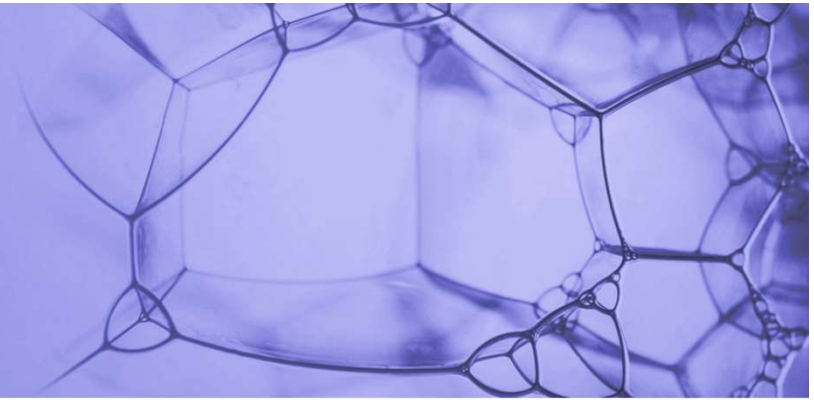


**LOSCHMIDT
LABORATORIES**



Protein expression & purification technologies

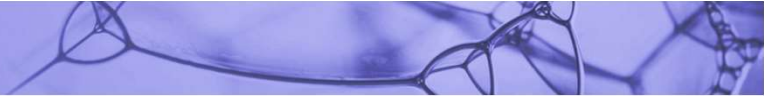
Manufacturing of recombinant proteins, protein complexes and virus-based vehicles for biotechnology and basic research

Dr. Martin Marek
Loschmidt Laboratories
Faculty of Science, MUNI
Kamenice 5, bld. A13, room 332
martin.marek@recetox.muni.cz



What will we talk about

- Introduction to recombinant protein production
 - definition, goals and applications
- Protein expression in microbial cells
 - *Escherichia coli*, yeasts
- Expression and multi-expression in eukaryotic cells
 - Mammalian and insect cells, baculovirus expression
- New emerging expression tools
 - CRISPR/Cas9 technology, cell-free protein expression
- Protein purification, characterization and storage
 - Chromatography separation & QC check

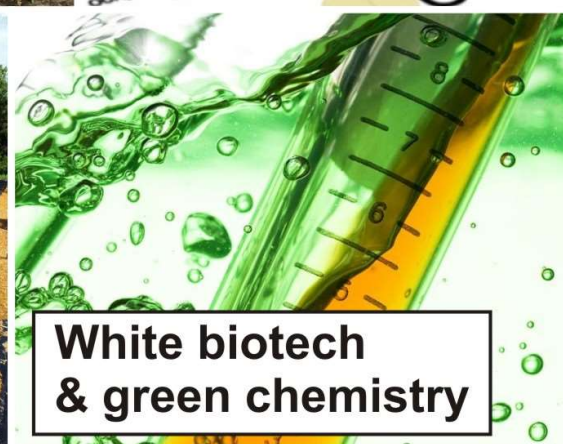
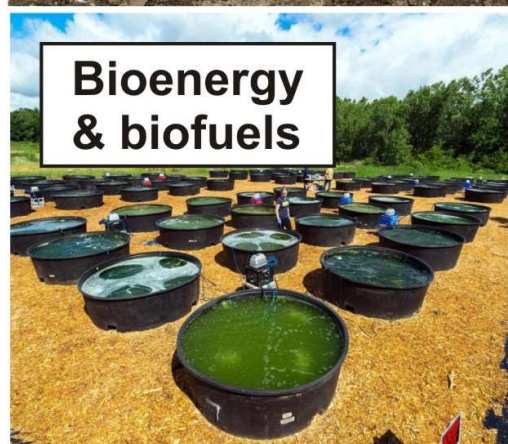


Why to purify a protein?

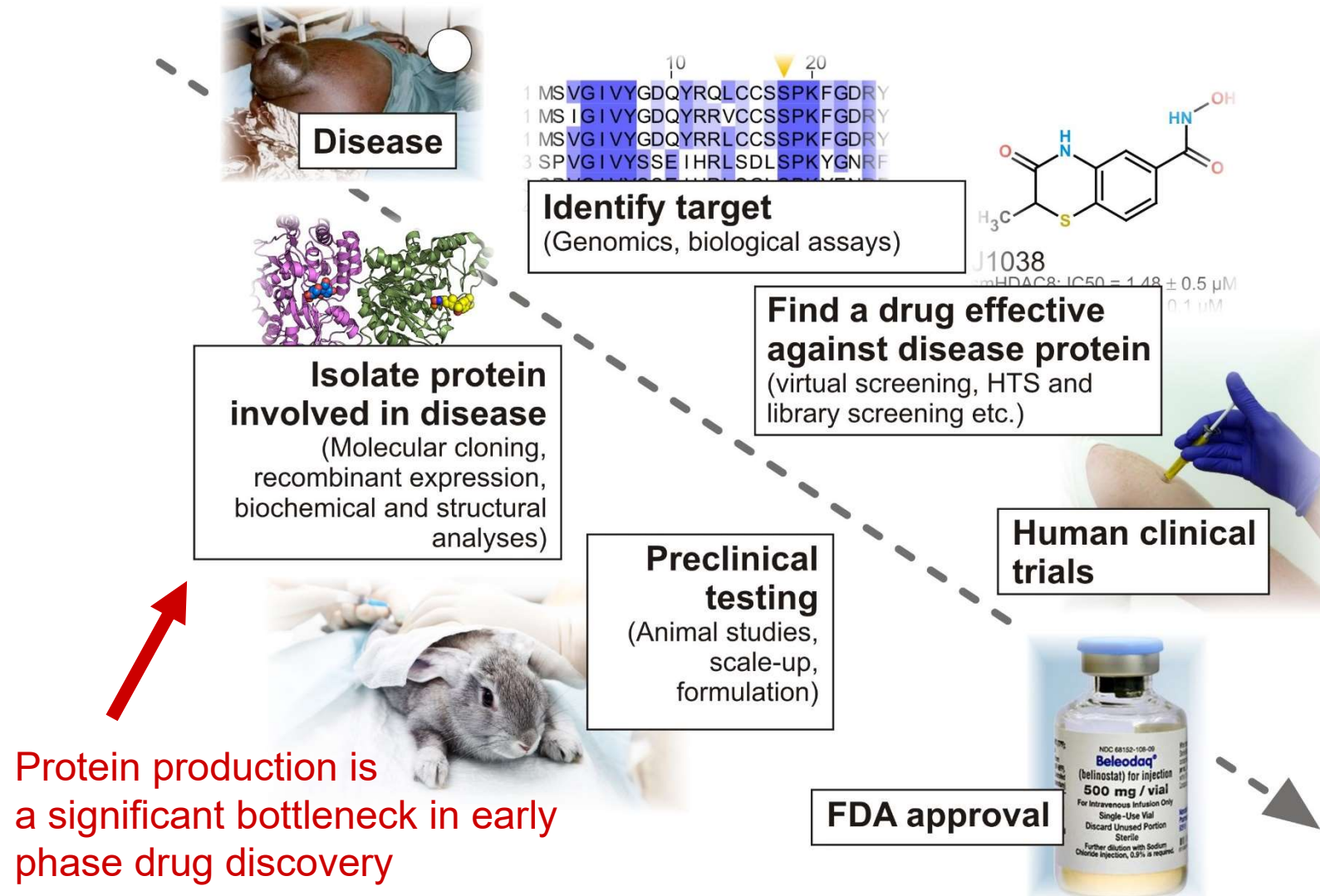
- To study its function
- To analyze its physical properties
- To determine its sequence
- To determine its three-dimensional structure
- For industrial or therapeutic applications

Applications of protein technologies

Why should we bother to think about protein technologies?



The key role of protein production in pharma industry

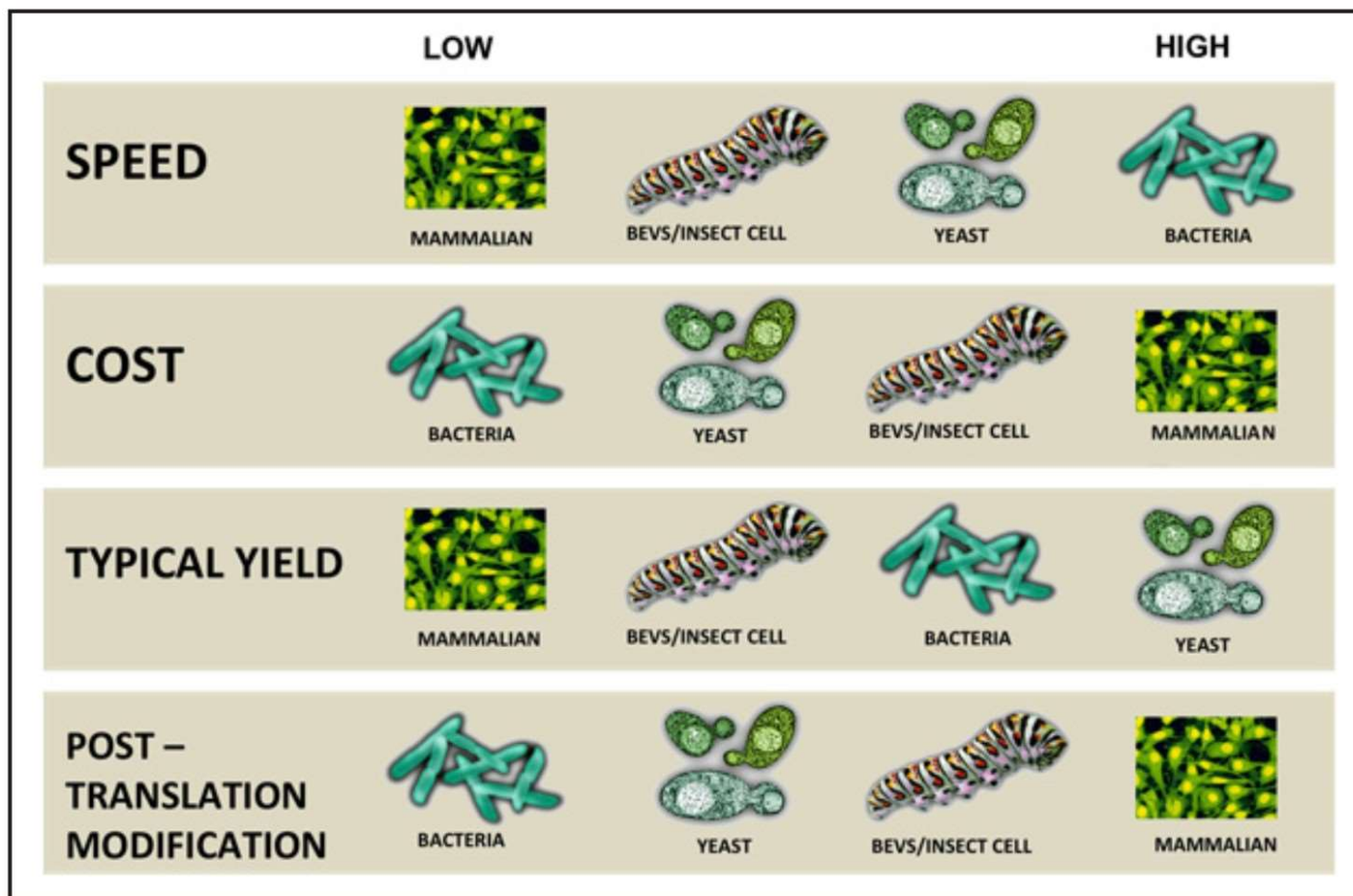


Recombinant protein production workflow

- Molecular cloning
- Protein expression
- Protein purification
- Protein characterization
- (Protein structure determination)

Cellular protein production: selection of a host

- *Escherichia coli*
- Yeasts (*Pichia*, *Kluyveromyces*)
- Insect cells (Baculovirus expression system)
- Mammalian cells (HEK293)



Bacterial expression system

Concepts
Methods
Applications



Recombinant protein expression in *Escherichia coli*

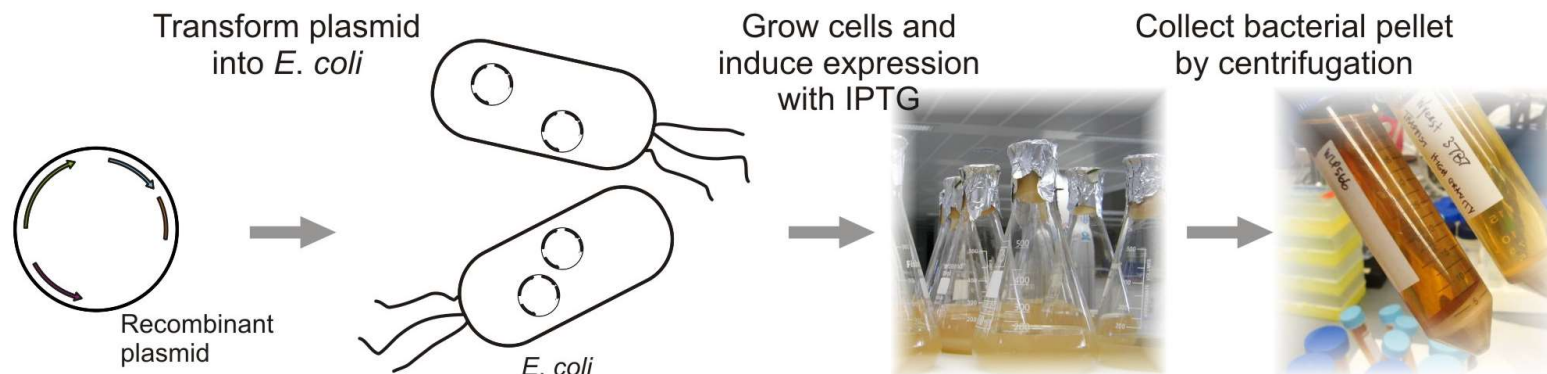
ADVANTAGES

- Inexpensive setup and running costs
- High recombinant protein production levels
- Short timeline from cloning to protein recovery (1 week)
- Limited technical knowledge required for culturing
- Scalability from small (2 mL) to very large culture (>10,000 L) volumes

DRAWBACKS

- Inability to perform post-translation modifications (PTMs)
- Limited formation of disulphide bond

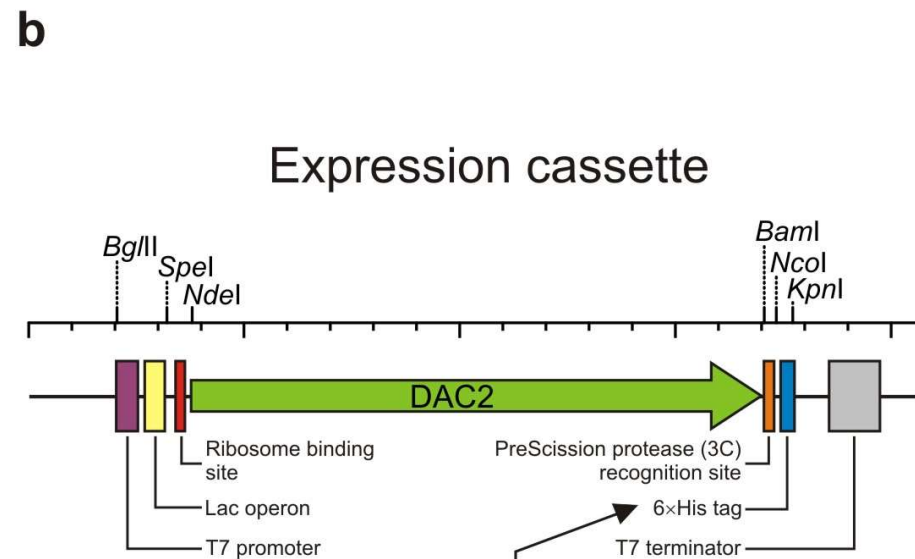
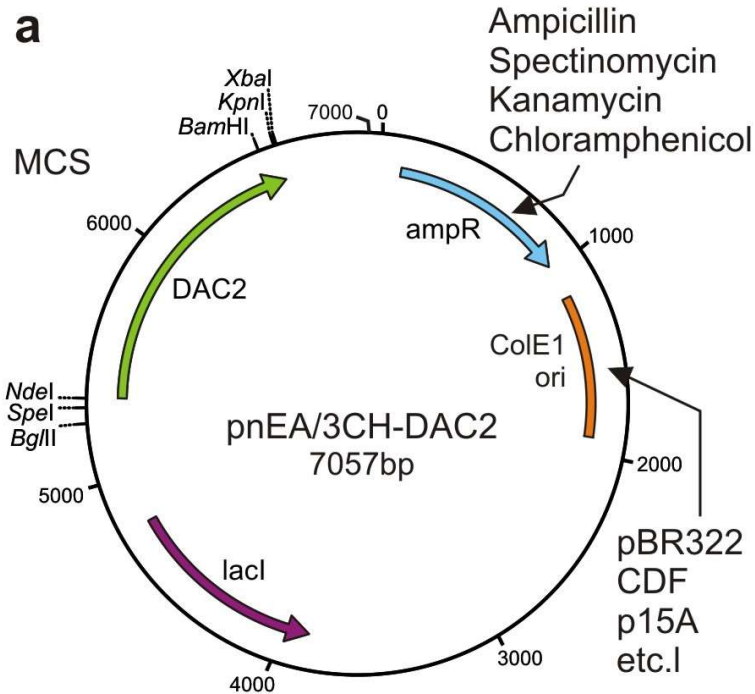
Workflow of protein expression in *E. coli*





Protein expression in *E. coli*: a plasmid backbone

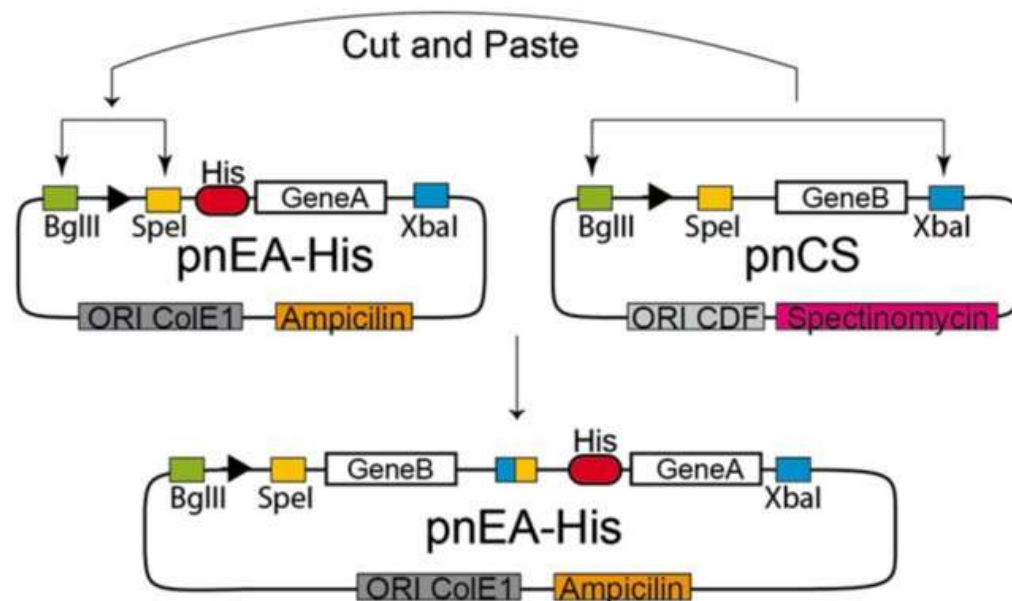
- pET
- pGEX



Polyhistidines (6xHis, 10xHis or 12xHis)
Thioredoxin (Thx)
Maltose-binding protein (MBP)
Glutathione S-transferase (GST)
Small ubiquitin-like modifier (SUMO)

Co-expression of protein complexes in *E. coli*

The functional units within cells are often macromolecular complexes rather than single species. Production of these complexes as assembled homogenous samples is a prerequisite for their biophysical and structural characterization and hence an understanding of their function in molecular terms. Co-expression in *Escherichia coli* can decipher the subunit composition, assembly, and production of whole protein complexes.



