



Cytologie II.



Buněčné inkluze, chromoplasty

begónie (kysala) královská (*Begonia rex*) č. *Begoniaceae* - kysalovité

- příčný řez řapíkem listu; **drůzy** - složené krystaly štavelanu vápenatého víceméně kulovitěho tvaru, z jejichž povrchu vyčnívají jednotlivé krystaly





Begonia rex, obj. 40×



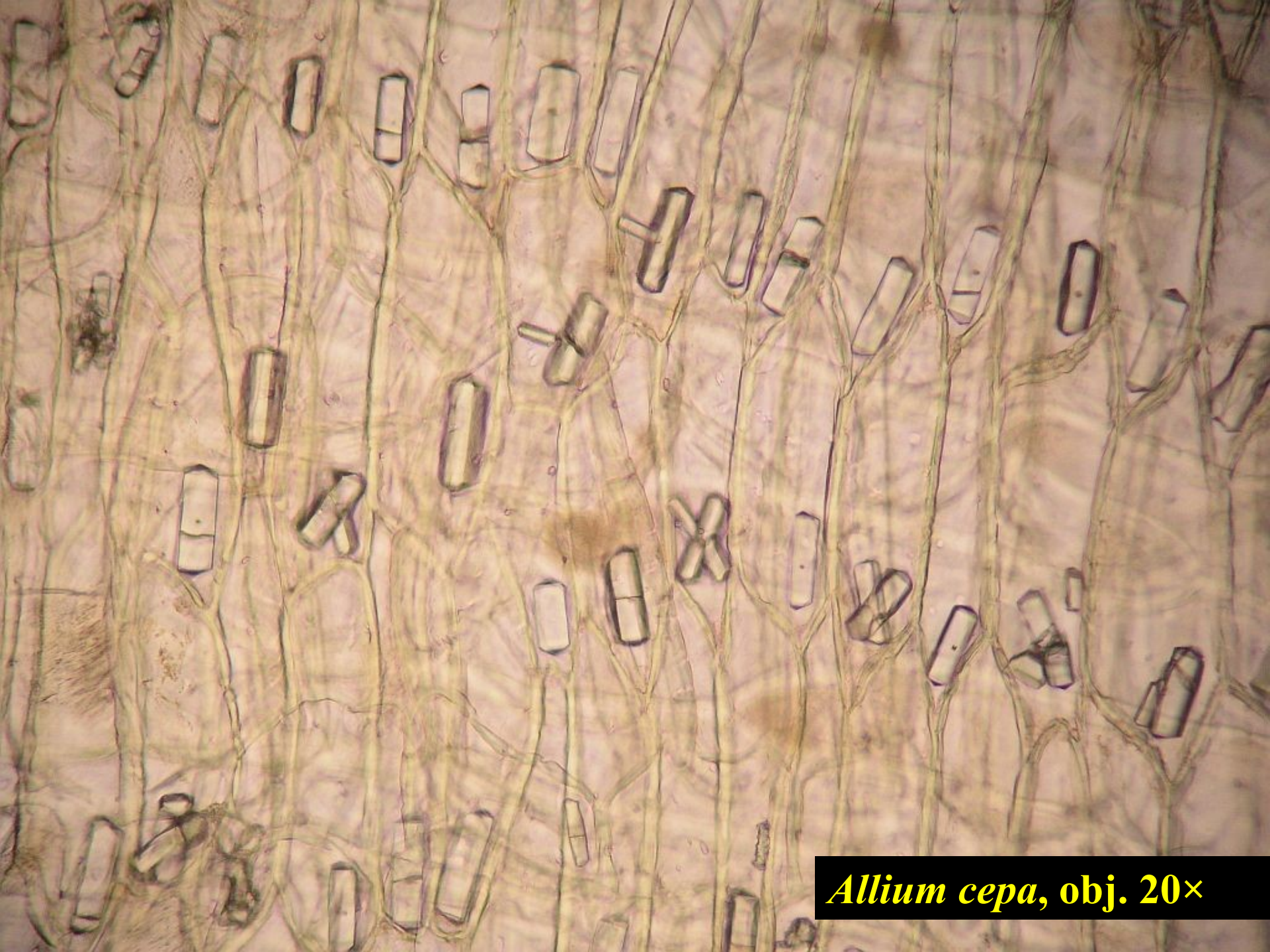
Begonia rex, obj. 40×



Begonia rex, obj. 40×

česnek cibule (*Allium cepa*) č. *Alliaceae* - česnekovité

- epidermis ze zevní strany vnějších vrstev suknic cibule
- několik dní v 70% et-OH; **styloidy**; šťavelan vápenatý

