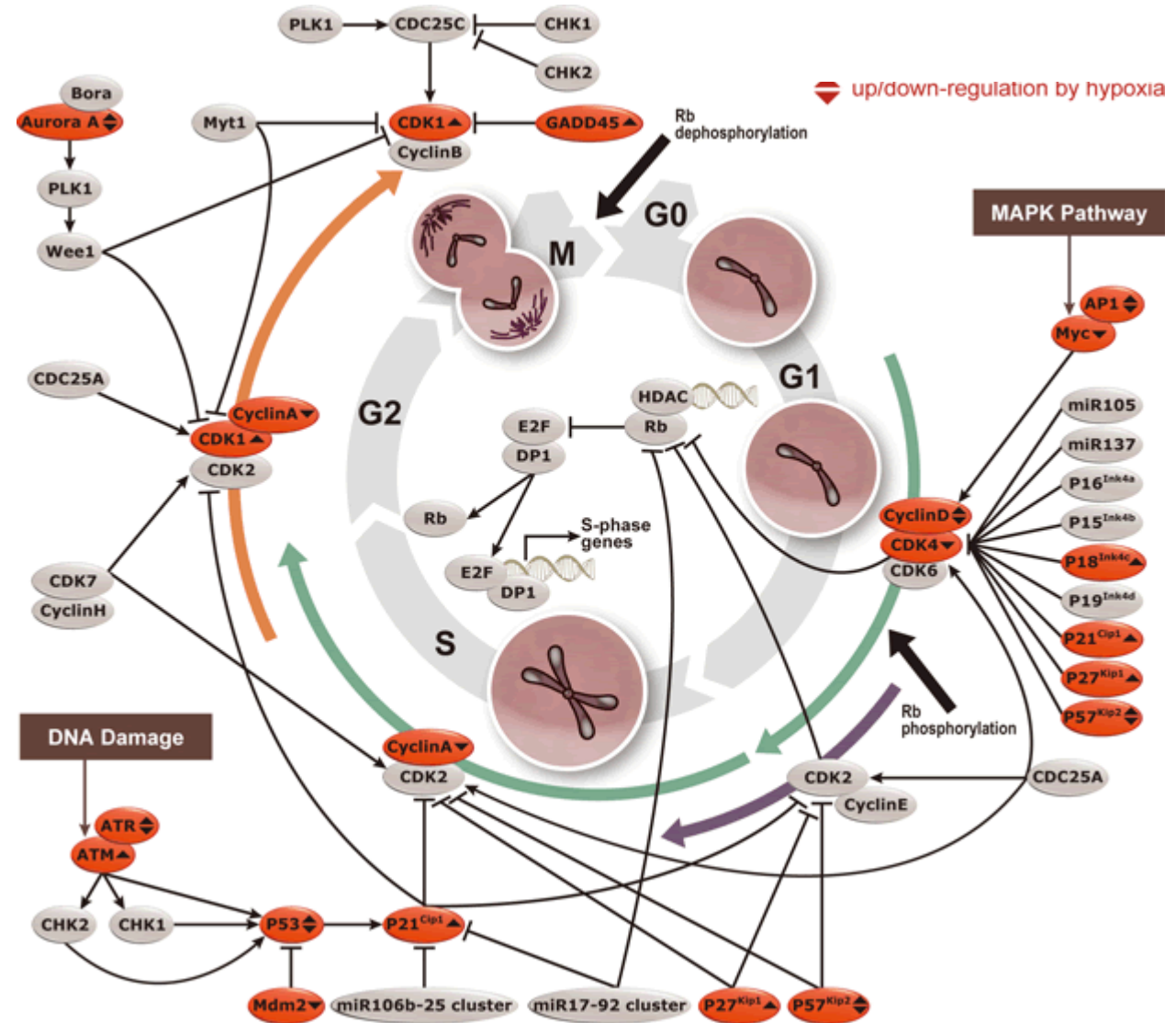


# Cell cycle

RNDr. Jan Škoda, Ph.D.  
Department of Experimental Biology



# Outline

- Definition and phases of the cell cycle
- The cell-cycle control system
- Molecular regulation of the cell cycle
- Defects in the cell cycle
- Models to study the cell cycle



# The cell cycle



# *Omnis cellula e cellula*

- **Law of cell lineage: each cell (stems) from another cell**
- *Pathologic conditions result from defects in cells*

– 1857 – Rudolf Virchow



– 1852 – Robert Remak



# The cell cycle

- Well-organized sequence of events in which the cell replicates by duplicating its content and dividing in two
  - Biogenesis of cell structures (and organelles)
  - Their distribution within the cell and to daughter cells
  - Unicellular organisms (bacteria, yeasts, protists) – new organism; Multicellular organisms: development, growth and tissue renewal/regeneration
- **A minimum set of spatiotemporally-organized processes** that a cell must perform **to pass the genetic information to the next generation of cells**: replication of genome, essential macromolecules and organelles; proper distribution into daughter cells



# The cell cycle

**Requires a precise control**

## All organisms

- Orchestrates **cell growth and division**: functional cell size

## Multicellular organisms

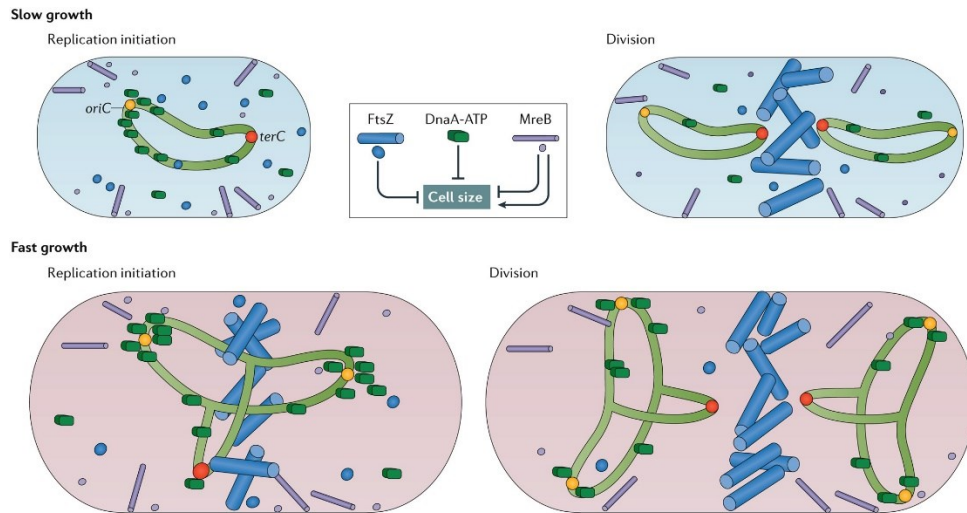
- Orchestrates **cell division with development/renewal**
- Homeostasis and a proper function of tissues
- Prevents uncontrolled proliferation



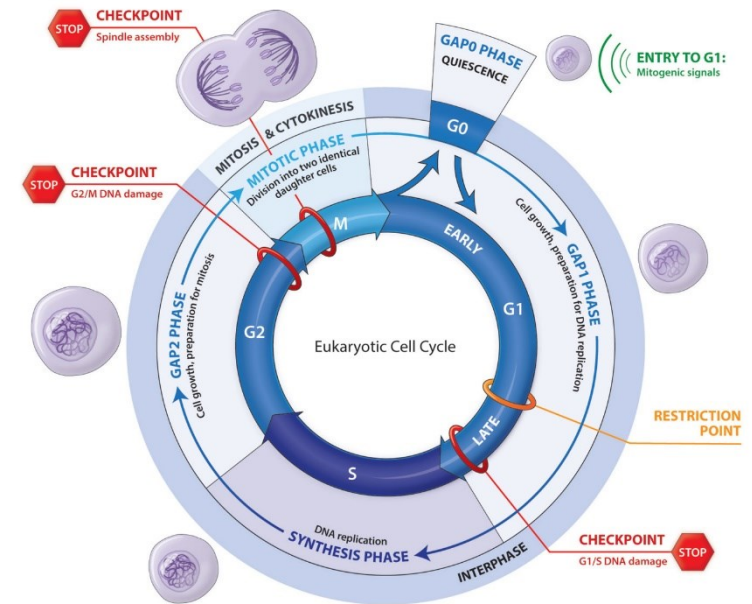
# The cell cycle: growth and division

– **Prokaryotic cells:** cell growth affects spatial localization of **cell cycle determinants**

– **Eukaryotic cells:** cell cycle entry regulated by **growth factors** (restriction point: "point of no return" )



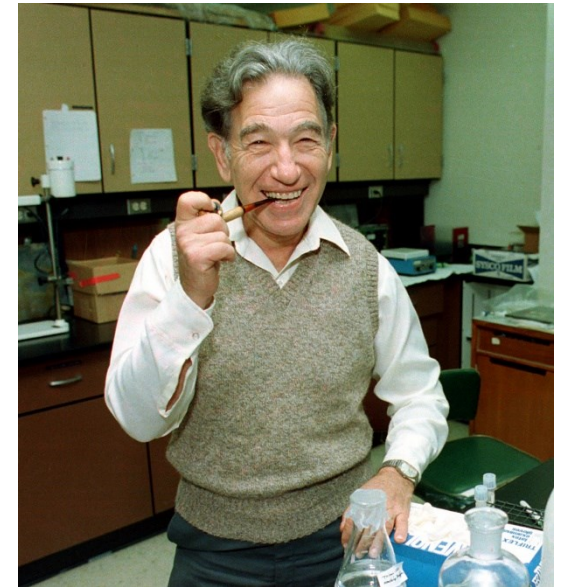
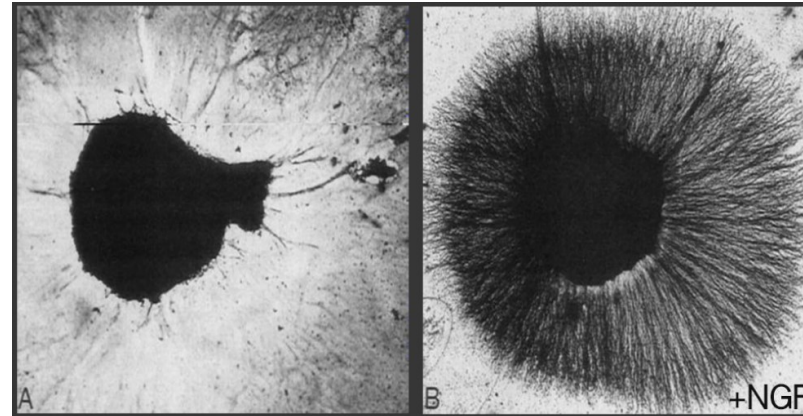
In media that support slow growth, replication initiation and Z-ring assembly occur sequentially (top panel), whereas in media that support fast growth, FtsZ can form the Z-ring a few minutes after cell birth, potentially coinciding with replication initiation (bottom panel).



