



MASARYKOVA UNIVERZITA

Design sekvence PCR primerů

Hana Konečná

CEITEC – MU Centrální laboratoř - Proteomika
Přírodovědecká fakulta NCBR

Tento projekt je spolufinancován Evropským sociálním fondem a státním rozpočtem České republiky.



INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ



SYNTEZICKÉ OLIGONUKLEOTIDY MASARYKOVA UNIVERZITA

OLIGONUKLEOTIDY

PCR primery

- definice
- aplikace
- modifikace
- syntéza

- design sekvence
- zásady navrhování
- software OLIGO 7
- praktická ukázka

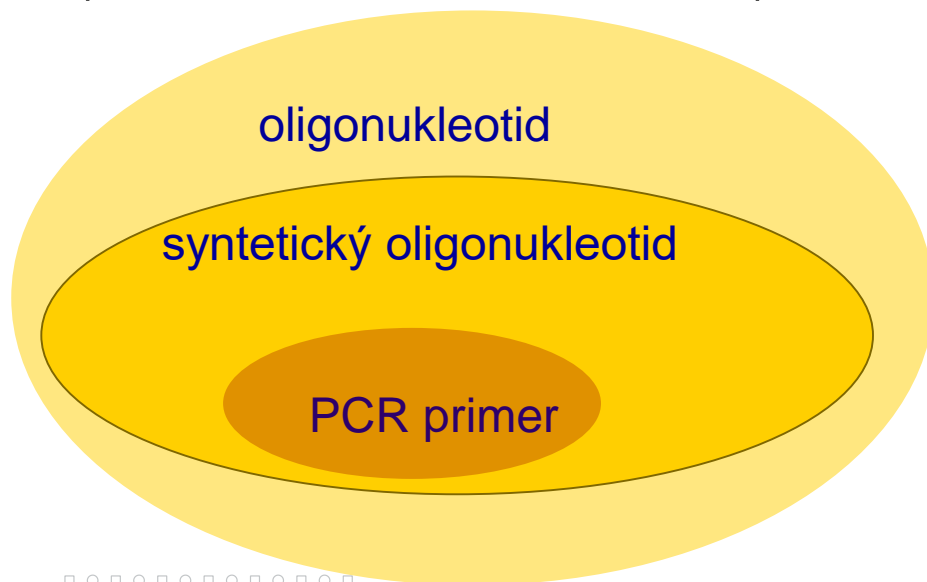
Tento projekt je spolufinancován Evropským sociálním fondem a státním rozpočtem České republiky.



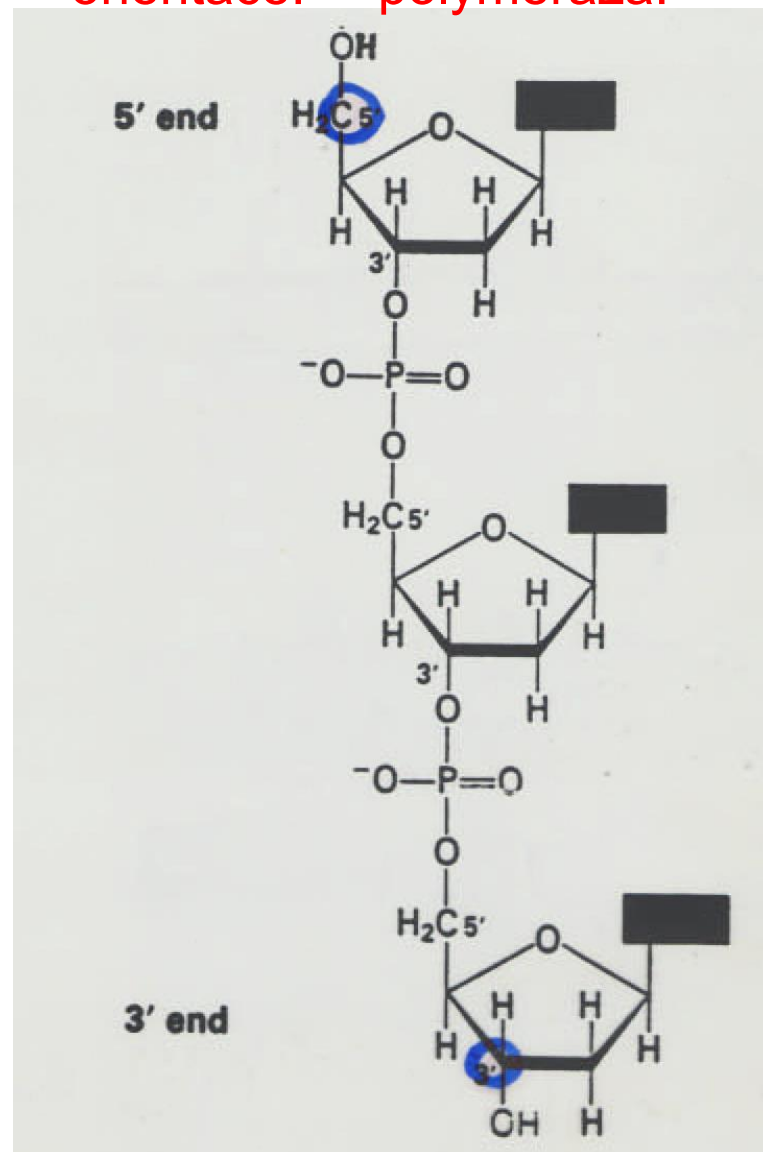
INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ

oligonukleotid

- krátká jednořetězcová struktura
- DNA nebo RNA (event. PNA, LNA...)
- **hydroxyl** na obou koncích (normálně na 5' - konci fosfát)



orientace! polymeráza!

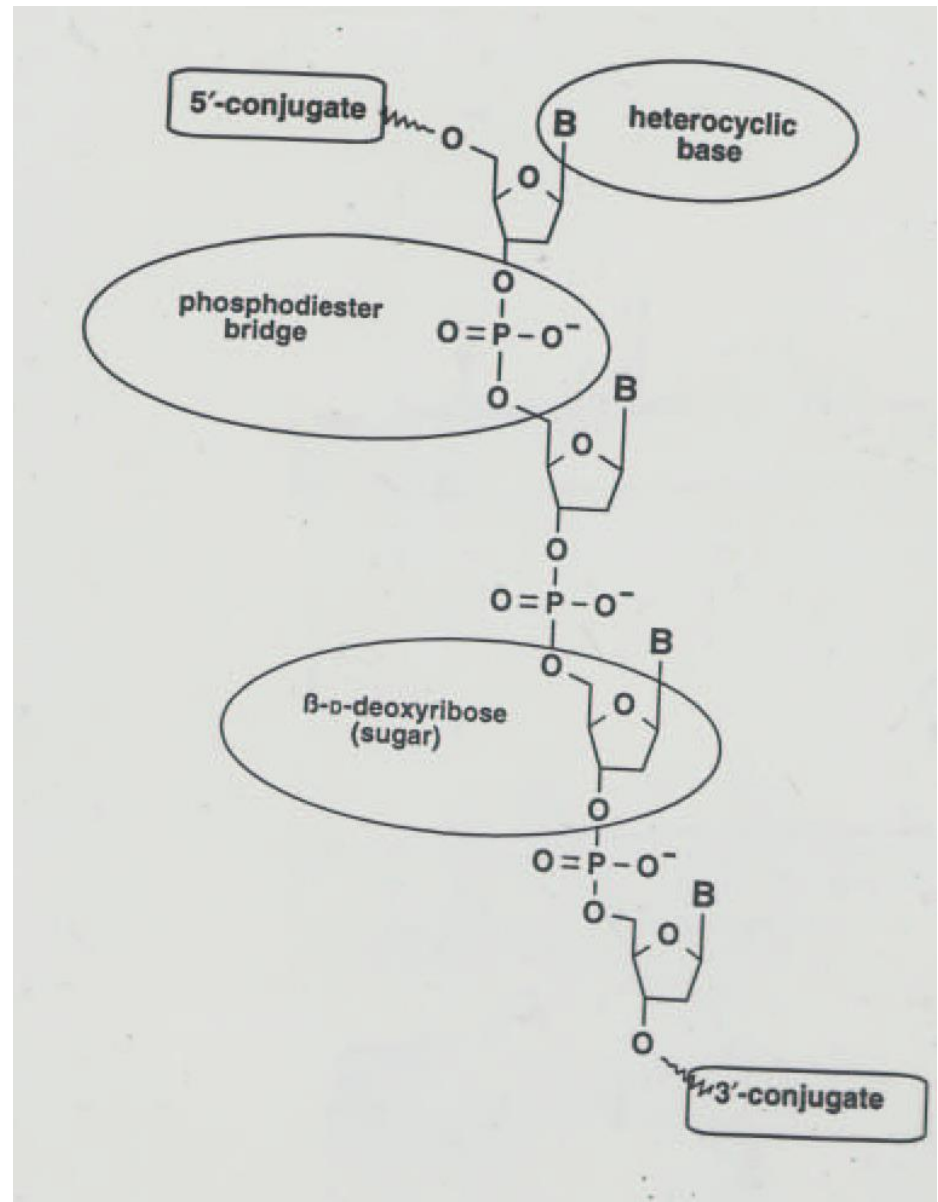


Aplikace syntetických oligonukleotidů

- primery pro syntézu komplementární DNA
PCR, Real-Time PCR
- syntéza genů a rekombinantní proteiny
- hybridizační sondy pro klonování
- místně cílená mutageneza
- sekvenování a genetické profilování
- diagnostika – testy a biosensory
- gene arrays
- blokace genové exprese *antisense oligo*
- potenciální léčiva a DNA vakcíny
- NMR studia interakcí DNA-protein
- strukturální rentgenová analýza NA