- Example of the concatenation of two vectors of the pET-MCN vector series: pNEA-His (vector encoding an N-terminal poly-histidine tag in front of protein A) and pNCS (vector expressing the native protein B).
- The pNEA-His (acceptor vector) is linearized by removing with the restriction enzymes BglIII and Spel part of its promoter (T7 promoter and lacO).
- The full promoter of the pNCS (donor vector) is cut out with the restriction enzymes BglIII and XbaI. After ligation of the pNCS promoter with the linearized pNEA-His vector, a new vector is obtained based on the backbone of the pNEA-His vector and whose promoter controls the genes of proteins A and B.

Wide range of *E. coli* strains

Strain	Features
BL21 (DE3)	Most common host strain, enables high-level recombinant protein expression
BL21 (DE3) pLysS	Enables high-level expression and suppression of T7 RNA Polymerase basal level expression
BL21-CodonPlus	Improved expression of genes with codons rarely used in bacteria
Rosetta	Improved expression of genes with codons rarely used in bacteria
Arctic Express	Contains a plasmid encoding chaperonins to aid in folding and allow expression at low temperature (12°C)
Shuffle T7 Express	Expresses DsbC to enable cytoplasmic disulphide bond formation

Expression in *E. coli*: tips and tricks

- **Growth temperature:** Typically 37°C, but lowering (25°C or 15°C) improves folding/solubility
- **Expression with a fusion tag:**
 - polyhistidenes (6xHis, 10xHis or 12xHis)
 - Thioredoxin (Thx)
 - Maltose-binding protein (MBP)
 - Glutathione S-transferase (GST)
 - Small ubiquitin-like modifier (SUMO)
- **Co-expression:** Chaperonins (folding) and foldases (disulphide isomerase, DsbC)
Co-expression with interaction partners (co-solubility effects)
- **Media:** LB Broth, 2xLB Broth (simple and cheap), Terrific Broth (TB), 2YT
- **Antibiotics:** Selection of recombinant clones and prevention of contamination
Concentration of antibiotics in large-scale expression is decreased
- **Codon optimization:** Modifying codons in a gene sequence to match the codon usage bias of the host cell used for expression

Expression in yeasts

Concepts
Methods
Applications

Expression in yeasts

- The yeast systems, such as *Saccharomyces cerevisiae* and *Pichia pastoris*, constitute another microbial systems in heterologous protein expression.

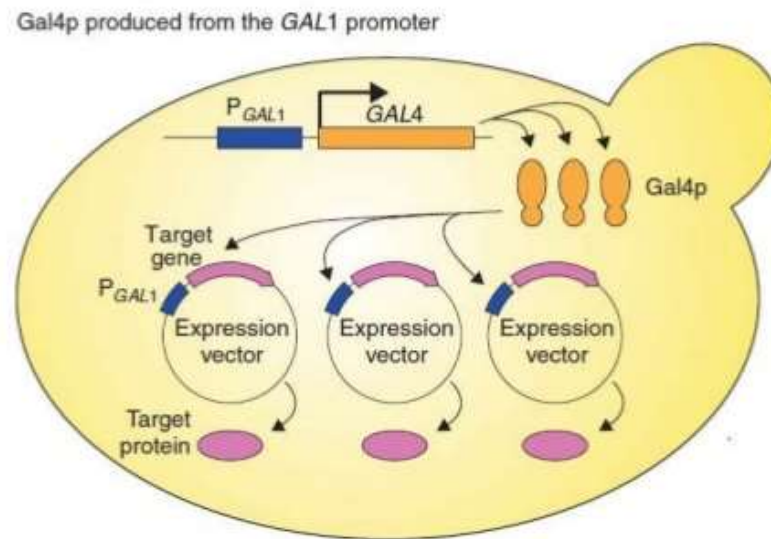
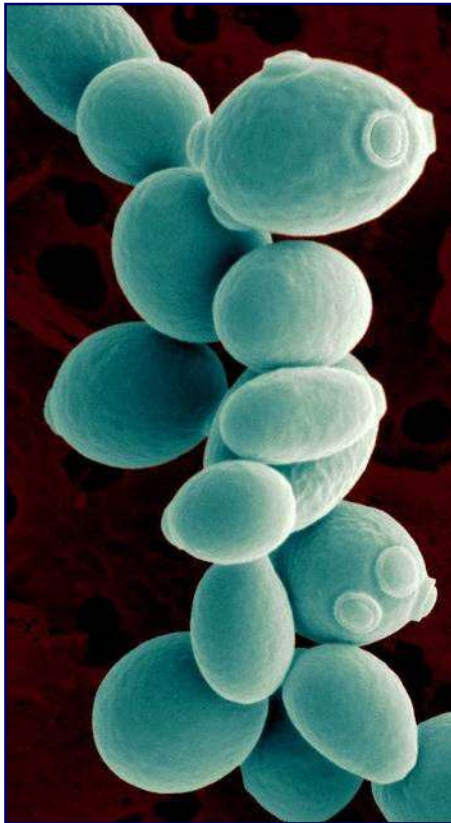
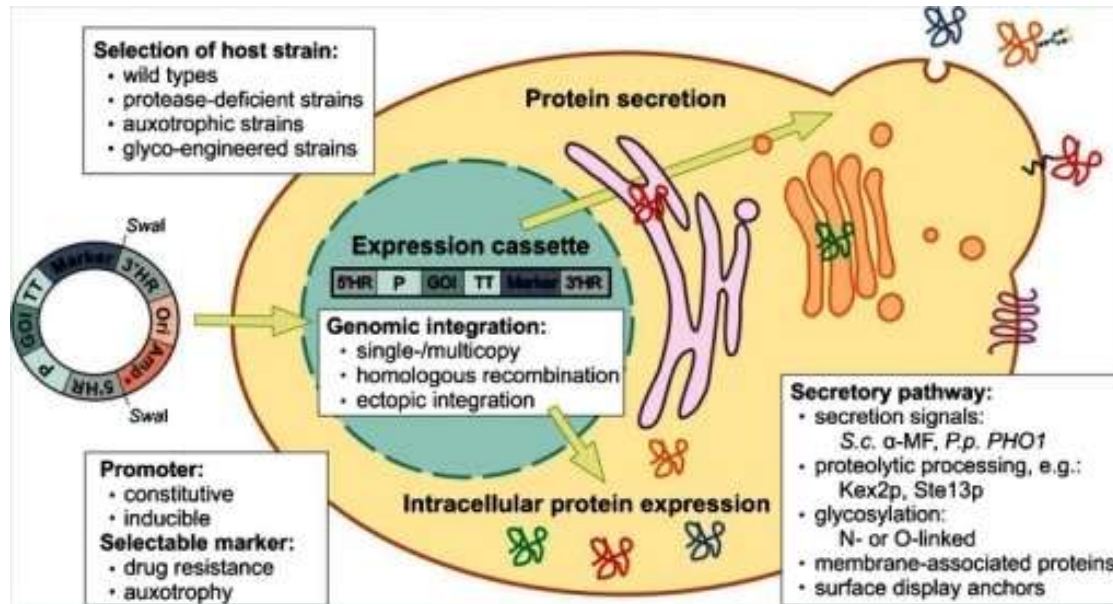
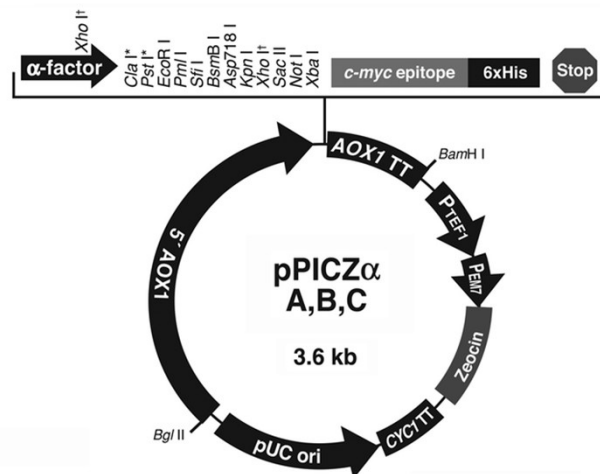


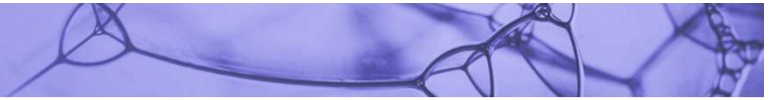
Figure 8.5. Galactose inducible gene expression in yeast. The expression of genes from multicopy vectors under the control of the $GAL1$ promoter (P_{GAL1}) can be increased substantially if the gene encoding the transcriptional activator of $GAL1$, $GAL4$, is also placed under the control of P_{GAL1} . In this case, induction by galactose will produce more Gal4p and consequently more of the target protein

Expression in *Pichia pastoris*



- *Pichia pastoris* (*P. pastoris*) is a widely used protein expression host for the production of biopharmaceuticals and industrial enzymes
- *P. pastoris* belongs to methylotrophic yeasts, which share a common pathway to metabolize one-carbon compounds as carbon and energy sources
- Vectors: The promoters for protein expression in *P. pastoris* include inducible promoters (AOX1, FLD1, ADH1, etc.) and constitutive promoters (GAP, TEF1, etc.). For *P. pastoris*, HIS4 (auxotrophic markers) and zeocin resistance (dominant markers) are the most popularly markers used. The heterologous proteins can be intracellular or secreted expression. *P. pastoris* has the ability to secrete high titres of proteins into culture media





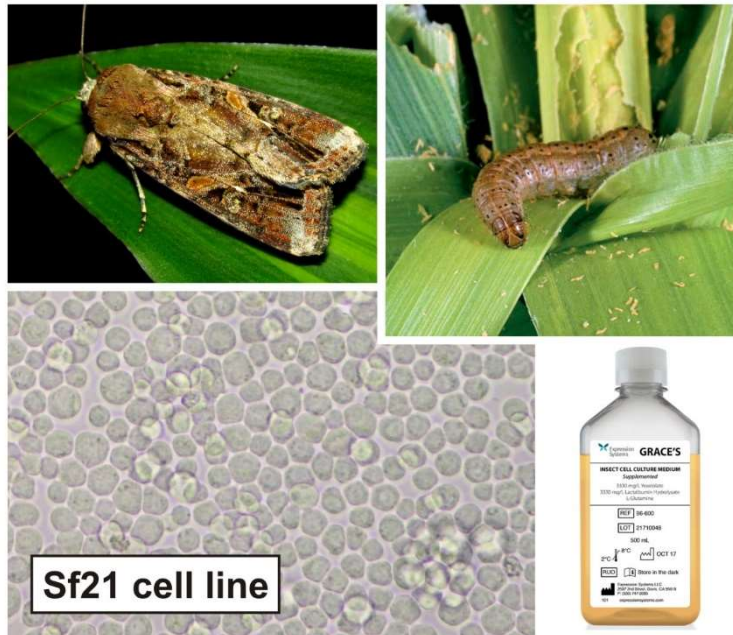
Expression in insect cells

Concepts
Methods
Applications

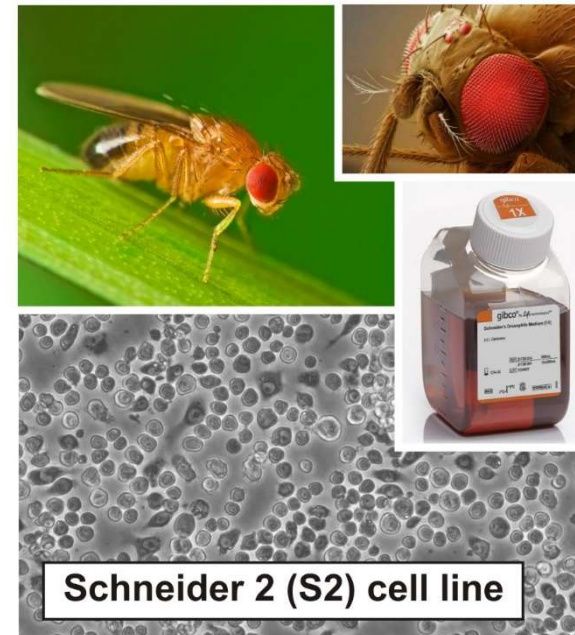
Production of proteins in insect cells

- Insect cells can efficiently express recombinant biologically active proteins and are mostly used for the development of virus-like particles and vaccines. It has been proven that insect cells are excellent platforms for the production of recombinant antibodies
- There are mainly three insect expression systems: **baculovirus expression vector system (BEVS)**, **InsectSelect (IS) system** and ***Drosophila* expression system (DES)**

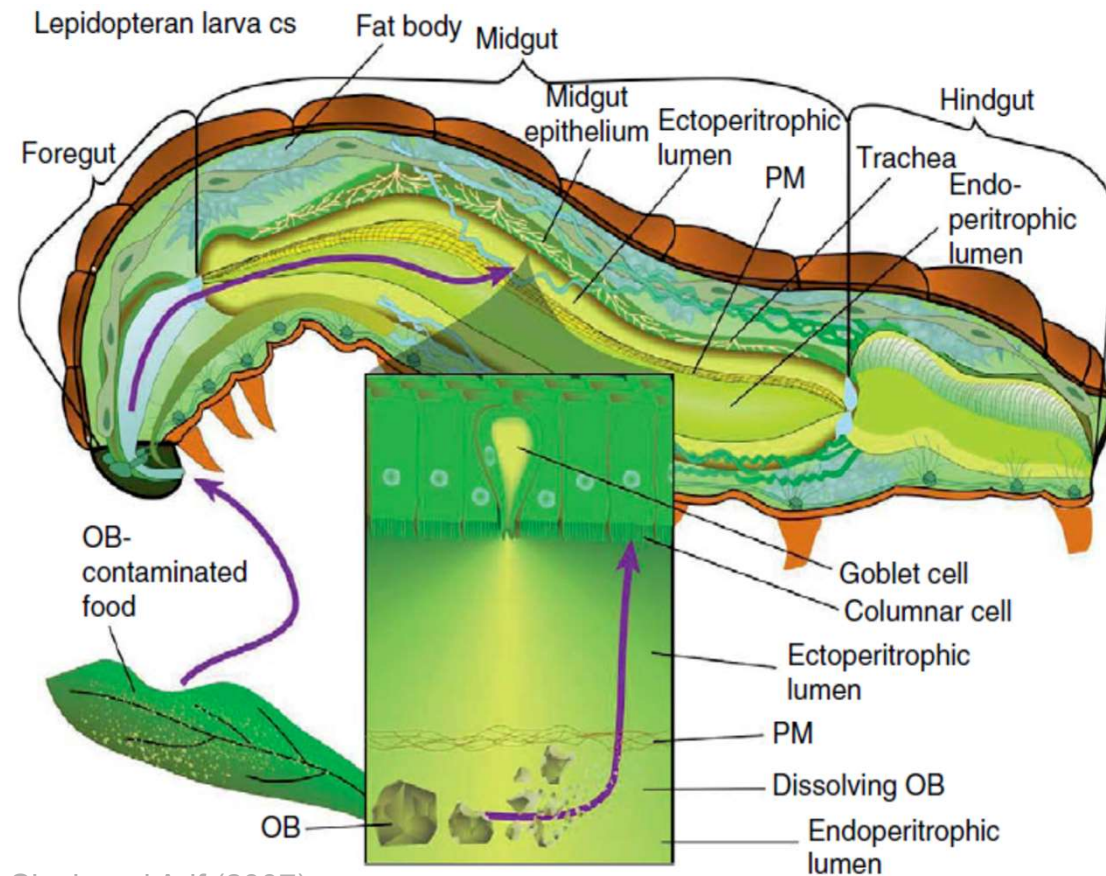
Spodoptera frugiperda, a fall army worm



Drosophila melanogaster



Introduction to baculovirus biology

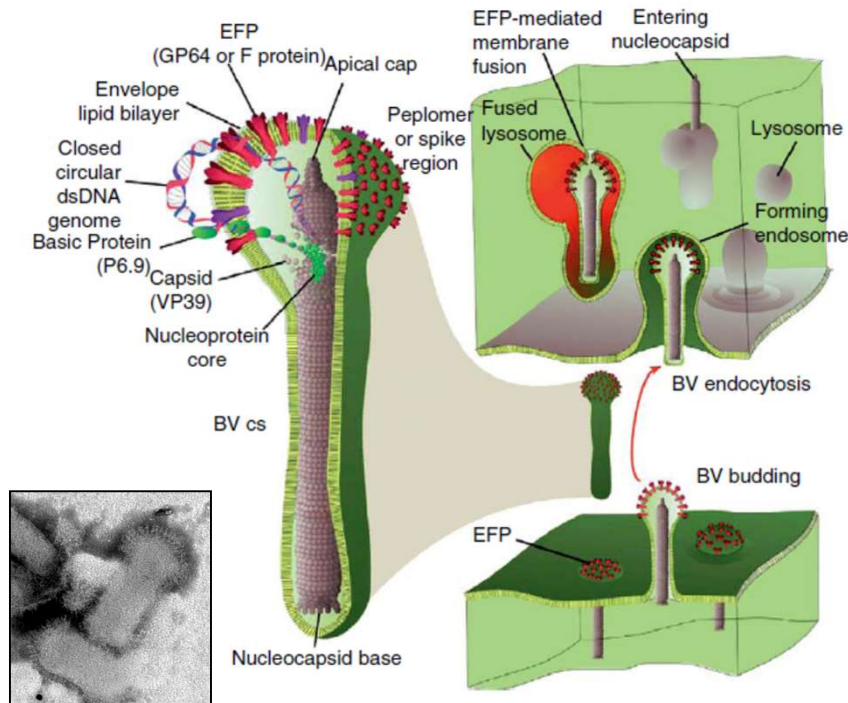


Slack and Arif (2007)

