

# CG920 Genomics

## Finishing Lesson 2

### Genes Identification

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INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ

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Evropským sociálním fondem  
a státním rozpočtem České republiky

# Outline

(finishing Lesson 02)

- **Forward and Reverse Genetics Approaches**
  - Differences between the approaches used for identification of genes and their function
- **Identification of Genes *Ab Initio***
  - Structure of genes and searching for them
  - Genomic colinearity and genomic homology
- **Experimental Genes Identification**
  - Constructing gene-enriched libraries using methylation filtration technology
  - EST libraries
  - Forward and reverse genetics

# Forward and Reverse Genetics

- Principles of experimental identification of genes using forward and reverse genetics
  - Alteration of phenotype after mutagenesis
    - **Forward genetics**
  - Identification of sequence-specific mutant and analysis of its phenotype
    - **Reverse genetics**
  - Analysis of expression of a particular gene and its spatiotemporal specificity

# Forward Genetics

- Principles of experimental identification of genes using forward and reverse genetics
  - Alteration of phenotype after mutagenesis
    - **Forward genetics**

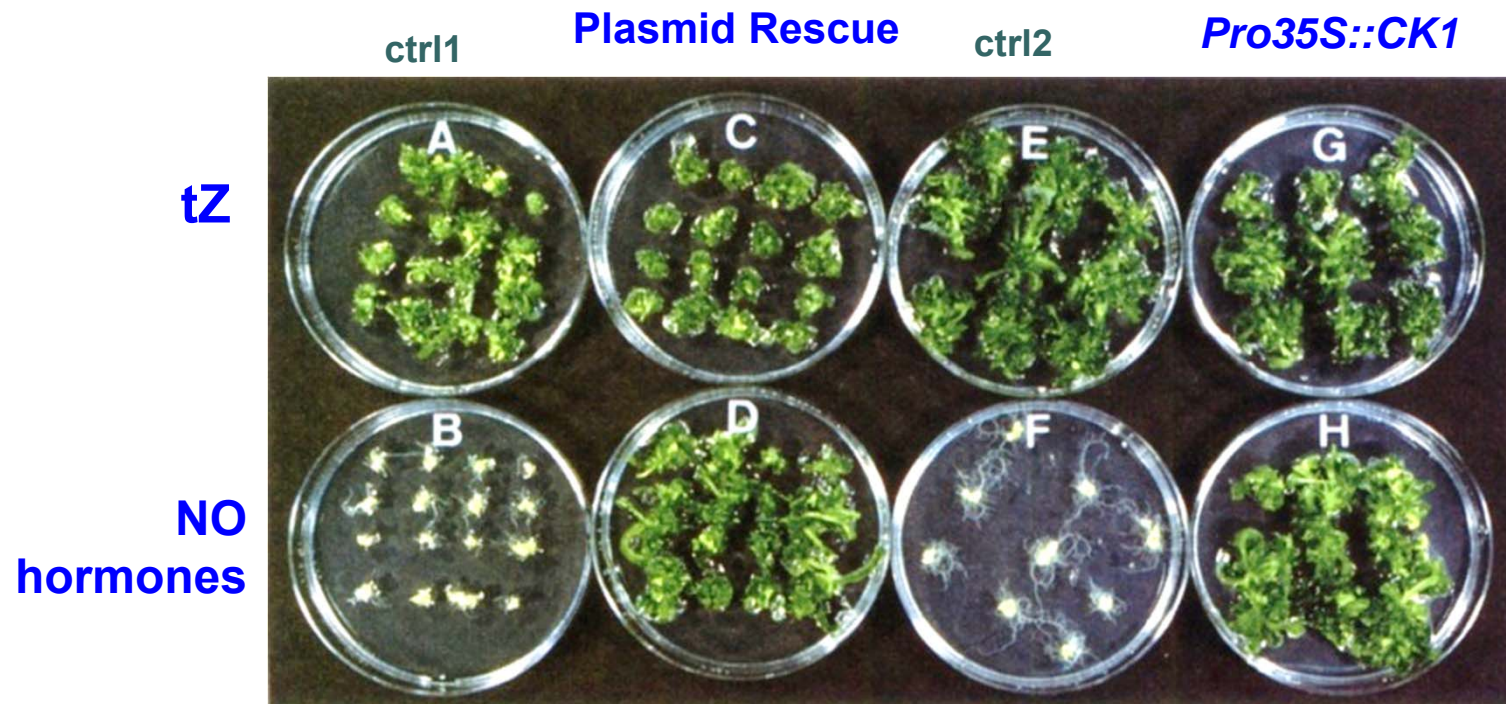


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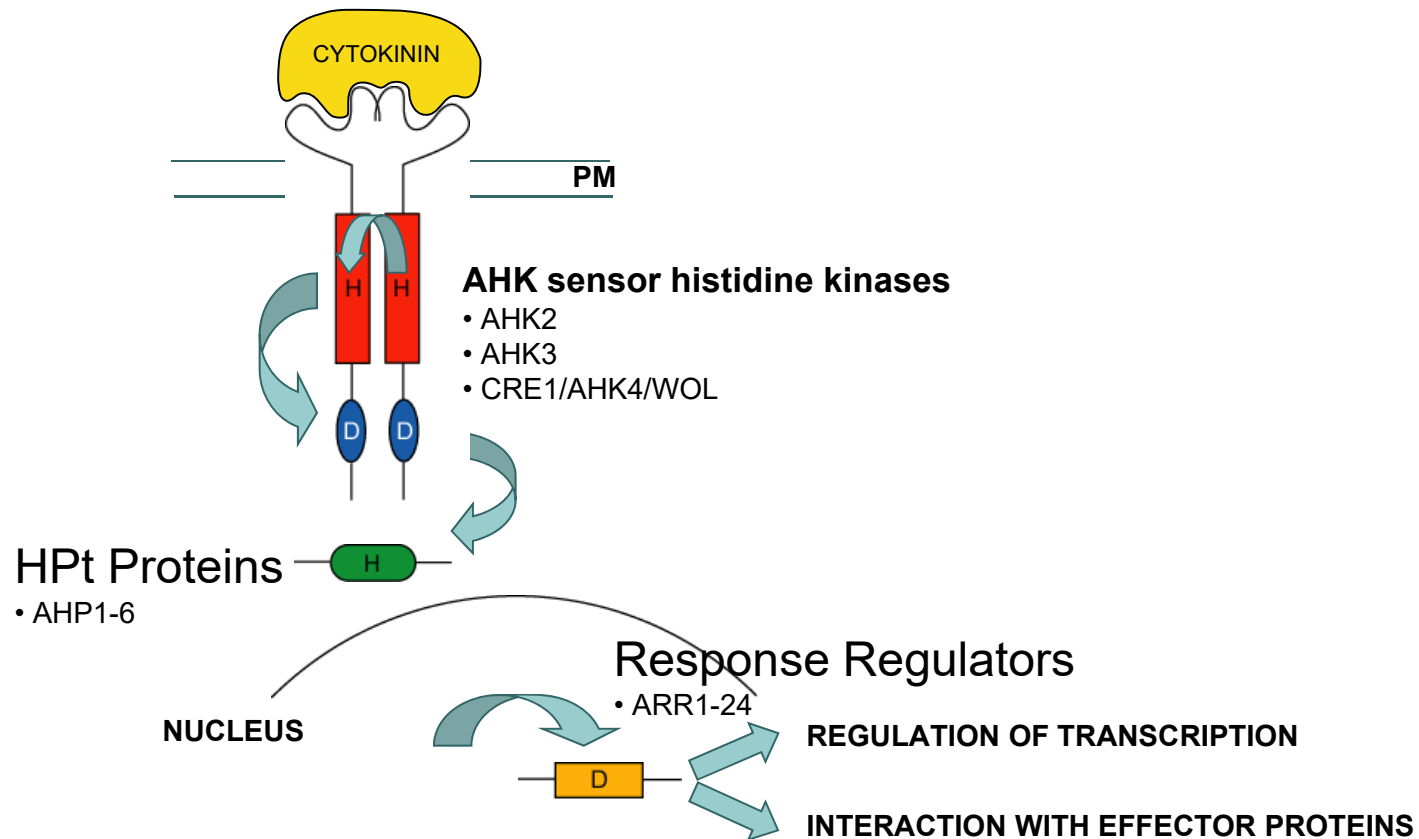
# Identification of *CKI1* via Activation Mutagenesis

- *CKI1* overexpression mimics cytokinin response



Kakimoto, *Science*, 1996

# Signal Transduction via MSP

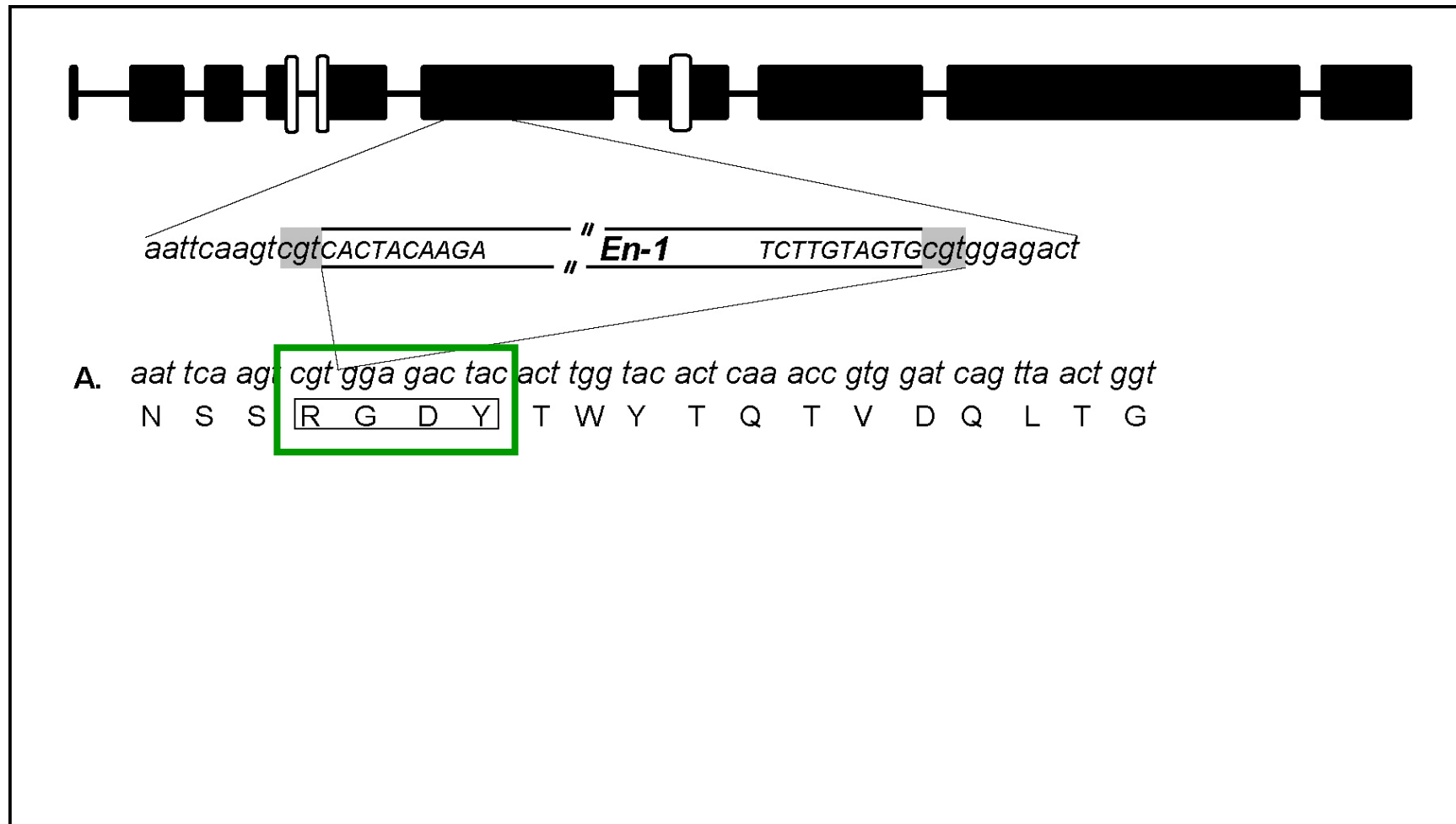




# Reverse Genetics

- Principles of experimental identification of genes using forward and reverse genetics
  - Alteration of phenotype after mutagenesis
    - **Forward genetics**
  - Identification of insertional mutant and analysis of its phenotype
    - **Reverse genetics**

# Identification of insertional *cki1* mutant allele

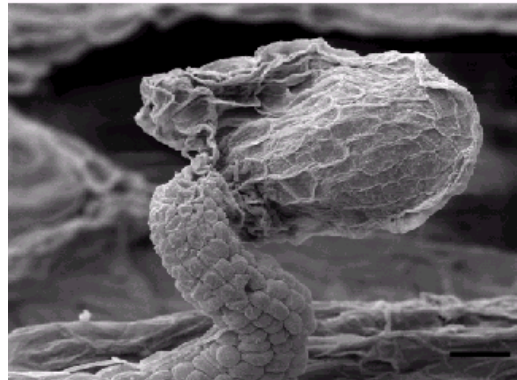




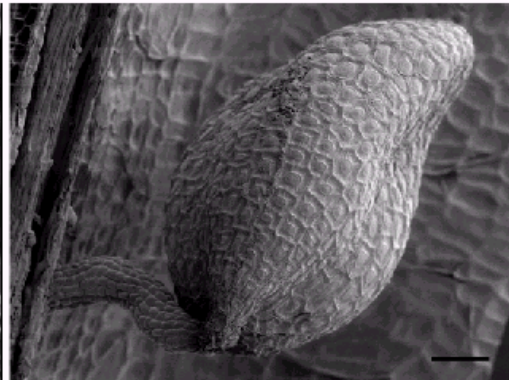
# CKI1 Regulates Female Gametophyte Development