# Growth factors and cell cycle

## Rita Levi-Montalcini and Stanley Cohen

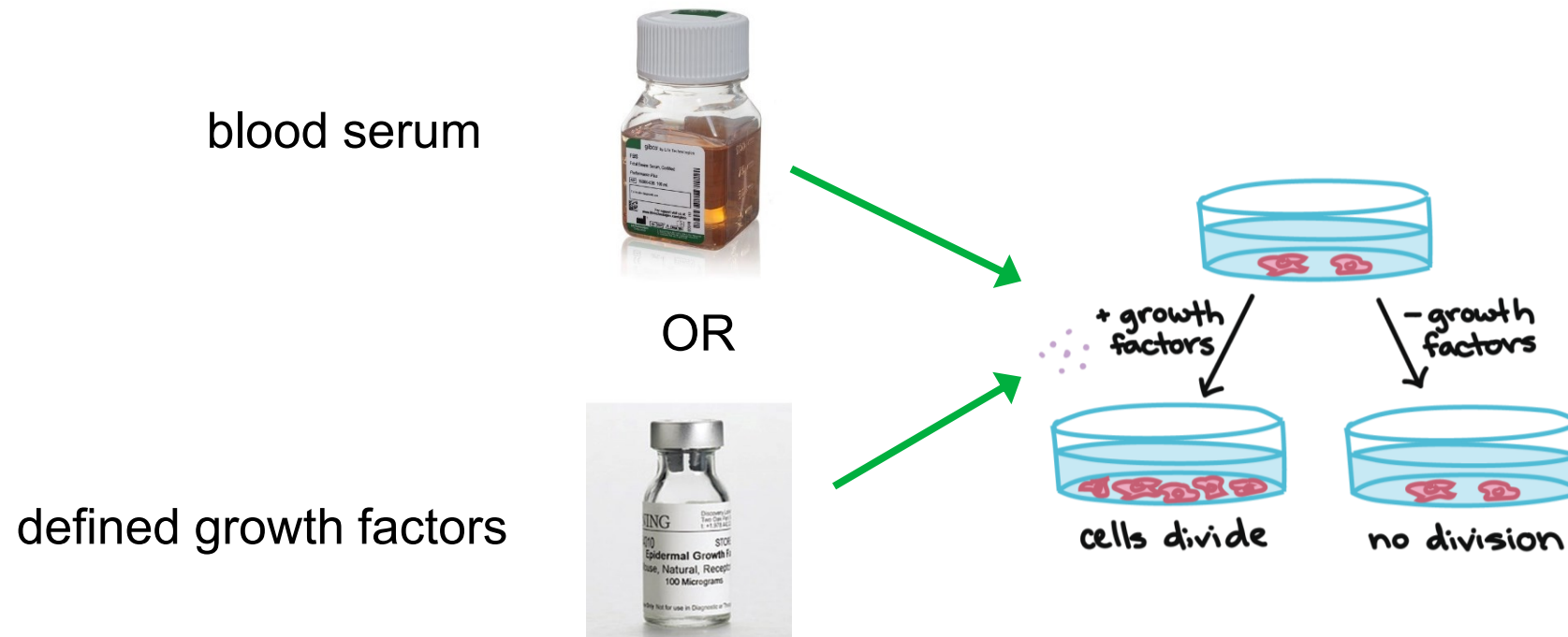
- 1950s discovered *nerve growth factor* (NGF) and *epidermal growth factor* (EGF)
- Nobel Prize in 1986 for “discoveries which are of fundamental importance for our understanding of the mechanisms which regulate cell and organ growth.”
- **Growth factors stimulate the proliferation of cells** (and regulate differentiation during development)





# Growth factors and cell cycle

– **Mammalian cell culture** protocols include growth factors in media:



# Phases of the eukaryotic cell cycle

## Interphase

### – (G<sub>0</sub> phase)

- Resting state (quiescence)
- Functional/metabolically active
- Terminally differentiated cells

### – G<sub>1</sub> phase

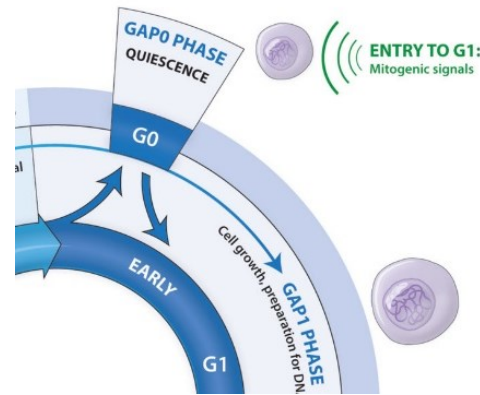
- Metabolic activity, cell growth
- Biosynthesis, doubling of organelles

### – S phase

- Replication of DNA

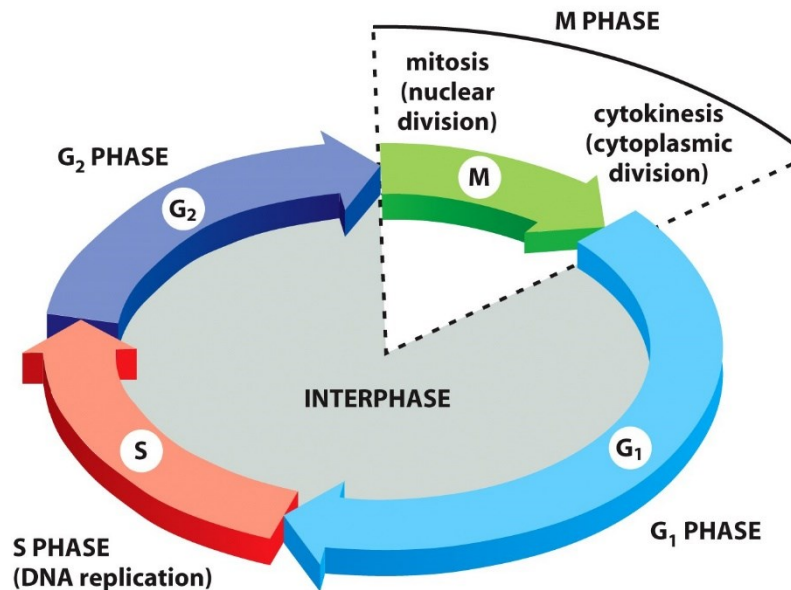
### – G<sub>2</sub> phase

- Biosynthesis of proteins essential for nuclear division and cytokinesis



## M phase

- Nuclear division (mitosis/meiosis)
- Cytokinesis

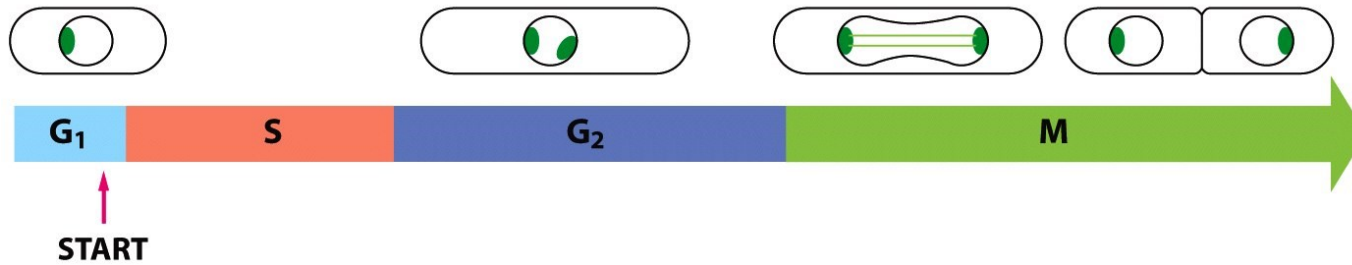


(“G” phase comes from “gap” phase)

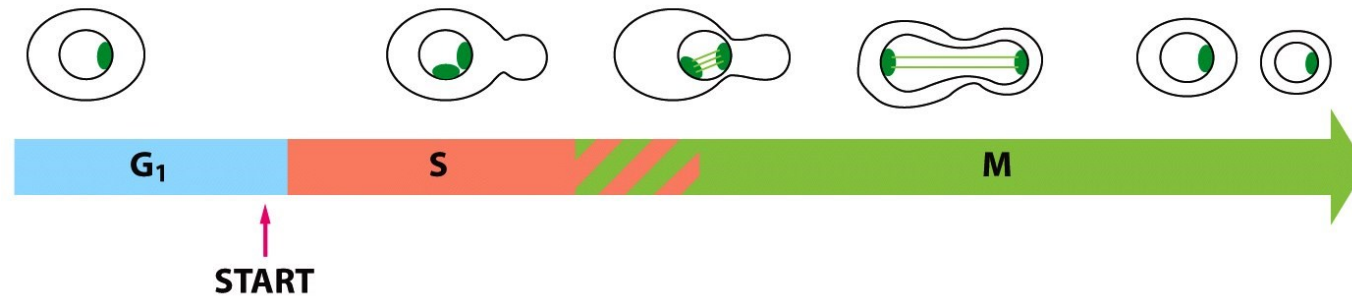


# Not all eukaryotic organisms follow the typical eukaryotic cell cycle

(A) FISSION YEAST (*Schizosaccharomyces pombe*)



(B) BUDDING YEAST (*Saccharomyces cerevisiae*)



- Budding yeasts: mitotic spindle forms during late S phase (no G2 phase)

# The cell-cycle control system



# Transition through different phases must be well controlled

## Maintenance of genome integrity

- Proper sequence of different phases
  - DNA replication followed by nuclear division (not replicated twice, prevents gene amplification)
- Fully replicated error-free DNA
  - Nuclear division must not start before DNA replication is completed
- Proper segregation of chromatids/chromosomes

## Homeostasis

- In coordination with developmental programs

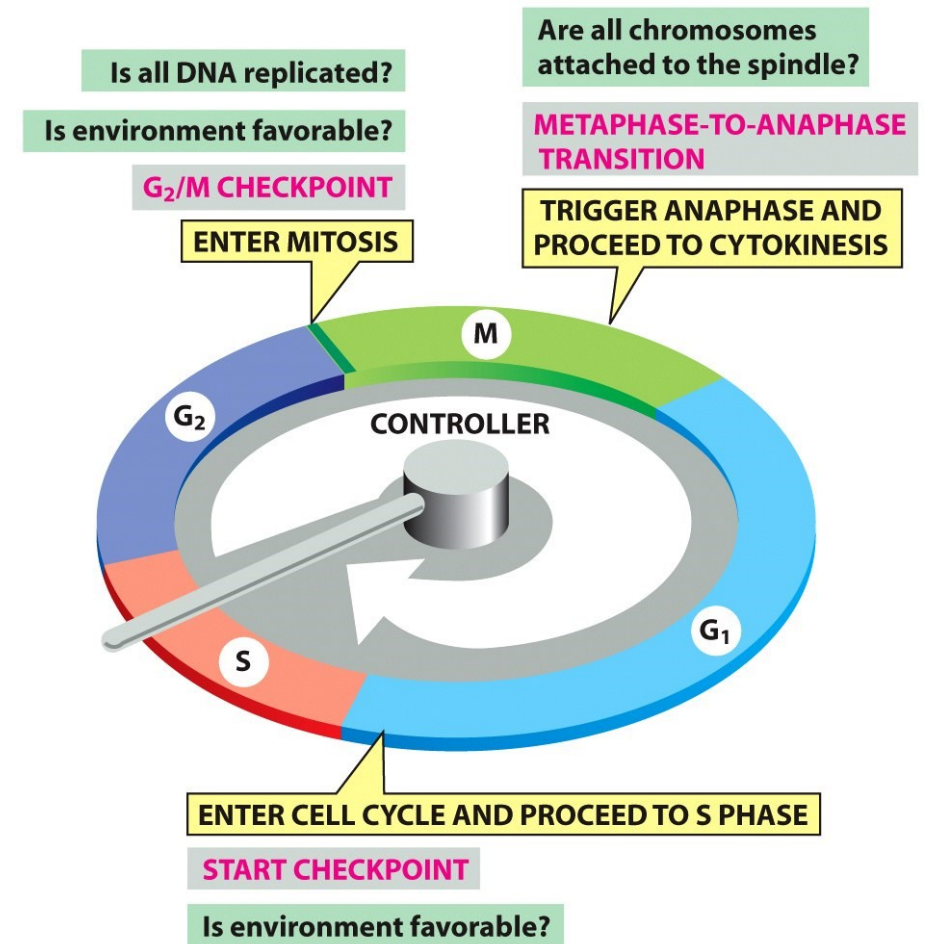


# Cell cycle checkpoints

– Regulate the cell cycle progression

## Three major checkpoints:

- Late G1 (G1/S):  
Restriction point (mammalian cells)  
Start (yeasts)
- Late G2 (G2/M)
- Metaphase-to-anaphase transition