- Insect-infecting enveloped DNA viruses
- Baculoviruses are a very diverse group of viruses with double-stranded, circular, supercoiled genomes, with sizes varying from about 80 to over 180 kb, that encode between 90 and 180 genes
- The genome is packaged in rod-shaped nucleocapsids that are 230-385 nm in length and 40-60 nm in diameter
- In the most well characterized baculoviruses, the virions are present as two types, occluded virions (ODV) and budded virions (BV).
- Although these two types of virions are similar in their nucleocapsid structure, they differ in the origin and composition of their envelopes and their roles in the virus life cycle

Baculovirus morphogenesis: two types of virions

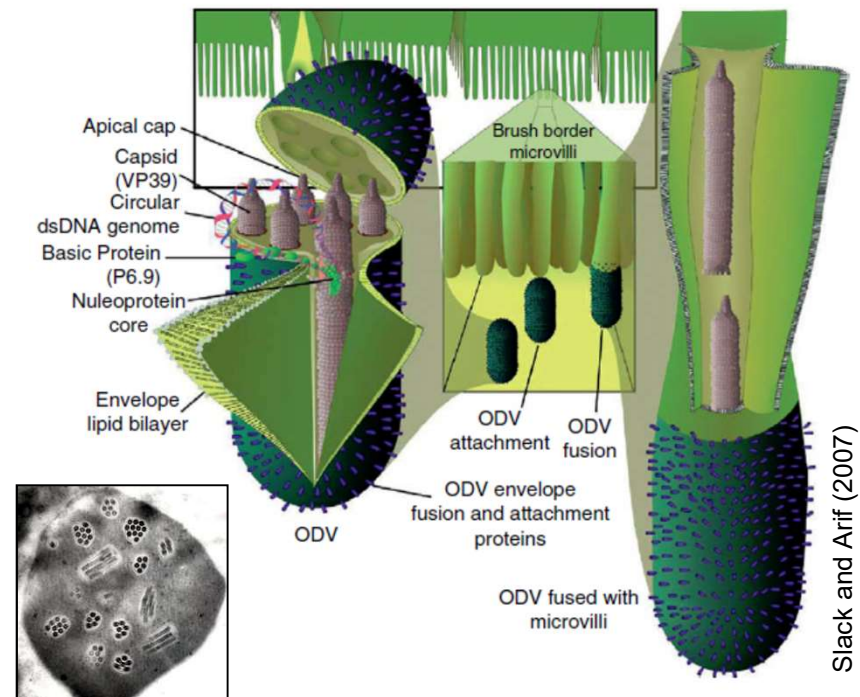
Budded virus (BV)



Circular dsDNA genome and associated proteins
Capsid shell (VP39, etc.)

BV envelope (GP64, F)

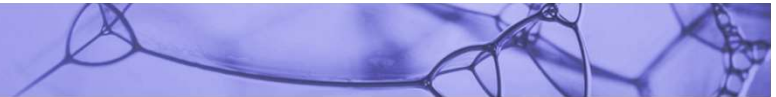
Occlusion-derived virus (ODV)



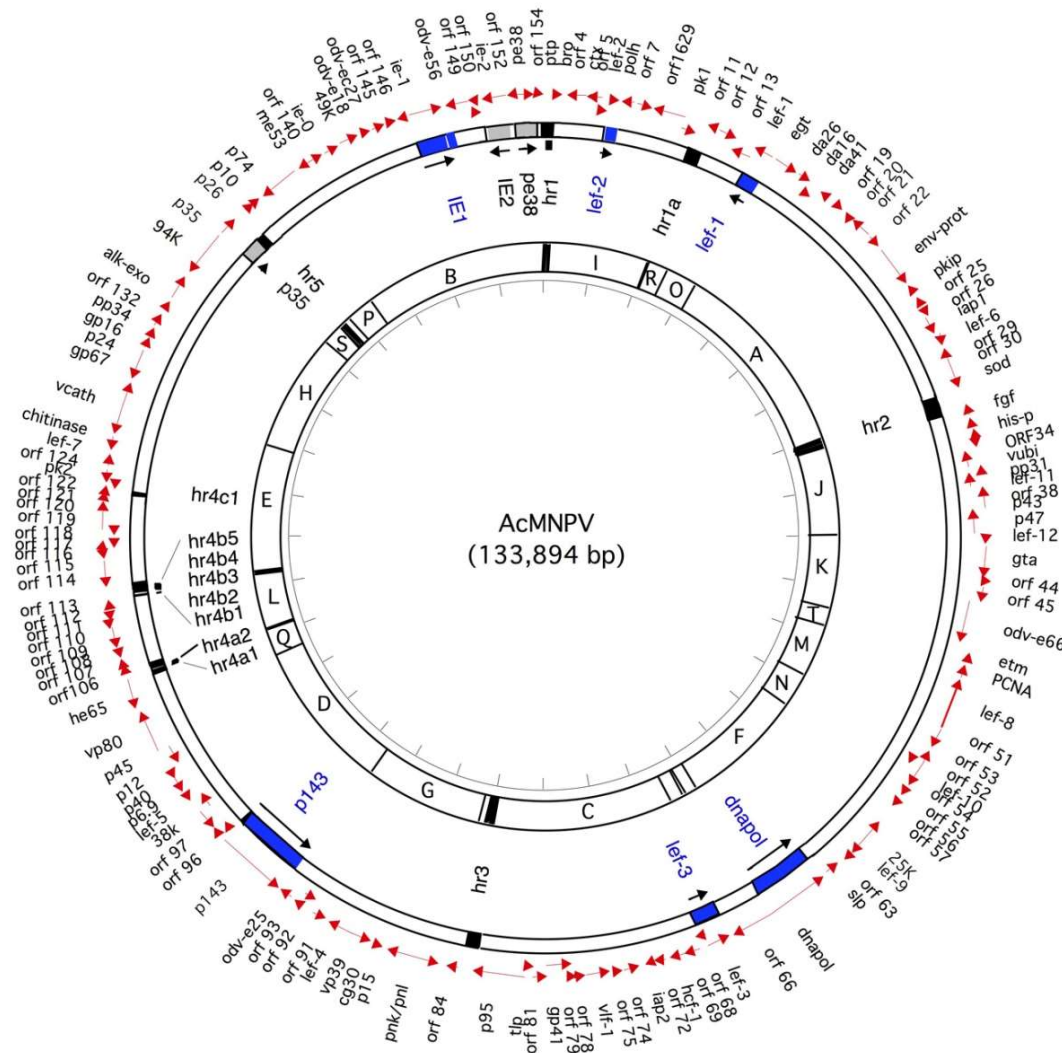
Circular dsDNA genome and associated proteins
Capsid shell (VP39, etc.)

ODV envelope (P74, ODV-E66, PIFs etc)

Tegument (GP41), occlusion body (Polyhedrin)

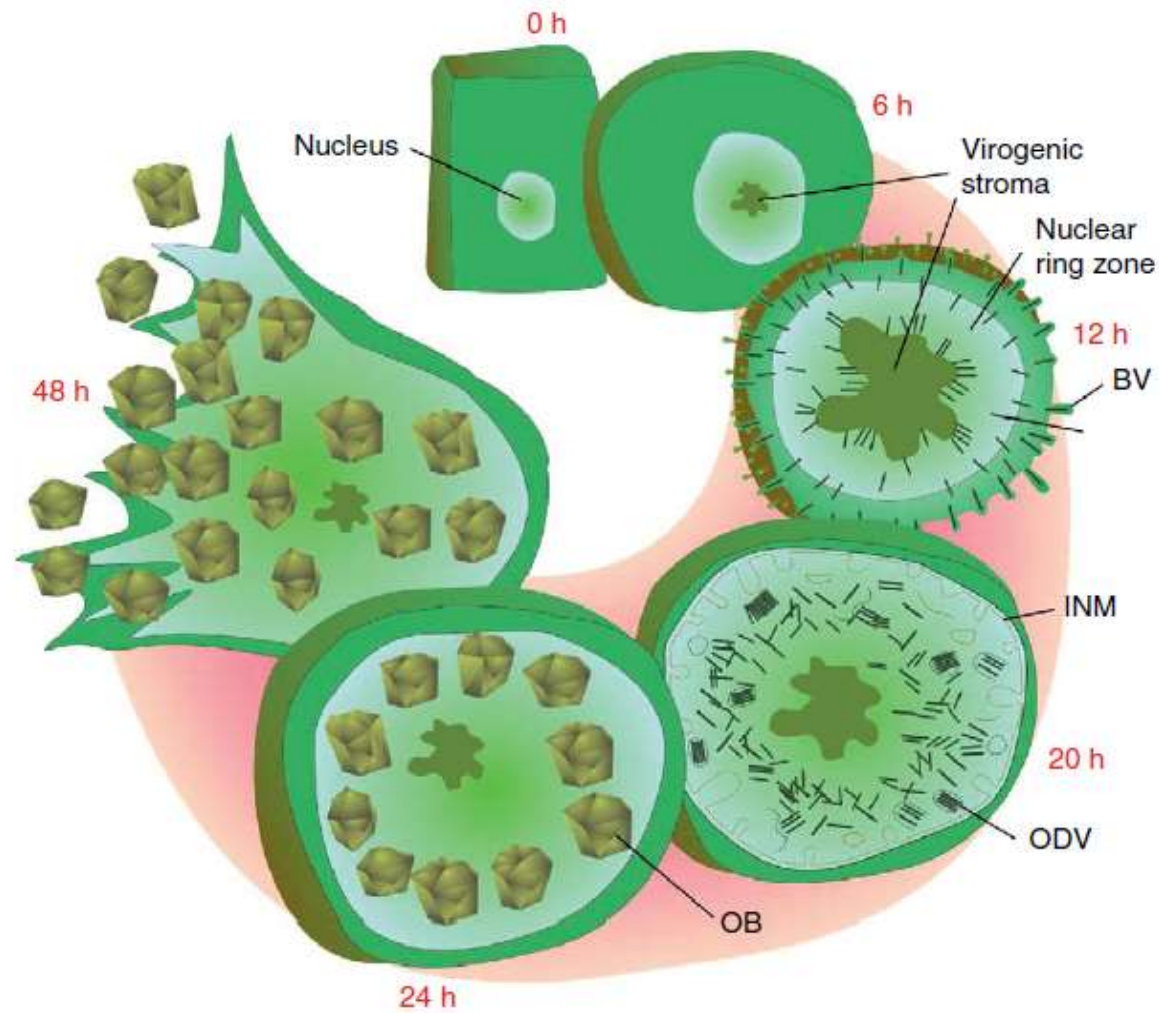


The baculovirus genome



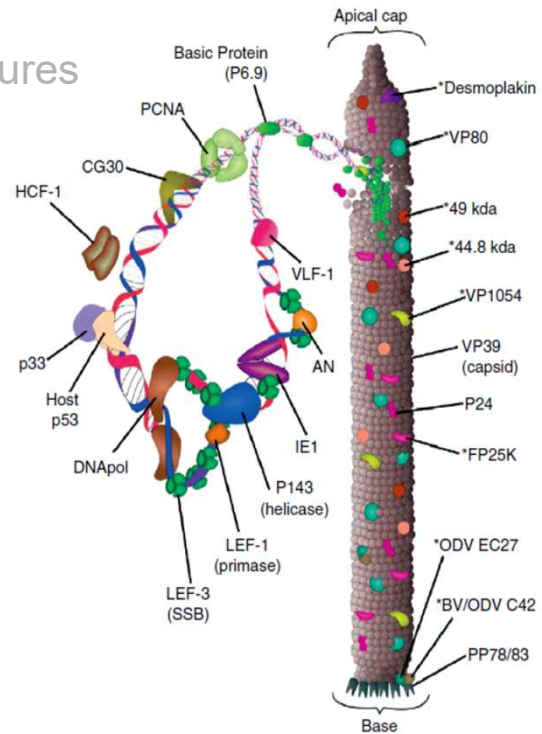
- Circular, supercoiled dsDNA genomes, with sizes varying from about 80 to over 180 kb, that encode between 90 and 180 genes
- The genome is packaged in rod-shaped nucleocapsids that are 230-385 nm in length and 40-60 nm in diameter

Time-course baculovirus infection

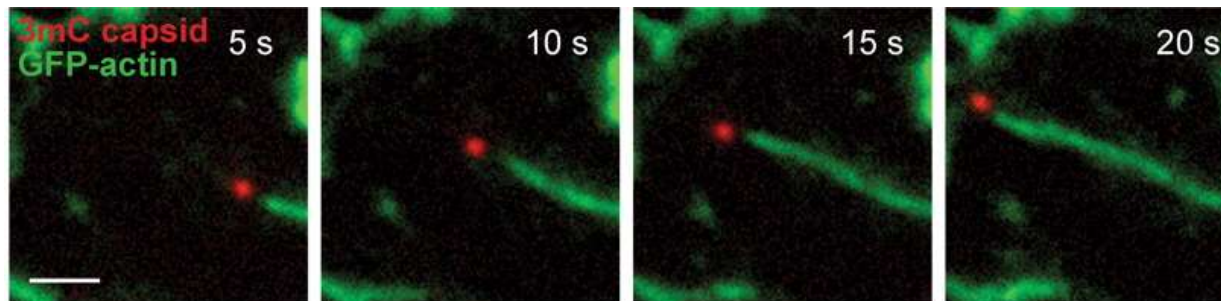


The structure of baculovirus nucleocapsid

- Rod-like capsid (VP39, etc.)
- Distinct apical and basal structures
- Circular dsDNA genome
- PP78/83: Wiskott-Aldrich syndrome protein (WASP)-like protein

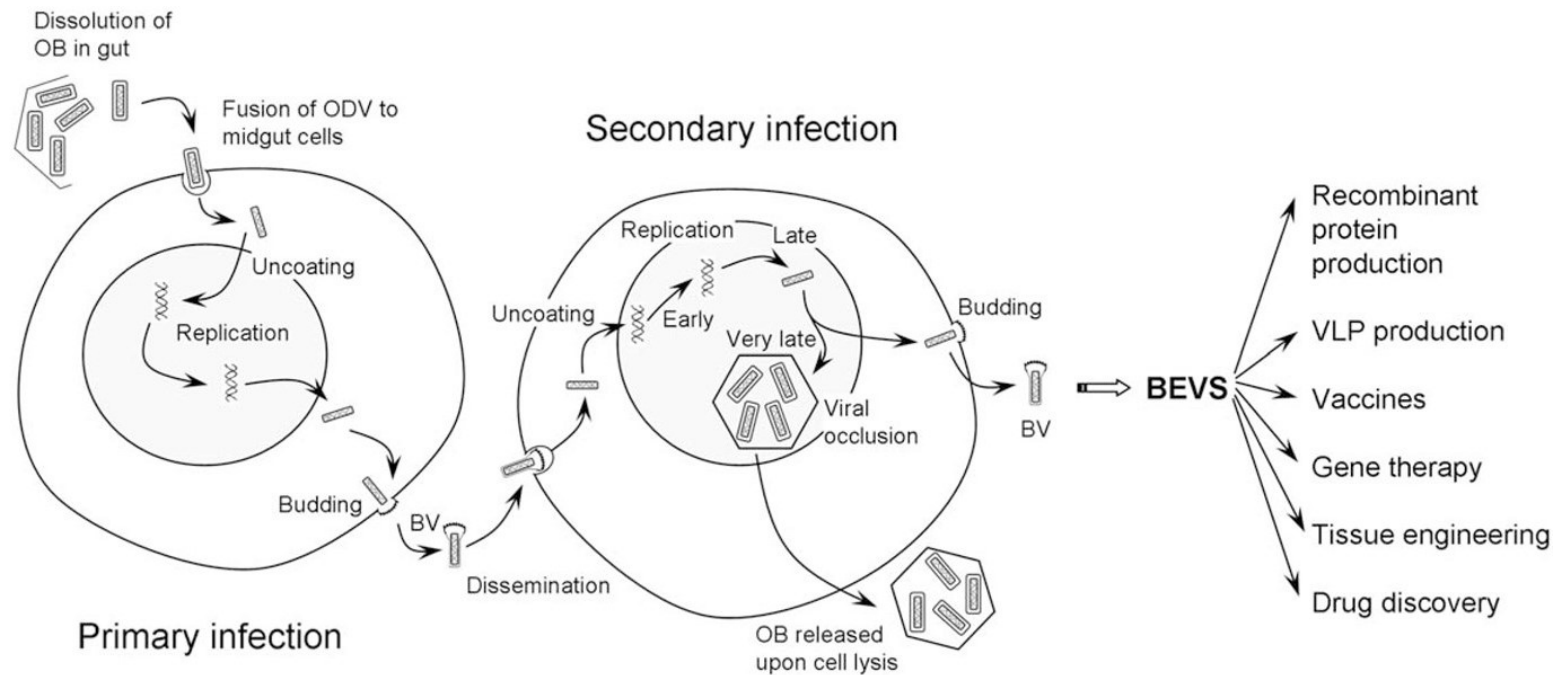


Dr. Taro Ohkawa
 University of California
 Berkeley, USA



Ohkawa *et al.* (2010)
 J. Cell Biol.