- CKI1 is necessary for proper megagametogenesis in *Arabidopsis*

*CKI1/cki1-i*



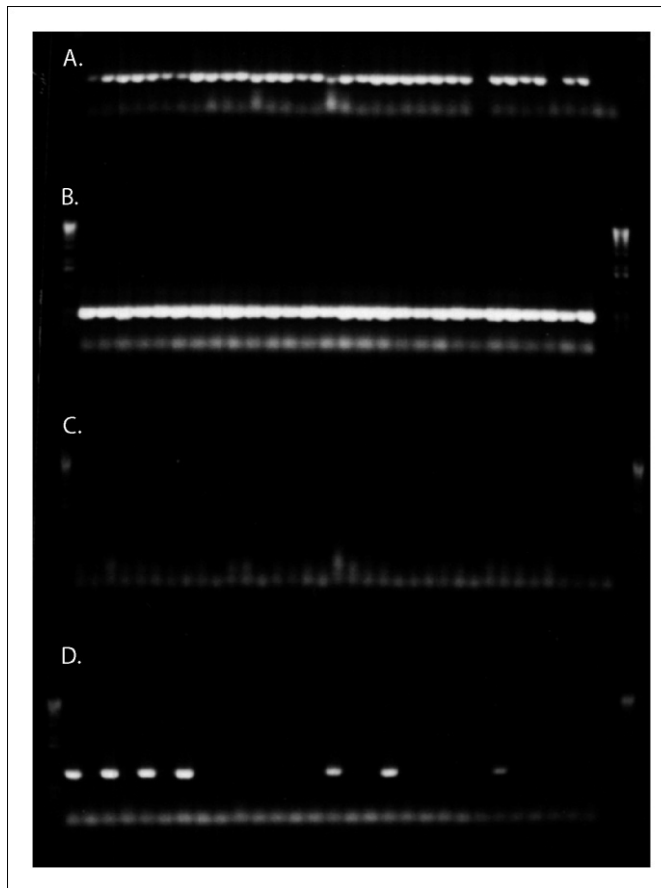
*CKI1/CKI1*



Hejátko et al., *Mol Genet Genomics* (2003)

# CKI1 and Megagametogenesis

- *cki1-i* is not transmitted through the female gametophyte



A. ♂ wt x ♀ **CKI1/*cki1-i***



CKI1 specific primers (PCR positive control)

B. ♂ **CKI1/*cki1-i*** x ♀ wt

C. ♂ wt x ♀ **CKI1/*cki1-i***

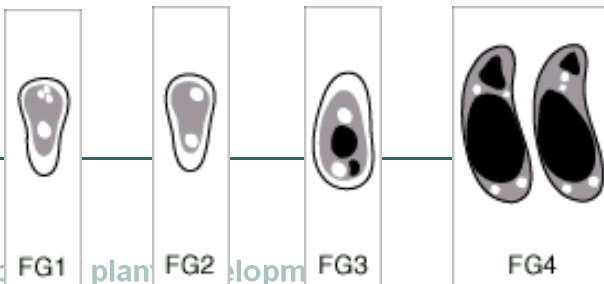
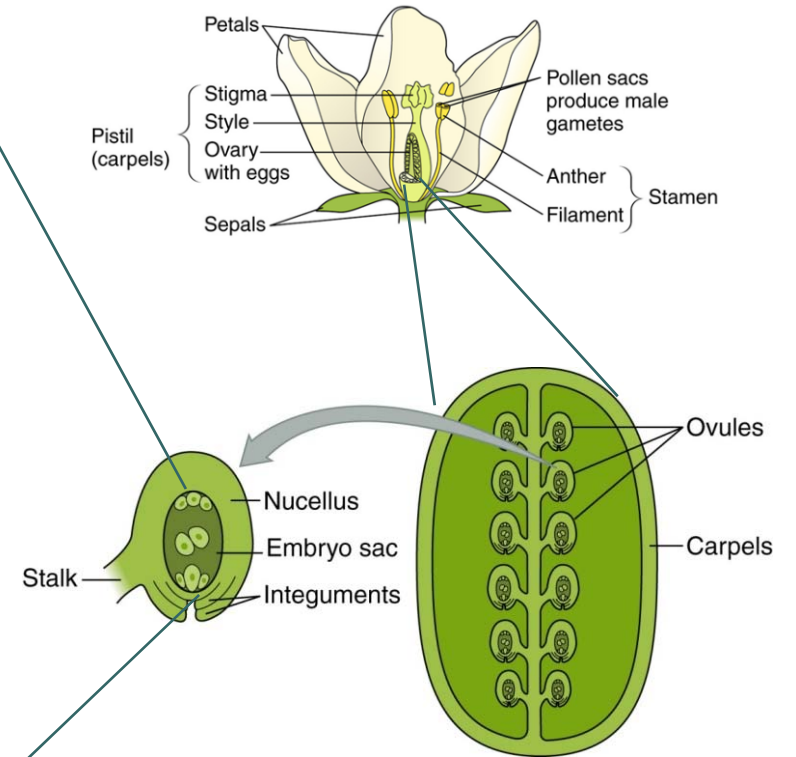
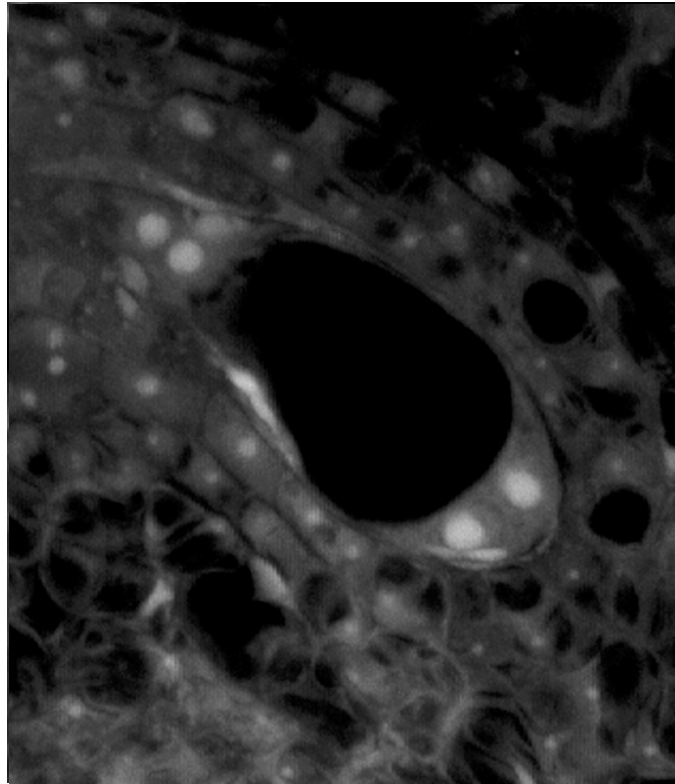


*cki1-i* specific primers

D. ♂ **CKI1/*cki1-i*** x ♀ wt

# CKI1 and Megagametogenesis

FG 4



Hormonal regulation of plant development



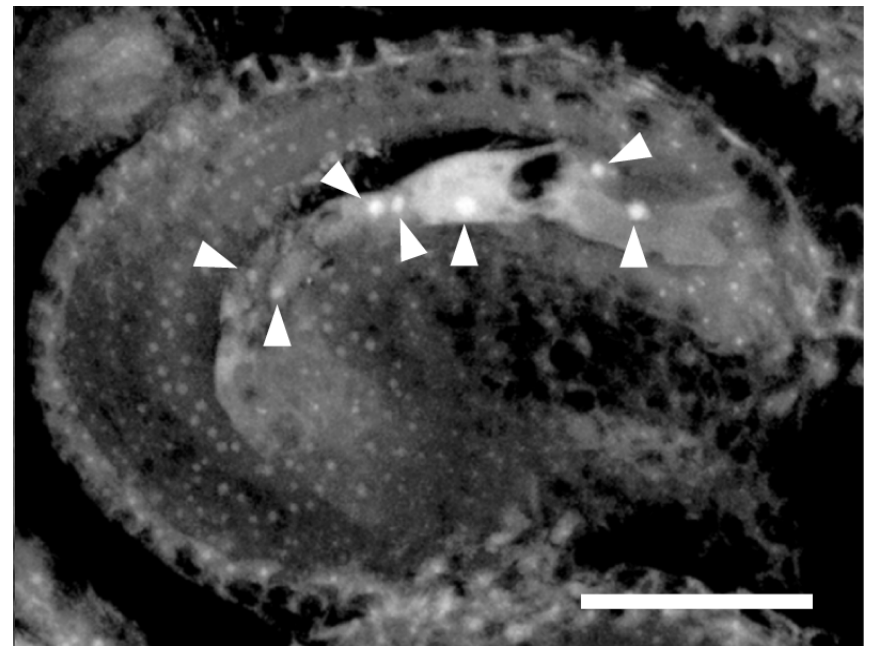
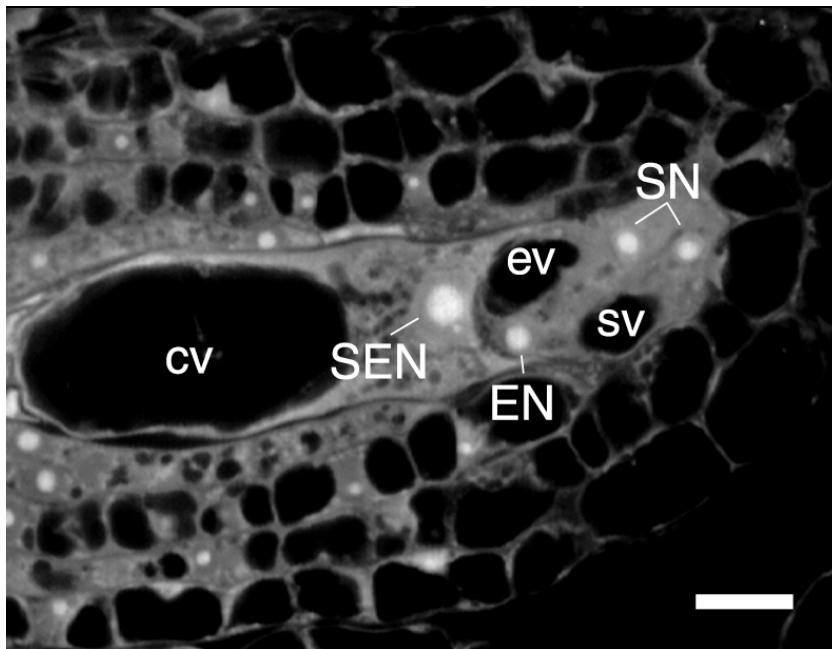
# CKI1 and Megagametogenesis

CKI1

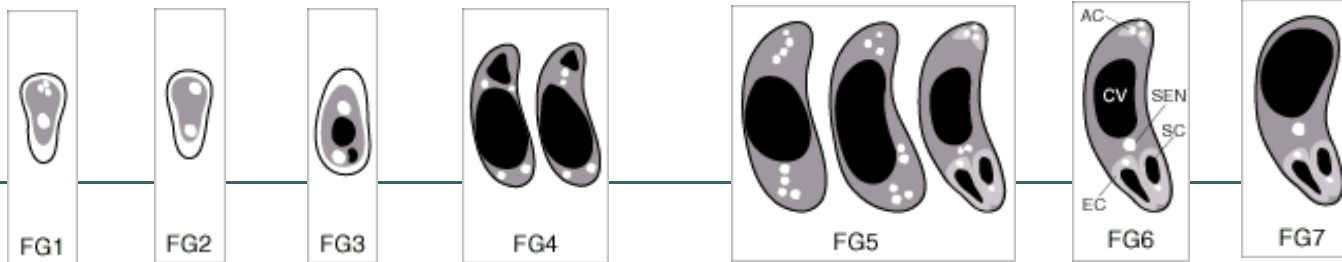
FG4 to FG5

*cki1-i*

28 HAE



Hejatko et al., *Mol Genet Genomics* (2003)

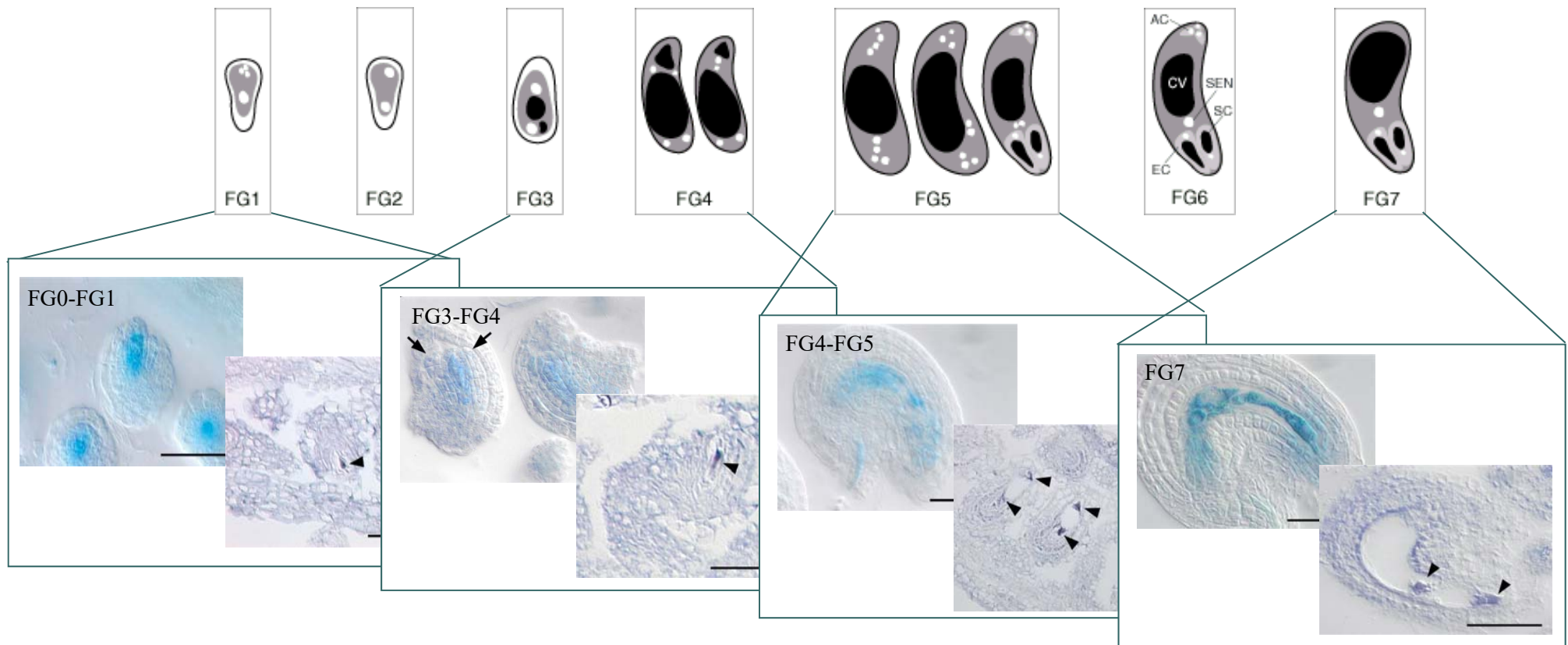




# Forward and Reverse Genetics

- Principles of experimental identification of genes using forward and reverse genetics
  - Alteration of phenotype after mutagenesis
    - **Forward genetics**
  - Identification of insertional mutant and analysis of its phenotype
    - **Reverse genetics**
  - Analysis of expression of a particular gene and its spatiotemporal specificity

