

What NMR can tell about proteins

For Application to Protein Characterization

by

Radovan Fiala, Karel Kubicek, and Pavel Kaderavek
CEITEC, Masaryk University



How to prepare and what to expect from NMR spectroscopy

A] Scientific question / I'd like to obtain:

- 1) Structure
- 2) Relaxation properties
- 3) Interaction at atomic level resolution
- 4) Analysis (NMR is also analytical method)
- 5) Image (MRI)

B] What do I need to provide / prepare?

C] How much would it cost?

D] How long would it take?

E] Can I do it on my own (or a student/colleague of mine)

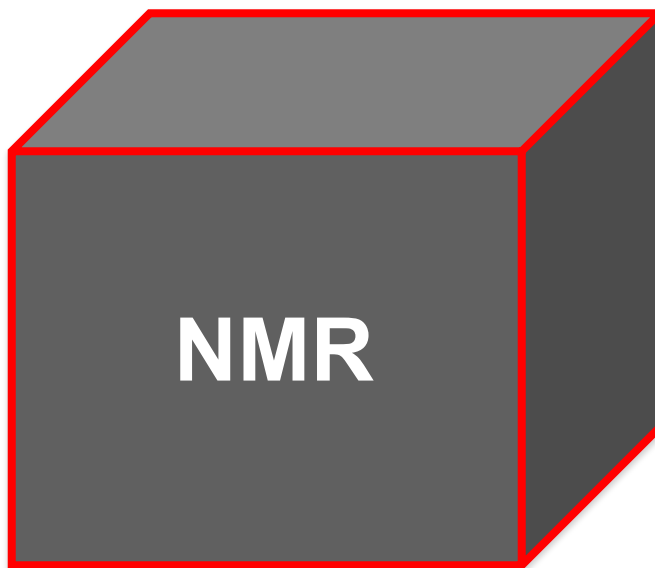


200-500 μl of
100-1000 μM
compound



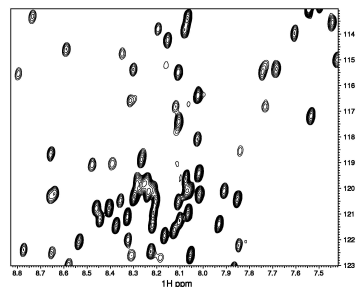
sample

Method of choice

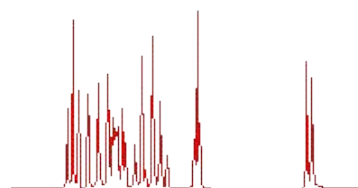


NMR

Data to be analyzed



spectrum



Results

- 1) Structure
- 2) Relaxation properties
- 3) Interaction at atomic level resolution
- 4) Analysis
- 5) Image



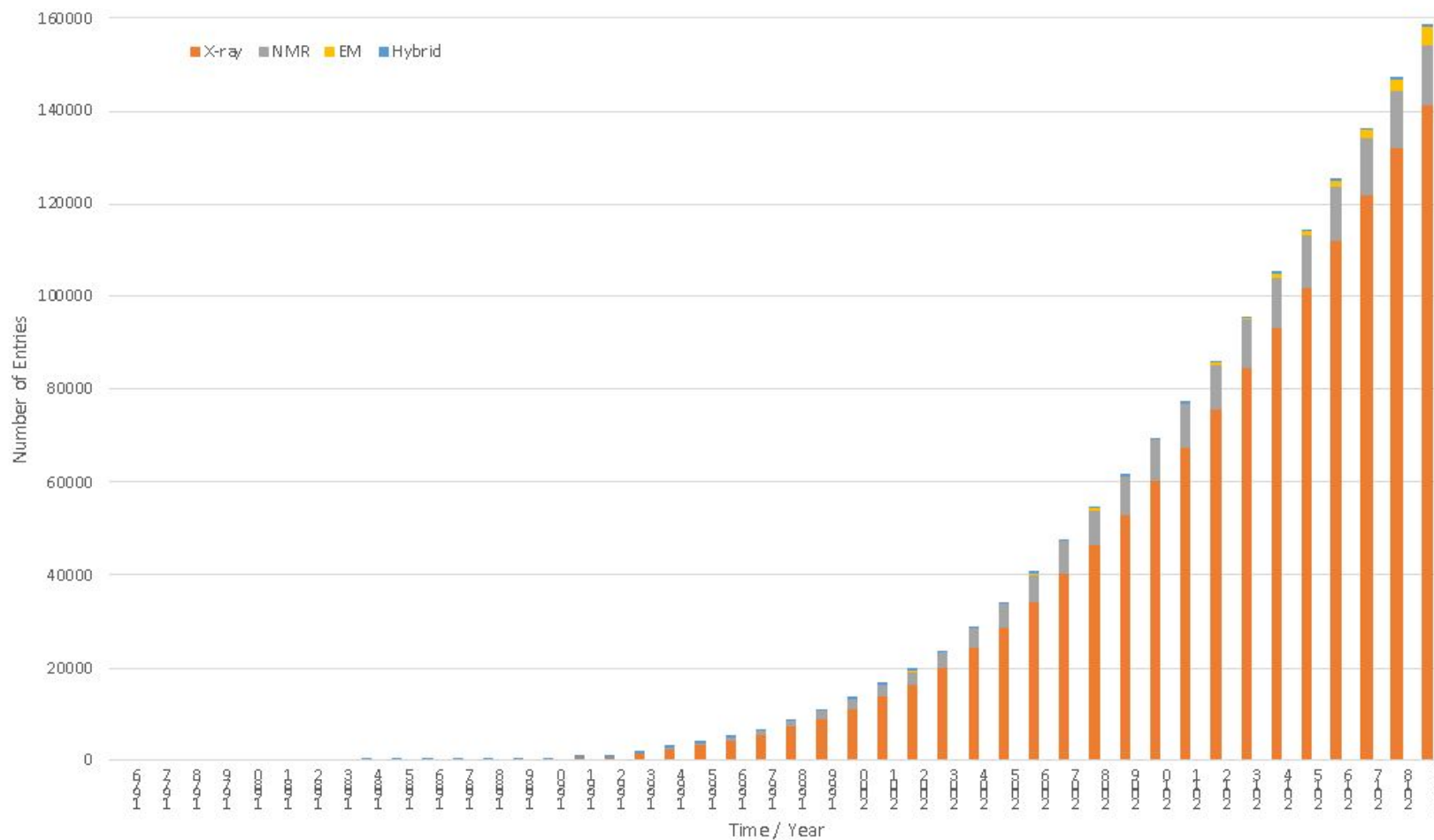
PDB Data Distribution by Experimental Method and Molecular Type

