# From discovery to technology explosion

1868: Discovery of DNA

1953: Watson and Crick propose double helix structure

1977: Sanger sequencing

1985: PCR

2000: Working draft human genome announced (Sanger method)

2005: 454 sequencer launch

(pyrosequencing)

2006: Genome Analyzer launched

(Solexa sequencing)

2007: SOLiD launched

(ligation sequencing)

2009: Whole human genome no longer merits Nature/Science

paper

2010: "third-gen" systems

\$ human Genome

\$3 billion

\$2-3 million

\$250k

\$50k

\$20k

<\$1k

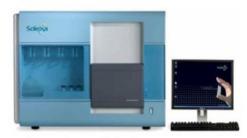




Applied Biosystems ABI 3730XL 1 Mb / day



Roche / 454 Genome Sequencer FLX 100 Mb / run



Illumina / Solexa Genetic Analyzer 2000 Mb / run



Applied Biosystems SOLiD 3000 Mb / run





### 





#### Sanger Sequencing Next generation sequencing A fragment gDNA **PCR** Dideoxy sequencing reaction | ligate adapters ddA dd-(G)dNTPs: ddT ddC ddNTPs: ddG adapters fragmented DNA ddprimer sequencing library denatured template DNA polymerase Capillary electrophoresis **C** Cluster generation clusters longest fragment CTTGGATGACATshortest fragment **D** Sequencing Sequencing read-out 1 read/sample 40 x 106 reads/sample\*

Sequencer	454 GS FLX	HiSeq 2000	SOLiDv4	Sanger 3730xl
Sequencing mechanism	Pyrosequencing	Sequencing by synthesis	Ligation and two-base coding	Dideoxy chain termination
Read length	700 bp	50SE, 50PE, 101PE	$50 + 35 \mathrm{bp}$ or $50 + 50 \mathrm{bp}$	400~900 bp
Accuracy	99.9%*	98%, (100PE)	99.94% *raw data	99.999%
Reads	1 M	3 G	$1200 \sim 1400 \mathrm{M}$	_
Output data/run	0.7 Gb	600 Gb	120 Gb	1.9~84 Kb
Time/run	24 Hours	3~10 Days	7 Days for SE 14 Days for PE	20 Mins~3 Hours
Advantage	Read length, fast	High throughput	Accuracy	High quality, long read length
Disadvantage	Error rate with polybase more than 6, high cost, low throughput	Short read assembly	Short read assembly	High cost low throughput

# Oxford Nanopore

Application Specific **Generic Platform** Electronic read-out system

# **DNA** degradation

Mechanical damage during tissue homogenization.

Wrong pH and ionic strength of extraction buffer.

Incomplete removal / contamination with nucleases.

Phenol: too old, or inappropriately buffered (pH 7.8 – 8.0); incomplete removal.

Wrong pH of **DNA solvent** (acidic water).

Recommended: 1:10 TE for short-term storage, or 1xTE for long-term storage.

Vigorous pipetting (wide-bore pipet tips).

**Vortexing** of DNA in high concentrations.

Too many **freeze-thaw** cycles (we tested 5, still Ok).

Debatable: sequence-dependent

#### Illumina, Inc. (ILMN)

Summary

☆ Add to watchlist

**Historical Data** 

NasdaqGS - NasdaqGS Real Time Price. Currency in USD

**342.68** +19.95 (+6.18%)

Conversations

Statistics

At close: November 7 4:00PM EST

Chart

Buy	Sell
-----	------

Profile

Financials

Analysis

Previous Close	322.73	Market Cap	50.374B
Open	325.00	Beta (3Y Monthly)	2.04
Bid	304.93 x 800	PE Ratio (TTM)	77.92
Ask	350.00 x 800	EPS (TTM)	4.40
Day's Range	318.38 - 343.04	Earnings Date	Jan 28, 2019 - Feb 1, 2019
52 Week Range	203.83 - 372.61	Forward Dividend & Yield	N/A (N/A)
Volume	1,273,158	Ex-Dividend Date	N/A
Avg. Volume	1,019,553	1y Target Est	359.75



**Options** 

Holders

Sustainability

Trade prices are not sourced from all markets

### Genome sequencing

### Two strategies

- Whole genome shotgun (bottom-top)
- Clone-by-clone (top-bottom)



### Sequencing without a limit?

 A rapid progress in next generation sequencing technologies promises to provide complete (reference) DNA sequences







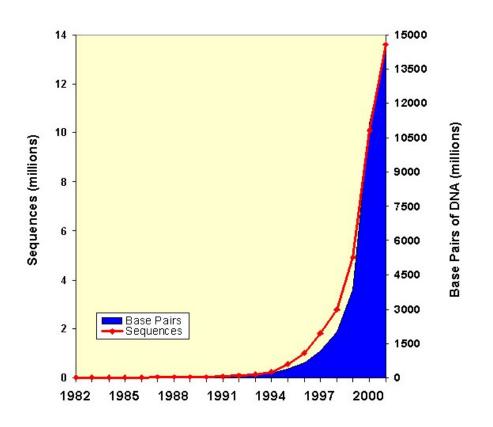


#### The bottleneck:

- NOT the sequencing capacity
- BUT the ability to assemble many short reads with prevalence of repeated DNA (and polyploidy)

# Genome sequencing

### **GenBank 1982 Los Alamos Sequence Database**





Walter Goad

# Frederick Sanger

1958 – Nobel prize – insuline structure

1975 - Dideoxy sequencing method

1977 – Φ-X174 (5,368 bp) sequence

1980 - second Nobel prize

λ phage sequence shotgun method (48,502 bp)



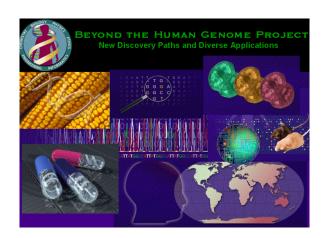
# Genome sequencing

• 1986 Leroy Hood: automatic sequencing machine



Leroy Hood

• 1986 Human Genome Initiative



# Genome sequencing

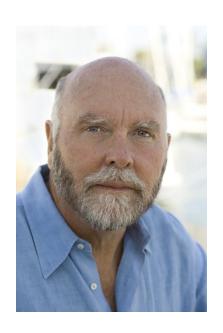
 1995 John Craig Venter first bacterial genome



John Craig Venter

### **Craig Venter**

### Global Ocean Sampling Expedition

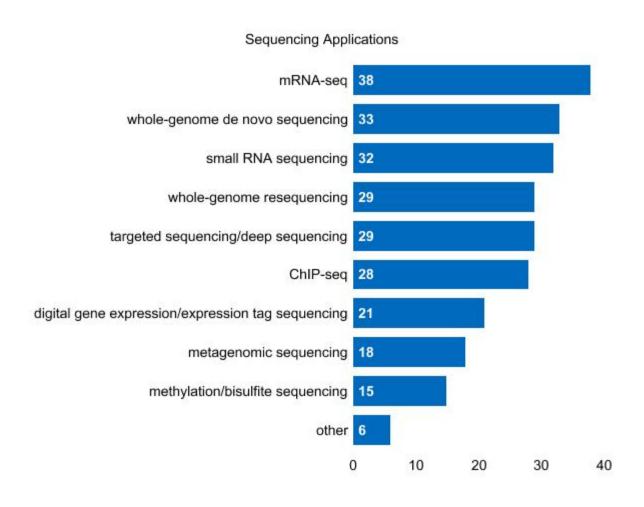


Synthetic genomics

Human Longevity Inc

http://www.youtube.com/watch?v=J0rDFbr hjtl

# Which applications are labs performing?



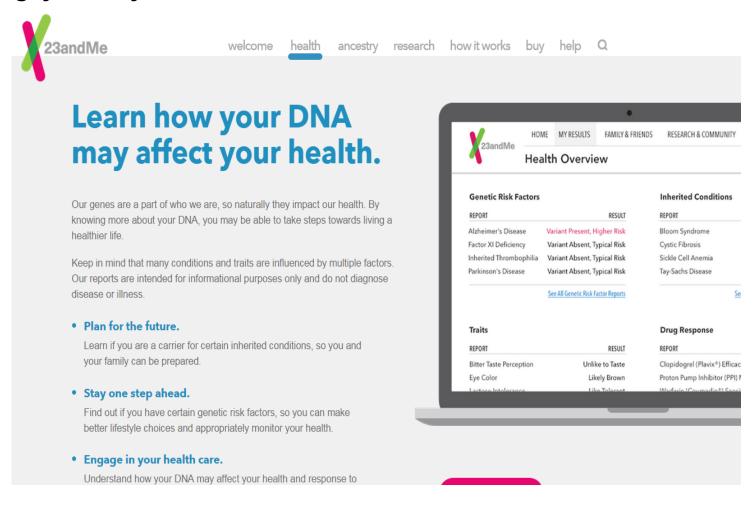
### 2010 Human genome reference

### 2010 Human genome reference



### 23andme (30% GSK)

# Anne Wojcicki CEO - manželka spoluzakladatele Google Sergey Mikhaylovich Brin



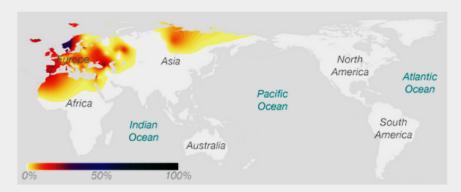


HOME MY RESULTS FAMILY & FRIENDS RESEARCH & COMMUNITY

MATERNAL LINE: H1

Overview History Haplogroup Tree Community

Locations of haplogroup H1 before the widespread migrations of the past few hundred years.