Modifikace

- degenerace
- konce řetězce
- báze
- fosfát
- cukr
- PNA



Modifikace na 5' - konci

postsyntetické modifikace →



sekvenování →
fragmentační analýza
gene arrays
Real-Time PCR

5'

fosforylace

aminoskupina

thioskupina

digoxigenin

biotin

enzymy

psoralen

akridin

cholesterol

fluoresc. barviva

zhášedla

2,4-dinitrofenyl

TBR-chelát

spacer

větvení

blokáda



Modifikace na 3'- konci

derivatizovaná matrice



3'

fosfát

thioskupina

aminoskupina

spacer

akridin



biotin



fluoresc.barviva



zhášedla

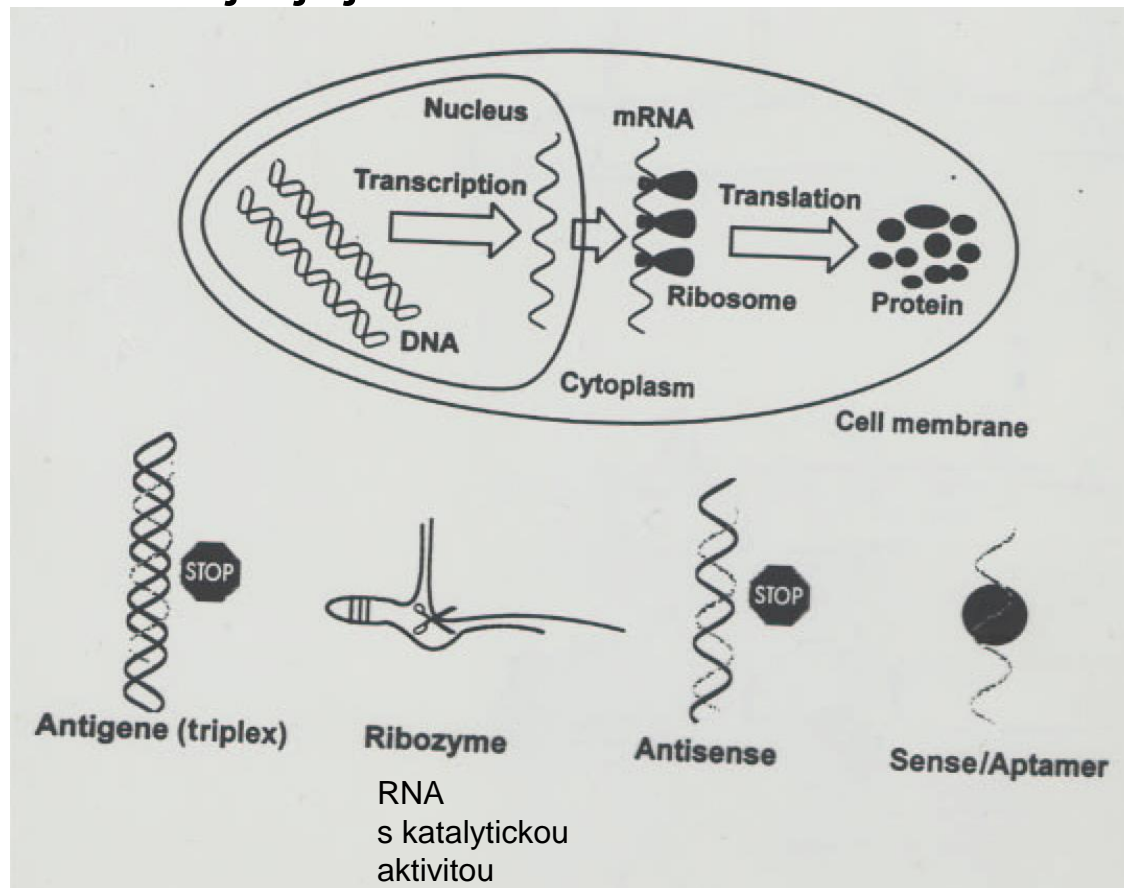
cholesterol

2,4-dinitrofenyl



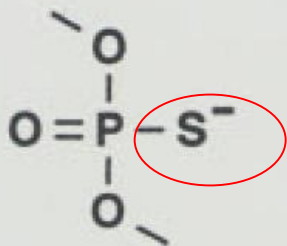
ANTISENSE oligonukleotid

- oligonukleotid nebo analog
- komplementární k segmentu RNA nebo DNA
- vazbou inhibuje jejich normální funkci

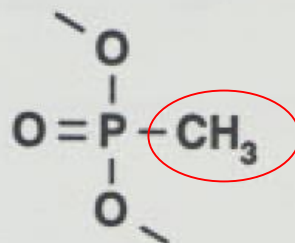


Terapeutika

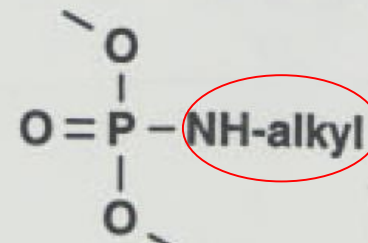
→ nedegradována nukleázami!
modifikace fosfodiesterové vazby



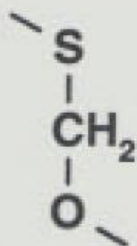
phosphorothioate



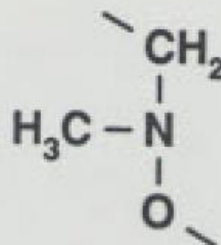
methylphosphonate



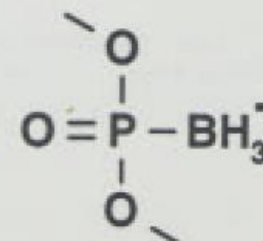
phosphoramidate



3'-thioformacetal



methylene(methyliminio)



boranophosphate

TIDES: Oligonucleotide and Peptide Therapeutics

2018

May 20-23, 2019
Manchester Grand Hyatt,
San Diego, CA

**THE LARGEST MEETING TO ACCELERATE
OLIGONUCLEOTIDE AND PEPTIDE PRODUCTS FROM
EARLY DISCOVERY TO LATE-STAGE DEVELOPMENT &
COMMERCIALIZATION**

- Messenger RNA Therapeutics
- CRISPR-Cas9 and Applications of Genome Editing
- Risk-based Approaches to CMC Development of Therapeutics Oligonucleotides
- Opportunities for Extending to Larger Disease Populations
- Stereochemical Control of Antisense Oligonucleotides Enhances Target Efficacy
- Mechanisms of Action and Quantification of Biodistribution of Oligonucleotides
- Alternative Manufacturing Technologies to Reduce Cost of Goods in Oligonucleotide Manufacturing: Application of Enzymatic Manufacturing
- Late Stage Drug Product Development and Drug Product Manufacturing
- Nonclinical Case Studies in Oligonucleotide Development
- Regulatory Authority Expectations

Degenerované oligonukleotidy

symbol AA nukleotidy

amino acid	amino acid symbol	nucleotide sequence (with degeneracy)	complement (for designing reverse primers)
methionine	M	ATG	TAC
tryptophan	W	TGG	ACC
cysteine	C	TGY	ACR
aspartic acid	D	GAY	CTR
glutamic acid	E	GAR	CTY
phenylalanine	F	TTY	AAR
histidine	H	CAY	GTR
lysine	K	AAR	TTY
asparagine	N	AAY	TTR
glutamine	Q	CAR	GTY
tyrosine	Y	TAY	ATR
isoleucine	I	ATH	TAD
alanine	A	GCN	CGN
glycine	G	GGN	CCN
proline	P	CCN	GGN
threonine	T	ACN	TGN
valine	V	GTN	CAN
leucine	L	YTN	RAN
arginine	R	MGN	KCN
serine	S	WSN	WSN

kód báze

zahrnuje

M	A or C
R	A or G
W	A or T
S	C or G
Y	C or T
K	G or T
V	A or C or G
H	A or C or T
D	A or G or T
B	C or G or T
N	G or A or T or C
X	G or A or T or C

Degenerované oligonukleotidy

CODEHOP

Consensus Degenerate Hybrid
Oligonucleotide Primers

HYDEN

HighLY DEgeNerate primers

2-deoxyinosin

M	A or C
R	A or G
W	A or T
S	C or G
Y	C or T
K	G or T
V	A or C or G
H	A or C or T
D	A or G or T
B	C or G or T
N	G or A or T or C
X	G or A or T or C