Exp. Method	Proteins	NA	Protein/NA Complex	Other	Total
X-Ray	133461	2098	7091	8082	150732
NMR	11493	1309	267	92	13161
EM	4181	47	1478	501	6207
Other	32	1	0	4	37
Multi Method	162	6	3	6	177

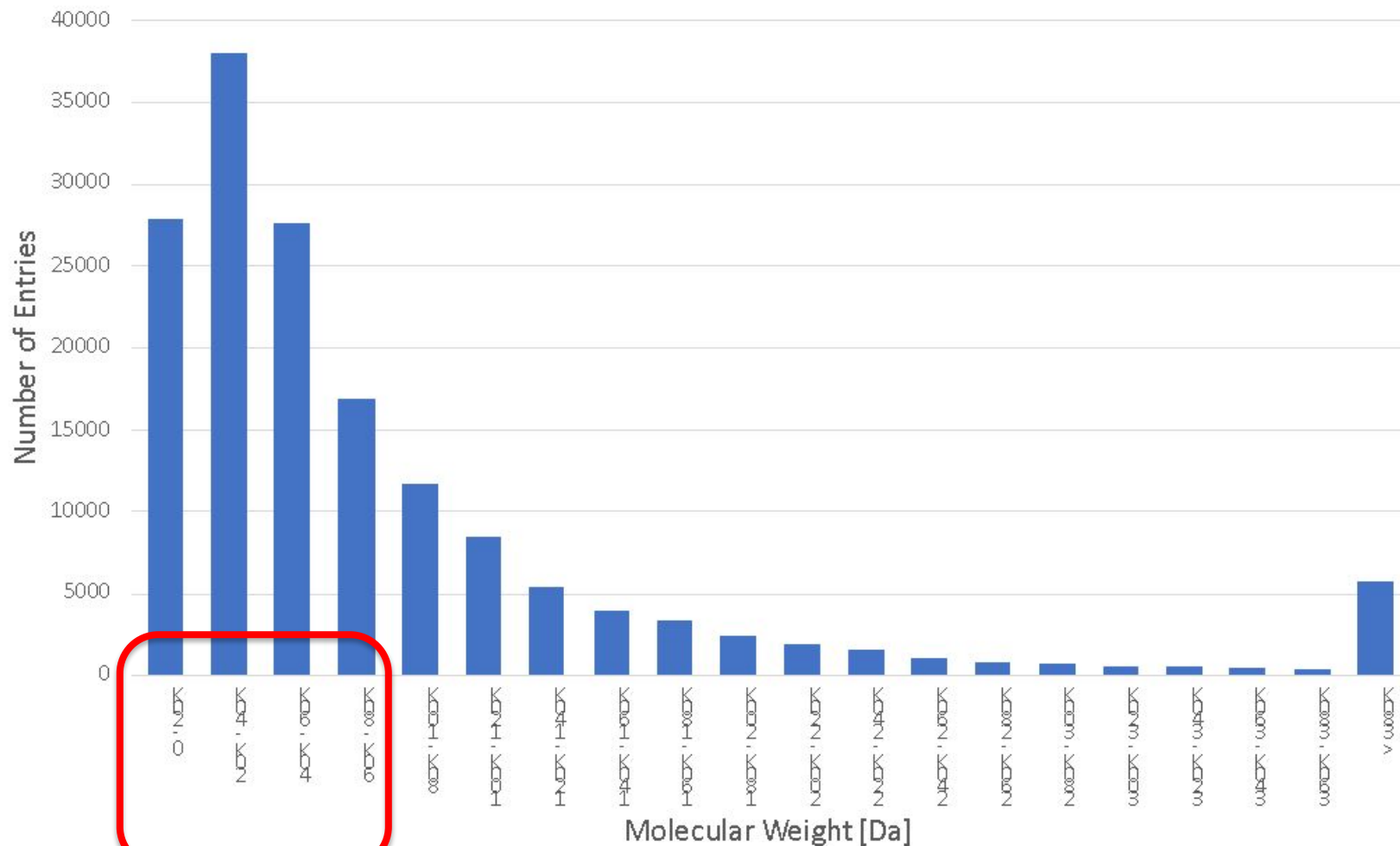


Source: <http://www.rcsb.org> as of 03Nov2020

Overall Growth of Released Structures Per Year



PDB Data Distribution by Molecular Weight of the Deposited Structure



NMR

Safety First



Composition of the Earth's Crust, Seawater, and the Human Body*

Earth's Crust		Seawater		Human Body [†]	
Element	%	Compound	mM	Element	%
O	47	Cl ⁻	548	H	63
Si	28	Na ⁺	470	O	25.5
Al	7.9	Mg ²⁺	54	C	9.5
Fe	4.5	SO ₄ ²⁻	28	N	1.4
Ca	3.5	Ca ²⁺	10	Ca	0.31
Na	2.5	K ⁺	10	P	0.22
K	2.5	HCO ₃ ⁻	2.3	Cl	0.08
Mg	2.2	NO ₃ ⁻	0.01	K	0.06
Ti	0.46	HPO ₄ ²⁻	<0.001	S	0.05
H	0.22			Na	0.03
C	0.19			Mg	0.01

*Figures for the earth's crust and the human body are presented as percentages of the total number of atoms; seawater data are millimoles per liter. Figures for the earth's crust do *not* include water, whereas figures for the human body do.

[†]Trace elements found in the human body serving essential biological functions include Mn, Fe, Co, Cu, Zn, Mo, I, Ni, and Se.



