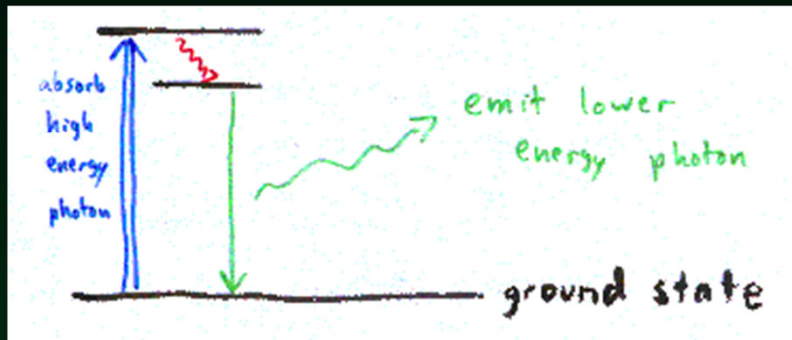


# Cellular Communication

Genomics Lectures

Kamil Růžička  
FGP CEITEC MU

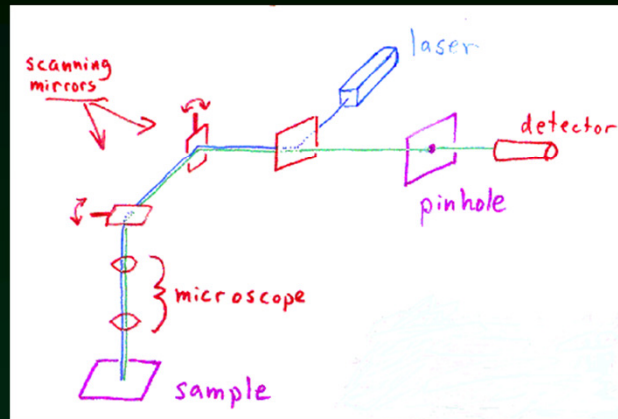
# How does fluorescence work?



## How does a fluorescence microscope work?



# How does a confocal microscope work?





# Protein localization

live imaging

GFP discovery - Nobel Prize 2008



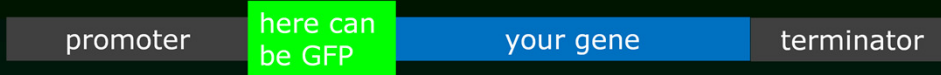
Osamu Shimomura

Martin Chalfie

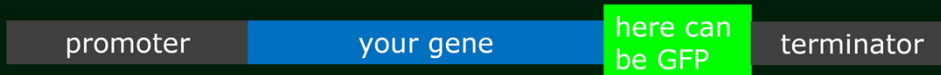
Roger Tsien

# GFP fusions

N-terminal fusion



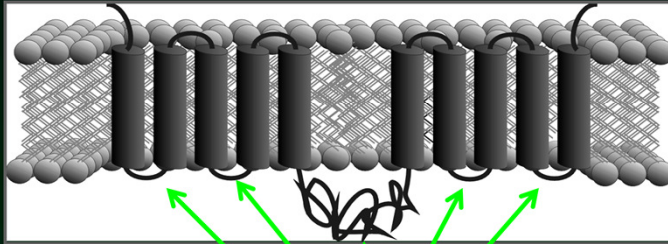
C-terminal fusion



fusion inside the coding sequence



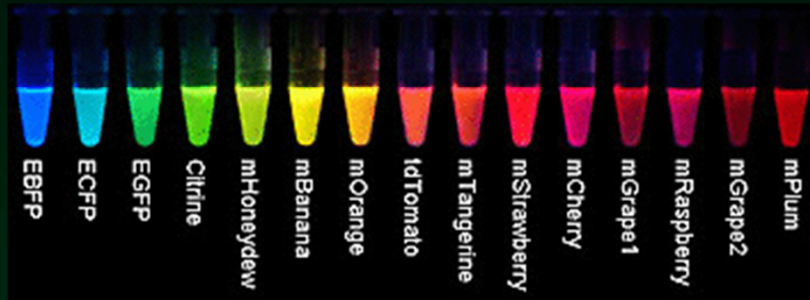
# GFP and membrane proteins



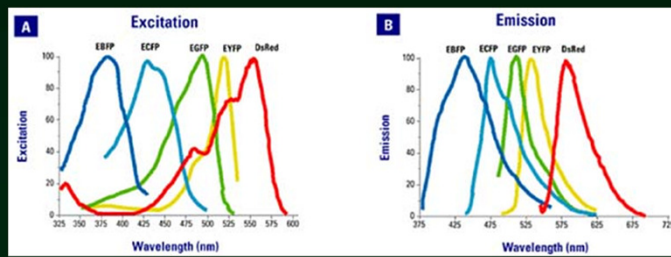
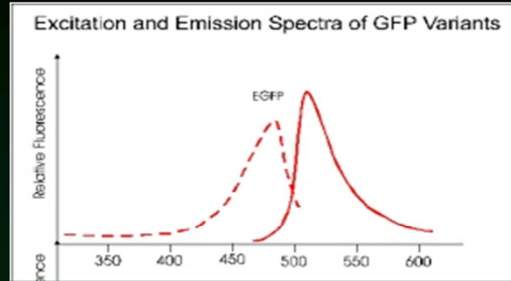
It is good to have GFP tag localized inside the cell

