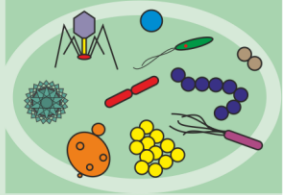


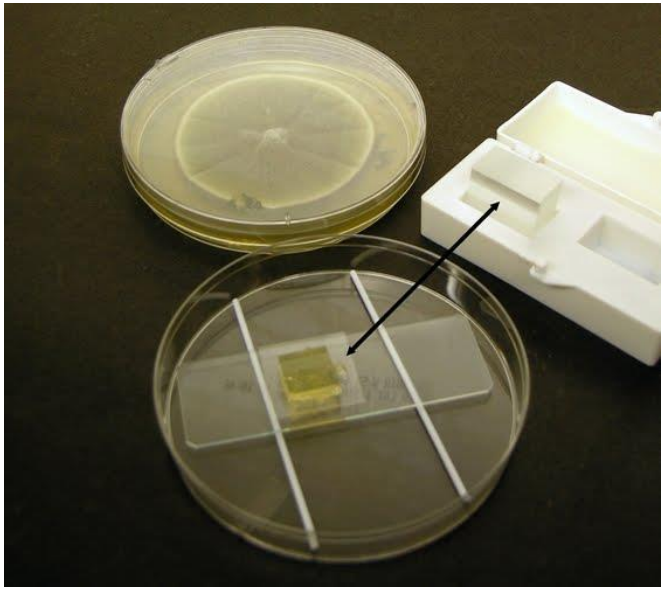
5. – 6. practice

Slide cultures and fluorescence

Fidrich (2018)

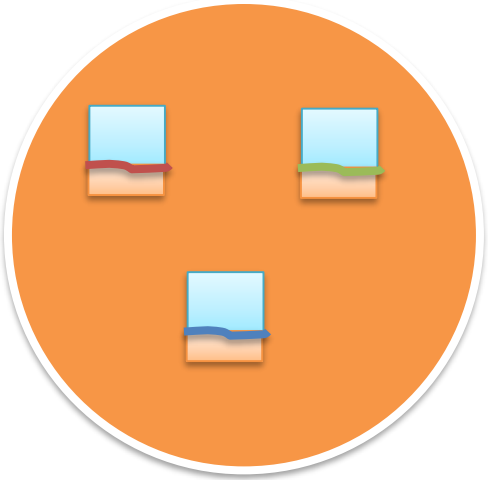
Slide cultures I.

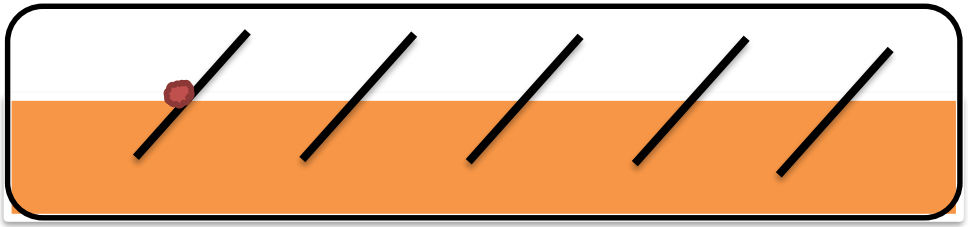
1. Place the microscopic slide on glass beads in sterile Petri dish
2. Place the thin square of agar on microscopis slide by sterilised scalpel
3. Innoculate edges of agar and cover it by sterile cover glass
4. Add circa 5 ml sterile distilled water on glass beads under microscopis slide
5. Close the Petri dish



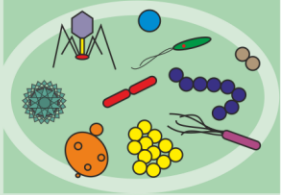
Slide cultures II.

1. Stab sterile cover glass to the agar (45°angle) – slanted cover glass
2. Innoculate the culture on agar along the cover glass
3. Close the Petri dish





What to observe ?



- 1x slide cultre on glass beads
- 1x slanted cover glass

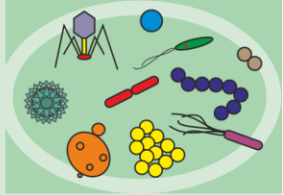
5 day cultures:

Rhodococcus erythropolis (M8 or MPA)

Nocardia carnea (M8)

Streptomyces griseus (M15)

Fluorescence - DAPI staining



1. Pour water inside the measuring cylinder (cca 5 ml)
2. Filter culture through the bacterial filter (25 µl of each culture)
3. Allowed to air dry

[redacted]

4. Pipette 20 µl of DAPI stain
5. Put in fridge for 10 min
6. Wash in water – ethanol – water
7. Let it dry
8. Drop imersion oil on microscopil slide, put the filter, imersion oil again, cover glass and imersion oil again