## NEWS AND COMMENTARY

## Allopolyploid genomes The complex nature of allopolyploid plant genomes

MJ Hegarty and SJ Hiscock

Heredity advance online publication, 3 June 2009; doi:10.1038/hdy.2009.61

olyploidy is a major creative force in plant evolution, from ferns to flowering plants. Estimates suggest that  $\sim 70\%$  of flowering plants have undergone at least one whole-genome duplication event during their evolutionary history (Levin, 2002), and that 2-4% of all speciation events involve polyploidisation (Otto and Whitton, 2000). The majority of polyploid plants arise through the combined effects of interspecific hybridisation and polyploidy (allopolyploidy), and much attention has focused on understanding what happens when divergent genomes are brought together by these processes. A recent study by Buggs et al. (2009) has shown that at a genomic level the possible outcomes of allopolyploidy are not always as predictable as we might expect. As it is known that many polyploid genomes eventually return to a diploid-like state through loss or divergence of duplicated genes, Buggs et al. (2009) set out to determine the timing of these events in the allotetraploid hybrid Tragopogon miscellus. This hybrid is an excellent system to examine the timing of genomic changes associated with allopolyploidy, as it is known to have arisen within the last 80 years, unlike many other natural hybrids that have formed many thousands or even millions of years ago, such as wheat (Peterson et al., 2006) or cotton (Adams et al., 2003). By assaying the detectability of cleaved amplified polymorphic sequence markers on both genomic DNA and cDNA, Buggs et al. (2009) were able to determine whether parental gene copies for 10 loci were being lost or silenced in 57 hybrids from five natural populations. They were also able to compare these hybrids to a resynthesised allotetraploid created under glasshouse conditions. They found that, across 10 homoeologous gene pairs in the 57 natural hybrids, 3.2% of loci had been rendered undetectable in the genomic DNA, suggesting loss of one parental gene copy. In addition, analysis of seven of these homoeologue pairs using cDNA as a template showed that expression of one

parental gene copy had been lost in 6.8% of loci. In none of these cases, silencing or allele loss was detected in the resynthesised allopolyploid.

These findings demonstrate that silencing and/or elimination of parental gene copies in Tragopogon allopolyploids is not an immediate consequence of interspecific hybridisation or genome duplication, but rather an ongoing process that varies in frequency among different hybrid populations. As the authors note, this is in stark contrast to the evidence from some other allopolyploid systems, for example, allohexaploid bread wheat (Triticum aestivum), where some gene loss occurs immediately upon interspecific hybridisation and again after polyploidisation (Ozkan et al., 2001; Shaked et al., 2001). Nevertheless, other allopolyploids are also known to be subjected to an ongoing process of recombination between homoeologous chromosomes (Figure 1), for example Brassica, which shows gradual loss of certain parental DNA fragments (Song et al., 1995; Gaeta et al., 2007). The timing of parental gene silencing in the Tragopogon hybrids also differs from that observed in other systems. Immediate changes to gene expression occur upon interspecific hybridisation in cotton (Flagel et al., 2008) in a manner that can be both temporally

or spatially dependent, even within a single cell (Adams et al., 2003; Hovav et al., 2008). Immediate changes to gene expression have also been observed in other hybrids, notably Arabidopsis suecica allotetraploids (Wang et al., 2006), and in Senecio allopolyploids (Hegarty et al., 2005) where distinct changes to gene expression are associated with both interspecific hybridisation and genome duplication (Hegarty et al., 2006). Such changes in gene expression are not simply the 'additive' product of the progenitor species because  $\sim 24\%$  of the affected genes display non-additive expression in both cotton (Flagel et al., 2008) and Senecio (Hegarty et al., 2008), with the percentage of microarray features displaying non-additive expression in Senecio being similar to that seen in A. suecica at 3 and 5%, respectively (Wang et al., 2006; Hegarty et al., 2008). In Senecio, 42% of genes showing non-additive changes in expression also showed 'trangressive' patterns of gene expression-that is, expression change significantly higher or lower than that observed in either parent (Hegarty et al., 2008). Buggs et al. (2009) suggest that the contrast between findings in Tragopogon and the other systems may be due to their methodology, which detects only the presence or absence of gene expression, and is not capable of distinguishing changes to expression level in cases where complete silencing does not occur. Nonetheless, they raise the important point that allopolyploidy may vary in its effects between species. This is certainly true at the level of chromosomal organisation in the case of allotetraploid cotton, which shows none of the largescale genetic rearrangements typically observed in plant hybrids (Liu et al.,



Figure 1 Loss of parental DNA due to recombination between homoeologous chromosomes. This figure shows a simplified representation of recombination between homoeologues in an allopolyploid, leading to loss of DNA from one parent. A reciprocal exchange between homoeologues is followed by segregation of the recombinants such that certain gametes (boxed) may not possess either the original parental chromatid type or both recombinants, resulting in loss of one homoeologous fragment in offspring derived from these gametes (Figure adapted from Nicolas et al., 2008).

2001), nor does it seem to undergo any widespread changes in the pattern of DNA methylation seen in systems such as *Spartina* (Liu *et al.*, 2001; Salmon *et al.*, 2005). As Buggs *et al.* (2009) note, the level of divergence between the progenitors of a given hybrid is likely to be a factor in differences between hybrid systems, and, therefore, must be taken into account when making predictions as to the likely genetic and epigenetic consequences of genome mergers and duplication events.