here can be GFP

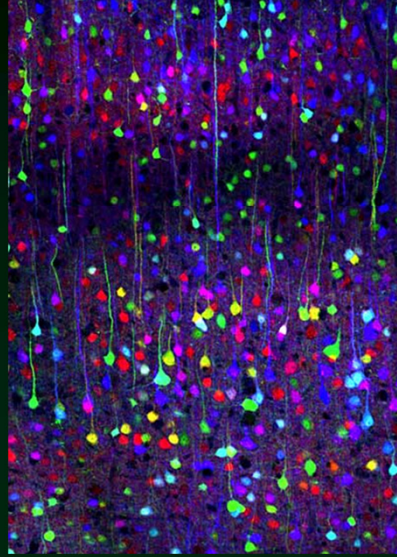
# Fluorescent proteins on the market



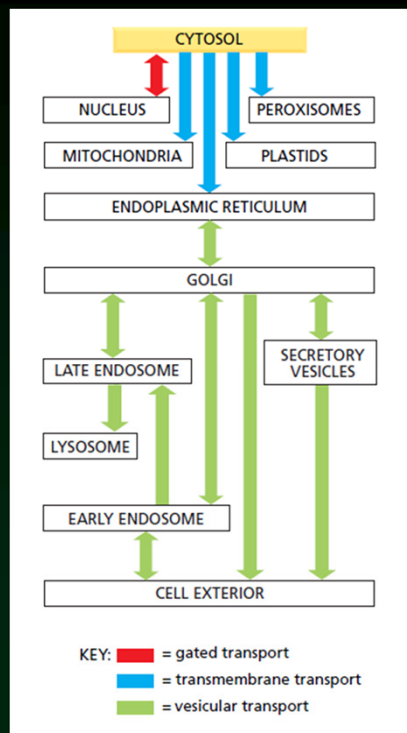
# Excitation and emission



## Multicolored fluorescent protein (neurones)

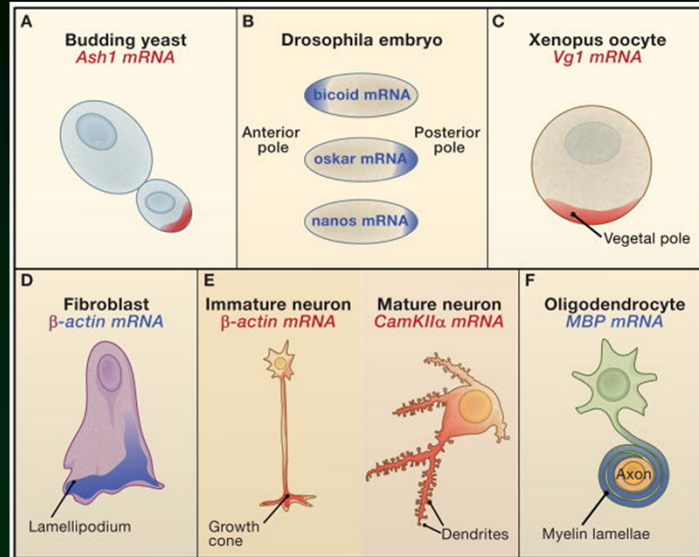


# Transport among compartments



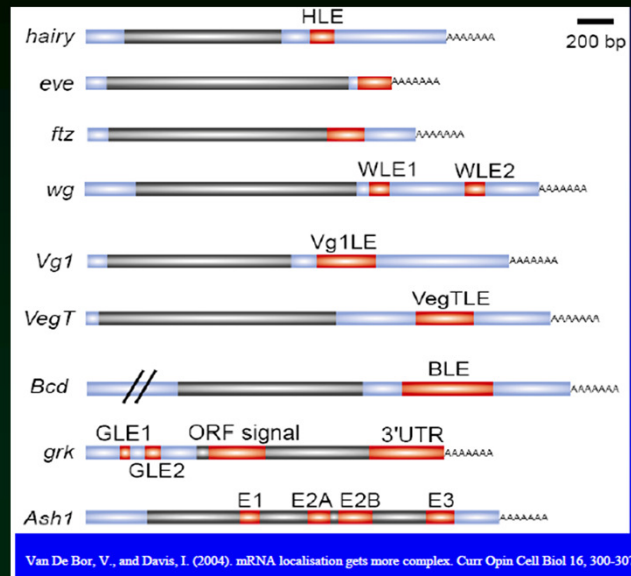
Alberts et al. 2008

# Also RNA can be differentially localized





# RNA ZIP codes for localization



Mikko Frilander

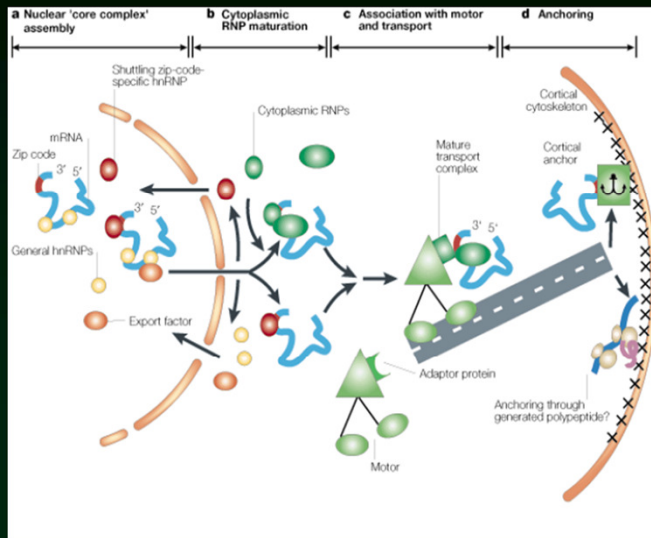
# ZIP codes often motor protein bound

Table 1 | **Cross-species comparison of proteins involved in cytoplasmic mRNA localization**

