

# Study of Interactions and Protein Structure Determination by NMR

For Application to Protein Characterization

by

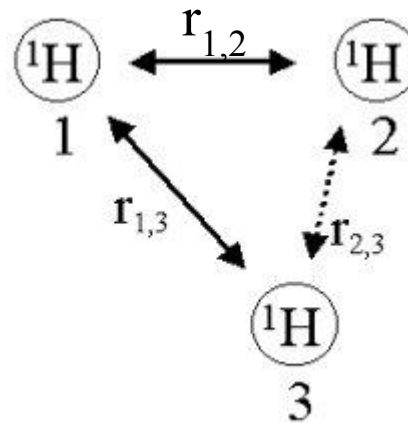
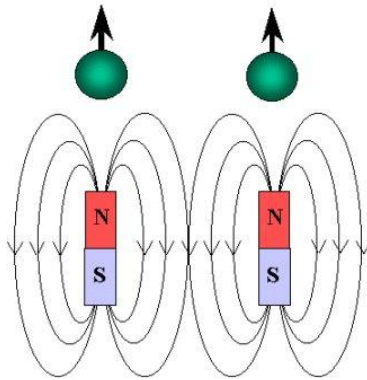
Radovan Fiala, Karel Kubicek, and Pavel Kaderavek  
CEITEC, Masaryk University



# NMR as a tool for study structure, dynamics and interactions of biomolecules

- 0) AA/NA sequence, resonance assignment, standard chemical shifts
- 1) Structure determination of proteins/NAs
- 2) NMR can provide detailed information about the structure at the atomic level resolution relying on the spatial proximity of two interacting protons – nuclear Overhauser enhancement (NOE)
- 3) Additional structural information can be obtained (residual dipolar couplings – RDCs,  $J$ -couplings, backbone chemical shifts - CSI)

NOE:



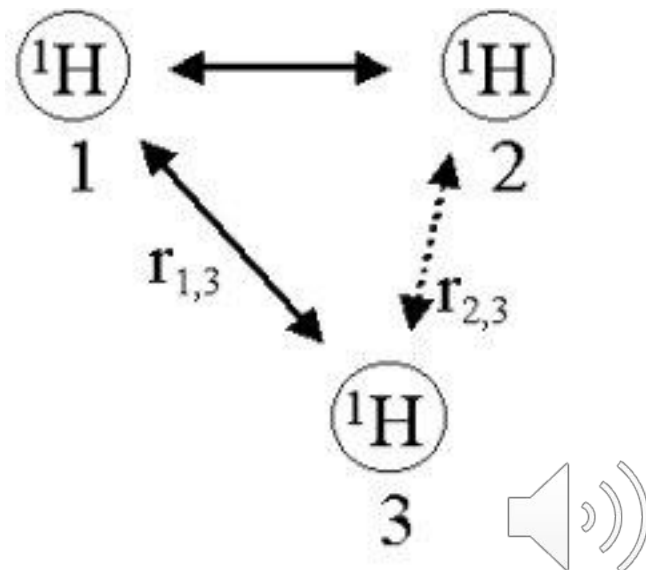
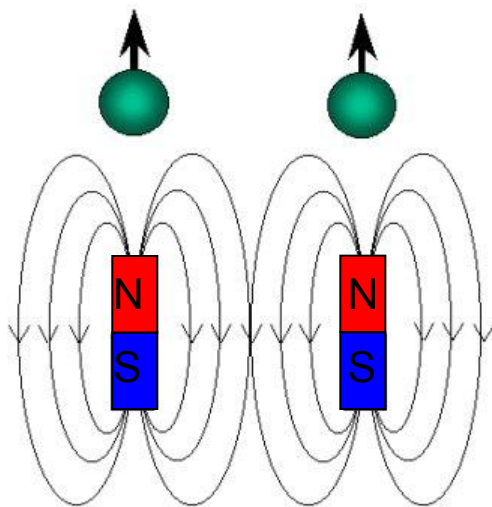
$$r_{1,2}; r_{1,3}; r_{2,3} \leq 6 \text{ \AA}$$

$$1 \text{ \AA} = 1.10^{-10} \text{ m}$$

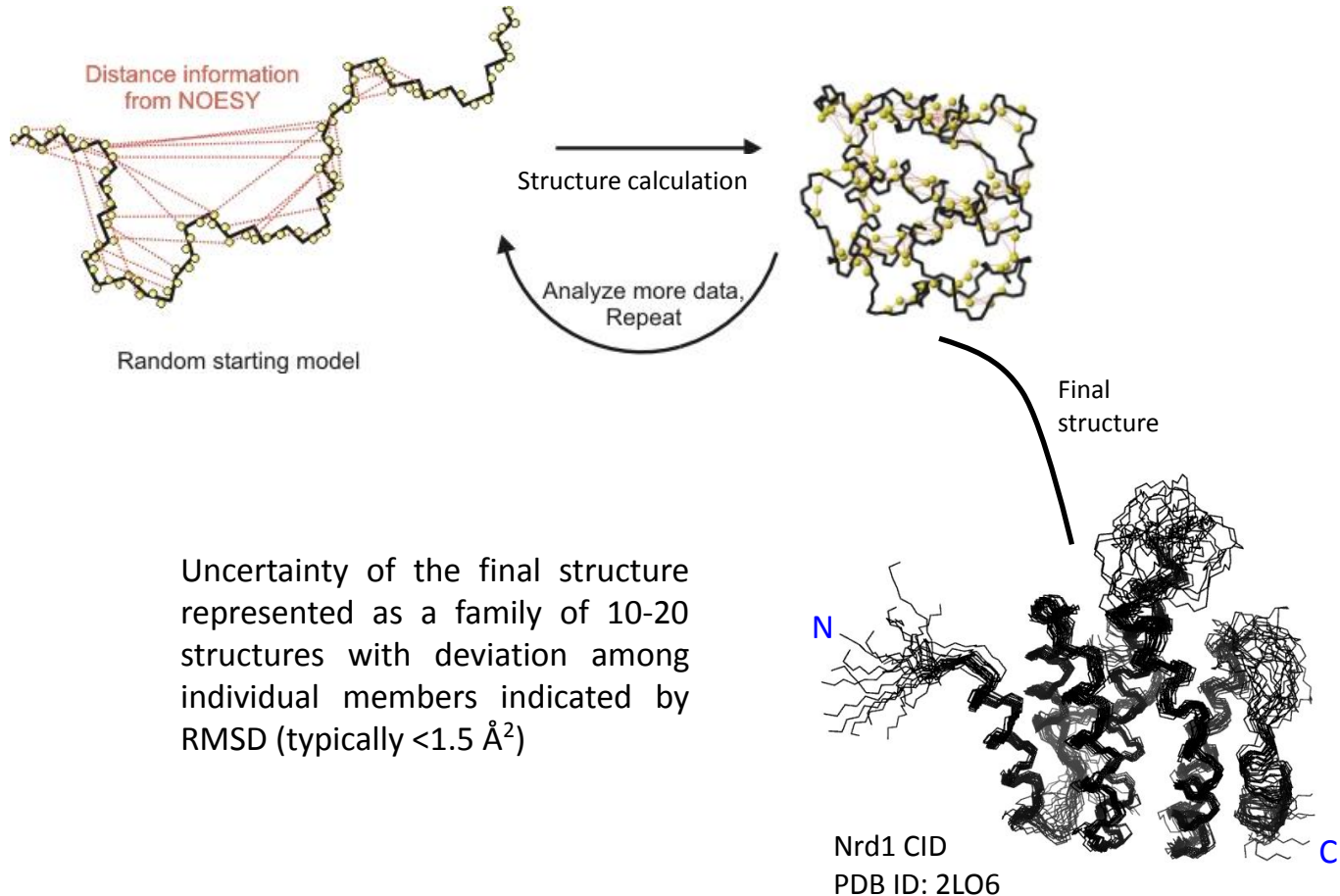


## Nuclear Overhauser Effect (Spectroscopy) = NOE(SY)

- i) caused by dipolar coupling between nuclei.
- ii) the local field at one nucleus is affected by the presence of another nucleus.
- iii) the result is a mutual modulation of resonance frequencies.
- iv) the NOE operates through space.
- v) the intensity of the interaction is a function of the distance between the nuclei according to the following equation:  $I = A(1/r^6)$ ,  $I$  is the intensity,  $A$  is a scaling constant, and  $r$  is the distance between the nuclei
- vi) **the NOE provides a link between an experimentally measurable quantity,  $I$ , and internuclear distance**
- vii) **NOE is only observed up to  $\sim 6\text{\AA}$**



## Iterative procedure of structure determination by NMR

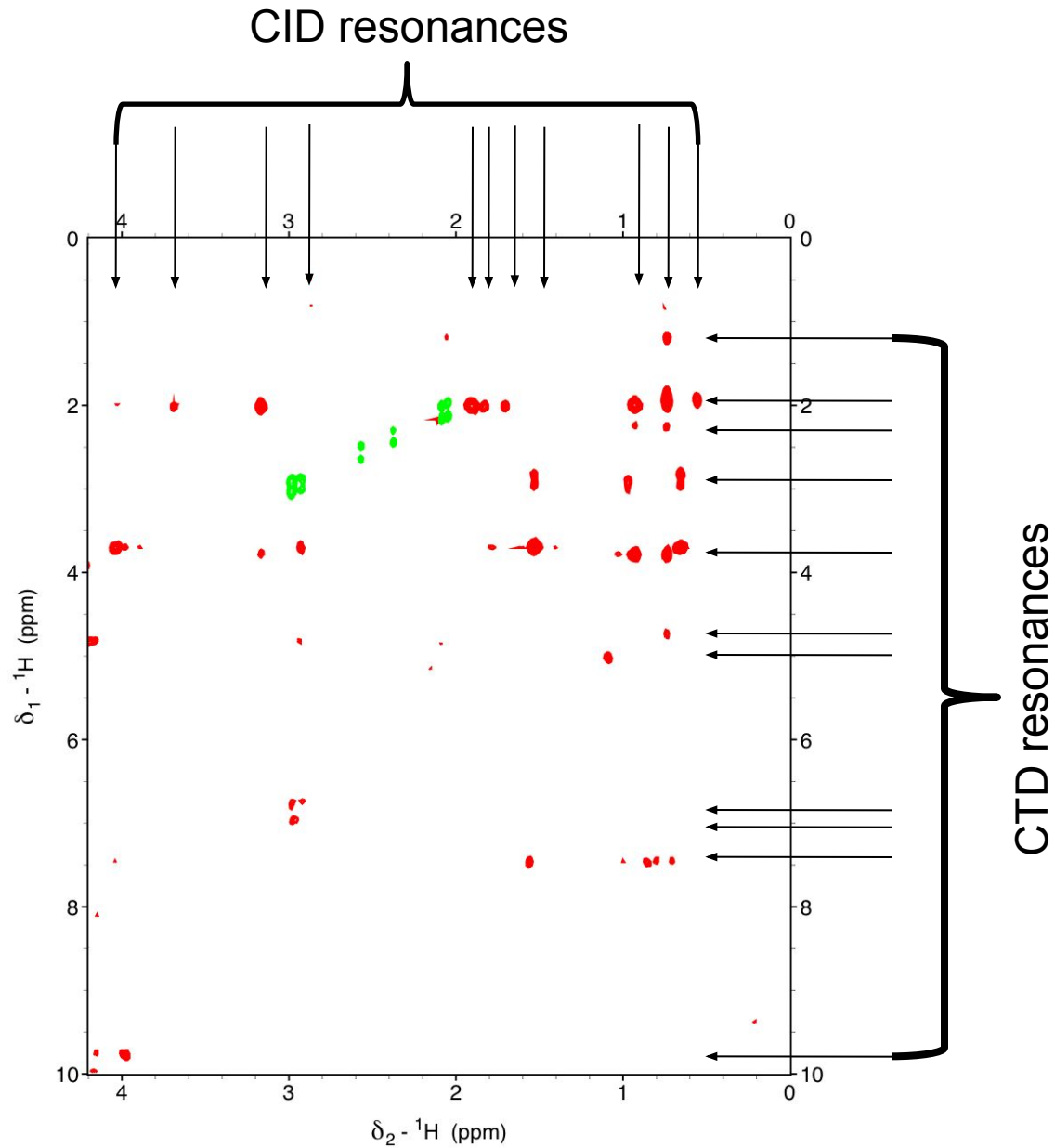


Uncertainty of the final structure represented as a family of 10-20 structures with deviation among individual members indicated by RMSD (typically  $<1.5 \text{ \AA}^2$ )