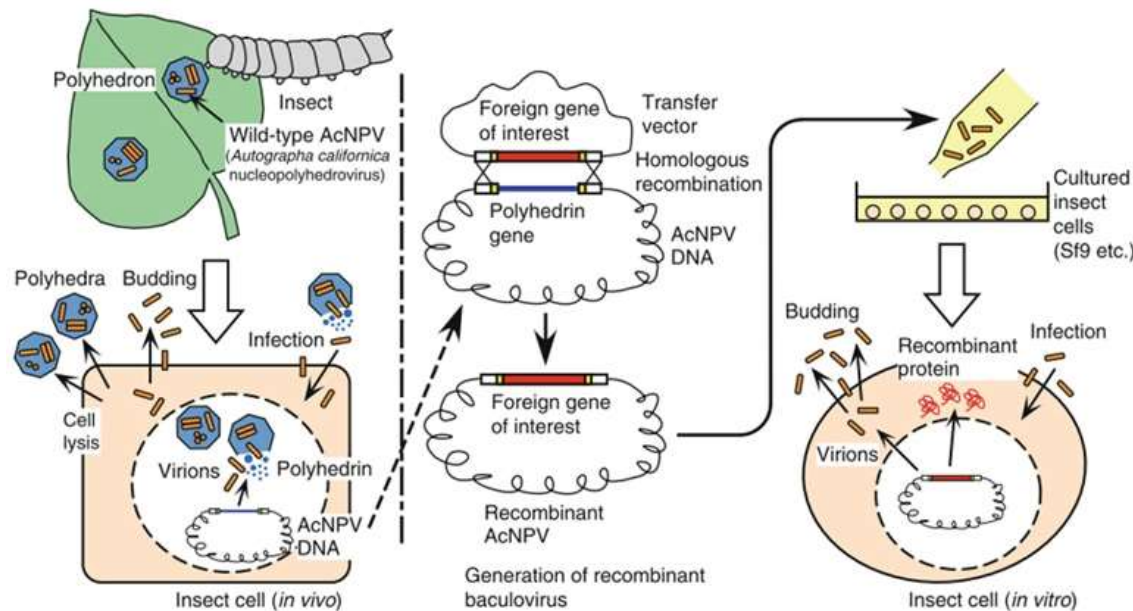
The replication cycle of baculovirus



- Baculovirus is a member of the insect-borne virus family and was originally used as a biopesticide with a large double-stranded circular DNA genome. The virus is enveloped, rod-shaped particles, ranging from 30 to 60 nm in diameter and from 250 to 300 nm in length.
- The natural hosts for these viruses are insects, and *Autographa californica* (AcMNPV) is the most studied member of the family, which was developed into a recombinant baculovirus vector and is still under application today.
- AcMNPV and other baculoviruses are able to generate “polyhedra” or “occlusion bodies” in the infected cells of insects. In the late phase of infection, dozens of polyhedra are formed in the host cell nuclei, which accommodate progeny virions encased by a protective paracrystalline array comprised of virus-encoded protein-polyhedrin.



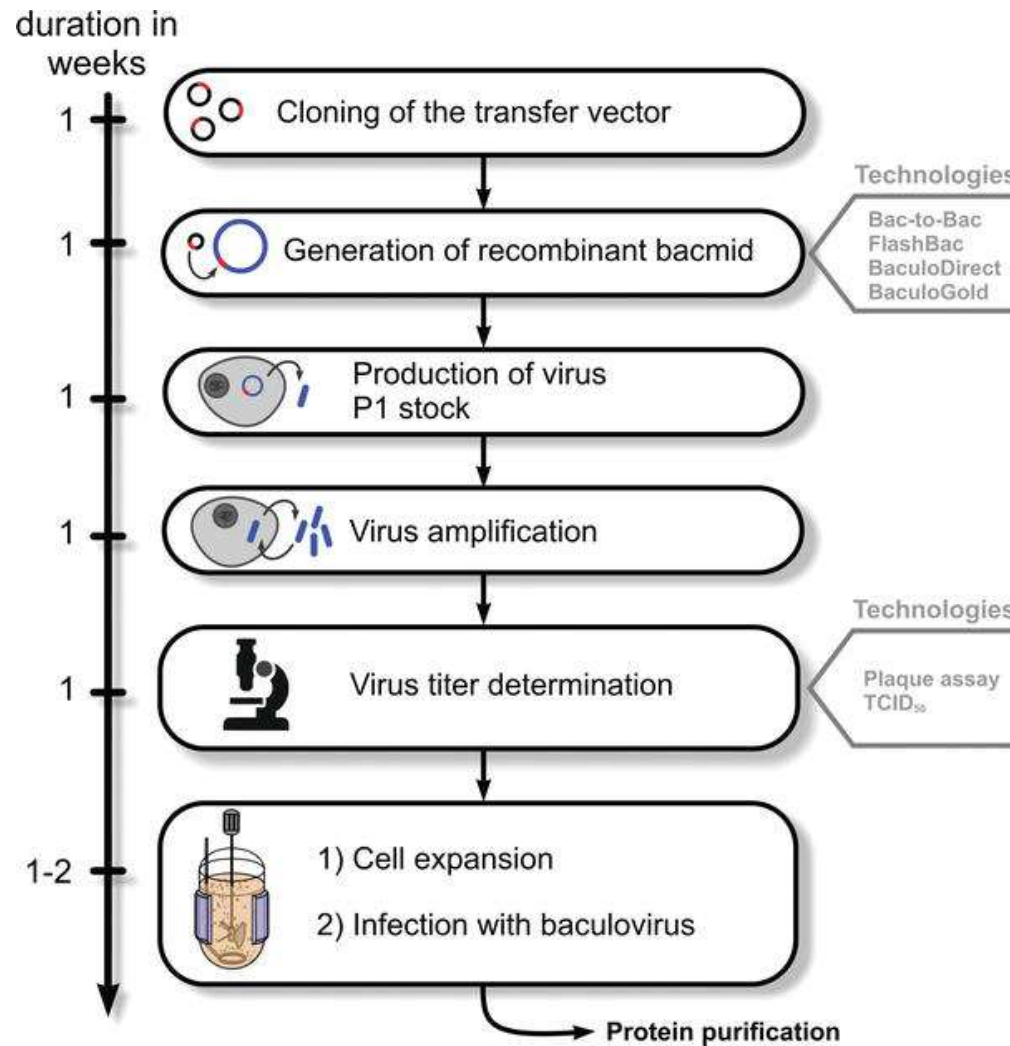
Baculovirus Expression Vector System (BEVS)



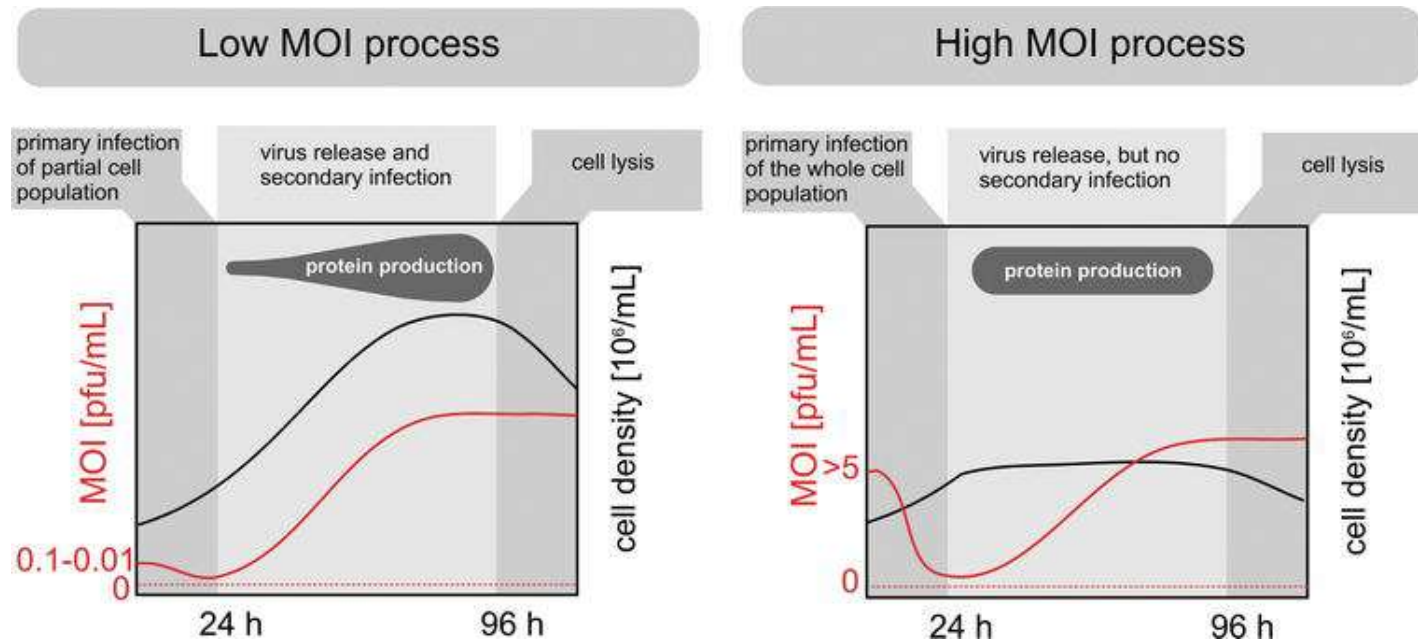
- Huge quantities of **polyhedrin** produced by baculoviruses are the fundamental features to develop the viruses as vectors for foreign protein production
- Later findings manifested that the polyhedrin protein is not necessary for baculovirus replication in infected insect cells
- So, the strategy for expressing recombinant protein by replacing viral DNA sequence encoding polyhedrin with the gene of interest is produced
- This theory became a reality when scientists performed homologous recombination of a polyhedrin region in the viral genome with a plasmid containing a foreign gene regulated by the polyhedrin promoter to produce a recombinant baculovirus, therefore triggering the birth of baculovirus-insect expression system



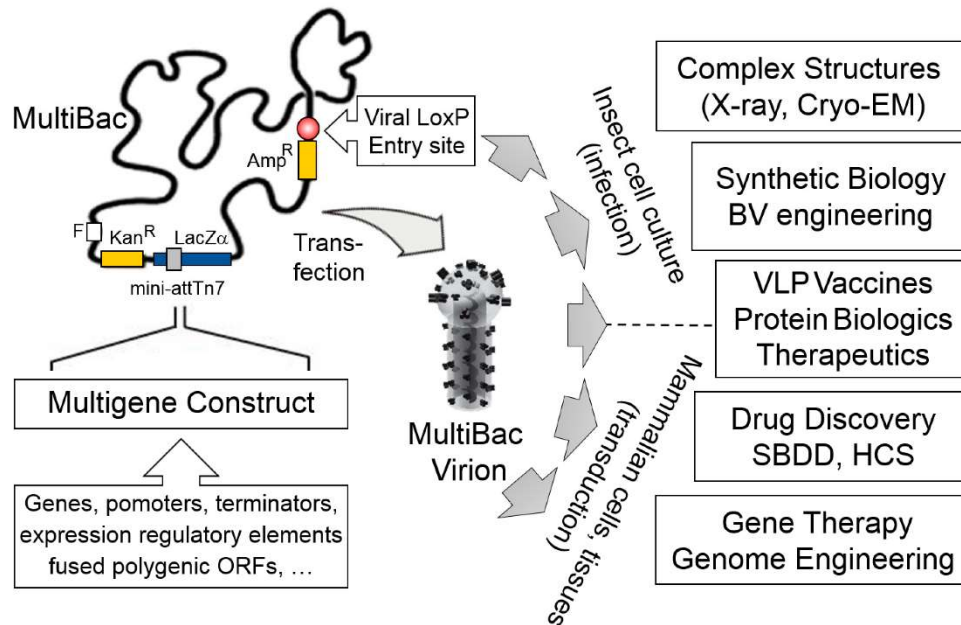
Workflow of protein production using BEVS