Function/feature	Yeast	<i>Drosophila melanogaster</i>	Vertebrates
Zip-code-binding hnRNP protein (located in nucleus and cytoplasm)	Not yet identified	Squid ( <i>grk</i> ) <sup>20</sup>	hnRNP A2 ( <i>MBP</i> ) <sup>21</sup> VgRBP60* ( <i>Vg1</i> ) <sup>62</sup> Vera/VgRBP* ( <i>Vg1</i> ) <sup>39,63</sup> ZBP-1* ( <i>β-actin</i> ) <sup>22</sup>
Cytoplasmic zip-code-binding RNP	She2 ( <i>ASH1, IST2</i> ) <sup>6,63,64</sup>	Staufen ( <i>osk, pros</i> ) <sup>65</sup> Swallow* ( <i>bcd</i> ) <sup>82</sup> Ypsilon-Schachtel* ( <i>osk</i> ) <sup>83</sup>	mStaufen* (?) <sup>68,79,80</sup> VILIP* ( <i>trk</i> ) <sup>69</sup> TB-RBP ( <i>CaMKII</i> ) <sup>98,99</sup>
Motor protein for RNP	Myo4 ( <i>ASH1, IST2</i> ) <sup>4,6</sup>	Kinesin I ( <i>osk</i> ) <sup>88</sup> Dynein* ( <i>bcd</i> ) <sup>82</sup>	Kinesin* ( <i>MBP, CaMKII</i> ) <sup>75,99</sup>
RNP motor adaptor	She3 ( <i>ASH1, IST2</i> ) <sup>6,63</sup>	Dynein light chain ( <i>bcd</i> ) <sup>82</sup>	Not yet identified
mRNA/RNP anchor	Bni1*, Bud6* ( <i>ASH1</i> ) <sup>32</sup>	Staufen ( <i>bcd</i> ) <sup>20</sup> Oskar ( <i>osk</i> ) <sup>37</sup>	<i>Xlsit</i> mRNAs* ( <i>Vg1</i> ) <sup>38</sup>

Mikko Frilander

# Delivering at the correct address



**1. Localisation complex assembly starts already in the nucleus**

**2. Cytoplasmic mRNP is "matured", nuclear export proteins removed, additional proteins attached.**

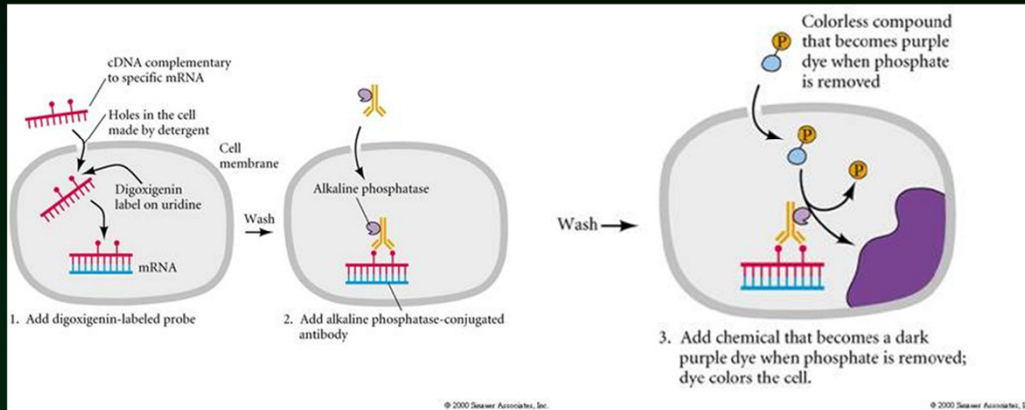
**3. mRNP is associated with motor and transport system and delivered to the destination**

**4. Delivered mRNP is anchored to the destination (for storage) or translated directly.**

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# Localization of mRNA

## RNA hybridization *in situ*



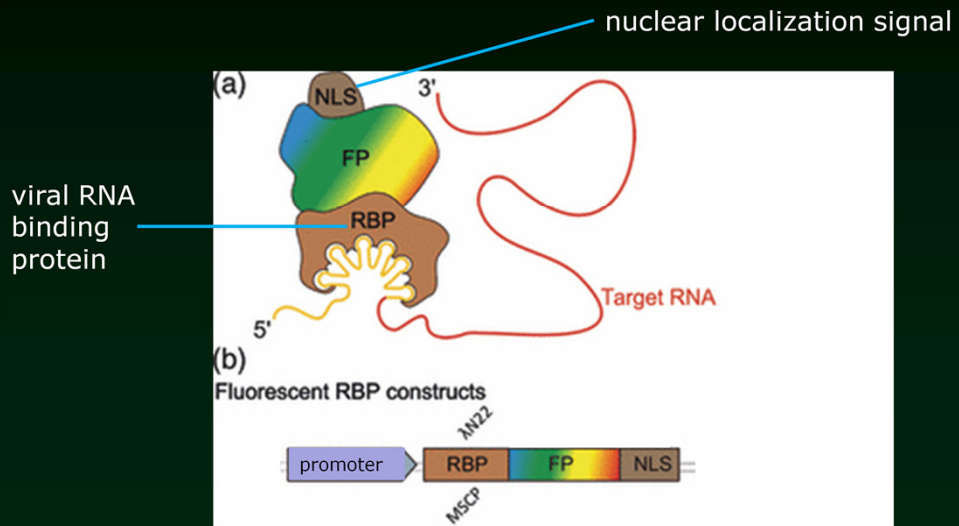
# Localization of mRNA

## RNA hybridization *in situ*

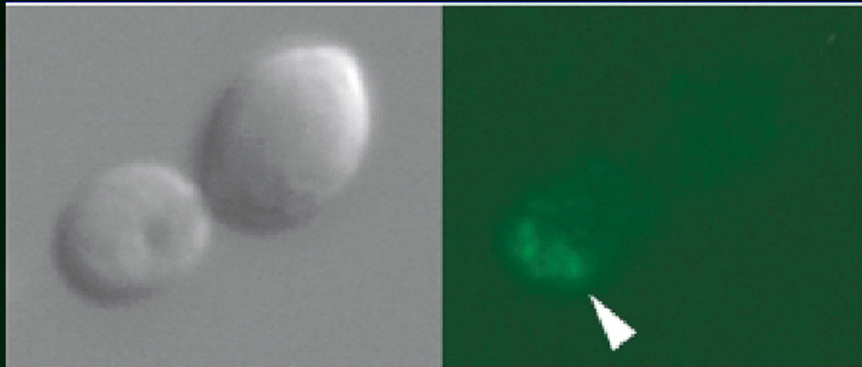
- classical technique, no alternative in developmental biology
- results often clear
- can be done without generating transgenic lines
- tedious
- only on "dead" samples