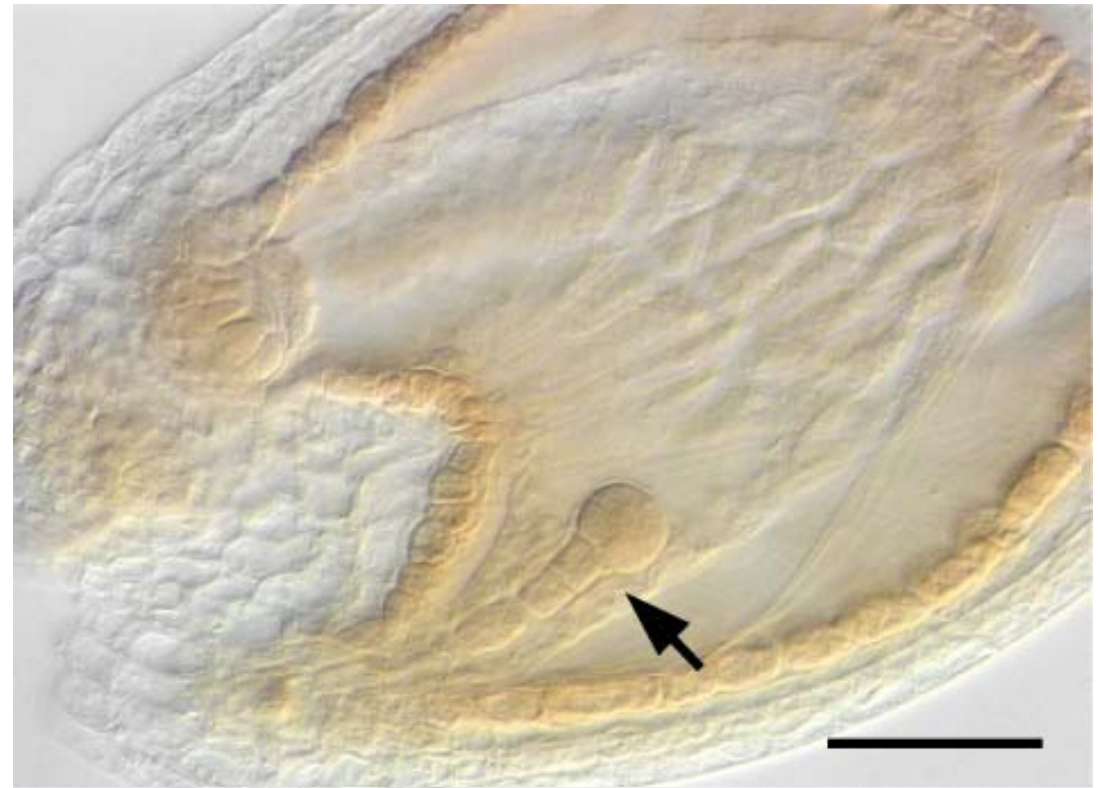
# CKI1 is Expressed During Megagametogenesis



# Paternal *CKI1* is Expressed in the *Arabidopsis* Sporophyte Early after Fertilization

♀ wt x ♂ Pro*CKI1*:*GUS*

**28 HAP**  
(hours  
after  
pollination)



Hejátko et al., *Mol Genet Genomics* (2003)



# CG020 Genomics

## Bi7201 Genomics – a basic course

### Lesson 3

#### Reverse Genetics

Jan Hejátko

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# Literature

- Literature sources for Chapter 03:
  - **Bioinformatics and Functional Genomics**, 2009, Jonathan Pevsner, Willey-Blackwell, Hoboken, New Jersey  
<http://www.bioinfbook.org/index.php>
  - **Plant Functional Genomics**, ed. Erich Grotewold, 2003, Humana Press, Totowa, New Jersey
  - Mello, C.C. and Conte Jr., D. (2004) Revealing the world of RNA interference. *Nature*, **431**, 338-342.
  - Klinakis et al.. (2000) Genome-wide insertional mutagenesis in human cells by the *Drosophila* mobile element *Minos*. *EMBO Rep*, **1**, 416.
  - Hansen et al.. (2003) A large-scale, gene-driven mutagenesis approach for the functional analysis of the mouse genome. *PNAS*, **100**, 9918.

# „Classical“ genetics *versus* „reverse genetics“ approaches in functional genomics

## RANDOM MUTAGENESIS

### „Classical genetics“ approach

1. IDENTIFICATION OF PHENOTYPE

2. GENE MAPPING

3. GENE IDENTIFICATION  
- position cloning

EMS



$h \times n$

T-DNA



### „Reverse genetics“ approach

1. ISOLATION OF SEQUENCE-  
-SPECIFIC MUTANT

2. IDENTIFICATION OF  
PHENOTYPE

3. PROOF OF CAUSAL RELATIONSHIP  
BETWEEN INSERTION AND  
PHENOTYPE

(retro)transposons



# Outline

- **Methods for Identification of Sequence-Specific Mutants**
  - Preparation of mutants collection
  - Searching for sequence-specific mutants using PCR
  - Searching for sequence-specific mutants in electronic databases
  - Knocking-out the gene using homologous recombination
- **Analysis of Phenotype and Confirmation of Causality Between Phenotype and Insertional Mutation**
  - Co-segregation analysis
  - Identification of independent insertional allele
  - Using unstable insertional mutagens and isolation of revertant lines
  - Mutant complementation by the transgene

# Outline

- Gene Silencing Using RNA Interference
  - Mechanism of RNA interference
- Genome Editing via CRISPR/Cas9

# Outline

- Methods of identification of sequence-specific mutants
  - Preparation of mutants collection



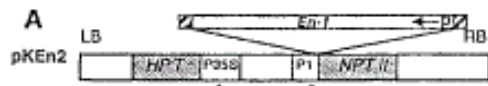
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# Types of Insertional Mutagens

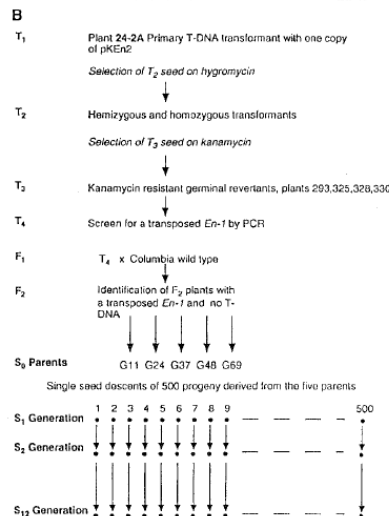
- Mobile elements
  - **Autonomous transposons (*En-1*)**
    - They contain a gene for transposase, enabling excision and reintegration into the genome
    - At both ends they contain short inverted repeat, which are recognized by transposase
- Stable elements
  - **Non-autonomous transposons (*dSpm*)**
    - mutant of *En/Spm* transposon, which has lost autonomy because of mutation in a gene for transposase
    - It can be activated by crossing with a line carrying the *En/Spm* transposon
  - **T-DNA**
    - completely stable, however, its insertion can lead to chromosome rearrangements (inversions, deletions, transpositions)

# Libraries of Insertional Mutants (plants)

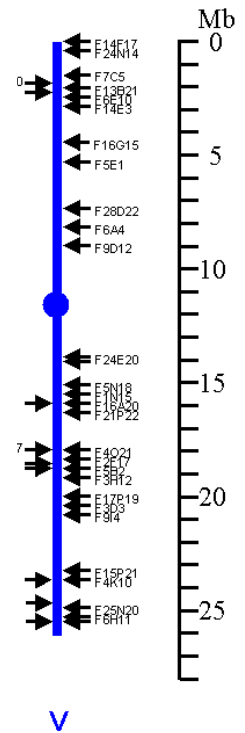
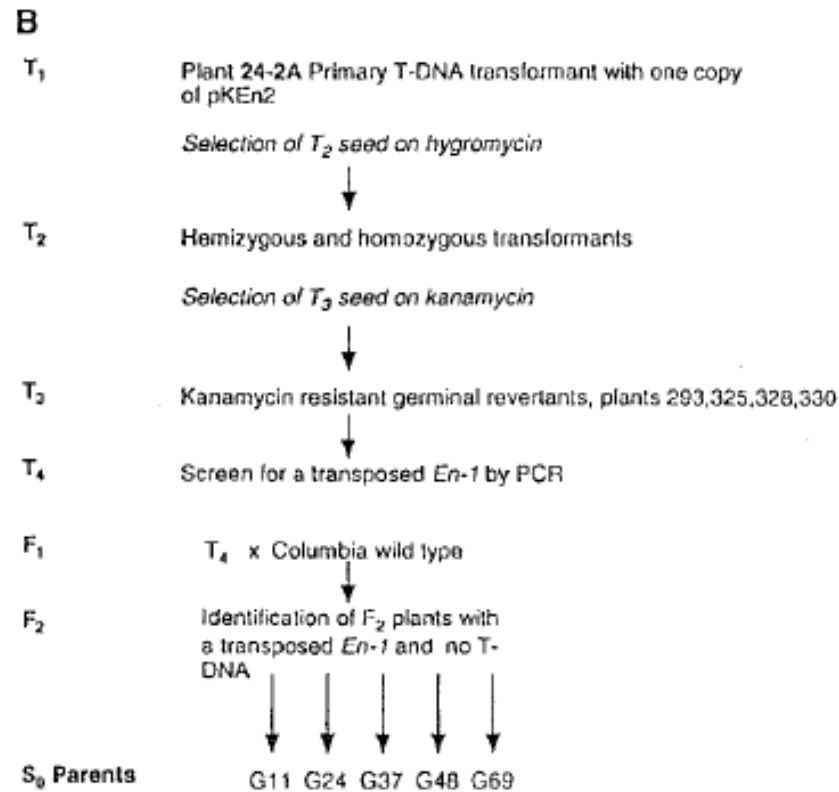


Preparation of transgenic plants

Creating the population of mutants

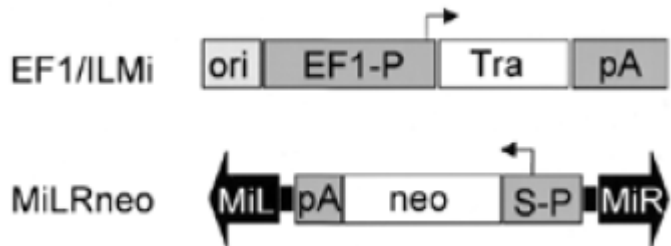


Searching for sequence-specific mutants by PCR



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je spolufinancována  
im sociálním fondem  
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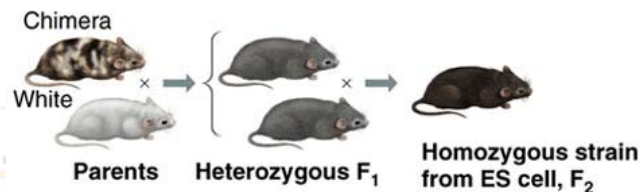
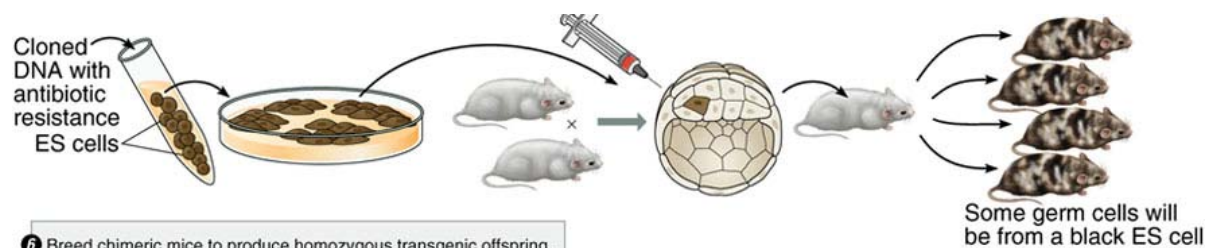
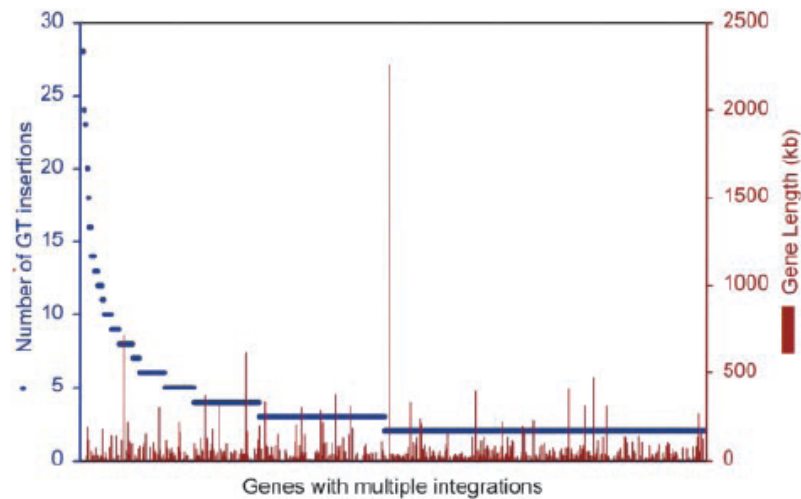
# Libraries of Insertional Mutants (animals)