A comparison of BEVS processes with low and high MOI



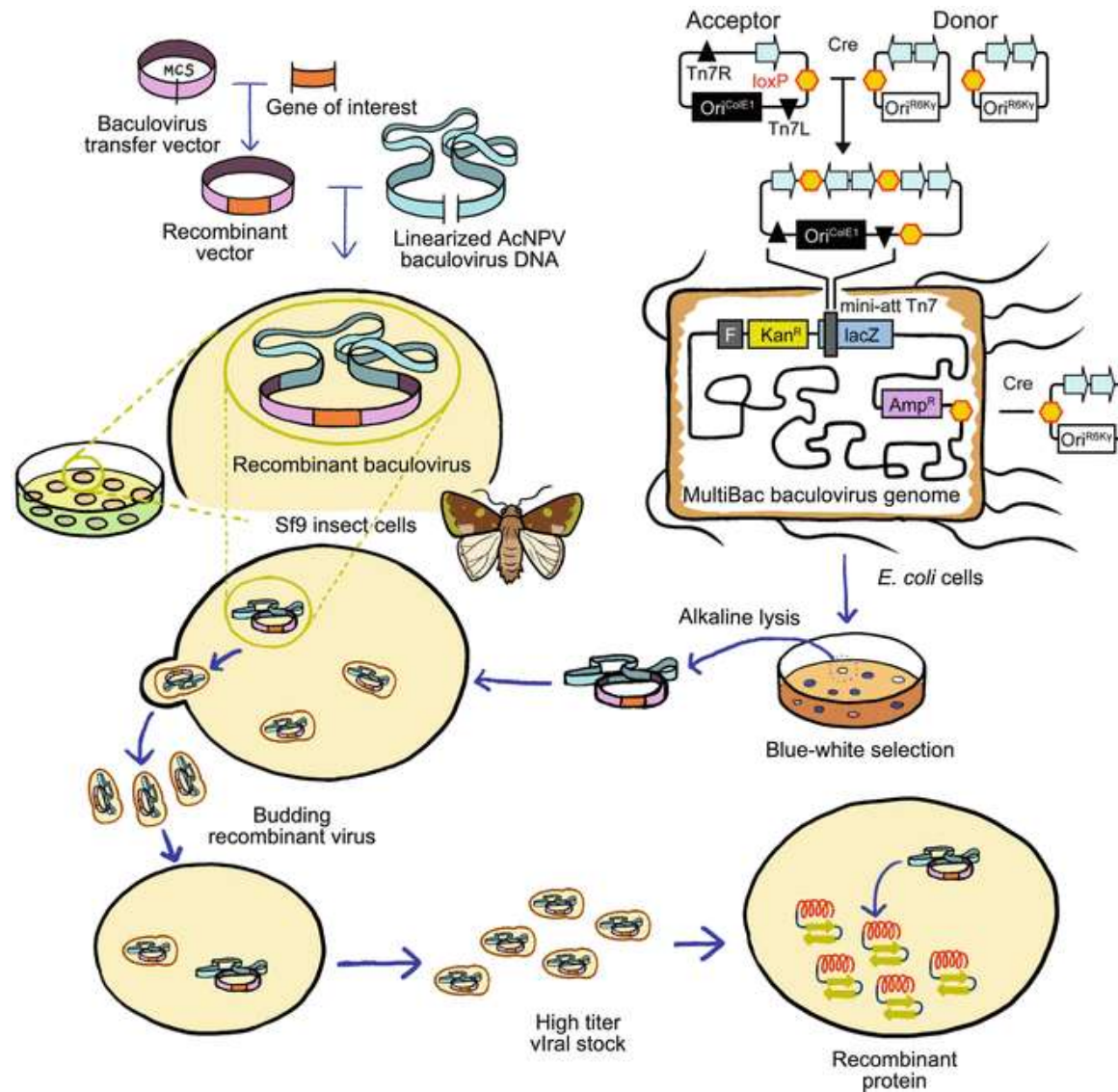
The MultiBac, a multi-expression system



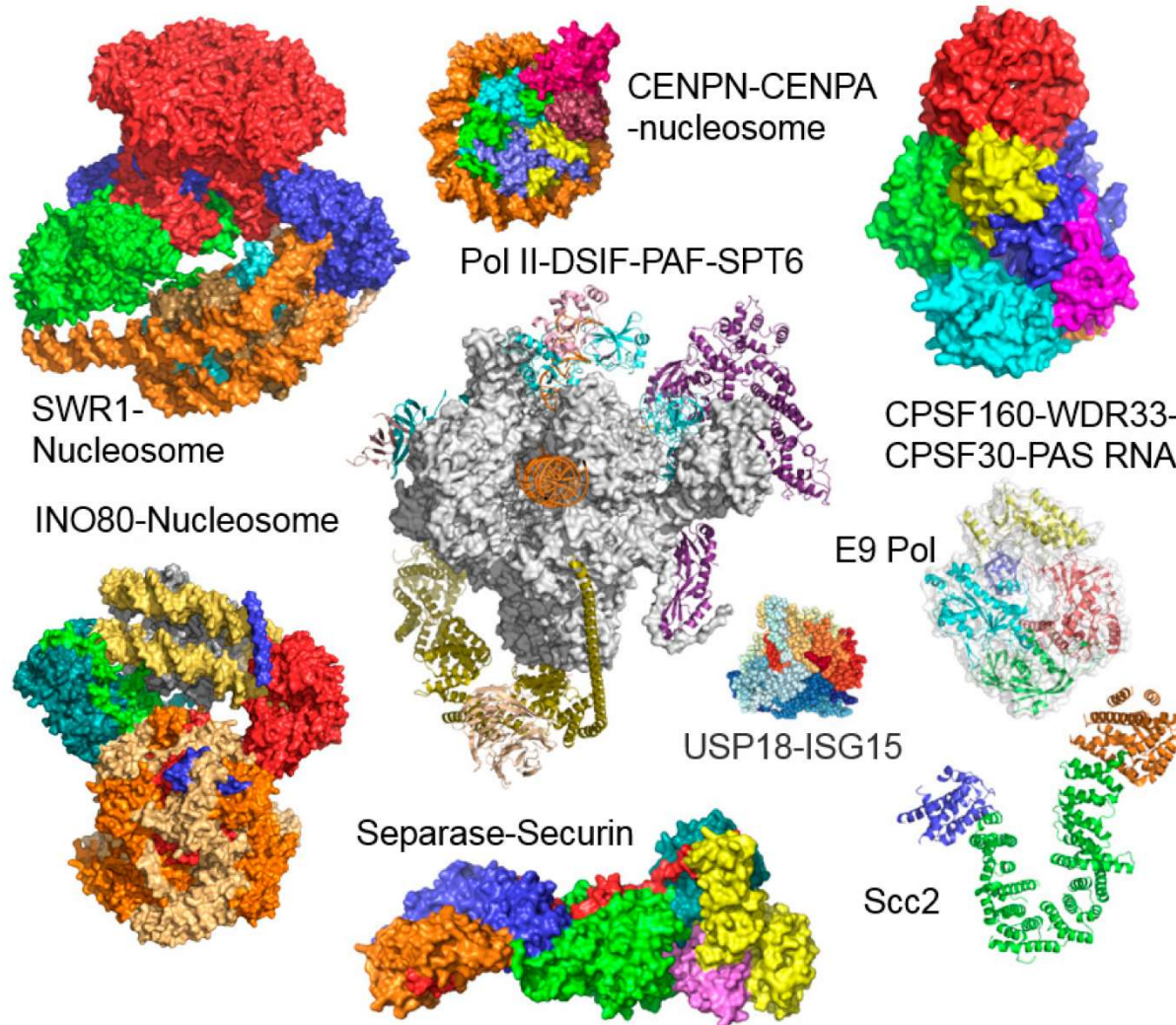
Prof. Imre Berger
 School of Biochemistry & Bristol Synthetic
 Biology Centre, University of Bristol, UK

- MultiBac consists of a baculoviral genome optimized for multigene delivery and protein complex expression (left).
- The genome is present as a bacterial artificial chromosome (BAC) in *E. coli* cells supplying the Tn7 transposase.
- Expression cassettes are assembled into the multi-gene expression constructs and inserted into the MultiBac genome by Tn7 transposition.
- A second entry option into the viral backbone is provided distal from the Tn7 site, relying on Cre recombinase catalysed site-specific integration into a LoxP sequence (circle filled in red).
- Composite MultiBac baculoviral DNA containing all DNA elements of choice is extracted from *E. coli* cultures, followed by transfection into insect cell cultures to manufacture functional MultiBac virions.
- These are then used for a wide range of applications (right), by the infection of insect cell cultures or transduction of mammalian cells, tissues and organisms.

Application of MultiBac system to large complexes

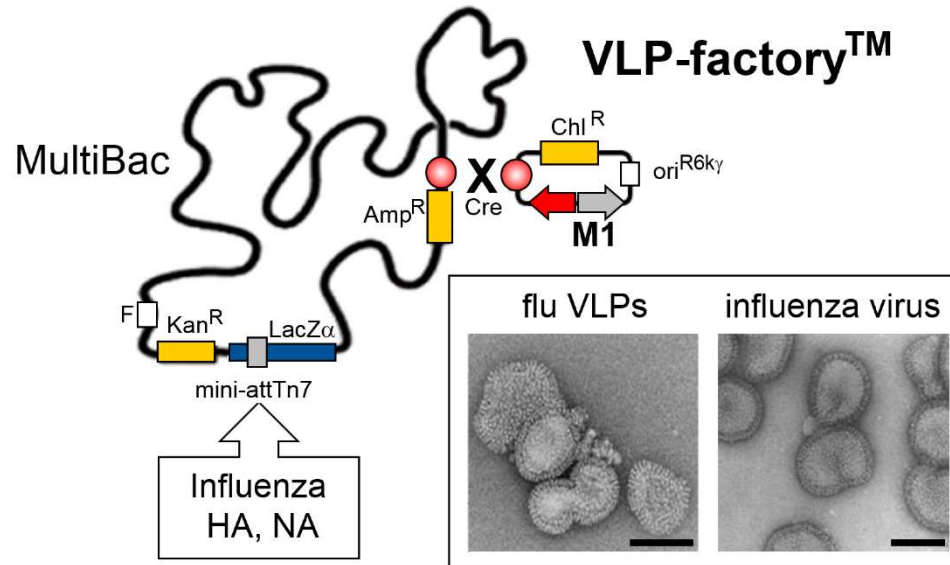


The MultiBac system: successful structural stories



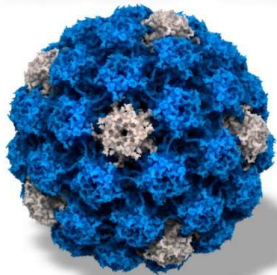
- Chromatin remodeling enzymes SWR1 (14 subunits) and INO80 (11 subunits)
- The yeast polII DSIF-PAF-Spt6 cryo-EM structure
- The CENPN-CENPA-nucleosome complex
- The human CPSF-160–WDR33–CPSF-30–PAS RNA quaternary complex
- The E9 polymerase
- The USP18-ISG15 complex
- The Separase-Securin complex
- The cohesion loader Scc

The MultiBac: a factory for synthetic virus-like particles



- A plasmid module comprising expression cassettes for the capsid-forming influenza H1N1 M1 protein (colored in grey) and a fluorescent protein marker, mCherry (colored in red), was introduced into the MultiBac baculoviral genome by Cre recombinase enzyme mediated plasmid fusion into the LoxP site (circle filled in red, gradient)
- Co-expression of HA, NA and M1 yields synthetic influenza virus-like particles (VLPs) resembling live influenza virus

WHAT YOU SHOULD KNOW ABOUT HPV VIRUS



Human papillomavirus-based vaccine produced by baculovirus expression system

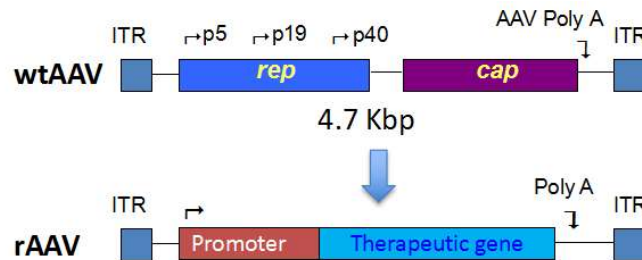
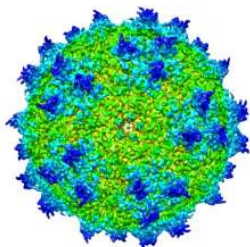
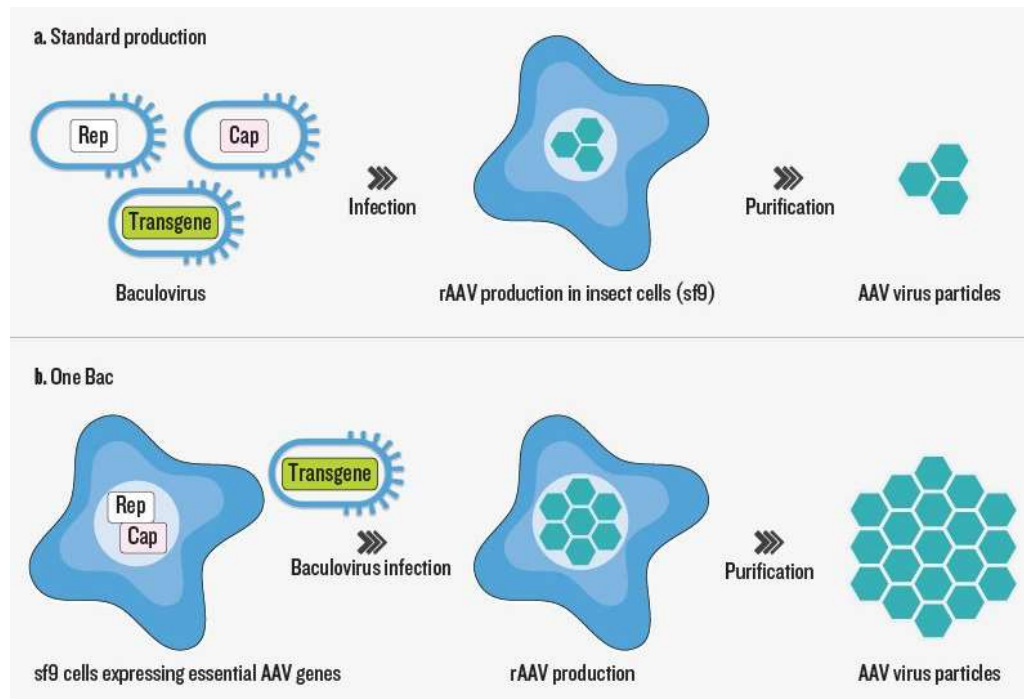


Selected vaccines produced by BEVS

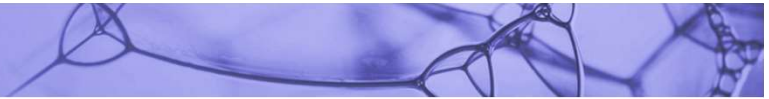
Application	Product name	Company	Stage	References
For human use				
Cervical cancer	CERVARIX [®]	GSK	Approved	[27]
Prostate cancer	PROVENGE [®]	Dendreon	Approved	[28]
Influenza	FluBlok [®]	Protein Sciences	Approved	[29, 30]
Influenza	A/H ₅ N ₁ Virus-like particle	Novavax	Phase I (NCT01596725)	[31]
For veterinary use				
Procrine circovirus 2 (PCV ₂)	Porcilis [®] PCV	Merck	Approved	[32]
PCV ₂	CircoFLEX [®]	Boehringer Ingelheim	Approved	[33]
Swine fever	Porcilis Pesti [®]	Merck	Approved	[34]

Baculovirus-mediated production of gene therapy vectors

Baculovirus-mediated production of adeno-associated viral (AAV) vectors as gene therapy vehicles



- In 2002, Prof. Robert Kotin and colleagues at the US National Heart, Lung, and Blood Institute first demonstrated its suitability for AAV manufacturing
- They infected Sf9 cell lines — derived from the fall armyworm — with three different baculoviruses: two containing essential genes for AAV particle production (rep and cap), and one containing the transgene sequence intended for delivery
- In this manufacturing process, the baculoviruses play a dual role, functioning as the ‘helper’ virus normally required for replication, as well as the vehicle for AAV genetic material
- In their initial demonstration, Kotin’s team achieved levels of productivity comparable with existing AAV manufacturing approaches — on the order of 50,000 functional viral particles per cell

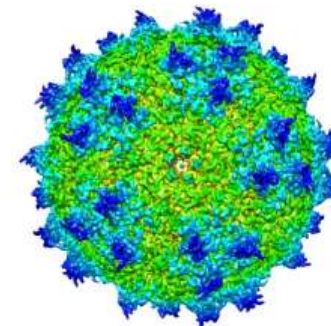


Baculovirus-mediated production of AAV vectors

Gene transfer and gene therapy are powerful approaches for many biological research applications and promising avenues for the treatment of many genetic or cancer diseases. The most efficient gene transfer tools are currently derived from viruses. Among them, the recombinant **adeno-associated viruses (AAVs)** are vectors of choice for many fundamental and therapeutic applications. The increasing number of clinical trials involving AAVs demonstrates the need to implement production and purification processes to meet the quantitative and qualitative demands of regulatory agencies for the use of these vectors in clinical trials. In this context, the rise of production levels on an industrial scale appeared essential. The introduction, in 2002, of an AAV process using a baculovirus expression vector system (BEVS) has circumvented this technological lock. The advantage of BEVS in expanding the AAV production in insect cells has been to switch the process to bioreactor systems, which are the ideal equipment for scaling up. We describe here a method for producing AAV vectors using the BEVS which can be easily used by research laboratories wishing to overcome the difficulties associated with the scaling up of production levels. The method provides sufficient quantities of AAV vectors to initiate preclinical projects in large animal models or for research projects where a single batch of vectors will consolidate the repeatability and reproducibility of in vitro and especially in vivo experimental approaches.

Development of AAV-based gene therapy products

- In 2017, FDA-approval of the first gene therapy product targeting a disease caused by mutations in a single gene
- This product, LUXTURNA™ (voretigene neparvovec-rzyl; Spark Therapeutics, Inc., Philadelphia, PA), delivers a normal copy of the *RPE65* gene to retinal cells for the treatment of *RPE65* mutation–associated retinal dystrophy, a blinding disease
- Many additional gene therapy programs targeting both inherited retinal diseases and other ocular diseases are in development



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LUXTURNA is the first FDA-approved
gene therapy for a genetic disease

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World's most efficient large-scale AAV manufacturing



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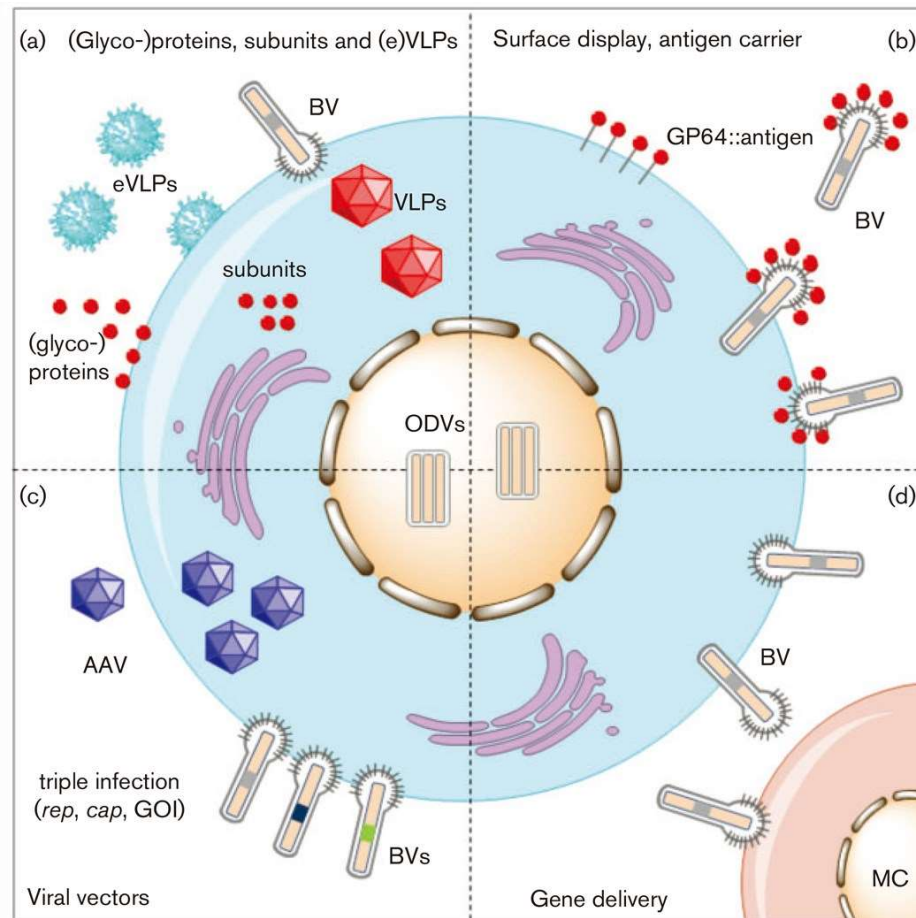
- Virovek has developed a patented BAC-to-AAV technology that utilizes the baculovirus expression system to produce AAV vectors in insect cells under serum-free condition
- The capability to generate over $3e+16$ vg of AAV vectors with a single production run, which is unmatched by any other AAV production system



**BAC-TO-AAV
TECHNOLOGY FOR
LARGE SCALE AAV
PRODUCTION**

[LEARN MORE](#)

Applications of BEVS



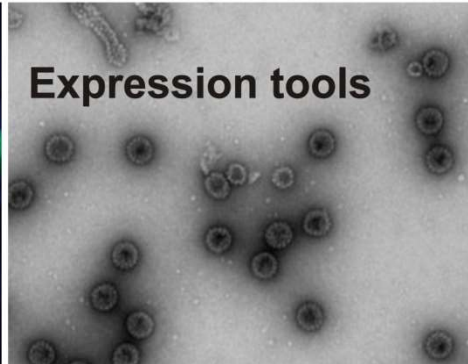
Overview of the various applications of the baculovirus expression system.

