



Institute of Biophysics
Department of Biophysical Chemistry and Molecular Oncology
Centre of Biophysical Chemistry, Bioelectrochemistry and Bioanalysis

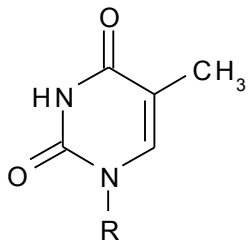


Electroactivity of DNA and effects of DNA structure

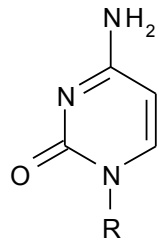
Miroslav Fojta

Olsztyn-Lańsk, September 17th, 2007

A pyrimidine bases

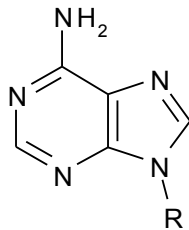


thymine (T)

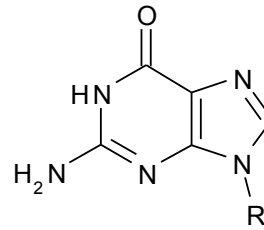


cytosine (C)

purine bases



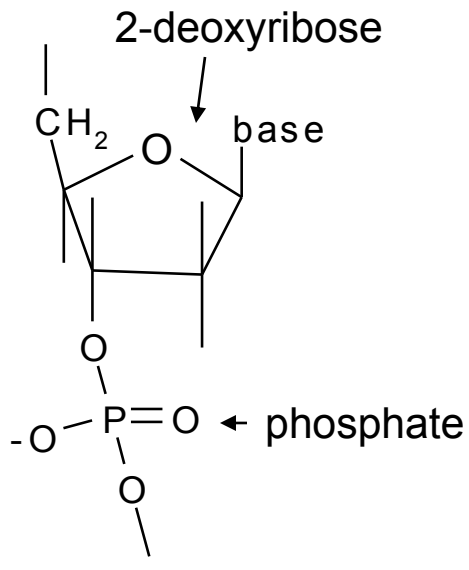
adenine (A)



guanine (G)

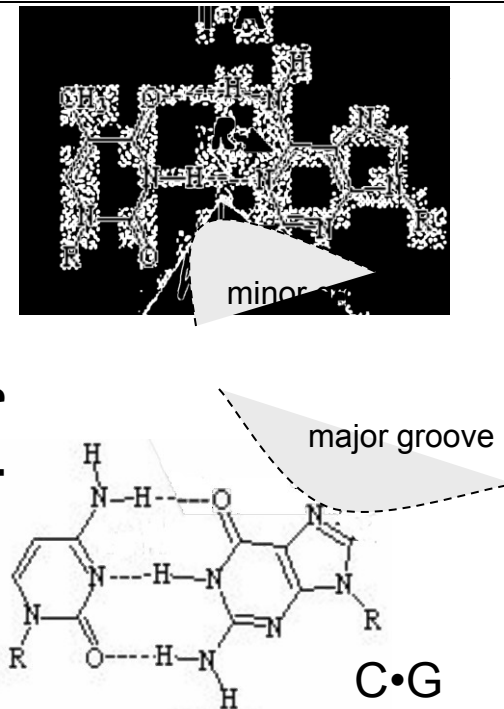
B

nucleotide



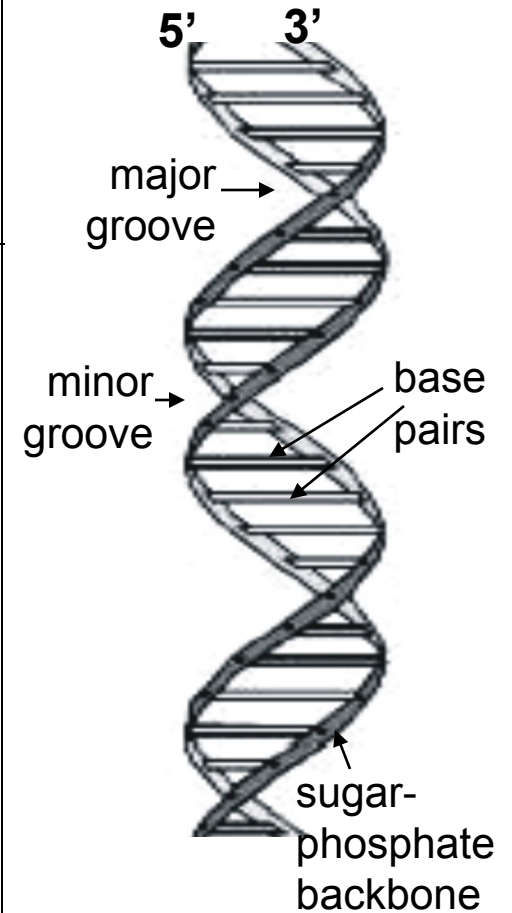
C

páry bazi



D

DNA double helix





late 1950s, Emil Paleček: DNA polarography



(Reprinted from *Nature*, Vol. 188, No. 4751, pp. 656-657, November 19, 1960)

Oscillographic Polarography of Highly Polymerized Deoxyribonucleic Acid

PROCEEDING from my findings^{1,2} that nucleotides, nucleosides and the bases of nucleic acids can be analysed by alternating current oscillographic polarography³⁻⁵, I have also tried to study polymerized deoxyribonucleic acid by this method.

The apparatus used was a Polaroskop P 524 (Křížek, Praha). With this apparatus it is possible to plot dE/dt against E (Fig. 1). The analysis was carried out by means of the dropping mercury electrode in the same electrolytes as were used in my previous work^{1,2}. All measurements were carried out with specimens of deoxyribonucleic acid from calf thymus.

I have established that in a medium of molar ammonium formate, deoxyribonucleic acid shows an anodic indentation at the same potential as deoxyglycolic acid (Fig. 2). Other characteristics of both indentations are also analogous (dependence on direct voltage, temperature, concentration of the electrolyte), which appears to indicate that that due to

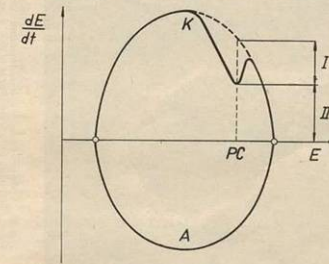


Fig. 1. Graph of dE/dt against E . The nature of the material analysed is characterized by the potential of the indentation (PC), which is somewhat similar to the polarographic half-wave potential. The quantity of the material is characterized by the depth of the indentation. For qualitative analysis, the height II , which can be measured much more easily, is generally measured. K , Cathodic part; A , anodic part

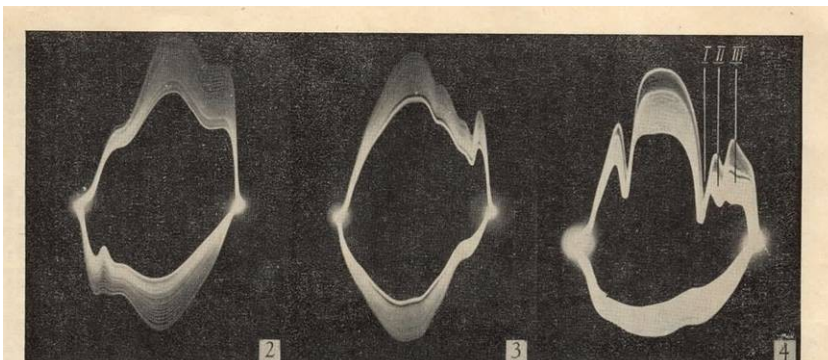


Fig. 2. 100 μ gm. deoxyribonucleic acid/ml. 1 M ammonium formate

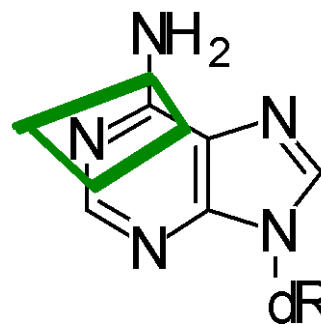
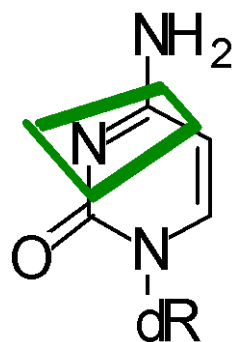
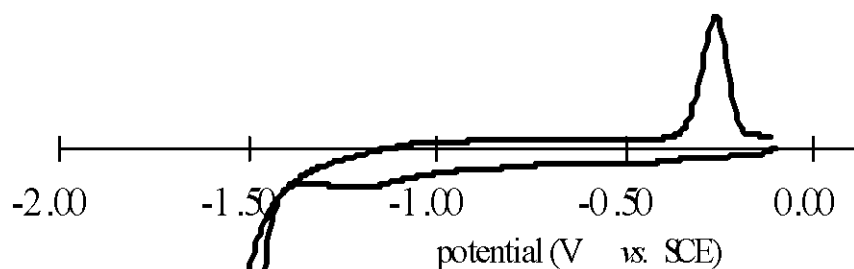
Fig. 3. Apurinic acid in 2 M ammonium formate (concentration corresponding to 2 mgm. of deoxyribonucleic acid)

Fig. 4. 900 μ gm. deoxyribonucleic acid + 5 μ gm. plasma albumin/ml. 10^{-3} M hexamine cobaltic trichloride in 0.1 M ammonium chloride-ammonium hydroxide. Indentations due to cobalt, I; deoxyribonucleic acid, II; protein, III

nucleic acids are electroactive

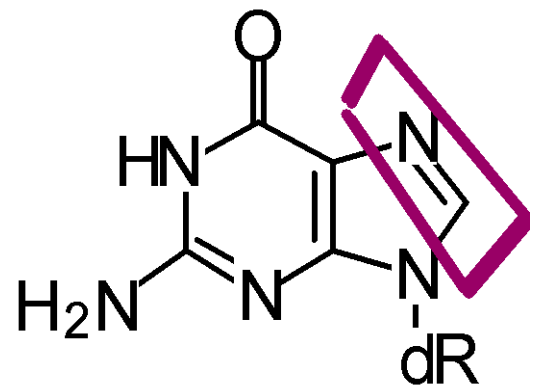
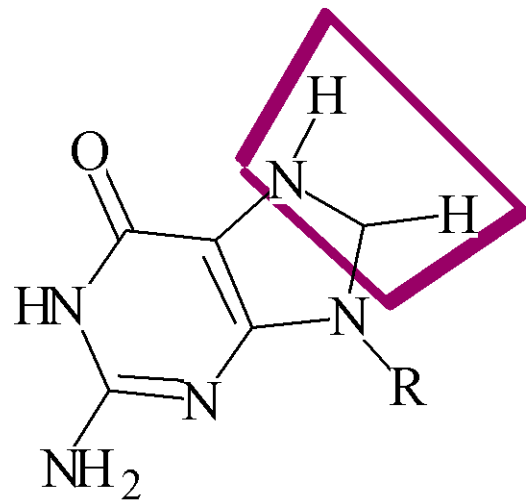
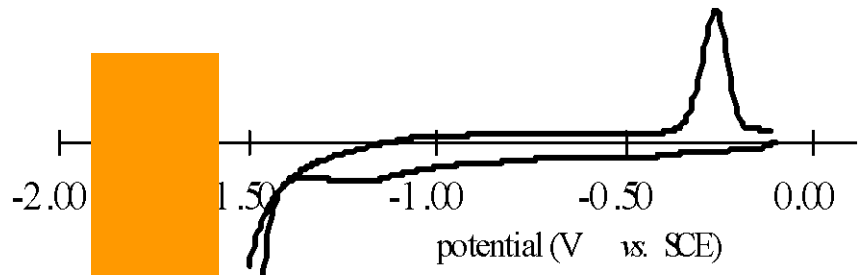
- at mercury electrodes, bases A,C and G undergo redox processes
- at carbon electrodes, purine bases can be oxidized
- sugar residues in nucleic acids can be oxidized at copper electrode Singhal, P.; Kuhr, W. G.: *Anal. Chem.* **1997**, 69, 3552-3557; *Anal. Chem.* **1997**, 69, 4828-4832.

Adenine and Cytosine are Reduced at the Mercury Electrode

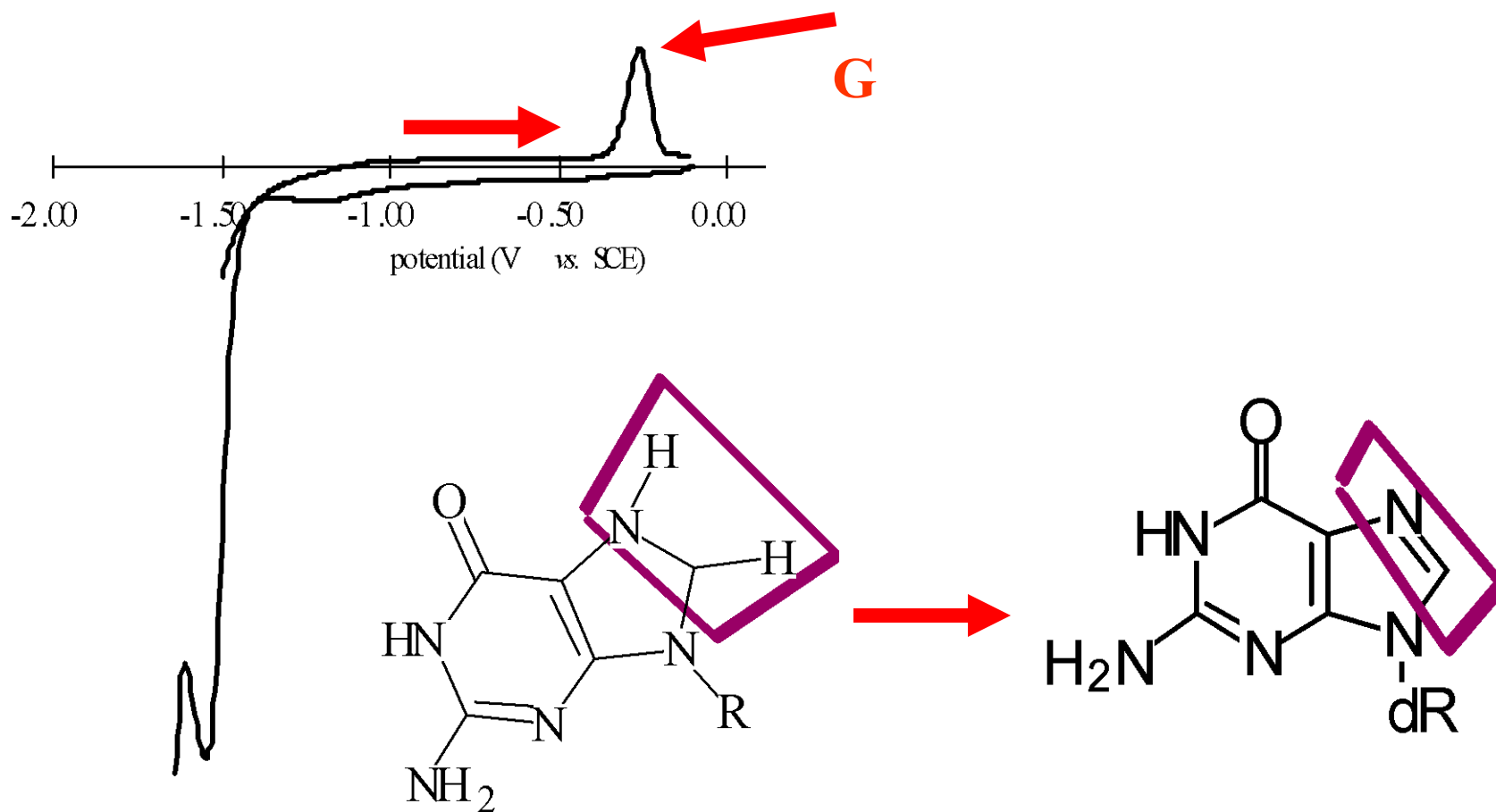


CA

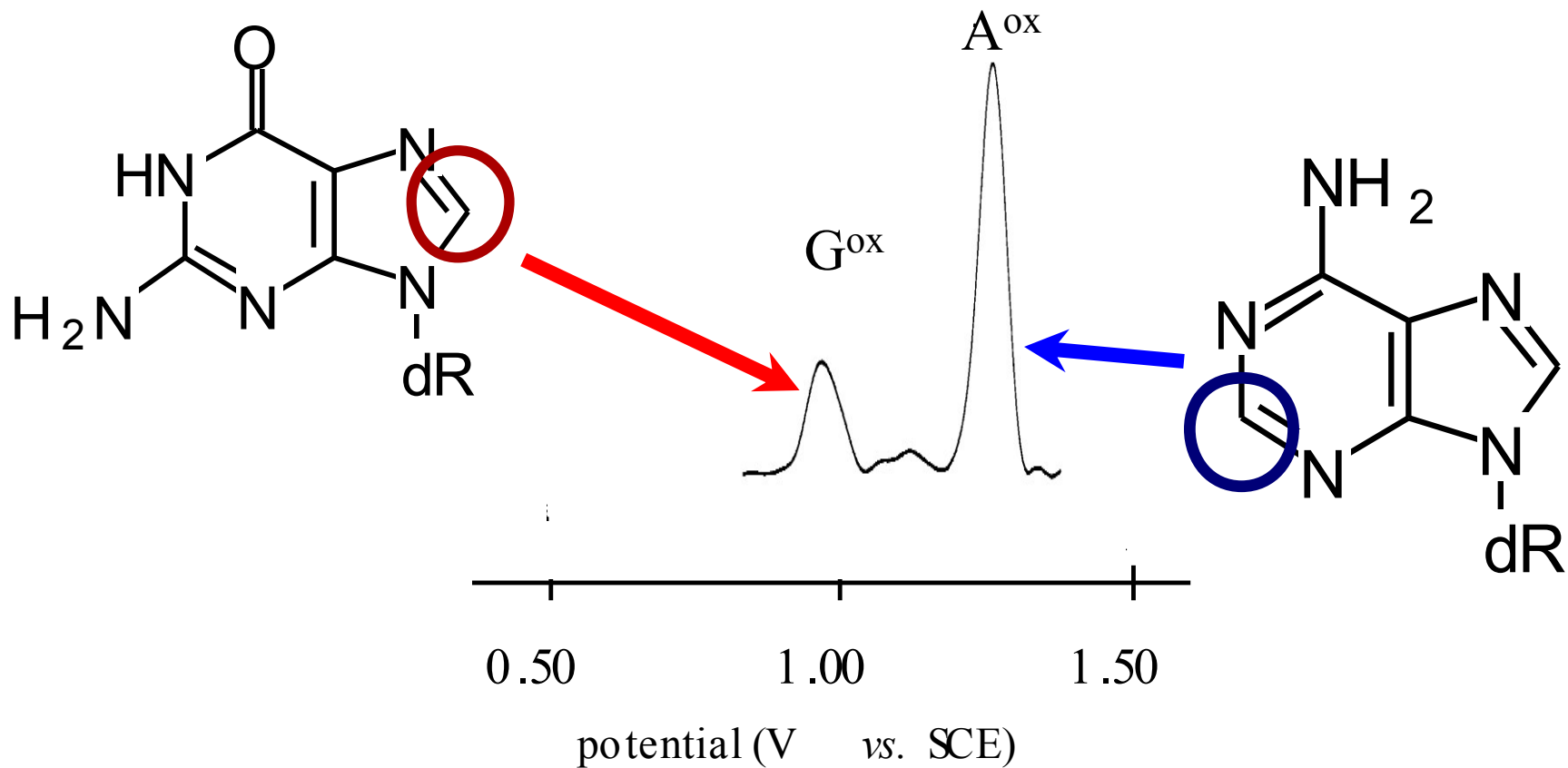
Guanine is reduced at the mercury electrode at highly negative potentials...