Haplogroup H1 is widespread in Europe, especially the western part of the continent. It originated about 13,000 years ago, not long after the Ice Age ended.

Maternal haplogroups are families of mitochondrial DNA types that all trace back to a single mutation at a specific place and time. By looking at the geographic distribution of mtDNA types, we learn how our ancient female ancestors migrated throughout the world.

Haplogroup: H1, a subgroup of H

Age: 13,000 years

Region: Europe, Near East, Central Asia,

Northwestern Africa

**Example Populations:** Spanish, Berbers, Lebanese **Highlight:** H1 appears to have been common in Doggerland, an ancient land now flooded by the

North Sea.

#### PATERNAL LINE: 11\*

Overview

History

Haplogroup Tree

Community

### I1\* is a subgroup of I1

Locations of haplogroup I1 before the widespread migrations of the past few hundred years.



Haplogroup I1 can be found at levels of 10% and higher in many parts of Europe, due to its expansion with men who migrated northward after the end of the Ice Age about 12,000 years ago. It reaches its highest levels in Denmark and the southern parts of Sweden and Norway.

Paternal haplogroups are families of Y chromosomes that all trace back to a single mutation at a specific place and time. By looking at the geographic distribution of these related lineages, we learn how our ancient male ancestors migrated throughout the world.

Haplogroup: I1, a subgroup of I

Age: 28,000 years

Region: Northern Europe

Example Populations: Finns, Norwegians, Swedes

Highlight: Haplogroup I1 reaches highest

frequencies in Scandinavia.

Haplogroups of You and Your Connections

11\*

Roman Hobza

Haplogroups of Example Profiles

These reports provide information about your possible risk for developing certain health conditions based on genetics. Environmental and lifestyle factors also often play a large role in your risk for developing these conditions.

#### Elevated Risk 🕜

NAME	CONFIDENCE	YOUR RISK	AVG. RISK	COMPARED TO AVERAGE
Venous Thromboembolism	***	41.8%	12.3%	3.39x
Gout	***	35.7%	22.8%	1.57x =
Melanoma	***	4.0%	2.9%	1.38x :
Restless Legs Syndrome	***	2.5%	2.0%	1.25x :
Exfoliation Glaucoma	***	2.2%	0.7%	2.90x <b>:</b>
Esophageal Squamous Cell Carcinoma (ESCC)	***	0.43%	0.36%	1.21x
Stomach Cancer (Gastric Cardia Adenocarcinoma)	***	0.28%	0.23%	1.22x ¦
Primary Biliary Cirrhosis	***	0.11%	0.08%	1.43x
Scleroderma (Limited Cutaneous Type)	***	0.08%	0.07%	1.24x

Show information for	Roman Hobza ▼	assuming	European ▼	ethnicity and an age range of	0-79	•



#### Roman Hobza 41.8 out of 100

men of European ethnicity who share Roman Hobza's genotype will develop Venous Thromboembolism between the ages of 0 and 79.



#### Average 12.3 out of 100

men of European ethnicity will develop Venous Thromboembolism between the ages of 0 and 79.

#### What does the Odds Calculator show me?

Use the ethnicity and age range selectors above to see the estimated incidence of Venous Thromboembolism due to genetics for men with Roman Hobza's genotype. The 23andMe Odds Calculator assumes that a person is free of the condition at the lower age in the range. You can use the name selector above to see the estimated incidence of Venous Thromboembolism for the genotypes of other people in your account.

The 23andMe Odds Calculator only takes into account effects of markers with known associations that are also on our genotyping chip. Keep in mind that aside from genetics, environment and lifestyle may also contribute to one's risk for Venous Thromboembolism.



#### **Understanding Your Results**

The heritability of venous thromboembolism is estimated to be 55%. This means that genetics (including unknown factors and known ones such as the SNPs we describe here) and environment play nearly equal roles in this condition. There are a number of environmental factors of various strengths that contribute to venous thromboembolism. Strong risk factors include hip or leg fractures, hip or knee replacement, major surgery or trauma, and spinal cord injury or surgery. Moderate risk factors include arthroscopic knee surgery, having central venous lines, congestive heart or respiratory failure, hormone replacement or oral contraceptive use, cancer, pregnancy, paralytic stroke, previous venous thromboembolism, and thrombophilia. Weak risk factors include bed rest for more than three days, immobility due to sitting (such as a long car or plane trip), specific types of chemotherapy, increasing age, laparoscopic surgery, obesity, and varicose veins. (sources)



#### What You Can Do

Assuming the ethnicity setting above is correct, your test results indicate you are at increased risk for venous thromboembolism based on genetics. Note that family history and non-genetic factors can also influence your risk for venous thromboembolism. Below are some steps you can take to reduce your risk.

#### Gene or region: F5 SNP: rs6025

	SNP used	Genotype	Adjusted Odds Ratio*		
Roman Hobza	rs6025	CT	European: 4.69		
* Odds ratios are reported for all available ethnicities.					

Factor V is the last clotting factor in the pathway before the activation step that turns prothrombin into thrombin. Clotting is usually kept from spiraling out of control by a feedback loop, similar to the way a thermostat operates. Once enough thrombin has been activated, it binds to a protein called "protein C." Protein C then inactivates factor V, thus cutting off activation of prothrombin into thrombin.

The SNP in the F5 gene causes a change in the protein sequence of factor V that prevents protein C from inactivating it. Since this version of factor V can still participate in the activation of thrombin, a situation results in which thrombin can be turned on but cannot be turned off. Once the clotting cascade is set off (whether appropriately or not), the riskier version of the SNP makes it more difficult to shut it off.

The riskiness of the T version of this SNP is further increased for women who also take hormonal birth control.

(The riskier version of this gene is also sometimes called Factor V Leiden, after the city in the Netherlands where this SNP and its effects on factor V's role in clotting were first discovered.)

The studies whose data we report as applicable to those of "European" ancestry confirmed the association between this SNP and VTE in samples from the Netherlands, Sweden, the United Kingdom, Brazil, Italy, and France.

African and Asian populations appear to have only one version of the SNP, meaning that association studies are very difficult to perform.

#### Citations

Rosendaal et al. (1995). "High risk of thrombosis in patients homozygous for factor V Leiden (activated protein C resistance)." Blood 85(6):1504-8.

Smith et al. (2007). "Association of genetic variations with nonfatal venous thrombosis in postmenopausal women." JAMA 297(5):489-98.

Emmerich et al. (2001). "Combined effect of factor V Leiden and prothrombin 20210A on the risk of venous thromboembolism--pooled analysis of 8 case-control studies including 2310 cases and 3204 controls. Study Group for Pooled-Analysis in Venous Thromboembolism." Thromb Haemost 86(3):809-16.

Bertina et al. (1994). "Mutation in blood coagulation factor V associated with resistance to activated protein C." Nature 369(6475):64-7.

Lane et al. (2000). "Role of hemostatic gene polymorphisms in venous and arterial thrombotic disease." Blood 95(5):1517-32.