- Nowadays, baculovirus-insect expression system has been relatively mature in many labs for protein production and is becoming a leading method to produce high quality proteins among the eukaryotic expression systems
- The use of BEVS to express the protein of interest requires two steps. In the first step, the insect cells are cultured to the desired concentration. In the second step, the baculovirus is applied to infect the insect cells. The virus that infects host cells will dominate the gene expression mechanism in the host cell and trigger the production process of the target protein
- Yet, several mature expressing vectors have been developed, including *Autographa californica* multiple nucleopolyhedrovirus (AcMNPV) that is the most extensively used
- The ability to produce biologically active mammalian proteins makes the BEVS a powerful tool over yeast and bacterial expression systems

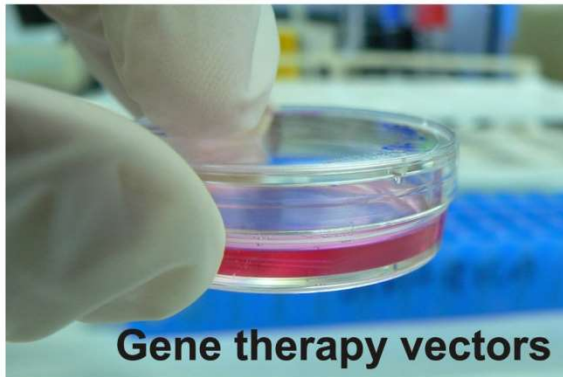
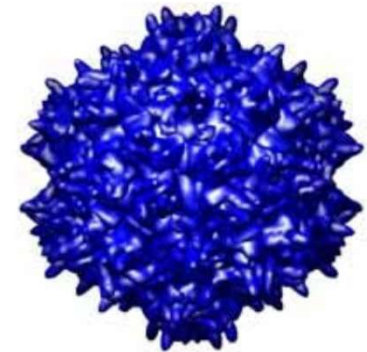
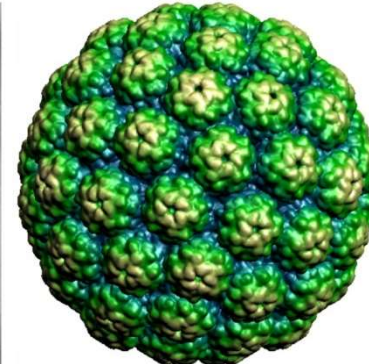
Applications of baculovirus technologies



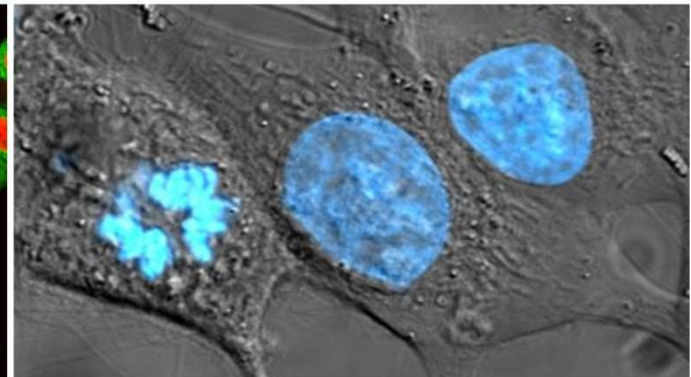
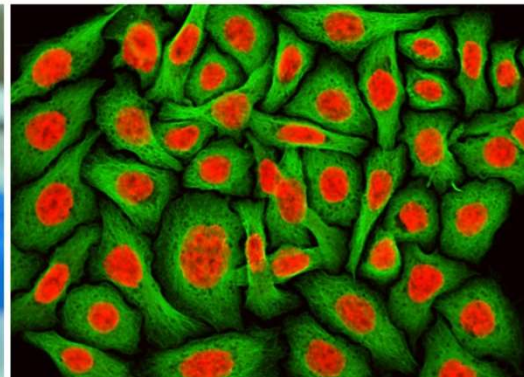
Biopesticides



Expression tools



Gene therapy vectors





Advantages and drawbacks of baculovirus expression

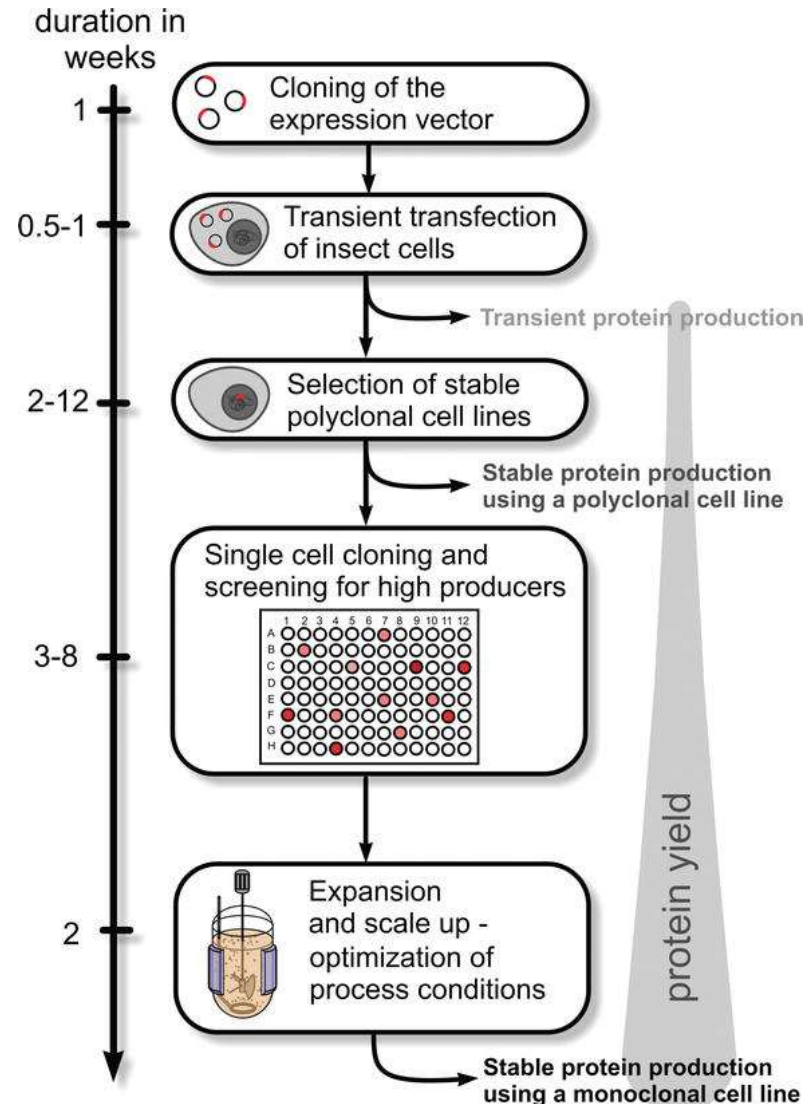
ADVANTAGES

- Recombinant protein has complete biological function, such as protein correct folding and disulfide bond
- Post-translation modification (glycosylation etc.)
- High level of expression, up to 50% of the total protein amount
- Accommodate large insert protein
- Simultaneously express multiple genes (multi-expression)

DRAWBACKS

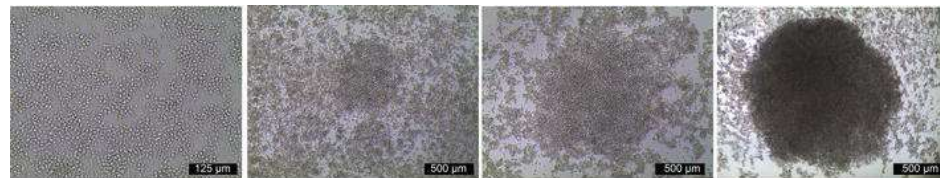
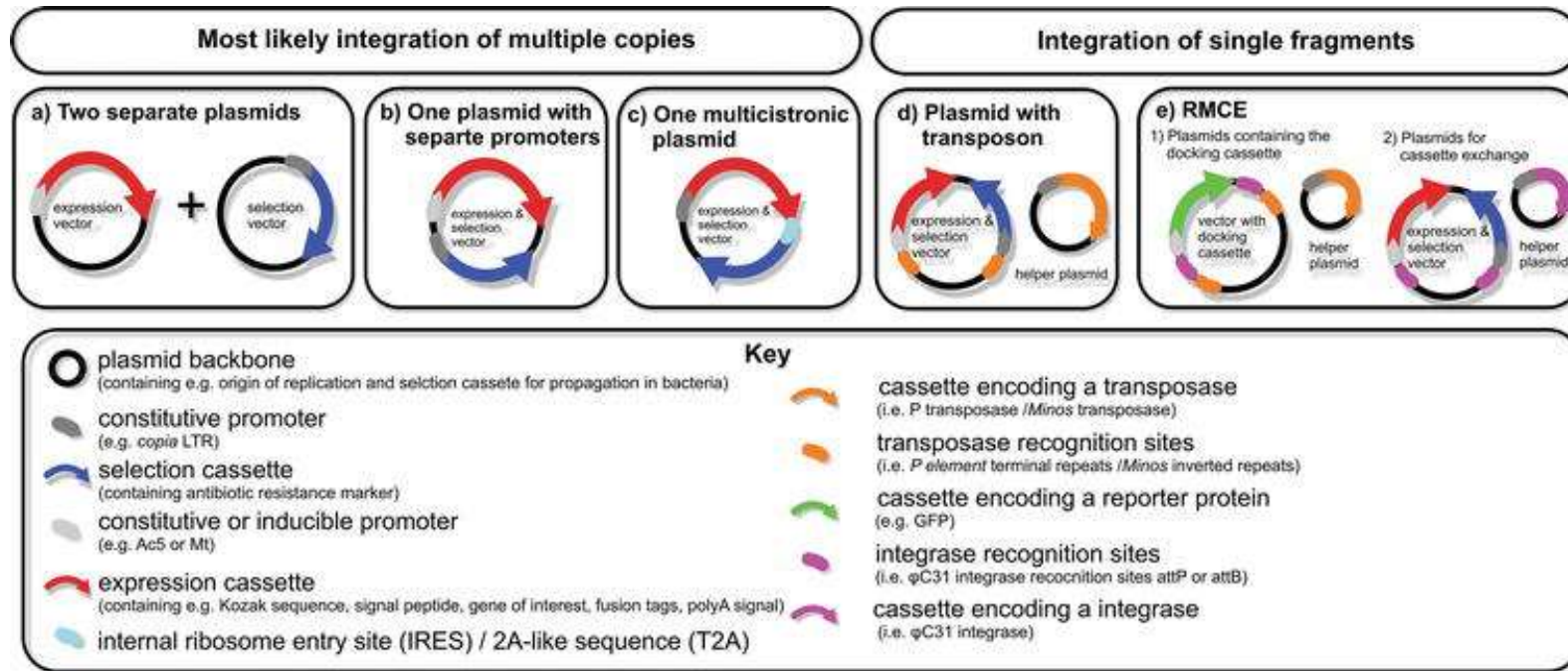
- The drawback is that exogenous protein expression is under control of the very late viral promoter, where the cells begin to die due to viral infection
- Insect expression system is normally used for production of membrane proteins, although the glycosylations may be different from those found in vertebrates
- In general, it is safer to use than mammalian virus, since it has limited host range and does not infect vertebrates without modifications.

Virus-free expression in insect cells



- Overview of the general procedure to produce stably transformed *D. melanogaster* S2 cell lines for recombinant protein expression
- Protein expression can be initiated at different points, starting with transient expression immediately after transfection followed by stable expression in a polyclonal cell line and finally the selection of a highly productive monoclonal cell line

The different plasmid sets to generate stable S2 cell lines



day 0 → day 20

Limiting dilution cloning

Co-cultivation of untransfected feeder cells with approximately one clone of the stable cell population in 96 well format

Start of selection

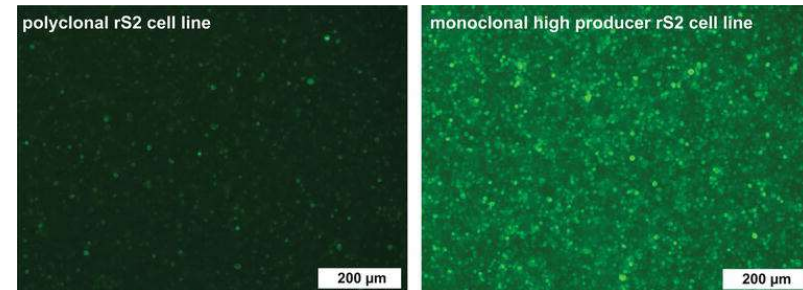
Addition of selection antibiotic to kill untransfected feeder cells and induce colony growth

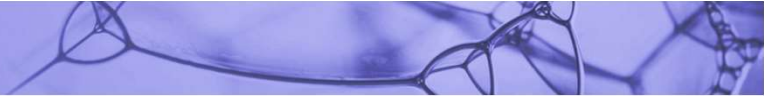
Maintaining selection pressure

Feed or medium exchange to keep the cells under selective pressure and maintain sufficient nutrient supply

Pick and expansion

Most of the feeder cells are dead and macroscopically visible monoclonal colonies can be picked and used for subsequent upscaling under selection pressure



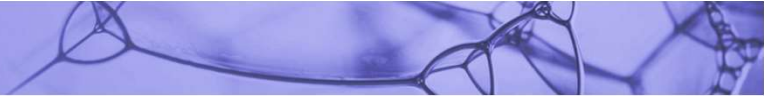


Expression in mammalian cells

Concepts

Methods

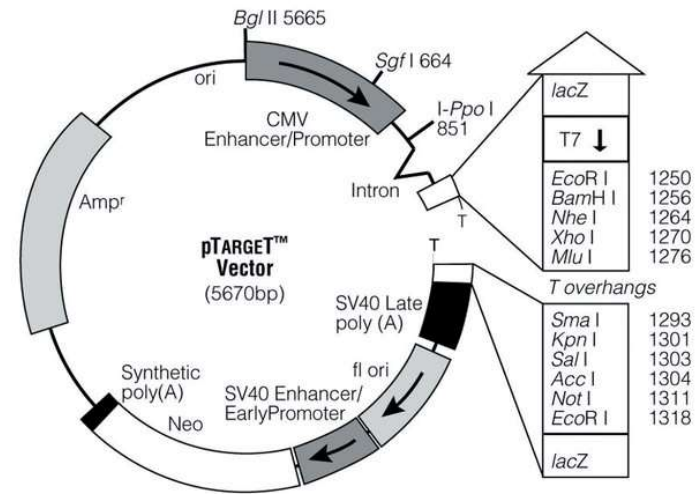
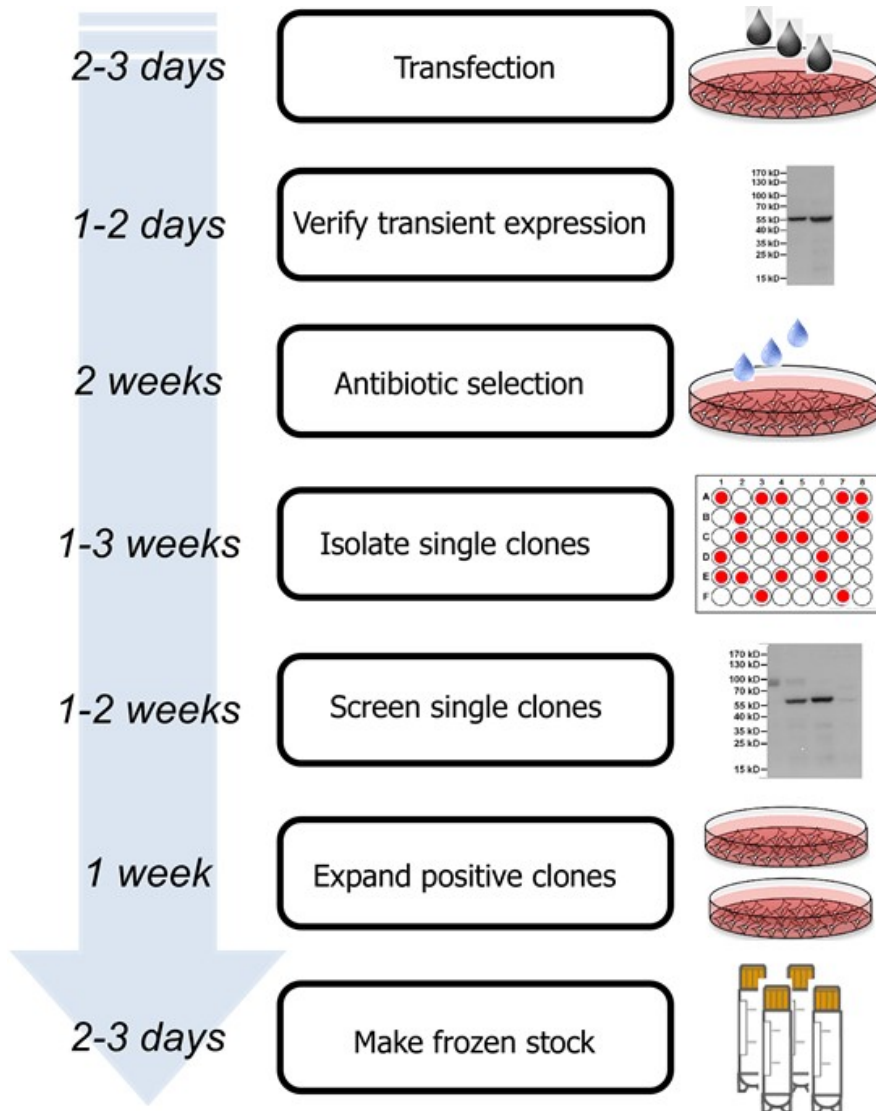
Applications