...and its reduction product yields anodic peak in cyclic voltammetry

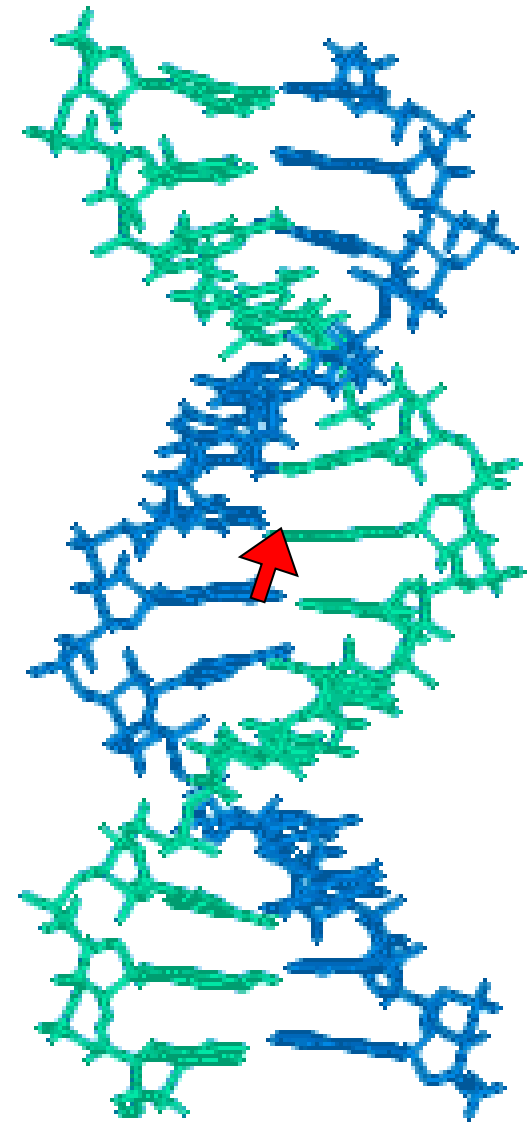
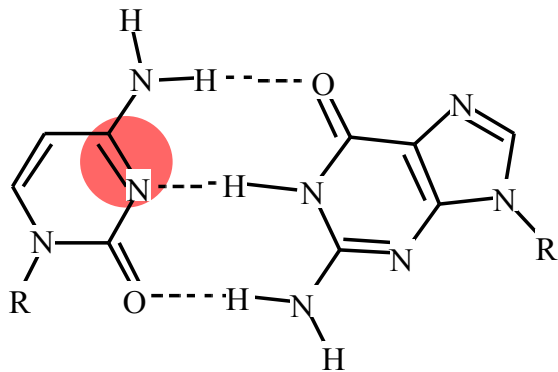
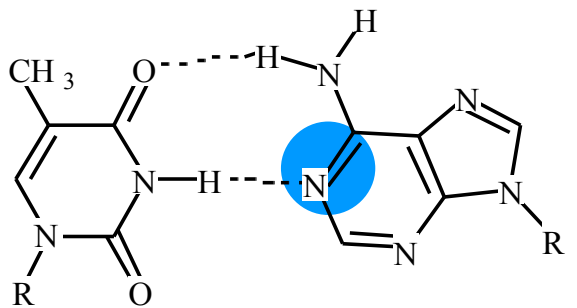


Guanine and adenine residues yield specific oxidation peaks at carbon electrodes

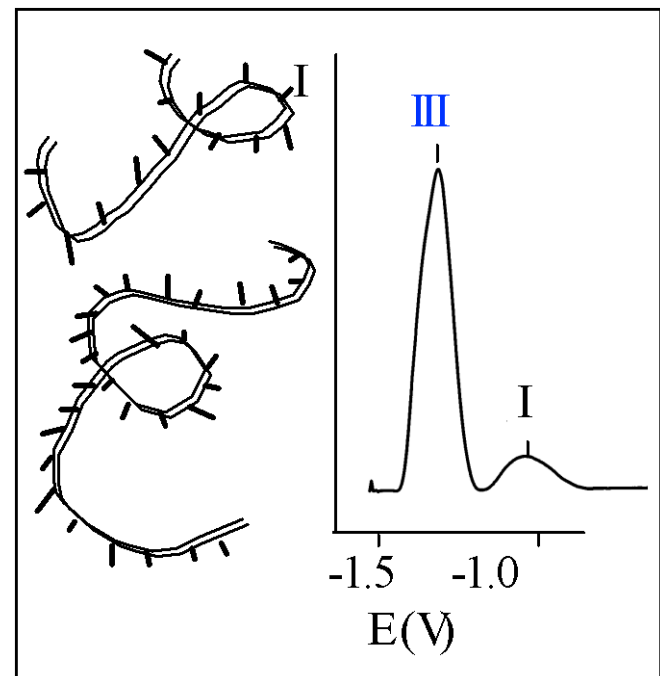
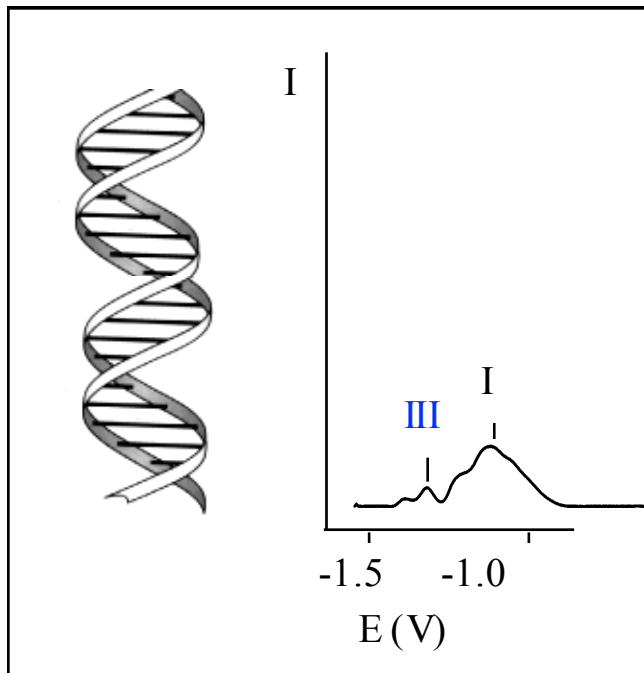


Reduction DNA signals at the 1 are strongly influenced by

- this is due to location of the **A** and **C** electroactive within the Watson-Crick hydrogen bonding system



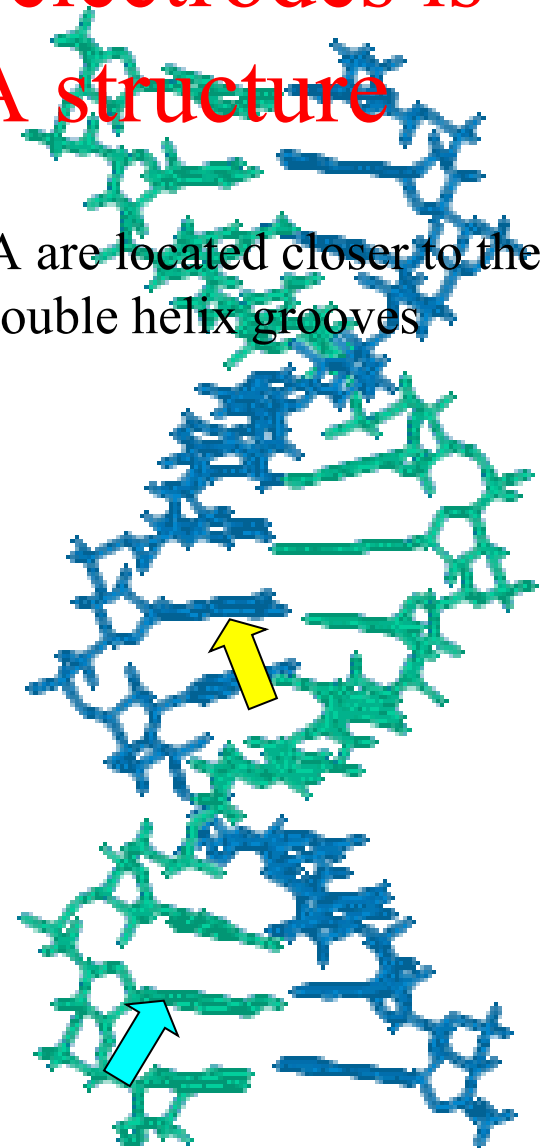
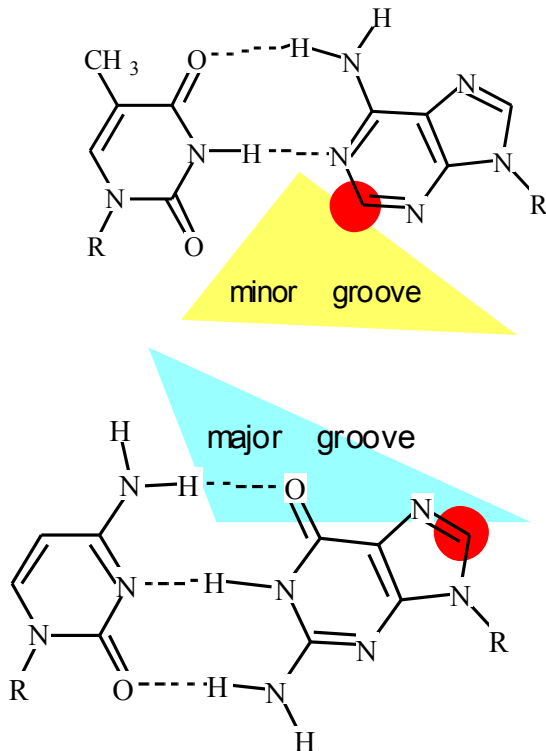
Reduction DNA signals at the mercury electrodes are strongly influenced by DNA structure



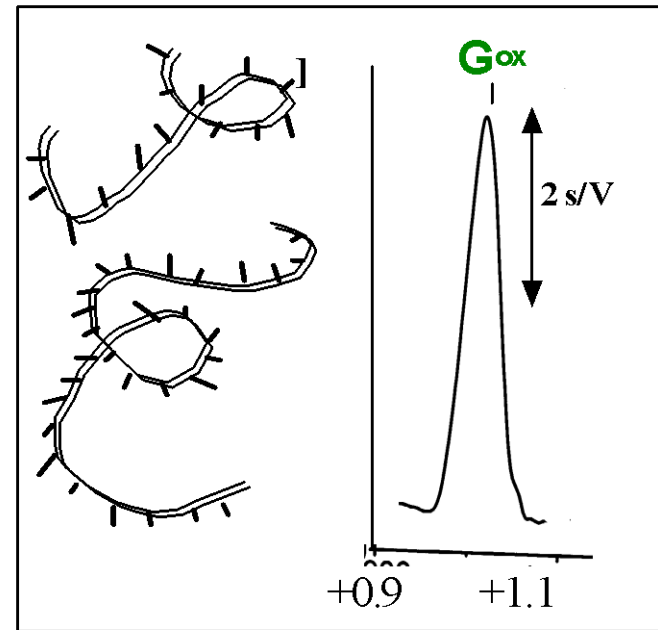
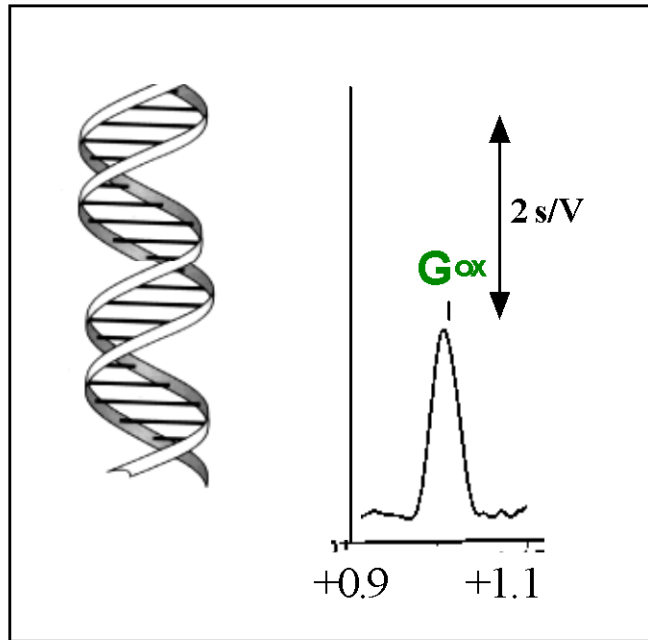
square-wave voltammetry

DNA oxidation at carbon electrodes is less influenced by DNA structure

- oxidation sites of guanine and adenine in dsDNA are located closer to the double helix surface and are accessible via the double helix grooves



DNA oxidation at carbon electrodes is less influenced by DNA structure

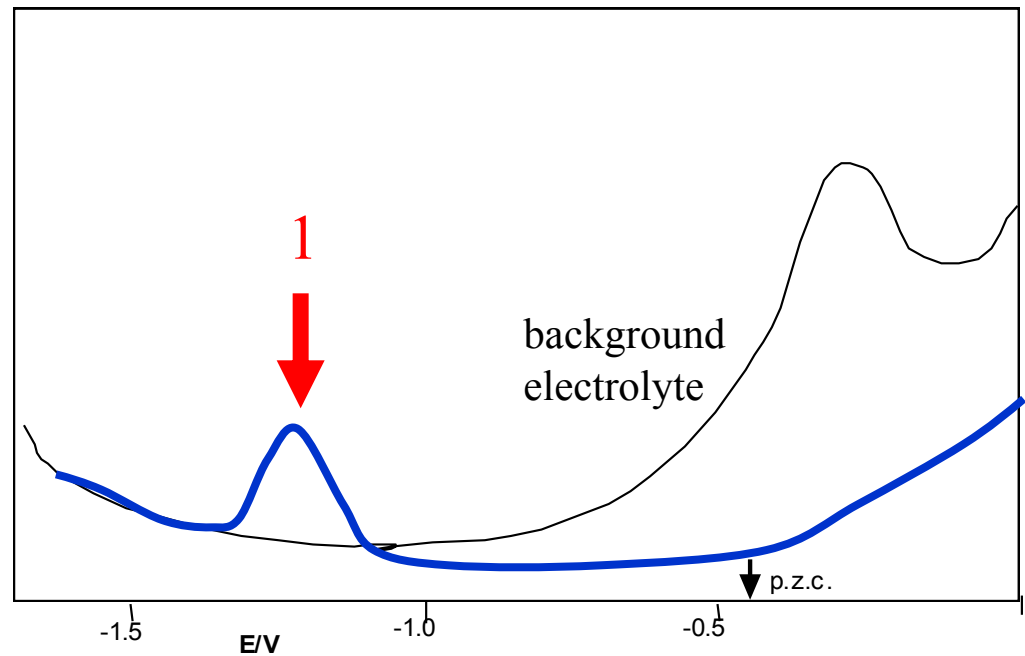
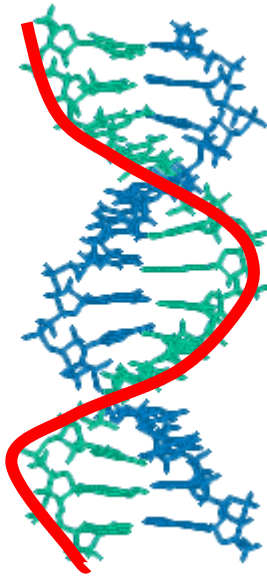