For most of modern NMR applications,
 ^{13}C , ^{15}N , and often ^2H needed

Isotope	Ground state spin	Natural abundance [%]	Rel. Sensitivity
^1H	$\frac{1}{2}$	~100	$1.00 \times 10^{+0}$
^{13}C	$\frac{1}{2}$	1.10	1.59×10^{-2}
^{15}N	$\frac{1}{2}$	0.37	1.04×10^{-3}
^{19}F	$\frac{1}{2}$	100	8.30×10^{-1}
^{31}P	$\frac{1}{2}$	~100	6.63×10^{-2}
^{12}C	0	98.90	-
^{16}O	0	~100	-



Samples:

- 1) Small organic molecules - synthesis
- 2) Peptides (10-40 aa) - synthesis
- 3) Proteins (40-200 aa) - $^{13}\text{C}/^{15}\text{N}$ enriched media
- 4) Large proteins > 200 aa - $^2\text{H}/^{13}\text{C}/^{15}\text{N}$ enriched media

If high concentrations (>1 mM) multidimensional spectra in natural abundance can be performed



NMR as a tool for structure determination of proteins and biological complexes:

From single proteins to large systems



Conventional scheme for structure determination by NMR

- 1) Sample preparation / protein expression
- 2) NMR data acquisition and processing
- 3) Backbone chemical shifts assignment**
- 4) Assignment of side-chain chemical shifts
- 5) Peak-picking of NOESY spectra
- 6) Structure calculation
- 7) (iterative improvement of 5 and 6)



Before we start with assignment of backbone chemical shifts we need to know:

1) Protein primary sequence

MQQDDDFQNF VATLESFKDL KSGISGSRIK KLTTYALDHI DIESKIISLI
IDYSRLCPDS HKLGSLYIID SIGRAYLDET RSNSNSSSNK PGTCAHAIN
LGEVIQELLS DAIAKSNQDH KEKIRMLLDI WDRSGLFQKS YLNAIRSKCF
AMDLEHHHHHH

2) Standard chemical shifts of $C\alpha$ and $C\beta$ (*vide infra*)

3) Secondary structure prediction (values of 2) will be affected accordingly)

4) Precise peak-picking (manually or semi-automatically)

5) $^{13}C/^{15}N$ uniformly labeled protein (6-30kDa)



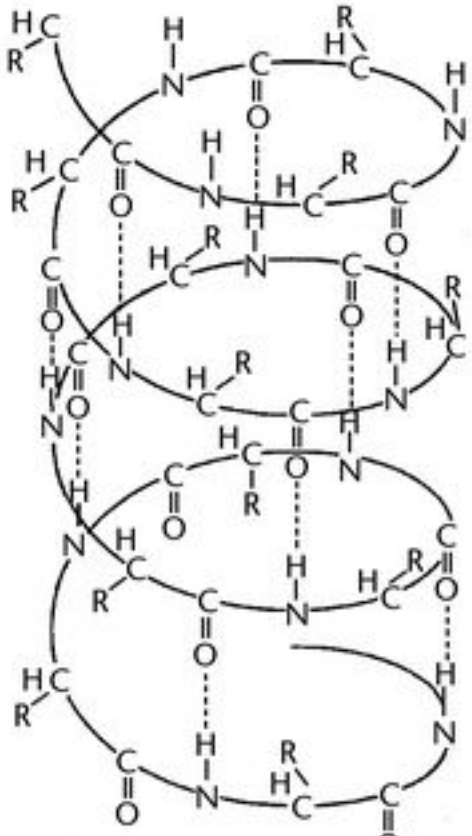
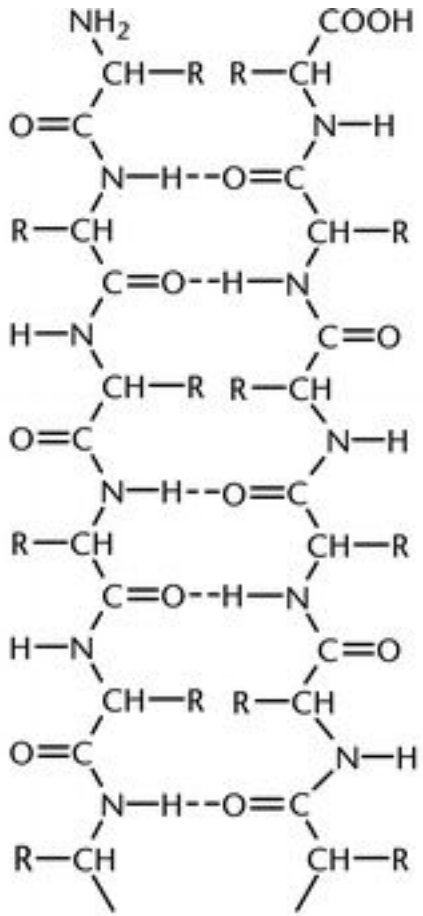
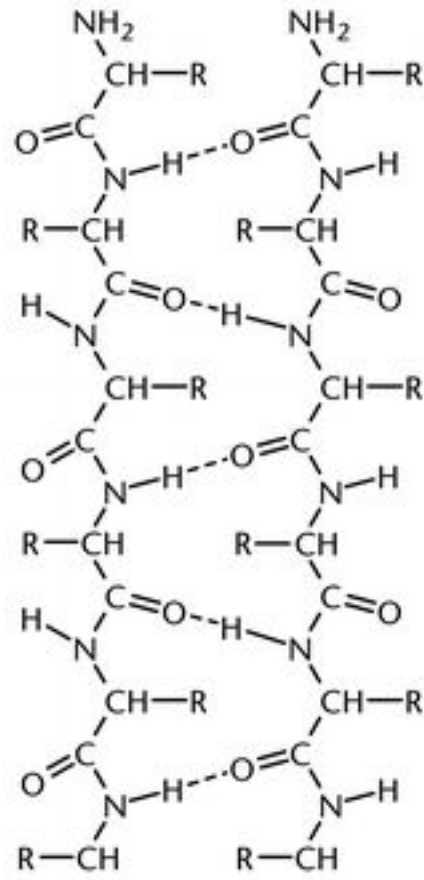
Protein backbone and secondary structure

