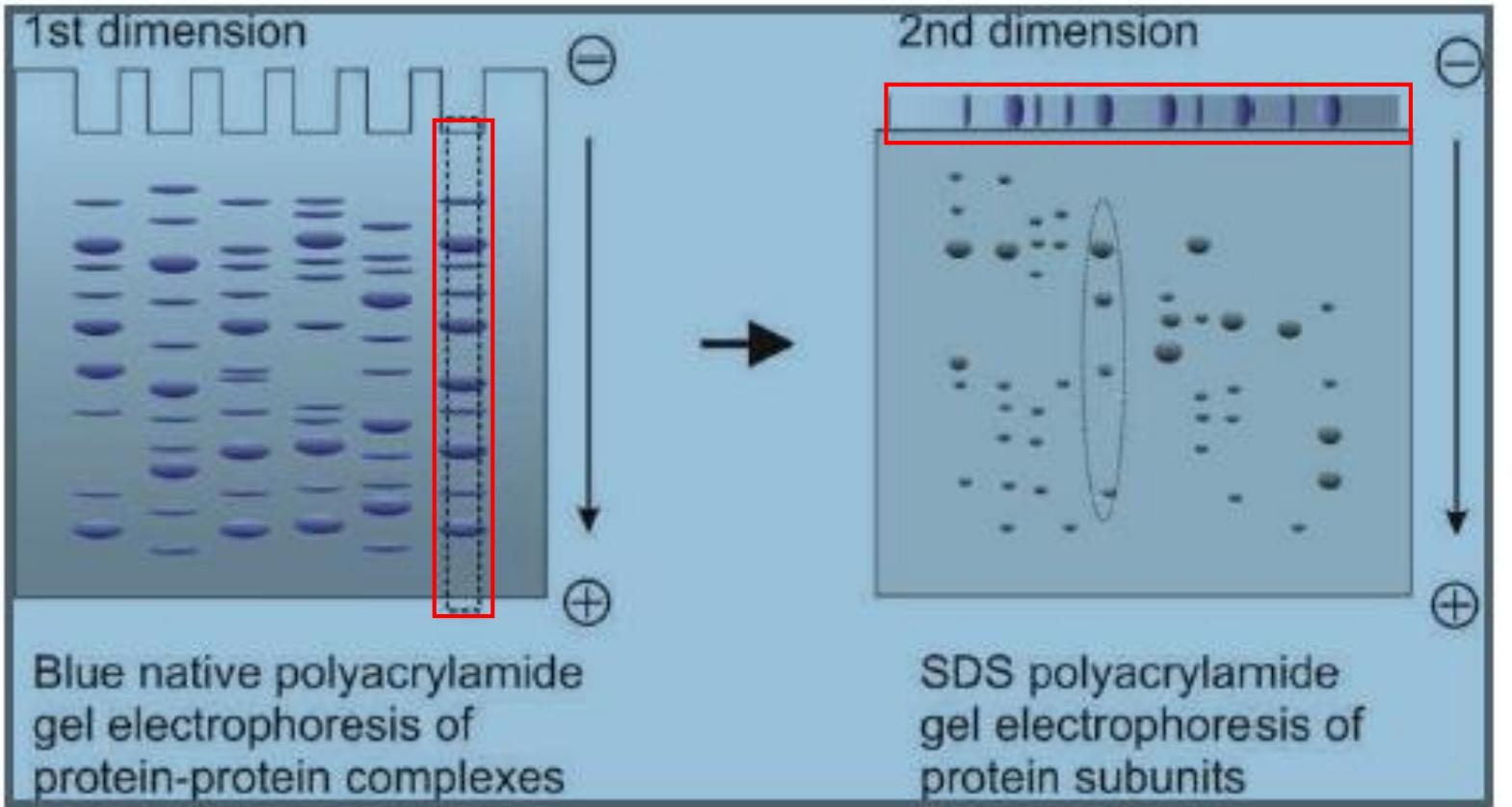


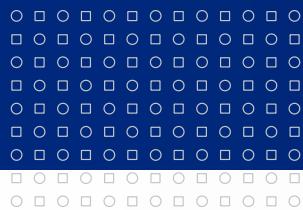


2DE

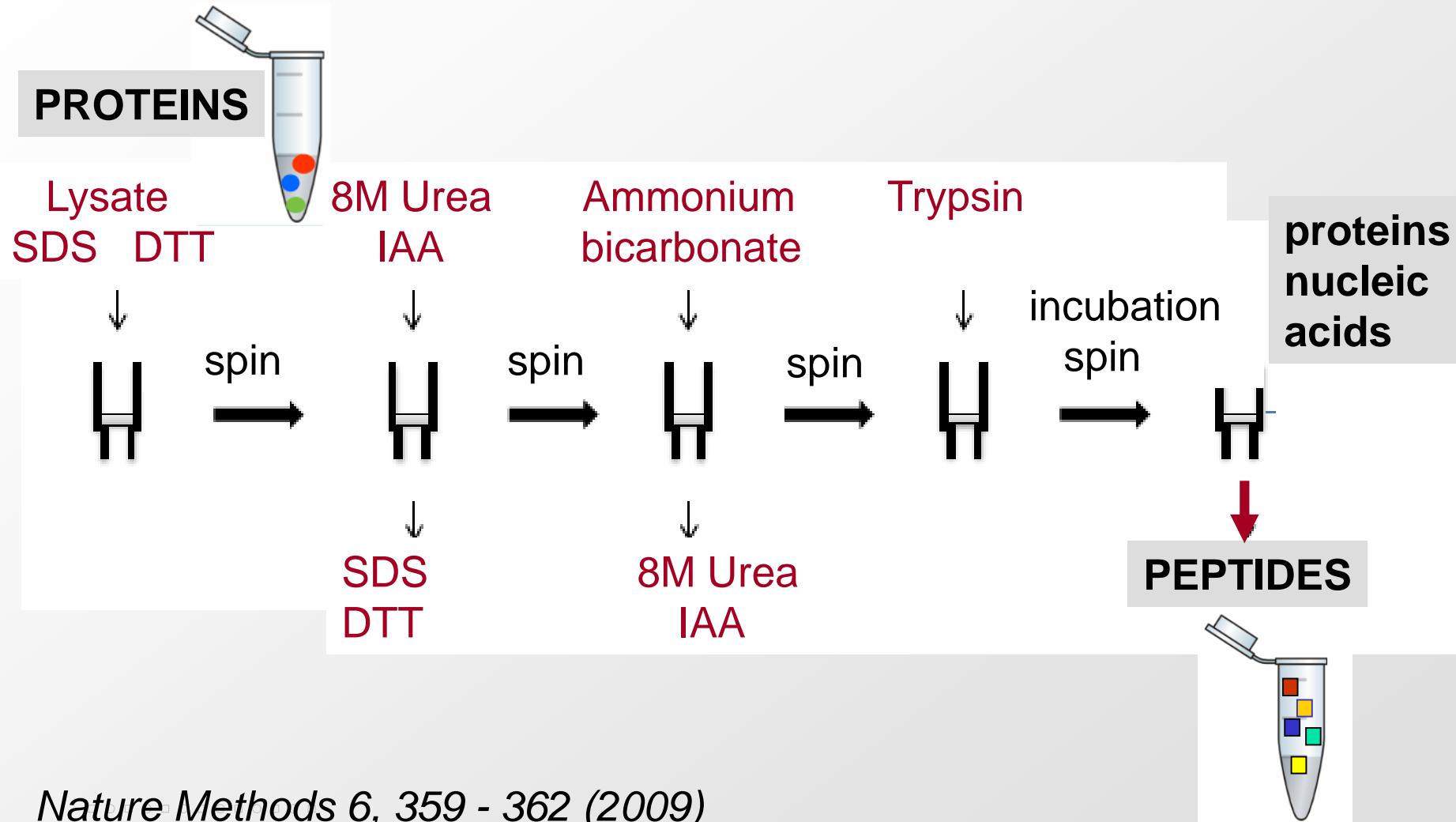
BNE

SDS-PAGE