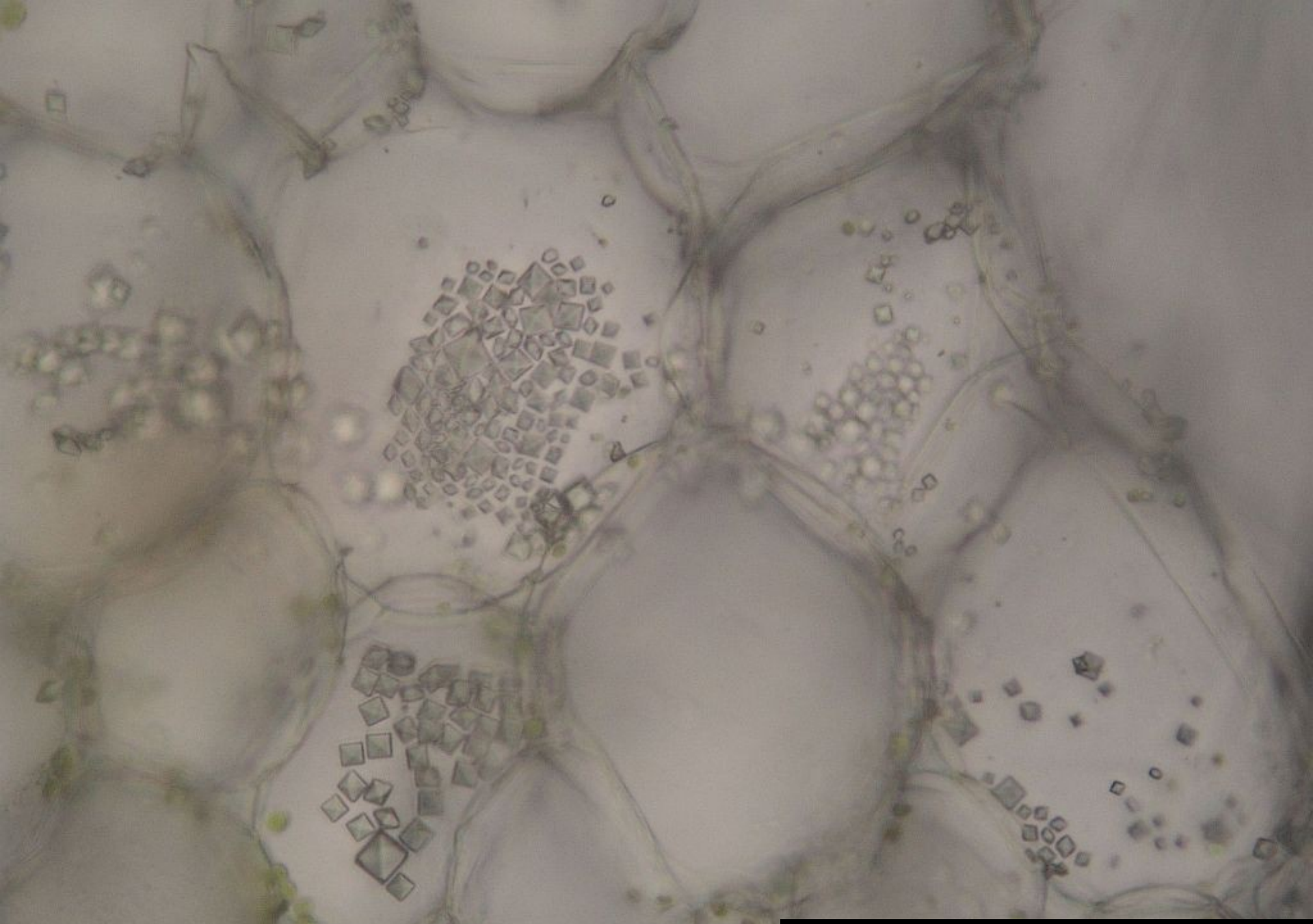


Allium cepa, obj. 20×

tradeskancie (voděnka, podénka, „blázen“) (*Tradescantia* sp.) č. Commelinaceae - křížatkovité