Degenerované oligonukleotidy

Příklady:

ACG TAC GTA CGT ACG TAC

nedegenerovaný

ACG T**M** GTA CGT ACG TAC

M = A/C

ACG TAC GTA C**D**T ACG TAC

D = A/G/T

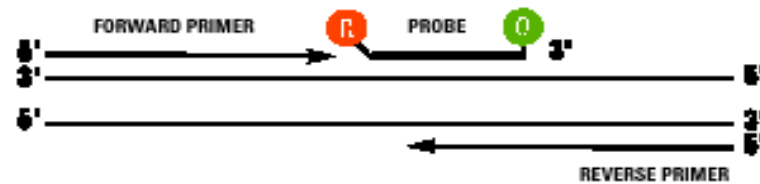
ACG TAC GTA CGT ACG **N**AC

N = A/C/G/T

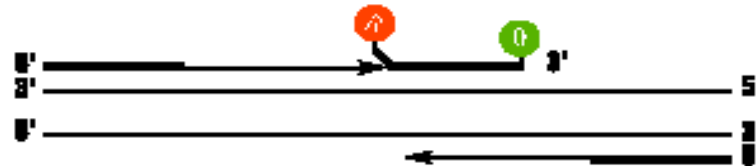


Real-Time PCR

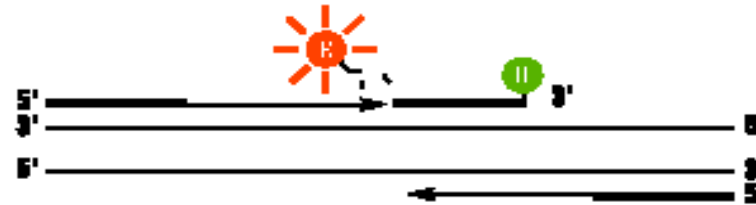
- 2x značená sonda
- REPORTER
- QUENCHER



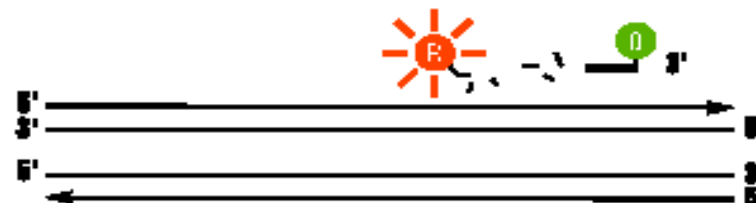
2. **Strand displacement:** When the probe is intact, the reporter dye emission is quenched.



3. **Cleavage:** During each extension cycle, the DNA polymerase cleaves the reporter dye from the probe.



4. **Polymerization completed:** Once separated from the quencher, the reporter dye emits its characteristic fluorescence.

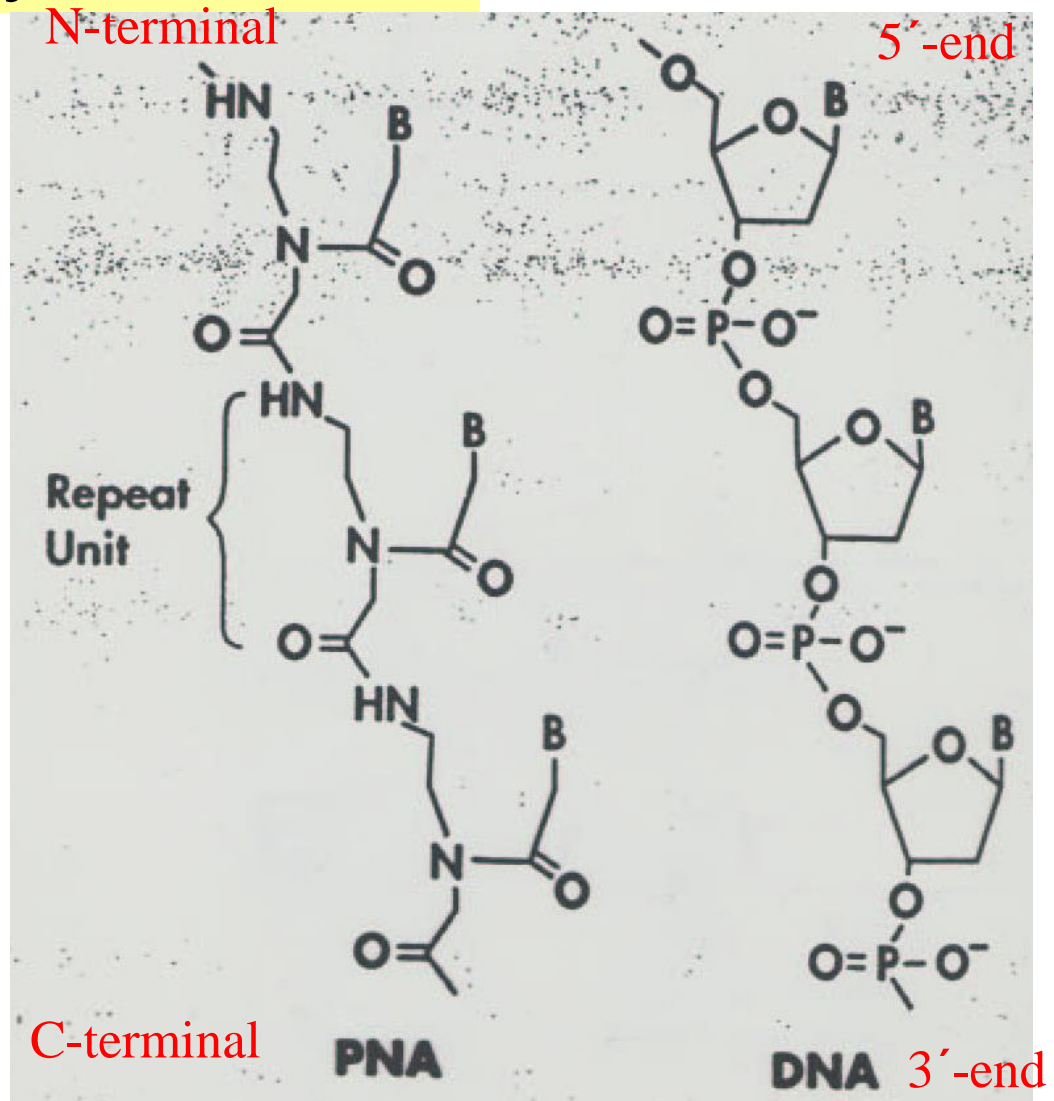


Peptidonukleová kyselina **PNA**

DNA

- nenabitá molekula
- vazba k DNA/RNA

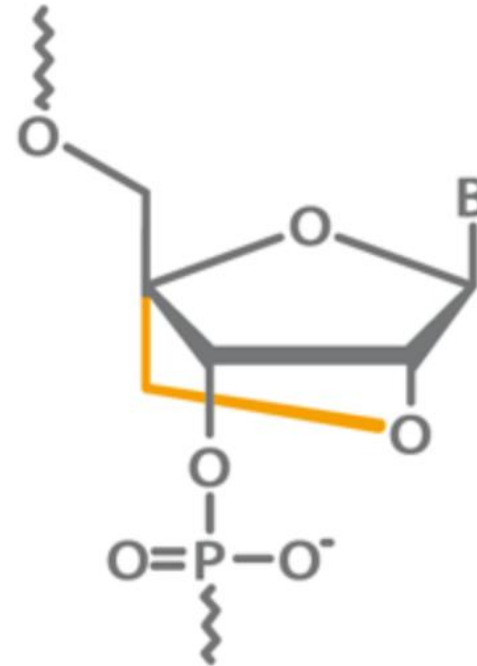
N-(2-aminoethyl)-glycin →



LNA

Locked Nucleic Acid

2'-O, 4'-C methylenový můstek
potlačená flexibilita ribofuranózového kruhu
struktura je **zamčena** do rigidní C3-endo konformace
zlepšená hybridizace
výjimečná biostabilita