Transfection into human cell cultures (HeLa) or mouse embryonic stem (ES) cells

Generating a population of mutant cell lines and frequency-analysis of insertions

*in vitro* analysis or preparation of library of insertional mutants by reintroduction ES into mouse embryos





# Outline

- Methods of identification of sequence-specific mutants
  - Preparation of mutants collection
  - Searching for sequence-specific mutants using PCR
    - PCR-based three-dimensional screening

# Isolation of sequence-specific mutants

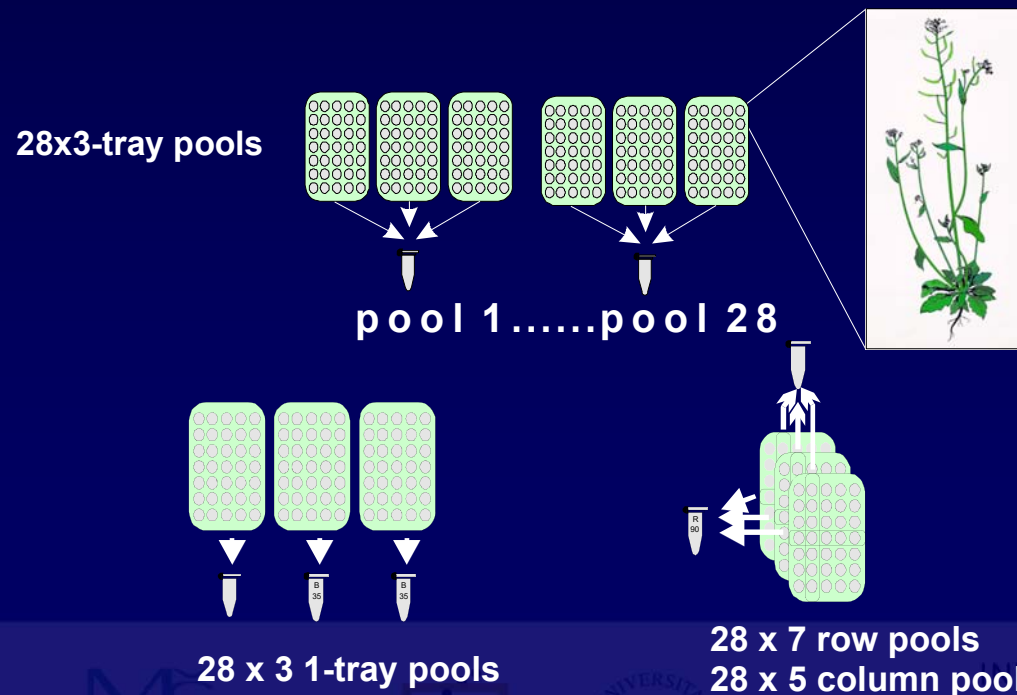
## 1. Library of *En-1* insertional mutants

- autonomous *En/Spm*, without selection
- 3000 independent lines
- 5 copies per line on average
- PCR-based three-dimensional screening

# Isolation of sequence-specific mutants

- PCR-based three-dimensional screening
  - Isolation of genomic DNA from the individual plants of mutant population and creating sets of DNA („triads“, rows and columns of triads and individual trays)

3.000 mutant lines of *A. thaliana* (5 copies of En-1/line)



# Isolation of sequence-specific mutants

- PCR-based three-dimensional screening
  - Isolation of genomic DNA from the individual plants of mutant population and creating sets of DNA („triads“, rows and columns of triads and individual trays)
  - Identification of positive „triad“ with PCR, blotting of PCR products and hybridization of the PCR products with gene-specific probe



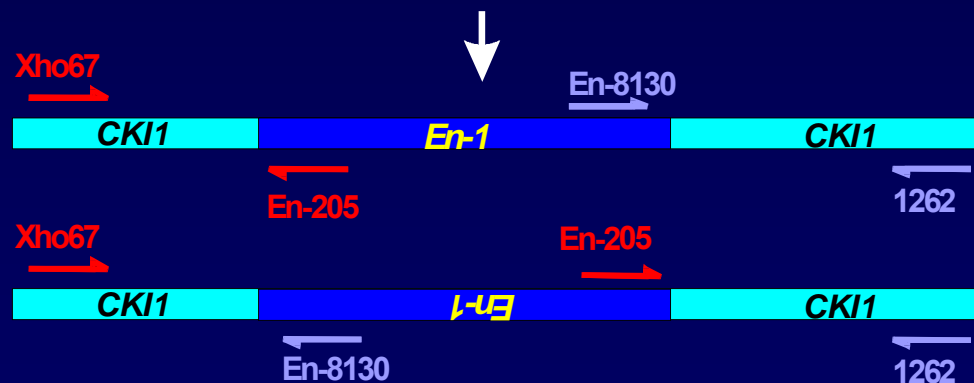
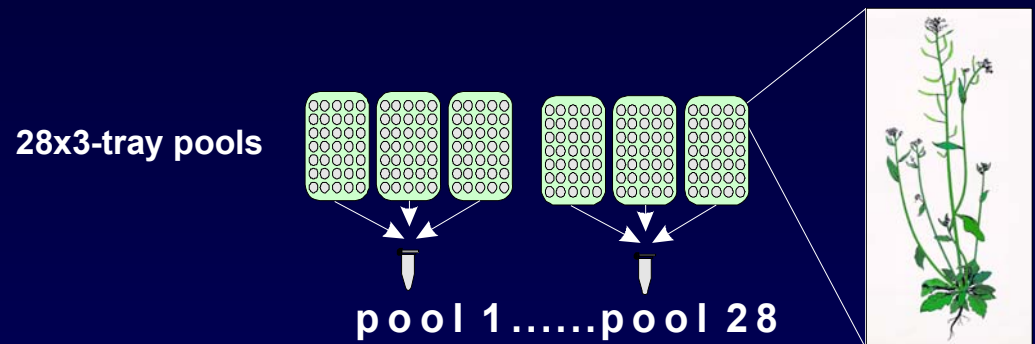
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# Isolation of sequence-specific mutants

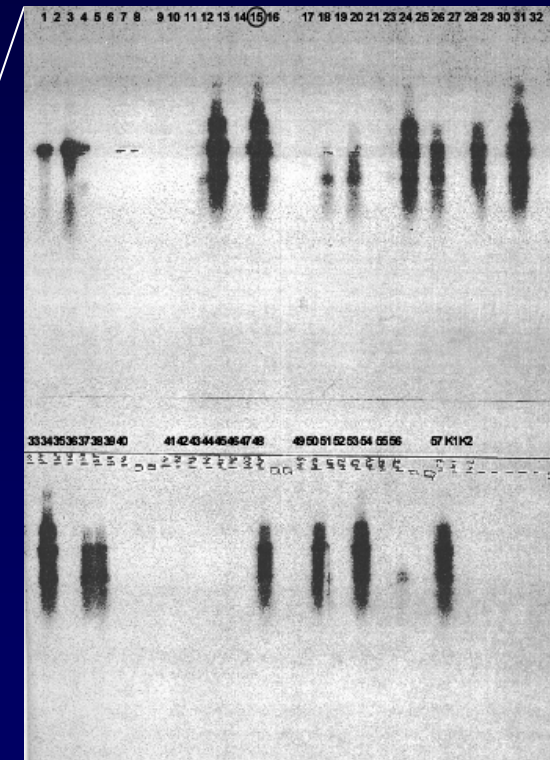
## 1. 3-tray pools screen

3.000 mutant lines of *A. thaliana* (5 copies of En-1/line)



(2x2x28=112 PCR reactions)

Identification of the PCR product by hybridization with a gene-specific probe



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# Isolation of sequence-specific mutants

- PCR-based three-dimensional screening
  - Isolation of genomic DNA from the individual plants of mutant population and creating sets of DNA („triads“, rows and columns of triads and individual trays)
  - Identification of positive „triad“ with PCR, blotting of PCR products and hybridization of the PCR products with gene-specific probe
  - Identification of the positive line through identification of positive tray, row and column



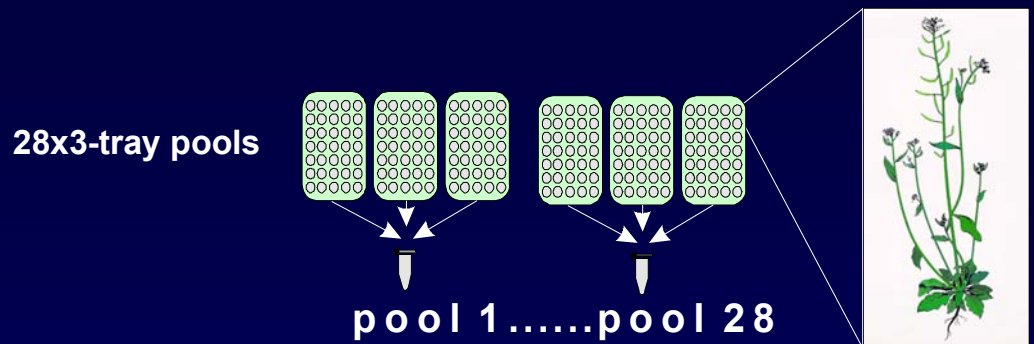
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# Isolation of sequence-specific mutants

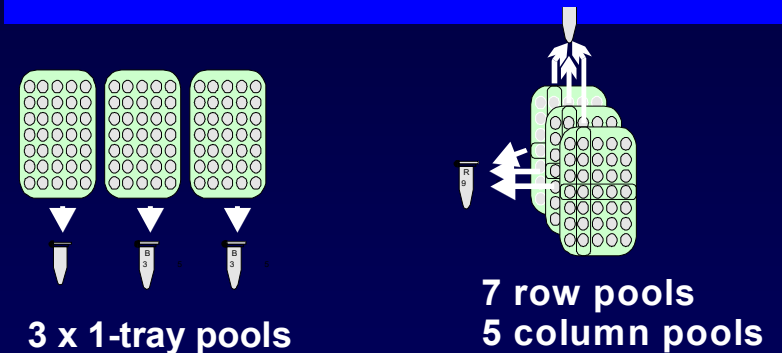
## 1. 3-tray pools screen

3.000 mutant lines of *A. thaliana* (5 copies of En-1/line)



(2x2x28=112 PCR reactions)

## 2. Identification of line carrying the insertion



(another 5+7+3=15 PCR reactions)

**In total: 112+15=127 PCR reactions**

Identification of the PCR product by hybridization with a gene-specific probe

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# Outline

- Methods of identification of sequence-specific mutants
  - Preparation of mutants collection
  - Searching for sequence-specific mutants using PCR
    - PCR-based three-dimensional screening
    - Hybridization with iPCR products on filters



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# Isolation of sequence-specific mutants

## Insertion library of dSpm mutants

- The Sainsbury Laboratory (SLAT-lines), John Innes Centre, Norwich Research Park
- DNA and seeds in Nottingham Seed Stock Centre
- 48.000 lines
- 1.2 insertion per line on average
- non-autonomous transposon
- PCR searching or hybridization with iPCR filters
- SINS (sequenced insertion sites) database

<http://nasc.nott.ac.uk>



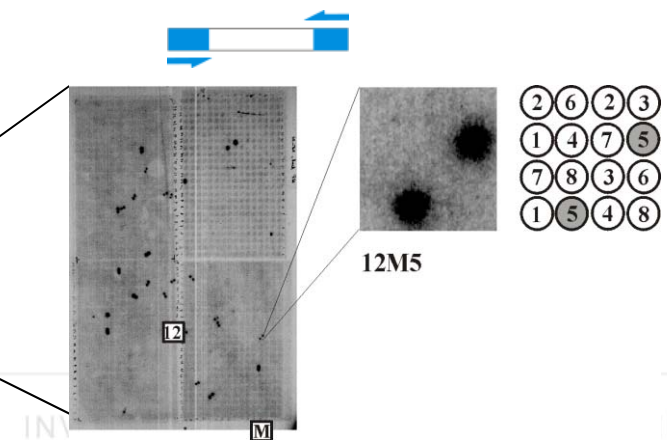
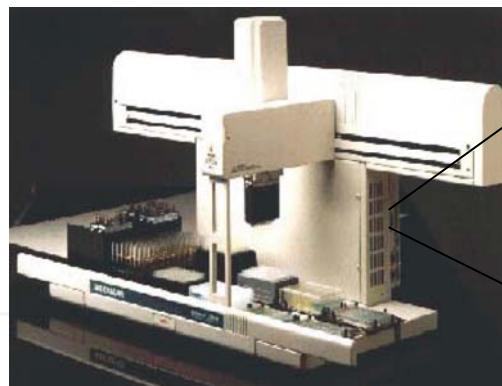
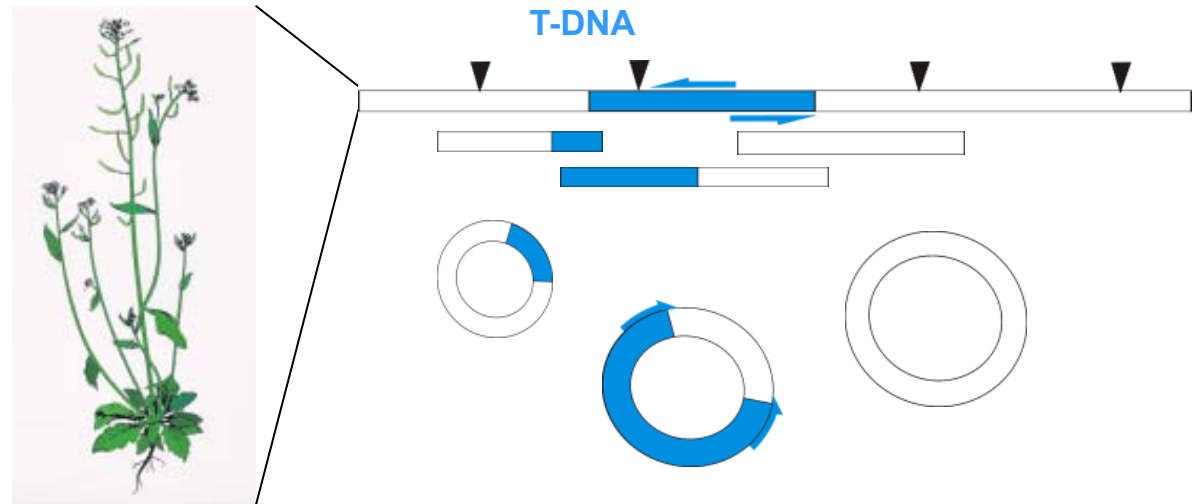
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# Isolation of sequence-specific mutants