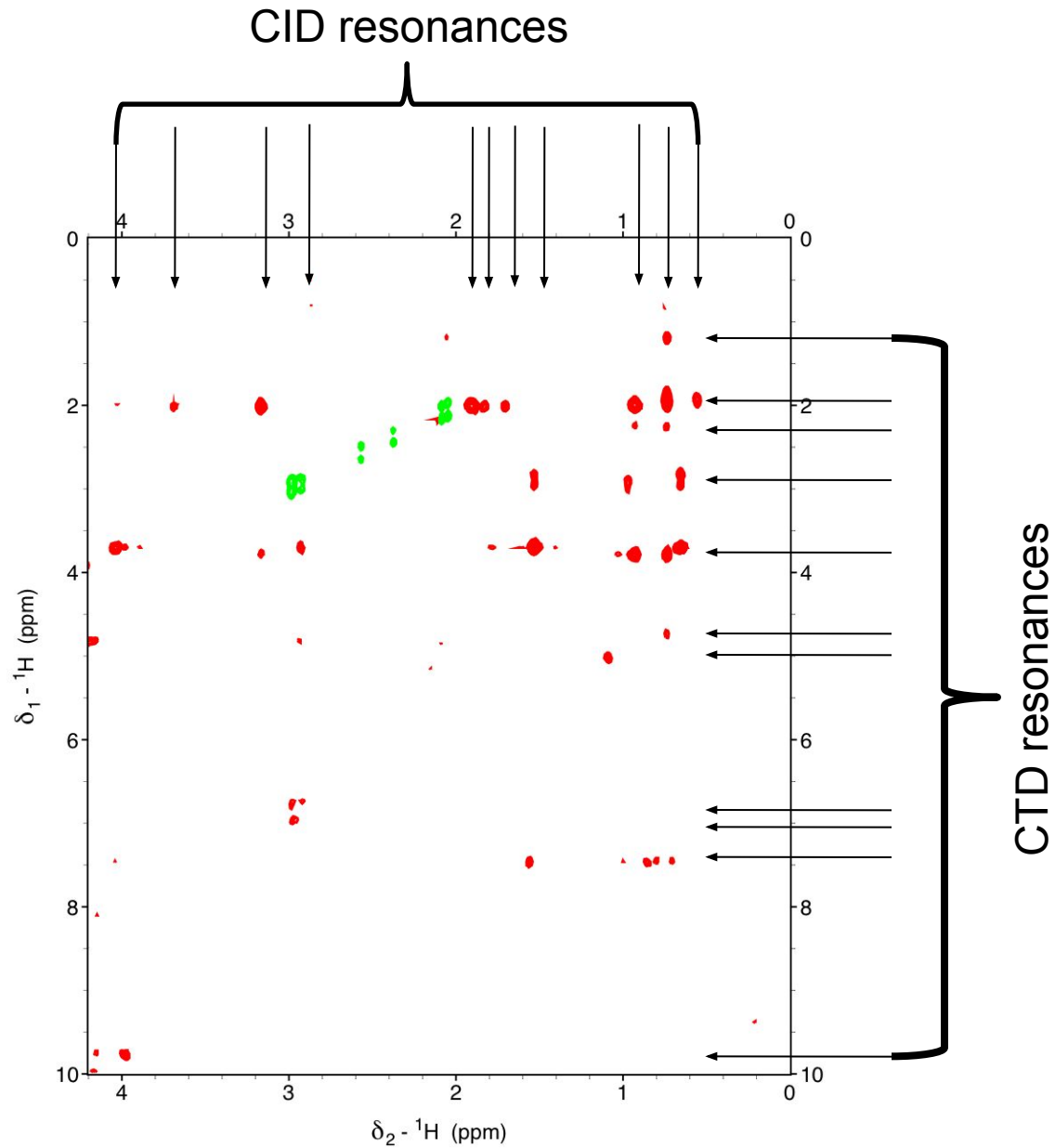
[http://www.fbregents.com/basics\\_nmr/9proteins.htm](http://www.fbregents.com/basics_nmr/9proteins.htm)



# Interligand NOEs between CID and CTD – 900MHz, 150ms, 293K

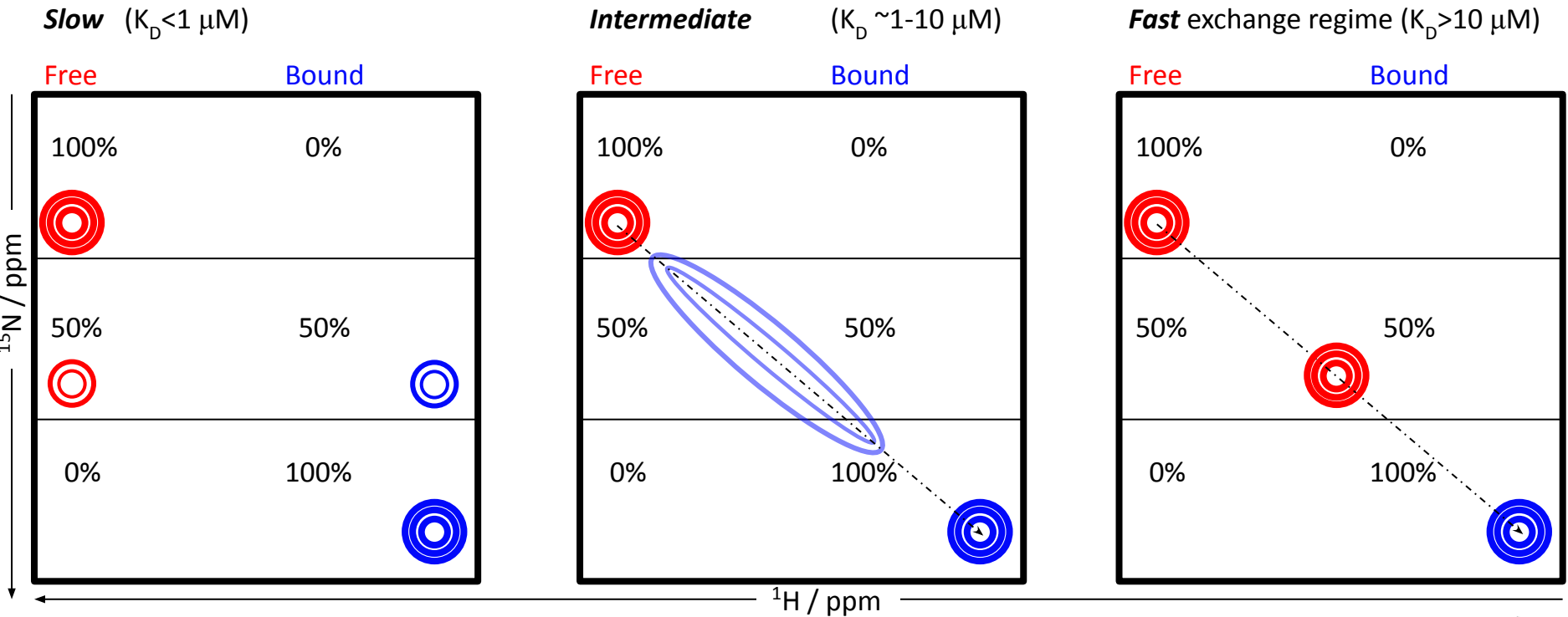


# Interligand NOEs between CID and CTD – 900MHz, 150ms, 293K

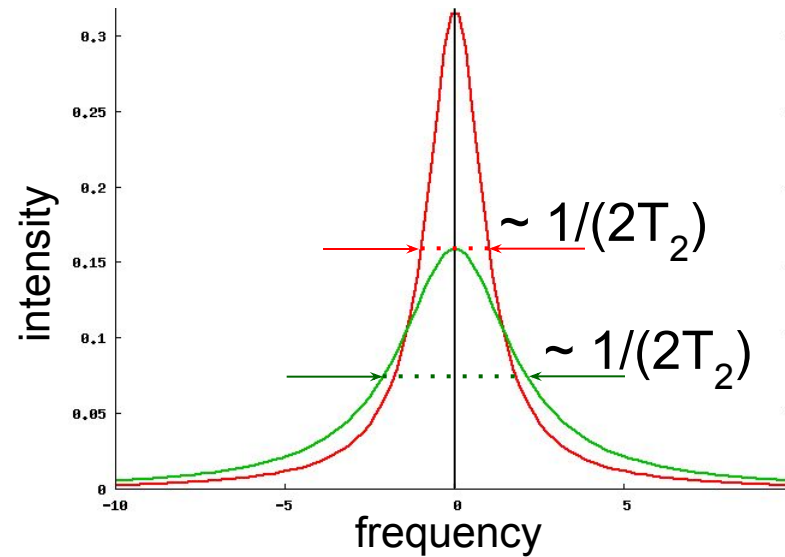
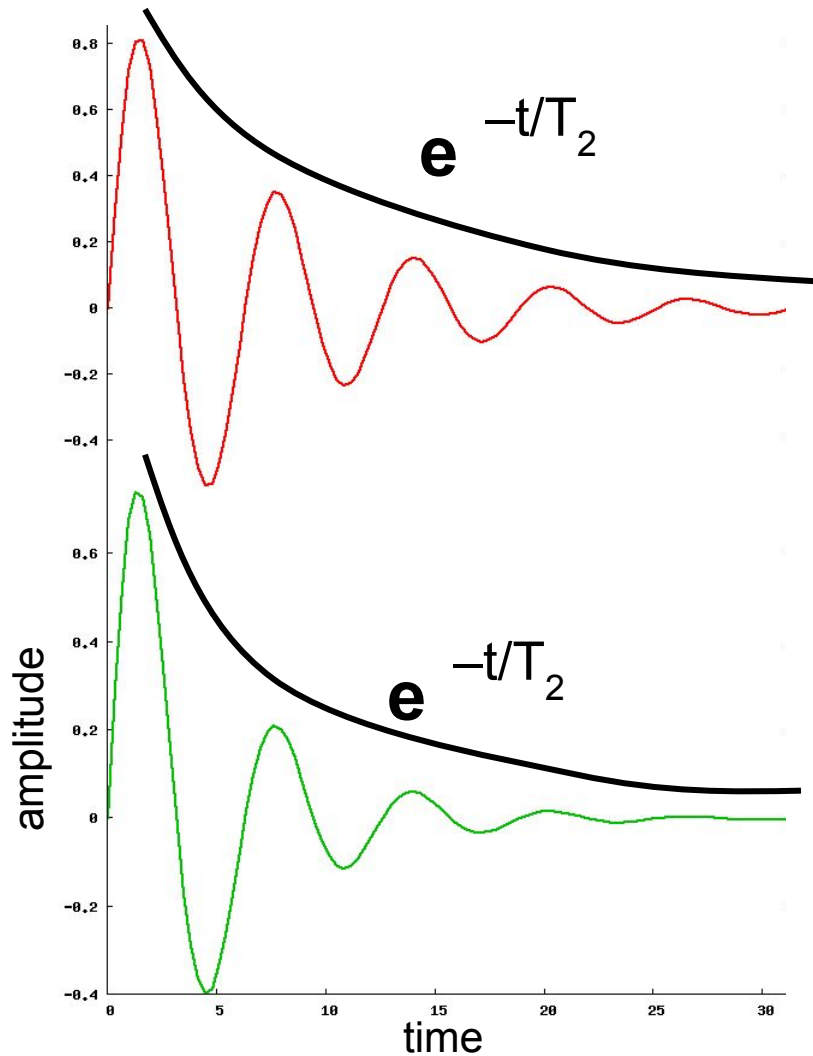