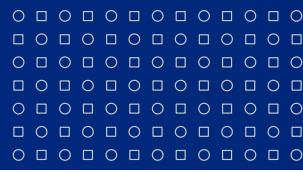




## FASP Filter aided sample preparation

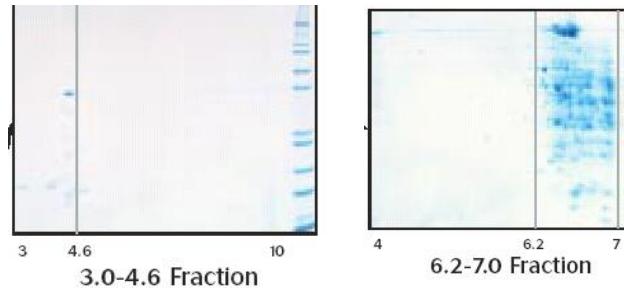


Nature Methods 6, 359 - 362 (2009)

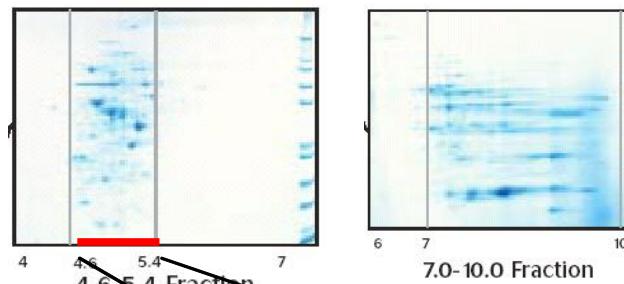
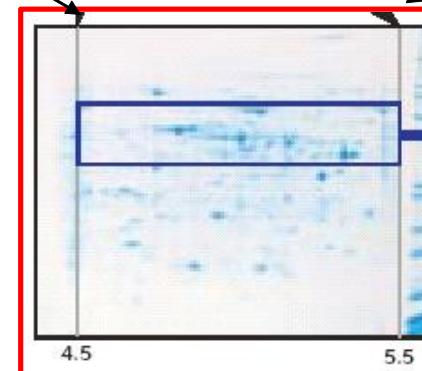
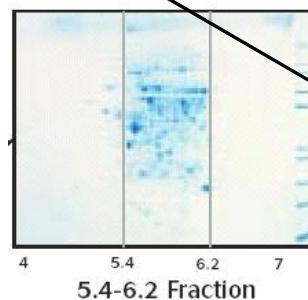
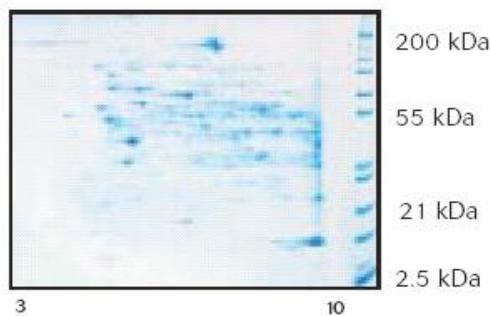