β -strand

parallel

antiparallel

α -helix

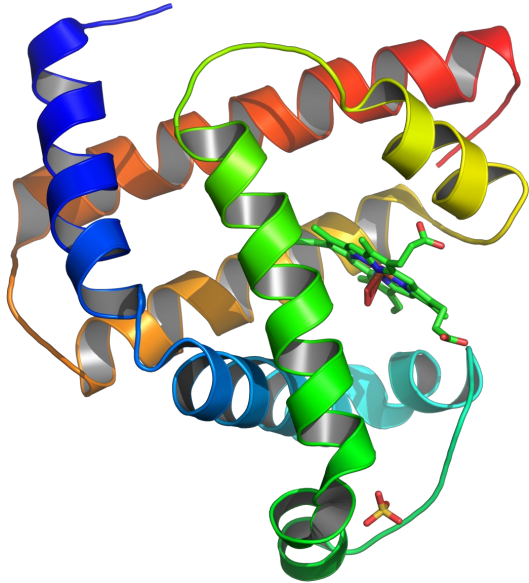


N- \longrightarrow C-terminus

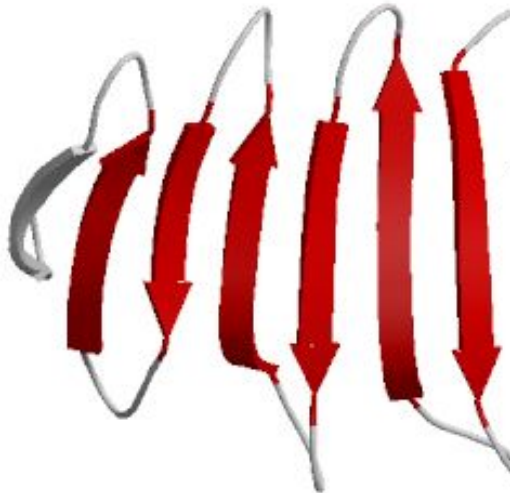
Hydrogen bond



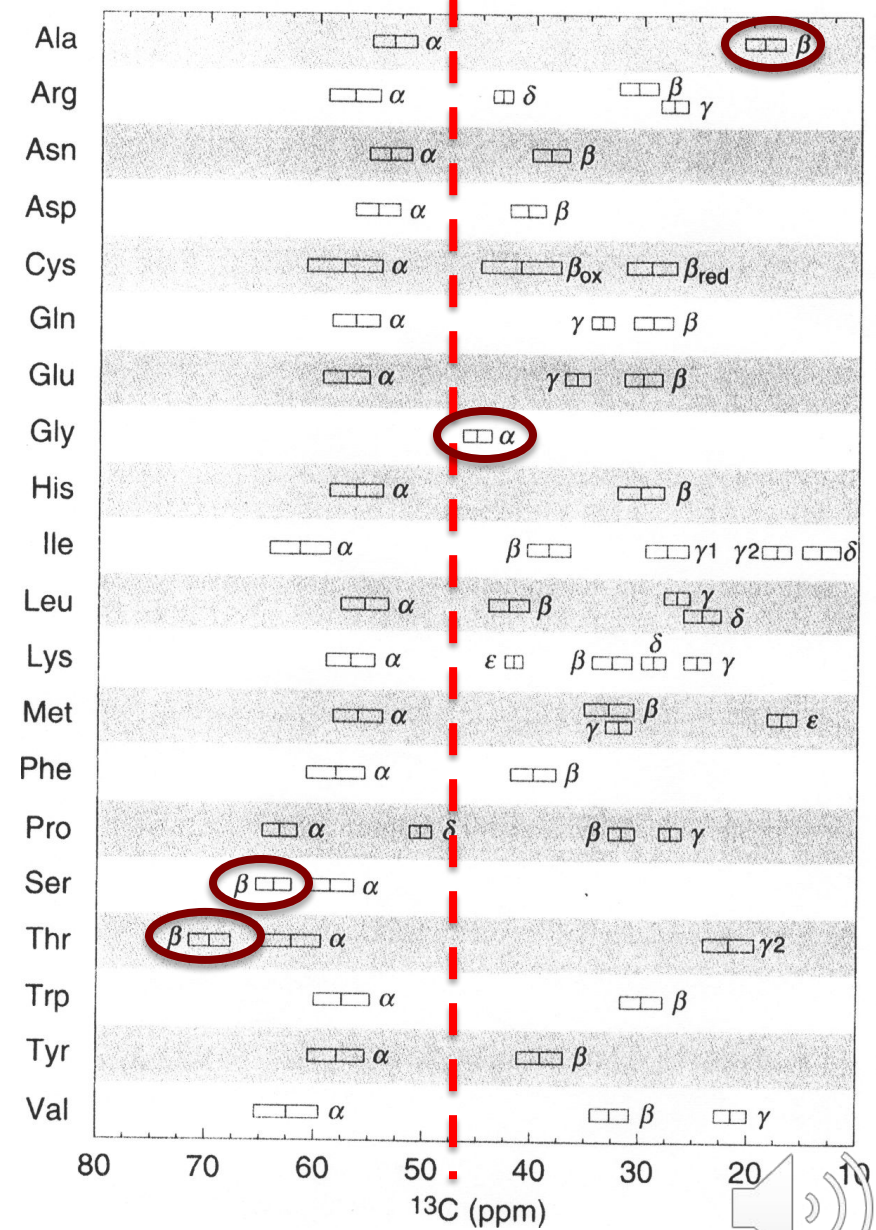
α -helix



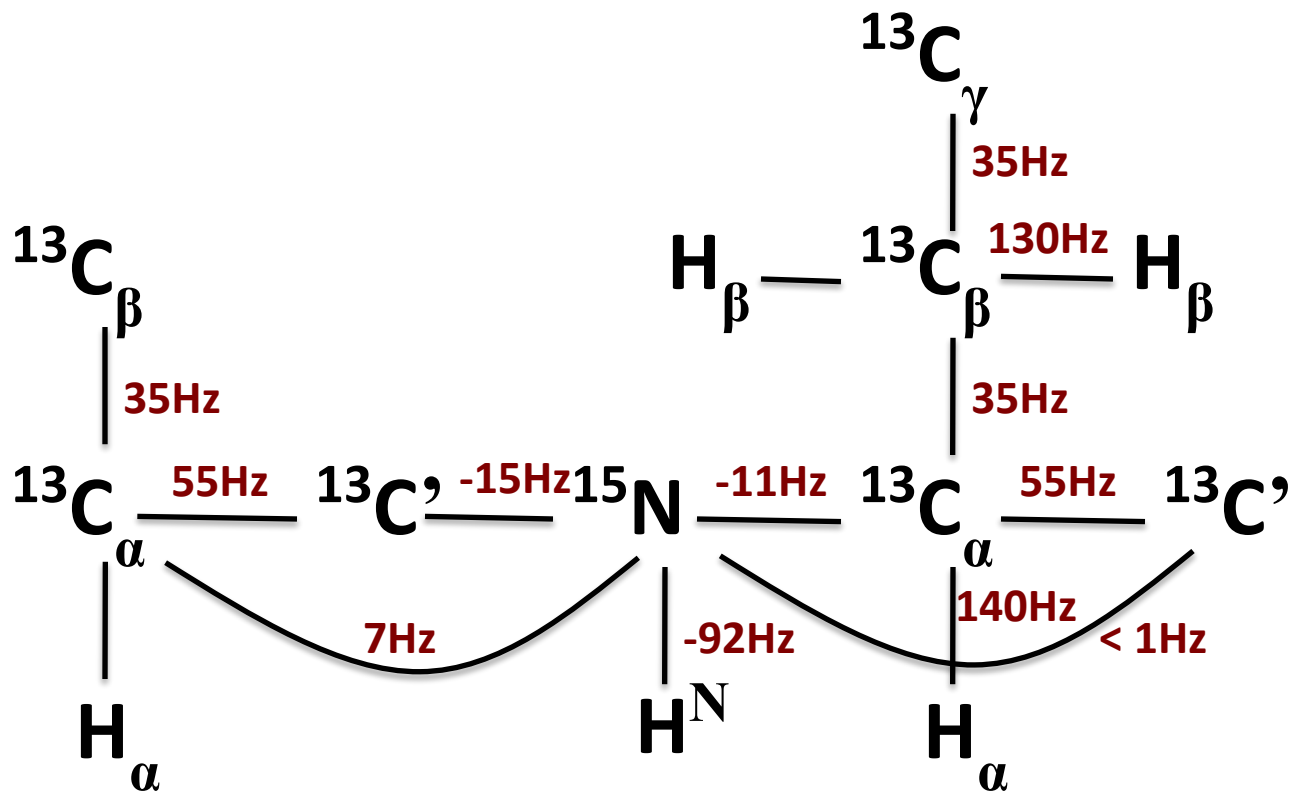
β -strand



