

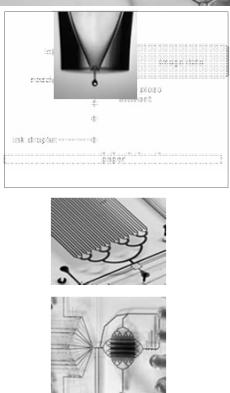
 LOSCHMIDT
LABORATORIES

6. Microfluidics – „Lab on a Chip“

Bi7430 Molecular Biotechnology

Microfluidics

- developed in the 1980s (IBM)
- multidisciplinary field
(engineering, physics, chemistry, material science, nanotechnology)
- integrate processes on chip
 - miniaturization
 - full automation
 - high throughput
 - low energy consumption
 - low sample consumption
 - less waste production



Microfluidics

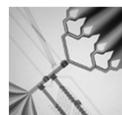
- narrow channels
(mm, μm)
- extremely small volumes
(nL, pL, fL)
- micro domain differs greatly from macroscopic fluids:
 - surface tension
 - capillary forces
 - fluidic resistance
 - fast thermal relaxation
 - laminar flow
 - diffusion



Microfluidic designs

continuous-flow microfluidics

manipulation of continuous liquid flow
through microfabricated channels



droplet-based microfluidics

manipulating discrete volumes of fluids
in immiscible phases



digital microfluidics

droplets manipulated on a substrate
using electrowetting



Lab on a Chip (LOC)

integration of laboratory assays on a chip

- sample preparation
- sample treatment
- detection



life science applications

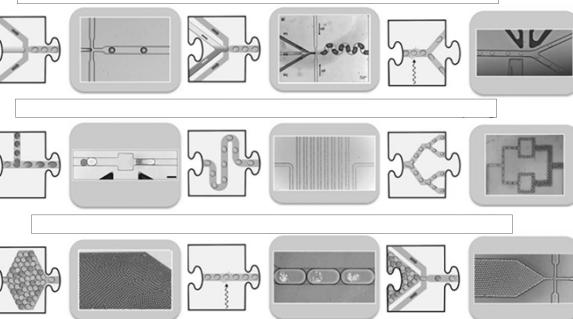
- molecular biology
- diagnostics
- sequencing
- DNA analysis
- proteomics
- clinical studies



Lab on a Chip (LOC)

large variability of designs in LOC toolbox

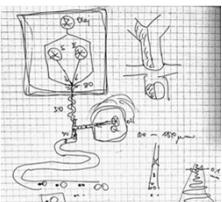
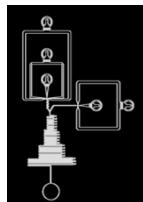
- limitation = back pressure



Chip design and manufacturing

DESIGN

- design softwares** (e.g., AutoCAD, DraftSight)
- printing the mask**

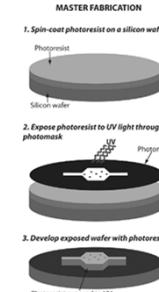
Chip design and manufacturing

FABRICATION

- soft photolithography**

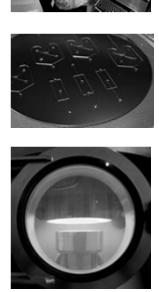
MASTER FABRICATION

1. Spin-coat photoresist on a silicon wafer
2. Expose photoresist to UV light through a photomask
3. Develop exposed wafer with photoresist



PDMS REPPLICATION MOLDING

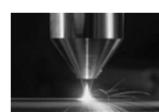
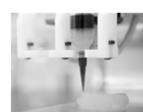
1. Pour PDMS monomer and cross-linker mixture onto master
2. Cure and peel-off PDMS
3. Cut devices, create access ports and bond to glass slide



Chip design and manufacturing

FABRICATION

- direct fabrication methods**
 - **laser cutting**
 - **3D printing**
 - **CNC micro-milling**