- příčný řez stonkem
- šťavelan vápenatý; **rafidy**





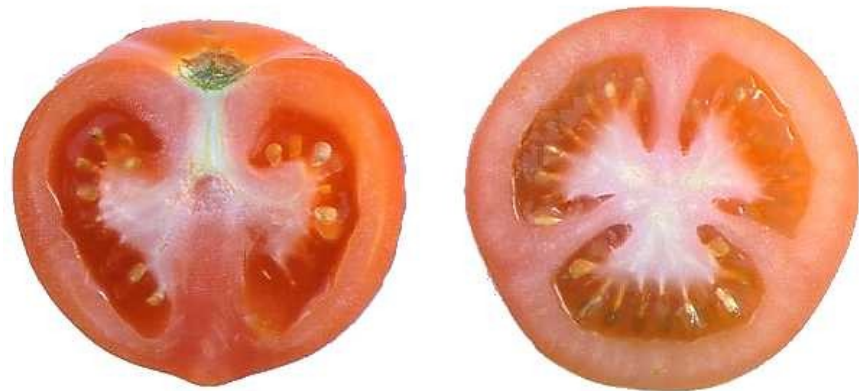
Tradescantia sp., obj. 40×

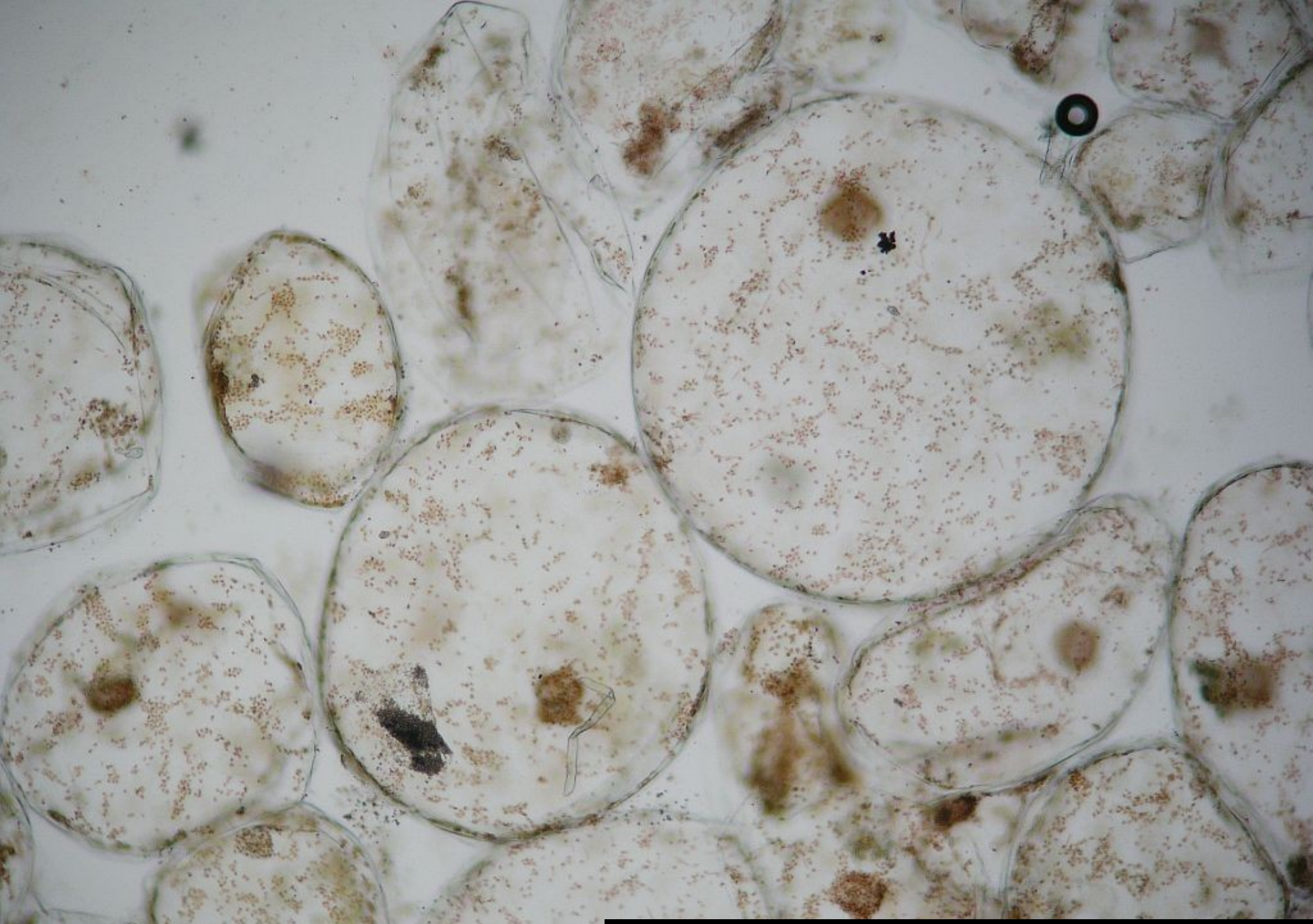