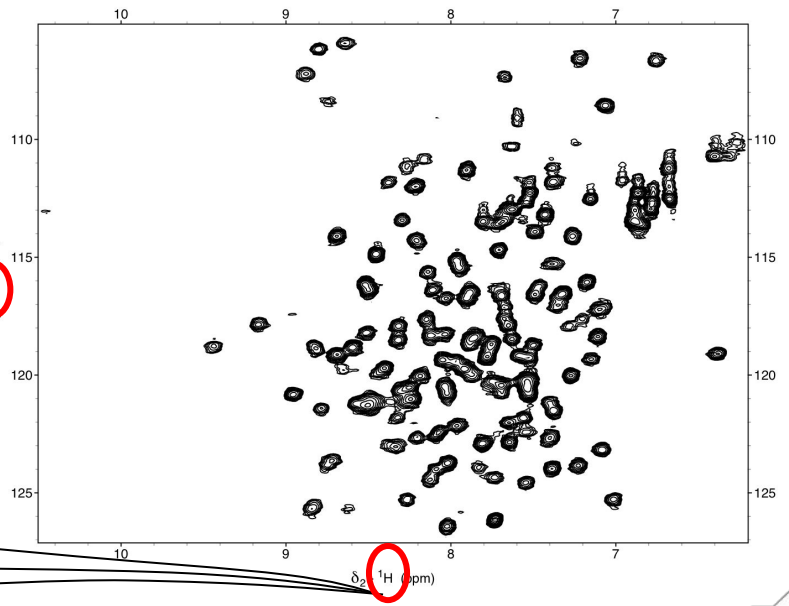
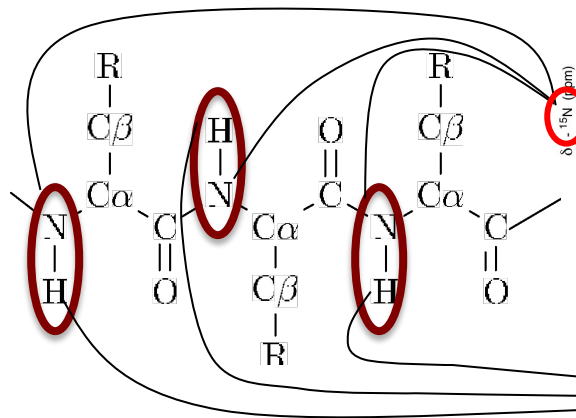
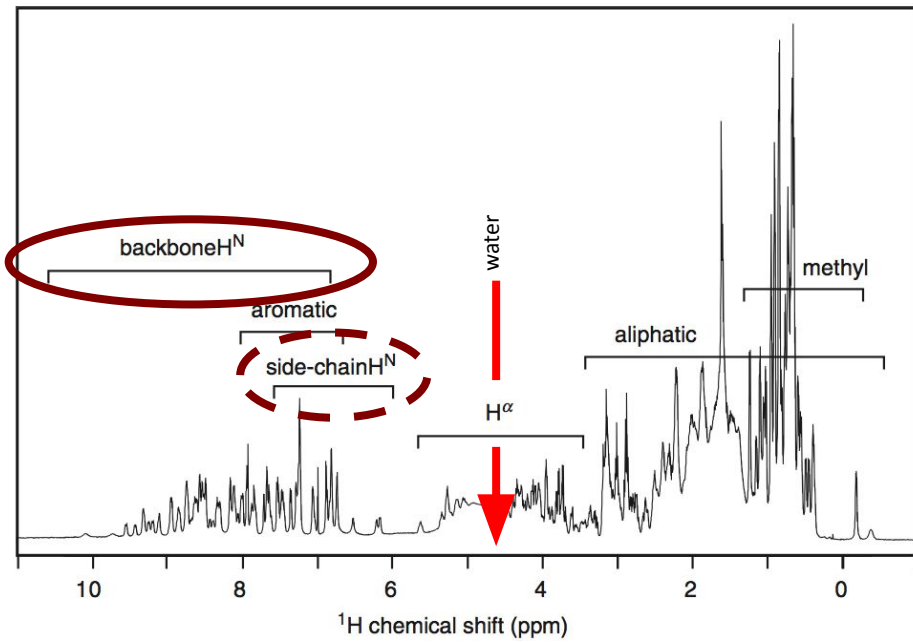
^{13}C chem. shift in proteins



In a $^{13}\text{C}/^{15}\text{N}$ uniformly labeled protein, correlation spectra can be measured through single- and double-bond couplings