chronopotentiometry at CPE

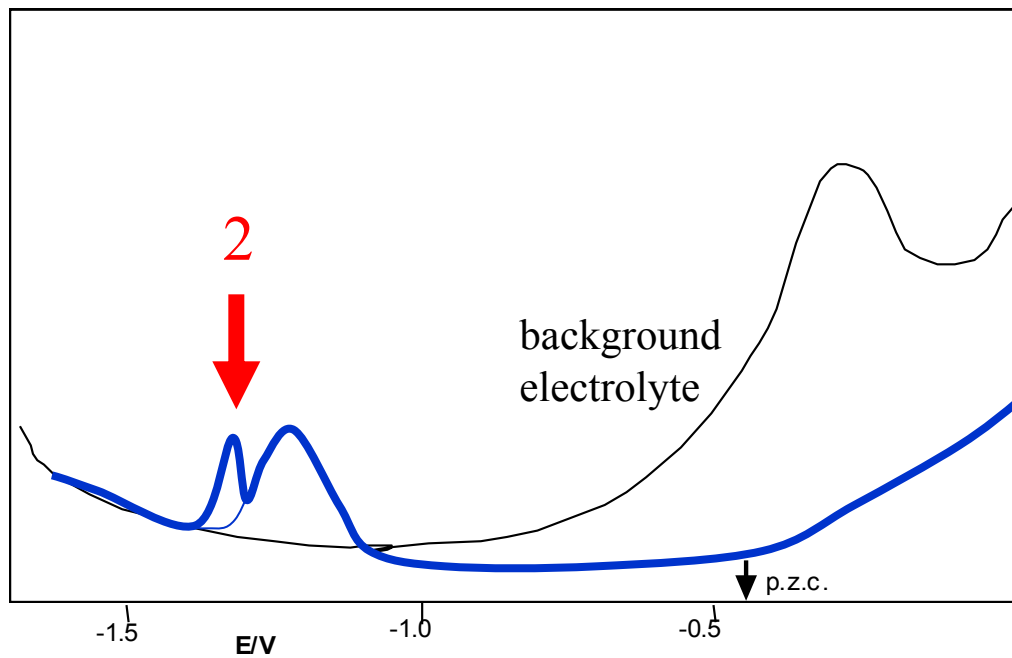
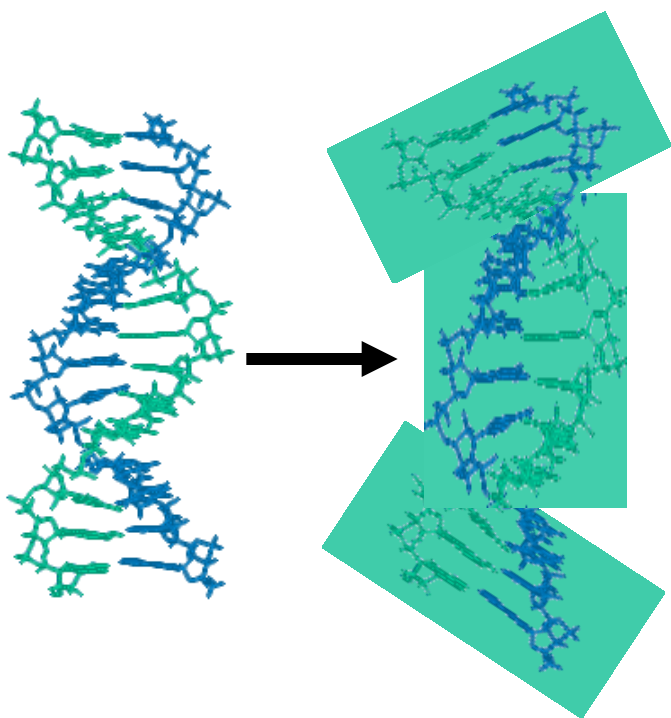
At mercury electrodes in weakly alkaline media,
adsorption-desorption (tensammetric) signals
of nucleic acids can be detected
(e.g., using AC polarography, voltammetry, AC Z)

- depending on the conditions and on **DNA structure**, individual components of the polynucleotide chains may be involved in adsorption/desorption processes

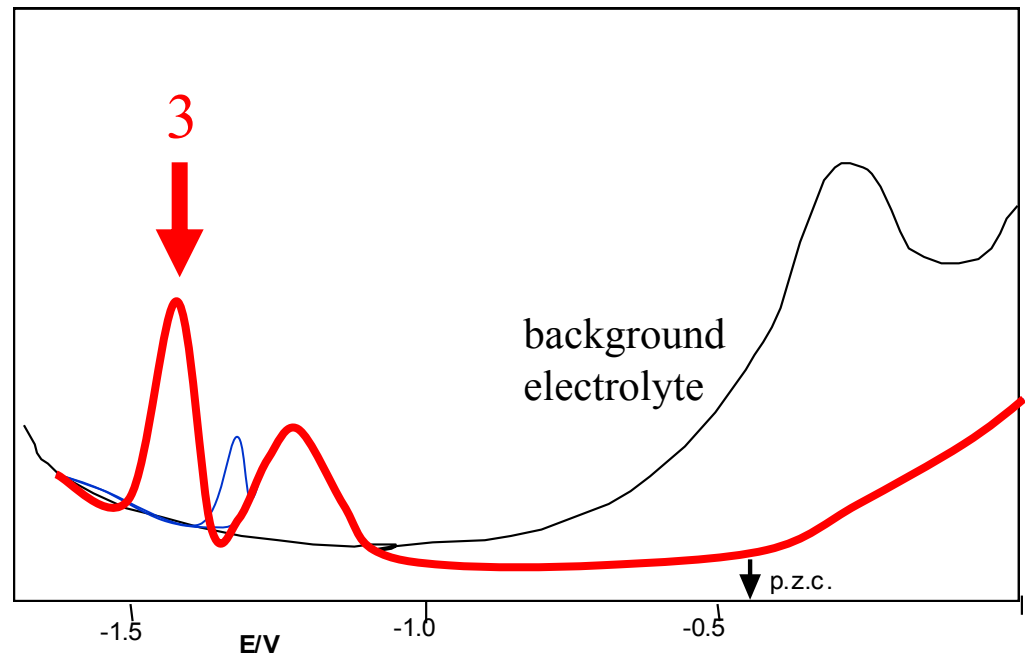
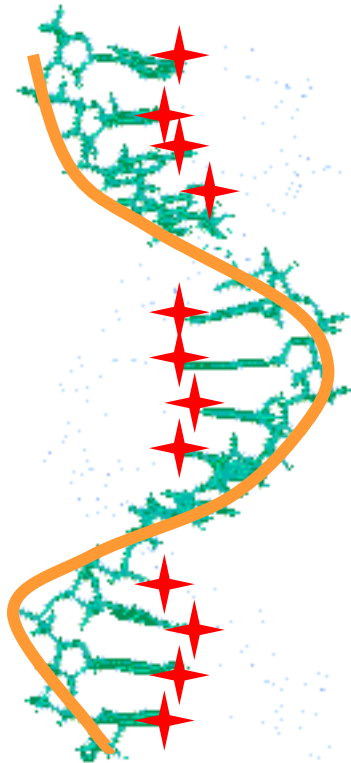
-at moderate ionic strength, **double-stranded DNA** yields **peak 1** due to desorption/reorientation of DNA segments adsorbed via the sugar-phosphate backbone



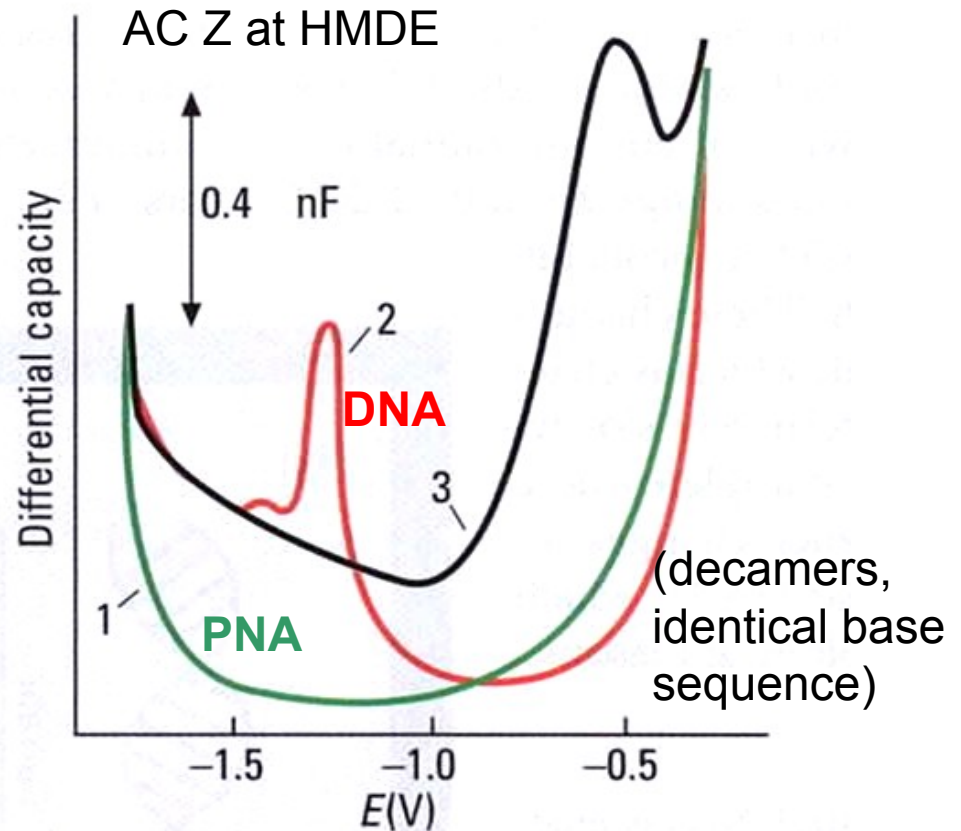
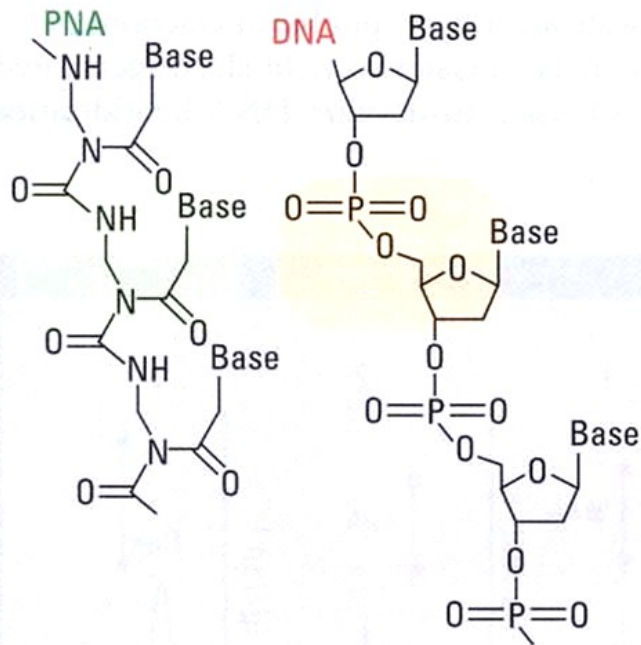
-distorted or regions of double-stranded DNA yield **peak 2**



single-stranded (denatured) DNA yields
peak 1 (due to the sugar-phosphate backbone) and **peak 3** due to
desorption/reorientation of DNA segments adsorbed via
freely accessible bases



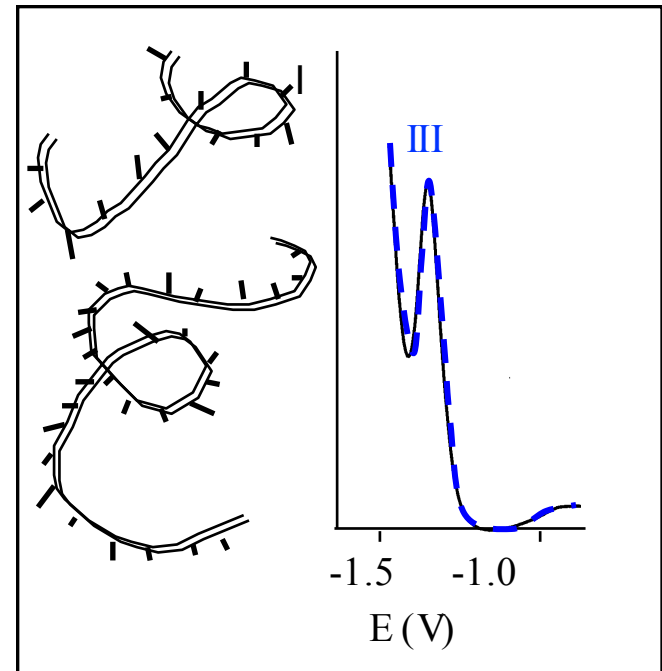
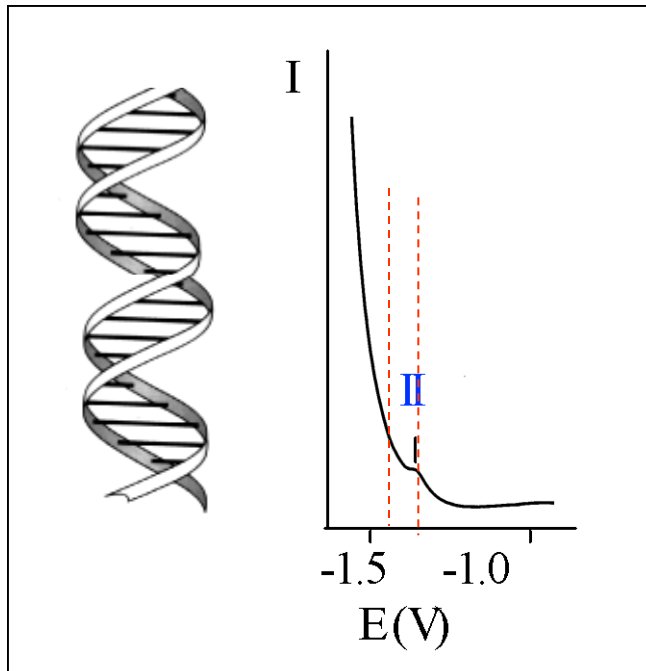
adsorption/desorption behavior of DNA at electrodes is strongly related to negative charge of its sugar-phosphate backbone (together with a strong adsorption of nucleobases via hydrophobic forces)



peptide nucleic acid: DNA analogue with neutral backbone

differential pulse polarography

- used in nucleic acid studies in the 60-70's
- discrimination between ss and dsDNA



differential pulse polarography

• peak II: high sensitivity to subtle changes of dsDNA structure (& dynamics)

• DNA premelting

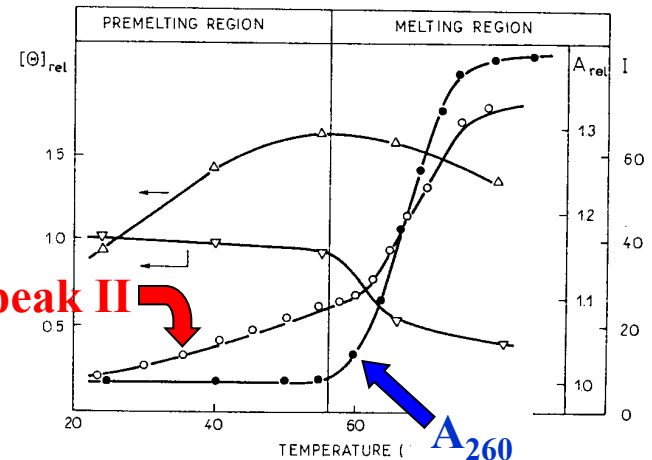
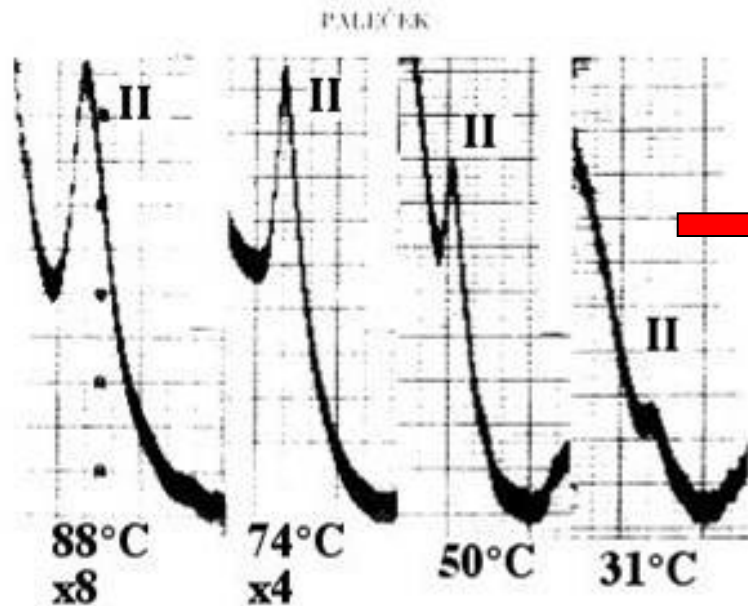
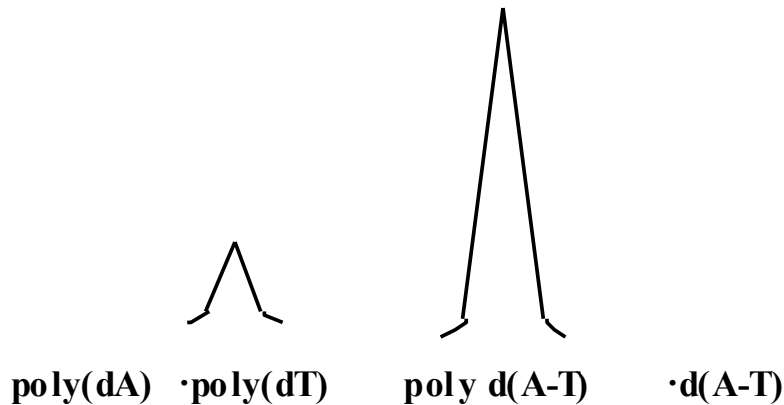


FIG. 2. Thermal transition of calf thymus DNA followed by circular dichroism (CD), derivative pulse polarography and spectrophotometry. CD: Δ — Δ , positive band (275 nm); ∇ — ∇ , negative band (245 nm); \circ — \circ , polarography; \bullet — \bullet , absorbance at 269 nm. θ_{rel} , the ratio between the ellipticity at the given temperature and at 25°C. A_{rel} , the ratio between the absorbance (260 nm) at the given temperature and the absorbance at 25°C. I, the height of the pulse-polarographic peak in divisions. Adapted from Paleček and Frič (21).