Tradescantia sp., obj. 40×

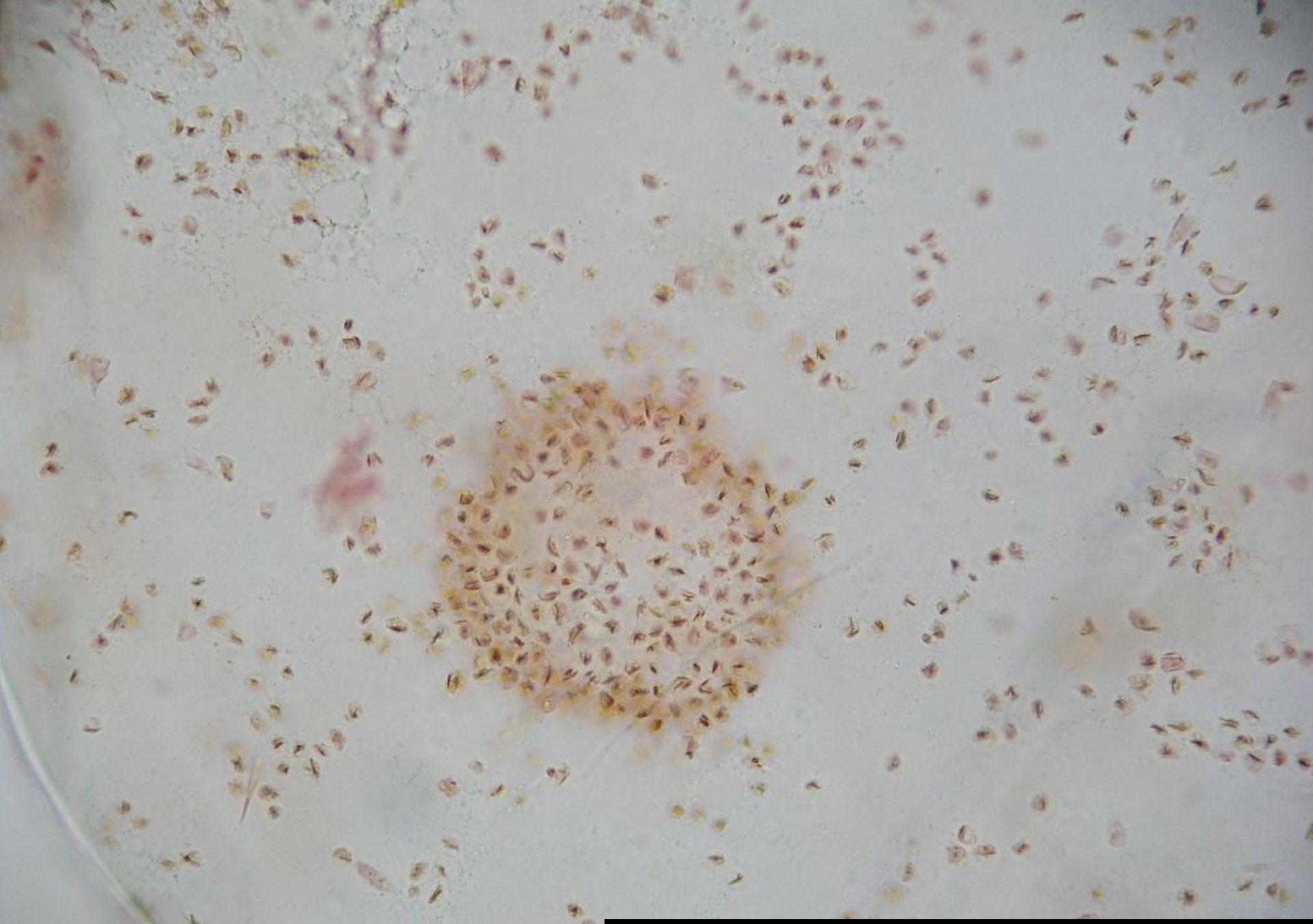
lilek rajče (*Lycopersicon esculentum*, syn. *Solanum lycopersicum*) č. *Solanaceae* - lilkovité