# Studying interactions by NMR titration

- 1) **Slow** exch. regime (on the NMR timescale) – individual peaks for each of the studied states (e.g. free / complexed forms of a protein), peak intensity representing population of a given state
- 2) **Intermediate** exchange regime – single peak whose chemical shift position is given by the molar ratio of the states present in solution
- 3) **Fast** exchange regime



## Not all molecules relax (decay) with the same rate



**Bigger molecules** (higher molecular weight) relax faster  $\Rightarrow$  **broad peaks**

**Small molecules** relax slower  $\Rightarrow$  **narrow peaks**





Size

Relaxation



slow (i.e. long  $t_2$  time)

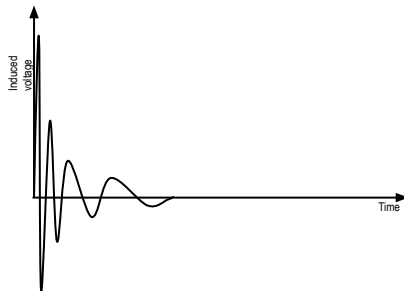
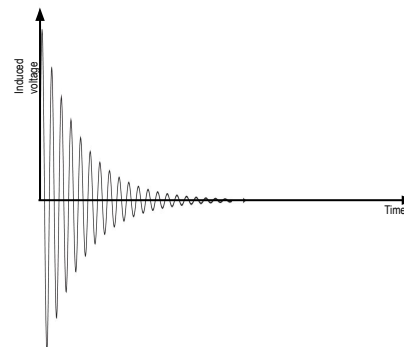
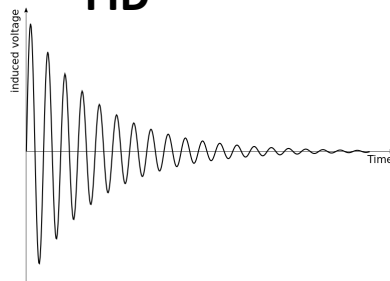


medium

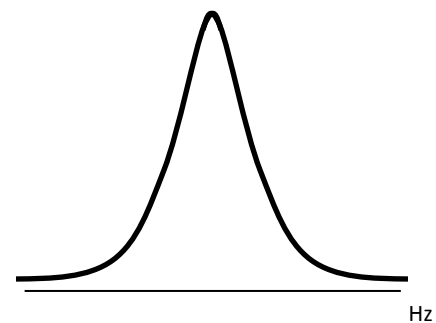
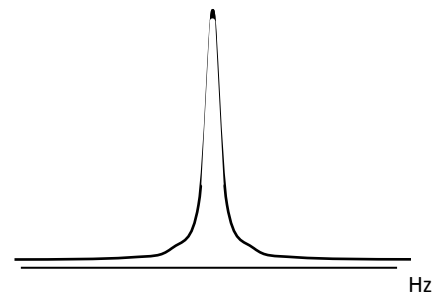


fast

FID



NMR line(width) after FT



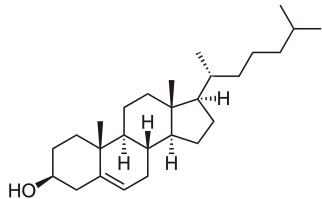
## Size

## Relaxation

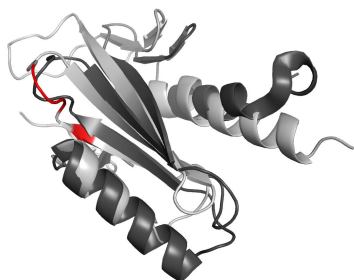
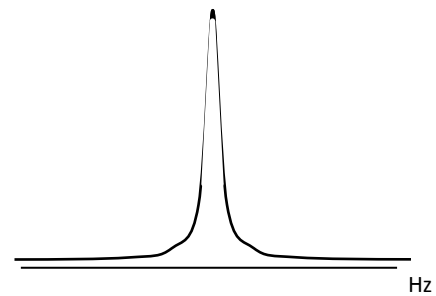
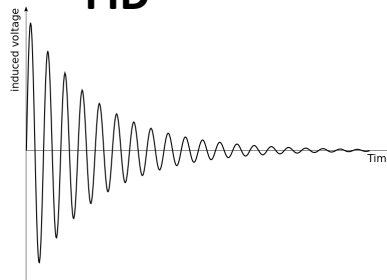
## FID

## NMR line(width) after FT

slow (i.e. long  $t_2$  time)

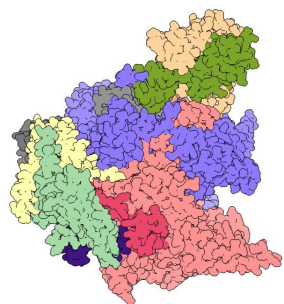
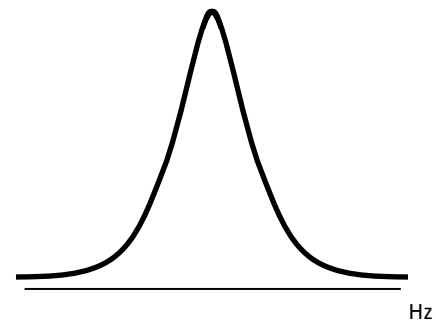
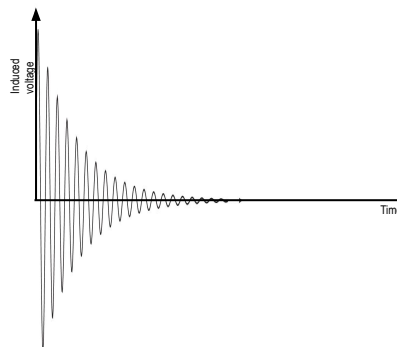


e.g. Cholesterol



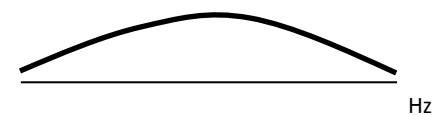
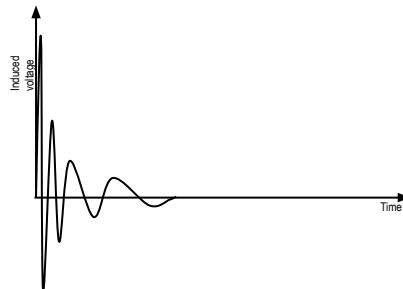
medium