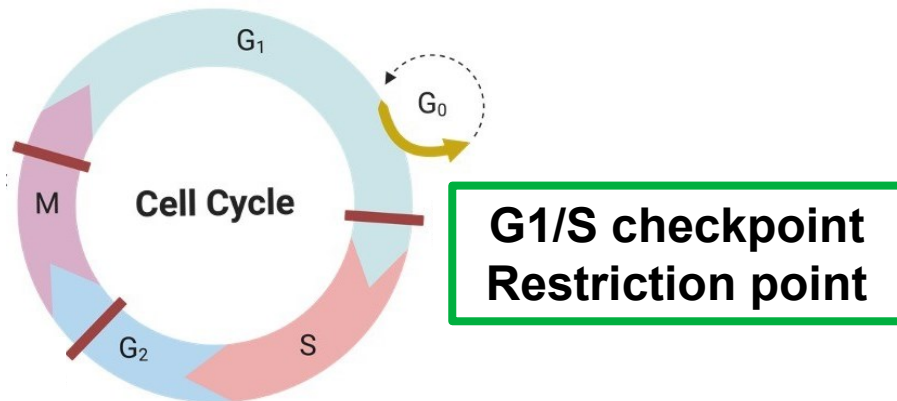


# G1/S checkpoint – Restriction point

## – Favorable environment?

- Enough nutrients, nucleotides
- **Extracellular mitogenic signals**  
(growth factors; homeostasis-related)

## – DNA undamaged?



## YES:

- **“Point of no return”** → committed to proceed through the cell cycle

## NO:

## – Progress delayed

- e.g., DNA repair

## – Or enters G<sub>0</sub> phase

- Terminally differentiated cells
- Quiescent cells (adult stem cells)
- Senescent (aged) cells

## – Or undergoes apoptosis

# G2/M and metaphase-to-anaphase

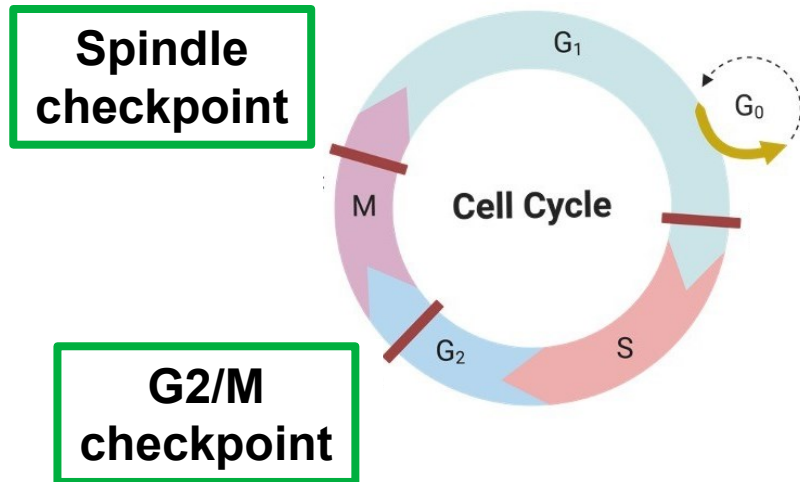
## G2/M checkpoint

- DNA fully replicated, no damage?
- Sufficient cell size? (Yeasts)

## Metaphase-to-anaphase

### transition (spindle checkpoint)

- Chromosomes (chromatids) attached to spindle?
- Aligned in the metaphase plate?



**Delay progression until problems solved  
/ Chronic activation leads to cell death**



# Molecular regulation of the cell cycle



# Central regulatory system

## Cyclins & cyclin-dependent kinases (Cdks)

### Cyclins

- Undergo **cycles of synthesis and degradation**
- **Regulate Cdk activity – positive regulators: bind to and activate Cdks**

### Cdks

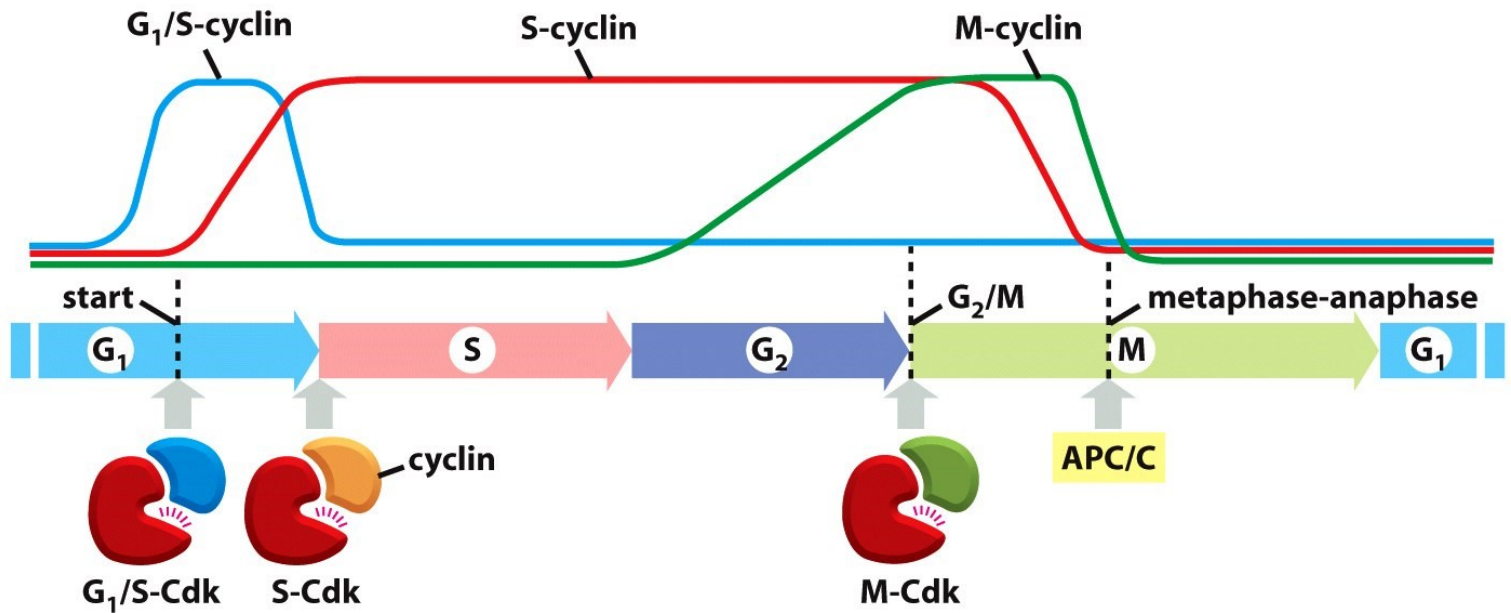
- **Protein kinases: active Cdks phosphorylate target proteins**
- **Govern progression through the cell cycle**

**Different cyclin-Cdk complexes = different target proteins**



# Central regulatory system

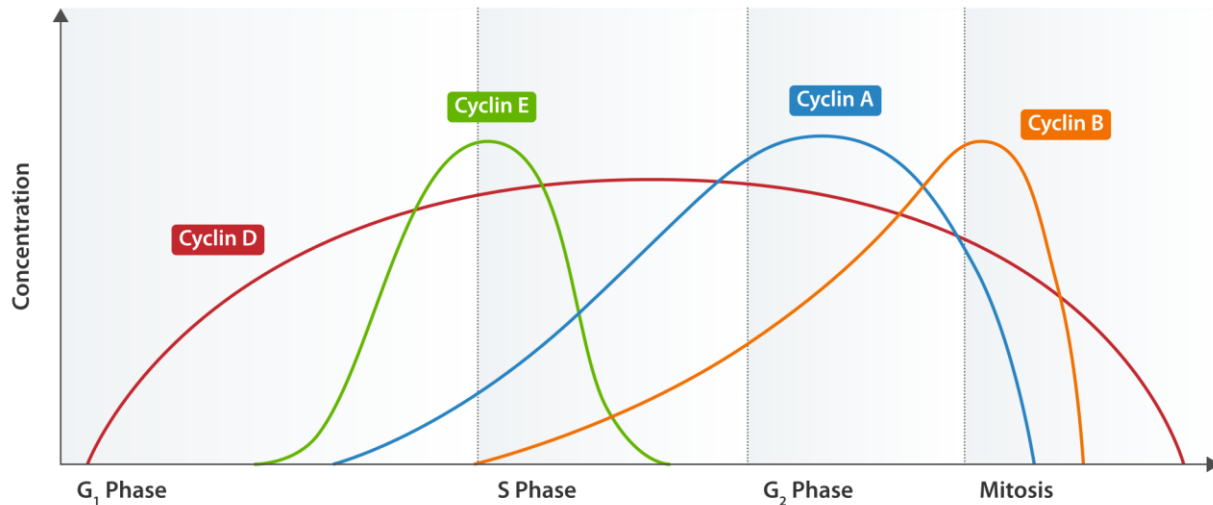
- Cdk's constantly present
- Cyclins synthesized and degraded during cell cycle
- Regulated by cell signaling and degradation systems



# Cyclin-Cdks complexes specific for each phase

CYCLIN-CDK COMPLEX	VERTEBRATES	
	CYCLIN	CDK PARTNER
G <sub>1</sub> -Cdk	cyclin D*	Cdk4, Cdk6
G <sub>1</sub> /S-Cdk	cyclin E	Cdk2
S-Cdk	cyclin A	Cdk2, Cdk1**
M-Cdk	cyclin B	Cdk1

- Help progression through G1 (Rb phosphorylation)
- **Progression through Restriction point**
- Mainly promote DNA replication
- Stimulate entry into the M-phase – **progression through G2/M checkpoint**



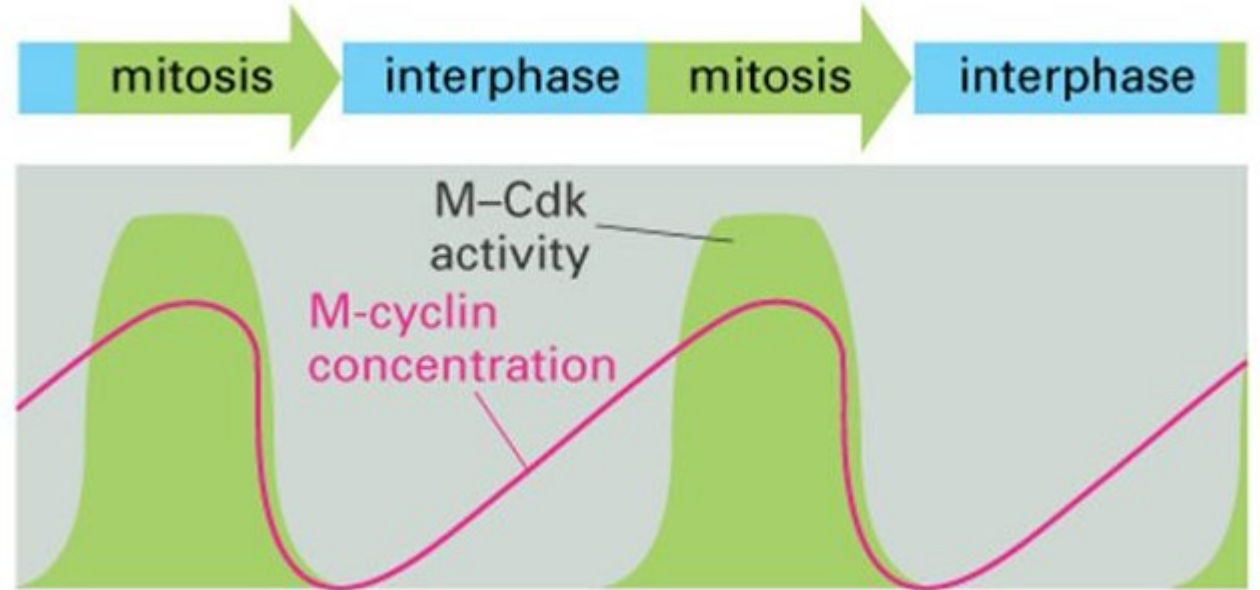
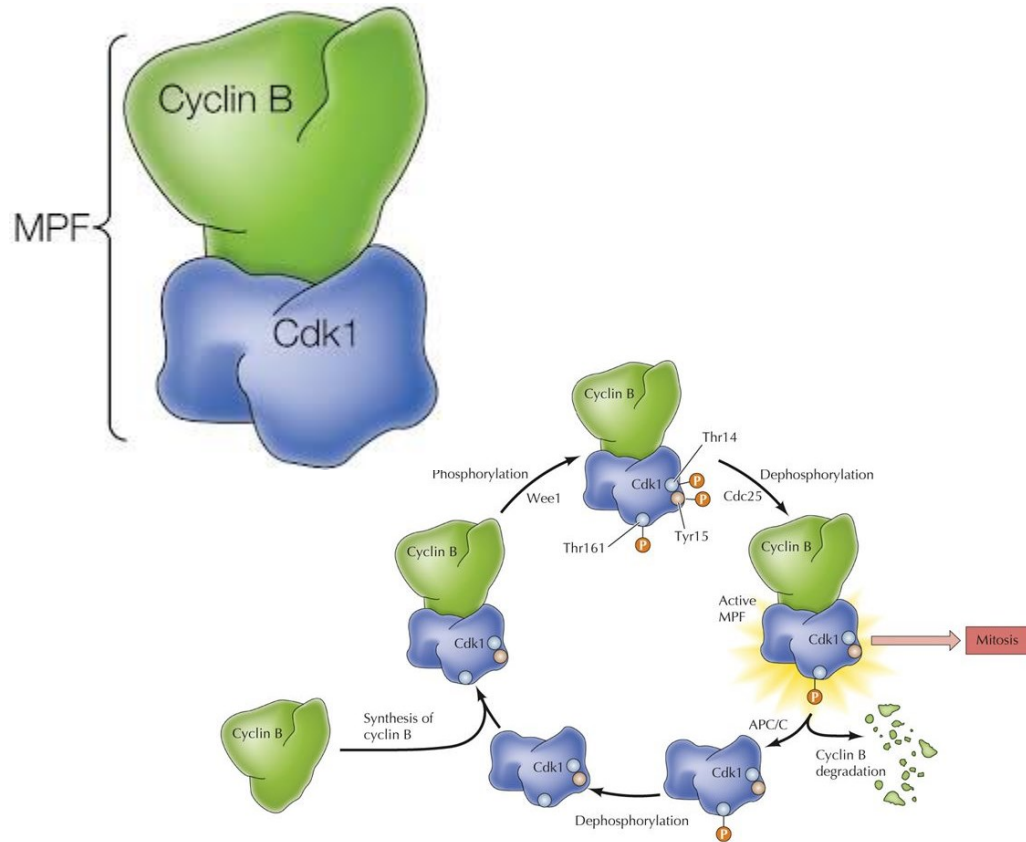
## Evolutionary conserved