- Hybridization with products of iPCR on filters

- Isolation of genomic DNA from the individual plants of mutant population
- Restriction endonuclease cleavage
- Ligation, formation of circular DNA
- Inverse PCR (iPCR) using the T-DNA specific primers
- Preparation of nylon filters with PCR products in the exact position using a robot
- Hybridization with a gene-specific probe



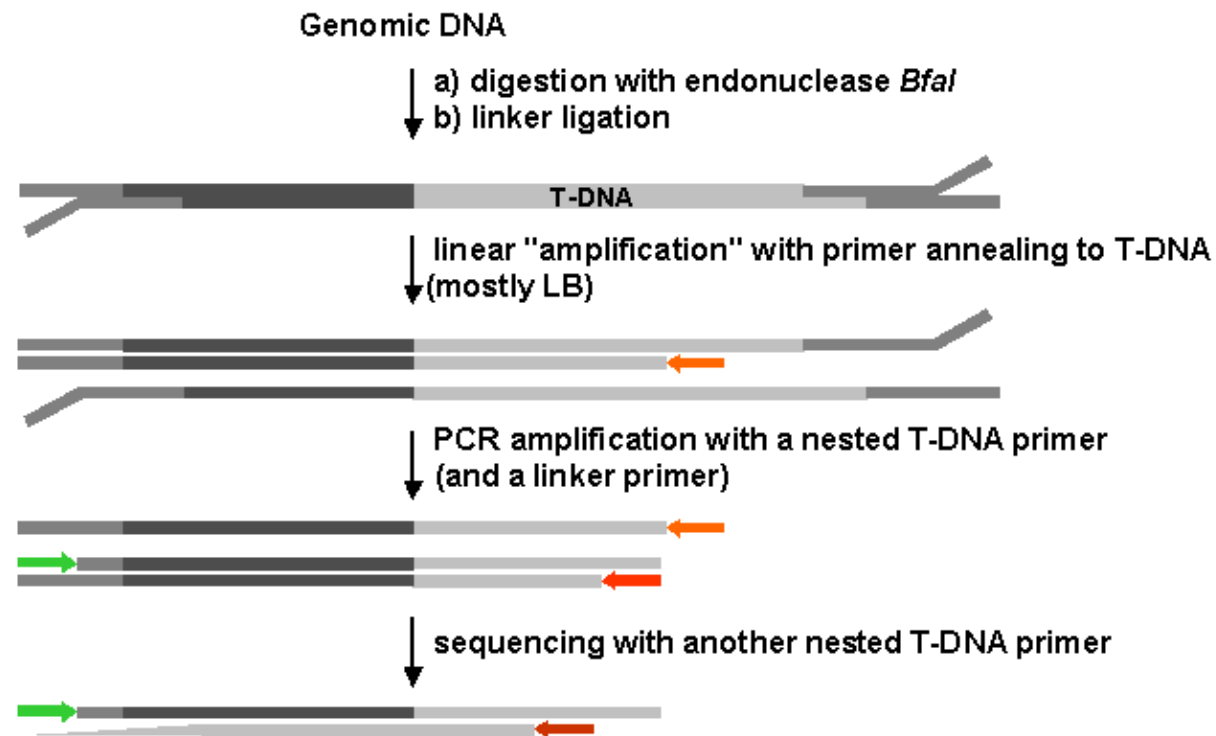
# Outline

- Methods of identification of sequence-specific mutants
  - Preparation of mutants collection
  - Searching for sequence-specific mutants using PCR
  - Searching for sequence-specific mutants in electronic databases

# Isolation of sequence-specific mutants

Preparation of libraries from population of *A. thaliana* mutated by T-DNA

## Sequencing of flanking sequence fragments



# Searching in electronic libraries of insertional mutants

>Insert\_SALK:029311: [Order line 029311](#) | [View in AGR](#)  
Length = 460

Score = 484 bits (244), Expect = e-135  
Identities = 250/252 (99%)  
Strand = Plus / Minus

Query: 1450 attagagtttgattgaagtgtgttttatatattgatagtgaggacattactataaaaaagc 1509  
|||||  
Sbjct: 459 attagagtttgattgaagcgcttttatatattgatagtgaggacattactataaaaaagc 400

Query: 1510 acaaggatacaacaatagagacagtcacatgtatatcacataaaggatggctcctcaatg 1569  
|||||  
Sbjct: 399 acaaggatacaacaatagagacagtcacatgtatatcacataaaggatggctcctcaatg 340

Query: 1570 tgttgctttaggacatttgtgagtatgtcaaaaaacttattcacatggtacactcatag 1629  
|||||  
Sbjct: 339 tgttgctttaggacatttgtgagtatgtcaaaaaacttattcacatggtacactcatag 280

Query: 1630 attagccccacttaggagtgctagaaaaagattgggactaaagtottgttggatcgaat 1689  
|||||  
Sbjct: 279 attagccccacttaggagtgctagaaaaagattgggactaaagtottgttggatcgaat 220

Query: 1690 atgattccaaac 1701  
|||||  
Sbjct: 219 atgattccaaac 208

Score = 111 bits (56), Expect = 8e-23  
Identities = 77/84 (91%)  
Strand = Plus / Plus

Query: 1923 tacattttctcgtacacaattaacgctatcaaatatatttataaaaccatttgcatttcac 1982  
|||||  
Sbjct: 13 tacattttctcgtacagattgacggtatcaaatatatttataaaaccgtagacatttcac 72

Query: 1983 ttcottaactaatcacataaatga 2006  
|||||  
Sbjct: 73 ttcottaactaatcacataaatga 96

Sbjct: 292 ccagcttctagaagcttcttggctcaagtttccagtagcgggacogattctcgagaateaca 233

[AGK insert page](#)

view detailed information on insert sequences in AGR

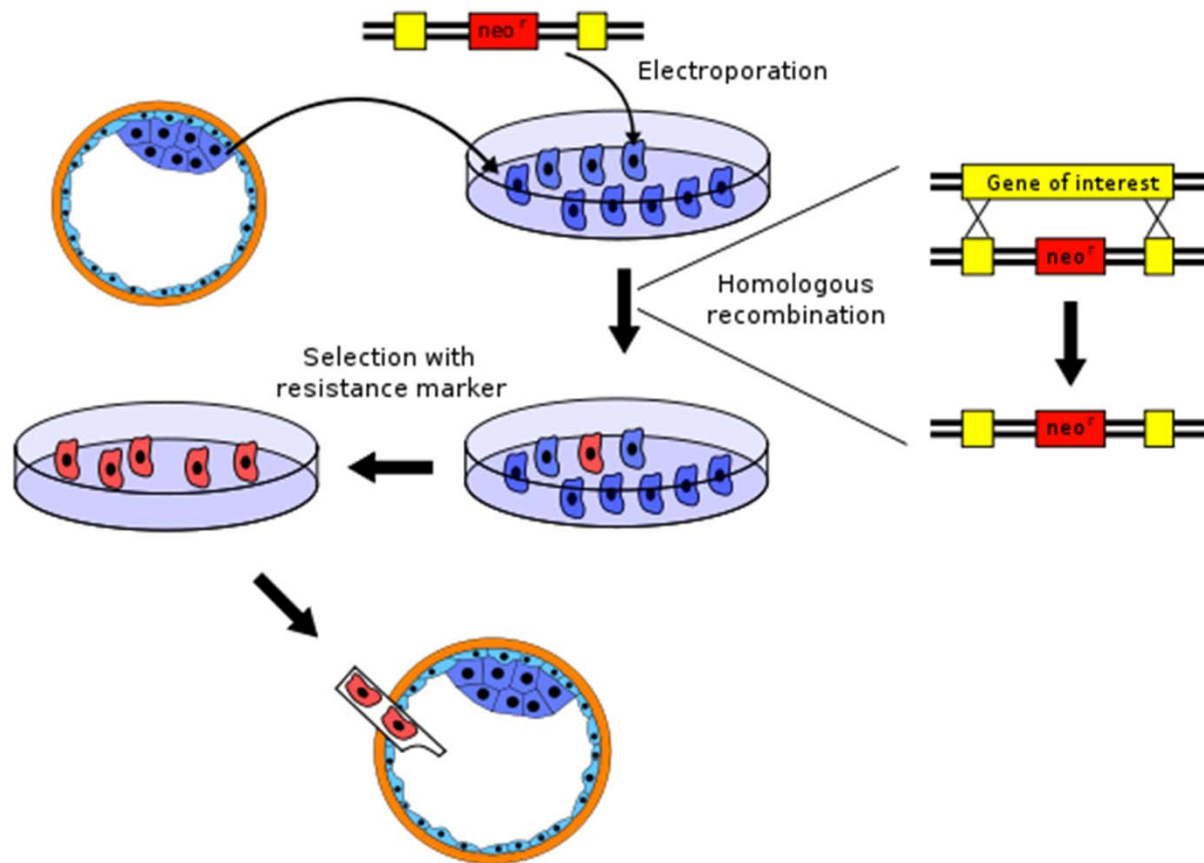




# Outline

- Methods for Identification of Sequence-Specific Mutants
  - Preparation of mutants collection
  - Searching for sequence-specific mutants using PCR
  - Searching for sequence-specific mutants in electronic databases
  - Knocking-out the gene using homologous recombination

# Knocking-Out the Gene





# Outline

- Methods for Identification of Sequence-Specific Mutants
  - Preparation of mutants collection
  - Searching for sequence-specific mutants using PCR
  - Searching for sequence-specific mutants in electronic databases
  - Knocking-out the gene using homologous recombination
- Analysis of Phenotype and Confirmation of Causality Between Phenotype and Insertional Mutation
  - Co-segregation analysis
  - Identification of independent insertional allele
  - Using unstable insertional mutagens and isolation of revertant lines
  - Mutant complementation by the transgene

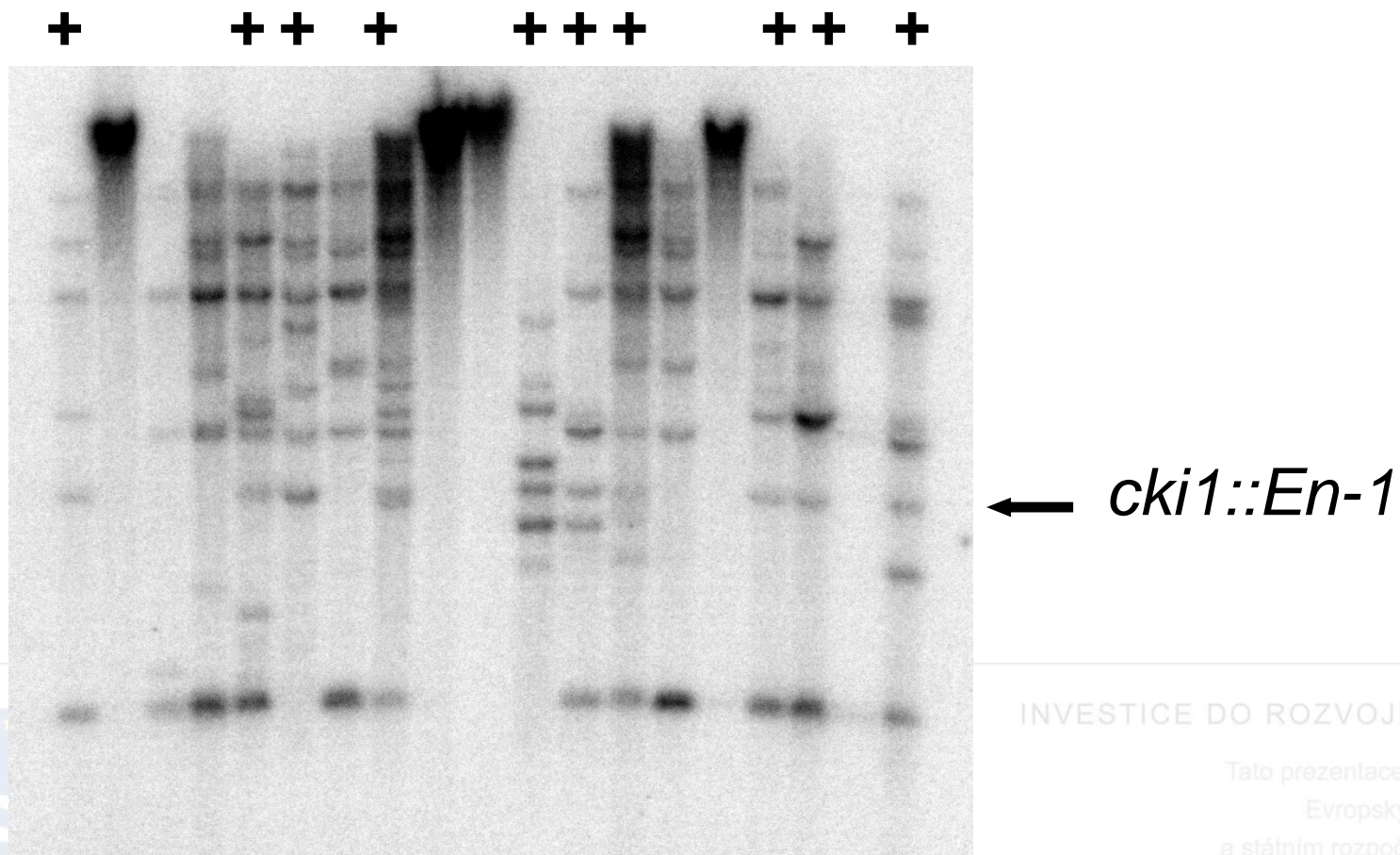
# Why is it necessary to analyze the causality between the insertion and the observed phenotype?

