

# Conformation, allostery

Petr Louša

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# Outline

- 1 Introduction
- 2 Allostery
- 3 Kinetics of conformational changes
- 4 Folding

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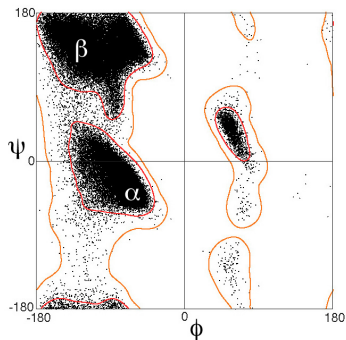
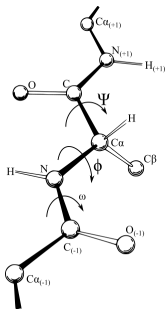
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# Conformation of proteins

**constitution** topology of molecule – *isopropanol*, *n-propanol*

**configuration** bond arrangement – *cis/trans*, *R/S*

**conformation** 3D structure – rotation around single bonds

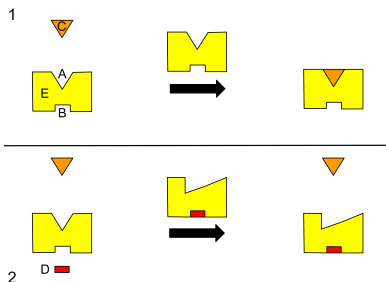


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# Allostery

- Change of conformation after ligand binding.
- $\Rightarrow$  Change of  $K_d$  for other ligands.
- Typical example **hemoglobin**.
- Often multimeric proteins with more equivalent active sites.



# Types of allostery

- By type of  $K_d$  change:
  - positive –  $K_d$  decreases – next substrate binds more easily – **activation**
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- By type of substrate:
  - **homotropic** – more same ligands bind to protein – e.g. hemoglobin
  - **heterotropic** – activator/inhibitor differs from next bound substrate – e.g. strychnine inhibits glycine receptor (inhibitive neurotransmitter)

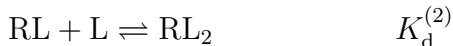


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- Non-regulatory allostery
  - protein needs other component for its function, but is not regulated by it – e.g. ions, vitamins

# Hill equation I

- Quantification of cooperativity



$$\vdots$$


- Constants  $K_d^{(i)}$  differ – system is cooperative.

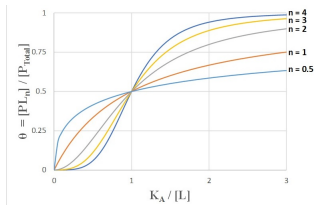
# Hill equation II

- Assumption –  $[RL]_i = 0$
- Only one equation remains  $\Rightarrow$  Hill analysis.



$$\hat{K}_d = \frac{[R][L]^n}{[RL]_n} \quad (2)$$

$$\hat{K}_d = (K_d)^n \quad (3)$$

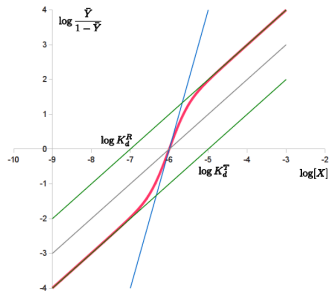


# Analysis of experiment

- Measure fraction of bound receptors and linearize:

$$y = \frac{[RL]_n}{[R_{\text{tot}}]} \quad (4)$$

$$\log\left(\frac{y}{1-y}\right) = n \log[L] - \log \hat{K}_d \quad (5)$$



# Microscopic models I – MWC model I

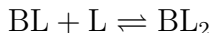
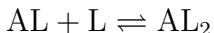
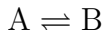
- Monod, Wyman, Changeux
- Two-state receptor system – can “switch” only in free form.
- Form A is dominant in free form.

$$[A] \gg [B]$$

- Affinity to B is significantly larger.
- Assumes change of protein structure after binding – “locking” in B state.

# Microscopic models I – MWC model II

- Assumes constant microscopic  $K_d$ .



- Ligand binds preferentially to B and shifts the equilibrium of free forms.
- Problem?

# Microskopik models II – KNF model

- Koshland, Nemethy, Filmer
- Generalization of MWC model.
- Assumes different  $B$  constants for sequential equilibria.
- $K_d$  constants are free parameters for fitting.
- Each ligand binding changes the binding site for other ligands.
  
- Disadvantage?

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- Disadvantage: too many free parameters ( $K_d$  constants) – better for experiment fitting than for predictions.

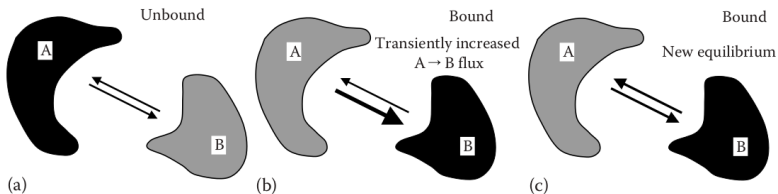


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# Kinetic view to allostery

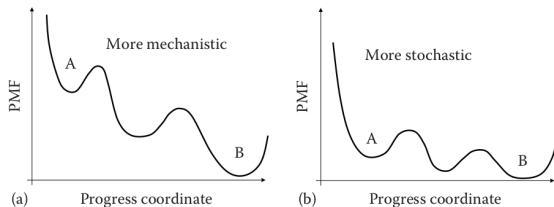
- Ligand binding changes equilibrium between two “states” (MWC model).
- After binding, structural ensemble changes – B forms dominate.
- Caused by changes of kinetic parameters of the transition.