¹ Nonstandard abbreviations: CD, circular dichroism; ORD, optical rotatory dispersion; ds, double-stranded; ss, single-stranded.

differential pulse polarography

• **polymorphy of DNA double helix: its structure depends on the nucleotide sequence**



B. subtilis and B. brevis DNAs have the same G+C content and different nucleotide sequence

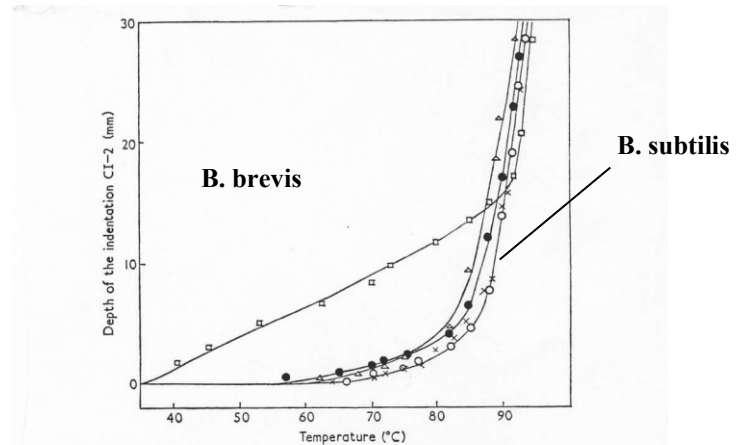


FIG. 12. Thermal transition of DNA's isolated from bacteria of the genus *Bacillus*. DNA at a concentration of 100 $\mu\text{g/ml}$. in 0.25 M-ammonium formate plus 0.025 M-sodium phosphate (pH 7.0).

—●—●—, *B. subtilis* 168; —×—×—, *B. natto*; —○—○—, *B. subtilis* var. *niger*;
—△—△—, *B. subtilis* var. *atterimus*; —□—□—, *B. brevis* (ATCC 9999).

P 524 polaroscope, dropping mercury electrode polarized with repeated cycles of a.c. The measurements were carried out in the laboratory of Prof. J. Marmur, Department of Biochemistry, Brandeis University, Waltham, Mass., U.S.A.

differential pulse polarography

- strand breaks

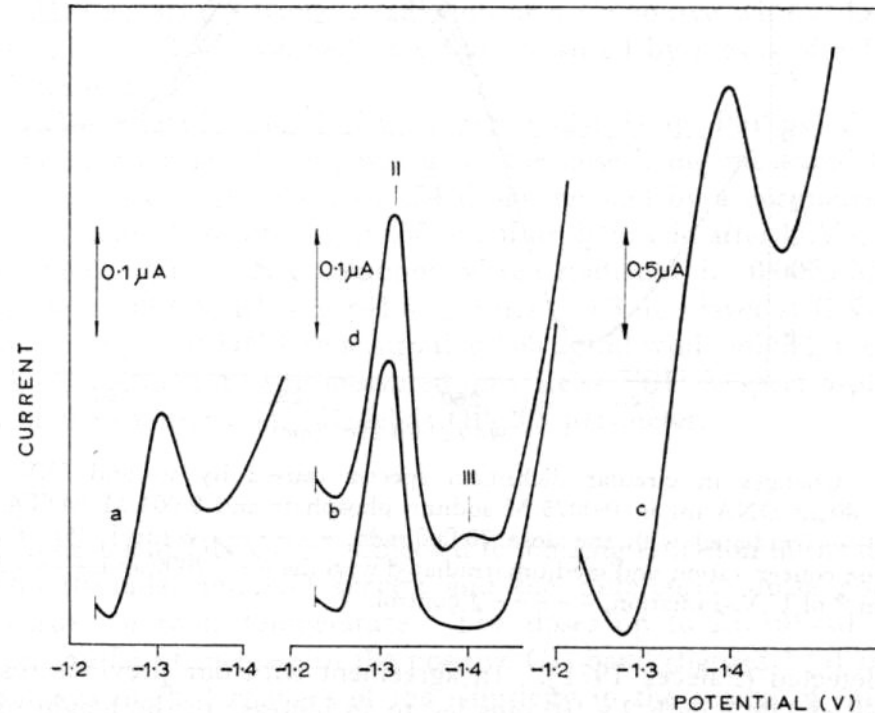


Figure 3. Differential pulse polarograms of DNA irradiated with ionizing radiation. DNA was irradiated in the concentration 460 $\mu\text{g}/\text{ml}$ in the medium given in figure 1. (a) a control; (b) 10^4 rads; (c) 6×10^5 rads; (d) the sample (b) heated at 50°C for 6 min and quickly cooled. The differential pulse polarograms were measured in 0.3 M ammonium formate, 0.1 M sodium phosphate, pH 6.9 at DNA concentration 400 $\mu\text{g}/\text{ml}$. Sensitivity of the apparatus was $1 \mu\text{A}$ in parts (a), (b) and (d), and $5 \mu\text{A}$ in part (c).

differential pulse polarography

➤ double helix distortions due to nucleobase photoadducts

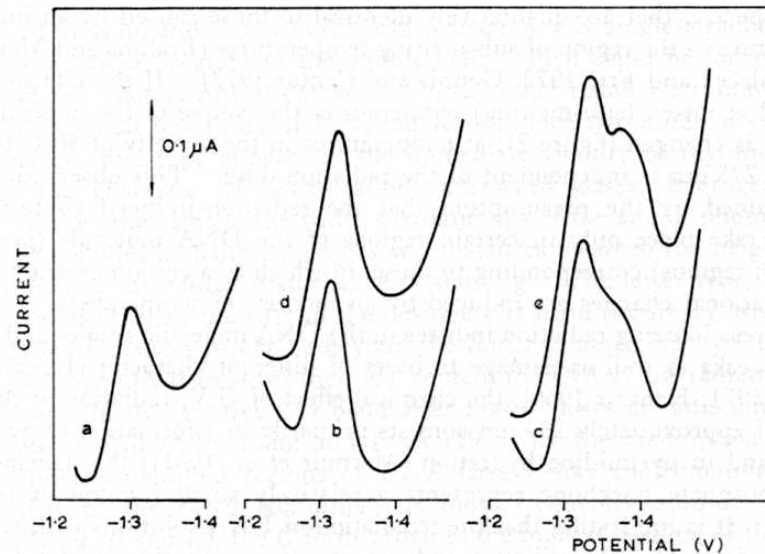


Figure 6. Differential pulse polarograms of DNA irradiated with U.V.-radiation. DNA was irradiated in the concentration $460 \mu\text{g/ml}$ in the medium given in figure 1. (a) a control; (b) $2.1 \times 10^4 \text{ erg mm}^{-2}$; (c) $6 \times 10^5 \text{ erg mm}^{-2}$; (d), (e) the samples (b) and (c), respectively, heated at 50°C for 6 min and quickly cooled. The differential pulse polarograms were measured under the conditions given in figure 3 with the apparatus sensitivity $1 \mu\text{A}$.

differential pulse polarography

➤ chemical modification of DNA: platinum adducts

distinction of the kind of structural change caused
by modification with different Pt complexes

peak II: conformation distortion, base pairing preserved

peak III: base unpairing

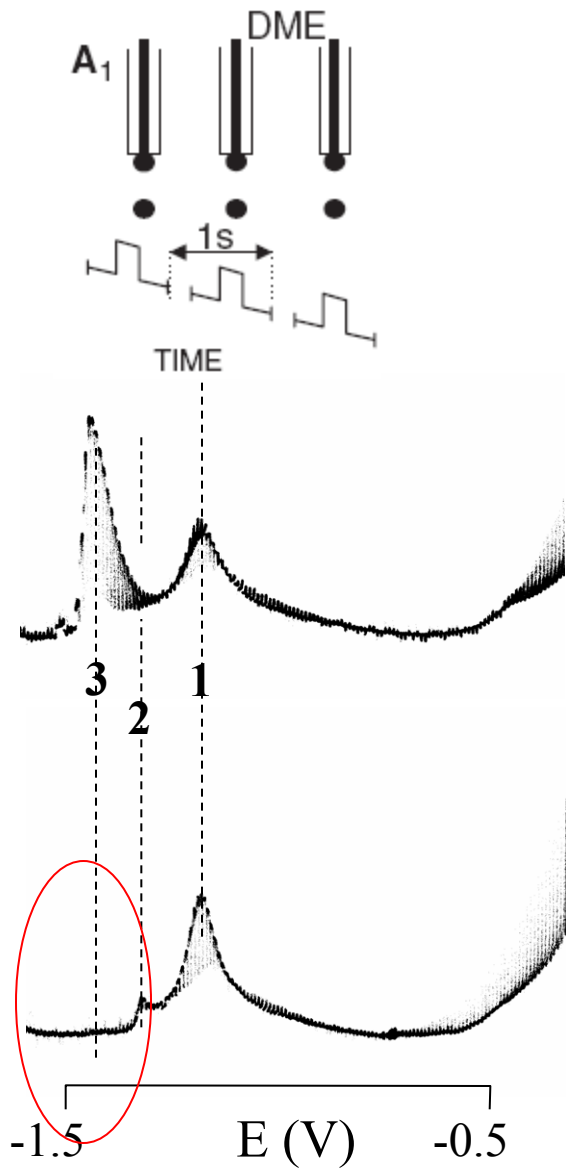


Fig. 7 Differential pulse polarographic analysis of CT-DNA modified by ORGANObisPt. DNA at a concentration of 0.4 mg/mL in 0.3 M ammonium formate with 0.01 M phosphate buffer, pH 6.8. *Curve 1*: control, unmodified DNA; *curves 2-6*: DNA modified by ORGANObisPt at $r_b = 0.001, 0.003, 0.005, 0.007, 0.01$, respectively; the *arrows* marked *II* and *III* indicate potentials E (against saturated calomel electrode) at which native or denatured DNA samples yielded DPP peaks II or III, respectively (see text)

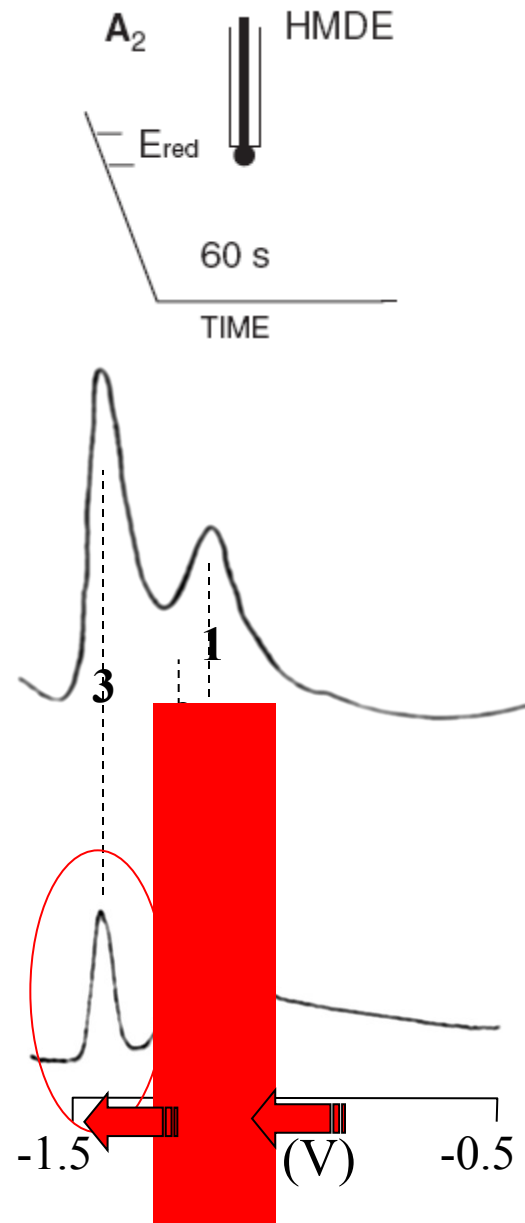
(Brabec et al.)

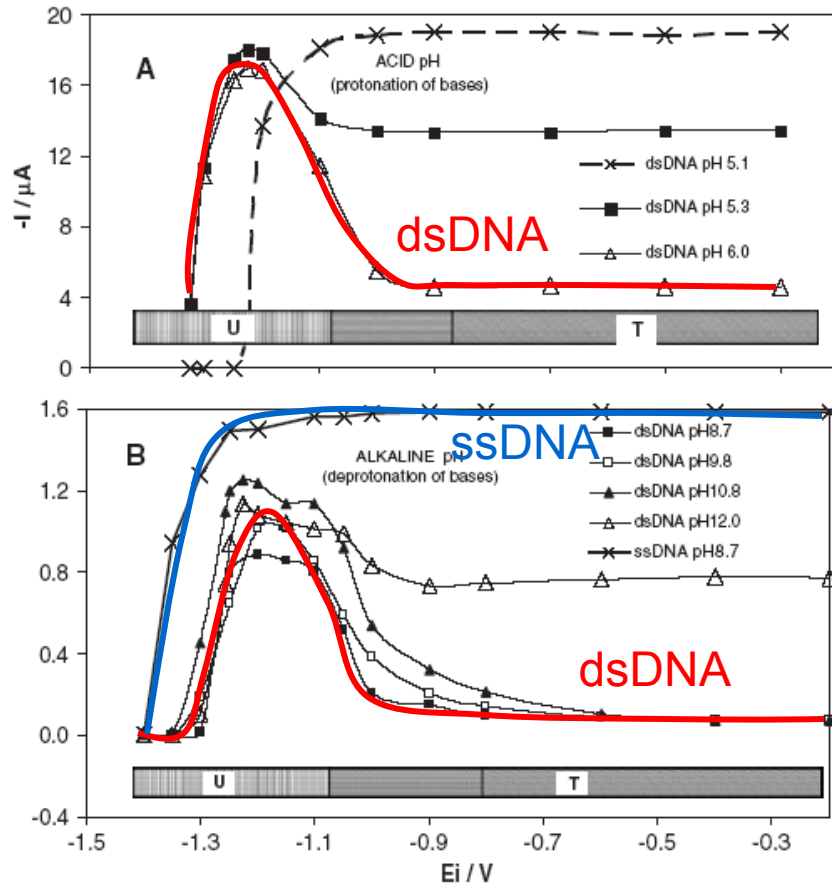
Changes of DNA structure at electrically charged surface

DME (SMDE)



HMDE





intensities of ssDNA-specific signals

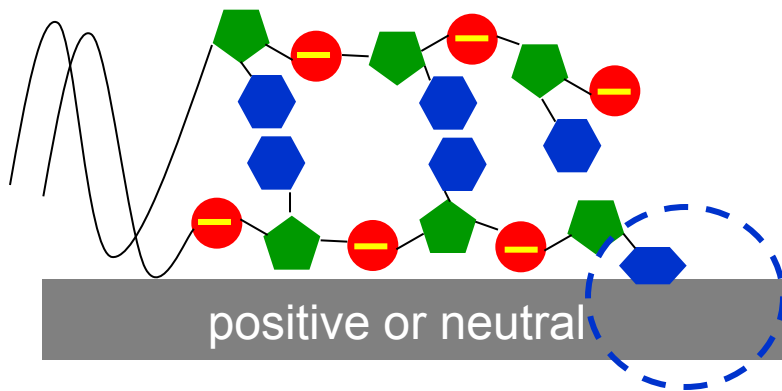
prolonged exposure to
(accumulation at)
potential given on x-axis




pH close to neutral (bases not ionized):

region T: – negligible structural changes
due to adsorption

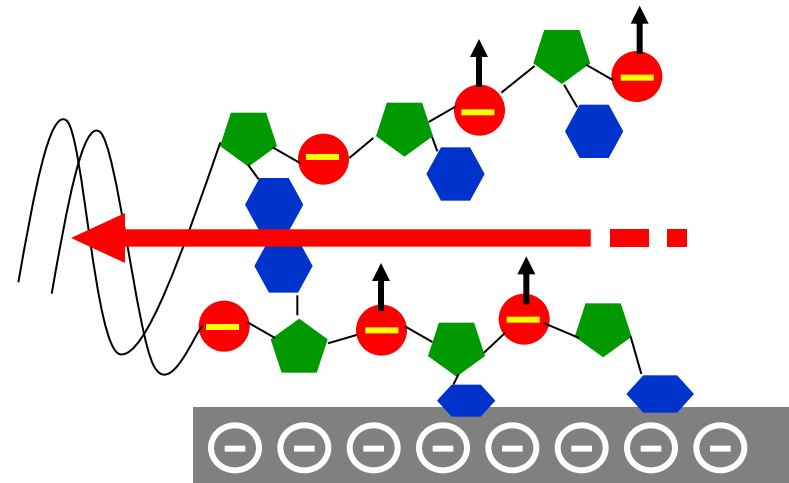
region U: surface denaturation

- close to the duplex ends (or single-strand breaks), some bases can be unpaired and make contact with the mercury surface