OLIGONUKLEOTIDY

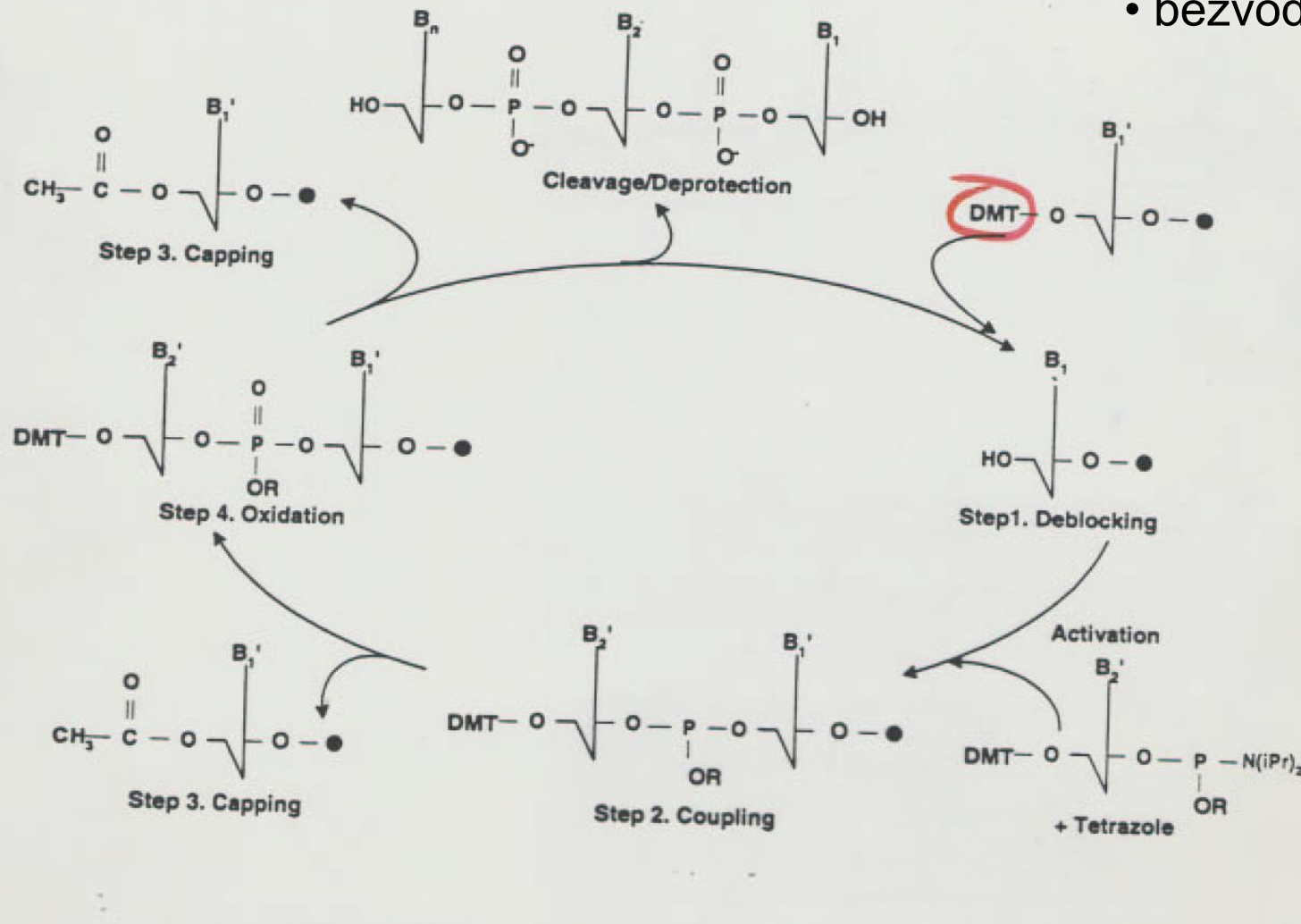
- organická syntéza
na pevné fázi **žádný enzym!**
- od 3'- konce k 5'- konci
- bezvodé prostředí



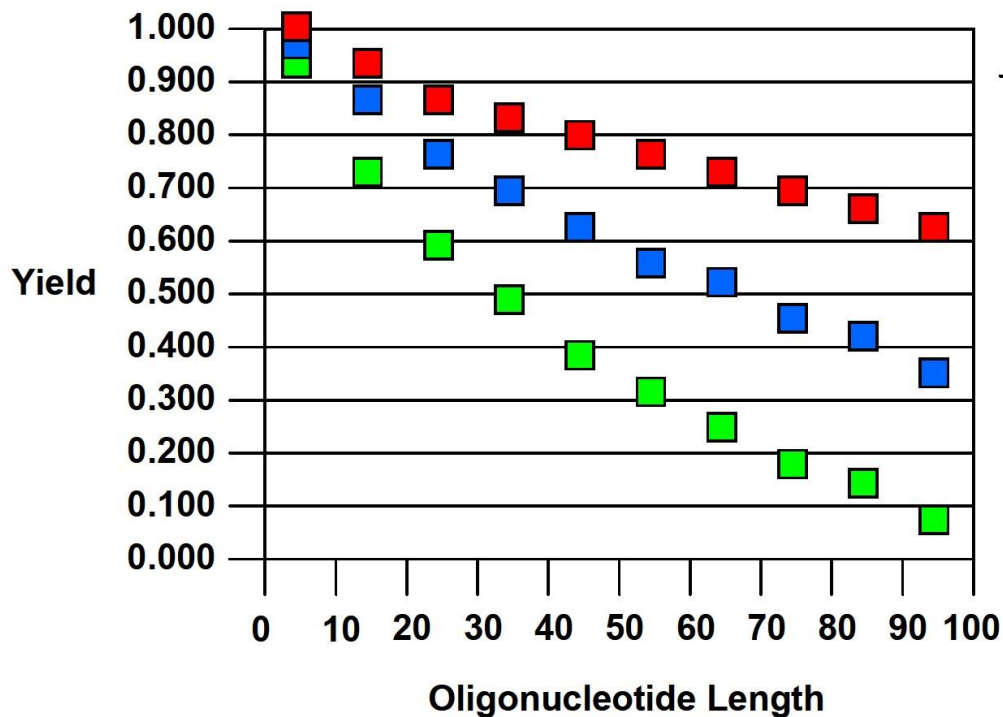
EXPEDITE 8909

Syntéza oligonukleotidu

- syntéza na pevné fázi
- od 3'-konce k 5'-konci
- bezvodé prostředí



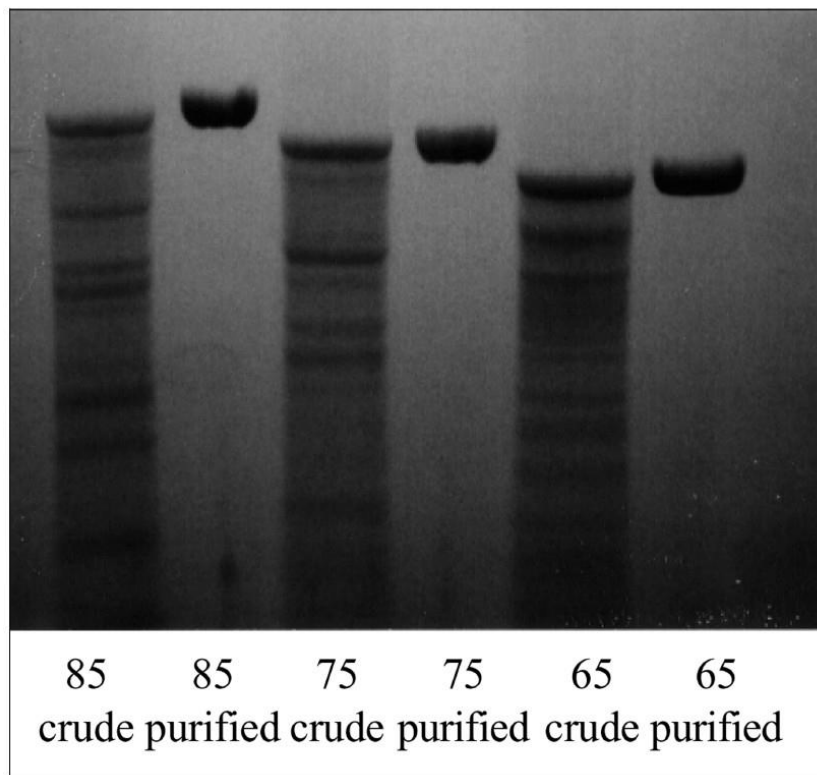
VÝTĚŽEK



Efficiency

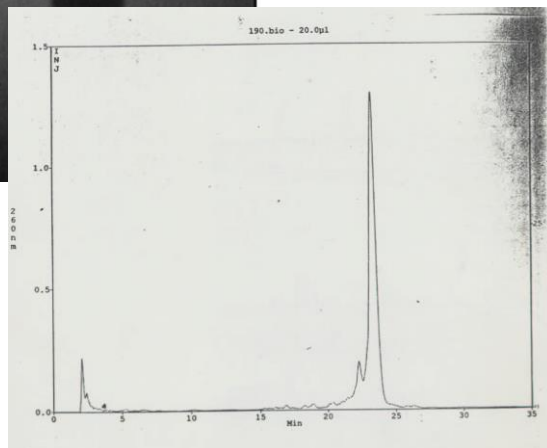
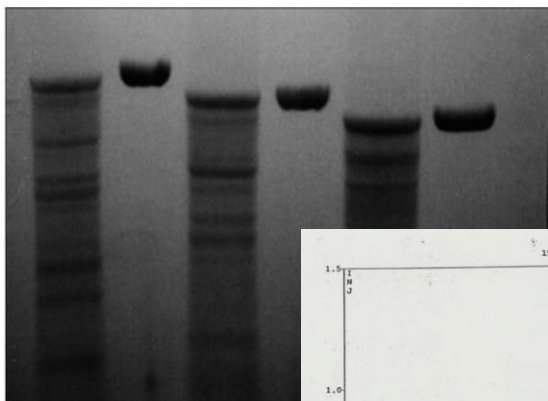
- 0.995
- 0.990
- 0.980