- Presence of **multiple insertions** in one line
- Possibility of **independent point mutation** occurrence
- Insertions of T-DNA are often associated with **chromosomal aberrations** (duplications, inversions, deletions)

# Causality between insertion and phenotype

- **Co-segregation analysis**

- Co-segregation of specific fragment, e.g. after insertion of T-DNA (or exposure to EMS etc.) into the genome of the observed phenotype



# Use of autonomous transposons for the isolation of new stable mutations and of revertant lines

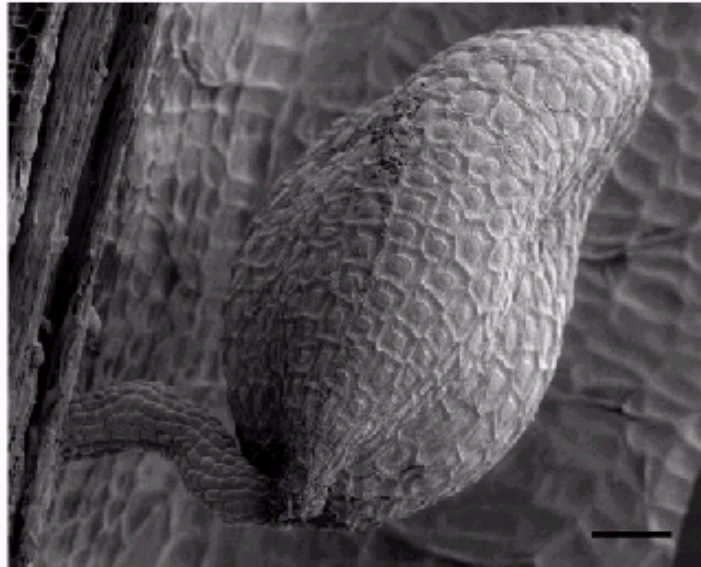
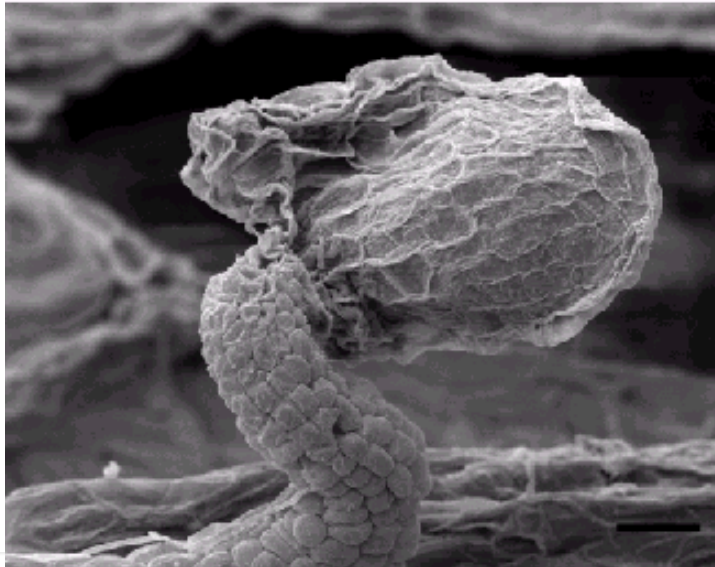
- Transposons are often characterized by excision and reinsertion into a nearby region – use for the isolation of new mutant alleles
- However, excision of transposons is not always entirely accurate – point mutations occur – isolation of revertant lines with silent mutation, or even isolation of the stable mutants

# Phenotype of silicles *cki1::En-1/CKI1*

*cki1::En-1/CKI1*



*CKI1/CKI1*



# Confirmation of phenotype *cki1::En-1/CKI1*

## 1. Isolation of revertant lines

- PCR-searching in 246 plants of segregating population
- from 90 *cki1::En-1* positive plants, 9 plants had both mutant and standard silicles



### Offspring analysis

- confirmation of absence of insertion using PCR
- PCR amplification and cloning the part of the genomic DNA at the insertion site
- sequencing

# Use of autonomous transposons for the isolation of new stable mutations and revertant lines



aattcaagtcgctCACTACAAGA " **En-1** TCTTGTAGTGcgtggagact

- A. aat tca agt **cg** **gga** gac tac act tgg tac act caa acc gtg gat cag tta act ggt  
 N S S **R G D Y** T W Y T Q T V D Q L T G
- B. aat tca agt **gg** **acg** act tgg tac act caa acc gtg gat cag tta act ggt  
 N S S **G T** T W Y T Q T V D Q L T G
- C. aat tca agt cgt **acg** gag act aca ctt ggt aca ctc aaa ccg tgg atc agt taa  
 N S S R T E T T L G T L K P W I S .
- D. aat tca agt cgc **gtg** gag act aca ctt ggt aca ctc aaa ccg tgg atc agt taa  
 N S S R V E T T L G T L K P W I S .

# Confirmation of phenotype *cki1::En-1/CKI1*

## 2. Isolation of a stable mutant line

- analysis of the phenotype of the segregating population (*CKI1/CKI1 CKI1/cki1::En-1*)
- PCR analysis of plants with the mutant phenotype – identification of plants without insertion
- PCR amplification and cloning the part of the genomic DNA at the insertion site
- sequencing



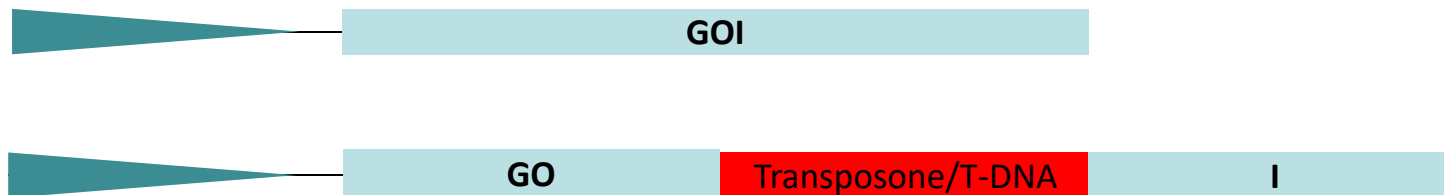
# Use of autonomous transposons for the isolation of new stable mutations and revertant lines



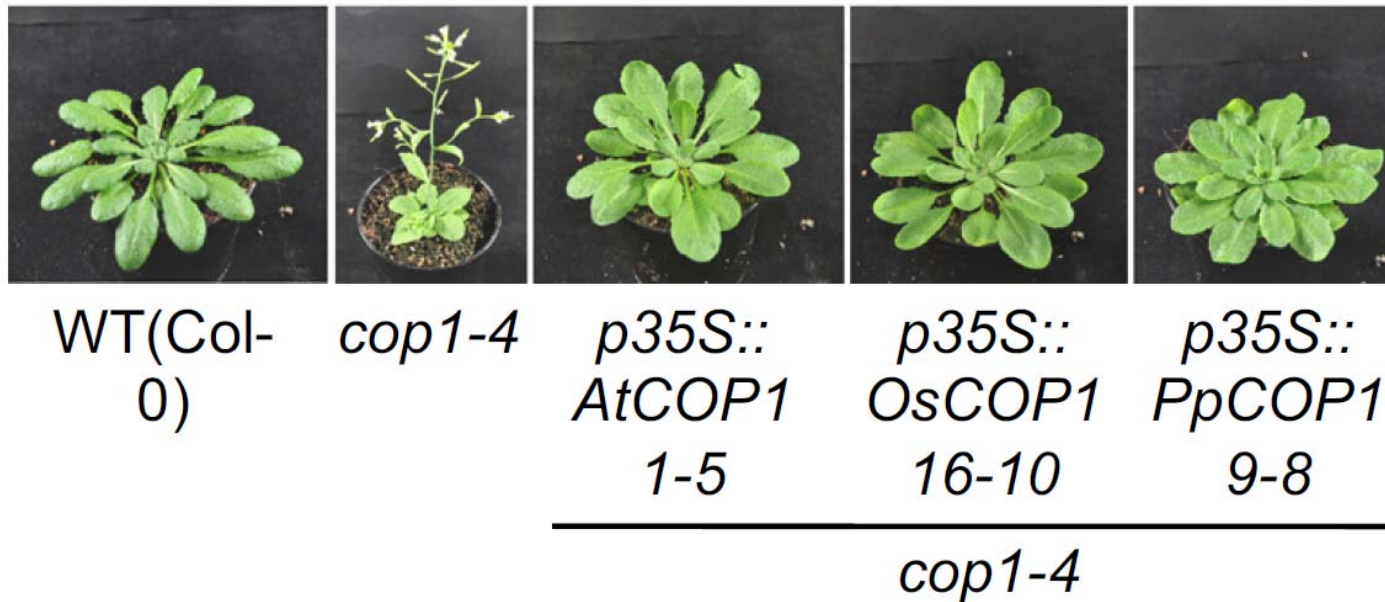
aattcaagtcgctCACTACAAGA " **En-1** TCTTGTAGTGcgtggagact

- A. aat tca agt **cg**t **gga** gac tac act tgg tac act caa acc gtg gat cag tta act ggt  
 N S S **R G D Y** T W Y T Q T V D Q L T G
- B. aat tca agt **gg**t **ac**g act tgg tac act caa acc gtg gat cag tta act ggt  
 N S S **G T** T W Y T Q T V D Q L T G
- C. aat tca agt **cg**t **ac**g gag act aca ctt ggt aca ctc aaa ccg tgg atc agt taa  
 N S S **R T** E T T L G T L K P W I S .
- D. aat tca agt **cg**c **gt**g gag act aca ctt ggt aca ctc aaa ccg tgg atc agt taa  
 N S S **R V** E T T L G T L K P W I S .

# Mutant Line Complementation



# Mutant Line Complementation



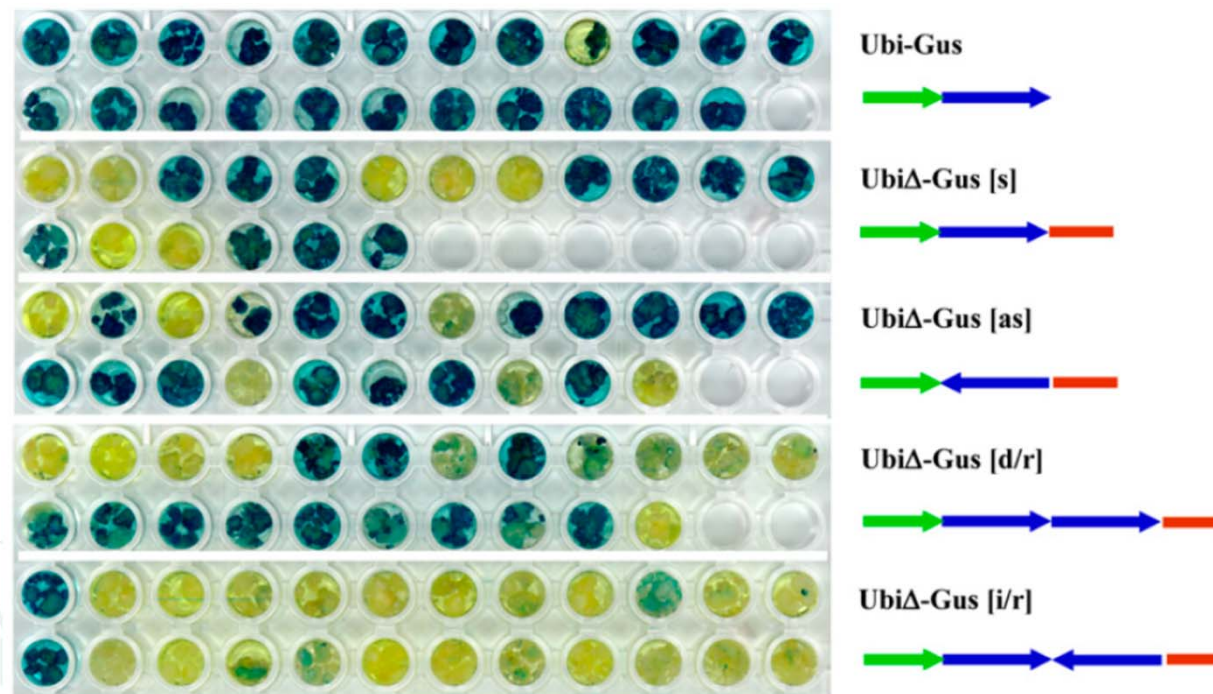
# Outline

- Gene Silencing Using RNA Interference
  - Mechanism of RNA interference

# RNA interference

- **Molecular basis of posttranscriptional gene silencing (PTGS)**

- RNAi found in plants and in *Coenorhabditis elegans*
  - Silencing was induced by both sense and antisense RNA (probably contamination by both during *in vitro* transcription)
  - dsRNA induced silencing about 10-100 times more effectively



Waterhaus et al., PNAS (1998)