Gene or region: F2 SNP: i3002432

### Decreased Risk @

NAME	CONFIDENCE	YOUR RISK	AVG. RISK	COMPARED TO AVERAGE
Type 2 Diabetes	***	17.7%	25.7%	0.69x <b>=</b>
Alzheimer's Disease	***	4.3%	7.2%	0.60x <b>1</b>
Rheumatoid Arthritis	***	1.6%	2.4%	0.68x :
Parkinson's Disease	***	1.2%	1.6%	0.73x :
Age-related Macular Degeneration	***	0.92%	6.55%	0.14x <b>L</b>
Crohn's Disease	***	0.31%	0.53%	0.58x ¦
Multiple Sclerosis	***	0.24%	0.34%	0.69x ¦
Type 1 Diabetes	***	0.12%	1.02%	0.12x :
Celiac Disease	***	0.05%	0.12%	0.44x ¦

BRCA Cancer Mutations (Selected)	***	Variant Absent
Beta Thalassemia	***	Variant Absent
Bloom's Syndrome	***	Variant Absent
Canavan Disease	***	Variant Absent
Congenital Disorder of Glycosylation Type 1a (PMM2-CDG)	***	Variant Absent
Connexin 26-Related Sensorineural Hearing Loss	***	Variant Absent
Cystic Fibrosis	***	Variant Absent
D-Bifunctional Protein Deficiency	***	Variant Absent
DPD Deficiency	***	Variant Absent
Dihydrolipoamide Dehydrogenase Deficiency	***	Variant Absent
Factor XI Deficiency	***	Variant Absent
Familial Dysautonomia	***	Variant Absent
Familial Hypercholesterolemia Type B	***	Variant Absent
Familial Hyperinsulinism (ABCC8-related)	***	Variant Absent
Familial Mediterranean Fever	***	Variant Absent
Fanconi Anemia (FANCC-related)	***	Variant Absent
G6PD Deficiency	***	Variant Absent

Reading Ability	***	Typical Nonword Reading Score
Response to Diet	***	See Report
Response to Exercise	***	See Repor
Sex Hormone Regulation	***	See Repor
Sweet Taste Preference 🔆	***	See Repor
Tooth Development	***	See Repor
Tuberculosis Susceptibility	***	See Repor
Breast Morphology 🍳 🔆	***	Not Applicable
Menarche 9	***	Not Applicable
Menopause 🍳	***	Not Applicable
Eating Behavior	**	Greater tendency to overea
HIV Progression	**	See Repor
Hair Thickness	**	Typical, if European or African
Longevity	**	See Repor
Measures of Intelligence	**	Lower Non-Verbal IC
Memory	**	Typical Episodic Memor
Odor Detection	**	Typical Sensitivity to Sweaty Odo
Pain Sensitivity	**	Increase
Avoidance of Errors	*	See Repor