- chromoplasty v subepidermálních parenchymatických buňkách plodu lilku rajčete





Lycopersicon esculentum, obj. 10×



Lycopersicon esculentum, obj. 40×

růže šípková (*Rosa canina*) č. *Rosaceae* - růžovité

- chromoplasty v subepidermálních parenchymatických buňkách plodu růže šípkové; drůzy





Rosa canina, obj. 40×



Rosa canina, obj. 40×

Seznam použitých rostlinných druhů

- begónie (kysala) královská (*Begonia rex*) – příčný řez řapíkem listu – drůzy
- česnek cibule (*Allium cepa*) – suchá zevní epidermis suknice cibule - styloidy
- voděnka (*Tradescantia sp.*) – příčný řez stonkem – rafidy (event. krystalický písek)
- lilek rajče (*Lycopersicon esculentum*) – subepidermální parenchymatické buňky oplodí – chromoplasty
- růže šípková (*Rosa canina*) – subepidermální parenchymatické buňky oplodí – vřetenovité chromoplasty, drůzy
- paprika setá (*Capsicum annuum*) - subepidermální parenchymatické buňky oplodí – chromoplasty

Crystals have been found widely in both the plant and animal kingdoms as far back as the seventeenth century by the pioneers of light microscopy. Although crystal formation in plants occurs in virtually all tissues, its highest concentration may be found within internal leaf idioblasts and external trichome cells ([Arnott & Pautard 1970](#), [Franceschi & Horner 1980](#)). Crystal concentration can vary considerably between species, occasionally reaching a truly enormous mass: for example, *Cactus senilis* has been found to have a dry weight mass of 85% crystalline calcium oxalate ([Cheavin 1938](#)).

Crystal composition is predominantly calcium oxalate (CaOx) but can occur as at least nine other salts. Despite a unique chemical composition, CaOx appears in several shapes (habits) such as: raphide needles, actinic druses, prisms, rhomboids and styloid columns ([Arnott & Pautard 1970](#)). Although the precise function of crystal formation is unclear, much attention has been focused on its role as a noxious anti-foraging device. In addition to making the host plant inedible to predators, crystals may also act as a detoxification mechanism by rendering excess reactive metabolites inert. Others have proposed roles for crystals as either simple unavoidable metabolic by-products or elegant cell regulators of osmotically active ions needed to maintain cell homeostasis ([Rasmussen & Smith 1961](#), [Raven & Smith 1976](#)). They may also have a role in essential nitrogen metabolism. Finally, it is possible that crystals may serve as a passive structural support for non-lignified tissue ([Franceschi & Horner 1980](#)).

A widely accepted function of crystal formation is as an anti-herbivory defense ([Pohl 1965](#), [Ward et al. 1997](#)). This study describes patterns of CaOx crystal accumulation and compares that to a recognized major leaf defense - toughness. Leaf crystal amount and toughness of two age classes in five tropical plants were quantified and correlated.