PAGE



PURIFIKACE a QC

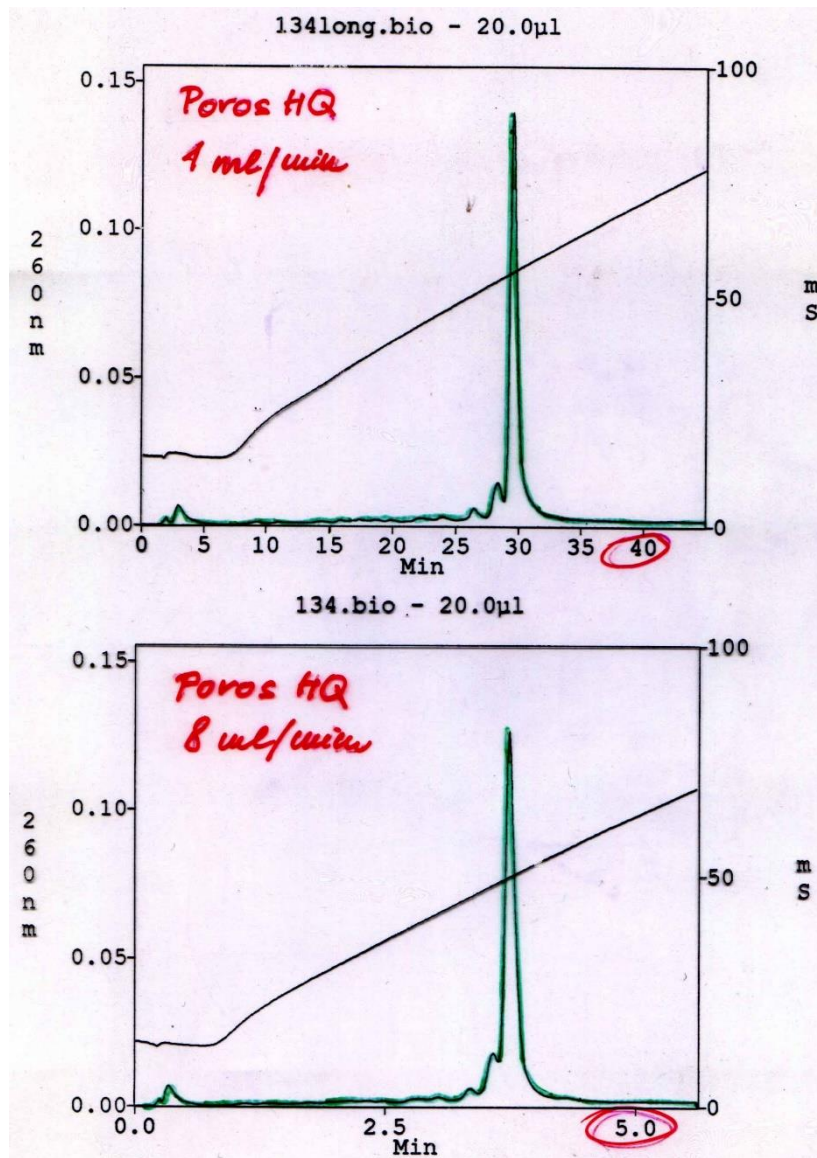
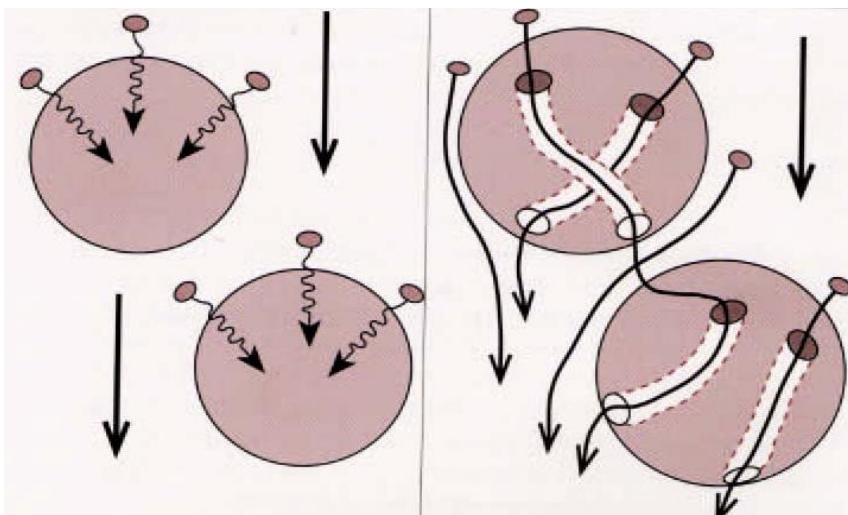
- Sephadex minispin kolonka
- RP cartridge
- HPLC
- Gelová elektroforéza



Perfúzní chromatografie

klasický HPLC sorbent

POROS



Jaký typ purifikace vybrat?

- aplikace oligo
- délka oligo
- modifikace oligo
- výtěžek oligo

PCR nebo sekvenování

standardní odsolení

Klonování . Mutageneza . Gel shift *

HPLC . PAGE

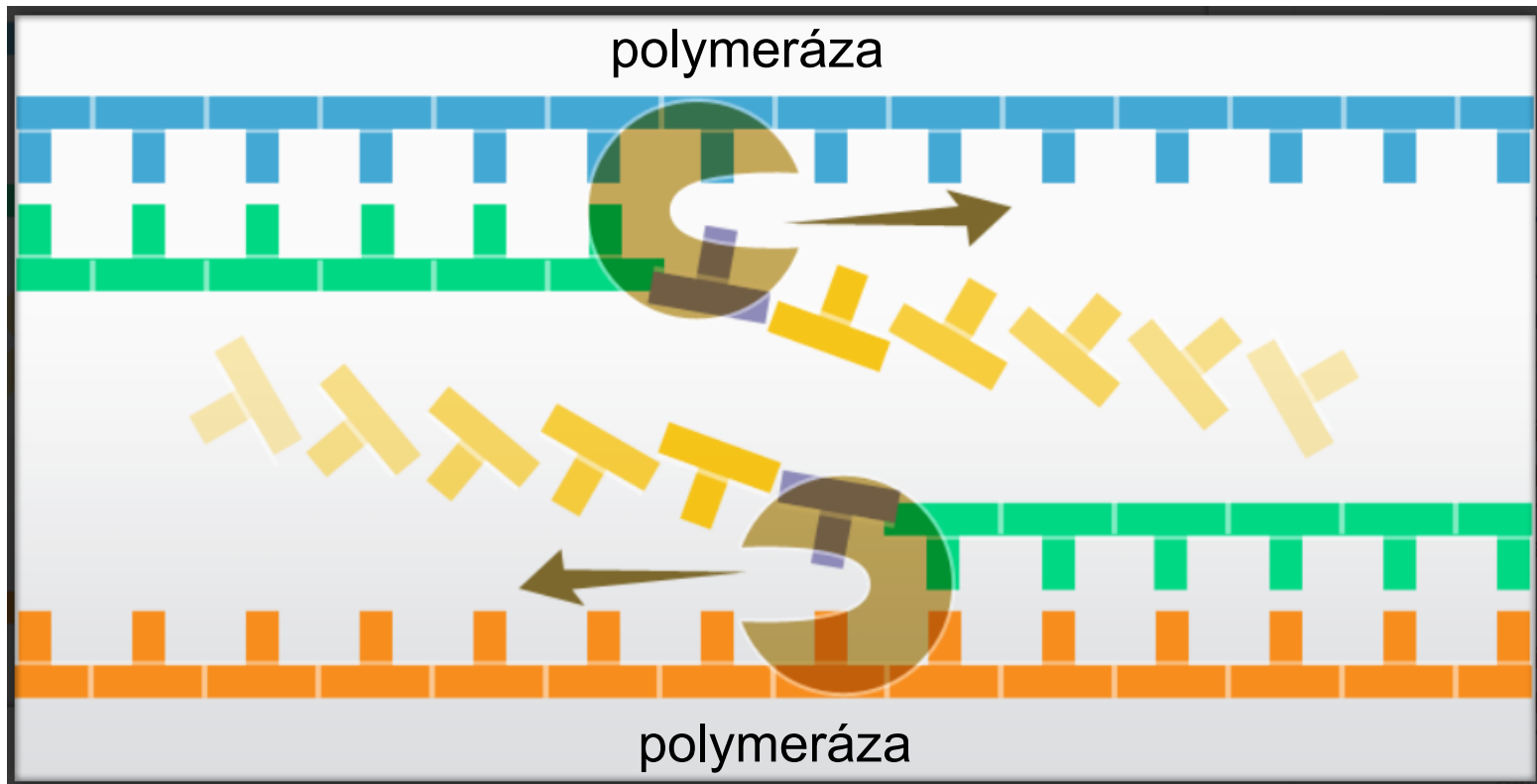
Modifikované oligonukleotidy

~~HPLC . PAGE~~

* affinity electrophoresis for protein-DNA or protein-RNA interactions

PCR primery

*Methods in Enzymology 529 (2013)
Laboratory Methods in Enzymology:
DNA Explanatory Chapter: PCR Primer Design*