# Genome Sequencer 20 System 454 pyrosequencing (2005)

http://www.454.com





### **DNA** library preparation

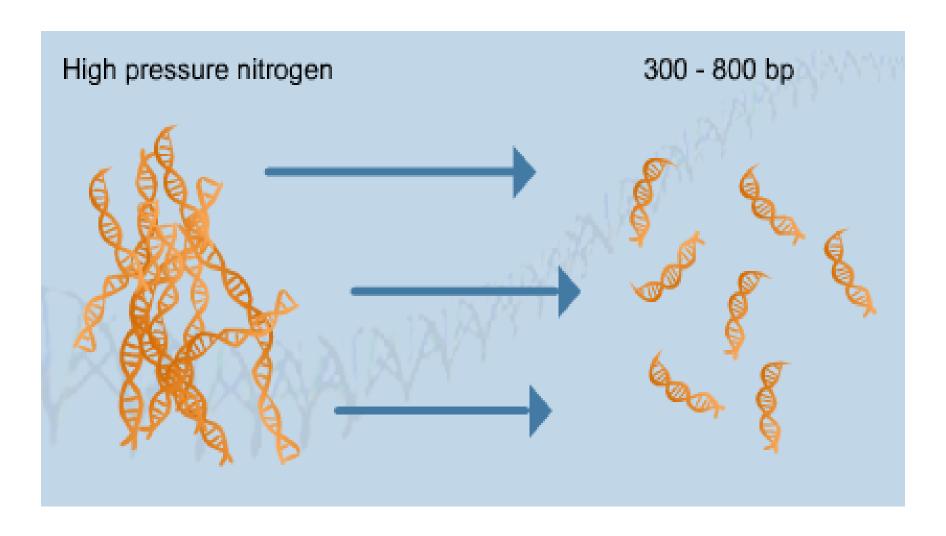
One sample preparation per genome

No Cloning

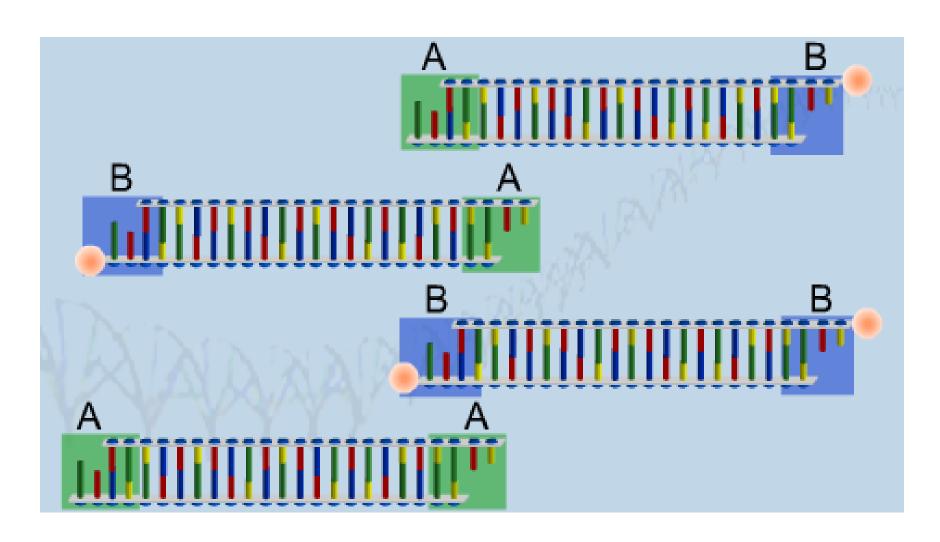
No Colony Picking



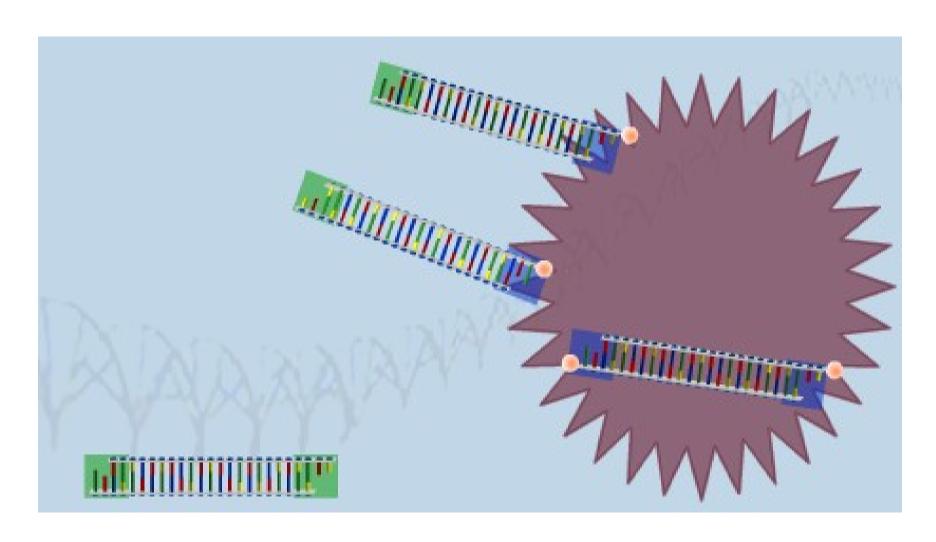
# Fragmentace DNA



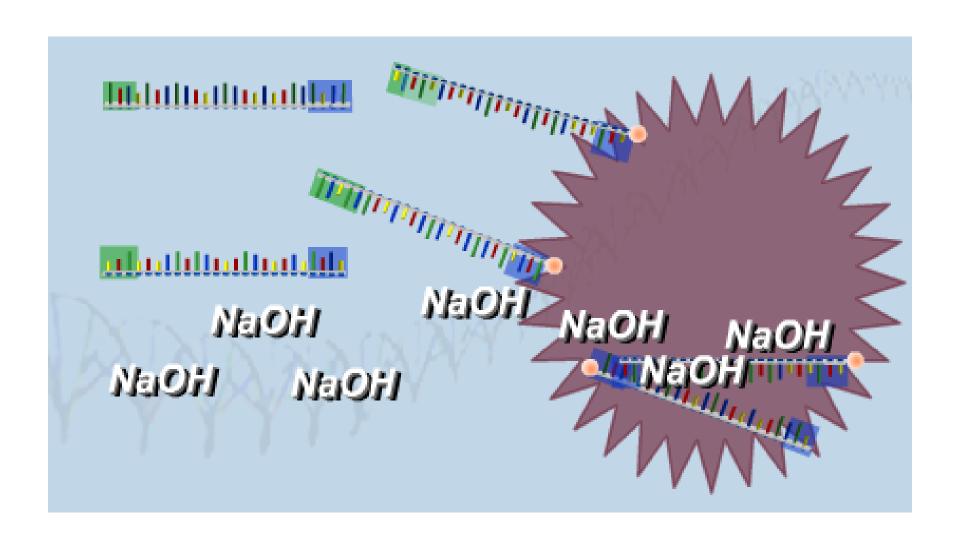
# Ligace adaptoru

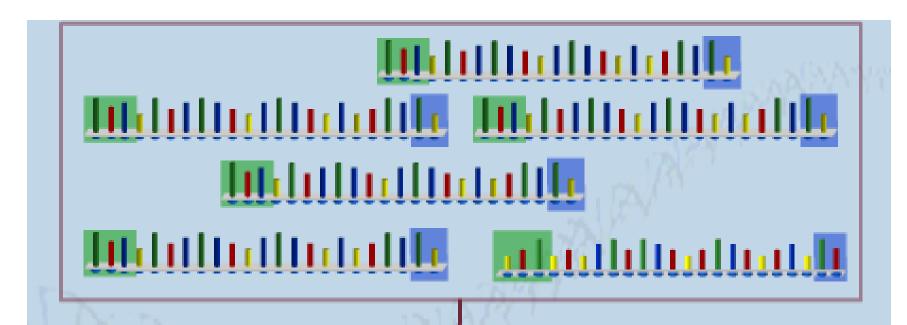


# Vychytání DNA molekul



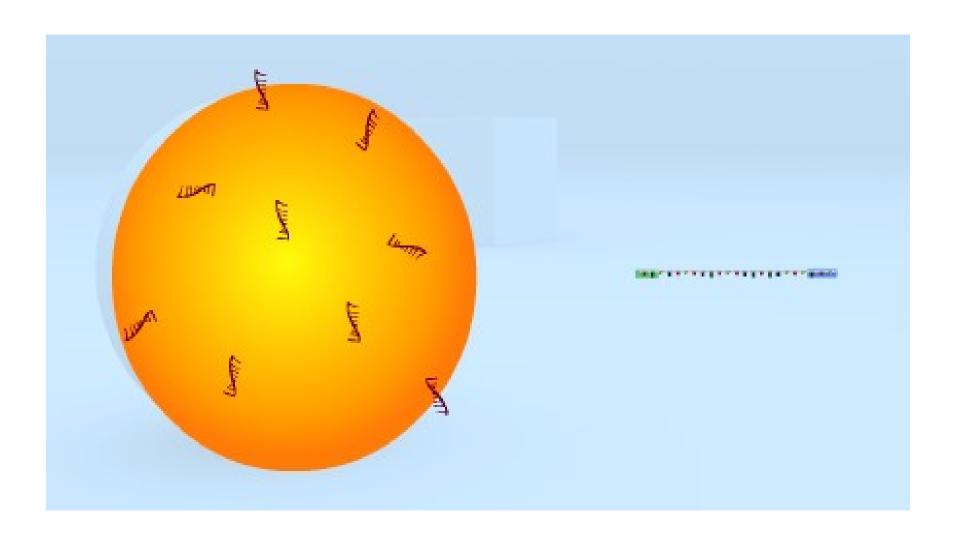
## denaturace



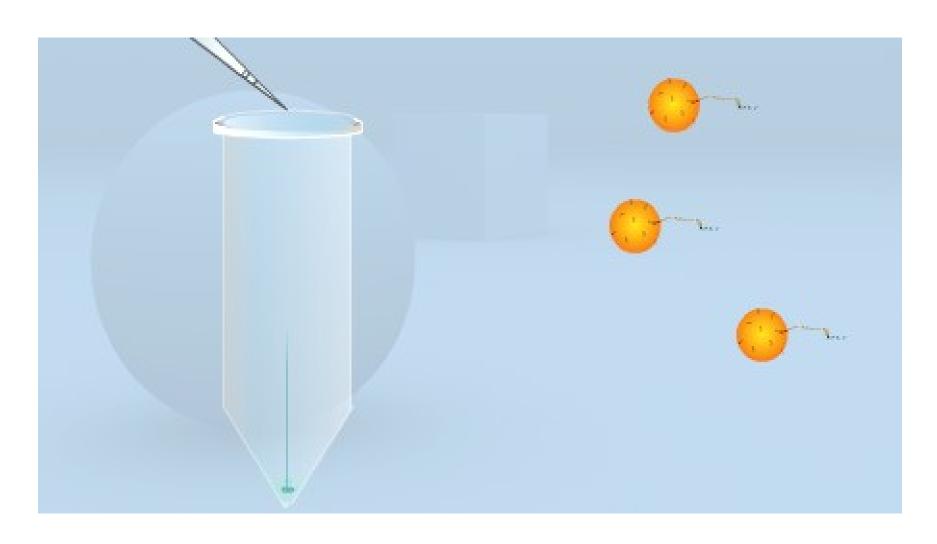


Single-stranded template DNA sstDNA

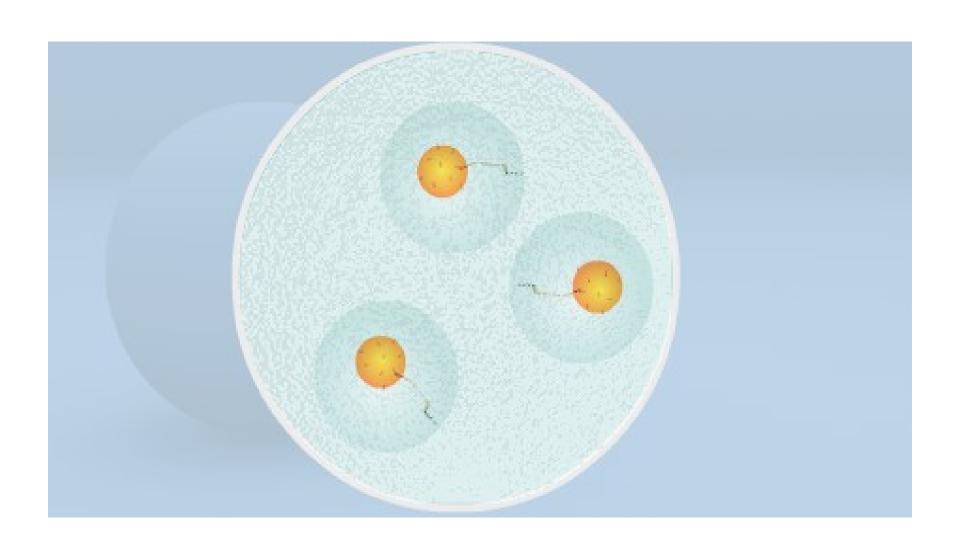
# emPCR



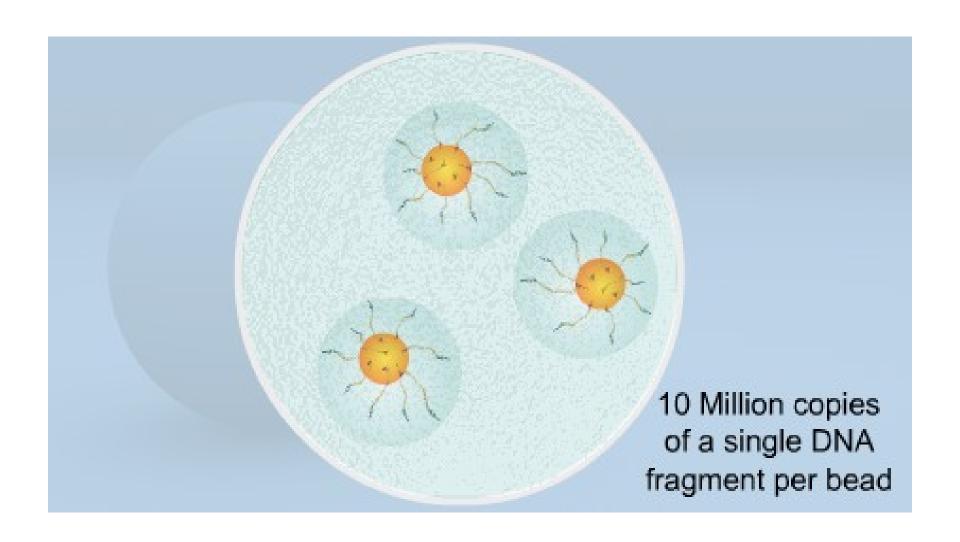
## Vznik emulze (olej)