# $\lambda N_{22}$ system

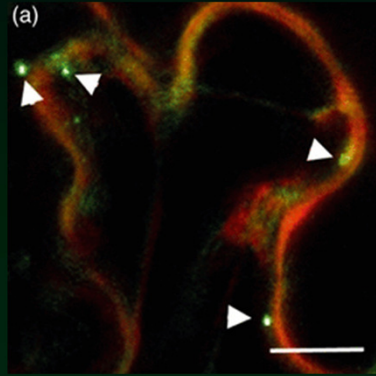


## Also mRNA can be differentially localized



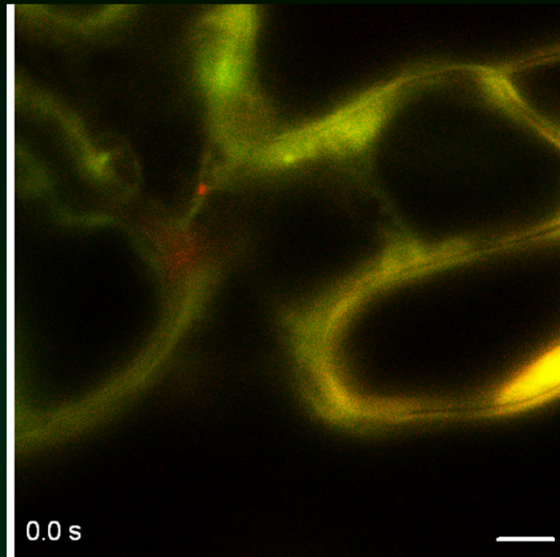
Ash1 mRNA localized to the tip of the daughter cell

Also mRNA can be differentially localized



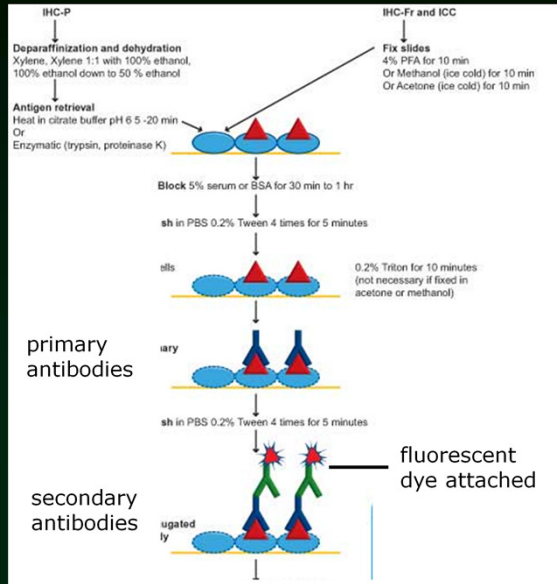
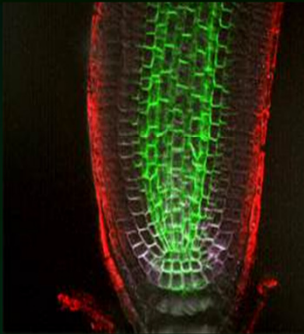


Also mRNA can be differentially localized



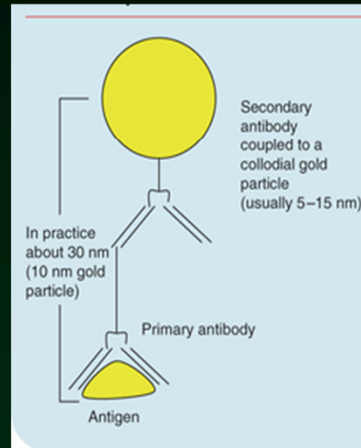
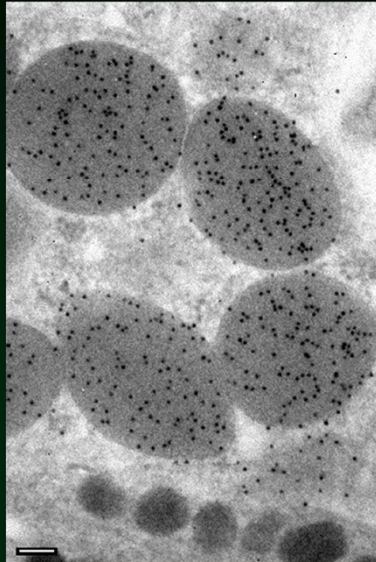
# Protein localization