<http://www.genomecompiler.com/tips-for-efficient-primer-design/>

DESIGN OLIGONUKLEOTIDU

- manuální
- počítačový

www.protocol-online.org/prot/Research_Tools/Online_Tools/PCR_Primer_and_Oligo_Design_/index.html

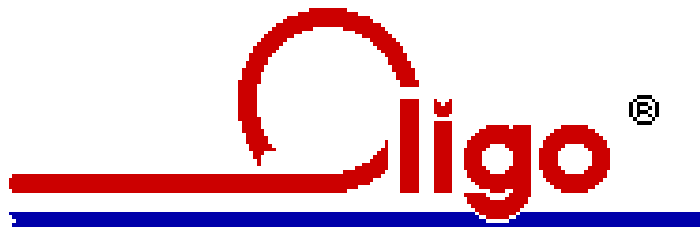
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1526718/pdf/1746-4811-2-4.pdf>

<https://academicjournals.org/journal/AJB/article-full-text-pdf/D84CFC59144>

Hlavní kritéria pro sekvenci PCR primeru

- vysoce specifické – zejména 3´konec
- netvoří dimery a vlásenky
- stabilní duplexy s aktivní sekvencí
- nepřiliš stabilní 3´-konec
- obvykle 18 – 25 nt
- 40% až 60% GC

www.genomecompiler.com/tips-for-efficient-primer-design/



OLIGO 6

- PCR primery,
- hybridizační sondy
- sekvenační primery

OLIGO 7 (od roku 2008)

- TaqMan sondy
- primery pro *nested PCR*
- *molecular beacons*
- siRNA

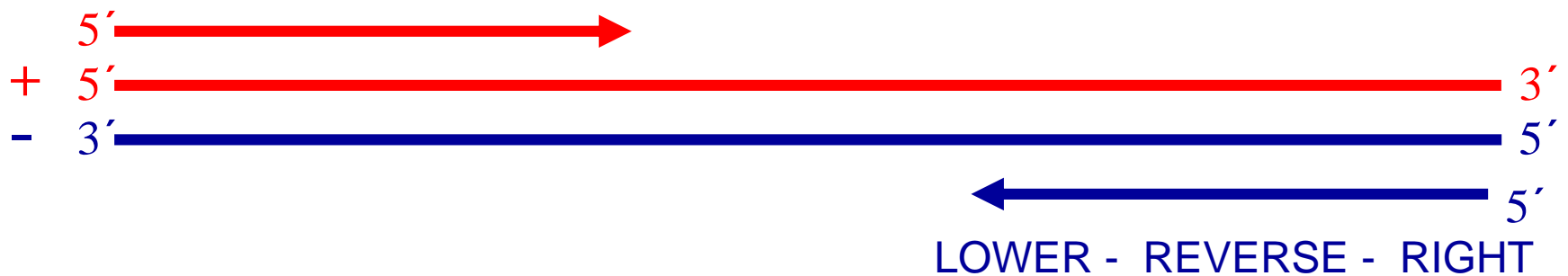
Terminologie PCR primerů

forward primer... část sekvence + vlákná

reverse primer... část sekvence - vlákná



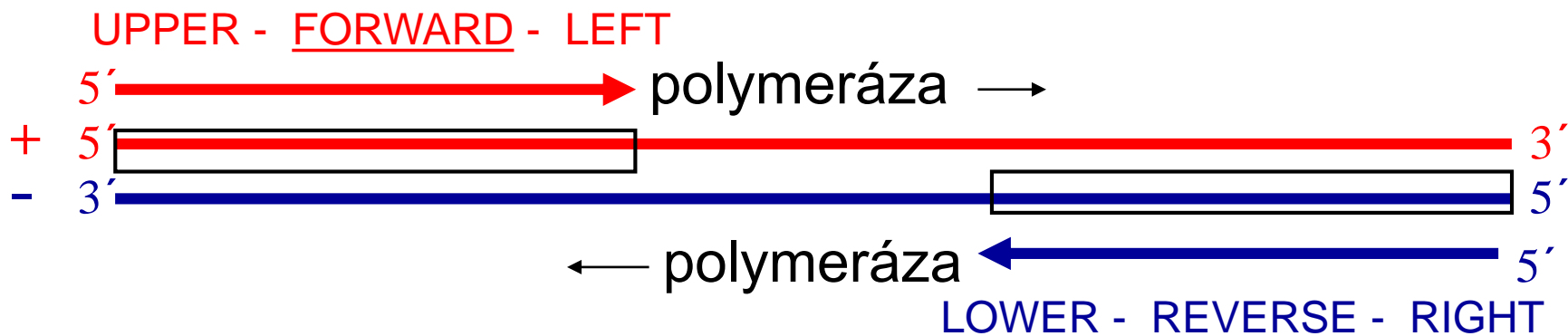
UPPER - FORWARD - LEFT



Terminologie

forward primer... část sekvence + vlákna

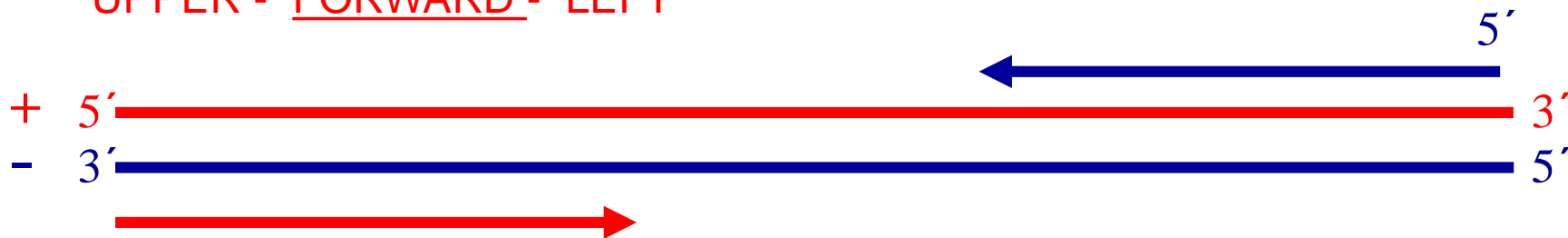
reverse primer... část sekvence - vlákna



Nasedání PCR primerů



UPPER - FORWARD - LEFT



LOWER - REVERSE - RIGHT



5' CTT CTG CTC AAT CTT TCT AC 3' FORWARD

+5'

1 ATGGCTTCTG CTCAATCTTT CTACAAACCAAGCTCTGTCT TGAAAATCAA
 51 TGTCATGGTT GGGACGAIG ATCATGTTTT CCTTGATATC ATGTCACGCA
 101 TGCTTCAACA CTCCAAATAC AGAGGTAATT AAATATTATT ATCATATTAT
 151 ATATAATATG TTATTGATTT TTTGTTTGTG ATTTCATTTA GATTTTTATT
 201 TCTATGATTT CTTAGCATGA AATACAATTT TTGGAGAAAC AACTAGCAGT
 251 TTTAAAAACA AAACCTGAAT TTTGAGAAAT TCAAAGATGT TATATATATA
 301 TGTCAAAATT TAACAATTAT TCTTCTAAAT CATCCGGATT CCGTTTACAT
 351 GTACACATCT ACAATTTTCA ATTGAGGTAT TCTTGTTTTG ATGCCTTTGA
 401 GACGAATAGT TTGATTGATA AAAAAAATTC TAACCAATAT GATATATAAA
 451 GTTTTTTTTT TTTTGTCAA ACCACTTTT ATACTATGTA ACTTTTTTAA
 501 GAGATTATTG AAAATAGTTT ATTTATAAAA TAGTAACCTA TTGTTGAATT
 551 AAAAAAAAAA AAAAAATTGT AAATCGTGTT TGCAAACGAC ATGTGATTTA
 601 TCTTAGTTTA AACTAGCTG ATATTCTTCA AATCGACTGT TCTTATAAGT
 651 AATCAACCAA TAGCATCAA TCACAATAAA TTGTAACAC TTCAATGAAA
 701 ATGGTGATTT TAAAGAATAT GTTTTACTTA TGTTATGAAC TATCTCAAAT
 751 TTGTGAAATA TTTCATAACT AATGTGGAAA ACTATATAAC CCCTCCATAC
 801 AAAACGTAAG TAAAATTTAT GAAATCCTAT CATTTTTAAA GGTTAAACCA
 851 ATCAAAAAGT AATAATTCTT GGTACTTGCA ATATTTTTGT CATTATATT
 901 TAGTTTATTA ATTTATTTT GATTAAATGG TTTTAGATCC ATCAGTTATG
 951 GAGATCGCAG TTATAGCTGT AGACGATCCG AAGAAAGCAT TATCTACTCT
 1001 AAAAATTCOA CGAGACAATA TAGATCTCAT AATCACAGAT TATTATATGC
 1051 CTGGTATGAA CGGTTTACAA CTCAAAAAAC AAATCACTCA GGAATTTGGA
 1101 AATTTACCGG TCTTAGGTAA CATTTTTTGT TCTTTACAAC TTAAATTA