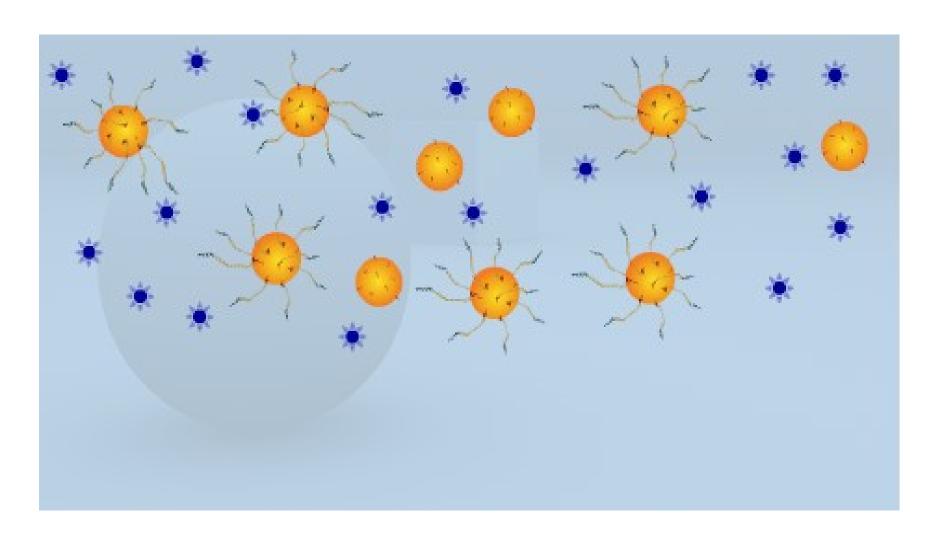
## emPCR



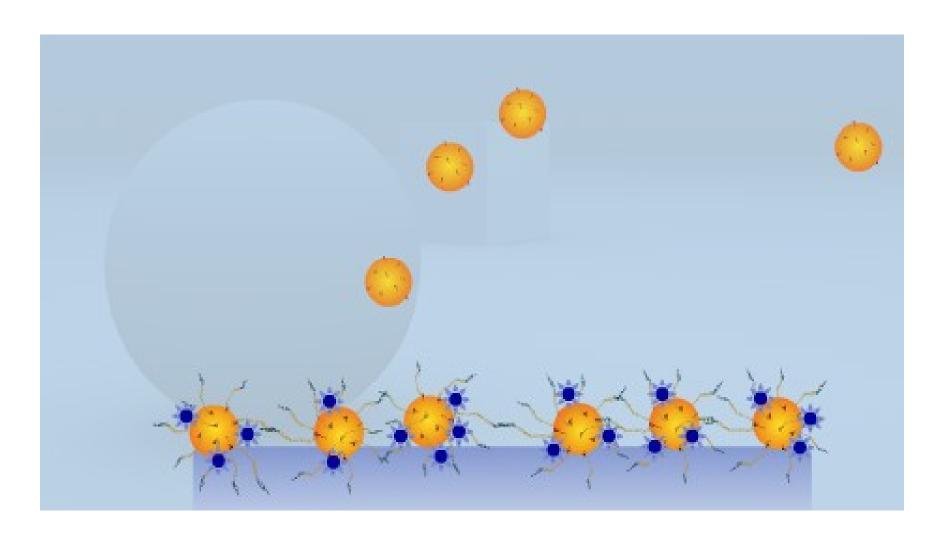
### emPCR



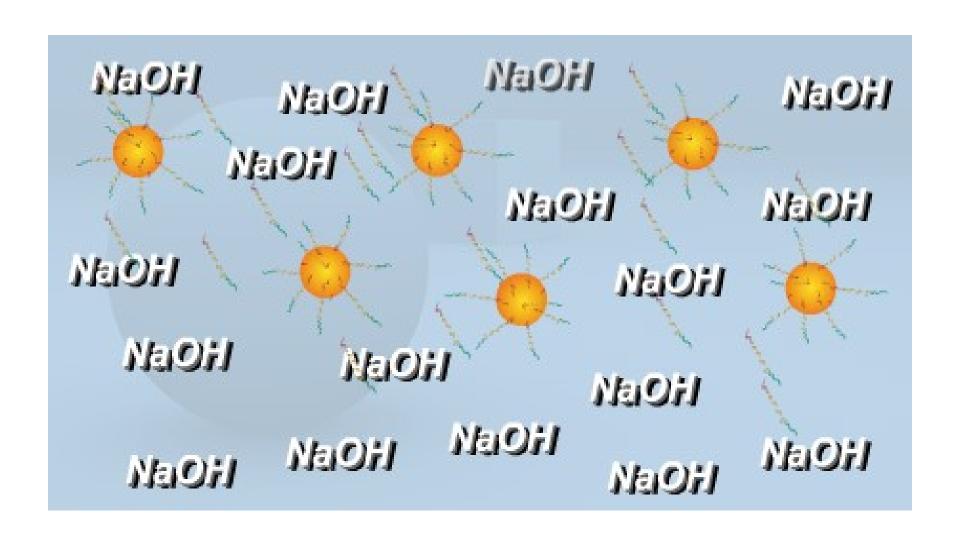
# Vychytání kuliček



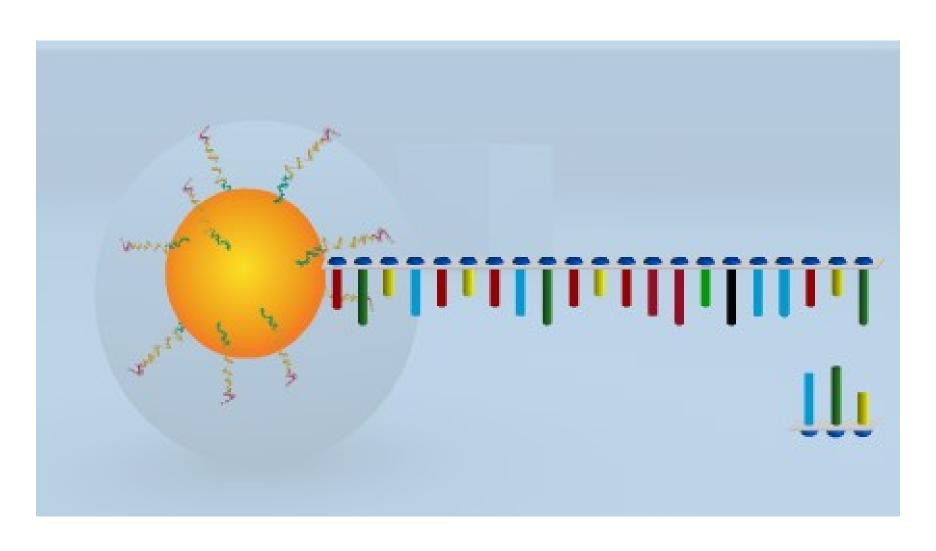
# Vychytání kuliček



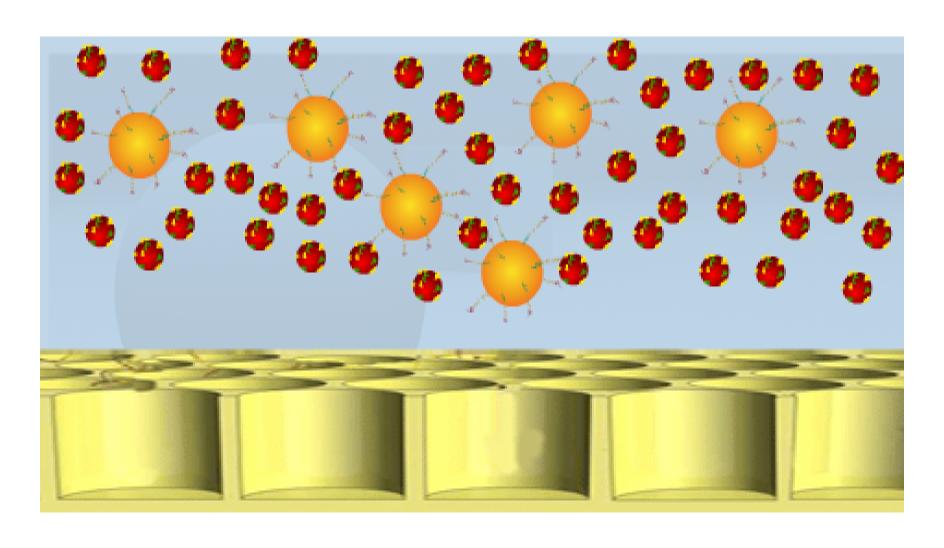
### denaturace



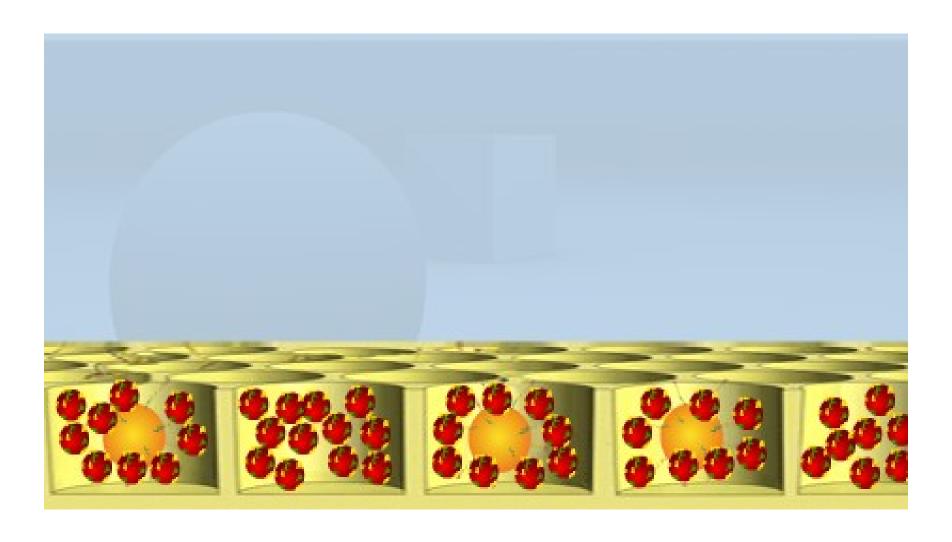
## Sekvenační primer



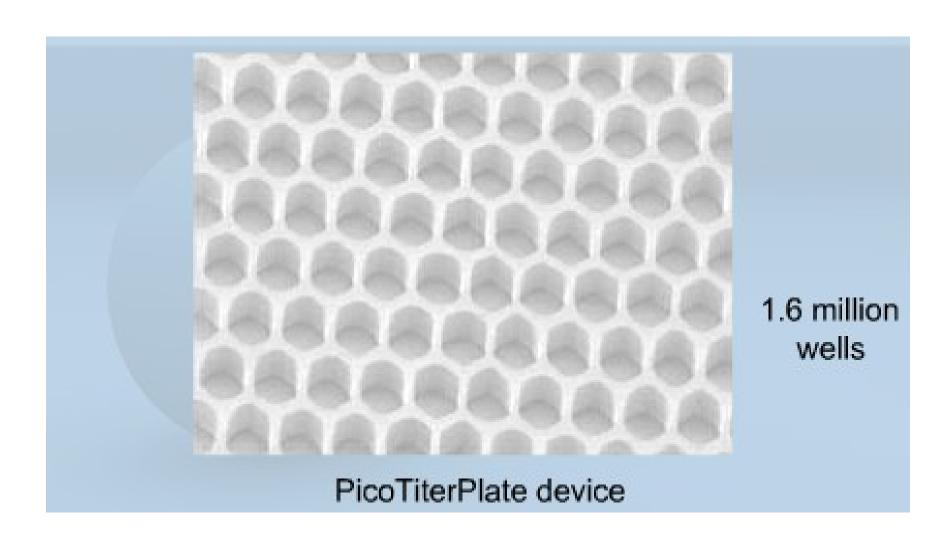
## Disperze na sklíčko



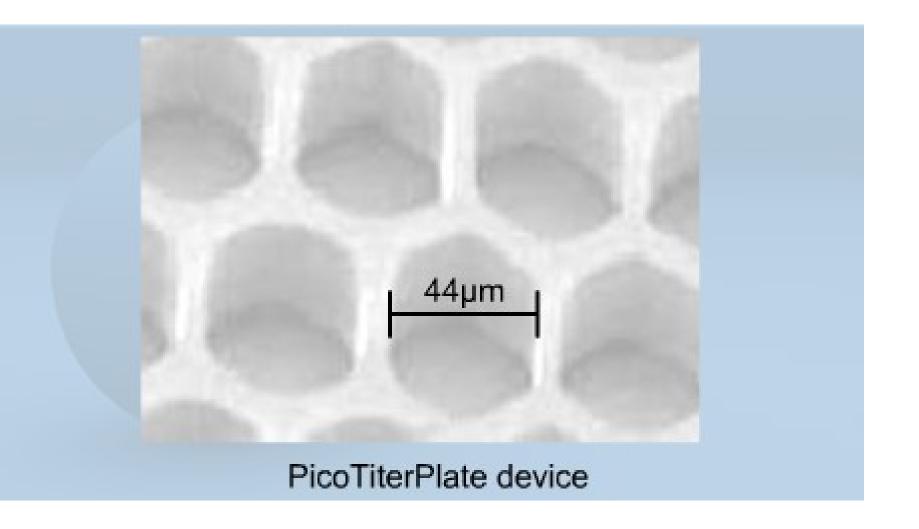
## Disperze na sklíčko

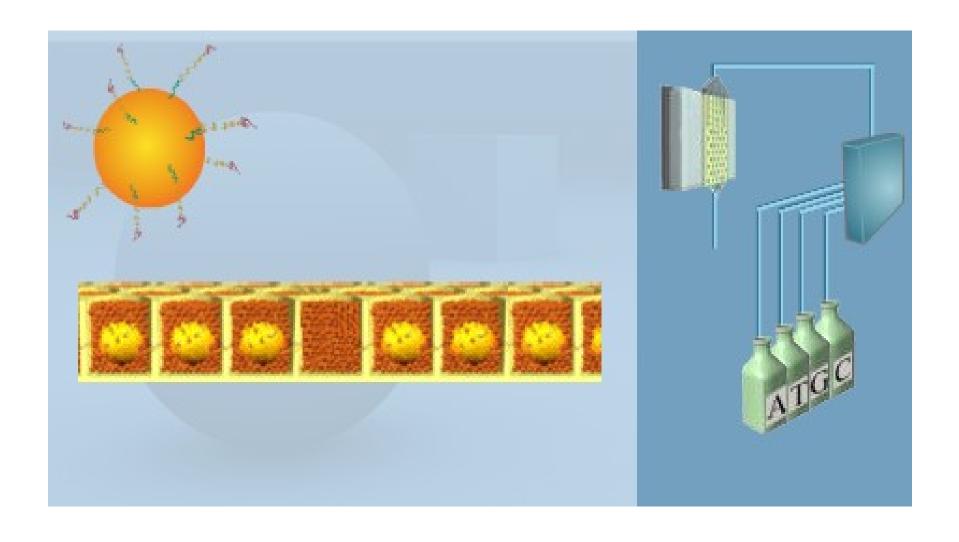