- Yeast cyclins/Cdks can be compensated by human homologs

CYCLIN-CDK COMPLEX	VERTEBRATES		BUDDING YEAST	
	CYCLIN	CDK PARTNER	CYCLIN	CDK PARTNER
G <sub>1</sub> -Cdk	cyclin D*	Cdk4, Cdk6	Cln3	Cdk1**
G <sub>1</sub> /S-Cdk	cyclin E	Cdk2	Cln1, 2	Cdk1
S-Cdk	cyclin A	Cdk2, Cdk1**	Cln5, 6	Cdk1
M-Cdk	cyclin B	Cdk1	Cln1, 2, 3, 4	Cdk1

# Mitosis promoting factor (MPF): M-Cdk

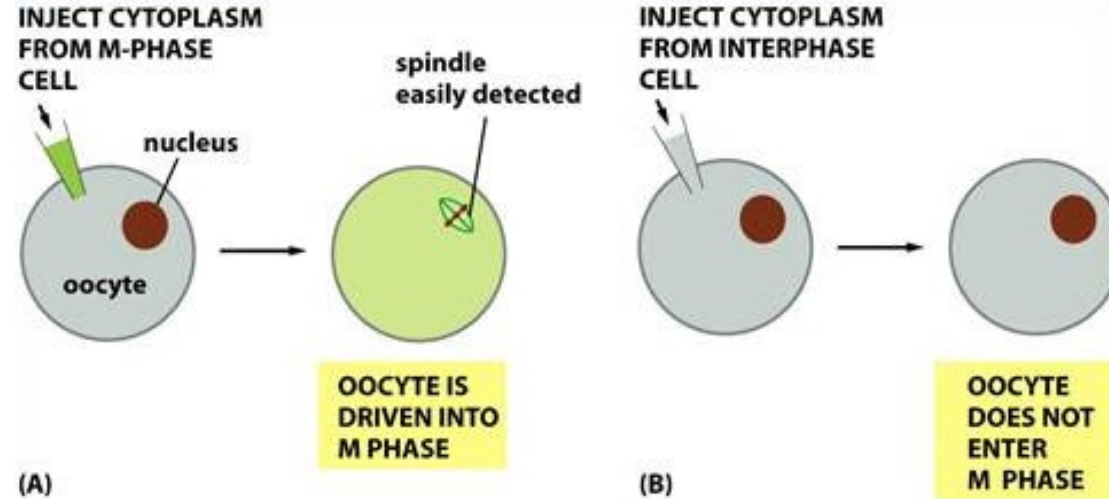
– Cyclin B + Cdk1 (Cdc2)



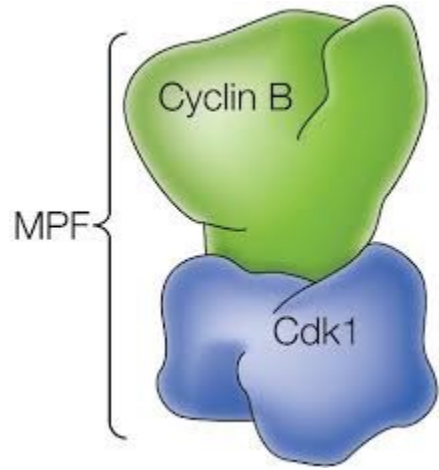
– M-Cdk (MPF) activity triggers progression to M-phase

# Discovery of MPF

- 1971: Masui & Markert, Smith & Ecker
- Microinjection of M-phase vs. interphase cytoplasm into oocytes of frog (*Rana pipiens*, northern leopard frog)
- **M-phase cytoplasm contains a factor inducing meiosis (M-phase) → MPF**



# Function of MPF (M-Cdk)



- **Phosphorylates target proteins → initiates processes essential for M-phase**
  - **Nuclear envelope breakdown:** phosphorylates nuclear pore complex subunits & proteins of nuclear lamina
  - **Initiates and promotes spindle assembly:** phosphorylation of motor proteins associated with microtubules
  - **Increases microtubule dynamics**
  - **Chromosome condensation:** phosphorylation of condensin proteins
- **Progression through G2/M checkpoint**

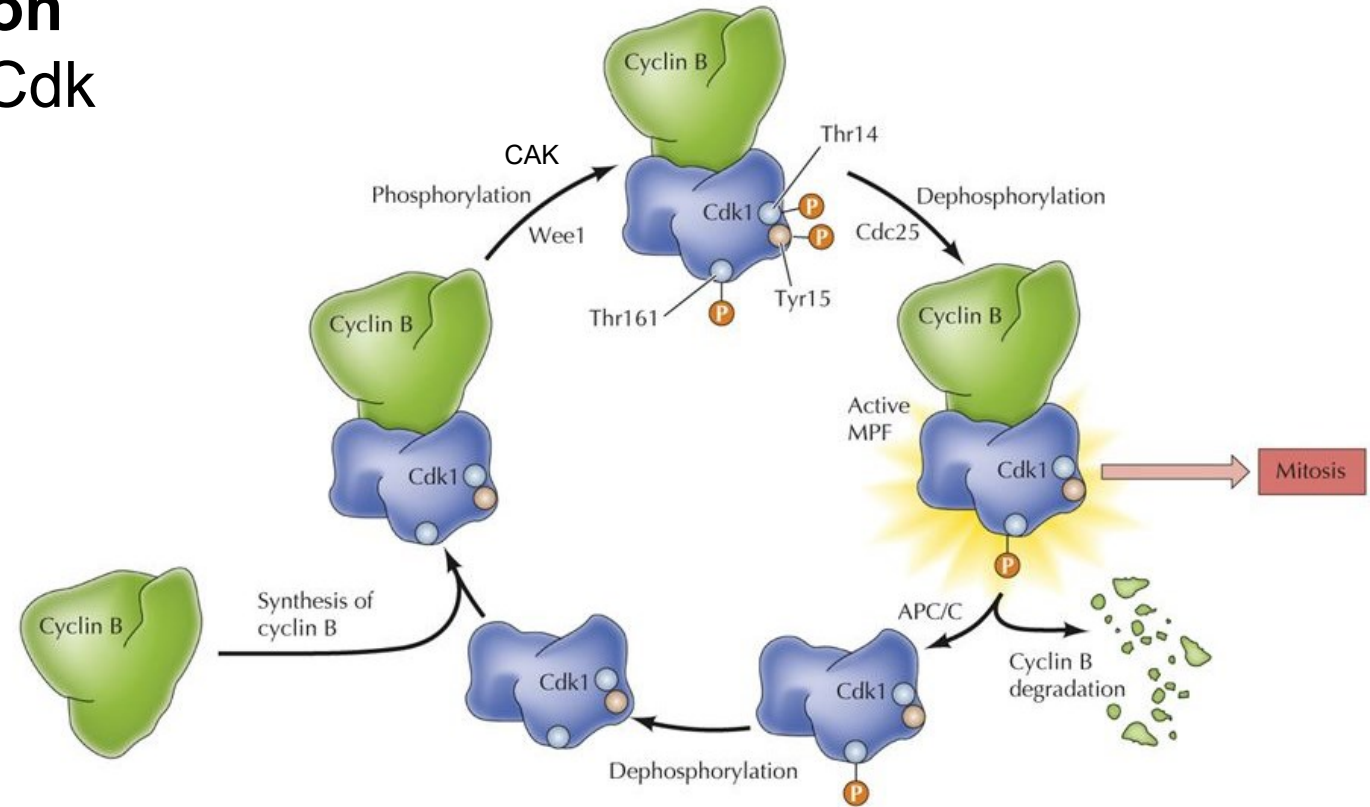
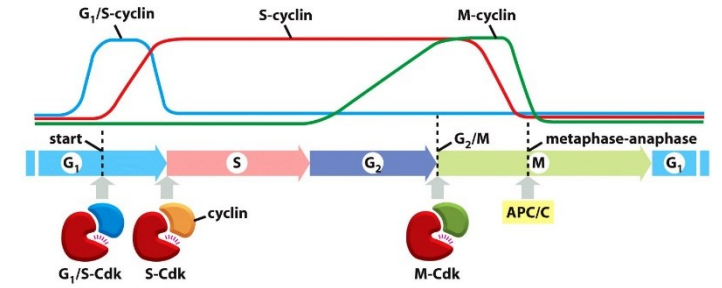
# Regulation of M-Cdk activity

## – Cyclin B levels & protein kinases

- Wee1: Inhibitory phosphorylation
- Activating phosphorylation by Cdk activating kinase (CAK)
- Cdc25: Activating dephosphorylation