3'

5' TGA AGA ATA TCA GCT AGT TT 3' REVERSE

File: Human 4E.seq
Sequence

DNA Sequence		Selected Oligo	Position	Length	#	Feature	Location
Sequence Length:	1868 nt	<input checked="" type="checkbox"/> Forward Primer	259	18	1	source	-18..1850
Reading Frame:	+1	<input checked="" type="checkbox"/> Reverse Primer	328	18	2	CDS	1..651
Current Oligo Length:	21 nt	<input type="checkbox"/> Upper Oligo	---	---			
Position:	356	<input checked="" type="checkbox"/> Lower Oligo	294	22			
t_m :	59.3°C	<input checked="" type="checkbox"/> PCR Product	87 nt				

pos:
tm:

260	270	280	290	300	310	320	330	340	350	360	370																														
CCTGGCTGTGACTACTCA >																																									
TTAATGCCTGGCTGTGACTACTCACTTTTAAAGGATGGTATTGAGCCTATGTGGGAAGATGAGAAAAACAACGGGGAGGACGATGGCTAATTACATTGAACAAACAGCAGAGACGAAGTGACCTC																																									
AATTACGGACCGACTGATGAGTGAAAAATTCCTACCATAACTCGGATACACCCCTTCTACTCTTTTTGTTGCCCTCCTGCTACCGATTAATGTAACCTGTTGTGCTCTCTGCTTCACTGGAG																																									
< ACTCGGATACACCCCTTCTACTC																																									
< CCTCCTGCTACCGATTAA																																									
L	M	P	G	C	D	Y	S	L	F	K	D	G	I	E	P	M	W	E	D	E	K	N	K	R	G	G	R	W	L	I	T	L	N	K	Q	Q	R	R	S	D	L



Search for Primers & Probes

Search Options Subsearches

Search in: + Strand - Strand
Search Mode: Select Verify

Complex Substrate

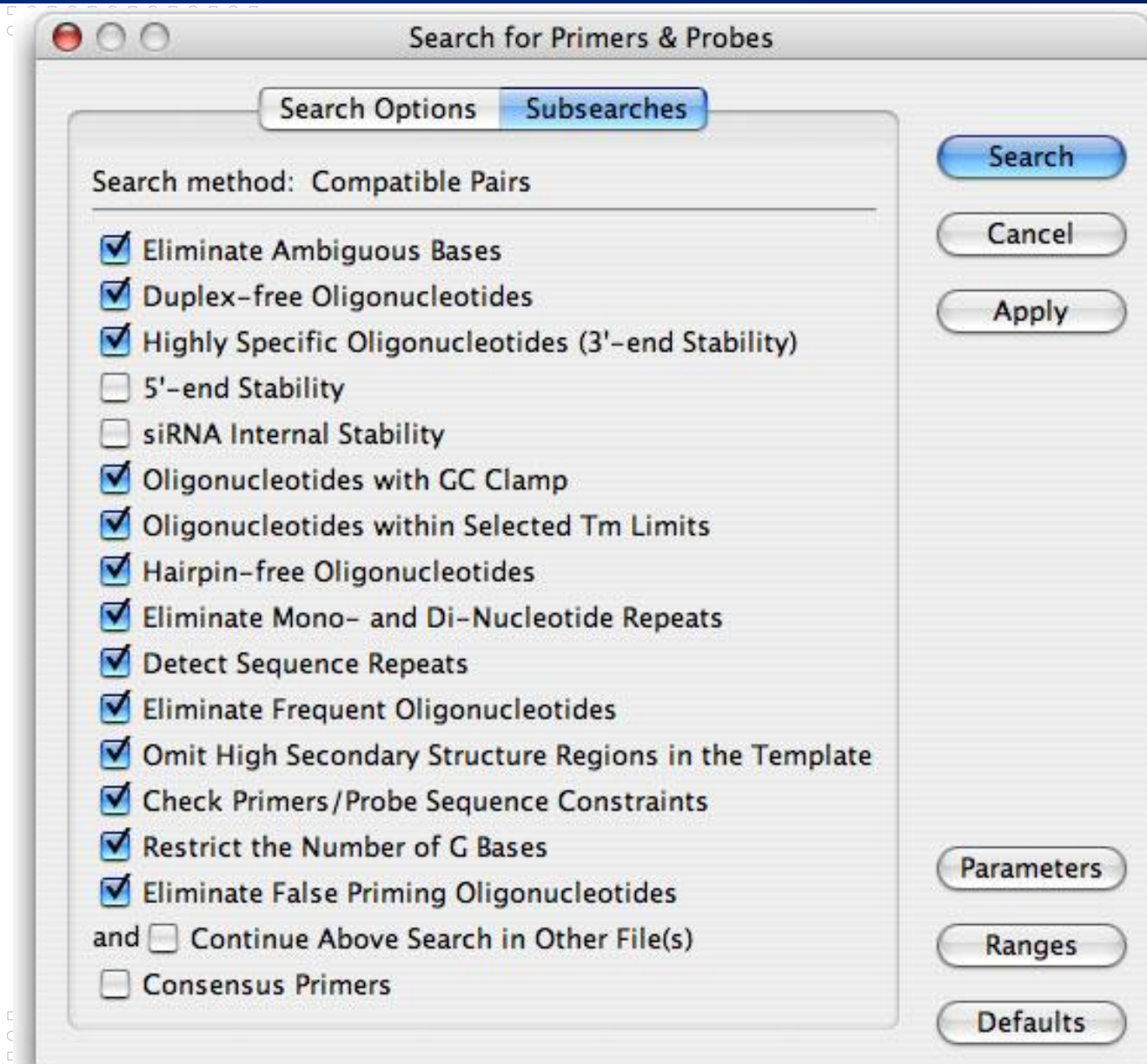
PCR Primers
Compatible with the Forward Primer Reverse Primer

TaqMan Probes & PCR Pairs
Compatible with the Upper Probe Lower Probe

Molecular Beacons & PCR Pairs
 Nested Primers
 Sequencing Primers
 Hybridization Probes
 siRNA Probes

After successfull search show: ▾

Search
Cancel
Apply
Parameters
Ranges
Defaults



PCR

File: Human 4E.seq

Optimal Annealing Temperature: 50.8 °C (Max: 66.3 °C)

	Position and Length		T _m [°C]	GC [%]	P.E.#	Score
Product	862		78.9	29.6	n/a	697
Forward Primer	918	22	56.9	45.5	471 / 471	840
Reverse Primer	1753	27	55.3	29.6	489 / 489	834
Upper Oligo	979	24	56.5	33.3	479 / 479	917
Lower Oligo	1694	23	55.4	39.1	457 / 457	841

Product T_m - Reverse Primer T_m : 23.6 °C
 Primers T_m difference: 1.6 °C Comments:

	Concentration	
Forward Primer	200.0	nM
Reverse Primer	200.0	nM
Upper Oligo	200.0	nM
Lower Oligo	200.0	nM
Monovalent Cation	50.0	mM
Free Mg[2+]	0.7	mM

Total Na[+] Equivalent: 155.8 mM

Selected Primers			
File: BRCA2 gene.seq			
AY436640:15438F22		AY436640:15917R20	
5' CAATATATACCGTAGTCCCCTA 3'		5' CAGCTACATATTACGCCAGA 3'	
Length:	22-mer	Length:	20-mer
Score:	802 points	Score:	914 points
5' Position:	15438	3' Position:	15917
T_m/t_m :	53.4 52.6 °C	T_m/t_m :	53.1 53.8 °C
$\Delta G/\Delta g$ (25 °C):	-30.5 -29.2 kcal/mol	$\Delta G/\Delta g$ (25 °C):	-28.6 -28.5 kcal/mol
$\Delta S/\Delta s$:	-472.1 -449.5 cal/°K * mol	$\Delta S/\Delta s$:	-430.5 -419.6 cal/°K * mol
$\Delta H/\Delta h$:	-171.3 -163.2 kcal/mol	$\Delta H/\Delta h$:	-157.0 -153.6 kcal/mol
3' ΔG :	-6.5 kcal/mol	3' ΔG :	-6.9 kcal/mol
Degeneracy:	1	Degeneracy:	1
P.E.#:	443/443	P.E.#:	477/477
1/E:	4.63 nmol/A ₂₆₀ 31.1 µg/A ₂₆₀	1/E:	5.05 nmol/A ₂₆₀ 31.0 µg/A ₂₆₀