immunolocalization - fluorescently



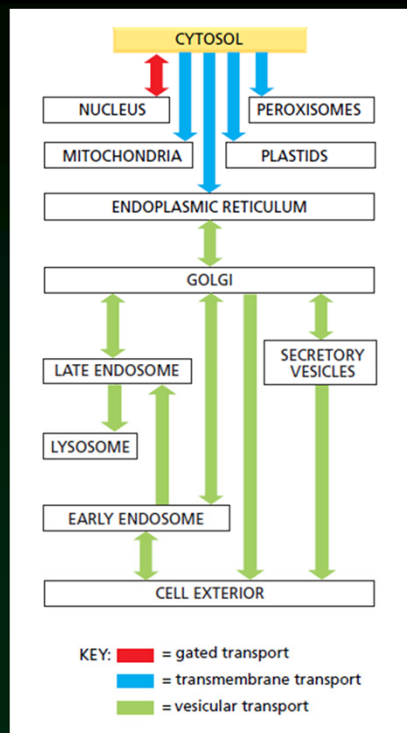
# Protein localization

immunolocalization - immunogold



electron microscope

# Transport among compartments

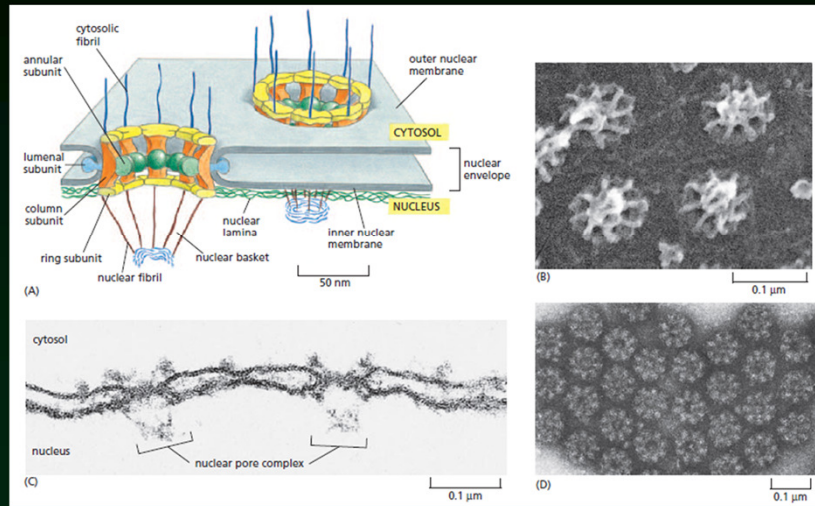


Alberts et al. 2008

# Protein sorting – target peptides

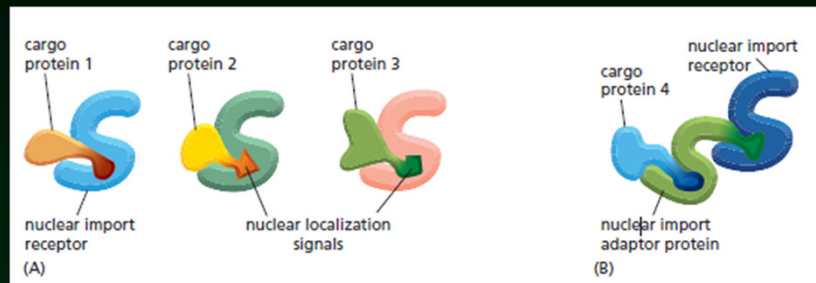
Location	Type of targeting signal	Properties
Nucleus	Nuclear localization signal (NLS)	Short clusters of basic amino acids
Endoplasmic reticulum	Signal peptide	Cleavable N-terminal presequence
Plastid	ER retention signal	C-termini, H/KDEL motif
	Transit peptide	Usually cleavable N-terminal presequence
Mitochondrion	Presequence/Transit peptide	Usually cleavable N-terminal presequence
Peroxisome	Peroxisome targeting sequence 1 (PTS1)	C-termini, a conserved short motif
	Peroxisome targeting sequence 2 (PTS2)	Cleavable N-terminal presequence
Tonoplast/vacuole	Signal peptide	Cleavable N-terminal presequence
	Vacuolar sorting signals	Internal short sequence at near N-terminal C-termini, targeting to protein storage vacuole
Apoplast	Signal peptide	Cleavable N-terminal presequence

# Nuclear transport

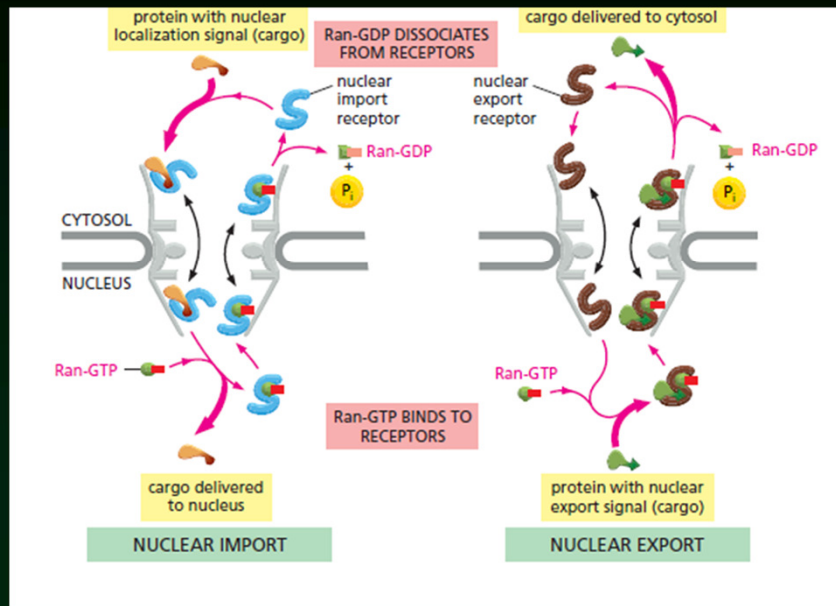


nucleoporins

# Nuclear import

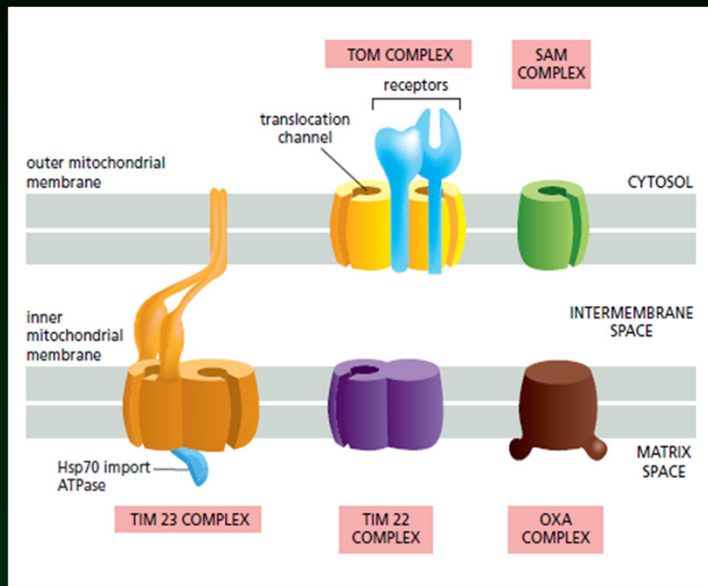


# Nuclear import





# Mitochondrial transport



TIM and TOM complexes decide at which side of mitochondria the protein will be transported.