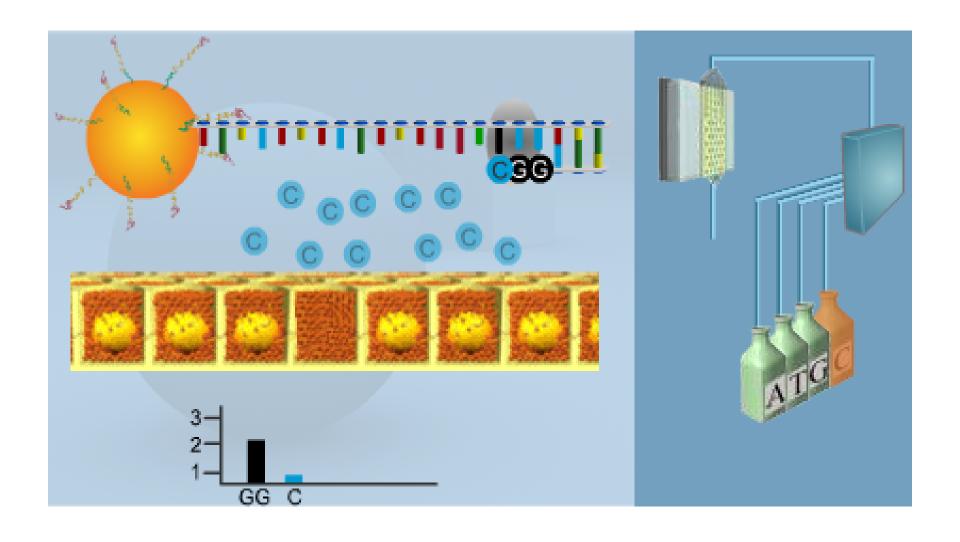
## Parametry mikroreaktorů

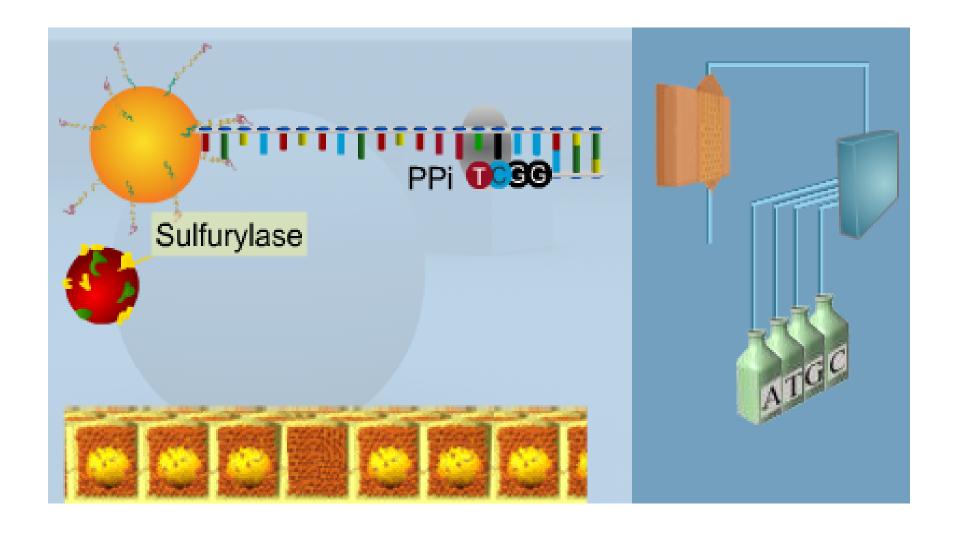


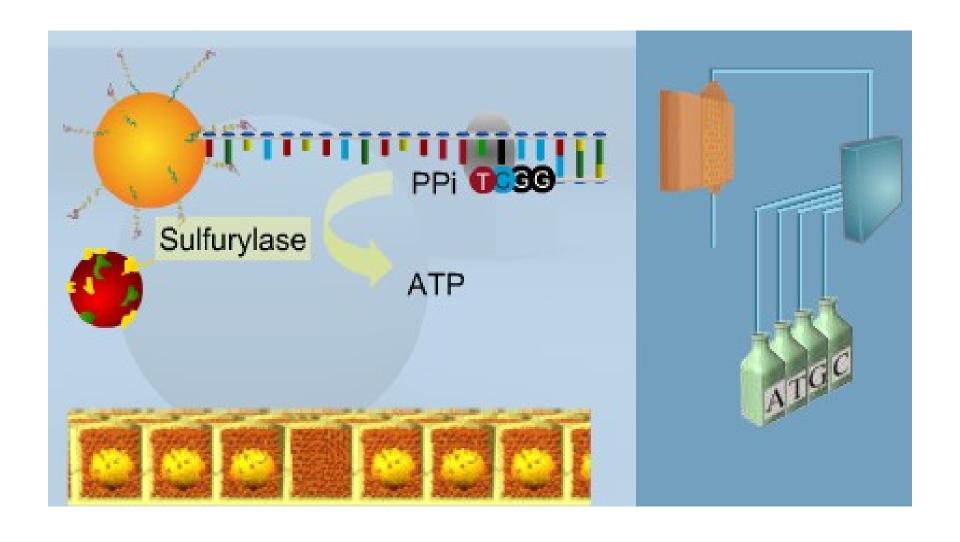
## Parametry mikroreaktorů

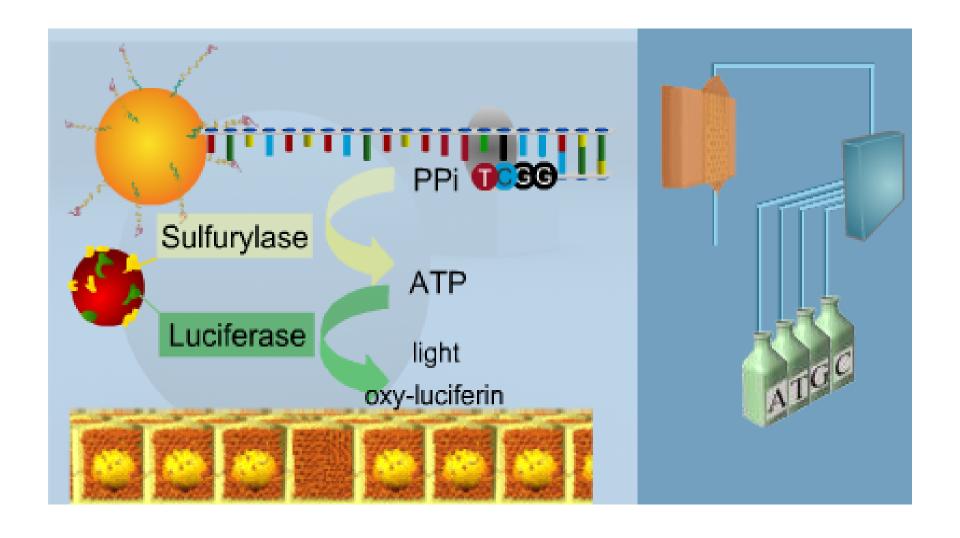


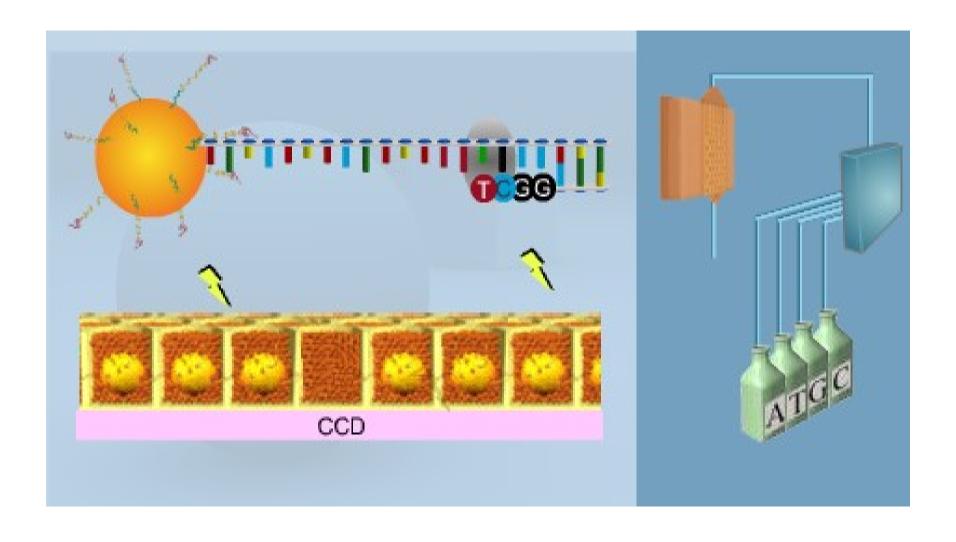


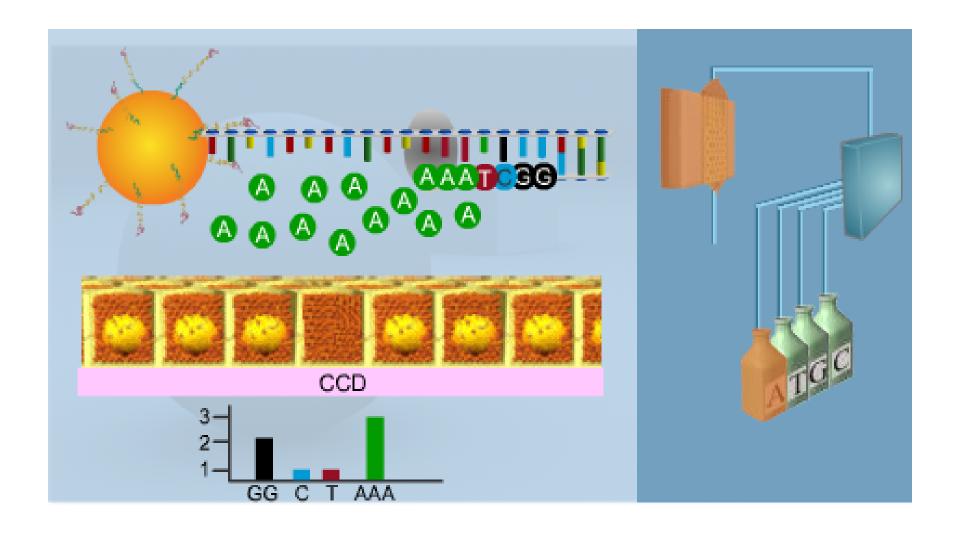


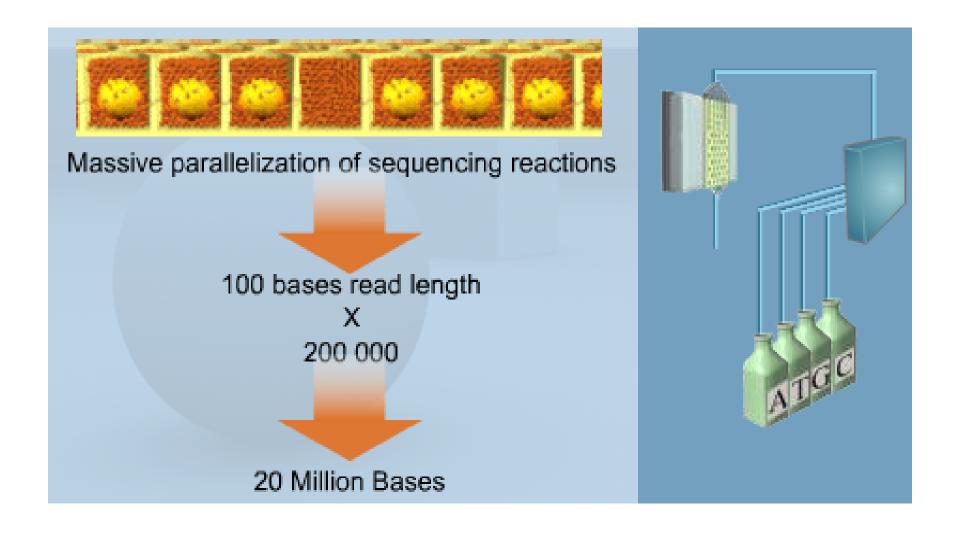






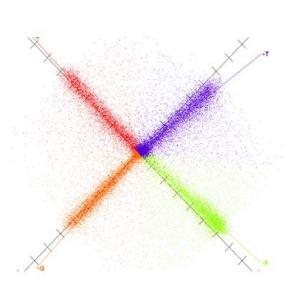






SOLID (Sequencing by Oligonucleotide Ligation and Detection) 2-base encoding sequencing (2007)