Biomolecules 5-30 kDa



fast

Large molecules 50+ kDa

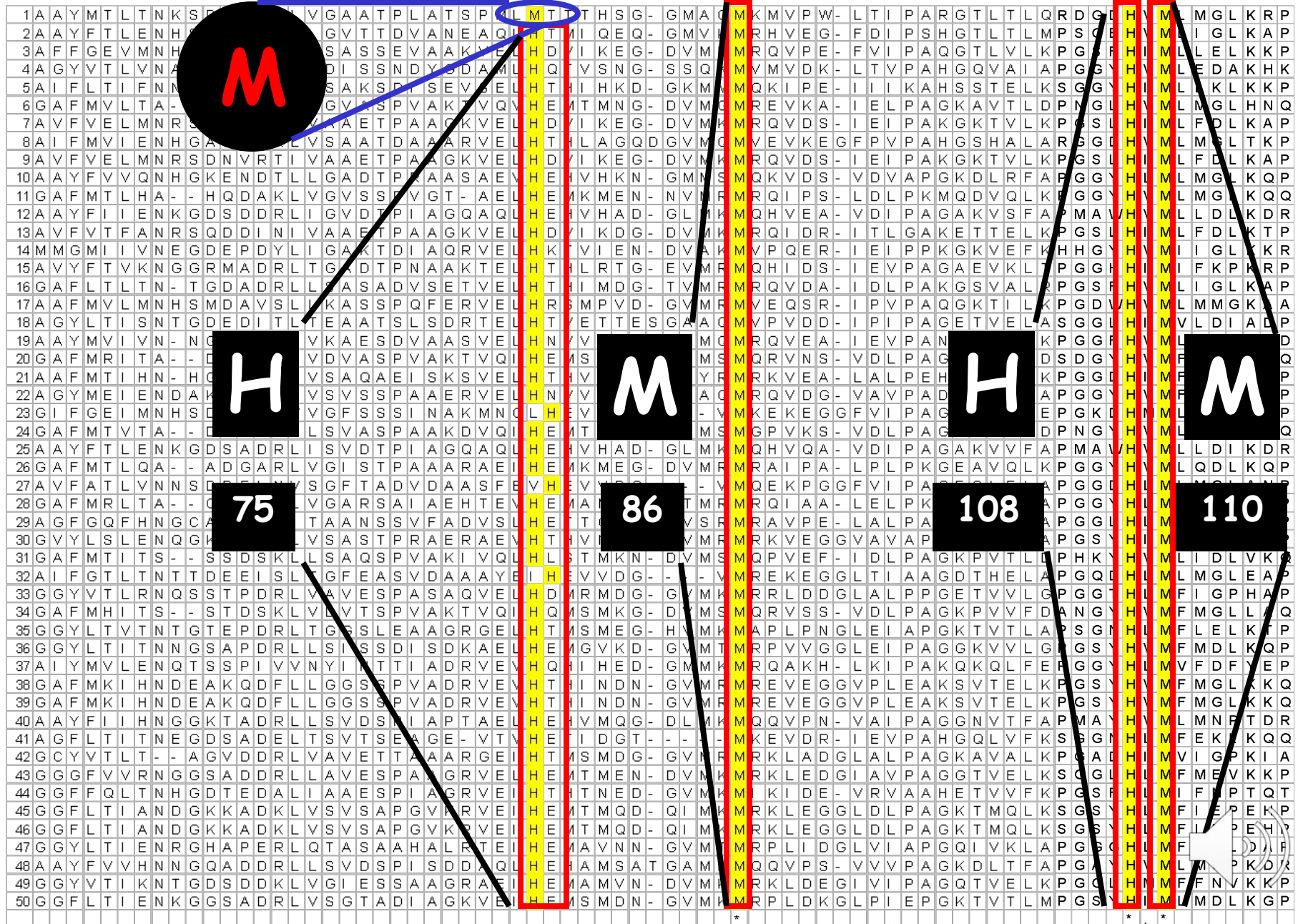


# **Protein – metal ion interaction**

slow exchange case



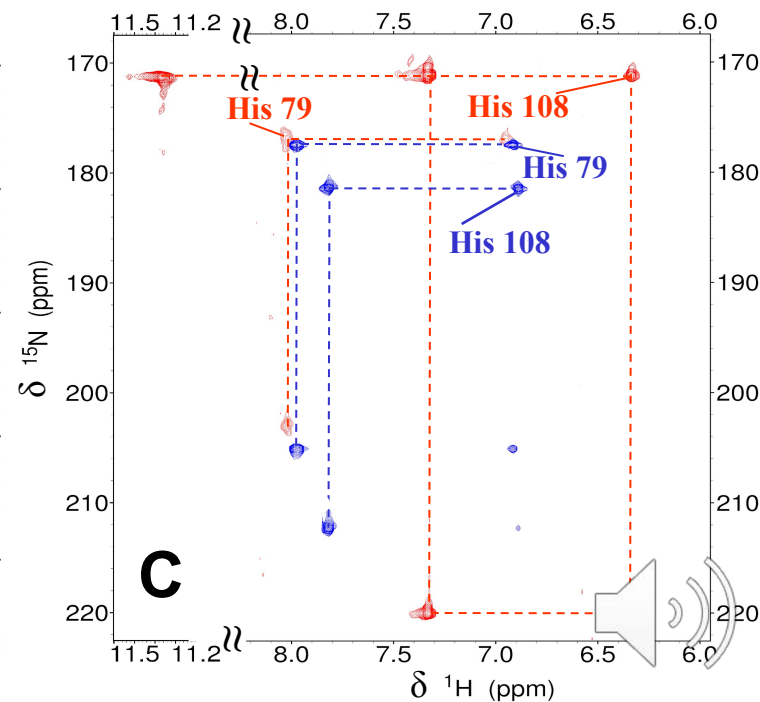
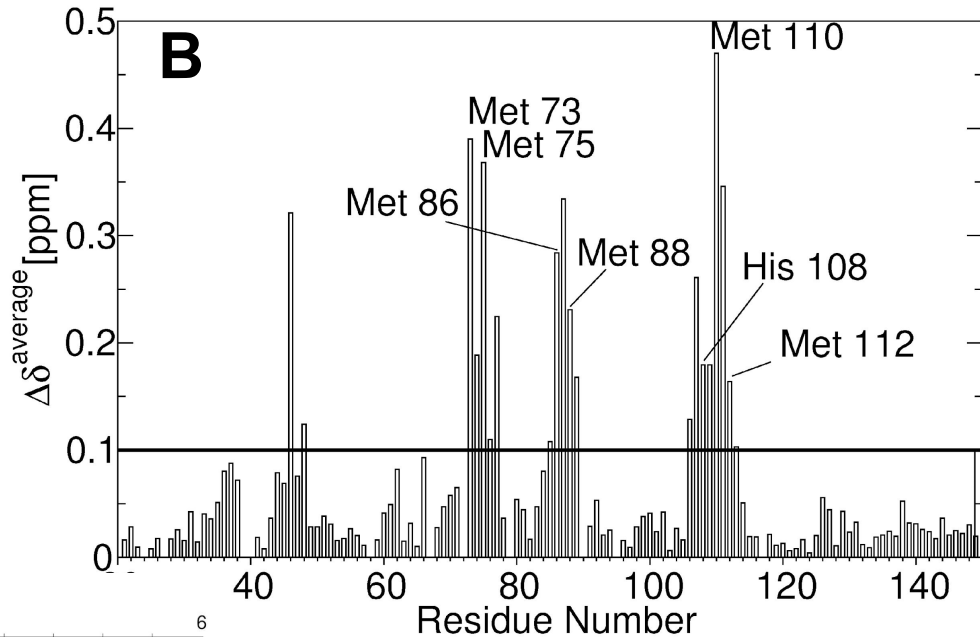
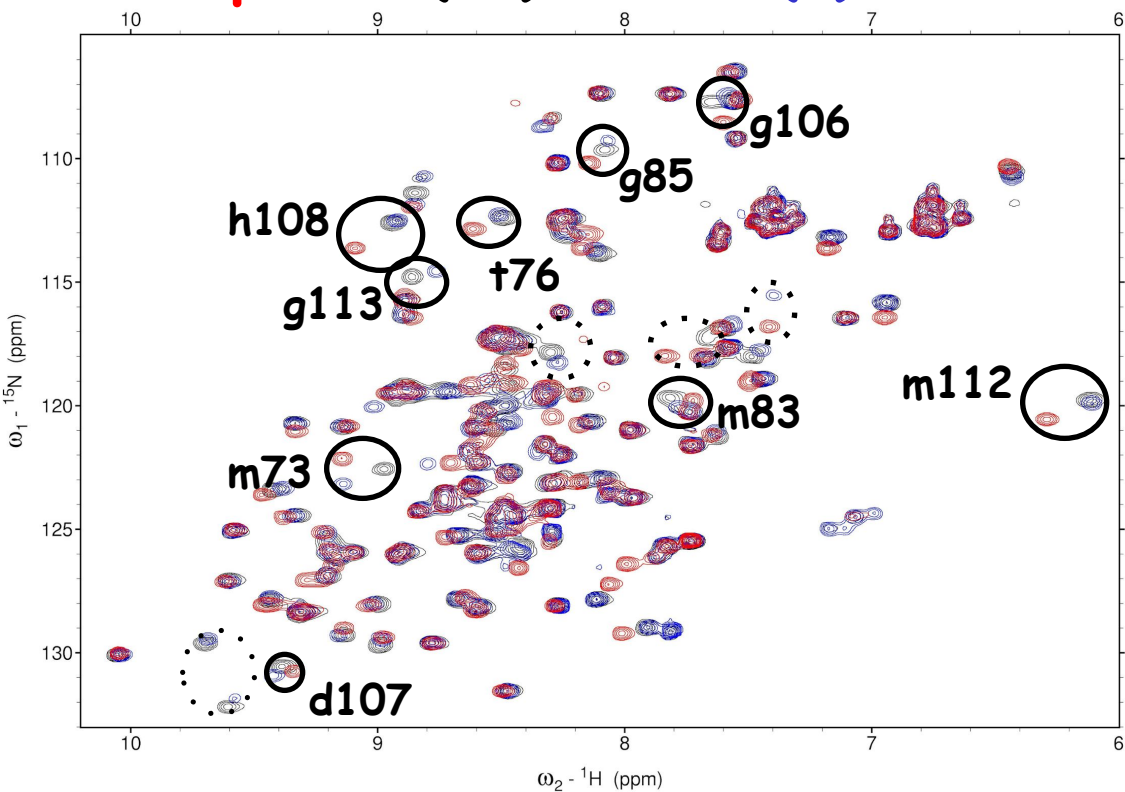
# DR1885 from *deinococcus radiodurans*

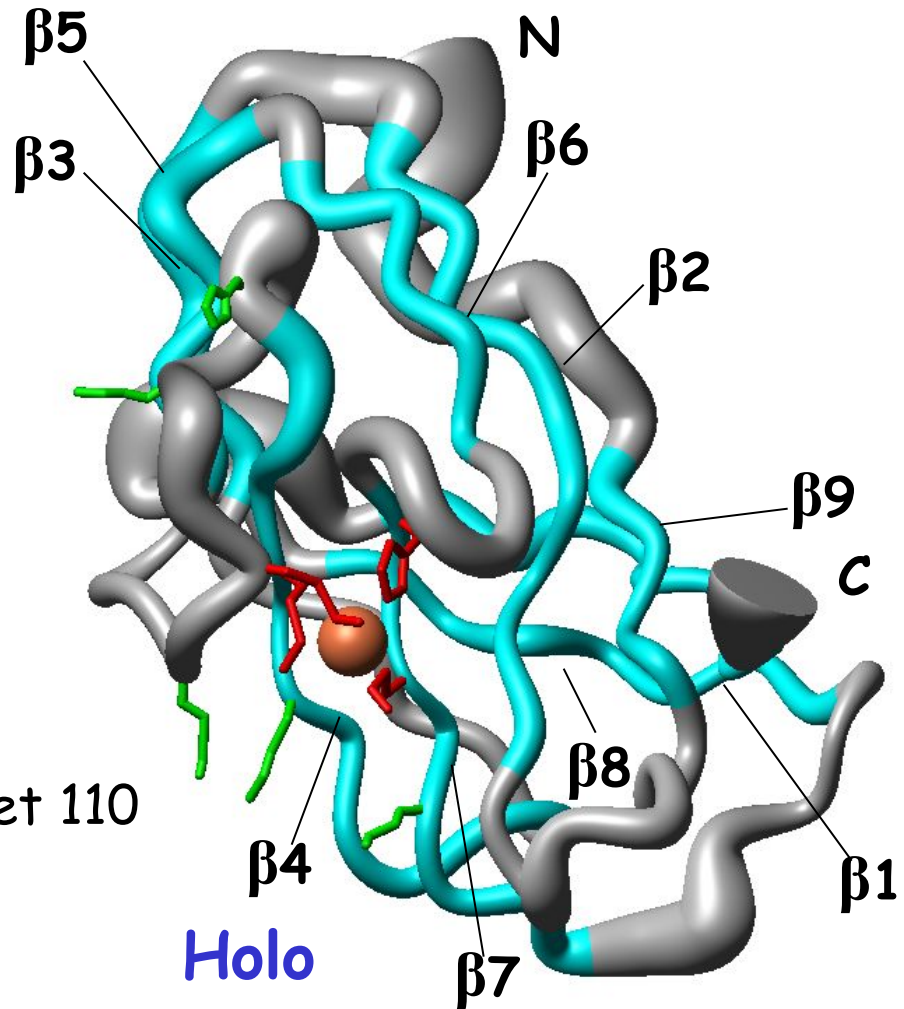
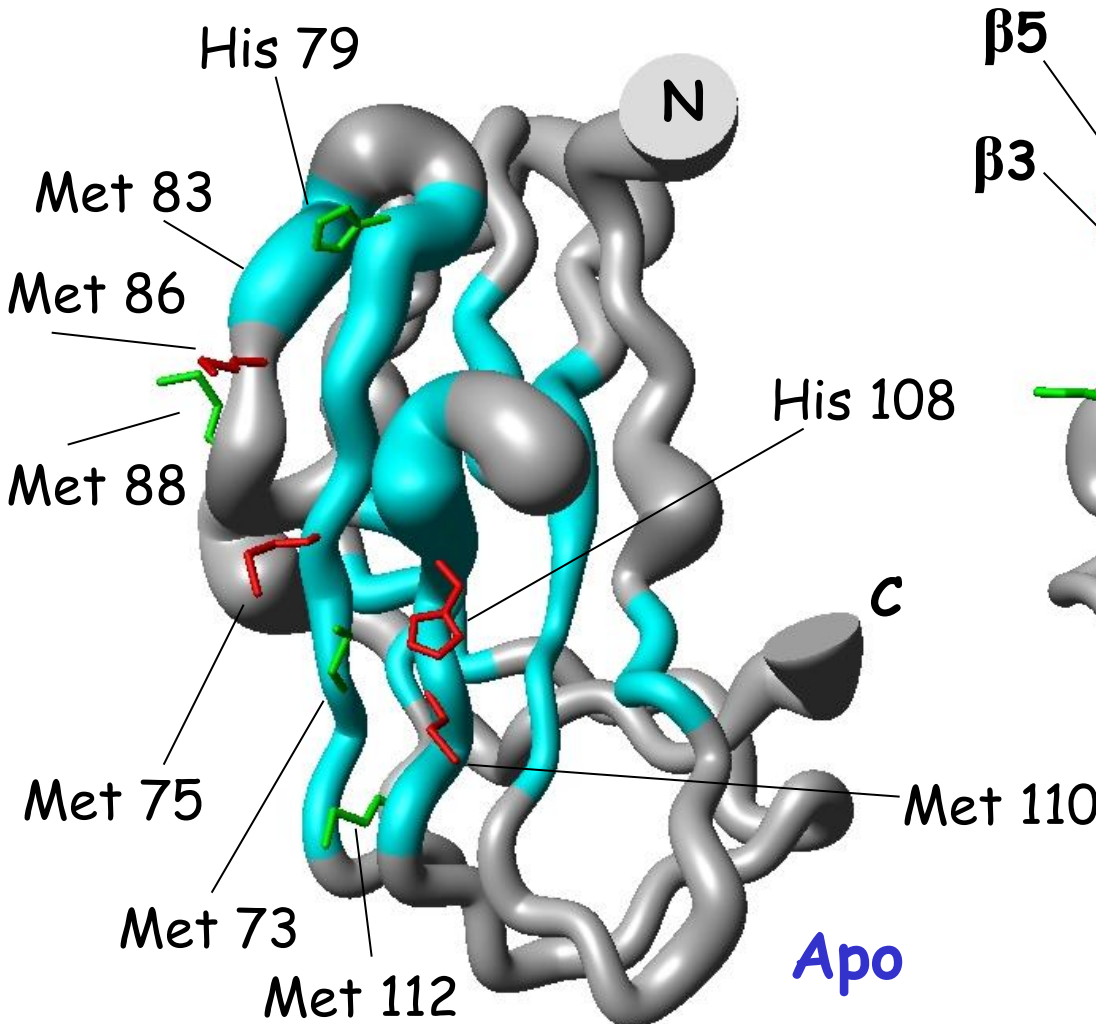