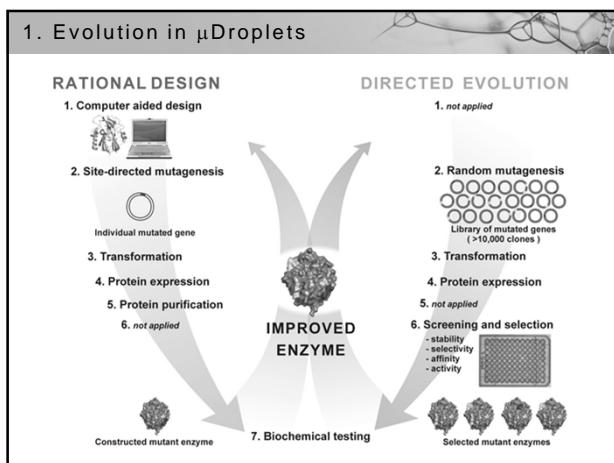

Example of LOC application

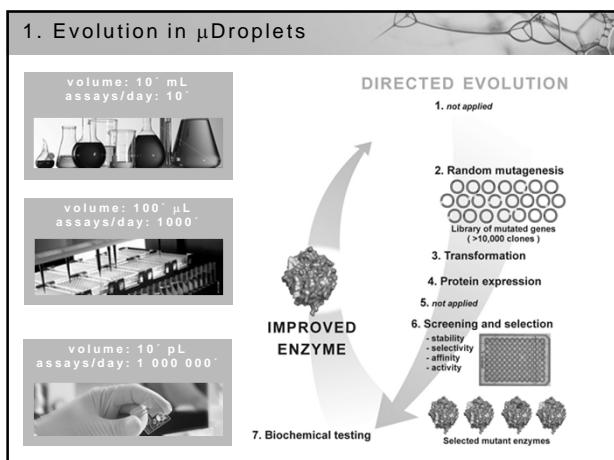
1. DIRECTED EVOLUTION IN μ DROPLETS

2. DRUG SCREENING IN CAPILLARY

3. TRANSIENT KINETICS ON CHIP

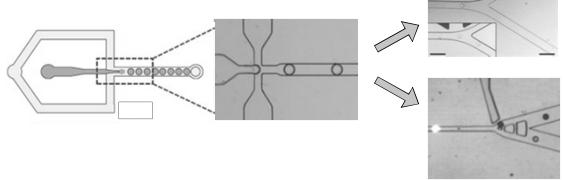
- small volume** → low sample consumption
- small design** → low demand for lab space
- fast operation** → high throughput, fast kinetics





1. Evolution in μ Droplets

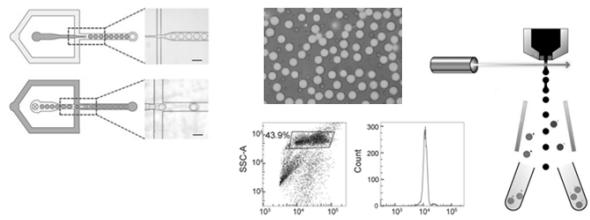
- monodisperse emulsion**
(50 pL, 10^7 droplets/hour)
 - on-chip fluorescence-activated droplet sorting (FADS)**
 - dielectrophoresis, membrane valves, laser manipulation
 - 10^3 events/hour**, 1-3 fluorescence dyes



Baret *et al.* 2009. Lab Chip 9: 1850-1858
Abate *et al.* 2010. Appl. Phys. Lett. 96: 203509

1. Evolution in μ Droplets

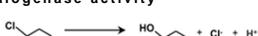
- ❑ monodisperse double emulsion
(2 pL, 10^7 droplets/hour)
 - ❑ off-chip fluorescence-activated cell sorting (FACS)
 - ❑ 10^8 events/hour, 8-12 fluorescence dyes



Lim et al. 2012 | Lab Clin Adv Oncol Article

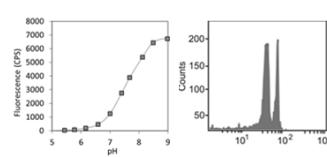
1. Evolution in μ Droplets

- engineering of dehalogenase activity



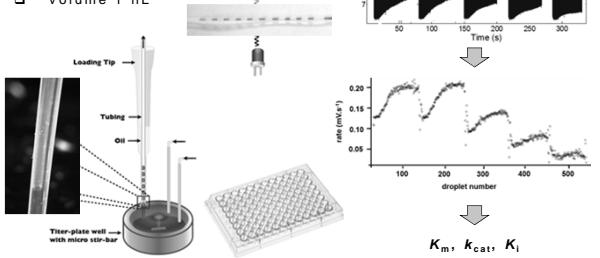
- large library ($>10^6$) in double emulsion
 - fluorescence based activity assay
(fluorescein, rhodamine)

The diagram illustrates the process of generating a double emulsion library. It starts with a single cell, which undergoes division and differentiation. This leads to a stage where the cell contains numerous small, distinct droplets. These droplets represent individual emulsion droplets within a larger spherical carrier, forming the characteristic double emulsion structure.