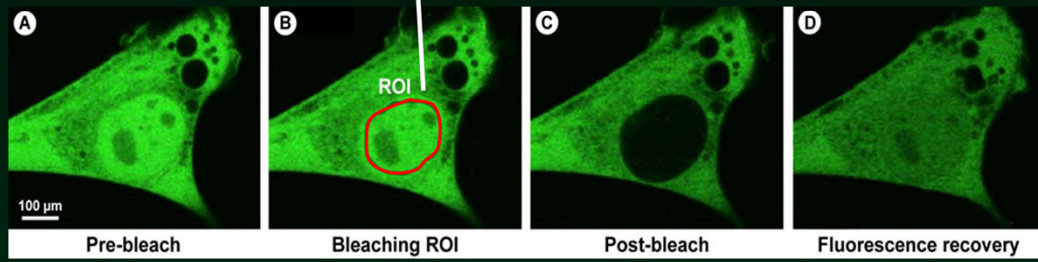
## Advanced confocal techniques

- FRAP
- photoactivatable FP
- FCS

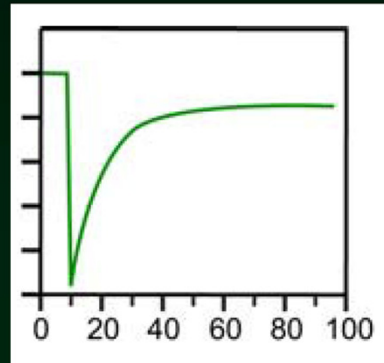
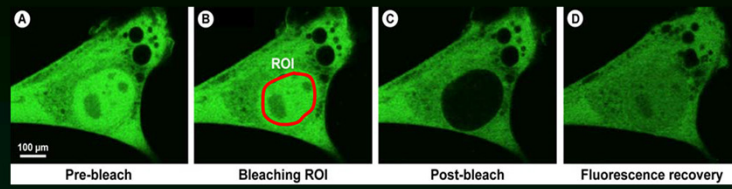
# FRAP

## Fluorescence Recovery After Photobleaching

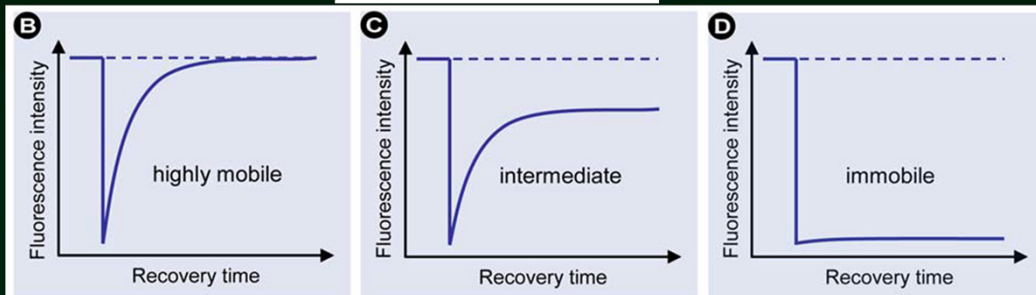
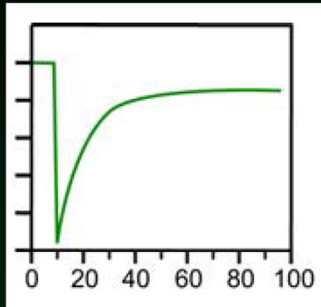
region of interest



# FRAP

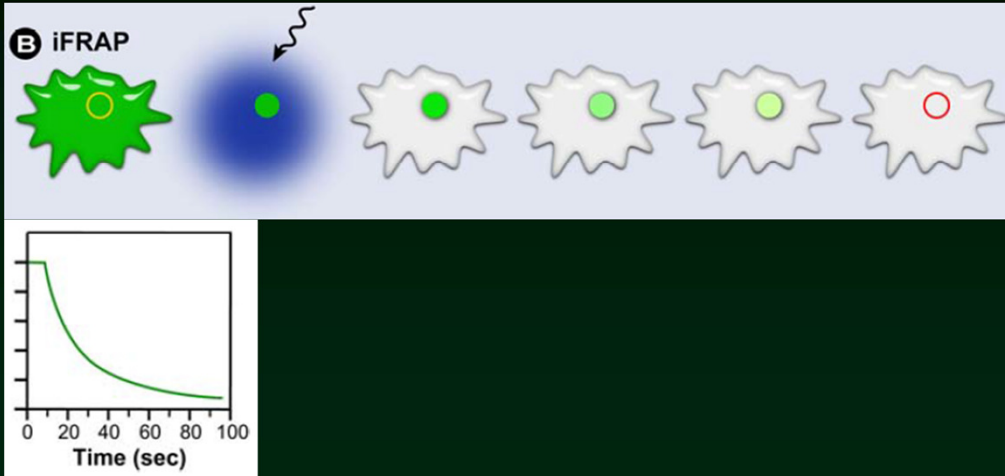


# FRAP – bleaching curve

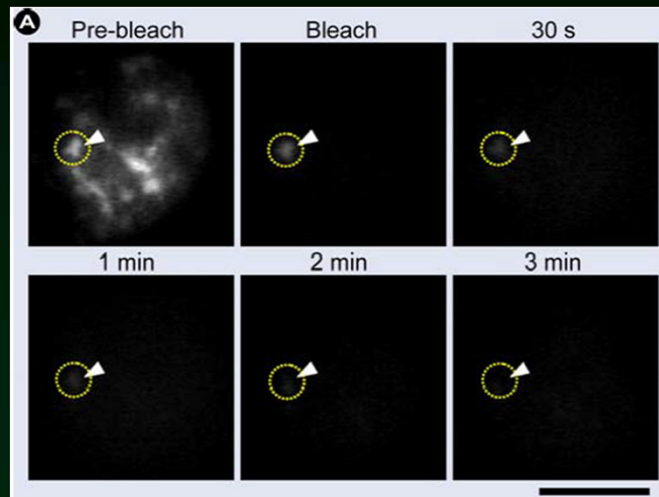


# iFRAP

inverse FRAP



# iFRAP – dissociation of premRNA from species



## FRAP - advantages

- not only proteins (also other dyes)