-  base
-  sugar
-  phosphate

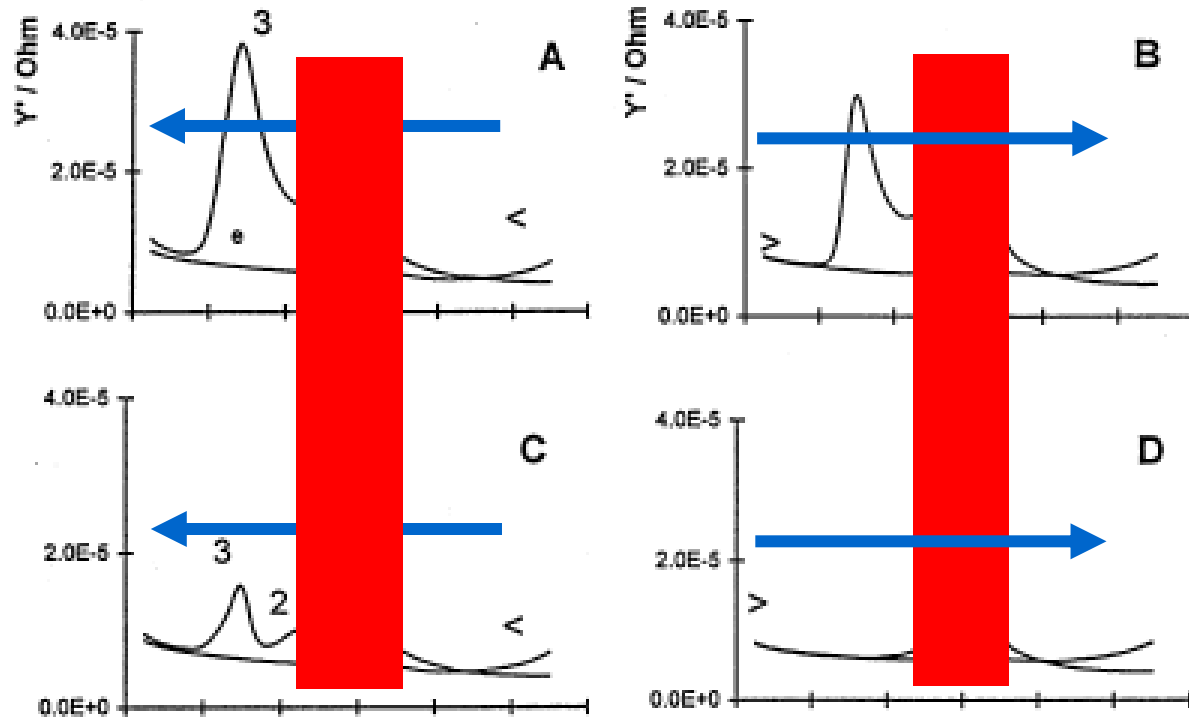
- phosphates repelled from negatively charged surface
- randomly adsorbed bases represent relatively firm anchor sites
- constraints in the double helix cause its (slow) unwinding
- more (unpaired) bases are coming into contact with the electrode



(in real situation the strand must rotate around one another; the process requires repeated adsorption/desorption events)

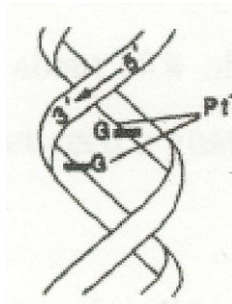


effects of initial potential and scan direction

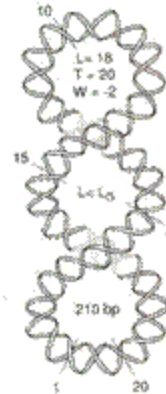


alternative models

- not double helix unwinding but conformation transition of dsDNA
- potential induced „ π -state“ of dsDNA involving B-A transition at the surface (Berg)
- „ladder DNA“ structure (Nurnberg)
- these models do not accord with numerous experimental data that support the unwinding model:



duplex with covalently cross-linked strands:
limited unwinding



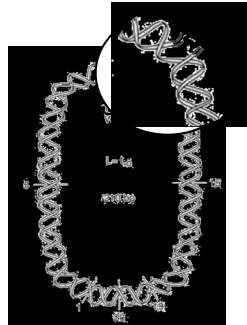
covalently closed circular DNAs:
limited unwinding

DNA with or without ends

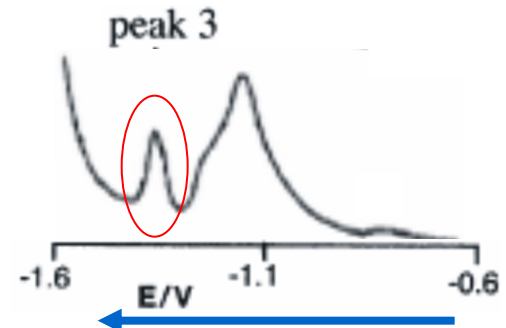
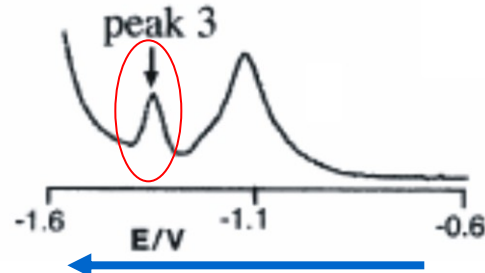
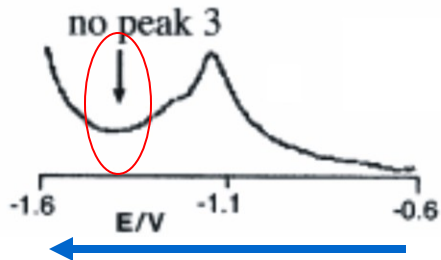
supercoiled



open (nicked) circular



linear



detection of DNA strand breaks using supercoiled DNA and mercury electrodes

supercoiled DNA

structural transitions induced by
DNA supercoiling

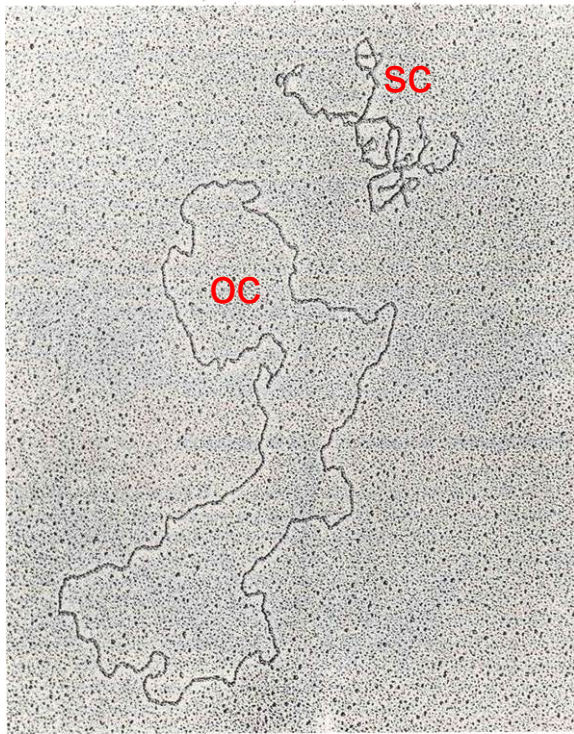
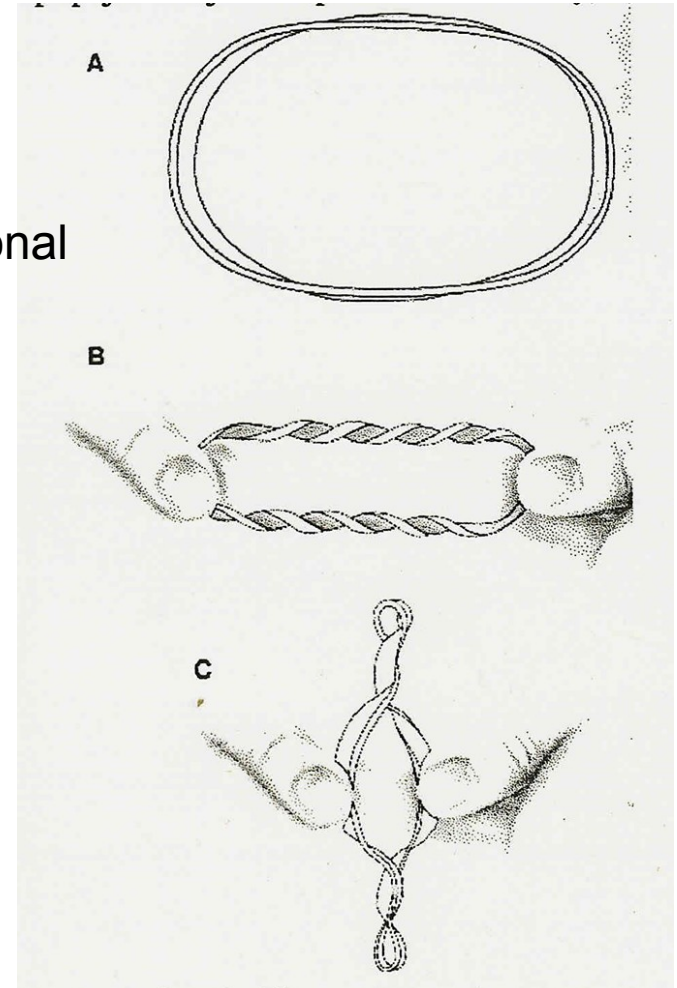


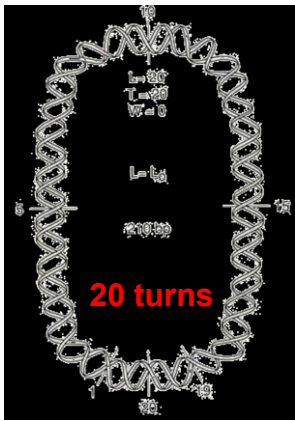
Figure 3.2 Electron micrograph of two forms of DNA. The tangled, twisted molecule is supercoiled DNA, originally called Form I DNA. When circular molecules are relaxed (or nicked) (Form II DNA), they lose the twists. A linear molecule (not shown) is called Form III. The plasmid molecules shown are 9000 bp in length. Courtesy of Jack D. Griffith.

ribbon model:

upon introducing torsional and/or bending stress, DNA behaves like an elastic ribbon

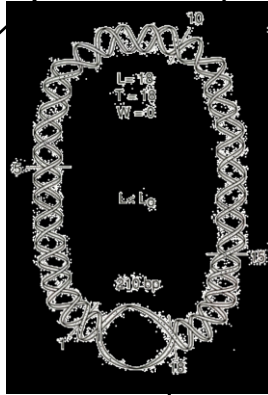
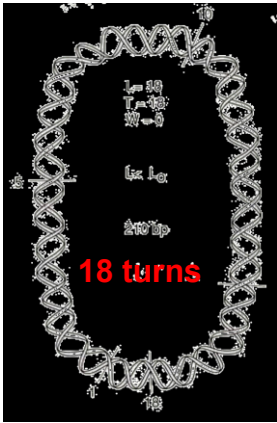


Vinograd, 1960's: **two forms of circular viral DNA**
(sedimentation velocity or sedimentation equilibrium studies)

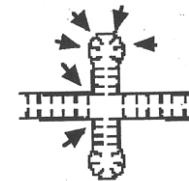


relaxed covalently closed circular DNA

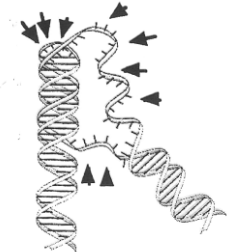
(constraint due to the twist deficit causes formation of the superhelix)



negatively supercoiled (sc) DNA

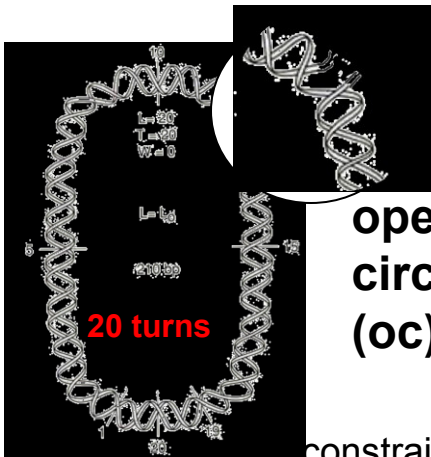


cruciform



triplex

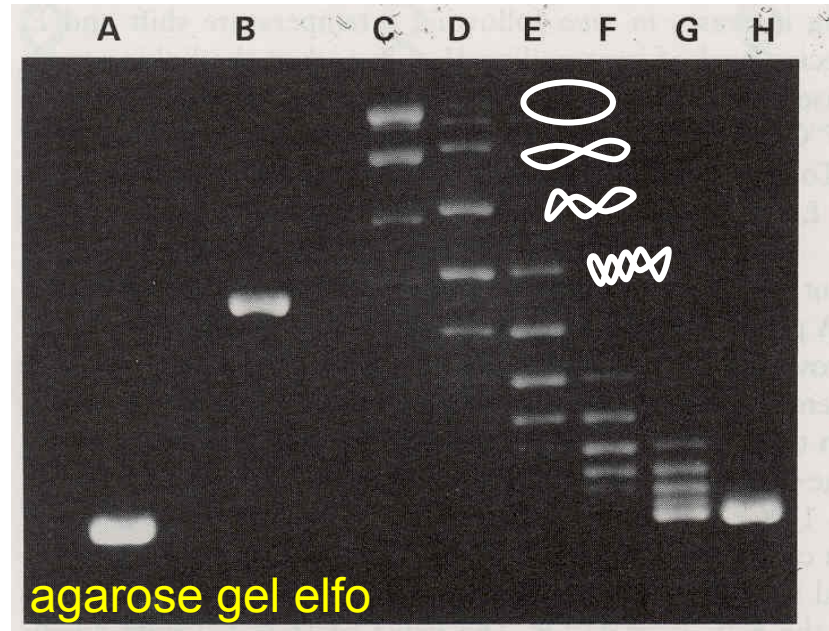
negative superhelicity is absorbed by local opening of the double helix



open circular (oc) DNA

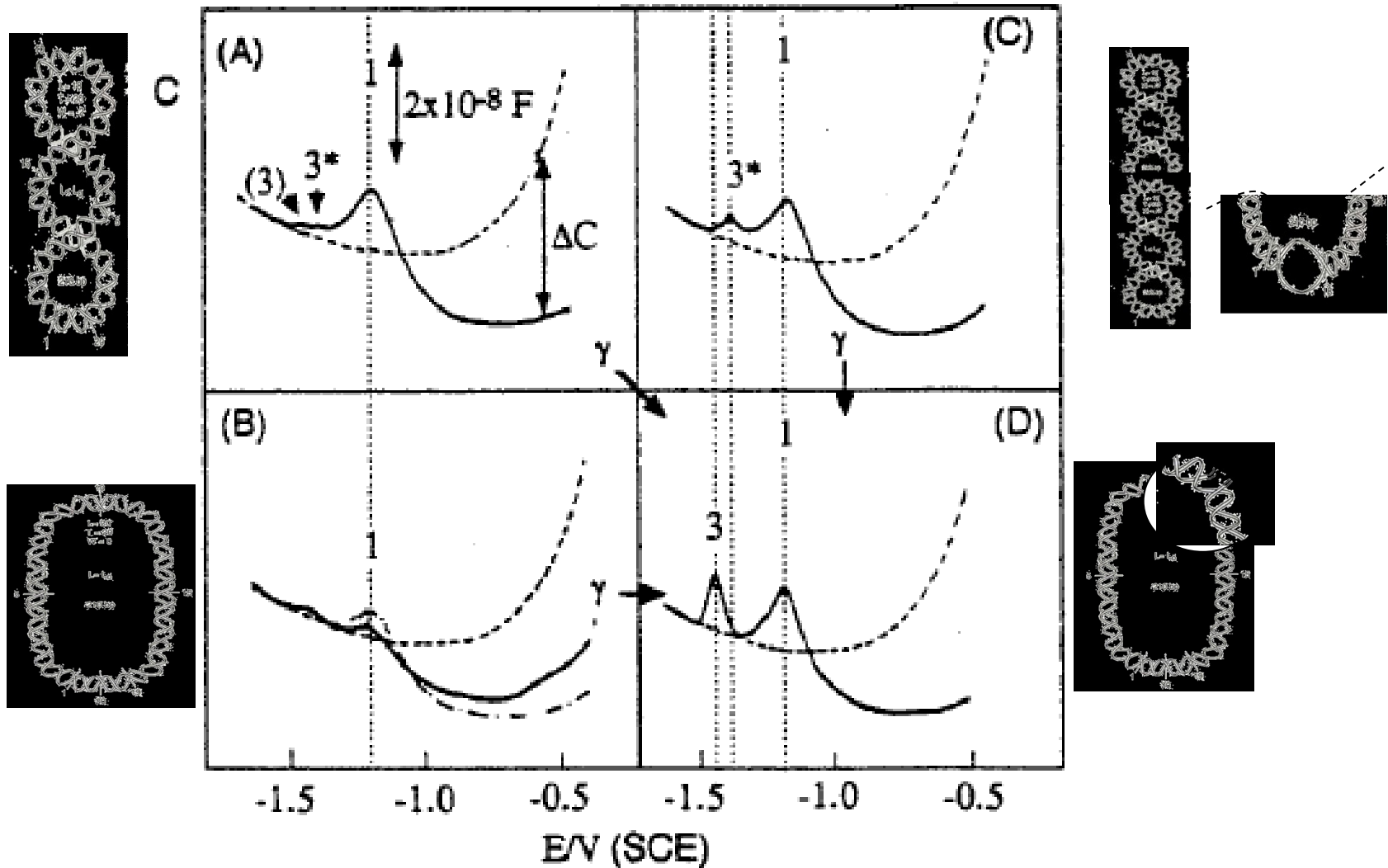
no constraint – free rotation at the strand break