2. Drug screening in capillary

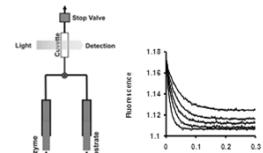
- high throughput generation of droplets (10^3 assay/day)
 - combinatorial measurements
 - concentration gradients
 - frequency 10'Hz
 - volume 1nl



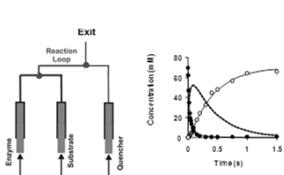
3. Kinetics on chip

RAPID MIXING TECHNIQUES

STORRED FLOW

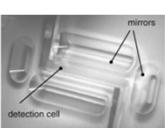
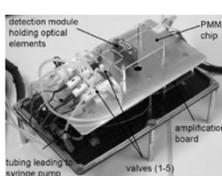


RABID QUENCH

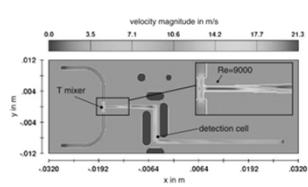


3. Kinetics on chip

□ continuous-flow microfluidics



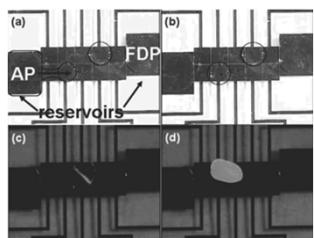
- dead time 8-9 ms (turbulence)
 - volume of reactants 20 μ L
 - poly-(methyl methacrylate) (PMMA;
Plexiglas® 7N) – UV transparent
 - LED (Tm fluorescence, 280 nm)



Bleul et al. 2011 Anal. Bioanal. Chem. 399:1117–1125

3. Kinetics on chip

digital microfluidics

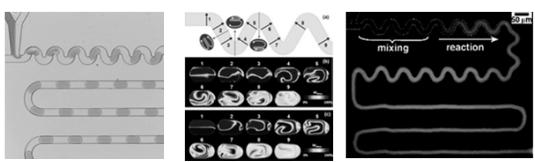


- dead time 15 s (diffusion)
 - volume of reactants 0.07 μ L

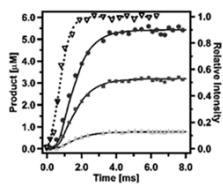
Miller et al. 2008 Anal. Chem. 80, 1614–1619

3. Kinetics on chip

droplet-based microfluidics



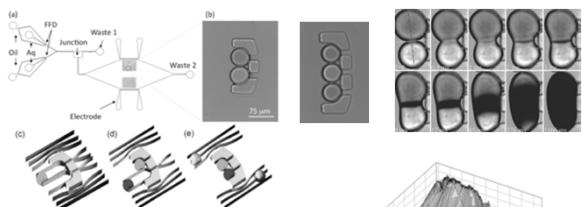
- ❑ dead time < 1 ms (chaotic advection)
 - ❑ volume of reactants 0.15 μ L
 - ❑ poly(dimethylsiloxane) – PDMS and glass
 - ❑ CCD camera and mikroskop



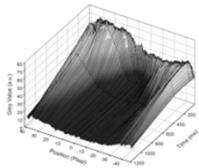
Song et al. 2003 J Am. Chem. Soc. 125:14613-14619

3. Kinetics on chip

□ droplet-based microfluidics



- ❑ dead time 0.1 ms (diffusion)
 - ❑ volume of reactants pL
 - ❑ poly-(dimethylsiloxane) – PDMS and glass
 - ❑ CCD camera and microscope



Microfluidics



- new field of development
- exponentially growing area
- large space for applications
- number of scientific journals
- commercial companies arrises