→ Active M-Cdk: M-phase

- Tagged for proteasomal degradation by APC/C





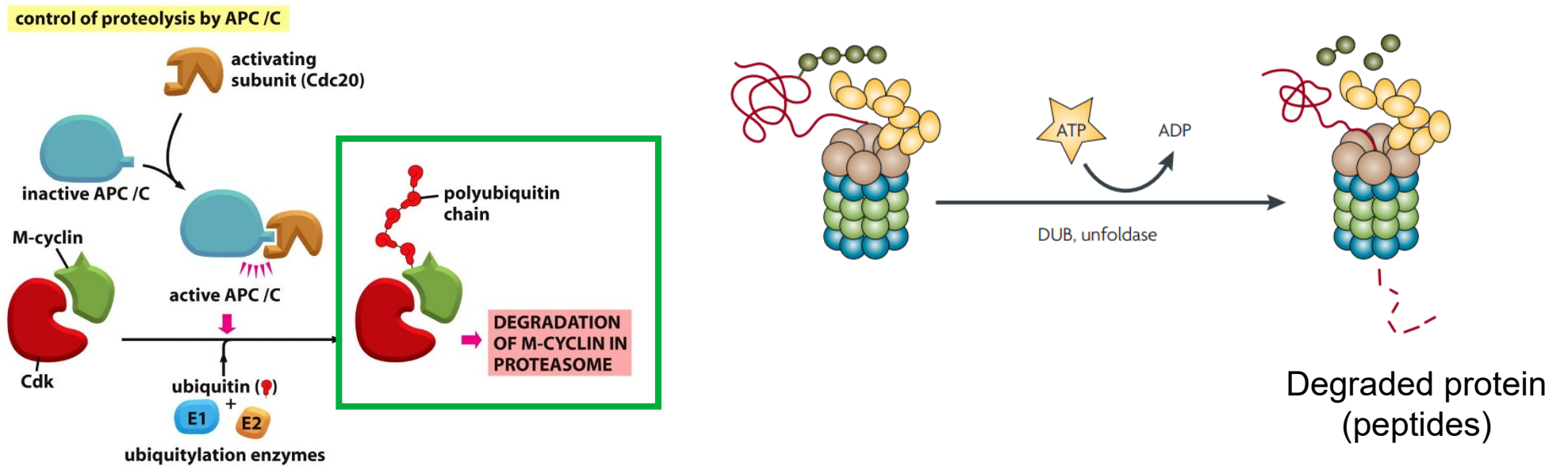
# APC/C (anaphase promoting complex/cyclosome)

- **Ubiquitin ligase:** adds multiple ubiquitin molecules to target proteins
- Polyubiquitylated proteins tagged for **degradation by proteasome**
- APC/C target specificity regulated by activating subunits
- Degradation targets
  - Securin (holds sister chromatid pairs) → promotes transition to anaphase
  - S- and M-cyclins → dephosphorylation and inactivation of Cdks (APC/C active to early G1)
- **Progression through spindle checkpoint**
- **Resets the cell-cycle control system (Cdks inactivation)**



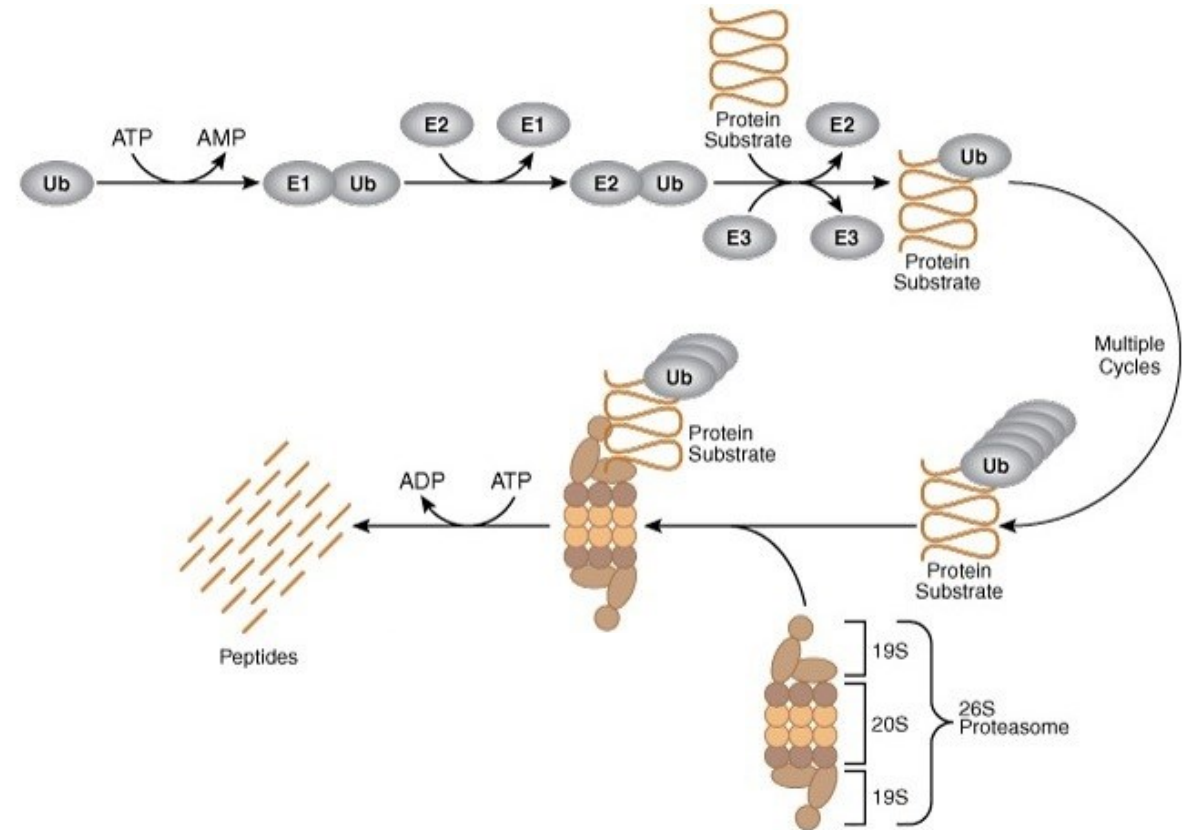
# Proteolysis by proteasome

- Ubiquitination (ubiquitylation) – tag for **proteasomal degradation**
- **Important regulatory role: Rapid degradation of cyclins** of preceding phases



# Ubiquitin-dependent proteolysis

- Multiple cycles of ubiquitination
- Ubiquitin transferred from ubiquitin conjugating enzyme, E2, to target protein by **E3 ubiquitin ligase**
- Ubiquitin covalently attached
- Polyubiquitinated proteins transferred to proteasome → Cleavage



# Signals regulating cell cycle entry

## Unicellular

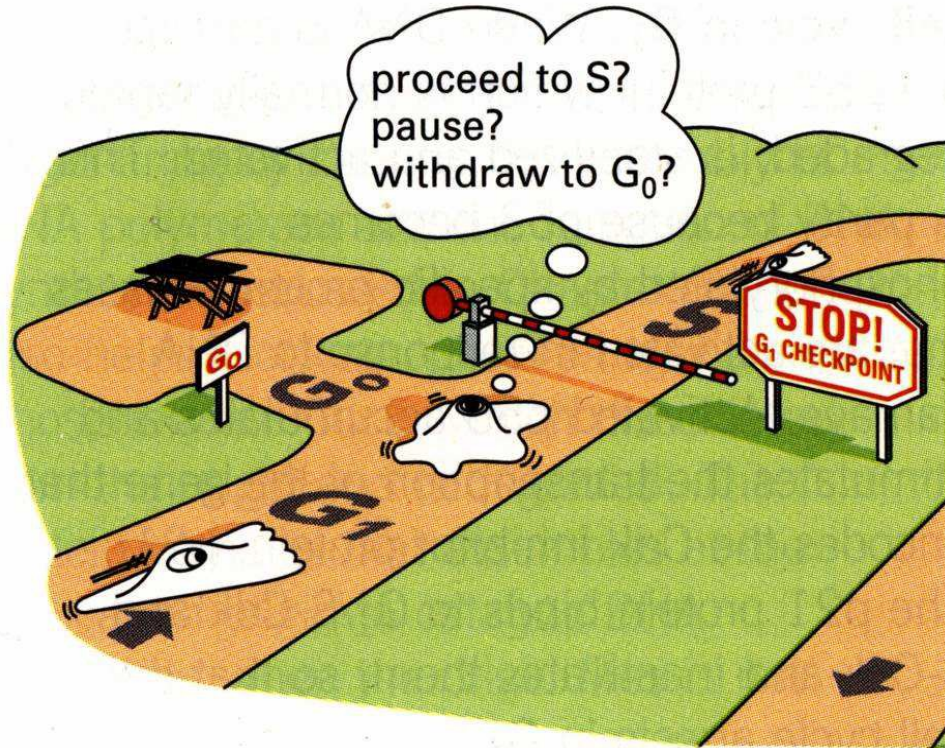
- Determined by nutrients availability (similar to prokaryotes)

## Multicellular

- **Mitogenic signals (growth factors)**
  - Produced by surrounding or distant cells within the organism
  - Homeostasis – **only when necessary to proliferate**: growth, renewal
- **Lack of mitogenic signaling**
  - Cell cycle halted/blocked (G0) – intrinsic braking mechanisms prevail



# Cell cycle arrest in G<sub>0</sub>



## – Lack of mitogenic signaling

- May be restored by mitogens

## – Terminally differentiated cells

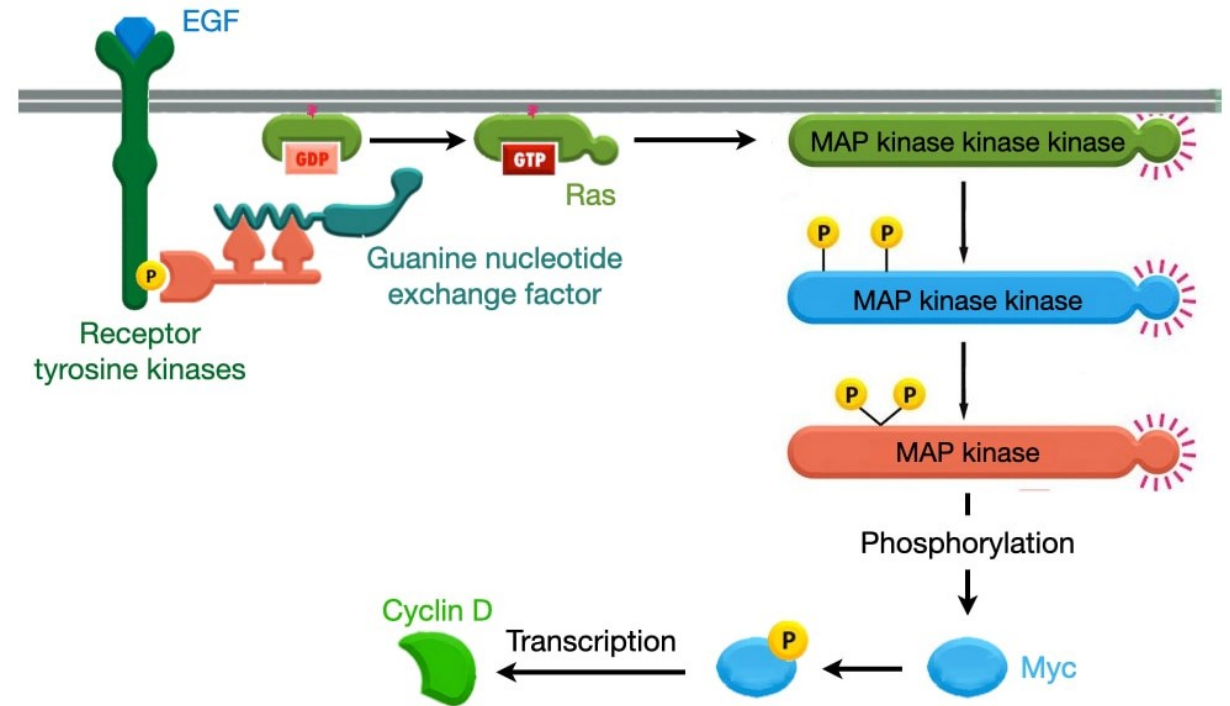
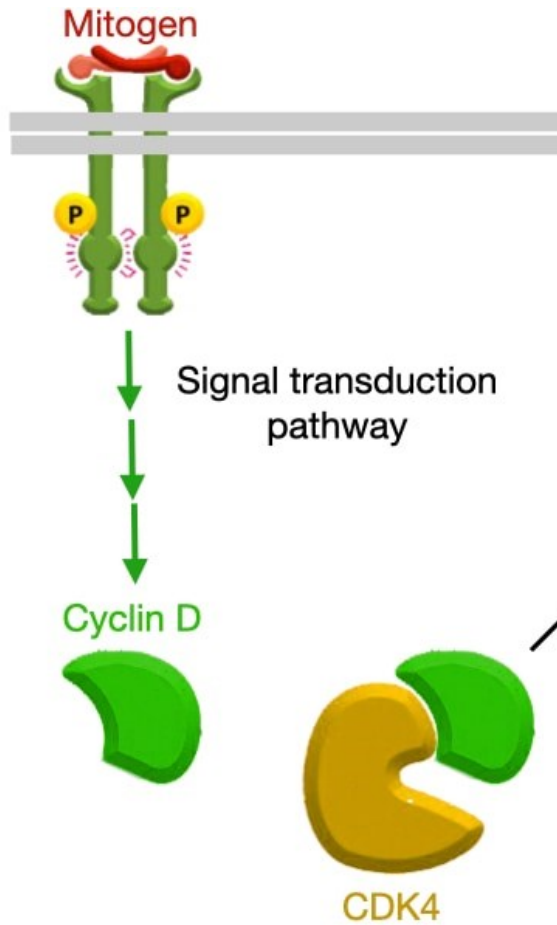
- Expression of Cdks and cyclins permanently turned off – cell cycle not responsive to mitogens
- e.g., neurons, skeletal muscle cells
- Reprogramming: dedifferentiation restores cell cycle