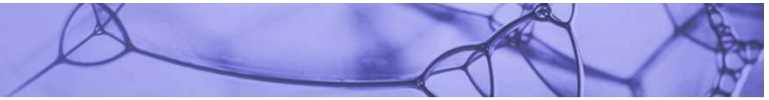


Mammalian expression system

- Recombinant proteins expressed in mammalian cell commonly use **plasmid transfection** and **viral vector infection**. Stable cell line using plasmid transfection takes several weeks or even months, while the virus vector can quickly infect cells within a few days
- Depending on the temporal and spatial differences in protein expression, the expression system can be divided into **transient**, **stable** and **induced expression** systems
- Transient expression system refers that host cell cultures without selection pressure and exogenous vector gradually lost while cell division. The target protein expression duration is short. The advantage of transient expression system is simple and short experimental period
- Stable expression system means that the carrier DNA replicate and express long time stably in host cell. Due to the need of select resistance and pressure steps, stable expression is relatively time-consuming and laborious
- Induction expression system refers that the target gene begins to express when induced by foreign small molecules. The use of heterologous promoters, enhancers and amplifiable genetic markers can increase protein production
- The mammalian expression system has unique advantage in protein initiation signals, processing, secretion, glycosylation and is suitable for expressing intact macromolecules

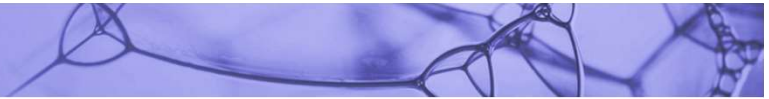
Mammalian stable cell line generation





Expression in *Leishmania tarentolae*

Concepts
Methods
Applications



LEXSY - Eukaryotic protein expression in *L. tarentolae*

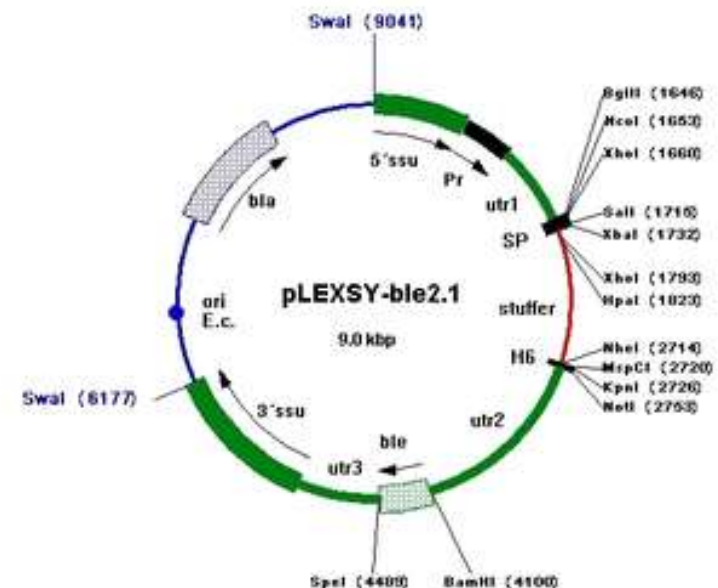
The unicellular kinetoplast protozoan *Leishmania tarentolae*, isolated from the Moorish gecko *Tarentola mauritanica*, not pathogenic to mammals (Biosafety level 1) – was turned into the protein-producing host of our eukaryotic protein expression system LEXSY:

- eukaryotic host as easy to handle as *E. coli*: no specific labware, no cell biology equipment required
- fully eukaryotic protein expression machinery with post-translational modifications, including glycosylation and disulfide bond formation
- shuttle vectors: cloning in *E. coli*, expression in LEXSY host
- constitutive or inducible, intracellular or secretory expression of target proteins
- stable expression strains for constant protein production

Leishmania tarentolae: a little-known expression host



- *Leishmania tarentolae*, which can be easily cultured and genetically manipulated, is harmless (unless you are a gecko from Africa)
- It is closely related to other kinetoplastida, including important human pathogens such as *L. donovani* (causing kala azar), *Trypanosoma brucei* (sleeping sickness) and *T. cruzi* (Chagas disease)

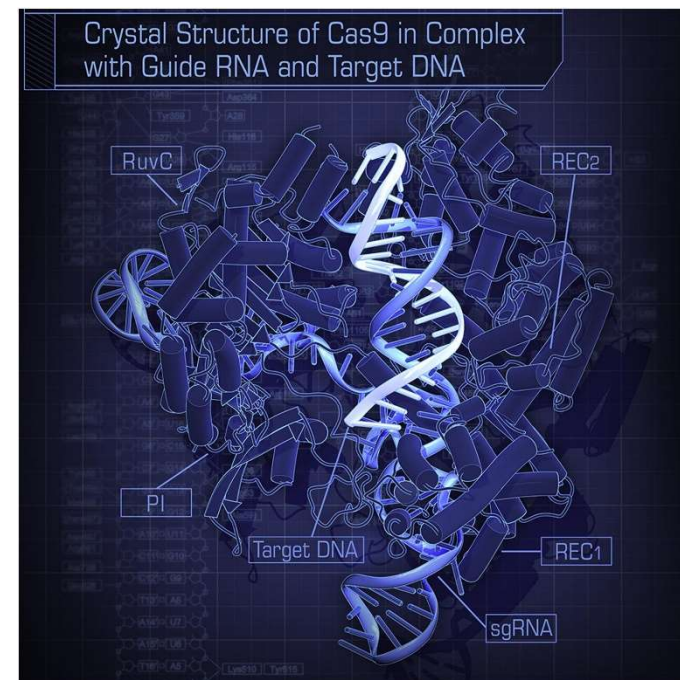
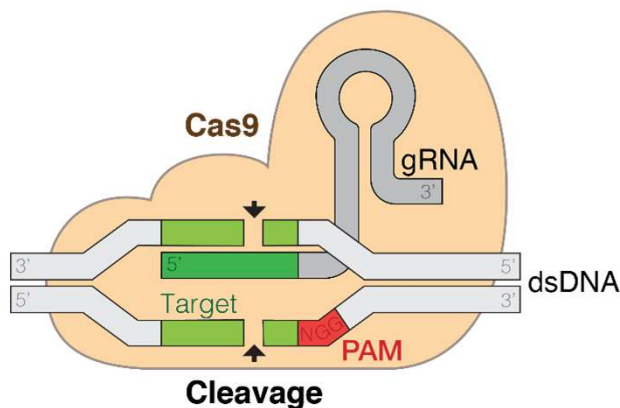


Emerging new expression tools

Concepts
Methods
Applications

The CRISPR/Cas9 system

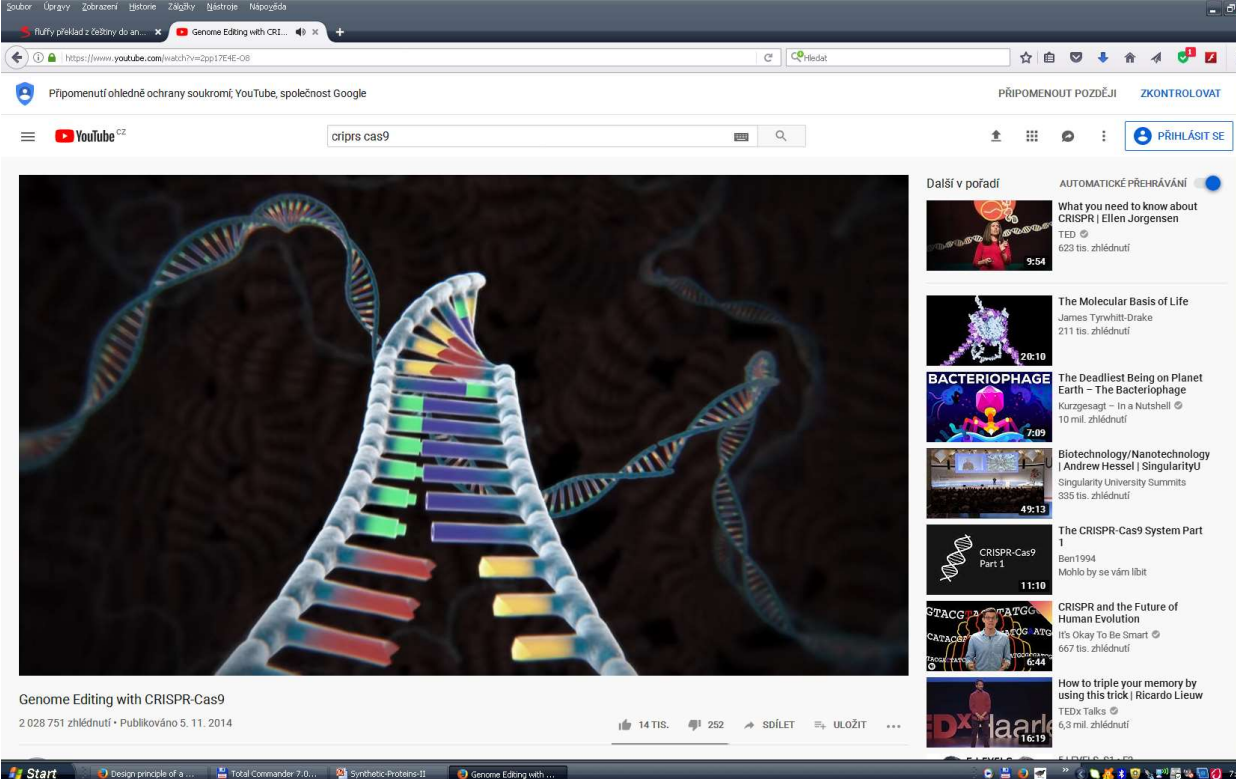
- The **Cas9** (CRISPR associated protein 9) is a protein which plays a vital role in the immunological defense of bacteria against DNA viruses, and which is used in genetic engineering. Its main function is to cut DNA and therefore it can alter a cell's genome
- Structurally, Cas9 is an **RNA-guided DNA endonuclease enzyme** associated with CRISPR (Clustered Regularly Interspaced Short Palindromic Repeats) adaptive immunity system in *Streptococcus pyogenes*
- Cas9 performs this by unwinding foreign DNA and checking for sites complementary to the 20 bp spacer region of the guide RNA
- If the DNA substrate is complementary to the guide RNA, Cas9 cleaves the invading DNA



Crystal structure of *S. pyogenes* Cas9 in complex with sgRNA and its target DNA at 2.5 Å resolution, Nishimazu *et al.*, Cell 156: 935–49 (2014)

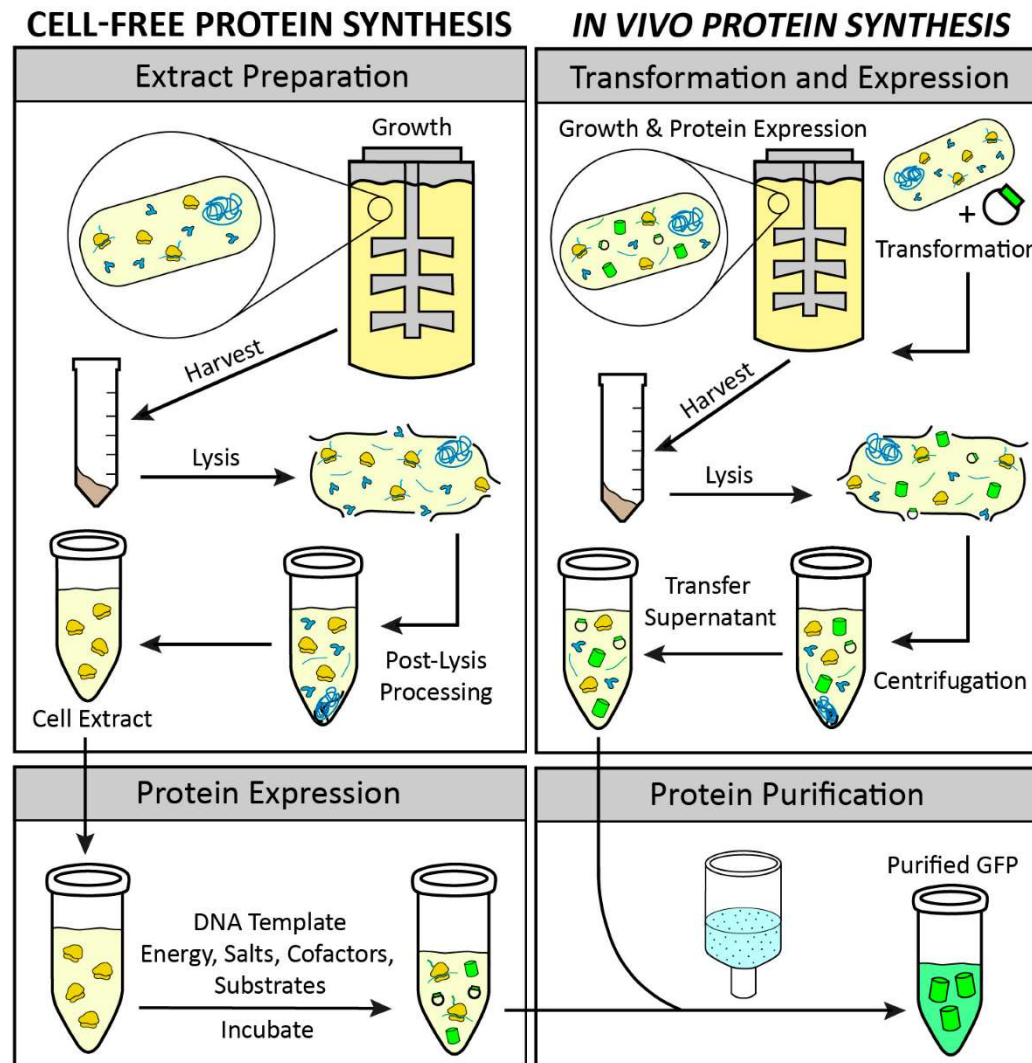
The CRISPR/Cas9 system on YouTube

<https://www.youtube.com/watch?v=bXnWIk8FgKc>
<https://www.youtube.com/watch?v=OjNrbPMXyMA>
<https://www.youtube.com/watch?v=0dRT7slyGhs>
<https://www.youtube.com/watch?v=2pp17E4E-O8>



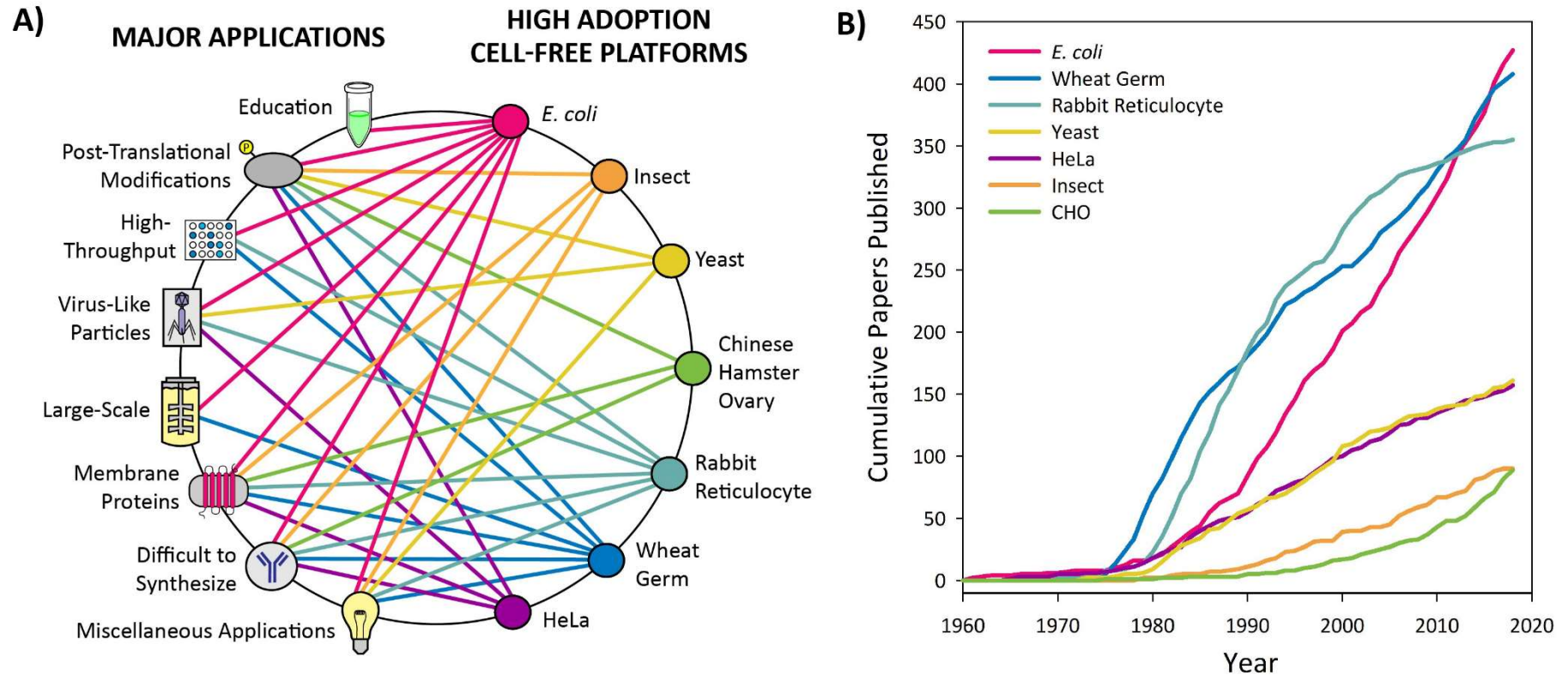
The screenshot shows a web browser window displaying a YouTube video. The video player is the central focus, showing a 3D model of a DNA double helix with a CRISPR-Cas9 system integrated into it. The video title is "Genome Editing with CRISPR-Cas9" and it has 2,028,751 views. The browser's address bar shows the URL "https://www.youtube.com/watch?v=2pp17E4E-O8". The YouTube interface includes a search bar with "crispr cas9" entered, a "PŘIHLÁŠIT SE" button, and a list of recommended videos on the right side. The Windows taskbar is visible at the bottom of the screen.