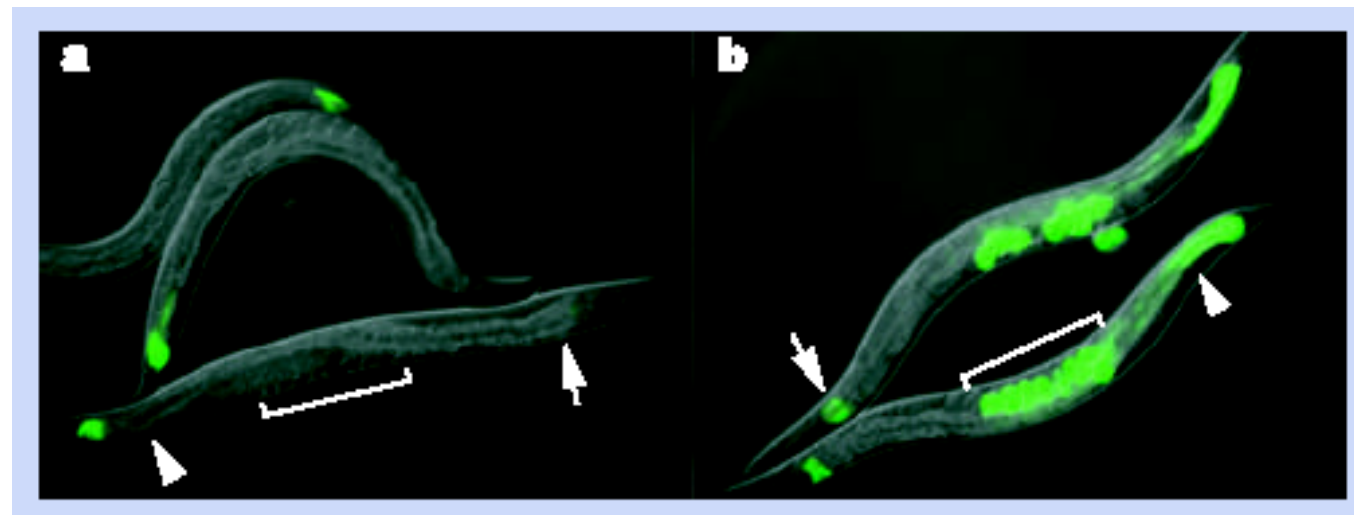
# RNA interference

- **Molecular basis of posttranscriptional gene silencing (PTGS)**

- dsRNA induction is dependent on its own genes – gene searching

*RNAi*

*rnai*

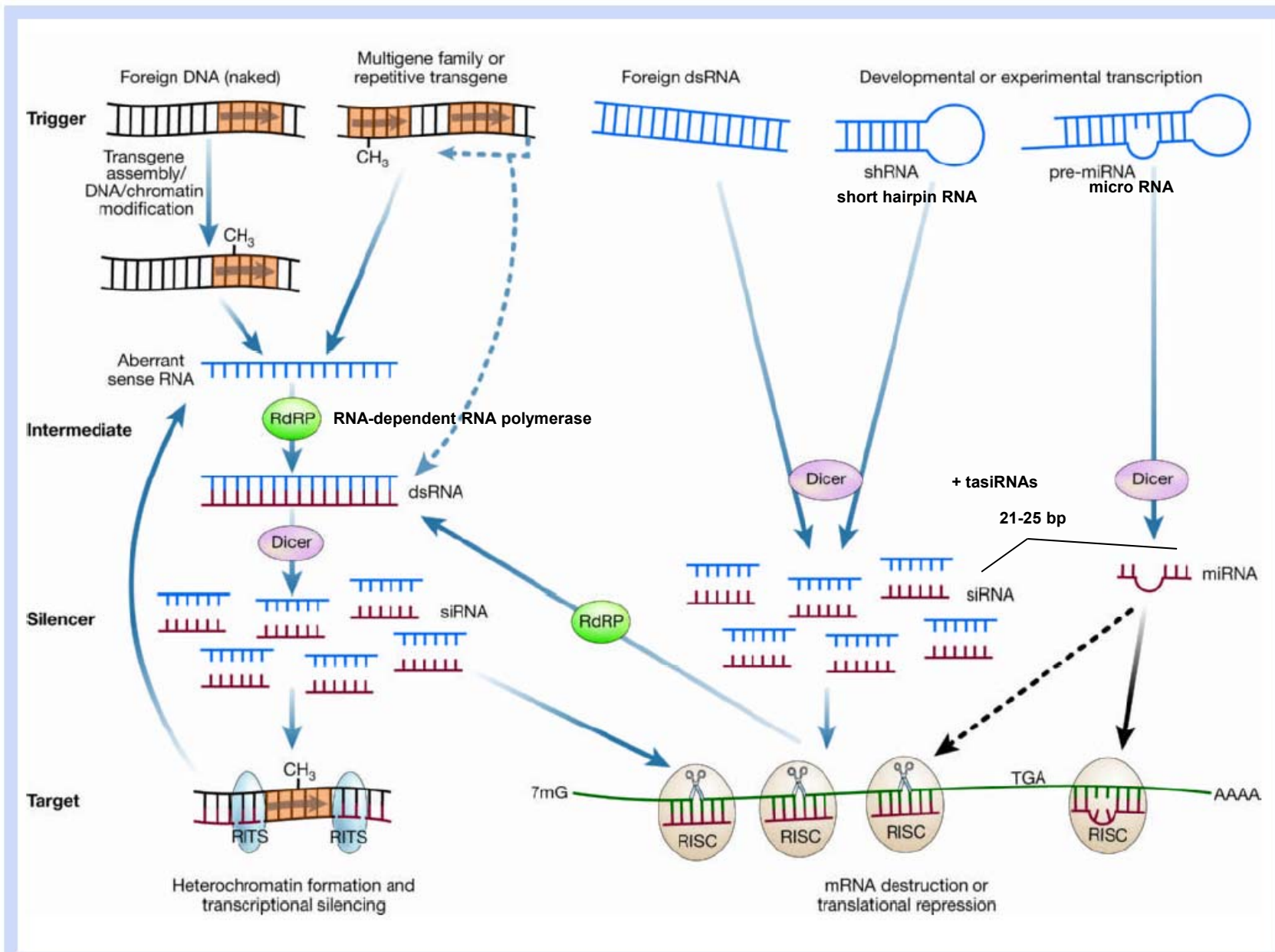


Mello and Conte, *Nature* (2004)

# RNA interference

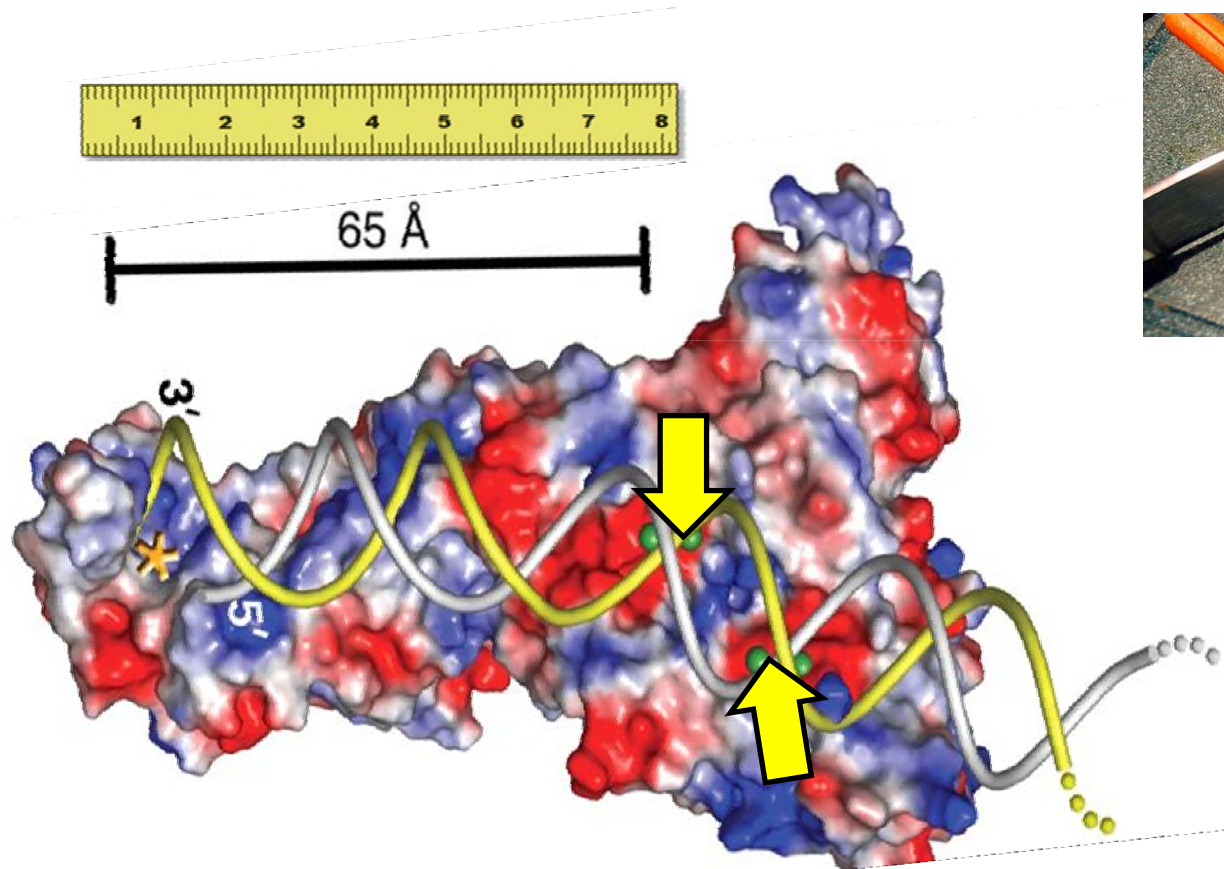
- **Molecular basis of posttranscriptional gene silencing (PTGS)**
  - RNAi found in *Coenorhabditis elegans* and in plants
  - It is a **natural mechanism** of regulation of gene expression in all eukaryotes
  - The principle is **creating dsRNA**, which can be triggered in several ways:
    - By presence of **foreign „aberrant“ DNA**
    - **Specific transgenes** containing **inverted repeats** of the cDNA parts
    - Transcription of own genes for **shRNA** (short hairpin RNA) or **miRNA** (micro RNA, endogenous hairpin RNA)
  - dsRNA is processed by enzyme complex (DICER), which leads to the formation of **siRNA** (short interference RNA), which is then bound to enzyme complex **RITS** (RNA-induced transcriptional silencing complex) or **RISC** (RNA-induced silencing complex)
  - **RISC** mediates either **degradation of mRNA** (in case of full similarity of siRNA and the target mRNA) or leads only to **termination of translation** (in case of incomplete homology, e.g. as in the case of miRNA)
  - **RITS** mediates **reorganization of genomic DNA** (heterochromatin formation and inhibition of transcription)

# Mechanism of RNA interference



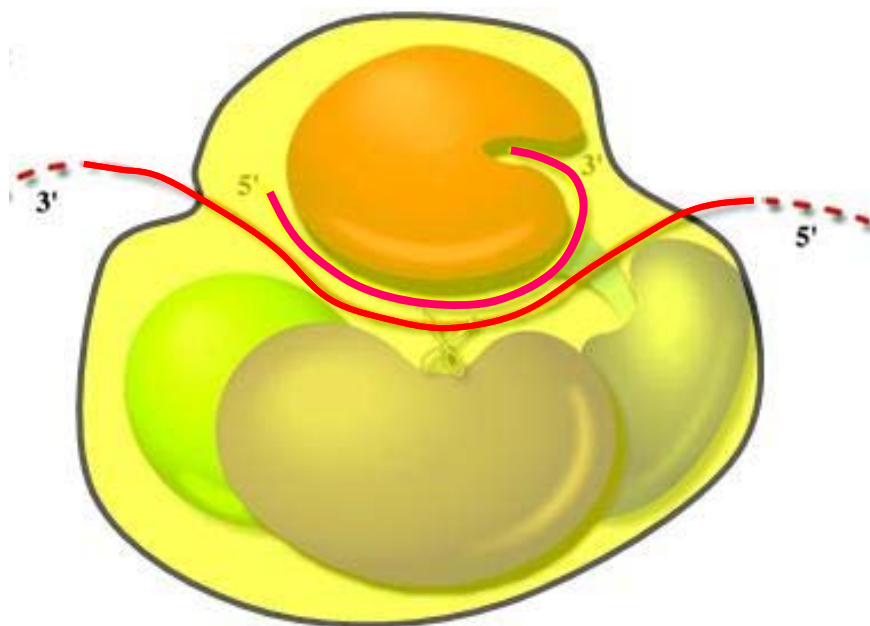


# Dicer and Dicer-like proteins



From MacRae, I.J., Zhou, K., Li, F., Repic, A., Brooks, A.N., Cande, W., Adams, P.D., and Doudna, J.A. (2006) Structural basis for double-stranded RNA processing by Dicer. *Science* 311: [195-198](#). Reprinted with permission from AAAS. Photo credit: [Heidi](#)

# Argonaute proteins



*ago1*



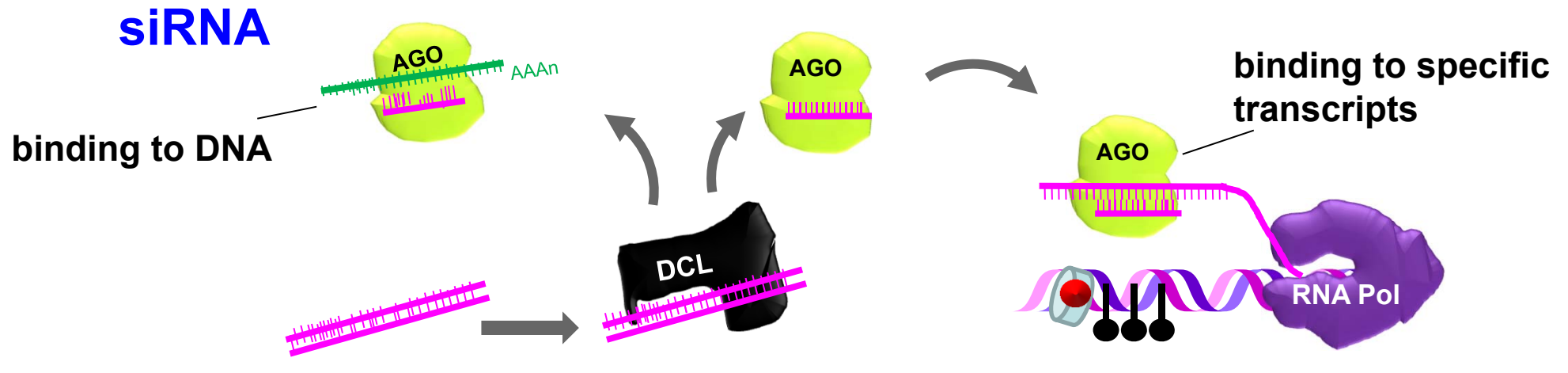
*Argonauta argo*



Reprinted by permission from Macmillan Publishers Ltd: EMBO J. Bohmert, K., Camus, I., Bellini, C., Bouchez, D., Caboche, M., and Benning, C. (1998) *AGO1* defines a novel locus of *Arabidopsis* controlling leaf development. EMBO J. 17: [170–180](#). Copyright 1998; Reprinted from Song, J.-J., Smith, S.K., Hannon, G.J., and Joshua-Tor, L. (2004) Crystal structure of Argonaute and its implications for RISC slicer activity. Science 305: [1434 – 1437](#). with permission of AAAS.