## – Replicative senescent cells

- Shortened telomeres (DNA damage) prevent progression through G<sub>1</sub>/S checkpoint
- Restricts number of cell divisions

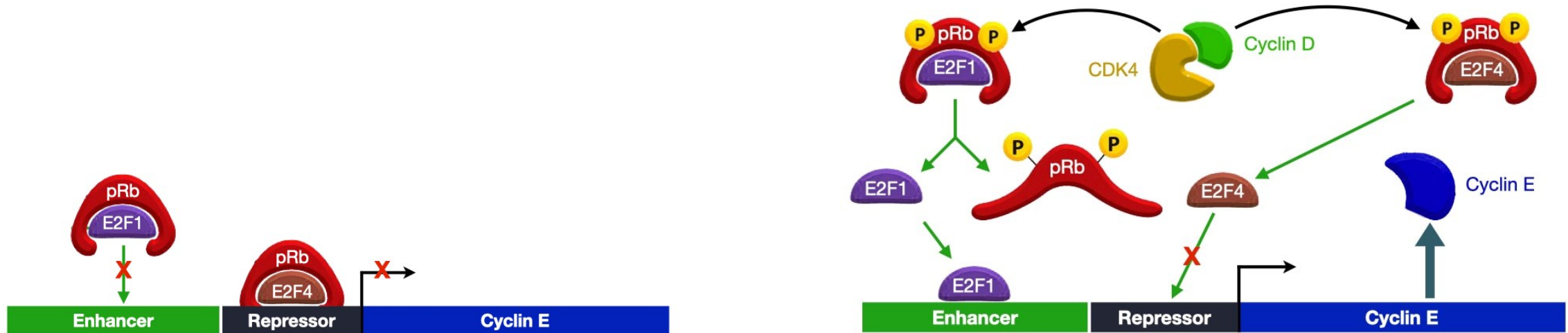
# Mitogenic signaling leads to cyclin D expression

- Epidermal growth factor (EGF) induced expression of cyclin D



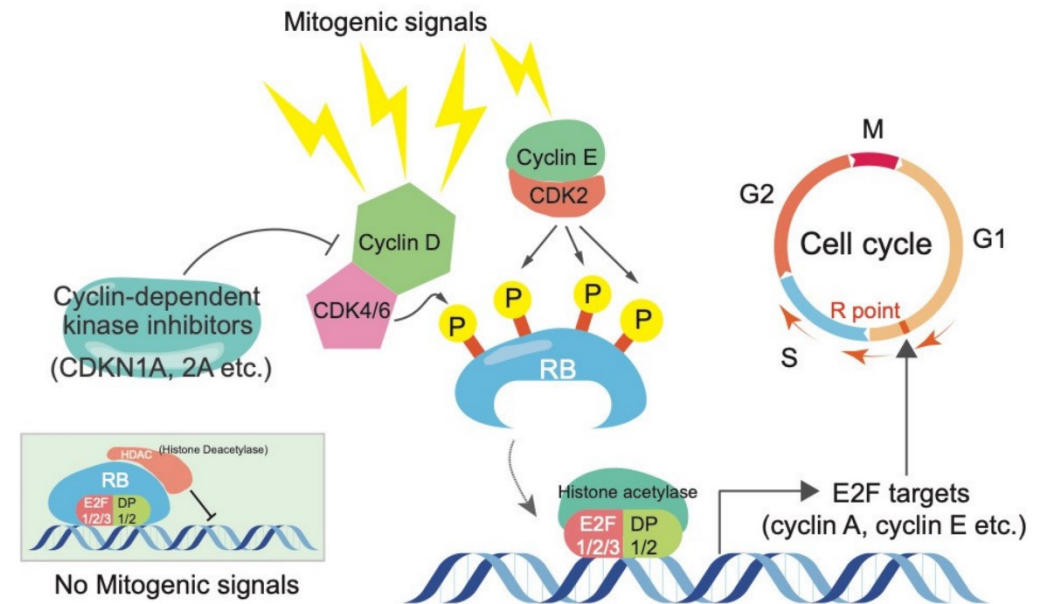
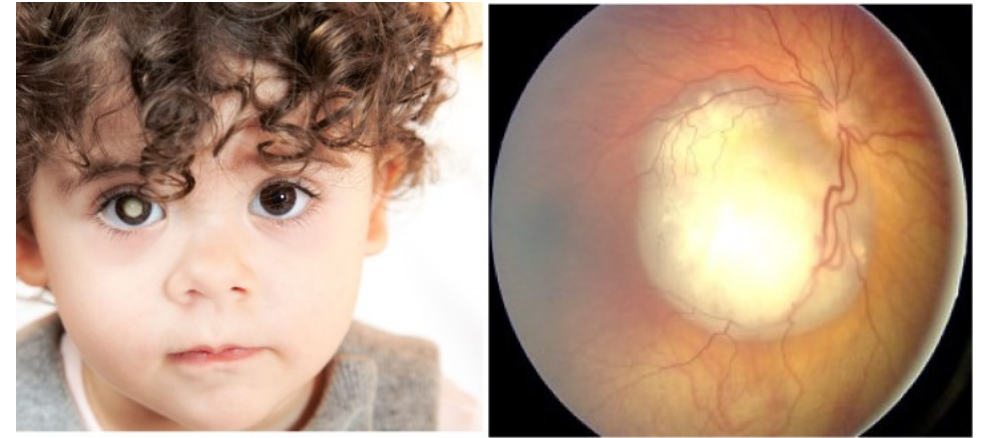
# How cyclin D helps to overcome restriction point?

- Cyclin D/CDK4 phosphorylates protein Rb – release of E2F transcription factors – expression of cyclin E and other S-phase proteins



# Protein Rb (retinoblastoma)

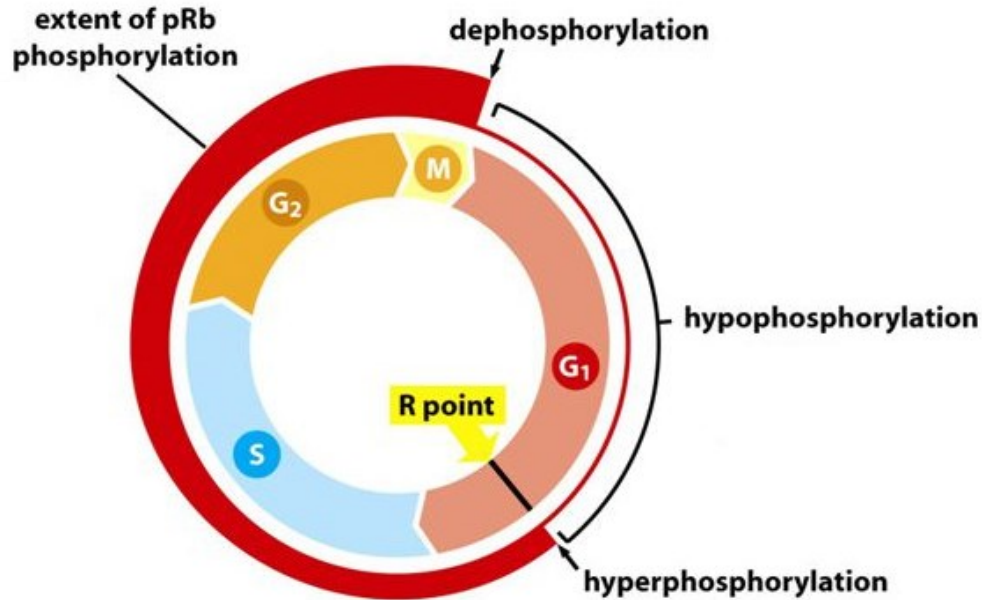
- Tumor suppressor
- Intrinsic negative cell cycle regulator
- Nuclear protein binding E2F
- Prevents transcription of E2F targets: genes encoding proteins involved mainly in DNA replication and cell cycle progression
- Rb phosphorylation – binding inhibited, E2F released



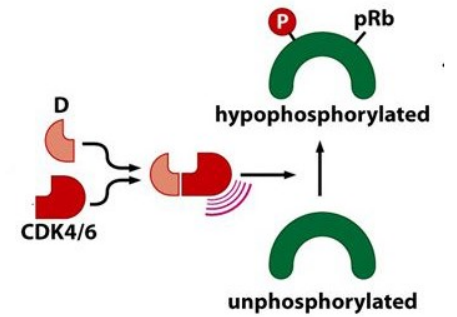


# Protein Rb phosphorylation & dephosphorylation

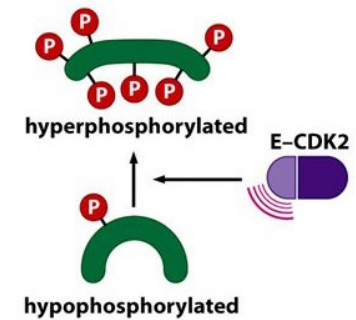
- Hyperphosphorylation – overcomes restriction point
- Dephosphorylated at late M-phase (protein phosphatase 1)