^1H 1D, Cavanagh et al., 2007

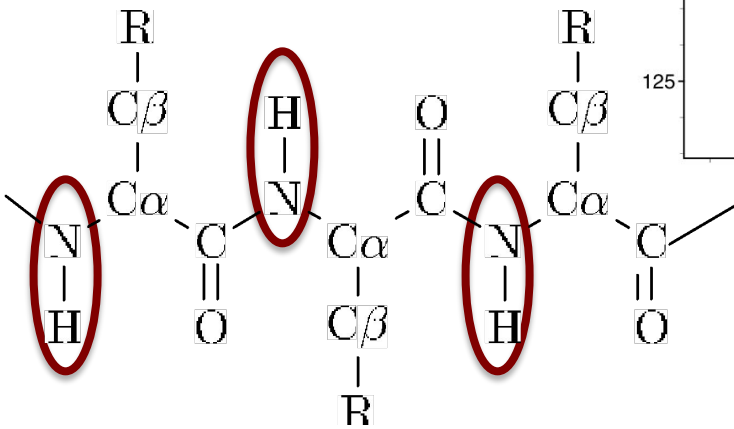
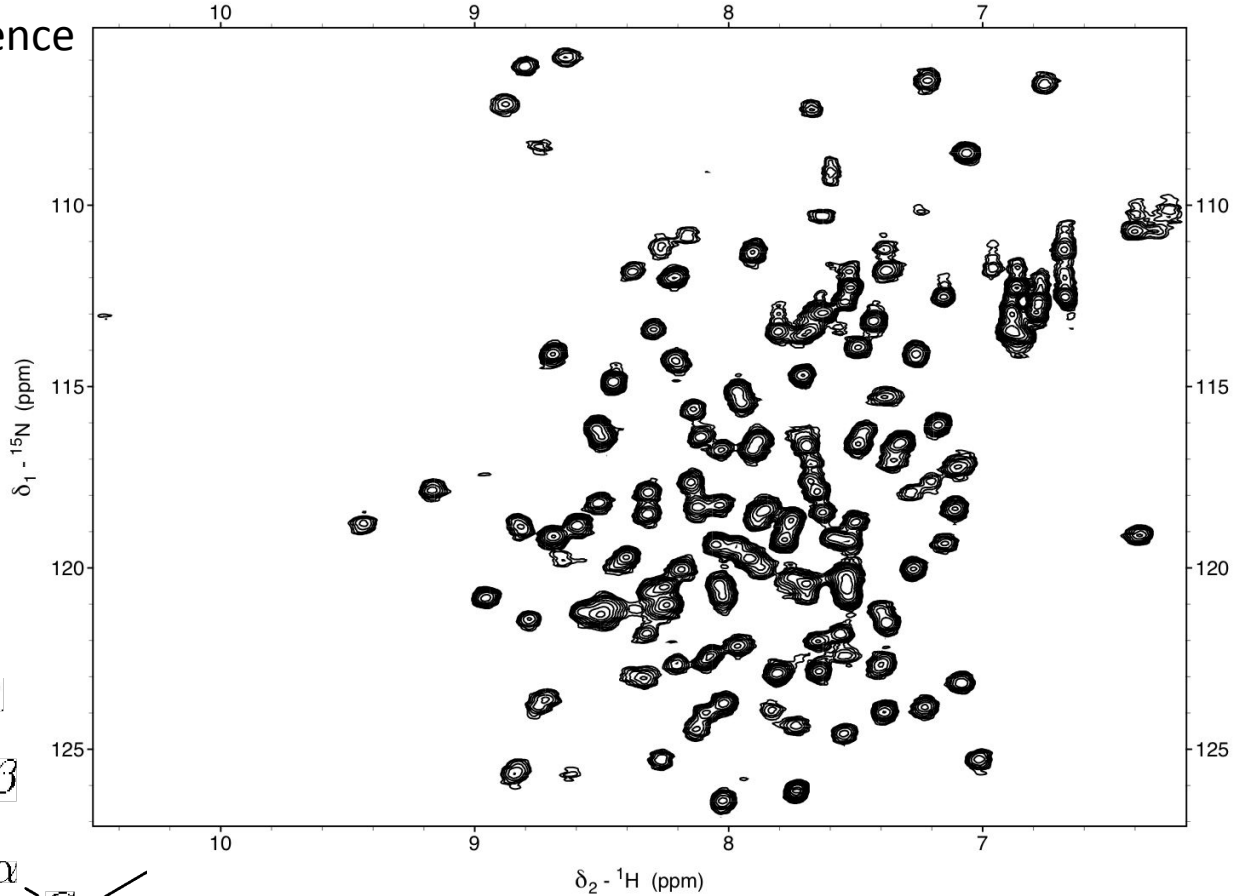