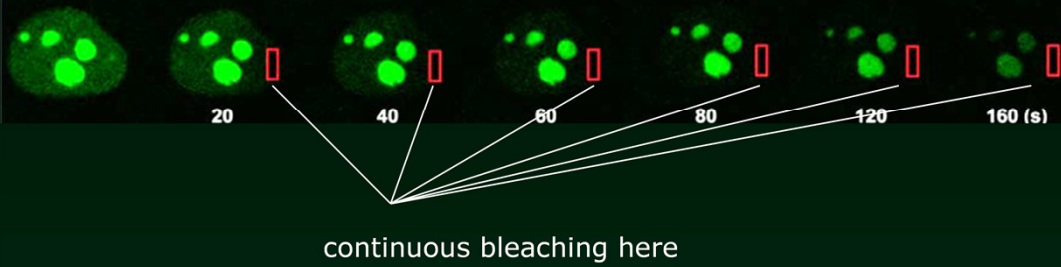
## FRAP – disadvantages

- your cells are moving
- high energy needed to bleach the ROI
  - can damage your material
  - long time needed to bleach
- usually only one ROI can be observed – time consuming

# FRAP derivatives

## FLIP

Fluorescence Loss After Photobleaching

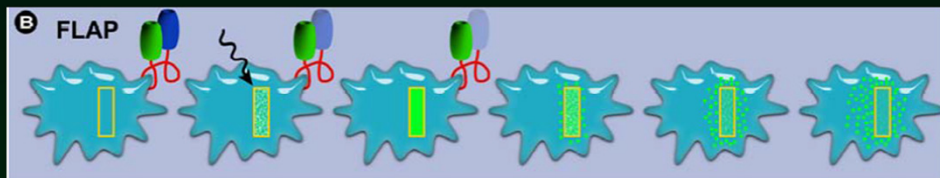


- bleaching process is repeated during the experiment
- for studying general protein turnovers in compartments
- less often used

# FRAP derivatives

## FLAP

Fluorescence Localization after Photobleaching

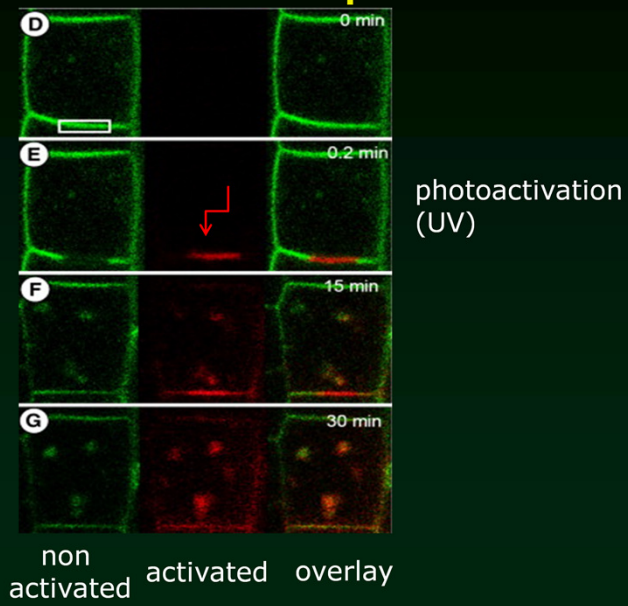


- two fluorochromes on one protein– one bleached, non bleached as control

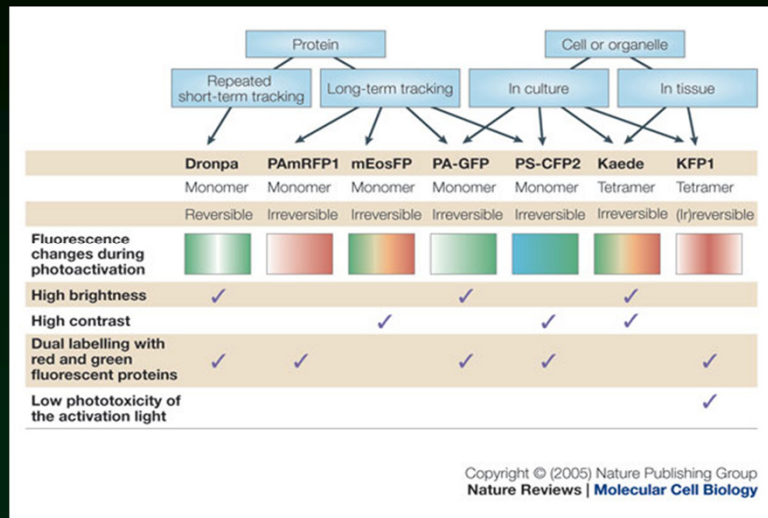
## Intermezzo: story from a conference

even top scientists can be wrong

# Photoactivatable proteins



# Photoactivable proteins



Dronpa, Kaede, Eos – probably most popular

# Photoactivable proteins

## Advantages:

- most elegant, most convincing

## Disadvantages:

- very weak signal
- each material needs optimization

## Remarks

- your material is 3D
- protein *de novo* synthesis in some experiments (e.g. cycloheximide stops translation)



# FLIM

Fluorescence Life Time Imaging Microscopy

## Fluorochromes

- excitation spectra
- emission spectra
- **unique lifetime**

Lifetime sensitive to almost everything:

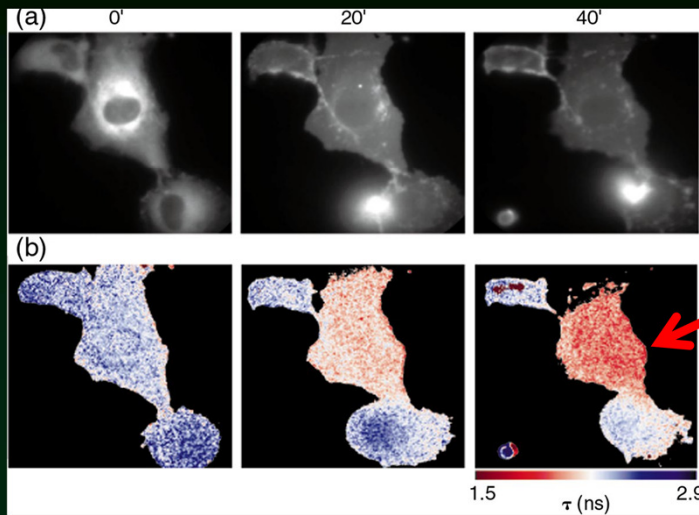
- pH
- ionic strength
- polarity
- other fluorochrome