DNA topoisomers differ in the superhelicity level

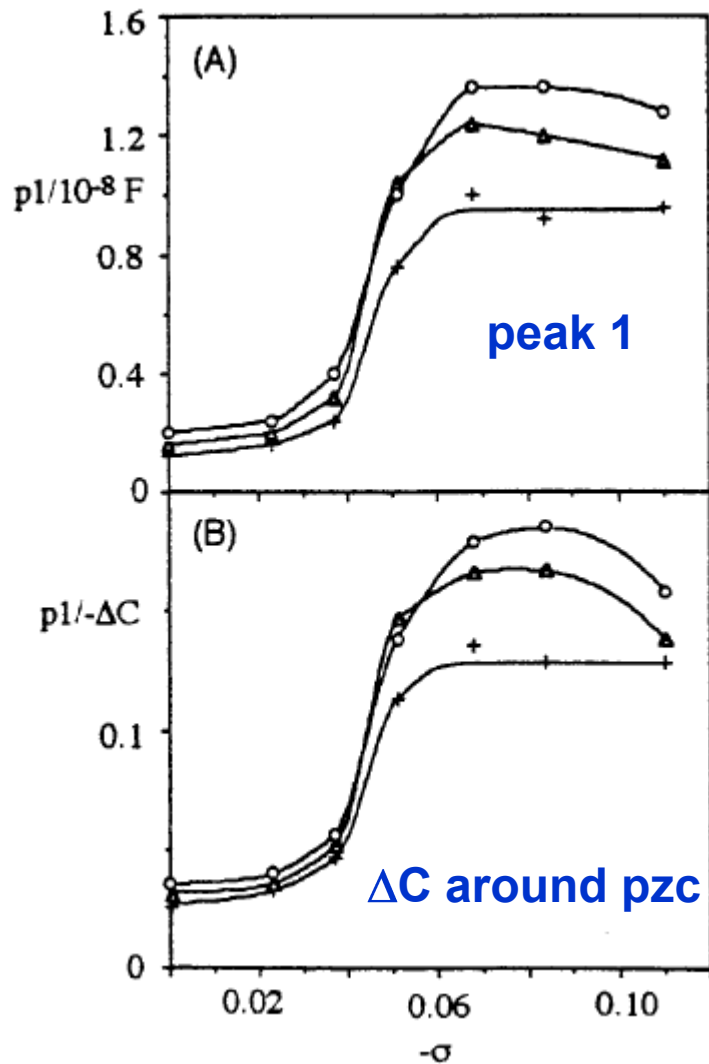


Two Superhelix Density-Dependent DNA Transitions Detected by Changes in DNA Adsorption/Desorption Behavior†

Miroslav Fojta,[‡] Richard P. Bowater,[§] Veronika Staňková,[‡] Luděk Havran,[‡] David M. J. Lilley,^{||} and Emil Paleček^{*,‡}

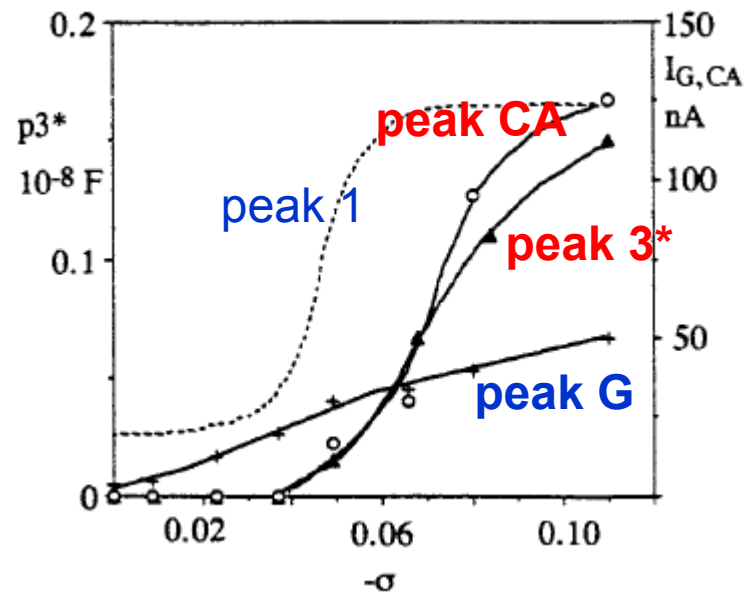


transition 1



overall shape of the molecules

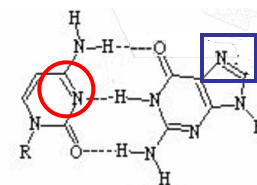
transition 2



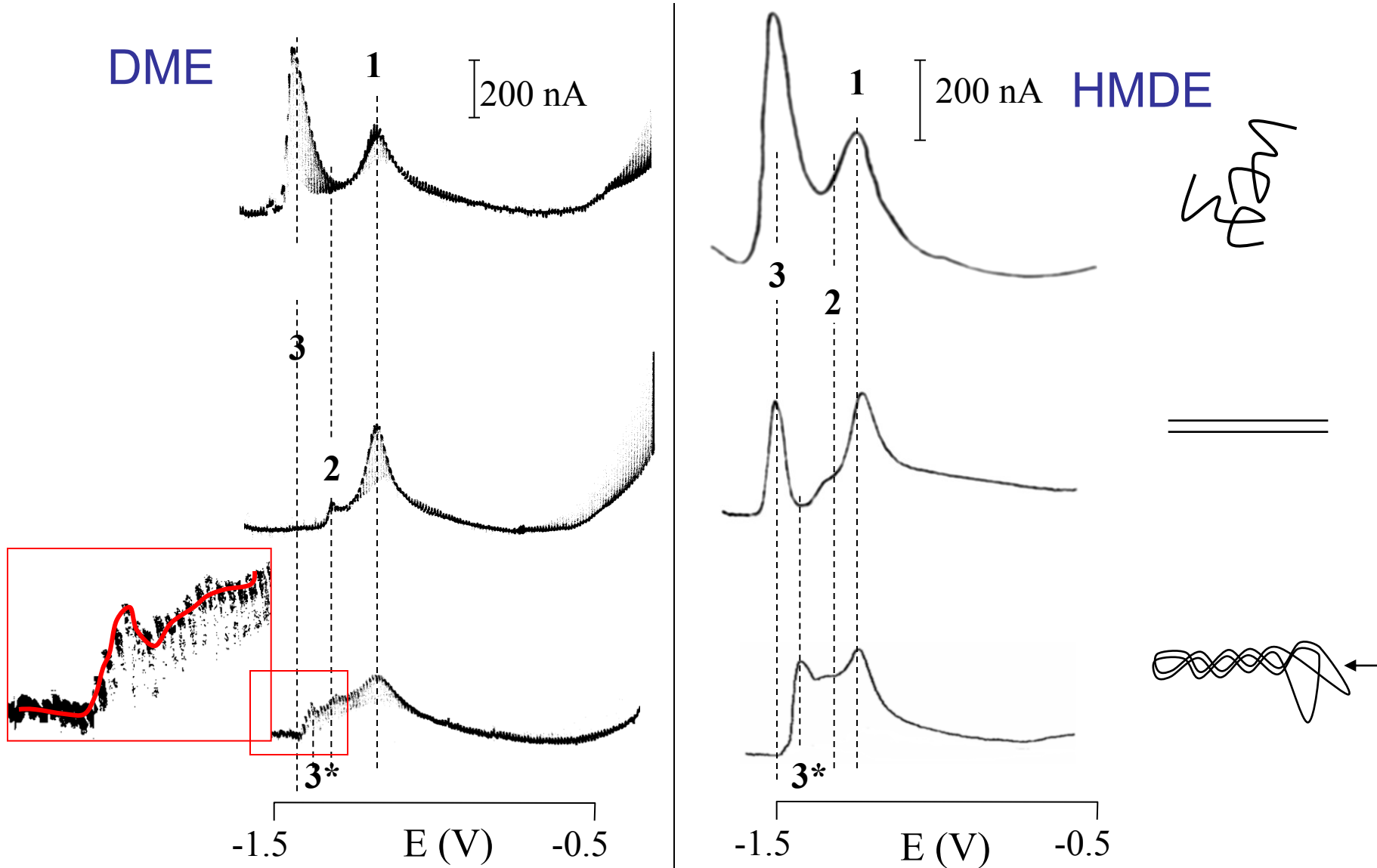
peak 3* is due to local helix opening (base unpairing)

peak CA (similar dependence on $-\sigma$ as peak 3*) is known to be sensitive to DNA denaturation

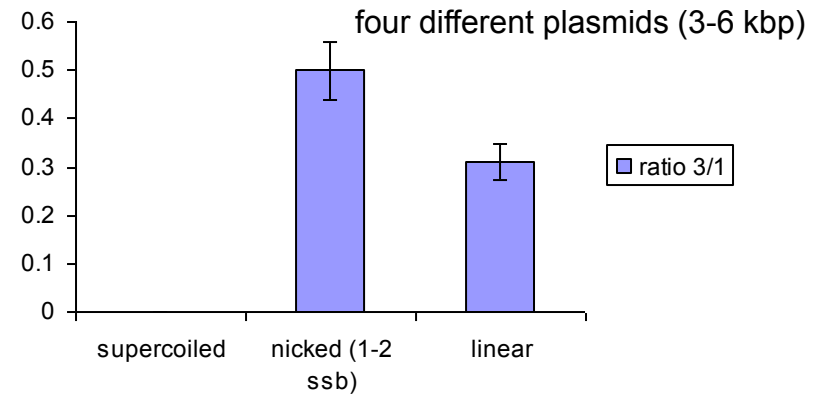
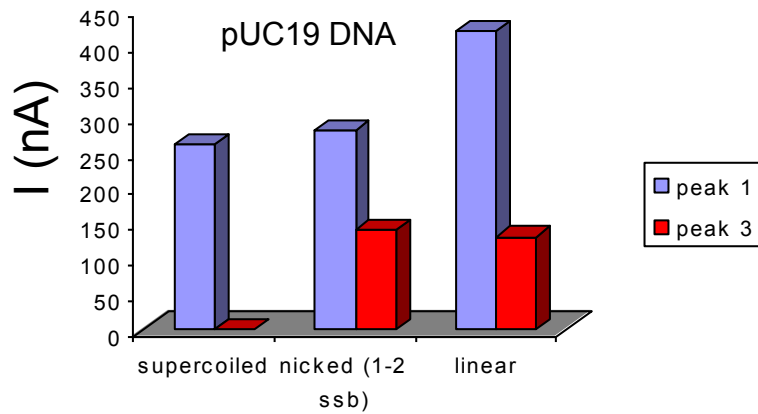
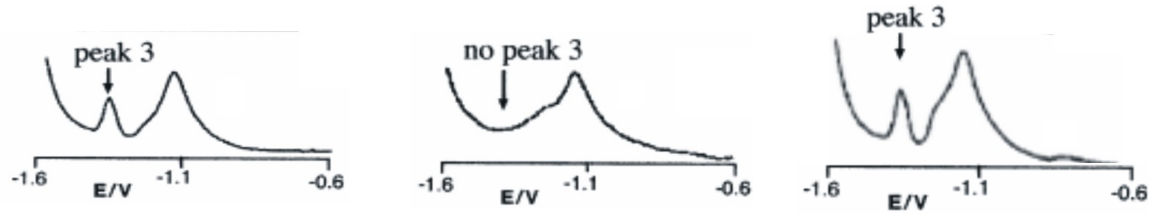
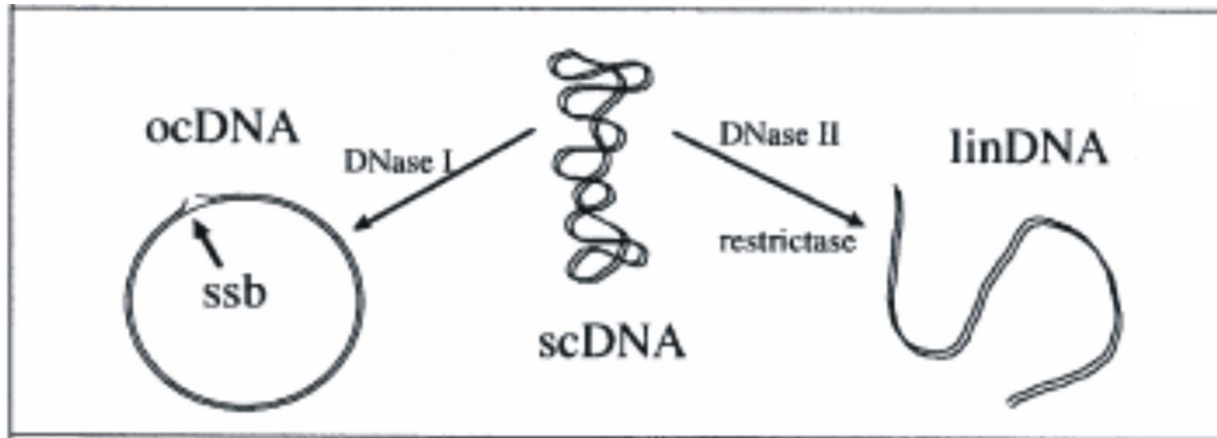
peak G (no transition 2) is less sensitive to DNA structure



Studies with DME: peak 3* is due to helix opening in solution (in difference to peak 3 produced by oc or linear dsDNA that is observed only at the HMDE)



circular and linear DNAs

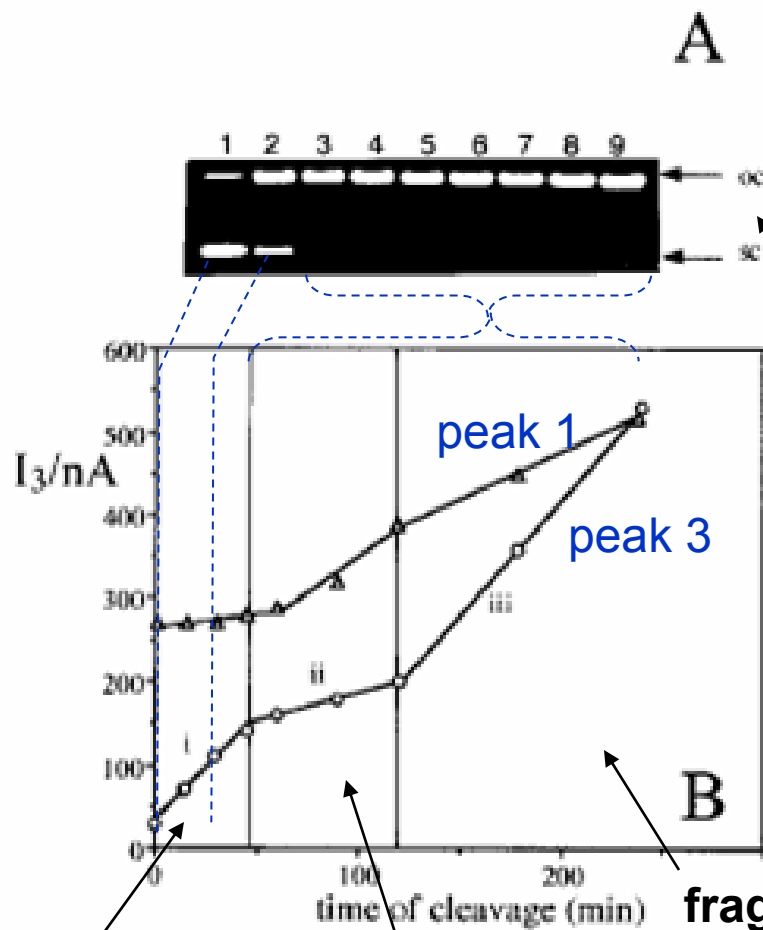


Cleavage of Supercoiled DNA by Deoxyribonuclease I in Solution and at the Electrode Surface

Miroslav Fojta,* Tatiana Kubičárová, and Emil Paleček

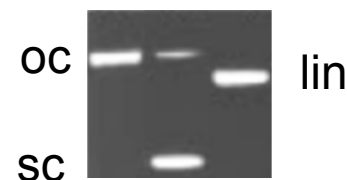
Institute of Biophysics of the Academy of Sciences of the Czech Republic, Královopolská 135, CZ-612 65 Brno, Czech Republic

monitoring of DNA cleavage in solution using electrochemistry



- DNase I introduces single strand breaks
- DNA treated with the enzyme, then adsorbed at HMDE

agarose gel elfo shows ocDNA as the only product of DNA cleavage during the experiment



fragmentation upon adsorption

linearization upon adsorption

nicked circular (sb accumulation)

Cleavage of Supercoiled DNA by Deoxyribonuclease I in Solution and at the Electrode Surface

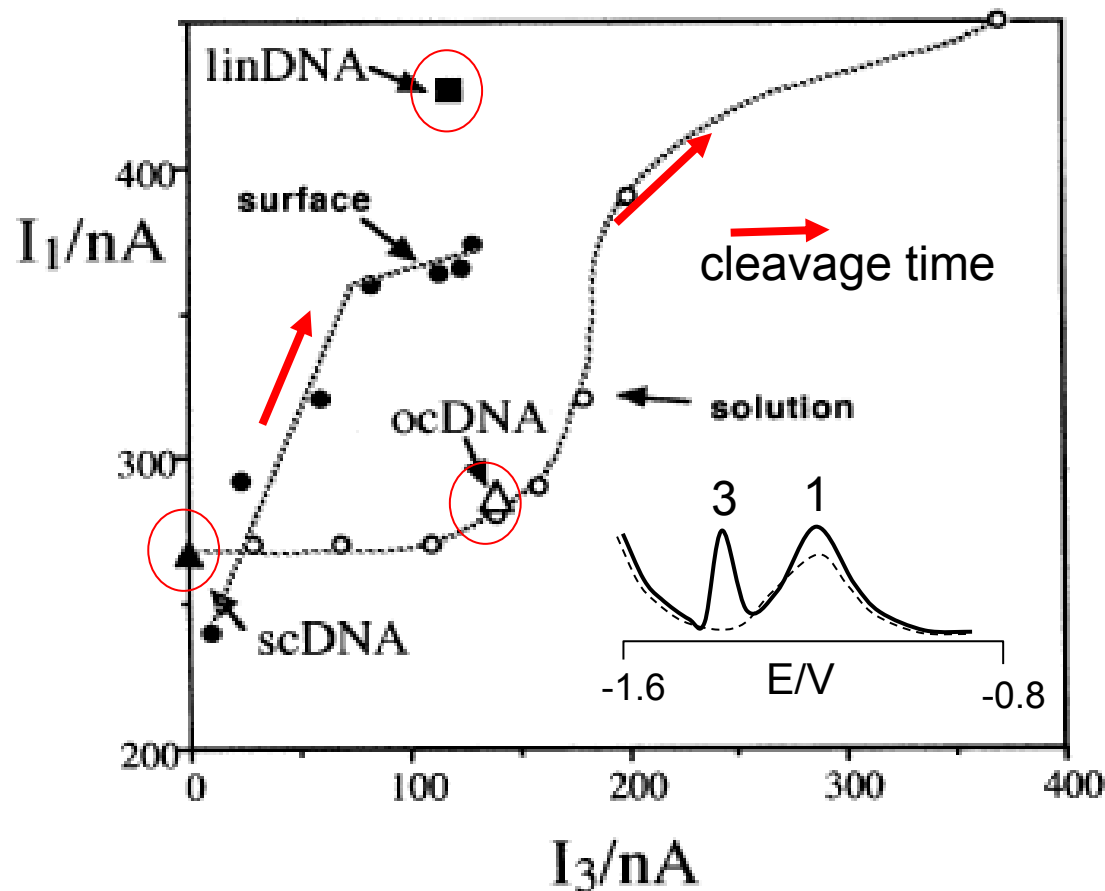
Miroslav Fojta,* Tatiana Kubičárová, and Emil Paleček

Institute of Biophysics of the Academy of Sciences of the Czech Republic, Královopolská 135, CZ-612 65 Brno, Czech Republic

different cleavage mechanisms in solution and at the surface?