# Processivity vs. Stochasticity

## Processivity

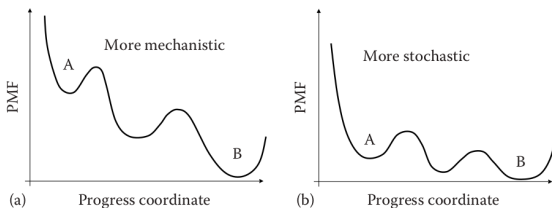
- Ability to run irreversibly in one direction.
- E.g. ATP synthase, motor proteins, polymerases
- Free energy decreases in larger jumps – irreversibility.
- Source of energy needed – ATP, GTP, proton gradient.
- ATP – ca  $20k_B T$  of energy.



# Processivity vs. Stochasticity

## Stochasticity

- Many reversible steps.
- E.g. glycolysis.
- Reversibility –  $\Delta G \approx k_B T$
- Even many reversible steps can lead to irreversible event –  $\Delta G$  adds up.



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# Fundamental questions

1. What structure does given sequence of amino acids take?
2. How does the protein fold?

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  - Homology modeling
  - Prediction *de novo*
    - Global minimum search  $\Delta G$
    - MD of stretched chain
    - Folding@home, Foldit
    - Evolution covariation – requires hundreds of homologous *sequences*
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## 2. How does the protein fold?

- Folding process
- Kinetics
- Transit states
- Intermediates



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  - Often capable of multiple de- and renaturation.
  - Anfinsen experiment – renaturation of ribonuclease A.
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  - Folding in membrane.
- Only with “helpers” – chaperons.
  - Hydrophobic boxes.
  - Proteins can search through the configuration space more easily.

# Folding as conformation change

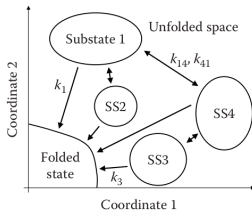
- Folding follows the same physics as other structural changes.

## Differences:

- Large ranges of equilibrium and nonequilibrium conditions.
- De-/renaturation can go very slowly and reversibly or “immediately”.
- “Unfolded state” is **not** a state.
- Many different substates – difficult to characterize.
- Differ also in the denaturation process – temperature, pH, chemical agents,...

# Overall folding rate

- Unfolded protein can have multiple substates.
- Only some allow transition to folded state.
- Depends on particular rate constants.
- Depends on substate populations.
- Population can differ based on the denaturation process.



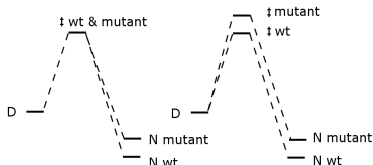
# $\Phi$ values analysis

- Alan Fersht
- Influence of individual residues on folding and transient states.
- Compare  $\Delta G$  profiles for wild type and mutant.

$$\Delta\Delta G_{ij} = \Delta G_{ij}(\text{mut}) - \Delta G_{ij}(\text{wt}) \quad (6)$$

$$\Phi_F = \frac{\Delta\Delta G_{D\dagger}}{\Delta\Delta G_{DN}} \quad (7)$$

- $\Phi_F = 0$  residuum is unfolded in transient state.
- $\Phi_F = 1$  residuum is folded in transient state.



# Conformation, allostery – exercise

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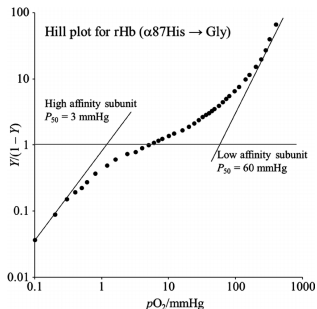


# Hill equation

1. Draw Hill plot for case of negative cooperativity.

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# Free energy differences

Assume free energy differences between states A and B of:

1.  $1 k_B T$
2.  $5 k_B T$
3.  $20 k_B T$

Calculate ratio of forward and backward reaction. Think, whether it is processive or stochastic process.

# Free energy differences – solution

$$\Delta G = -RT \ln K \quad (8)$$

$$\ln K = \frac{\Delta G}{RT} \quad (9)$$

$$K = \frac{k_{\text{on}}}{k_{\text{off}}} = \exp \frac{\Delta G}{RT} \quad (10)$$

1.  $1 k_{\text{B}}T \Rightarrow K = 2.7$
2.  $5 k_{\text{B}}T \Rightarrow K = 150$
3.  $20 k_{\text{B}}T \Rightarrow K = 4.85 \cdot 10^8$

# References

- Zuckerman, Daniel M. *Statistical Physics of Biomolecules. An Introduction*
- Atkins, Peter; de Paula, Julio. *Physical Chemistry*
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- Wikipedia