**Hypo**-phosphorylation is catalyzed by *cycD*-CDK4/6



**Hyper**-phosphorylation is catalyzed by *cycE*-CDK2

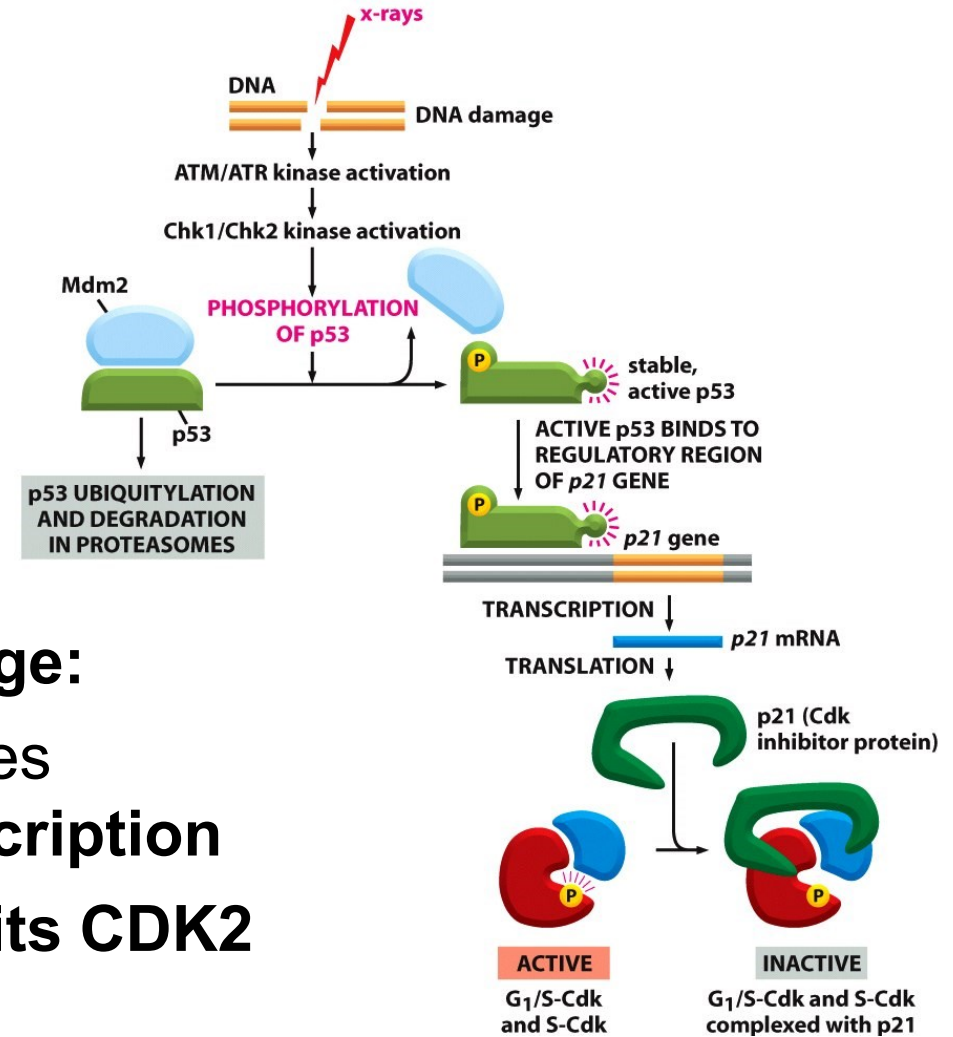
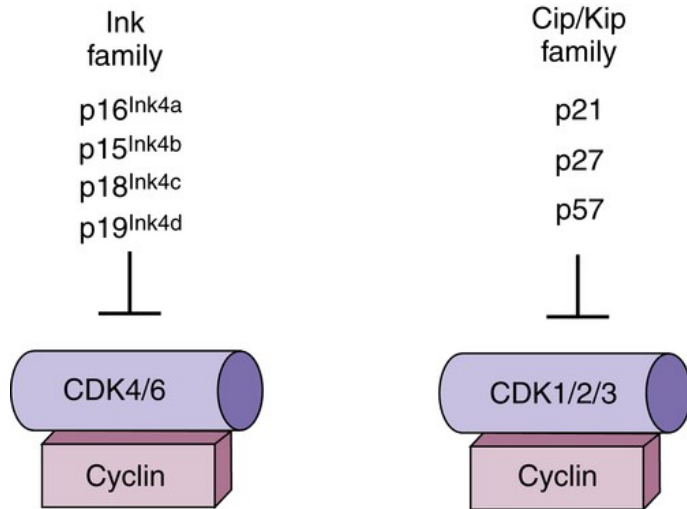


# Defects in the cell cycle



# Cell cycle arrest: Cdk inhibitor proteins

- Cell intrinsic Cdk inhibitors
- **Transcription induced upon stress stimuli, e.g., DNA damage**

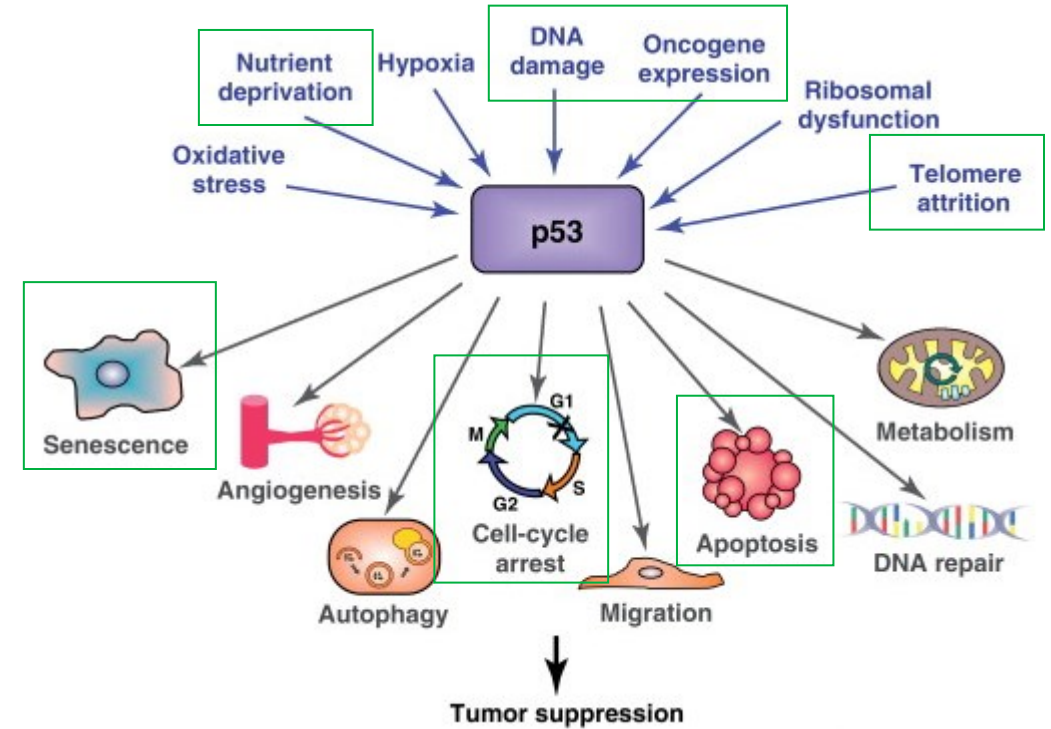


## DNA damage:

- p53 induces p21 transcription
- P21 inhibits CDK2 and CDK1

# p53 – guardian of the genome

- Tumor suppressor
- Integrates various stress signals
- Regulates expression of downstream targets
- Cell context-dependent response
  - Repair mechanisms, normal function restored, cell-cycle arrest, or apoptosis



- p53 mutated in ~50% of human cancers

# Cancer: A cell cycle defect

- Cell cycle positive regulators – typical **proto-oncogenes**
  - **Cyclin D/E, growth factors**, mitogenic signaling kinases and transcription factors (all promote cyclin D/E expression)
  - Amplification, overexpression, constitutive activity of kinases → **oncogenes: promote (uncontrolled) cell cycle progression**
- Cell cycle negative regulators – typical **tumor suppressors**
  - **Intrinsic Cdk inhibitors, protein Rb, protein p53** and DNA damage recognizing kinases
  - Inactivating mutations, deletions → **cell cycle checkpoint mechanisms disabled**



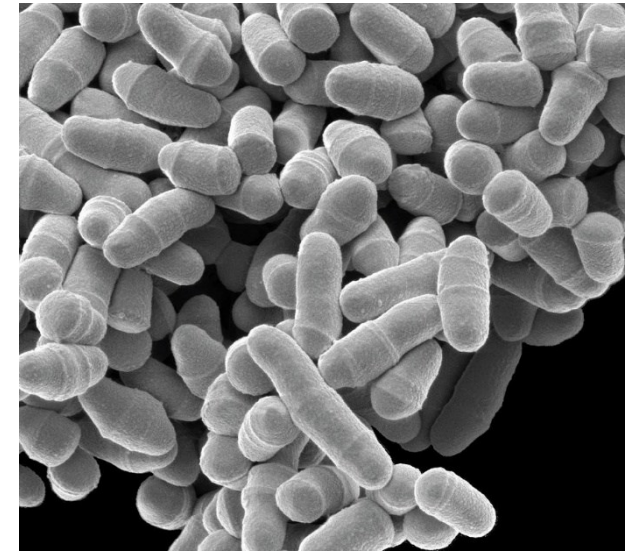
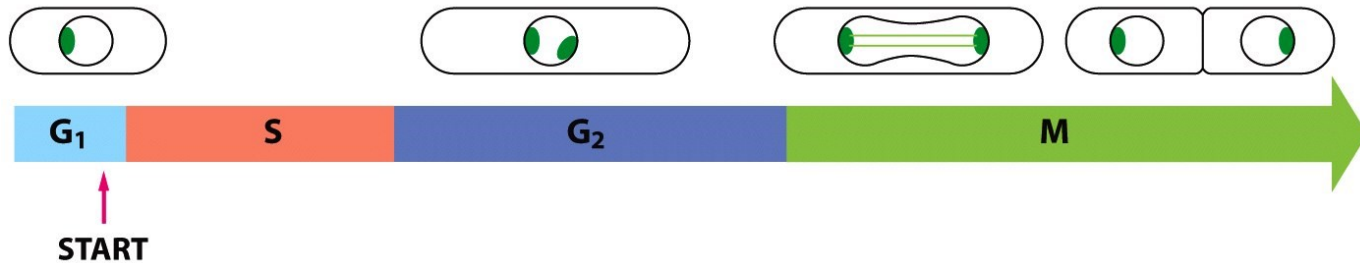
# Models to study the cell cycle



# *Schizosaccharomyces pombe*

- Fission yeast; Cell cycle: 4 phases

(A) FISSION YEAST (*Schizosaccharomyces pombe*)



- **Small genome:** 4,824 genes, ~70% human orthologs
- Non-pathogenic, **fast cell cycle** (90 min)
- Can exist as haploid: **easy loss-of-function studies** (only one allele)

# Schizosaccharomyces pombe

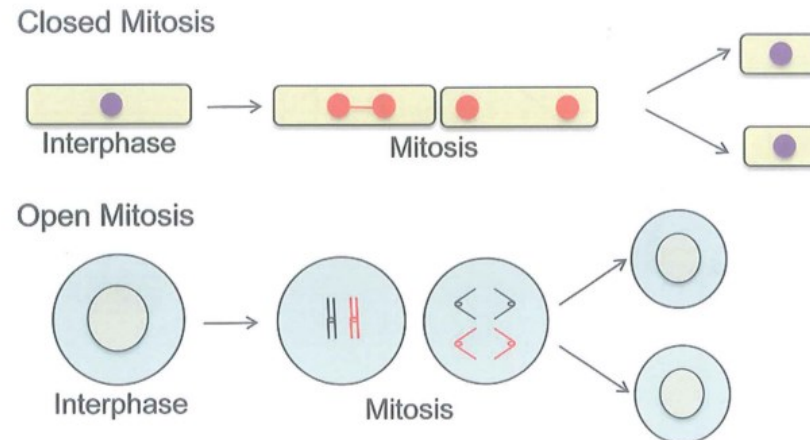
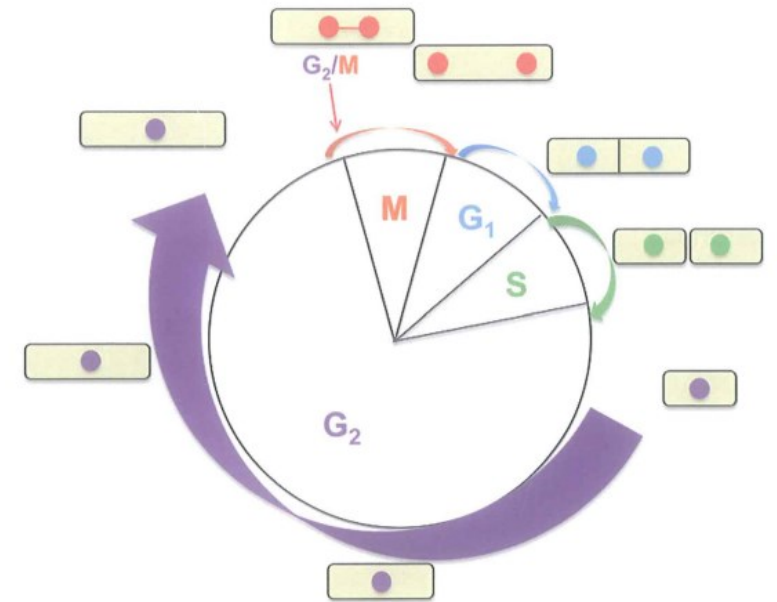
## – Optimal to study G2/M transition

- G2 is the longest – size is a parameter of progression
- Cdc2 (Cdk1), Wee1, Cdc25 identified in *S. pombe*