Cell-Free Protein Synthesis (CFPS)



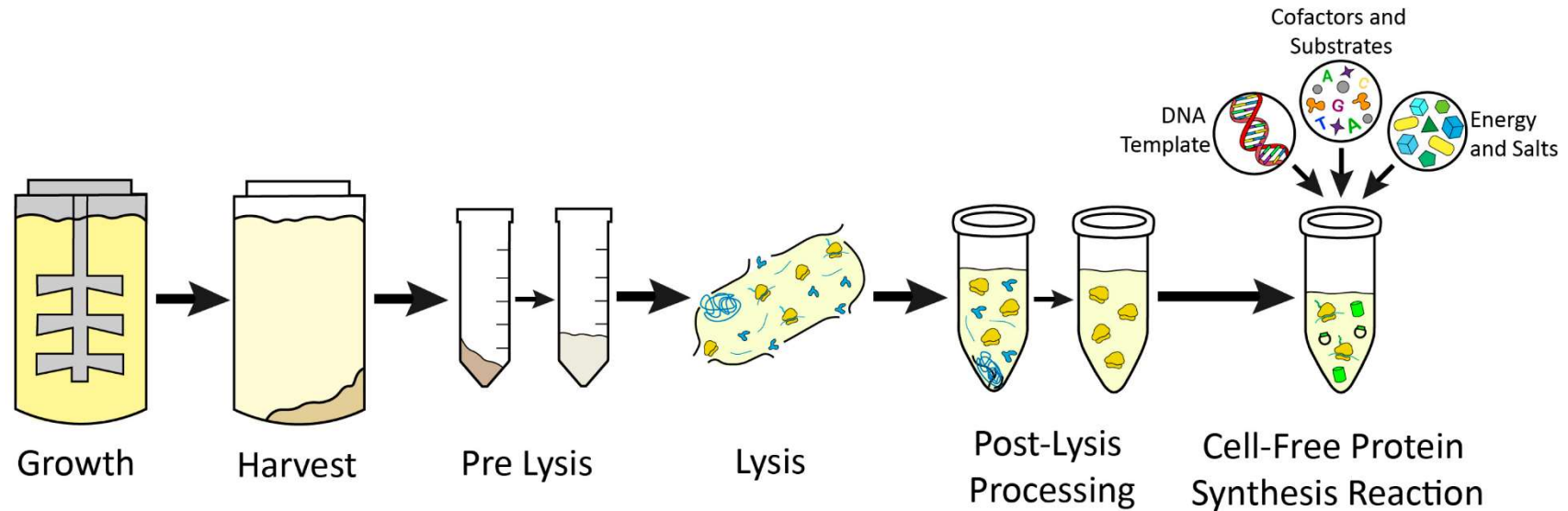
- Cell-free protein synthesis (CFPS) is a platform technology that provides new opportunities for protein expression
- The advantages of CFPS over in vivo protein expression include its open system, the elimination of reliance on living cells, and the ability to focus all system energy on production of the protein of interest

Cell-free platforms and their applications



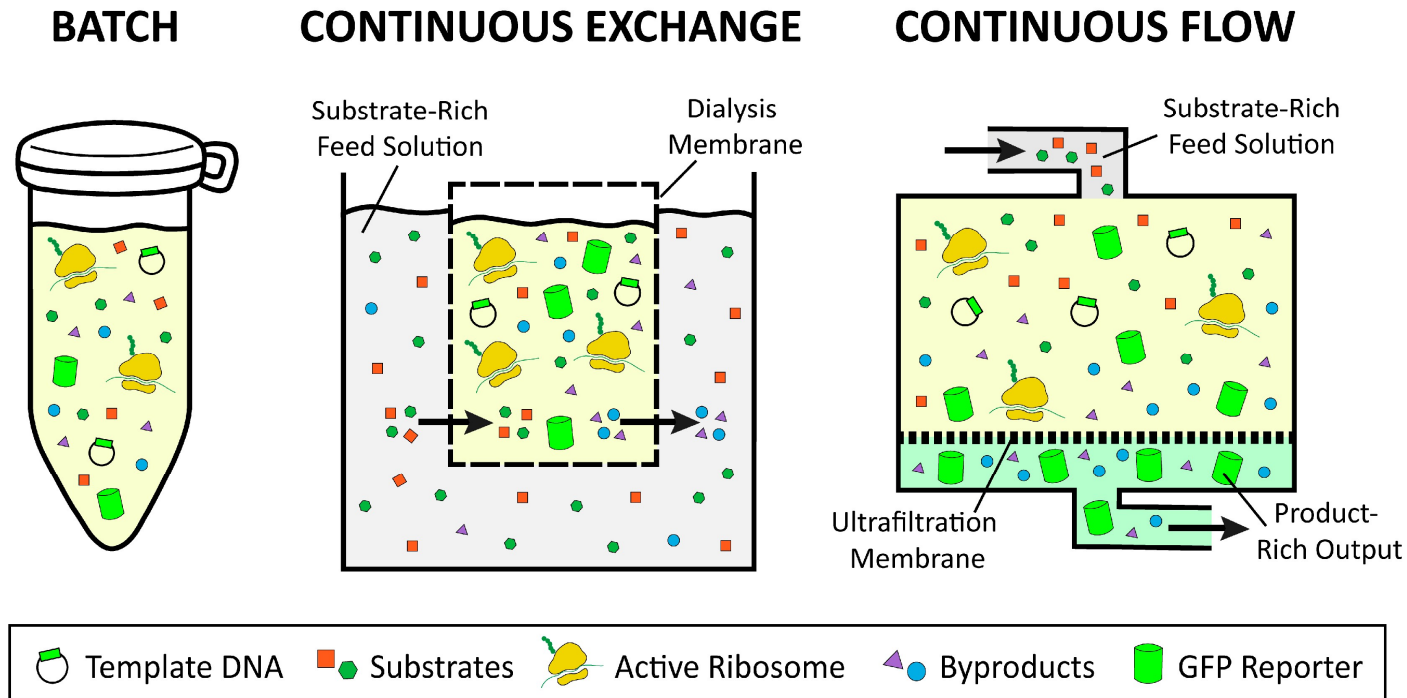
- (A) Web of the applications enabled by low adoption cell-free platforms. Connections shown are based on applications that have been published or that have been proposed in publications.
- (B) Cumulative number of peer-reviewed publications over the last 60 years for cell-free platforms.

Preparation of cell-free extract and set-up of CFPS reactions



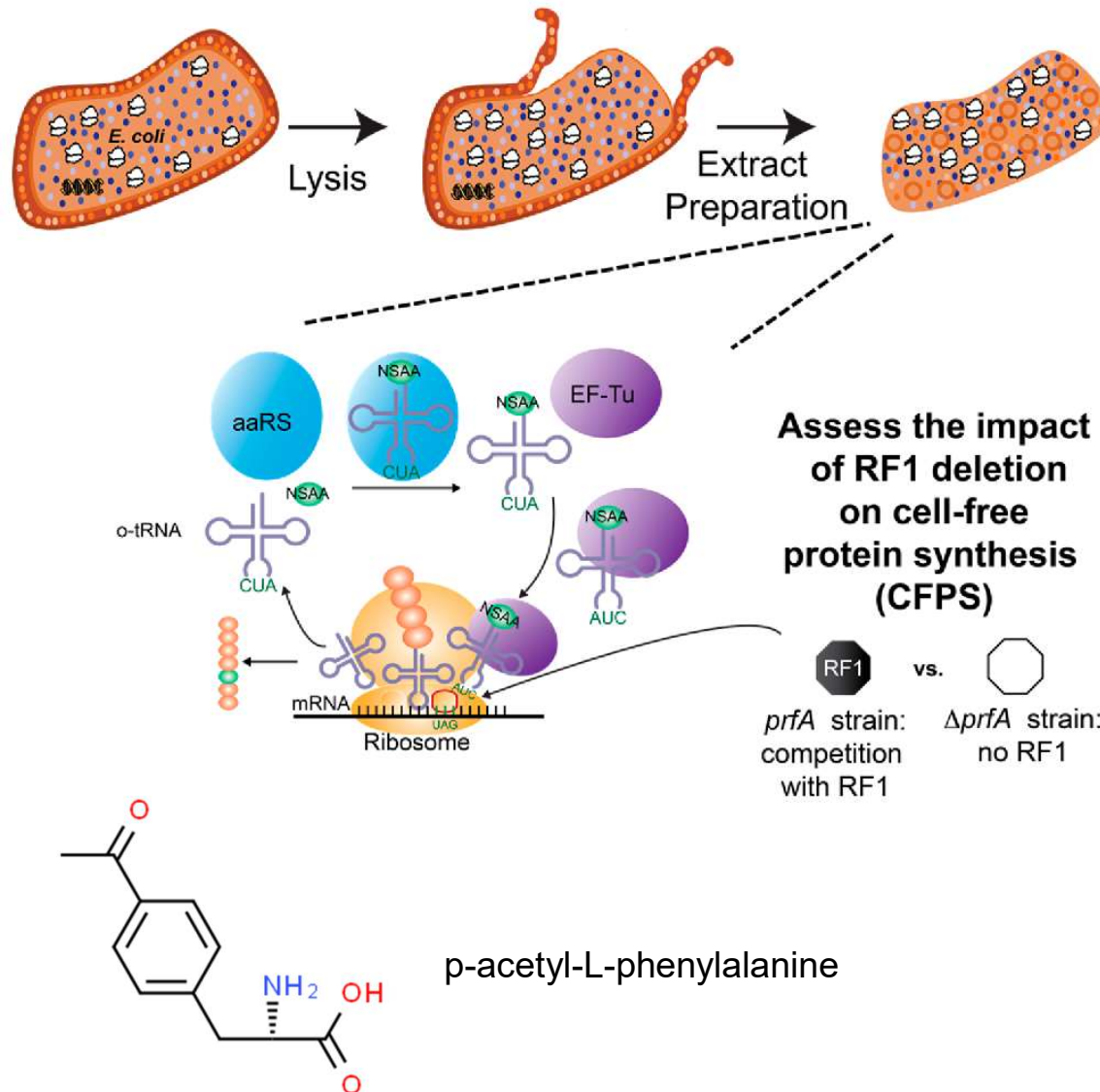
General workflow for preparation of cell-free extract and set up of CFPS reactions. A visualization from cell growth to the CFPS reaction is depicted above for a new user, highlighting the main steps involved.

Comparison of CFPS platforms



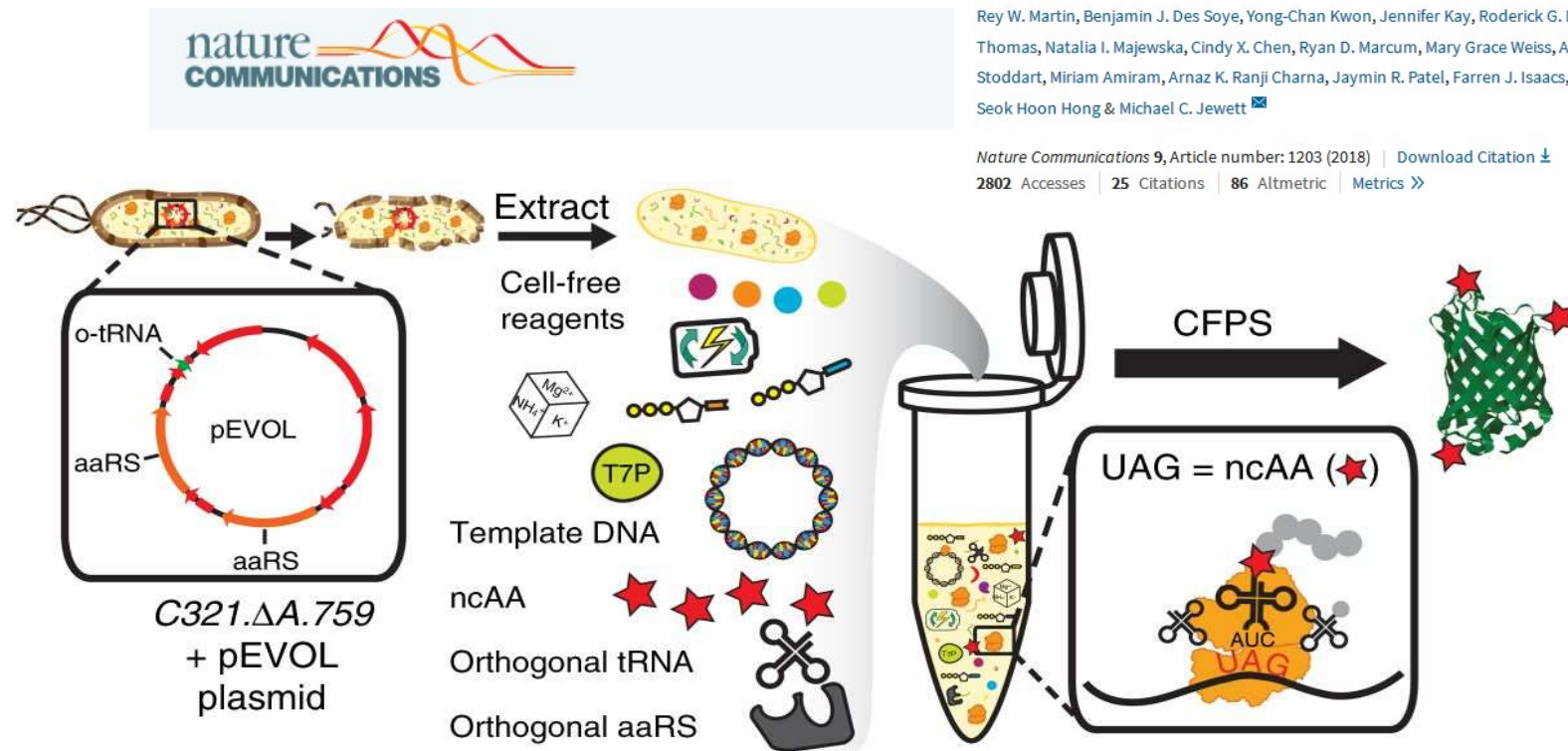
- Batch reactions contain all the necessary reactants within a single reaction vessel.
- Continuous exchange formats utilize a dialysis membrane that allows reactants to move into the reaction and byproducts to move out, while the protein of interest remains in the reaction compartment.
- Continuous flow formats allow a feed solution to be continuously pumped into the reaction chamber while the protein of interest and other reaction byproducts are filtered out of the reaction.

CFPS reaction incorporating nonstandard amino acids

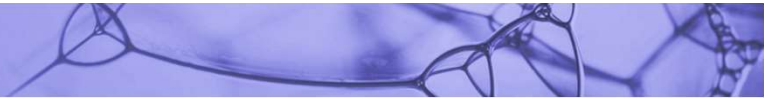


- Scheme of cell-free protein synthesis reaction incorporating nonstandard amino acid to investigate the effect of RF1
- Cell extracts containing transcription and translation machinery are prepared from rEc.E13 or rEc.E13. $\Delta prfA$ strains
- Plasmid DNA template of sfGFP containing single or multiple **amber codon** sites, **orthogonal tRNA/aaRS, NSAA**, T7 RNA polymerase, and other cofactors are added as necessary to activate the cell-free protein synthesis (CFPS) reaction

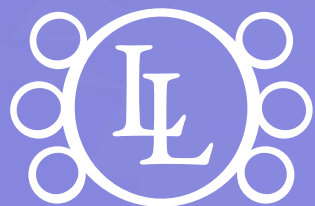
CFPS from extracts of a genomically recoded organism



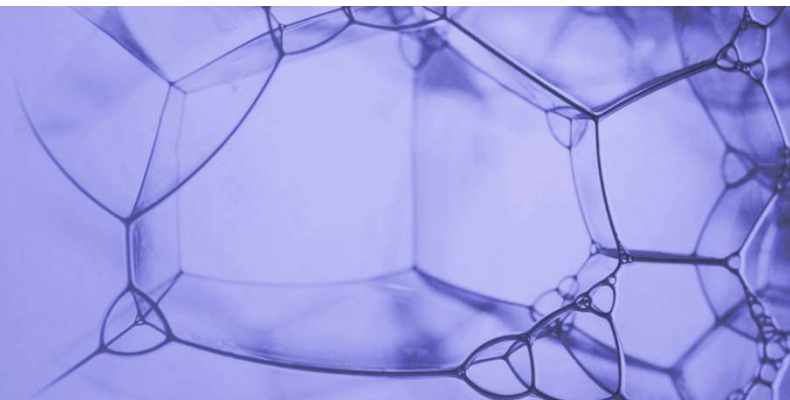
- Schematic of the production and utilization of crude extract from genomically recoded organisms with plasmid overexpression of orthogonal translation components for cell-free protein synthesis
- CFPS reactions are supplemented with the necessary substrates (e.g., amino acids, NTPs, etc.) required for in vitro transcription and translation as well as purified orthogonal translation system (OTS) components to help increase the ncAA incorporation efficiency
- aaRS, aminoacyl tRNA synthetase; ncAA, non-canonical amino acid; T7P, T7 RNA polymerase; UAG, amber codon



Questions



LOSCHMIDT
LABORATORIES



Dr. Martin Marek
Loschmidt Laboratories
Faculty of Science, MUNI
Kamenice 5, bld. A13, room 332
martin.marek@recetox.muni.cz