Priming Efficiency PE Score



Sekundární struktury

- HAIRPIN intramolekulární
- DIMER intermolekulární

Hairpin

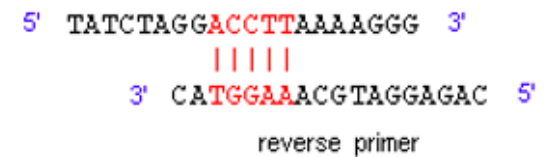


Self-Dimer



Dimer

forward primer



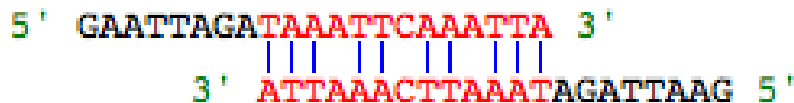
- HAIRPIN intramolekulární
- DIMER intermolekulární

Current Oligo Duplexes

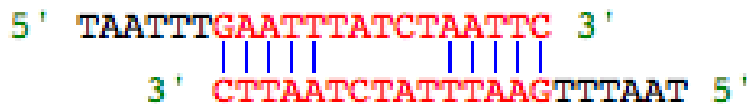
File: BRCA2 gene.seq

Current Oligo 21-mer [5042]

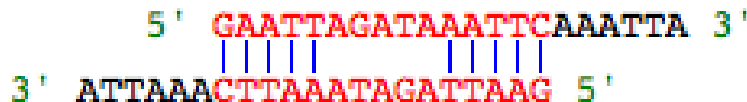
[Current+ Oligo] - The most stable 3'-dimer: # of hydrogen bonds = 10; $\Delta G = -0.7$ kcal/mol



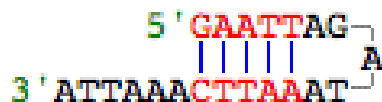
[Current- Oligo] - The most stable 3'-dimer: # of hydrogen bonds = 10; $\Delta G = -7.3$ kcal/mol; $T_m = 2.9^\circ\text{C}$



The most stable dimer overall: # of hydrogen bonds = 10; $\Delta G = -7.4$ kcal/mol; $T_m = 2.2^\circ\text{C}$



Hairpin: loop = 5 nt; $\Delta G = -3.0$ kcal/mol; $T_m = 54.6^\circ\text{C}$



Reverse Primer False Priming Sites

File: M13MP18

Reverse Primer M13MP18:6310R19 (positive strand)

Priming efficiency of the perfect match is 482 (above the threshold)

Priming efficiency: 482 (above the threshold)

```

5' (6328) GGTTTCCAGTCACGACG (6310) 3'
          |||||
3' (6328) ccaaaagggtcagtgctgc (6310) 5'
    
```

Priming efficiency: 244 (above the threshold)

```

5' (6328) GGTTTCCAGTCACGACG (6310) 3'
          ||| ||| |||||
3' (626)  agcaaagggtc--tgctgc (610) 5'
    
```

Priming efficiency: 193 (above the threshold)

```

5' (6328) GGTTTCCAGTCACGACG (6310) 3'
          | | | ||||| |||
3' (5125) tctaagggtcagtg-tgc (5108) 5'
    
```

AHP2 cDNA (TAIR database)

Sequence: AT3G29350.1 Date last modified 2007-04-17 Name AT3G29350.1 Tair
Accession Sequence:4010737427 Sequence Length (bp) 827

1 ACAATTCGCG AGAAAGACAA AACACAAGTT TCTTCTTCTT GGGATTGGCT
51 ATTTCCAGAA ATCCAAGTCA ATAATCAAAG TCCAAACAAA AAAATCCTCT
101 CCCAATCTCC GCTTCACTCT TCTCATGGAC GCTCTCATTG CTCAGCTTCA
151 GAGACAATTT CGTGATTACA CCATTTCTCT CTACCAACAG GGGTTTTTGG
201 ATGATCAATT TACTGAGTTG AAAAAGCTAC AAGATGATGG AAGTCCTGAT
251 TTTGTGTCTG AAGTGCTTTC ACTTTTCTTT GAAGATTGTG TGAAGCTTAT
301 CAGTAACATG GCTAGAGCTT TGGACACGAC AGGAACTGTA GATTTTAGTC
351 AGGTAGGTGC TAGTGTGCAT CAATTGAAGG GTAGTAGCTC AAGTGTTGGT
401 GCCAAGAGGG TCAAAACTTT GTGTGTTAGC TTCAAGGAAT GTTGTGAAGC
451 TAAGAACTAC GAAGGGTGTG TGAGATGTTT GCAGCAAGTG GATATTGAGT
501 ACAAGGCGTT AAAGACAAAG CTTCAAGATA TGTTCAATCT TGAGAAACAG
551 ATCATTCAAG CTGGTGGTAT AGTTCCTCAA GTGGATATTA ACTAAAGAGA
601 CTAGTCCATA AGAAGAAAAA AGATGATGAC TTTCTTTCTT TAGTTTCTCT
651 TCTAAATTAT TTTGGATTTG GTGTTTGCTC AAAAACTCAA TAAAATATGT
701 GCAAAAAGAA ACAAAAACAA GTGATGGTTG TTTATAAATC AGTAGTATGT
751 ATTGTTTGAT CTCATCCGAG AAAATTGAAA CCATTGGACT AATGAATGTG
801 ATGATAATAT ATATTGGTTT GCTTCTG

101 CCCAATCTCC GCTTCACTCT TCTCATGGAC GCTCTCATTG CTCAGCTTCA
 151 GAGACAATTT CGTGATTACA CCATTTCTCT CTACCAACAG GGGTTTTTGG
 201 ATGATCAATT TACTGAGTTG AAAAAGCTAC AAGATGATGG AAGTCCTGAT
 251 TTTGTGTCTG AAGTGCTTTC ACTTTTCTTT GAAGATTGTG TGAAGCTTAT
 301 CAGTAACATG GCTAGAGCTT TGGACACGAC AGGAACTGTA GATTTTAGTC
 351 AGGTAGGTGC TAGTGTGCAT CAATTGAAGG GTAGTAGCTC AAGTGTTGGT
 401 GCCAAGAGGG TCAAAACTTT GTGTGTTAGC TTCAAGGAAT GTTGTGAAGC
 451 TAAGAACTAC GAAGGGTGTG TGAGATGTTT GCAGCAAGTG GATATTGAGT
 501 ACAAGGCGTT AAAGACAAAG CTTCAAGATA TGTTCAATCT TGAGAAACAG
 551 ATCATTCAAG CTGGTGGTAT AGTTCCTCAA GTGGATATTA ACTAAAGAGA

EcoRI restriction site

5'.....G|AATTC.....3'

3'.....CTTAA|G.....5'

|

Design of primers

AHP2ex_up

5'- CCG GAA TTC ATG GAC GCT CTC ATT GCT CAG – 3'

AHP2ex_low

5'- CCG GAA TTC TTA GTT AAT ATC CAC TTG AGG – 3'

101 CCCAATCTCC GCTTCACTCT TCTC**ATGGAC GCTCTCATTG CTCAGCTTCA**
 151 GAGACAATTT CGTGATTACA CCATTTCTCT CTACCAACAG GGGTTTTTGG
 201 ATGATCAATT TACTGAGTTG AAAAAGCTAC AAGATGATGG AAGTCCTGAT
 251 TTTGTGTCTG AAGTGCTTTC ACTTTTCTTT GAAGATTGTG TGAAGCTTAT
 301 CAGTAACATG GCTAGAGCTT TGGACACGAC AGGAACTGTA GATTTTAGTC
 351 AGGTAGGTGC TAGTGTGCAT CAATTGAAGG GTAGTAGCTC AAGTGTTGGT
 401 GCCAAGAGGG TCAAAACTTT GTGTGTTAGC TTCAAGGAAT GTTGTGAAGC
 451 TAAGAACTAC GAAGGGTGTG TGAGATGTTT GCAGCAAGTG GATATTGAGT
 501 ACAAGGCGTT AAAGACAAAG CTTCAAGATA TGTTCAATCT TGAGAAACAG
 551 ATCATTCAAG CTGGTGGTAT AGTT**CCTCAA GTGGATATTA ACTAA**AGAGA

EcoRI restriction site

5'.....G|AATTC.....3'
 3'.....CTTAA|G.....5'

Design of primers

AHP2ex_up

5'- CCG **GAA TTC** ATG GAC GCT CTC ATT GCT CAG – 3'

AHP2ex_low

5'- CCG **GAA TTC** TTA GTT AAT ATC CAC TTG AGG – 3'

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Discovery is not in seeking new landscapes,
but in having new eyes...

Marcel Proust

Tato prezentace vznikla s podporou projektu **OP VK** „Rozvoj týmu pro výuku, výzkum a aplikace v oblasti funkční genomiky a proteomiky“ (CZ.1.07/2.3.00/09.0132)

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INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ