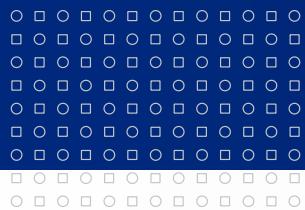
## PREFRACTIONATION

### MICRO RANGES

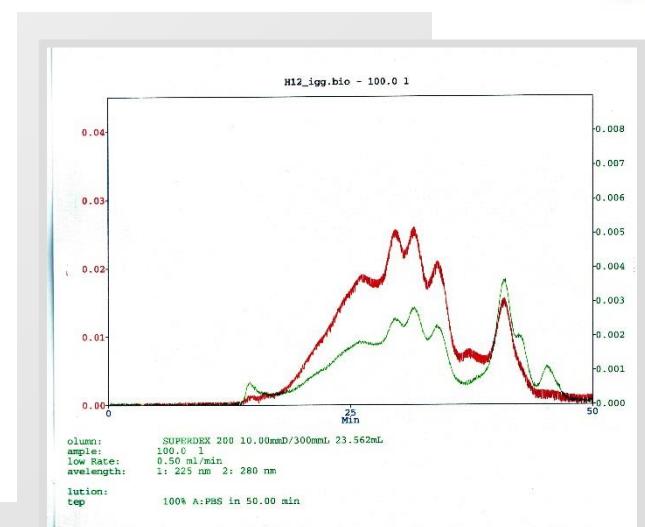
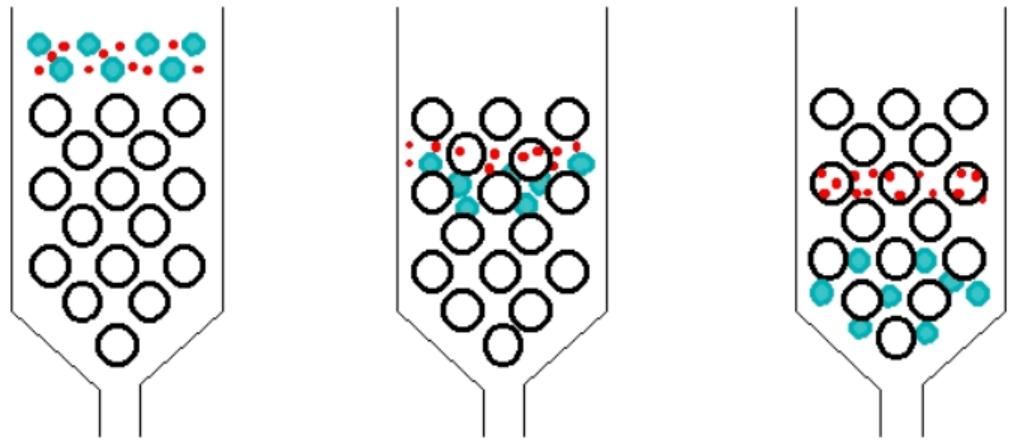
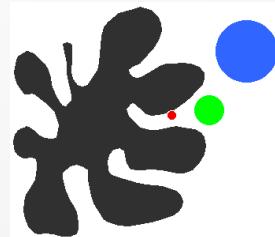


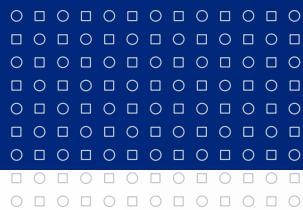
pl

**Unfractionated**



## GEL CHROMATOGRAPHY





## MOTIVATING LITERATURE FOR ADVANCED READERS

### **Two-dimensional gel electrophoresis in proteomics: A tutorial**

Thierry Rabilloud et al. *Journal of Proteomics* 2011

### **Two-dimensional gel electrophoresis in proteomics: past, present and future**

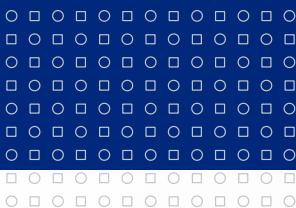
Thierry Rabilloud et al. *Journal of Proteomics* 2010

### **Proteomic biomarker discovery: It's more than just mass spectrometry**

Josip Blonder et al. *Electrophoresis* 2011

### **Basics and recent advances of two dimensional – polyacrylamide gel electrophoresis**

Sameh Magdeldin et al. *Clinical Proteomics* 2014



For all the complex problems and difficult questions  
there is always one simple, easily comprehensible  
**w r o n g** answer.