## – ! Differences from multicellular organisms

## – Closed mitosis

- Nuclear envelope does not break down

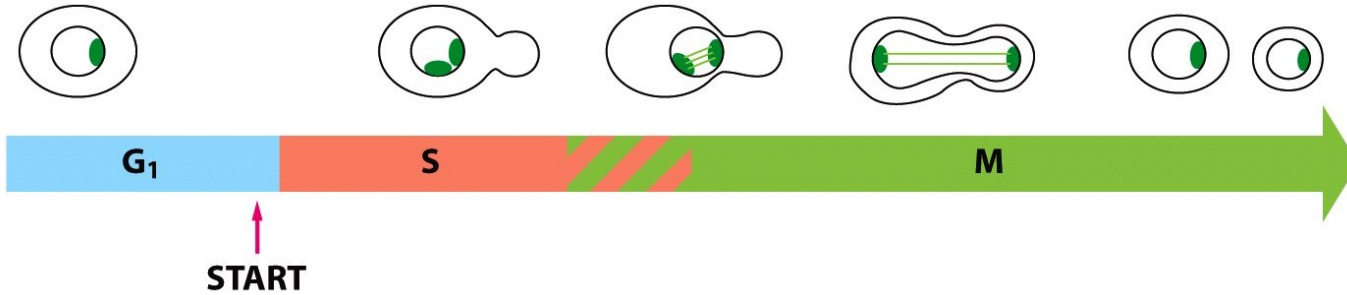




# Saccharomyces cerevisiae

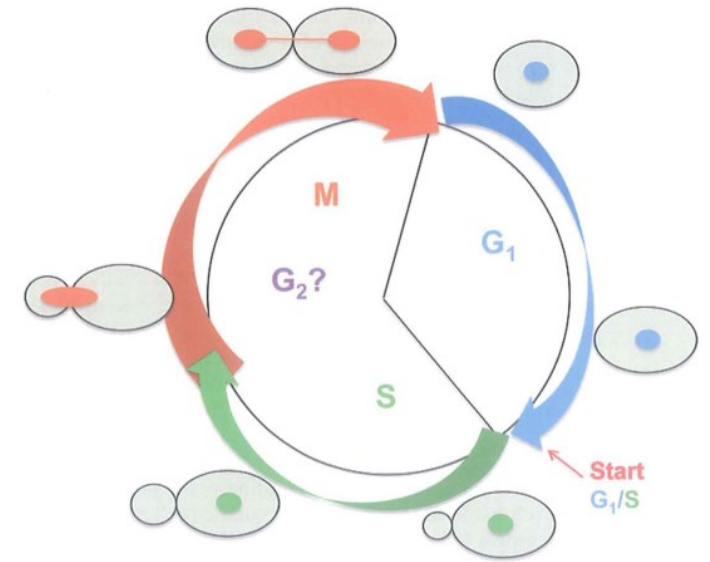
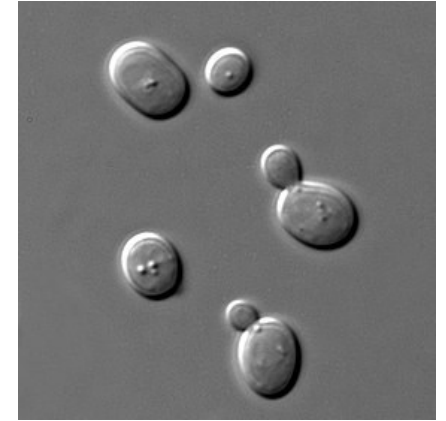
– Budding yeast

(B) BUDDING YEAST (*Saccharomyces cerevisiae*)



– Bud appears in S-phase, grows through cell cycle

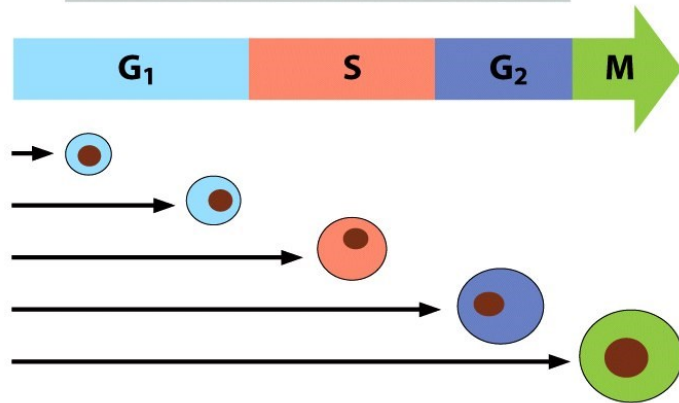
– **Optimal to study G<sub>1</sub>/S transition**



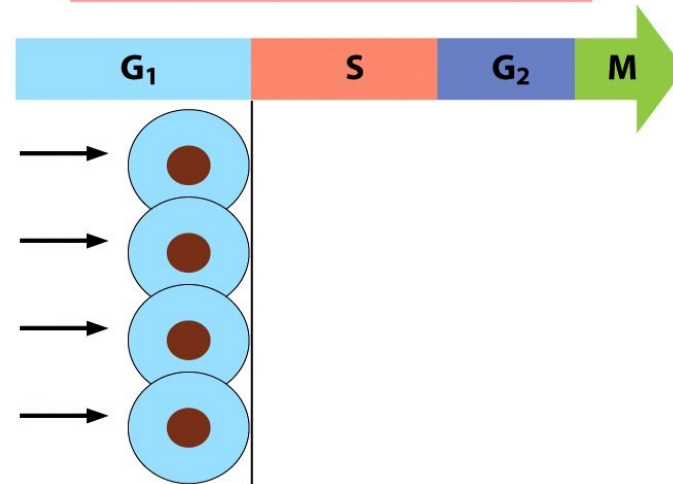
# How to study loss-of-function effects

- Loss of cell cycle – no cell division, non-viable culture
- **Conditional mutants – temperature sensitive**
  - Low temperature – protein functional vs. High temperature – protein not functional

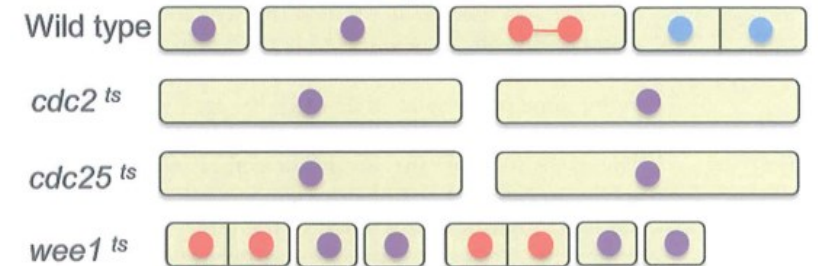
(A) PERMISSIVE (LOW) TEMPERATURE



(B) RESTRICTIVE (HIGH) TEMPERATURE



Fission yeast *S. pombe*



# Mammalian cell cultures

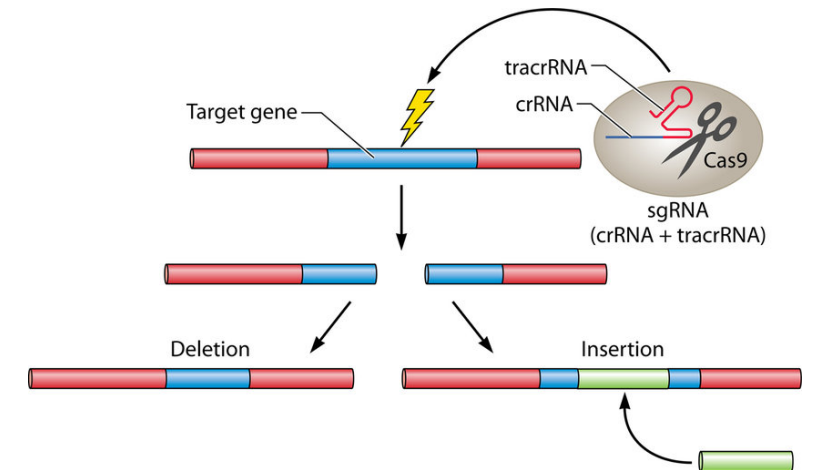
## – Primary (non-cancerous) cells

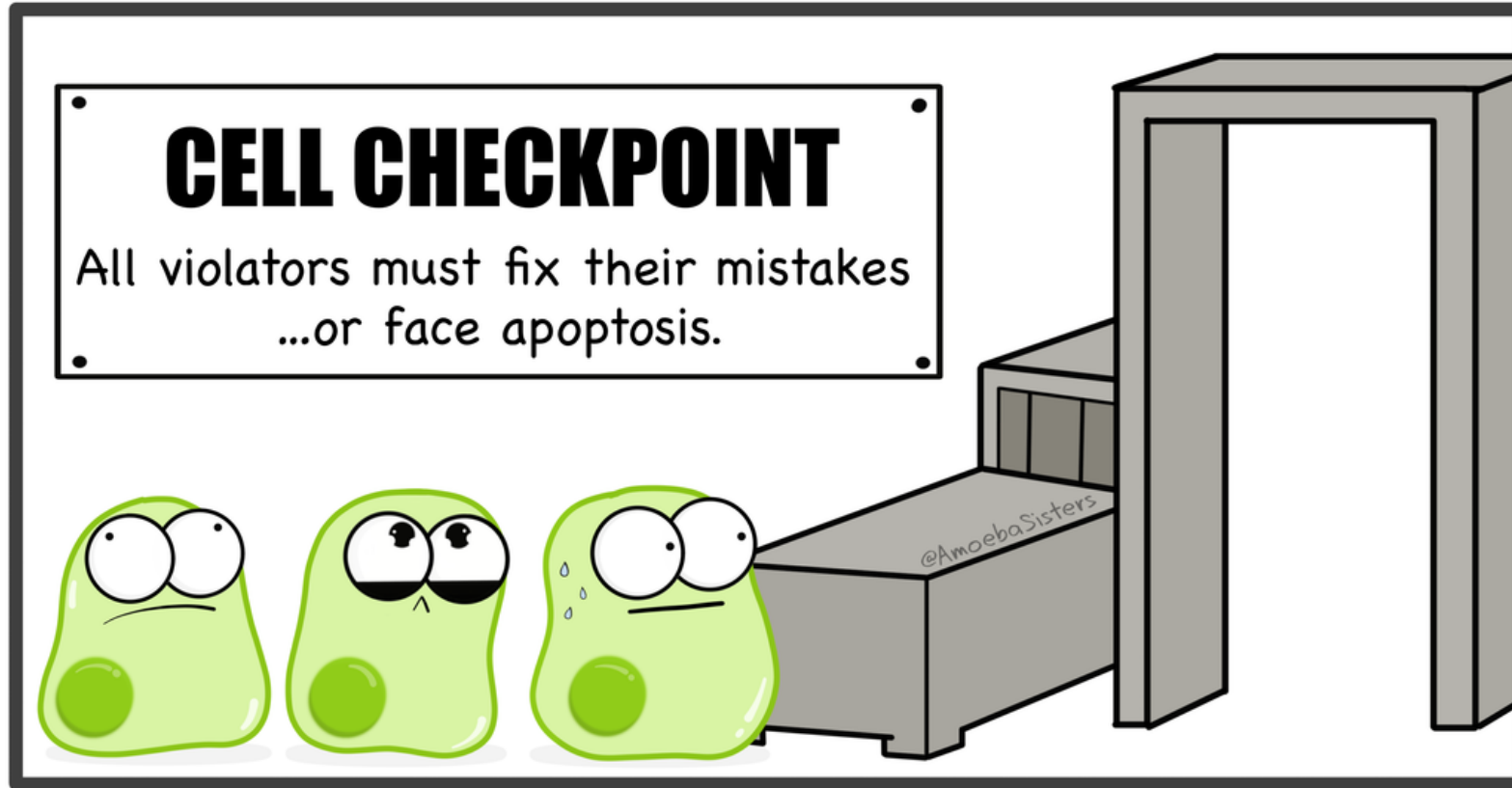
- Replicative senescence studies
- Normally 25-50 (max. 80) divisions – telomere shortening

## – Immortalized/cancer cell lines

- Not restricted by replicative senescence
- Approximate mimic of in vivo regulation in normal healthy cells

## – Genetic manipulation by RNA silencing, CRISPR-Cas9 editing





The cell checkpoints were always a site of intense scrutiny.