SOLiD™ System Sequencing by Oligonucleotide Ligation and Detection

### Properties of the Probes

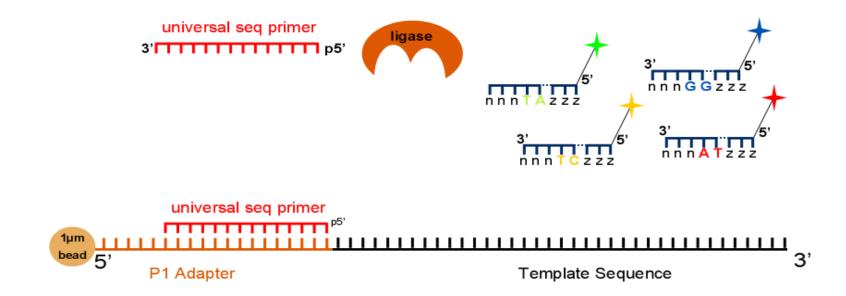
Spatial separation among dye, ligation & cleavage sites

Cleavage site, Fluorescent dye 3' Ligation site

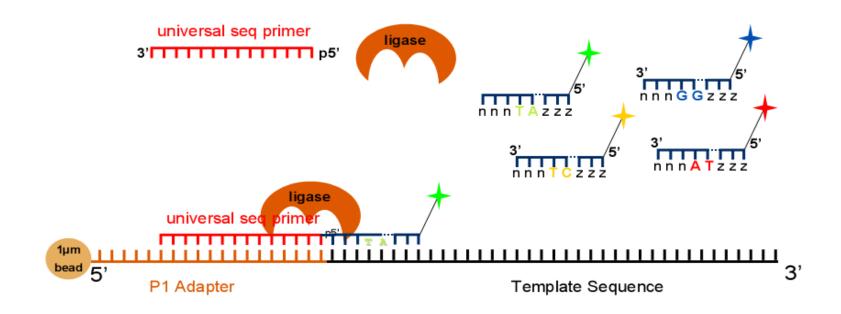
1,024 Octamer Probes (4<sup>5</sup>)

4 Dyes, 4 dinucleotides, 256 probes per dye N= degenerate bases Z= Universal bases

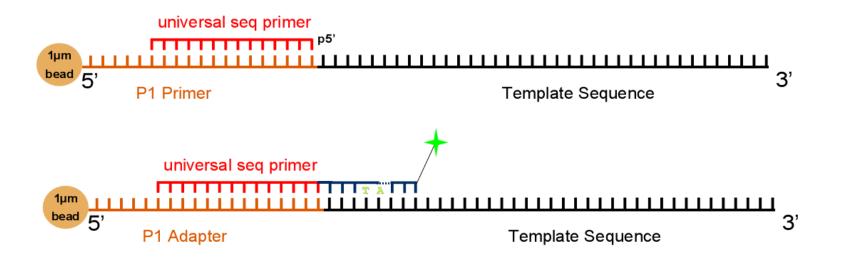
### SOLiD Chemistry System 4-color ligation Ligation reaction



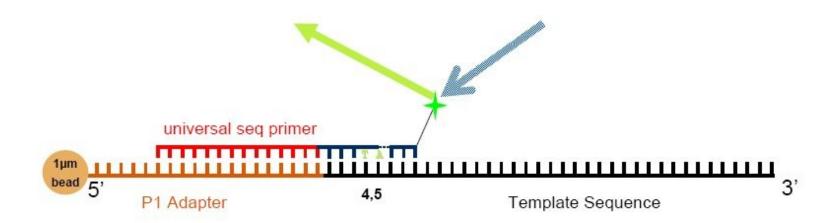