„solution“: as previous slide

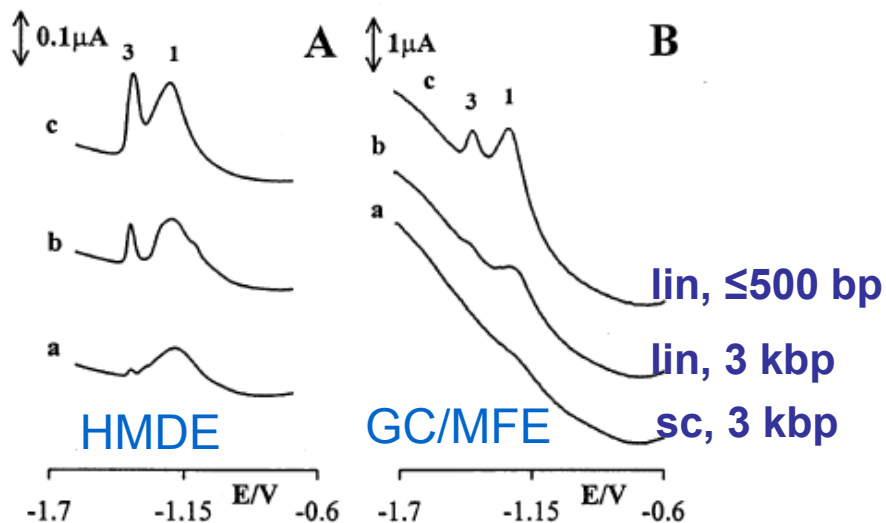
„surface“: DNA adsorbed at HMDE, then treated with the enzyme



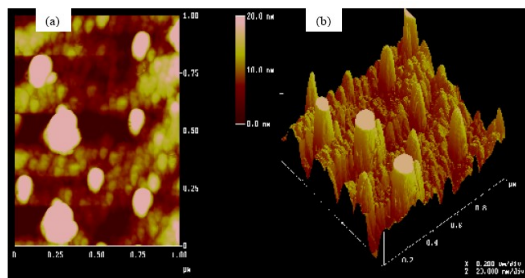
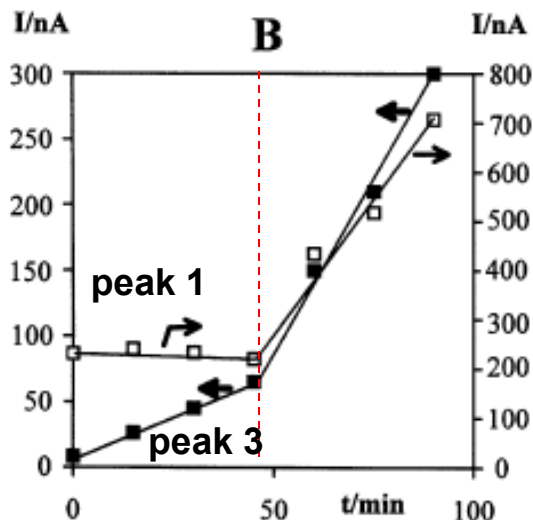
correlation between peak 3 and peak 1 peak heights

Mercury Film Electrode as a Sensor for the Detection of DNA Damage

Tatiana Kubičárová,⁺ Miroslav Fojta,^{**} Jasmina Vidic,⁺⁺ Luděk Havran,⁺ and Emil Paleček⁺

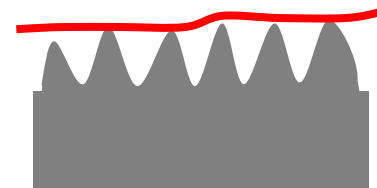


- at the GC/MFE: more pronounced effects of the DNA molecule length
- poor responses of long dsDNAs
- steeper dependence of peak heights on DNA cleavage extent (from the second phase)

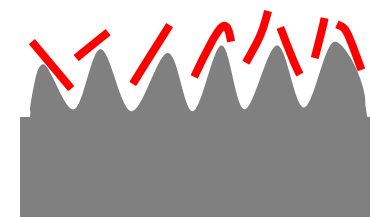


GC/MFE: not a smooth surface like HMDE

long rigid ds DNA: poor contact with the electrode surface



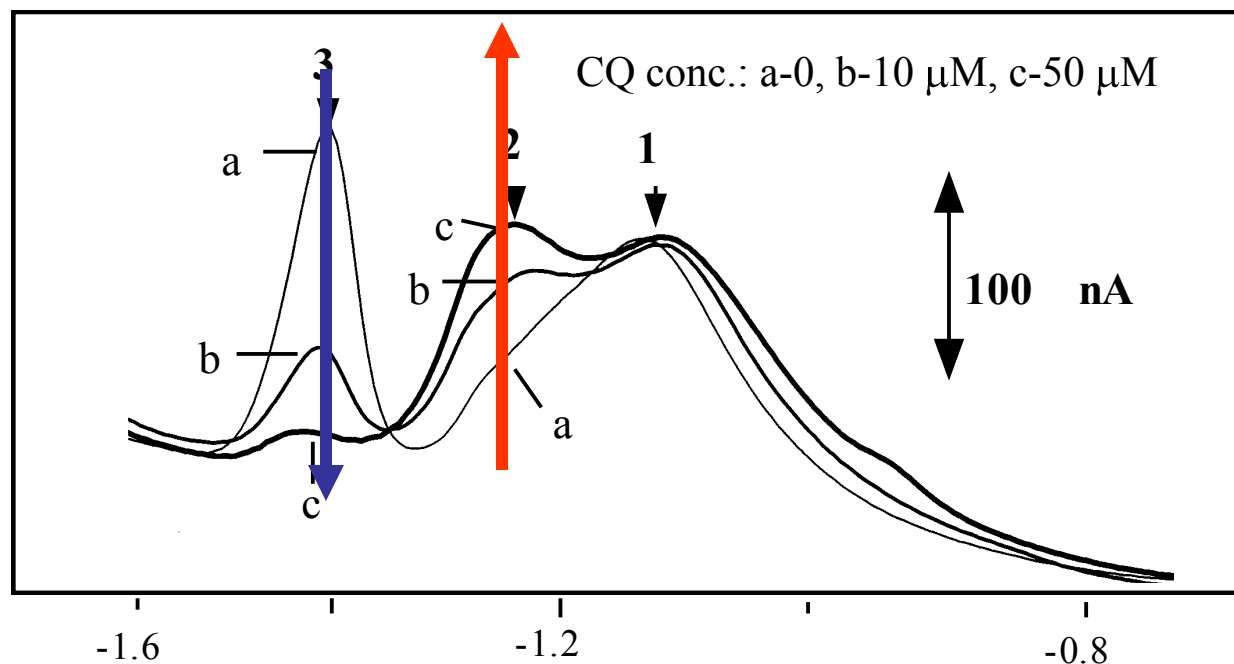
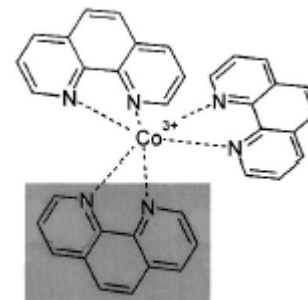
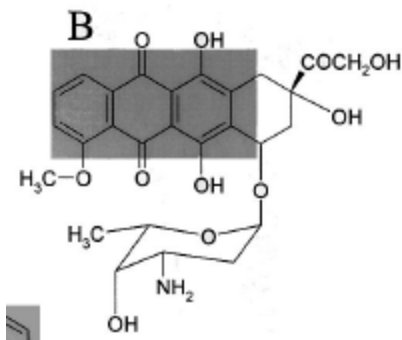
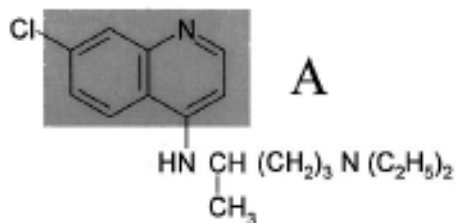
shorter DNA fragments can fit better the surface shape



DNA structural changes due
to intercalation

Adsorptive Transfer Stripping AC Voltammetry of DNA Complexes with Intercalators

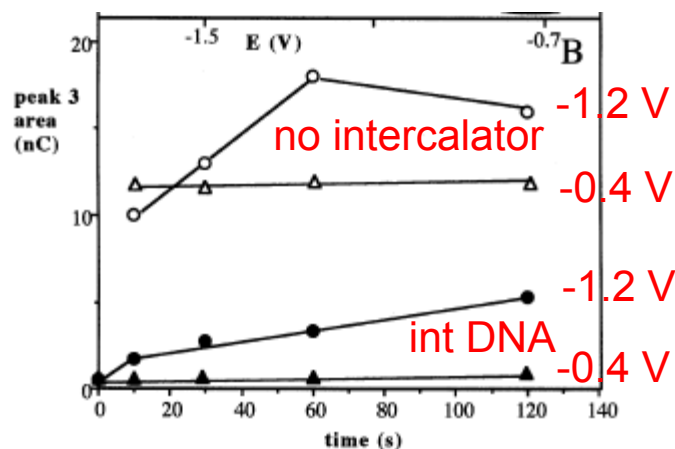
Miroslav Fojta,* Luděk Havran, Jana Fulnečková, and Tatiana Kubičárová



I

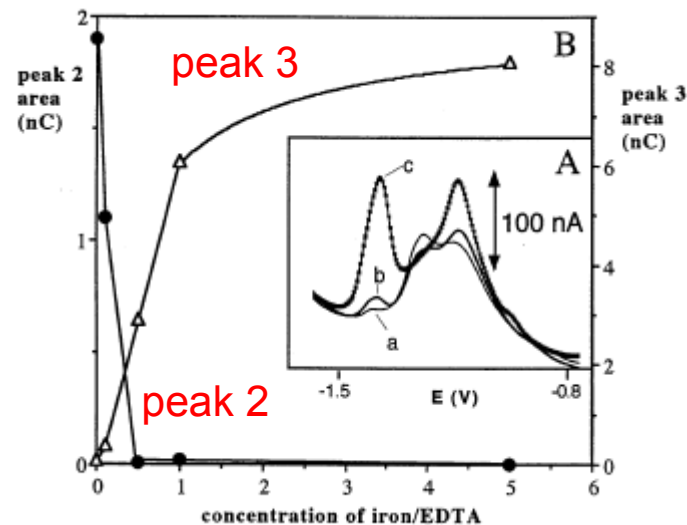
Adsorptive Transfer Stripping AC Voltammetry of DNA Complexes with Intercalators

Miroslav Fojta,* Luděk Havran, Jana Fulnečková, and Tatiana Kubičárová



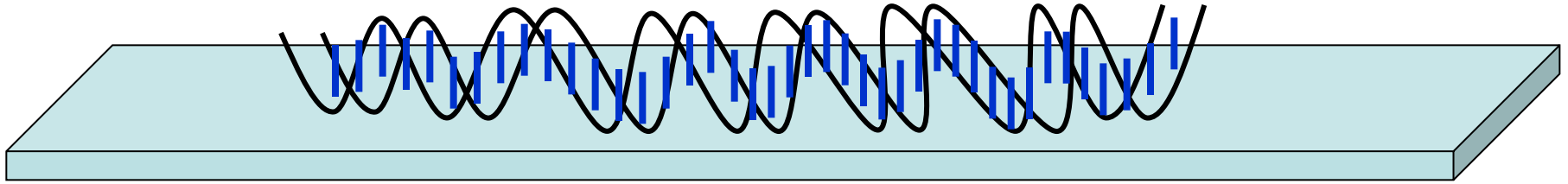
time of exposure to the potential

intDNA is more resistant to surface denaturation within the region U

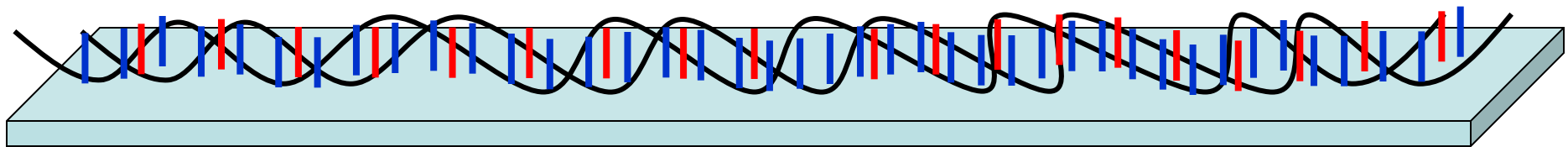


strand breaks abundance

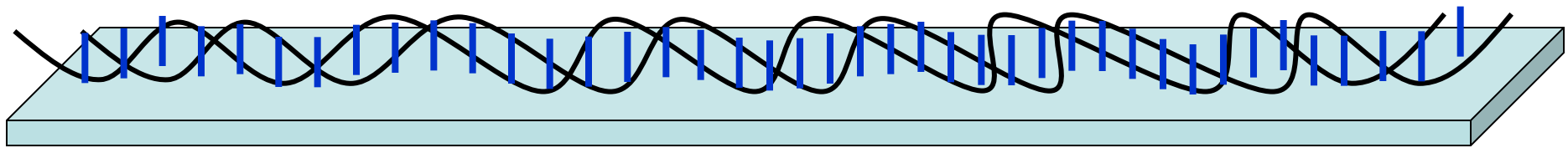
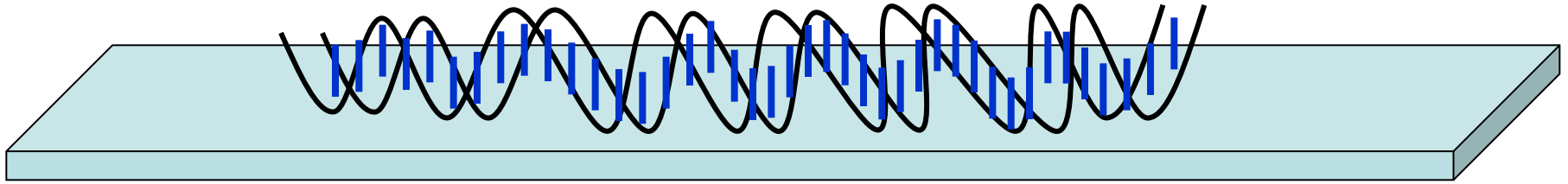
DNA containing numerous strand breaks, adsorbed in the presence of an intercalator, does not yield responses characteristic for intDNA



DNA in the absence of intercalator: B-form, bases hidden in the double-helix interior, relatively far from the electrode surface



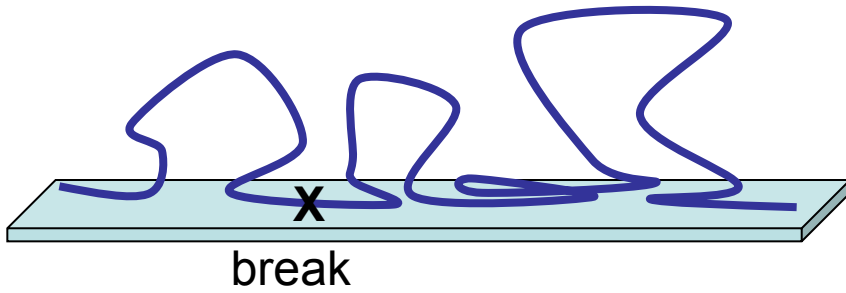
DNA saturated with the intercalator (**intDNA**): untwisted and lengthened double helix, less deep grooves – bases closer to the surface, contacts between the surface and the base pair edges