$^1\text{H}-^{15}\text{N}$ HSQC, cca 155 aa, well folded, 600MHz, 29



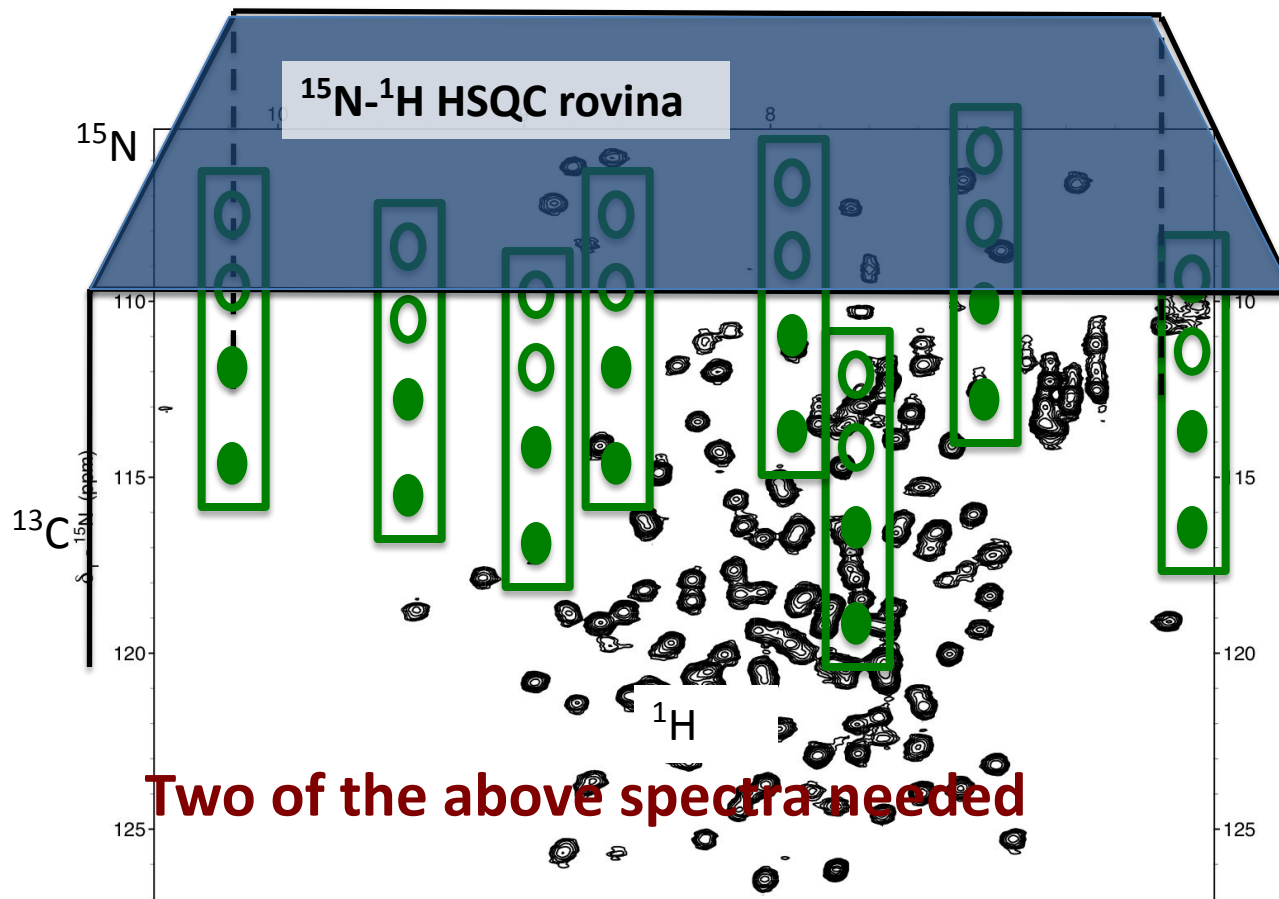
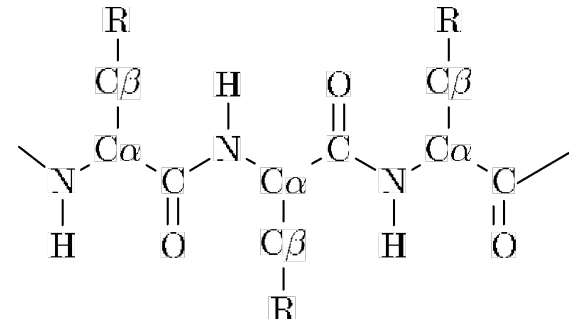
^{15}N - ^1H HSQC – Heteronuclear SingleQuantum Correlation

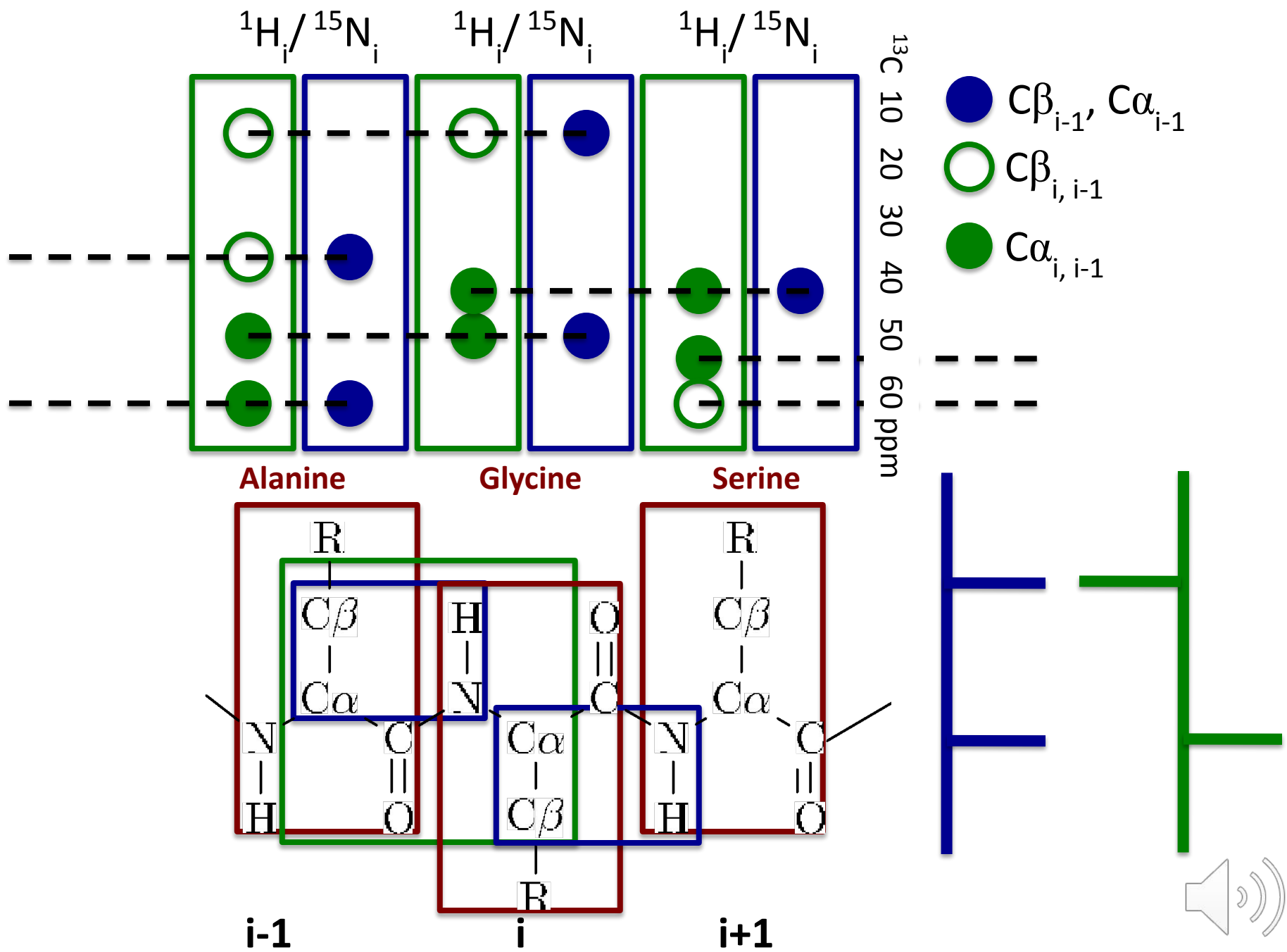
- 1) 1 peak \cong 1 amino acid
- 2) Excellent info about the protein fold
- 3) No info about primary sequence