# Interaction of DR1885 with copper

-titration (A,B)  
-<sup>2</sup>J HSQC (C)

**Apo**, **Cu(II)** and **Cu(I)**





— Conserved **Met&His**  
— Other **Met&His** residues

cyan -  $\beta$ -sheets  
grey - random coil



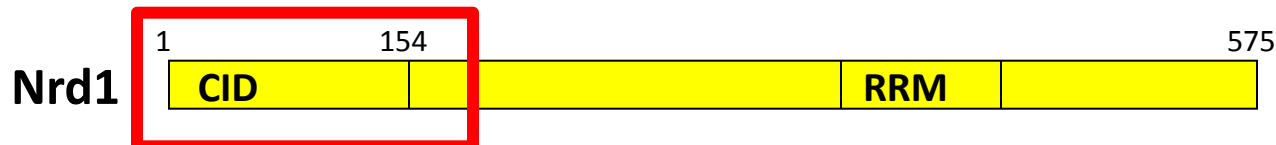
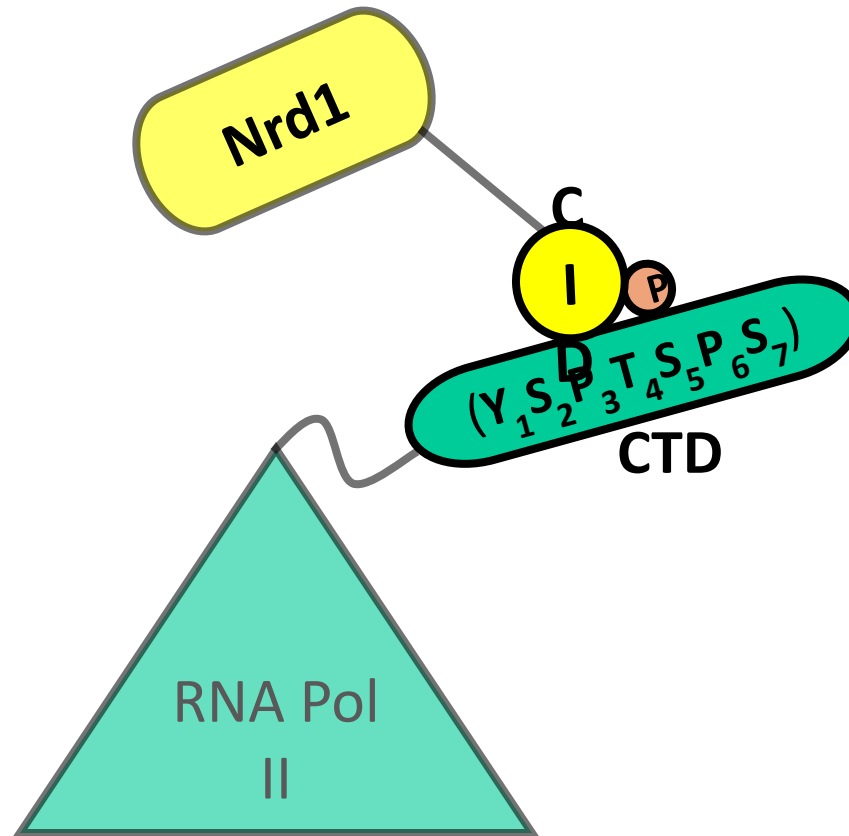
# **Protein – peptide interaction**

fast exchange case





# Interaction of Nrd1-CID with CTD

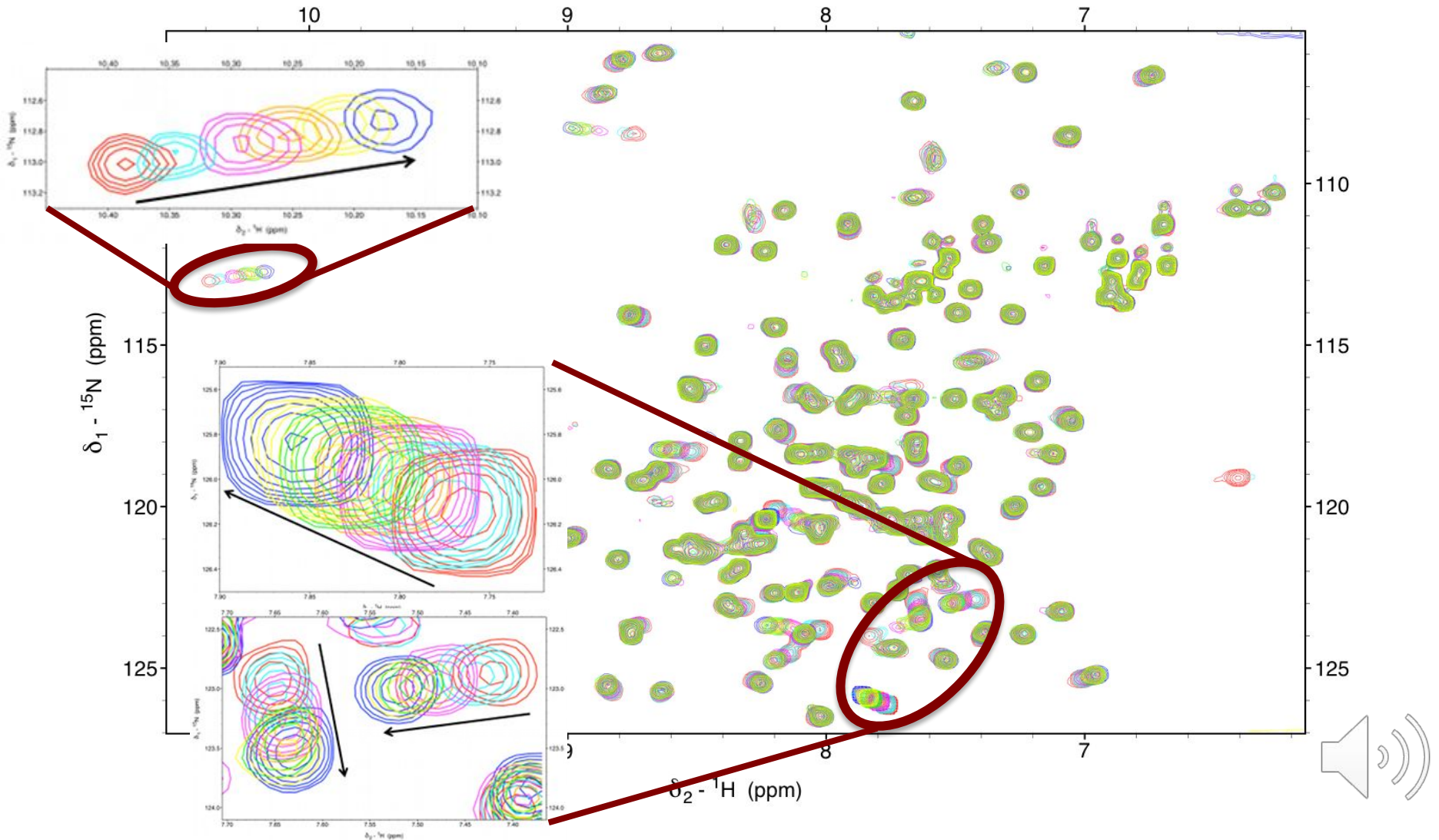


RRM: RNA recognition motif; CID: CTD interaction domain; CTD: C-terminal domain



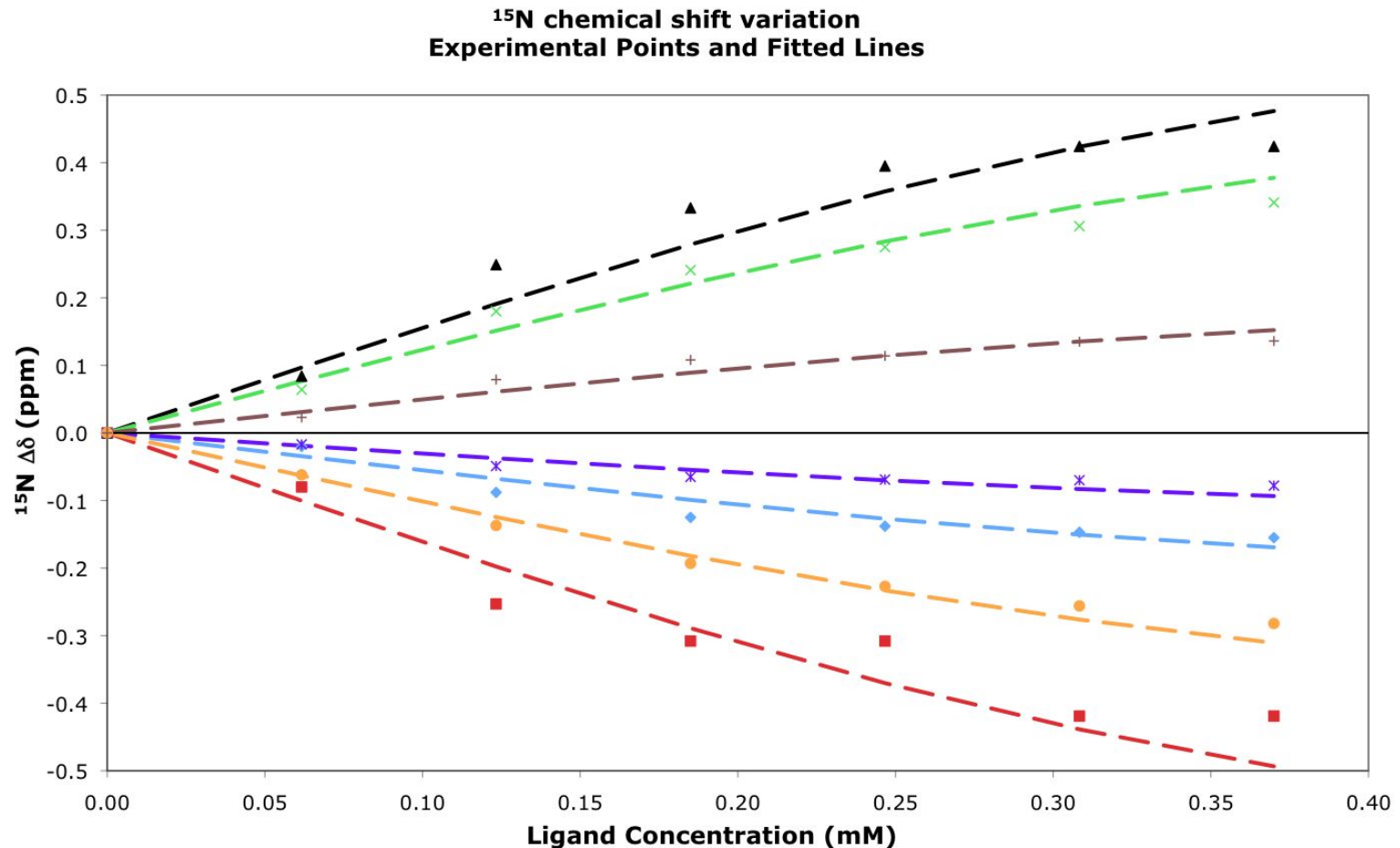


Interaction of  $^{15}\text{N}$  enriched CID with unlabeled CTD-Ser5P in  $n$ -steps,  $n=6$  in our case - peaks corresponding to the interacting residues of CID change their chemical shift (position in the spectrum) => interaction surface, binding constant, stoichiometry