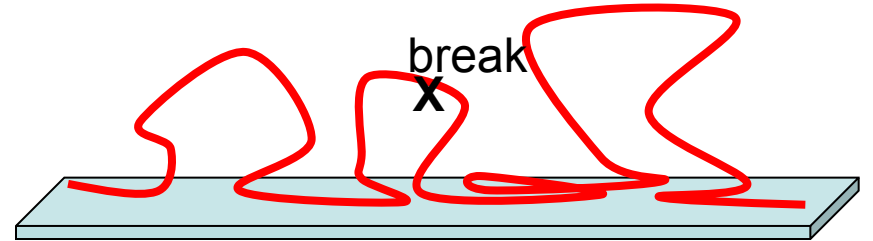
after removal of the intercalator, the intDNA conformation is preserved

the adsorbed untwisted regions of intDNA yield the AV voltammetric peak 2

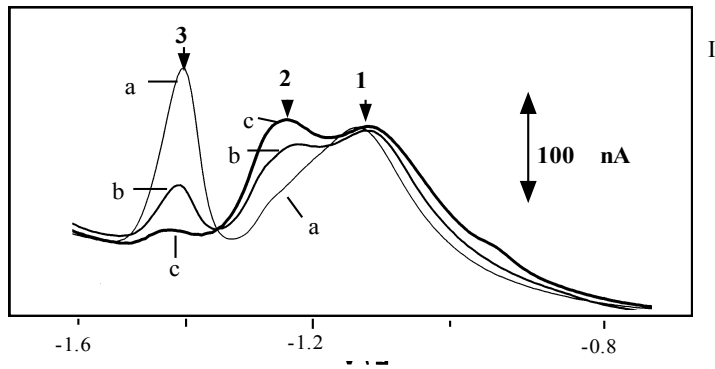
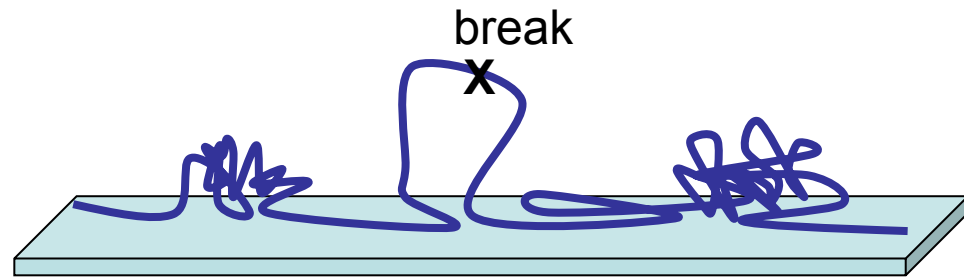
dsDNA without intercalator



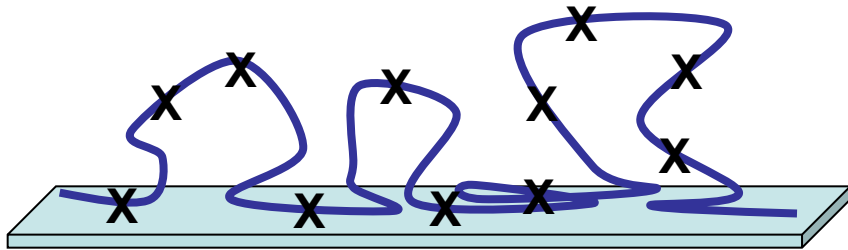
intDNA with intercalator



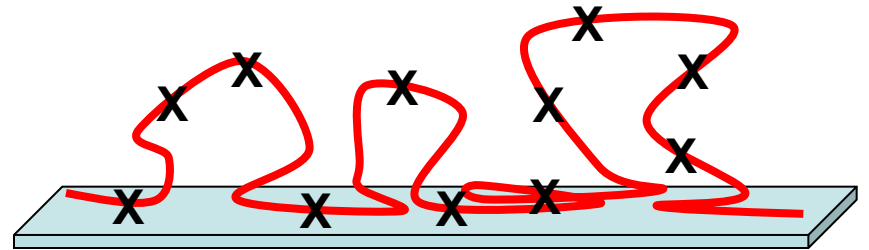
intDNA after intercalator removal



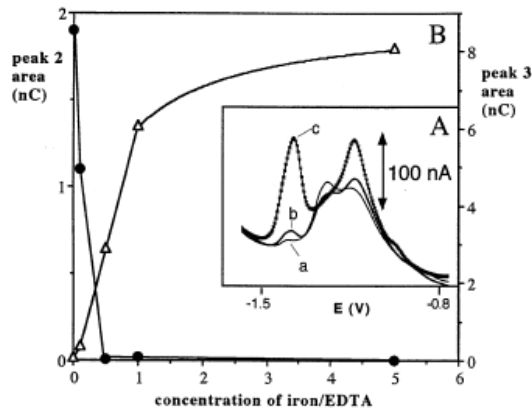
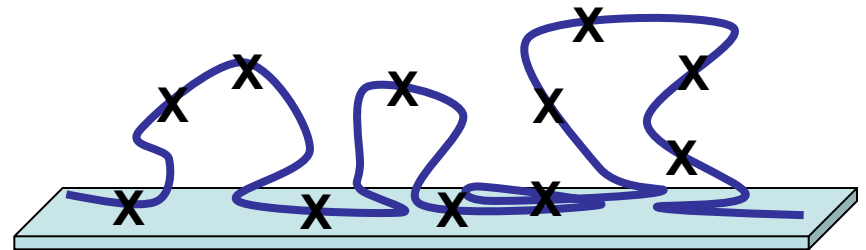
dsDNA without intercalator



with intercalator



intDNA after intercalator removal

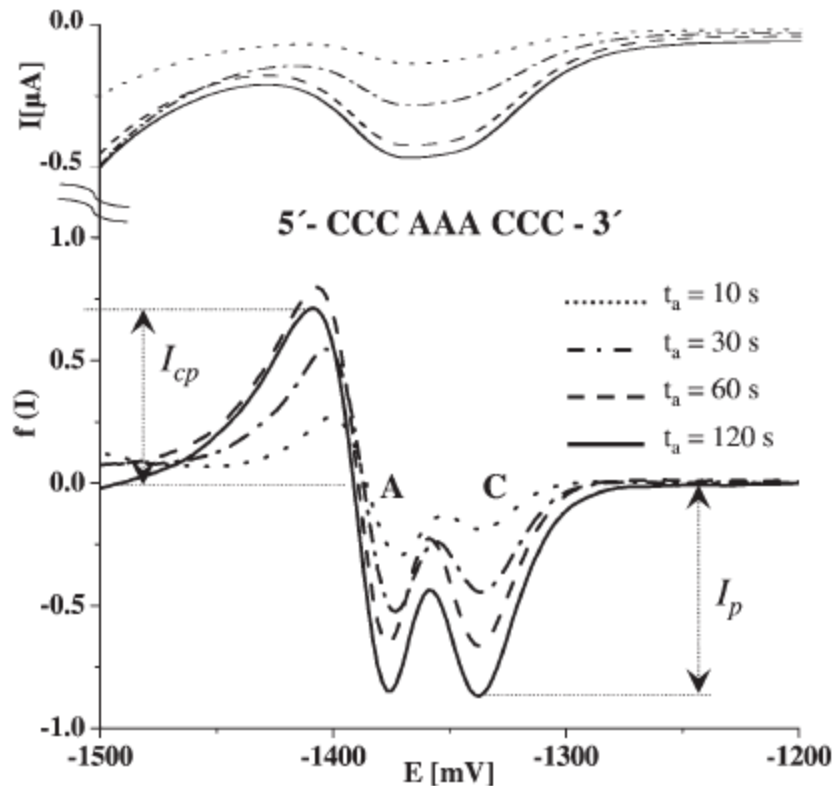


single-stranded oligonucleotides:

effects of nucleobase
composition and/or nucleotide
sequence

Resolution of Overlapped Reduction Signals in Short Hetero-oligonucleotides by Elimination Voltammetry

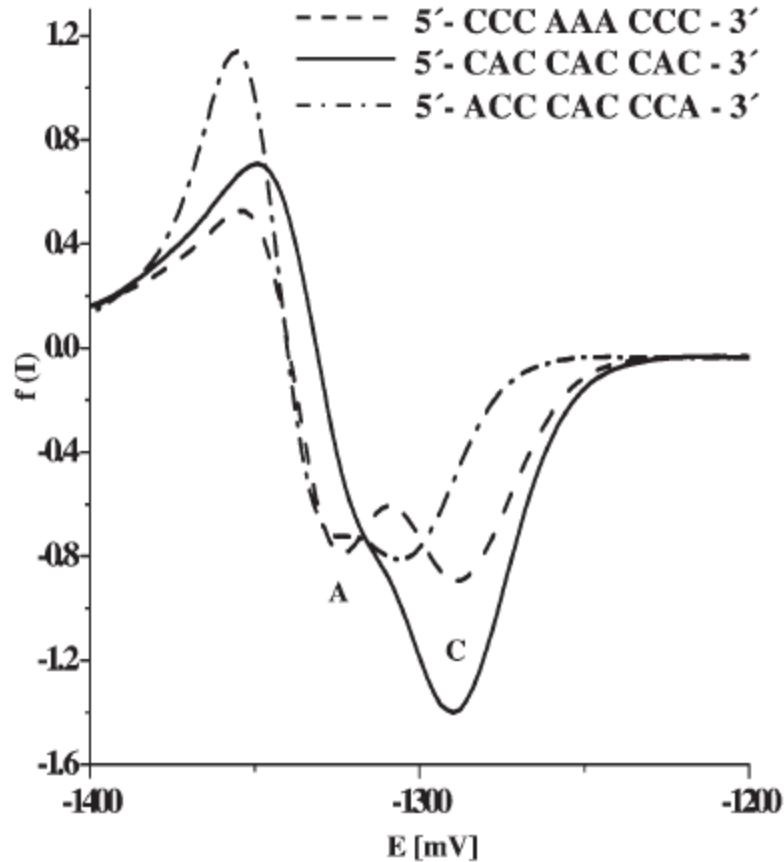
Radka Mikelova,^a Libuse Trnkova,^{a*} Frantisek Jelen,^b Vojtech Adam,^c Rene Kizek^c



- Dračka, Trnková
- current measured in voltammetry is a sum (linear combination) of partial current components (diffusion, capacitive, kinetic...)
- these components depend diversely on scan rate
- possibility of numerical elimination of any component based on measurements at several different scan rates
- studies of electrode processes
- separation of overlapped signals

Resolution of Overlapped Reduction Signals in Short Hetero-oligonucleotides by Elimination Voltammetry

Radka Mikelova,^a Libuse Trnkova,^{a*} Frantisek Jelen,^b Vojtech Adam,^c Rene Kizek^c



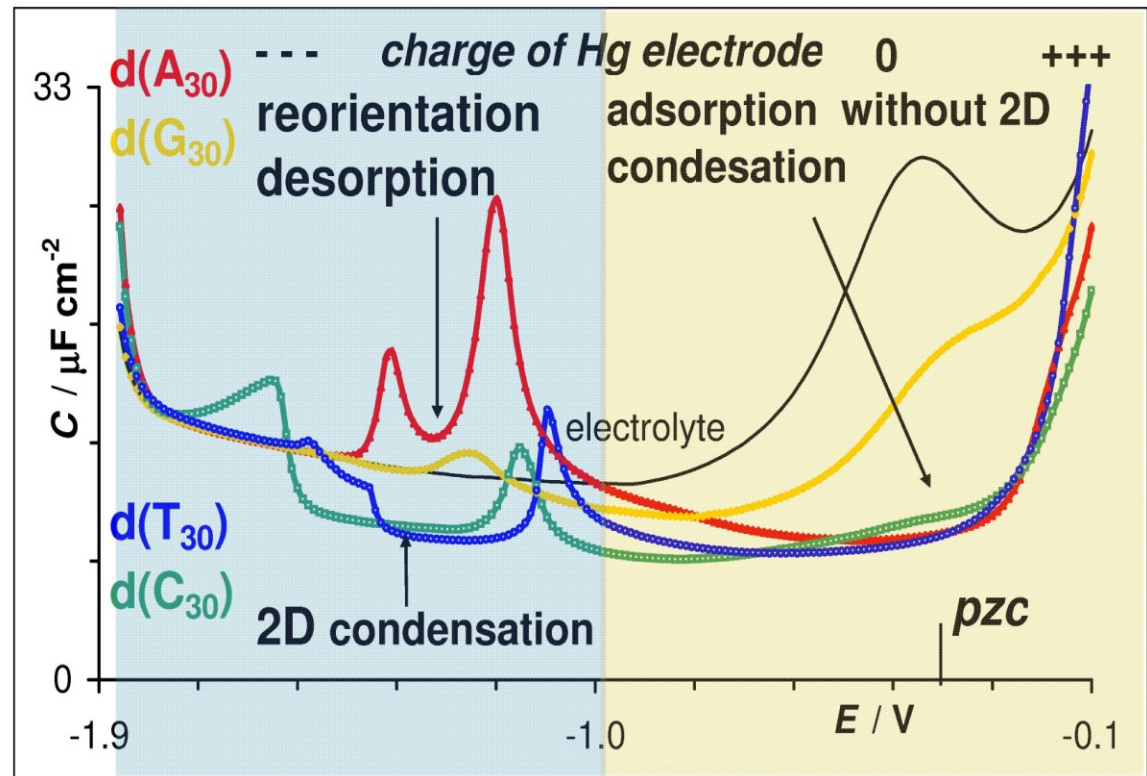
- different EVLS responses for ODNs with identical base content but differing in their sequence

2D condensation of homopyrimidine oligos at mercury-based electrodes

Two-Dimensional Condensation of Pyrimidine Oligonucleotides during

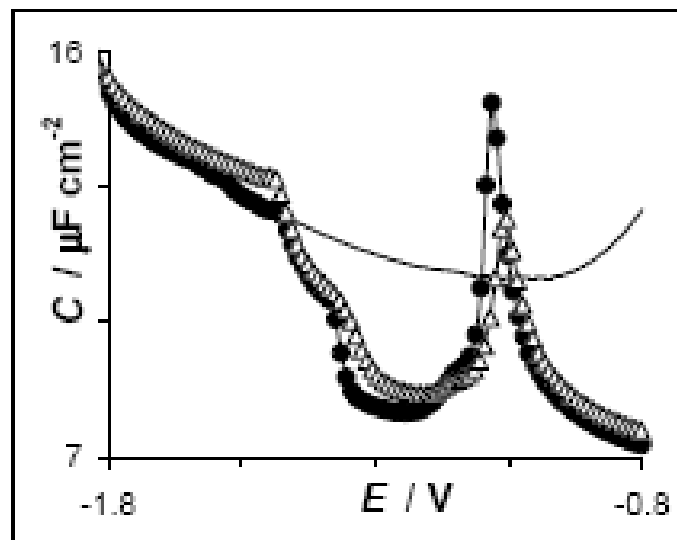
Their Self-Assemblies at Mercury Based Surfaces

Stanislav Hason*, Vladimír Vetterl¹, Miroslav Fojta*

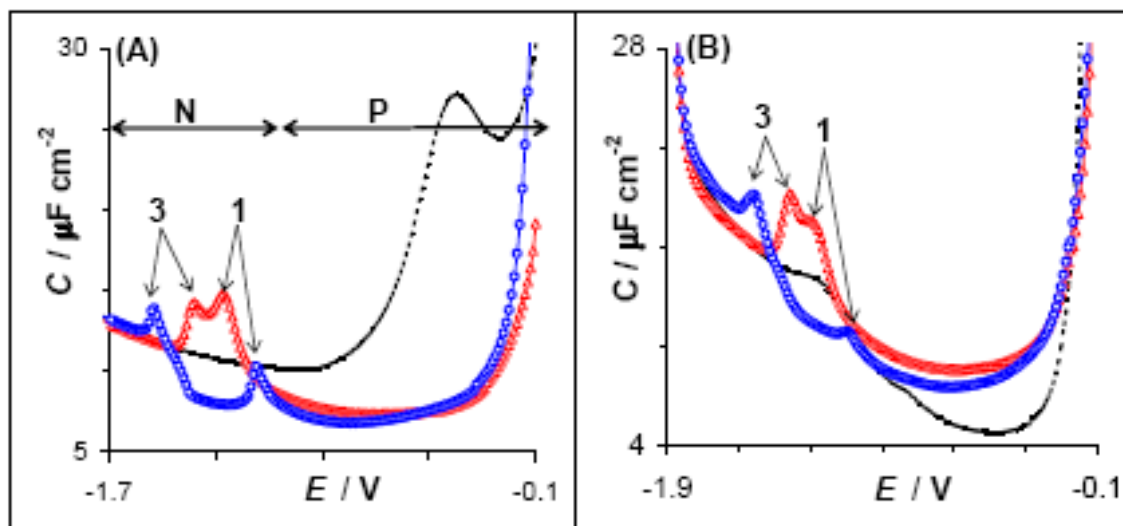


- capacitance pits indicate formation of condensed films
- NA bases, nucleosides, nucleotides for decades known to form such 2D-condensed layers (V. Vetterl)
- up to recently, no observations with DNA, polynucleotides or ODNs
- S. Hason: homopyrimidine ODNs (30 to 90-mers studied) can form such condensed films at negatively charged mercury or amalgam surfaces

transfer (ex-situ) experiment:
the condensed film is formed of reoriented ODN molecules already adsorbed (not due thickening the adsorbed layer by extra molecules form the bulk)



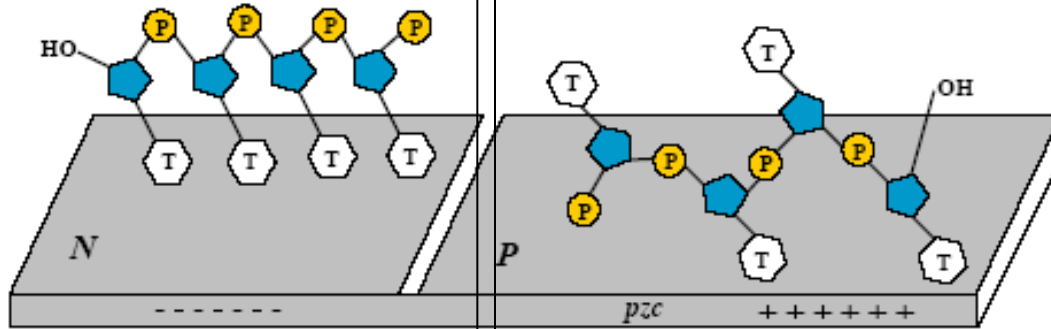
mixed **A+G** or **C+T** ODNs



HMDE

silver amalgam electrode

around pzc: random orientation of the ODN chain



negatively charged surface:
sugar-phosphate backbone
repelled
pyrimidine bases undergo self-assembly

these phenomena may affect
behavior of ODNs anchored
via terminal thiol group