# FLIM - applications

Protein-protein interactions  
(FRET-FLIM) (other lecture)

# FLIM - applications

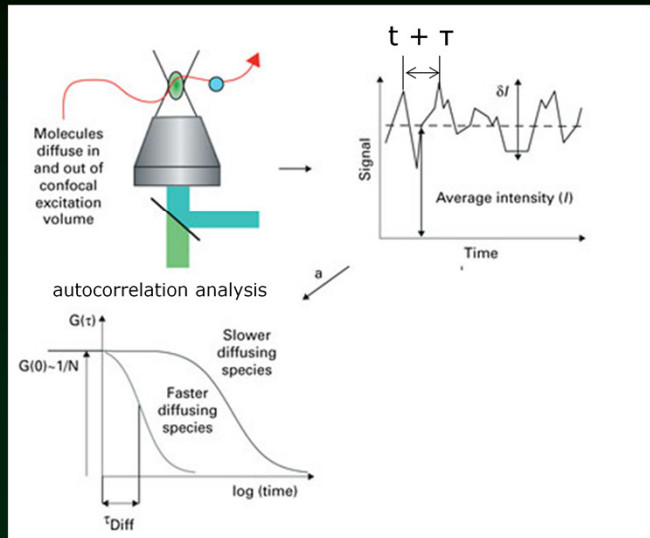
phosphorylation assay



lifetime decreased by  
site specific IgG  
injection

# FCS

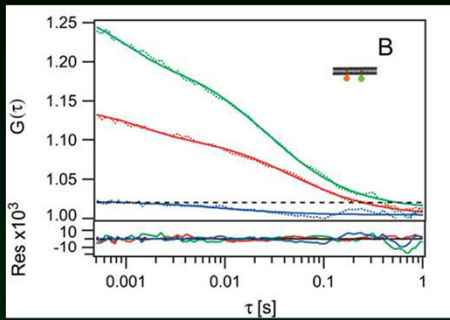
## Fluorescence Correlation Spectroscopy



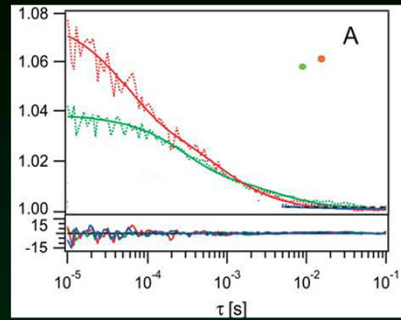
It is counted, how many times the fluorescent molecule comes through the focal plane.

Autocorrelation analysis: the way how to discriminate the diffusions speeds of particles.

# FCS

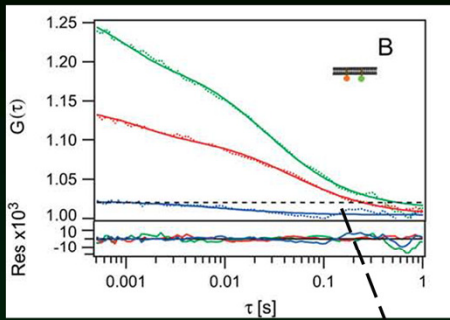


control membrane bound  
GFP and RFP  
(crosscorrelation curve)



free GFP and RFP

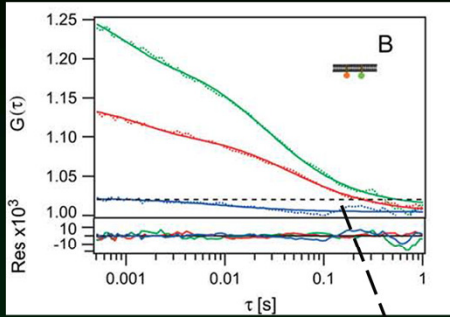
# FCS



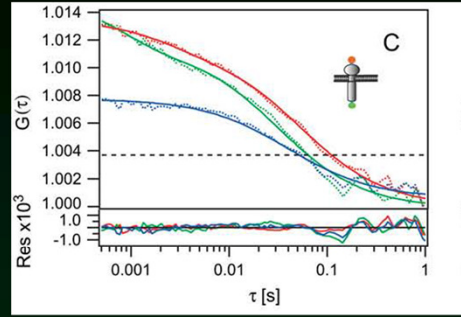
control membrane bound  
GFP and RFP  
(crosscorrelation curve)

channel crosstalk threshold

# FCS



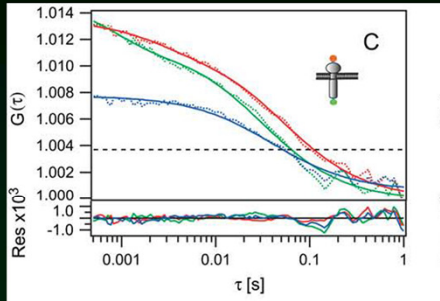
control membrane bound  
GFP and RFP  
(crosscorrelation curve)



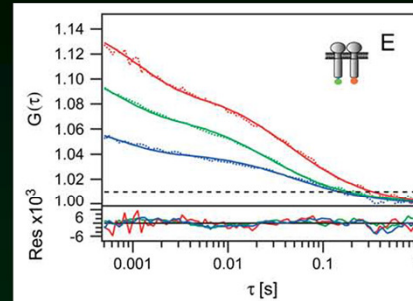
receptor with two labels

channel crosstalk threshold

# FCS



receptor with two labels



the crosscorrelation curve is above threshold -> EGFR protein dimerizes

Liu et al., 2007