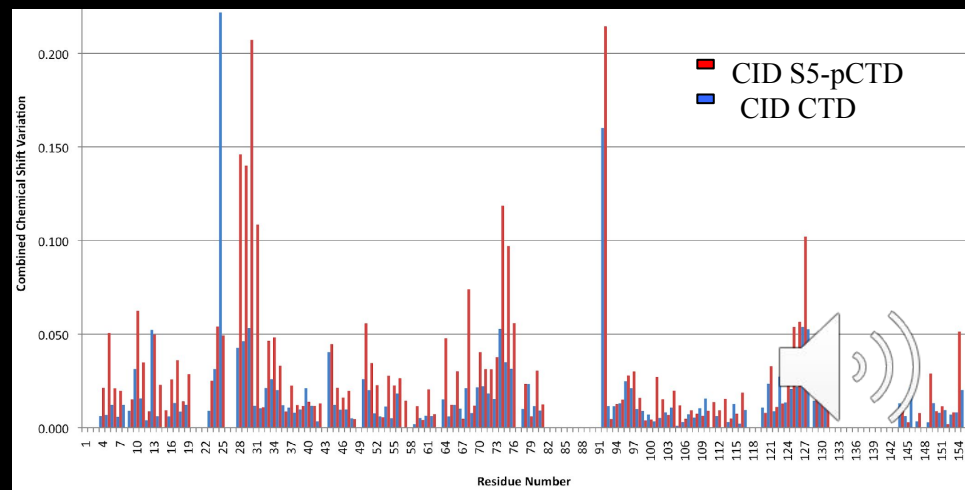
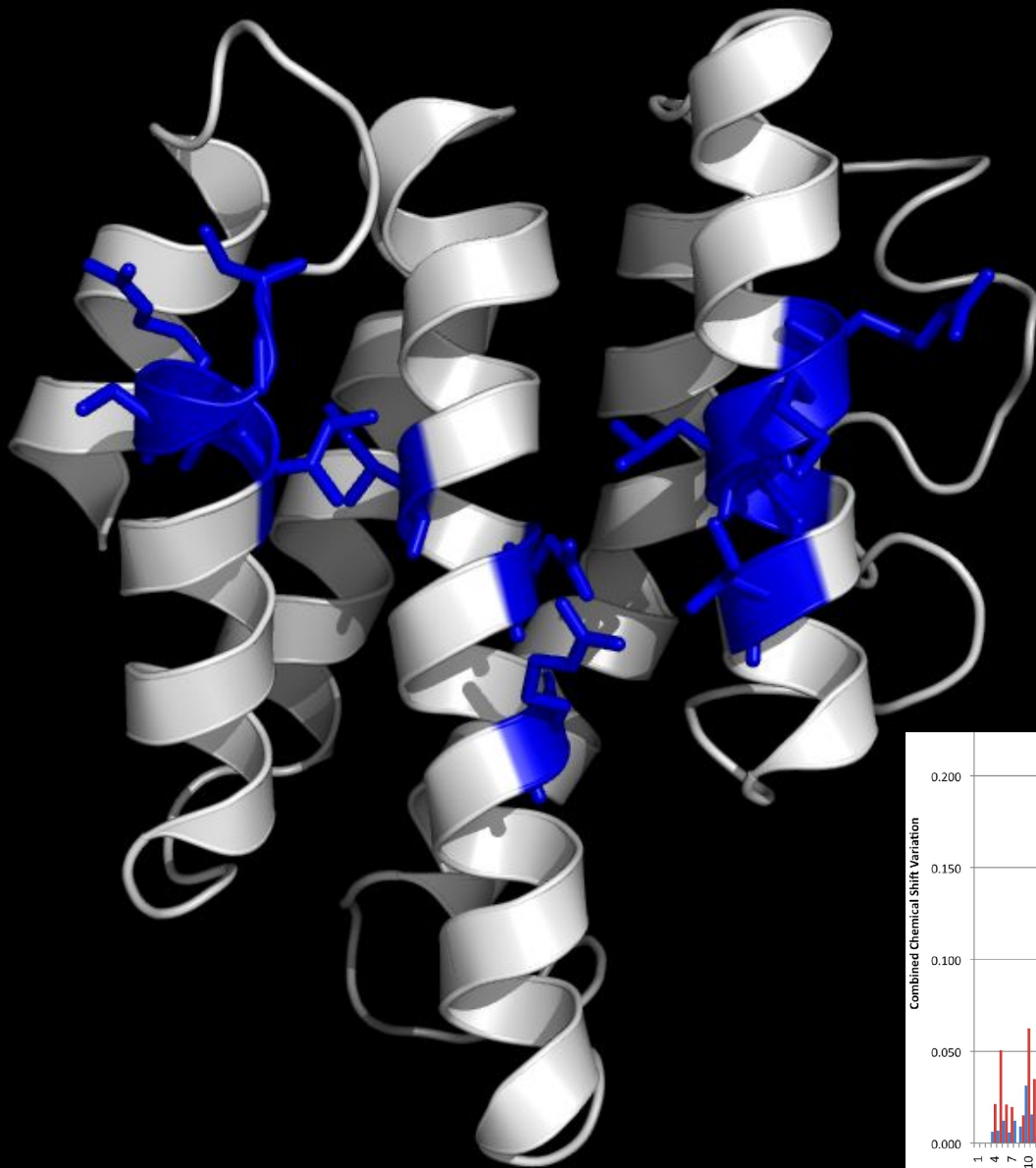


# Interaction of Nrd1-CID with CTD

- NMR Titration:  $\sim 0.6 \text{ mM } ^{15}\text{N}$  enriched CID +  $\sim 0.8 \text{ mM } (\text{YSPT}_{\text{pS}}\text{PS})_2$   
 $\sim 0.6 \text{ mM } ^{15}\text{N}$  enriched CID +  $\sim 0.8 \text{ mM } (\text{YSPT}_{\text{S}}\text{PS})_2$
- $\mu\text{M}$ - $\text{mM}$  range of interaction ->
  - > fast exchange regime on NMR time-scale
  - NMR-derived  $K_d = 0.080 \text{ mM}$  and  $35 \text{ mM}$



# Nrd1 CID interaction surface — CID residues experiencing the largest chemical shift variations upon the interaction with 5-phospho-Ser CTD shown in blue with side-chains in stick representation