Traditional triple resonance NMR spectra for protein backbone assignment

- 1) HNCO – chem. shifts of C=O
- 2) HNCA – chem. shifts of $C_{\alpha,i}$, $C_{\alpha,i-1}$
- 3) HNCOCA – chem. shifts of $C_{\alpha,i-1}$
- 4) **HNCACB** – chem. shifts of $C_{\alpha,i}$, $C_{\alpha,i-1}$, $C_{\beta,i}$, $C_{\beta,i-1}$
- 5) **HNCOCACB** – chem. shifts of $C_{\alpha,i-1}$, $C_{\beta,i-1}$

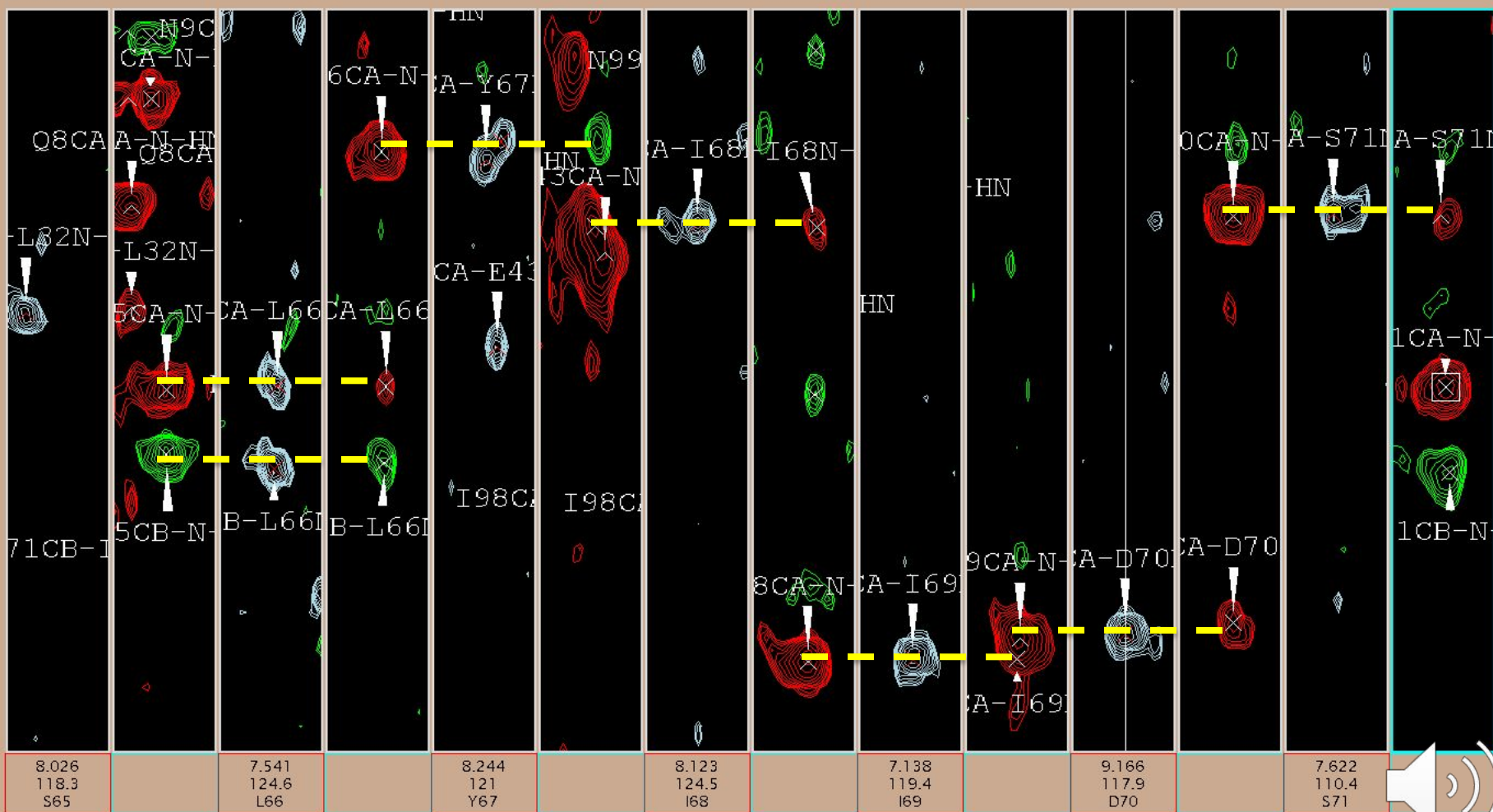




Backbone chemical shifts assignment example:

- fragment **S65-S71** (50-66ppm ν ^{13}C)

Show Find



Automatic chemical shifts assignment

- 1) With a known 3D protein structure (e.g. X-ray, Modeller)
- 2) Without prior knowledge of a structure
- 3) **e.g. Automated Projection Spectroscopy** APSY (S. Hiller, F. Fiorito, K. Wüthrich & G. Wider, *PNAS* (2005) 102, 10876-81)

Advantages

- 1) Time

Disadvantages

- 1) With increasing size of the protein (increasing spectral overlap) the reliability is decreasing



How to prepare and what to expect from NMR spectroscopy

A] Scientific question / I'd like to obtain:

- 1) Structure
- 2) Relaxation properties
- 3) Interaction at atomic level resolution
- 4) Analysis (NMR is also analytical method)
- 5) Image (MRI)

B] What do I need to provide / prepare?

C] How much would it cost?

D] How long would it take?

E] Can I do it on my own (or a student/colleague of mine)



Thank you for your attention

For Application to Protein Characterization

by

Radovan Fiala, Karel Kubicek, and Pavel Kaderavek
CEITEC, Masaryk University