The processes by which two genomes adapt to coexistence within the same nucleus are complex and can differ markedly between species. Ongoing studies of natural hybrid systems, such as Tragopogon and Senecio-where hybridisation and polyploidy have arisen relatively recently from an evolutionary perspective-are therefore crucial to identifying these processes and the factors that drive them. Given the recent resurgence of interest in recreating allopolyploid crop species to improve genetic diversity, the more data we can gather on the consequences of allopolyploidisation, the better.

Dr MJ Hegarty is at the Institute of Biological, Environmental and Rural Sciences, Aberystwyth University, Edward Llwyd Building, Penglais, Aberystwyth, Ceredigion SY23 3DA, UK and Professor SJ Hiscock is at the School of Biological Sciences, University of Bristol, Woodland Road, Bristol BS8 1UG, UK

## e-mail: ayh@aber.ac.uk

Adams KL, Cronn R, Percifield R, Wendel JF (2003). Genes duplicated by polyploidy show unequal contributions to the transcriptome and organ-specific reciprocal silencing. *Proc* Natl Acad Sci USA 100: 4649–4654.

- Buggs RJA, Doust AN, Tate JA, Koh J, Soltis K, Feltus FA et al. (2009). Gene loss and silencing in Tragopogon miscellus (Asteraceae): comparison of natural and synthetic allotetraploids. *Heredity* (this issue).
- Flagel LÊ, Udall J, Nettleton D, Wendel J (2008). Duplicate gene expression in allopolyploid Gossypium reveals two temporally distinct phases of expression evolution. BMC Biol 6: 16.
- Gaeta RT, Pires JC, Iniguez-Luy F, Leon E, Osborn TC (2007). Genomic changes in resynthesised *Brassica napus* and their effect on gene expression and phenotype. *Plant Cell* 19: 3403–3417.
- Hegarty MJ, Barker GL, Brennan AC, Abbott RJ, Edwards KJ, Hiscock SJ (2008). Changes to gene expression associated with hybrid speciation in plants: further insights from transcriptomic studies in *Senecio. Phil Trans Roy Soc Lond B Biol Sci* 363: 3055–3069.
- Hegarty MJ, Barker GL, Wilson ID, Abbott RJ, Edwards KJ, Hiscock SJ (2006). Transcriptome shock after interspecific hybridisation in Senecio is ameliorated by genome duplication. *Curr Biol* 16: 1652–1659.
- Hegarty MJ, Jones JM, Wilson ID, Barker GL, Coghill JA, Sanchez-Baracaldo P et al. (2005). Development of anonymous cDNA microarrays to study changes to the Senecio floral transcriptome during hybrid speciation. Mol Ecol 14: 2493–2510.
- Hovav R, Udall JA, Chaudhary B, Rapp R, Flagel L, Wendel JF (2008). Partitioned expression of duplicated genes during development and evolution of a single cell in a polyploid plant. *Proc Natl Acad Sci USA* 105: 6191–6195.
- Levin DA (2002). The Role of Chromosomal Change in Plant Evolution. Oxford University Press: New York.
- Liu B, Brubaker CL, Cronn RC, Wendel JF (2001). Polyploid formation in cotton is not accompanied by rapid genomic changes. *Genome* 44: 321–330.
- Nicolas SP, Leflon M, Liu Z, Eber F, Chelysheva L, Coriton O et al. (2008). Chromosome 'speed

dating' during meiosis of polyploid *Brassica* hybrids and haploids. *Cytogenet Genome Res* **120**: 331–338.

- Otto SP, Whitton J (2000). Polyploid incidence and evolution. *Annu Rev Genet* 34: 401–437.
- Ozkan H, Levy AA, Feldman M (2001). Allopolyploidy-induce rapid genome evolution in the wheat (*Aegilops-Triticum*) group. *Plant Cell* **13**: 1735–1747.
- Peterson G, Seberg O, Yde M, Berthelsen K (2006). Phylogenetic relationships of *Triticum* and *Aegilops* and evidence for the origin of the A, B, and D genomes of common wheat (*Triticum aestivum*). *Mol Phylogenet Evol* **39**: 70–82.
- Salmon A, Ainouche ML, Wendel JF (2005). Genetic and epigenetic consequences of recent hybridisation and polyploidy in *Spartina* (Poaceae). *Mol Ecol* **14**: 1163–1175.
- Shaked H, Kashkusk K, Ozkan H, Feldman M, Levy A (2001). Sequence elimination and cytosine methylation are rapid and reproducible responses of the genome to wide hybridisation and allopolyploidy in wheat. *Plant Cell* 13: 1749–1759.
- Song K, Lu P, Tang K, Osborn TC (1995). Rapid genome change in synthetic polyploids of *Brassica* and its implications for polyploid evolution. *Proc Natl Acad Sci USA* 92: 7719–7723.
- Wang J, Tian L, Lee HS, Wei NE, Jiang H, Watson B *et al.* (2006). Genomewide nonadditive gene regulation in Arabidopsis allotetraploids. *Genetics* 172: 507–517.

## Editor's suggested reading:

- Kalisz S, Kramer EM (2008). Variation and constraint in plant evolution and development. *Heredity* **100**: 171–177.
- Sjödin P, Hedman H, Shavorskaya O, Finet C, Lascoux M, Lagercrantz U (2007). Recent degeneration of an old duplicated flowering time gene in *Brassica nigra*. *Heredity* **98**: 375–384.
- Wu CA, Lowry DB, Cooley AM, Wright KM, Lee YW, Willis JH (2008). Mimulus is an emerging model system for the integration of ecological and genomic studies. *Heredity* 100: 220–230.