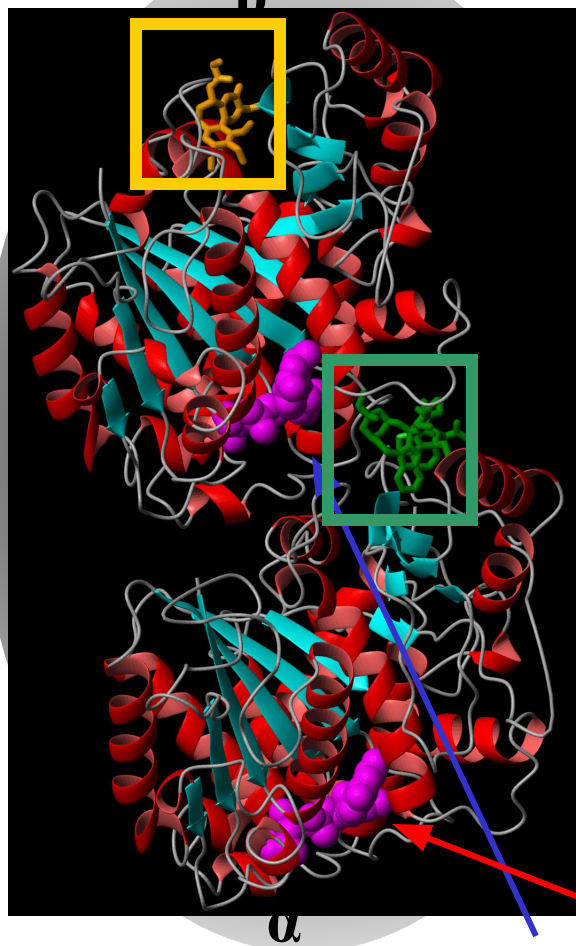
# **Protein – peptide interaction**

drug-receptor case



# *Tubulin* - successful target for anticancer therapy

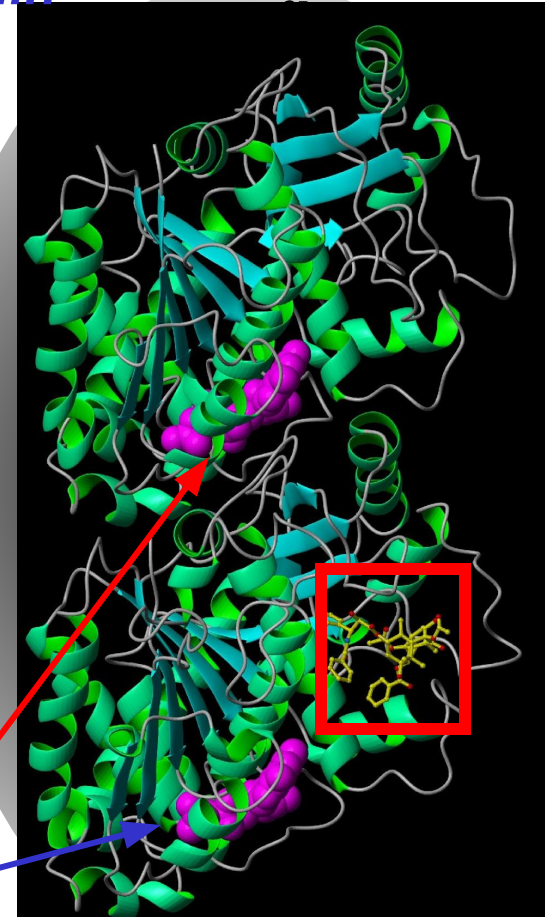
## *Binding sites on tubulin*



**Colchicine**

**Vinca**

**Taxane**



**GDP**

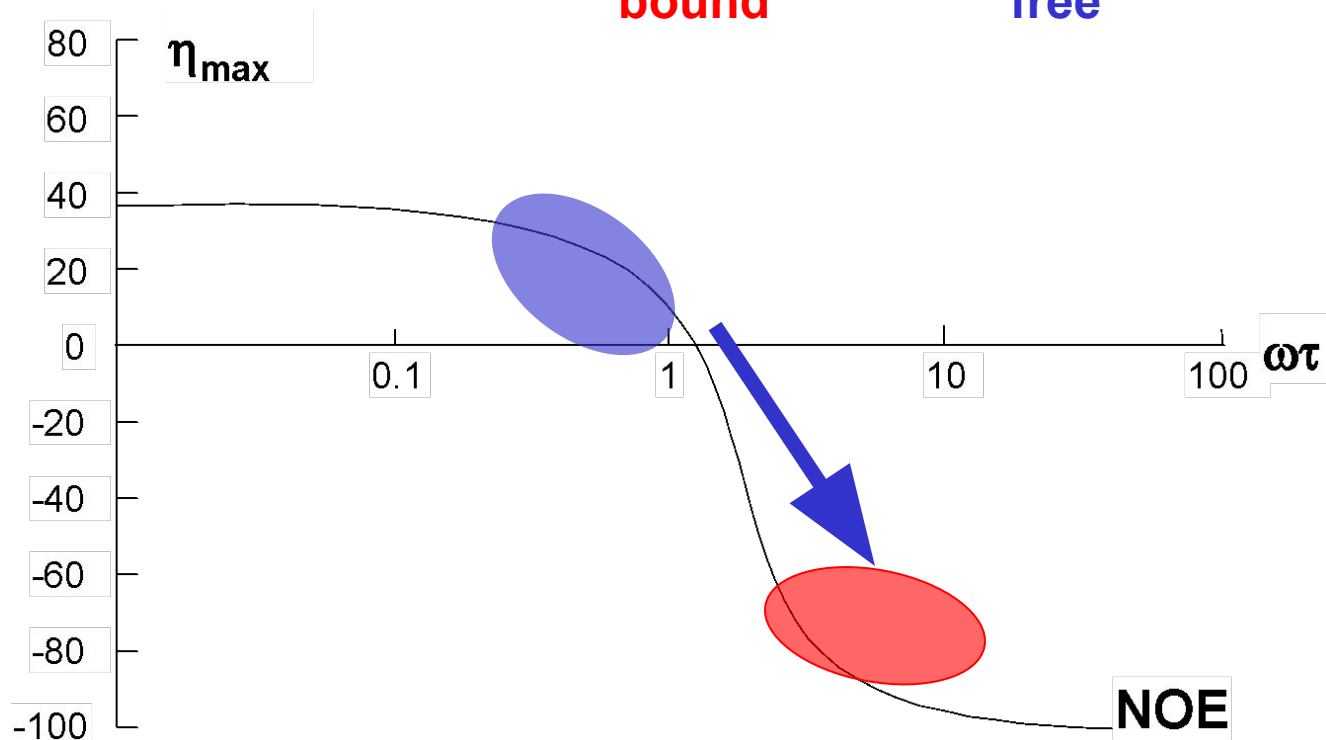
**GTP**

# Transferred-NOE

$$\text{NOE} = p_{\text{bound}} \cdot \text{NOE}_{\text{bound}} + p_{\text{free}} \cdot \text{NOE}_{\text{free}}$$

$$\tau_{\text{c,bound}} \gg \tau_{\text{c,free}} \quad (\text{and } p_{\text{L,free}} \gg p_{\text{L,bound}})$$

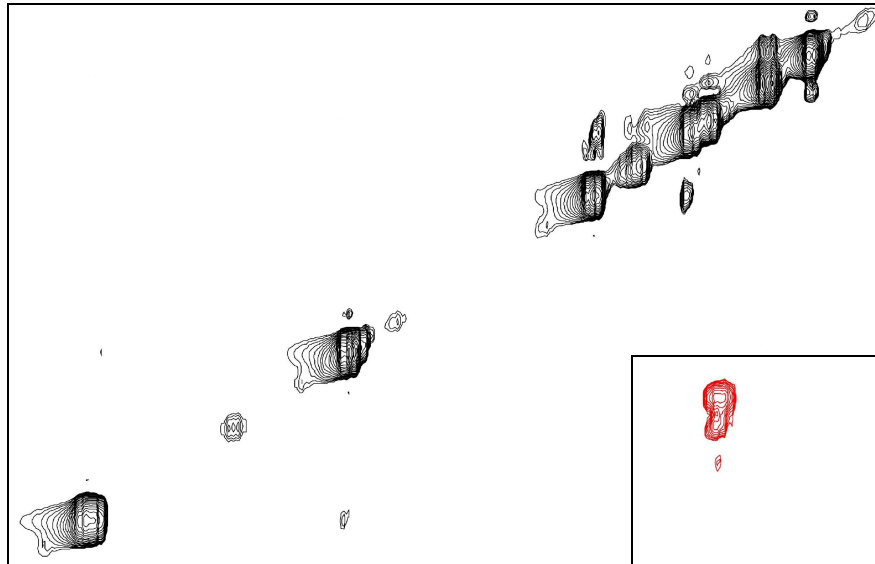
$$\text{NOE}_{\text{bound}} > \text{NOE}_{\text{free}}$$



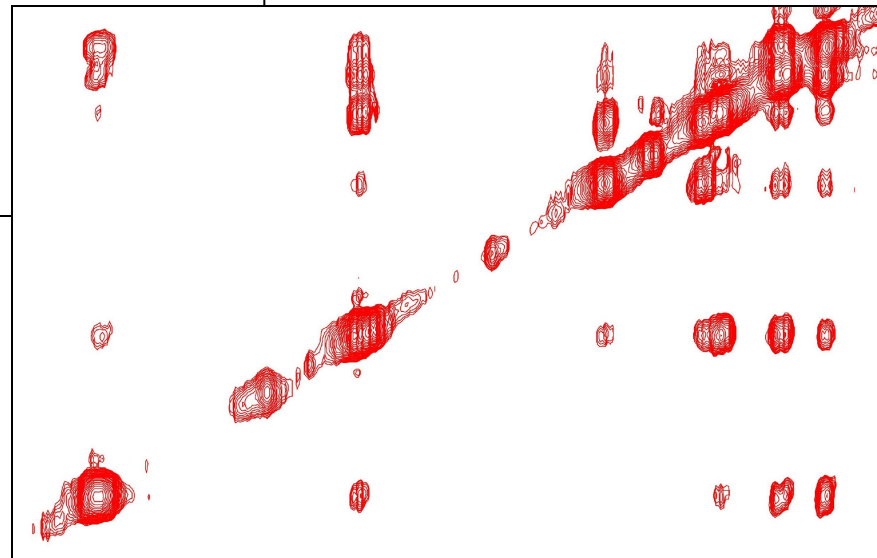


## Transferred NOE Experiments

tr-NOESY ~500 $\mu$ M tubulysin (TBS) **without** and **with** ~10 $\mu$ M tubulin



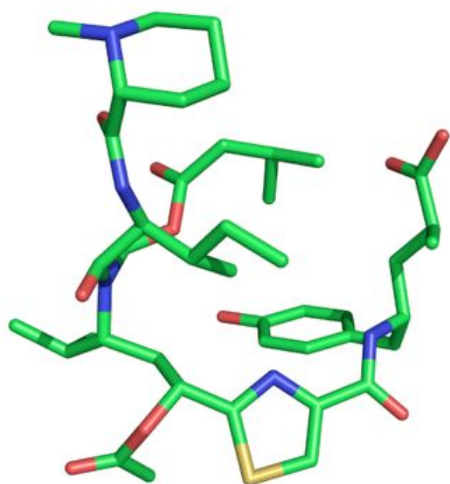
900MHz,  
mixing time=100ms



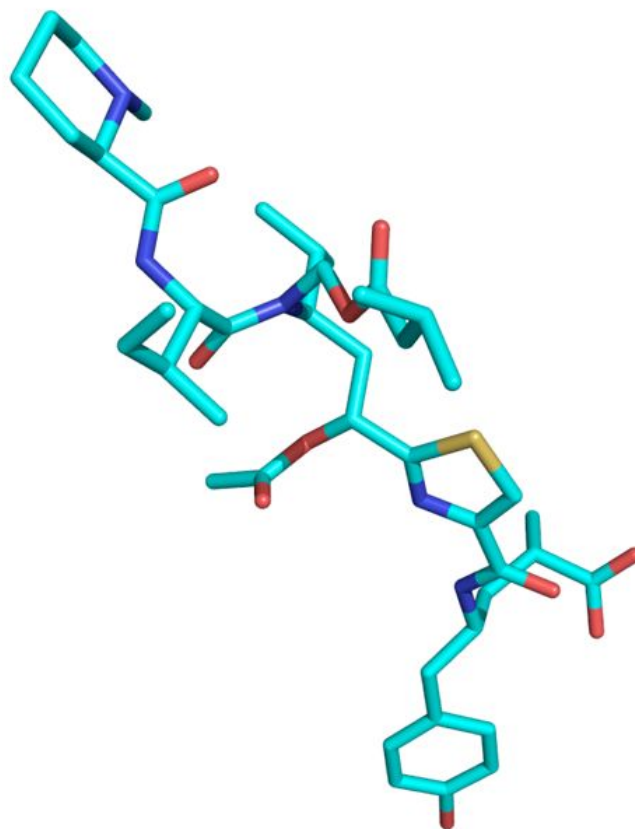
↑  
tbs w/o tub  
tbs:tub 50:1



A



B



Conformation of the **tubulin-bound** - NMR (**A**) and **free** – X-Ray TBS (**B**)



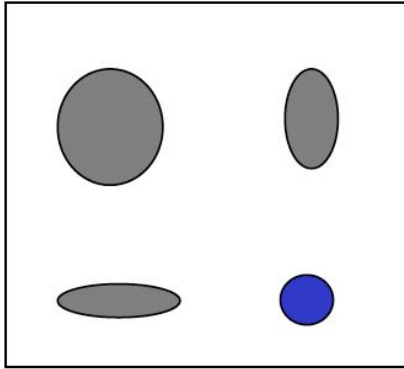


# **Large biomolecules and their interactions**

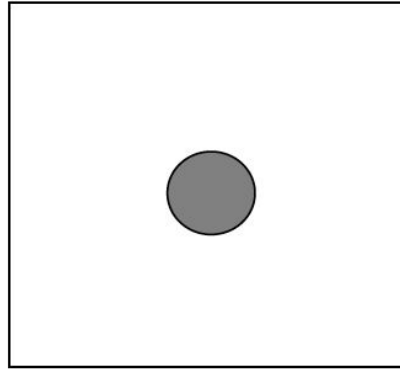


# TROSY-based NMR experiments to study complexes up to 900kDa

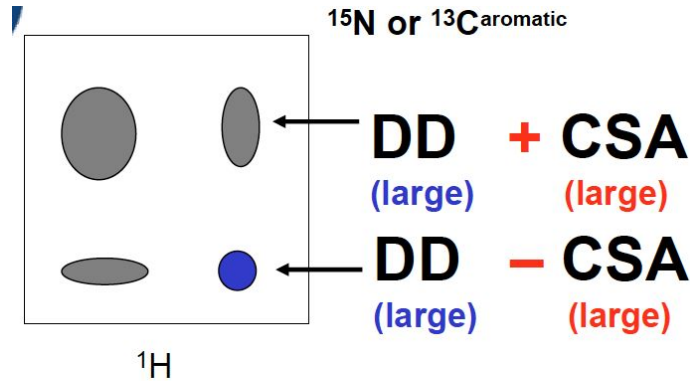
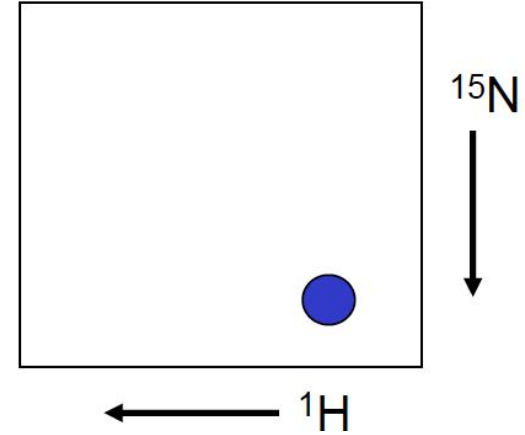
Coupled HSQC