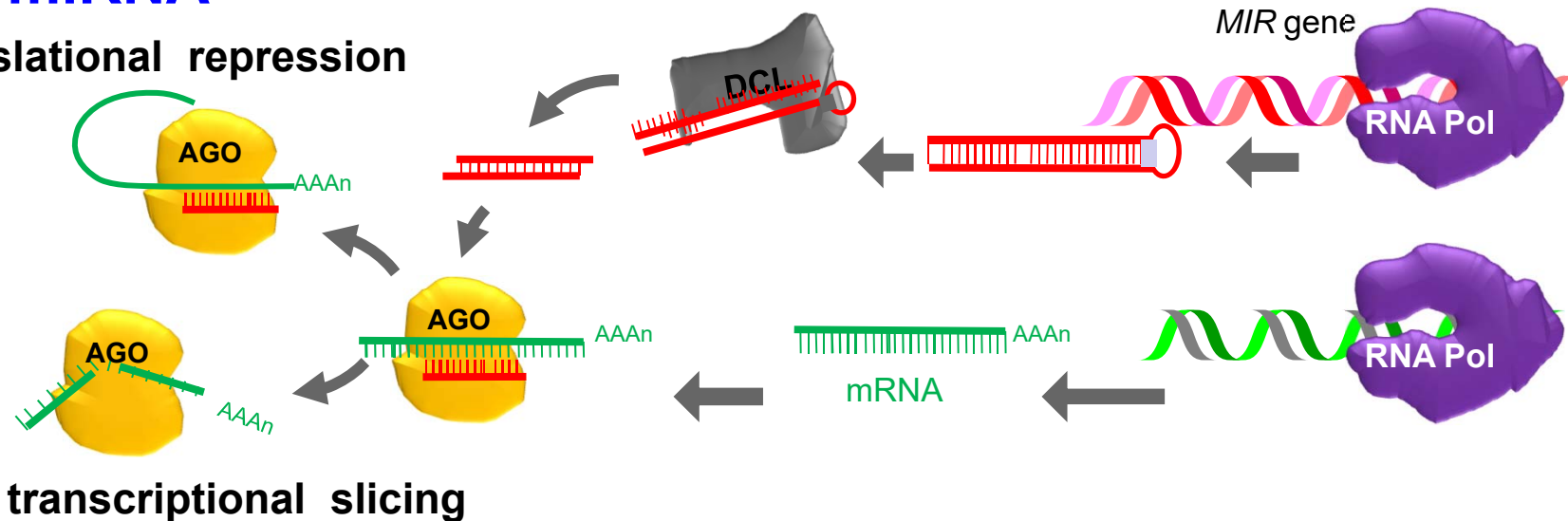
## transcriptional gene silencing

## post-transcriptional gene silencing



## miRNA

### translational repression



# The Nobel Prize in Physiology or Medicine 2006



**Andrew Z. Fire**

USA

Stanford University  
School of Medicine  
Stanford, CA, USA

b. 1959



**Craig C. Mello**

USA

University of  
Massachusetts Medical  
School  
Worcester, MA, USA

b. 1960

# Outline

- Gene Silencing Using RNA Interference
  - Mechanism of RNA interference
- Genome Editing via CRISPR/Cas9



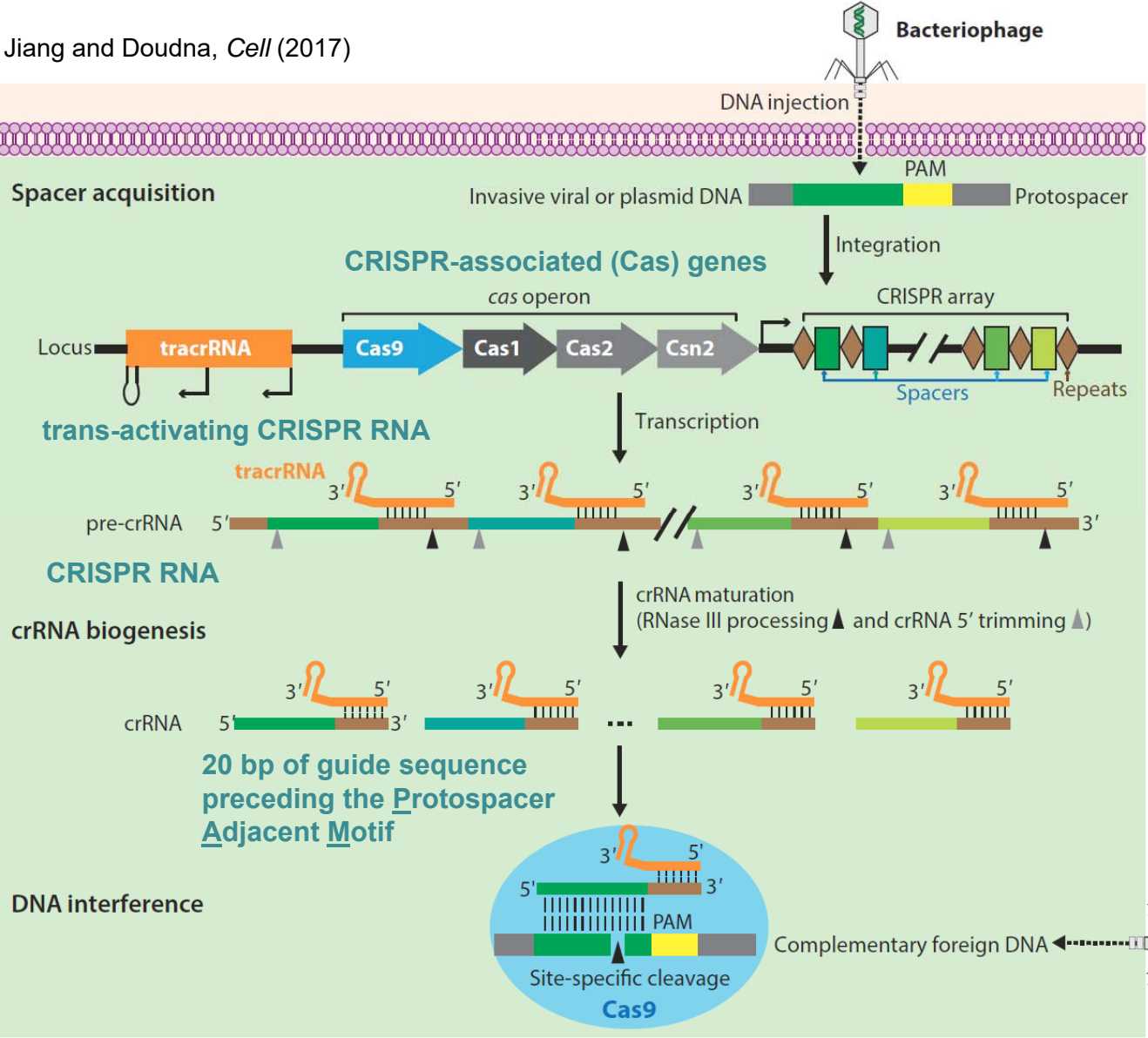
INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ

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Evropským sociálním fondem  
a státním rozpočtem České republiky

# CRISPR/Cas9 - Mechanism

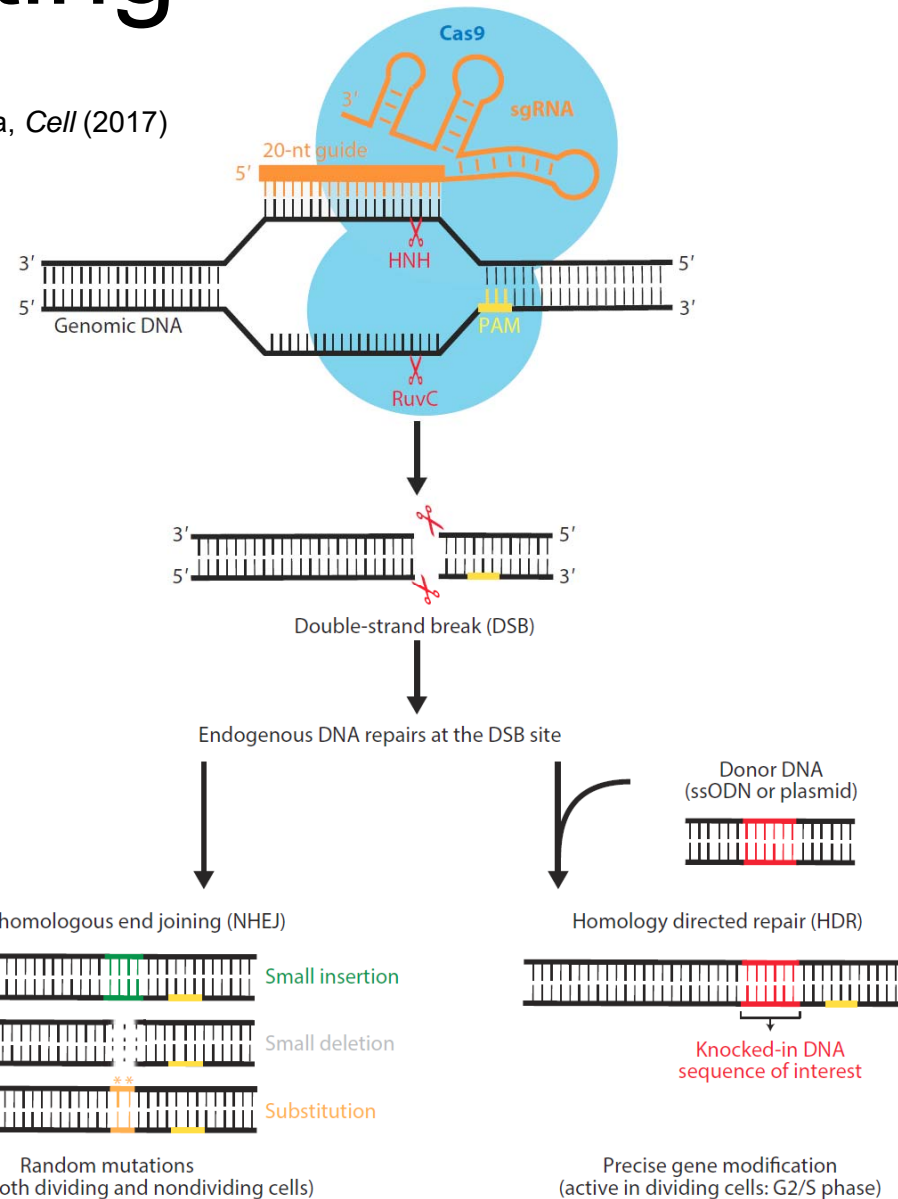
- Clustered Regularly Interspaced Short Palindromic Repeats

Jiang and Doudna, *Cell* (2017)



# CRISPR/Cas9 – Genome Editing

Jiang and Doudna, *Cell* (2017)



# CRISPR/Cas9 – Nobel Prize in 20..19?



Francisco Mojica



Emmanuelle Charpentier



Jenifer Doudna



Martin Jinek

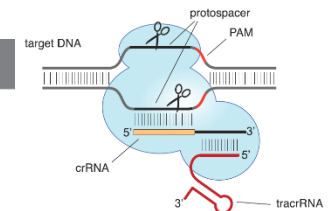
## RESEARCH ARTICLE

### A Programmable Dual-RNA-Guided DNA Endonuclease in Adaptive Bacterial Immunity

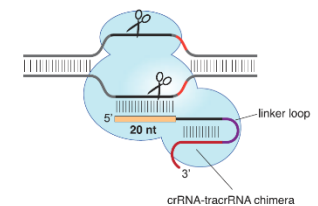
Martin Jinek,<sup>1,2\*</sup> Krzysztof Chylinski,<sup>3,4\*</sup> Ines Fonfara,<sup>4</sup> Michael Hauer,<sup>2,†</sup> Jennifer A. Doudna,<sup>1,2,5,6,‡</sup> Emmanuelle Charpentier<sup>4,‡</sup>

Jinek et al, *Science* (2012)

Cas9 programmed by crRNA:tracrRNA duplex



Cas9 programmed by single chimeric RNA





# Summary

- **Methods for Identification of Sequence-Specific Mutants**
  - Preparation of mutants collection
  - Searching for sequence-specific mutants using PCR
  - Searching for sequence-specific mutants in electronic databases
  - Knocking-out the gene using homologous recombination
- **Analysis of Phenotype and Confirmation of Causality Between Phenotype and Insertional Mutation**
  - Co-segregation analysis
  - Identification of independent insertional allele
  - Using unstable insertional mutagens and isolation of revertant lines
  - Mutant complementation by the transgene

# Summary

- Gene Silencing Using RNA Interference
  - Mechanism of RNA interference
- Genome Editing via CRISPR/Cas9

# Discussion



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Evropským sociálním fondem  
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