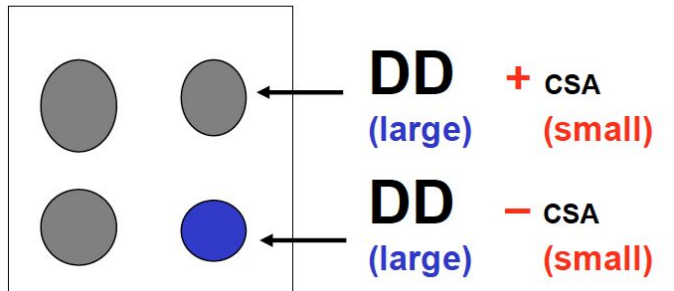
Decoupled HSQC



TROSY-HSQC



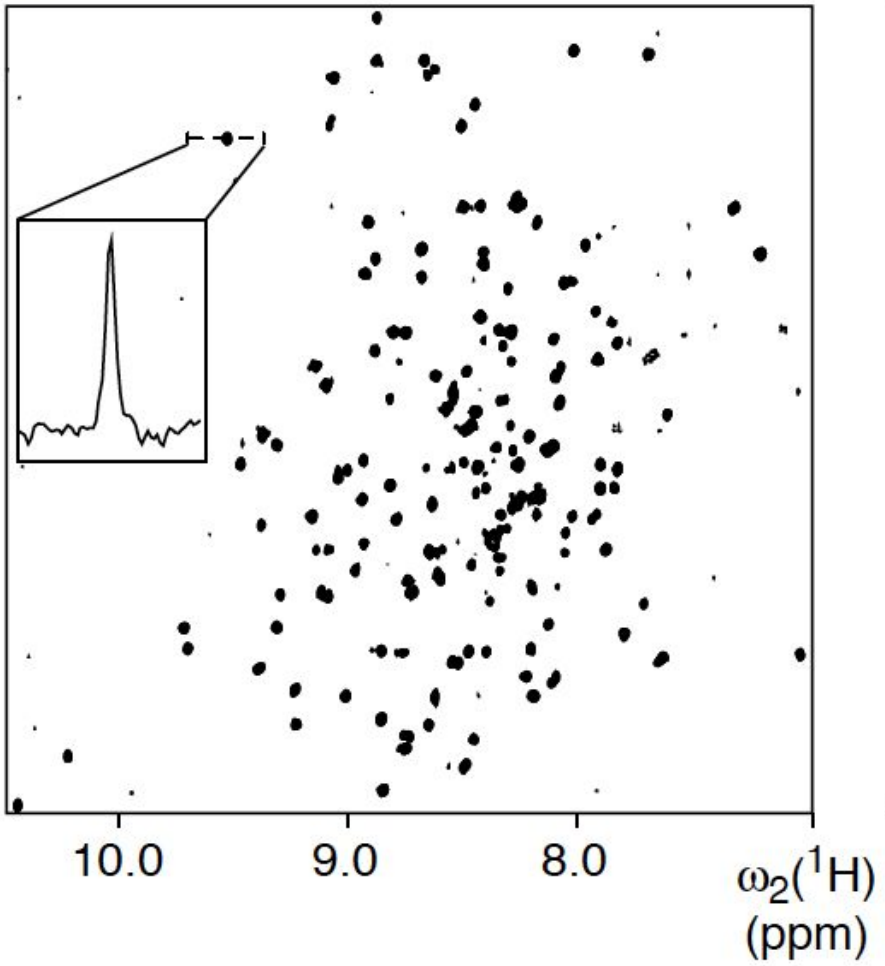
high mg. field >700 MHz



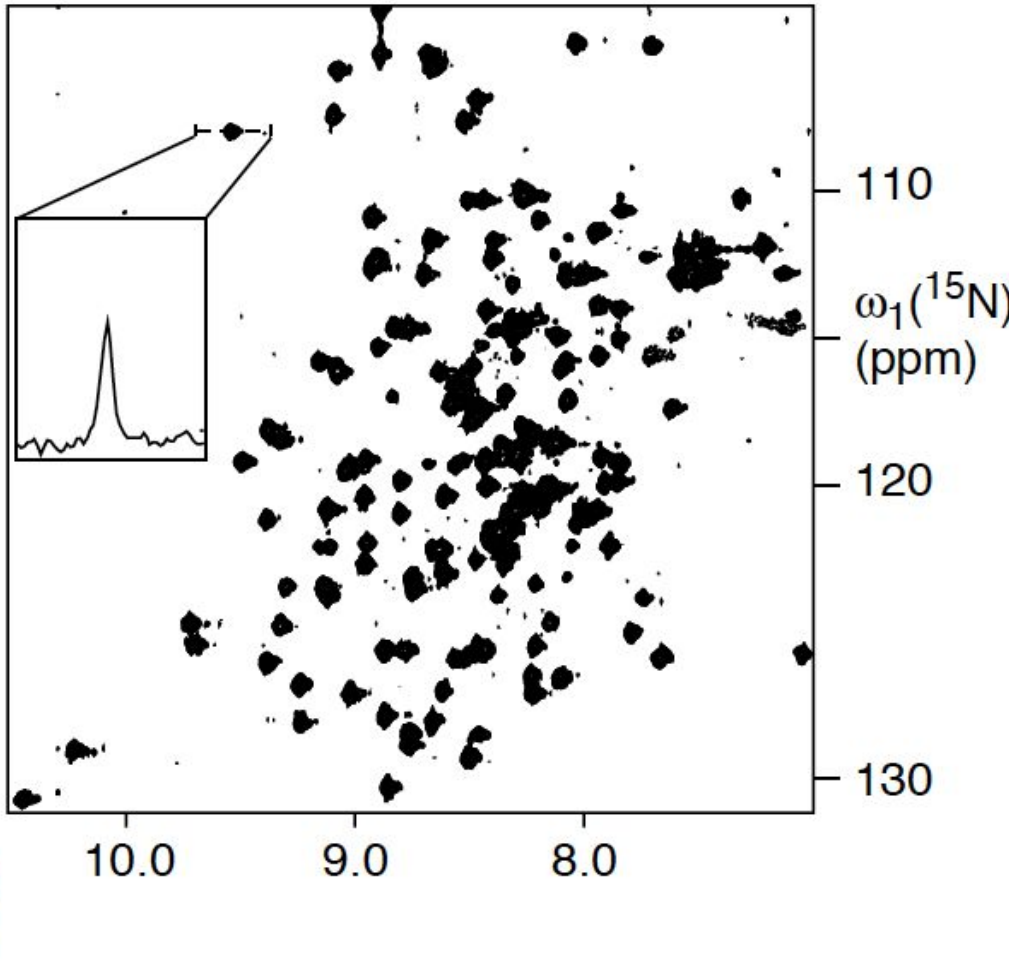
low mg. field <700 MHz



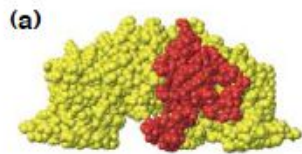
**(a)** 2D [ $^{15}\text{N}$ ,  $^1\text{H}$ ]-TROSY



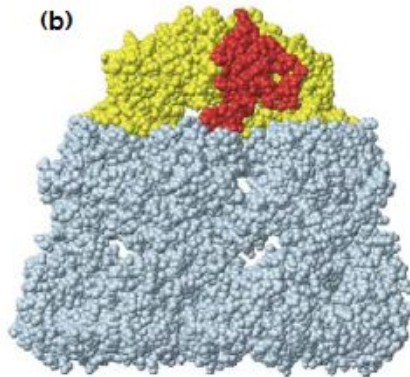
**(b)** 2D [ $^{15}\text{N}$ ,  $^1\text{H}$ ]-COSY



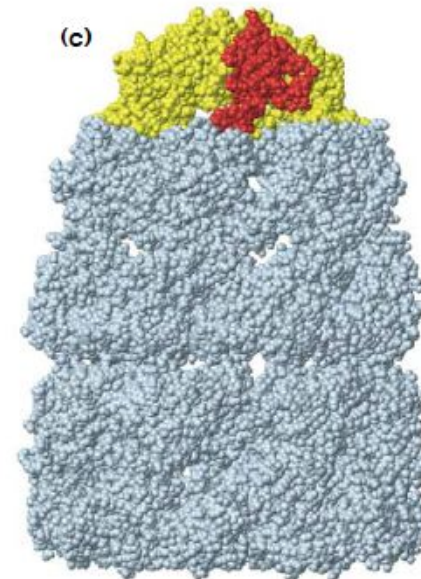
70%  $^2\text{H}$ ,  $^{13}\text{C}$ ,  $^{15}\text{N}$  sample in  $\text{H}_2\text{O}$ , 700MHz  
CRINEPT-TROSY



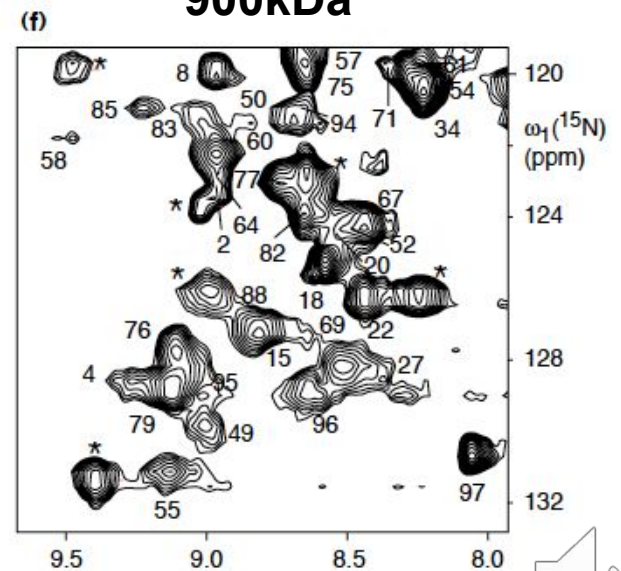
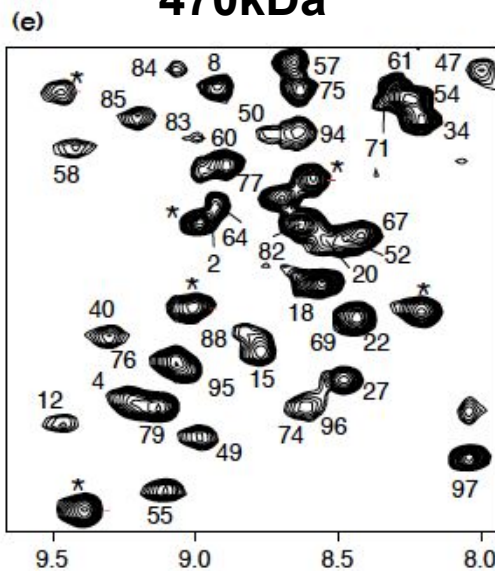
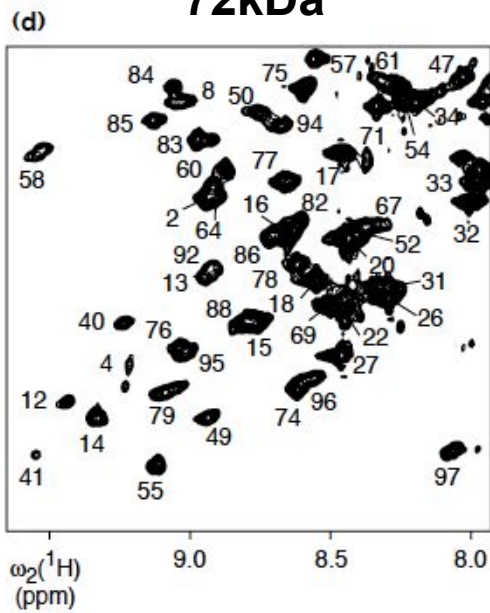
**72kDa**



**470kDa**



**900kDa**



# Summary

NMR is a robust tool for studying structural properties and interaction properties of biomolecules of variable molecular size at various levels of resolution.



# Thank you for your attention

For Application to Protein Characterization

by

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