# **SOLiD Chemistry System 4-color ligation Ligation reaction**



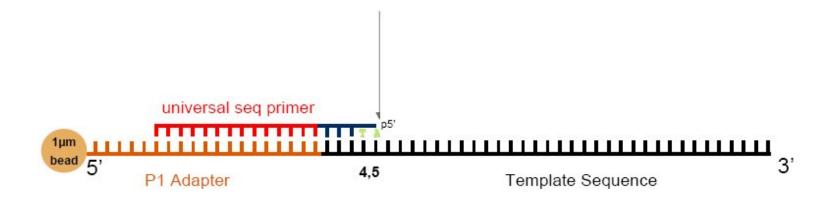
### SOLiD Chemistry System 4-color ligation De-Phosphorylation



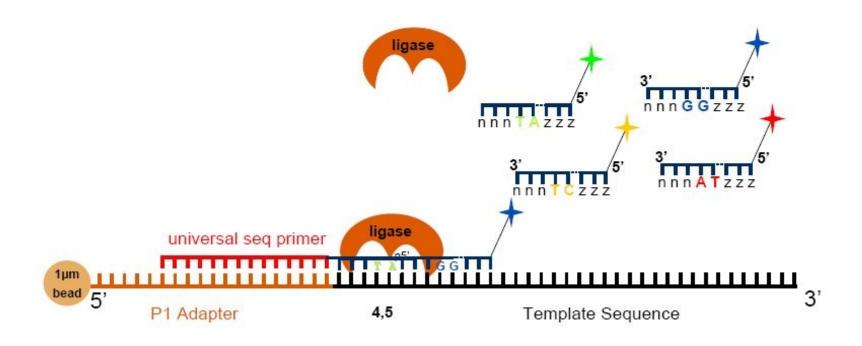
### SOLiD Chemistry System 4-color ligation Visualization



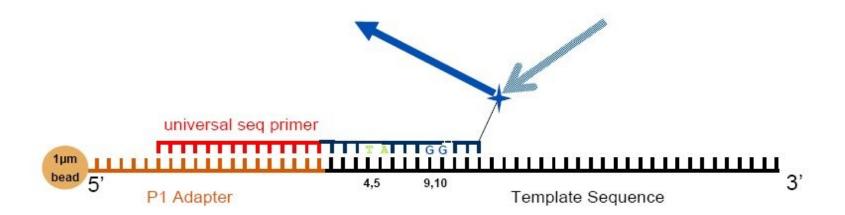
# **SOLiD Chemistry System 4-color ligation Cleavage**



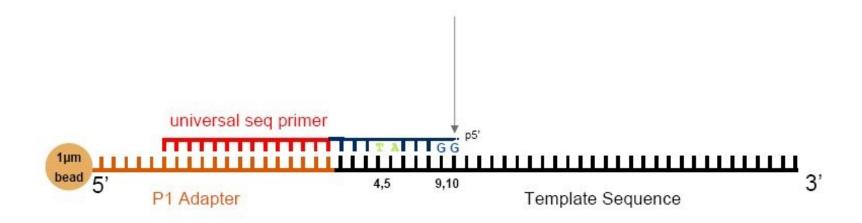
# SOLiD Chemistry System 4-color ligation Ligation (2<sup>nd</sup> cycle)



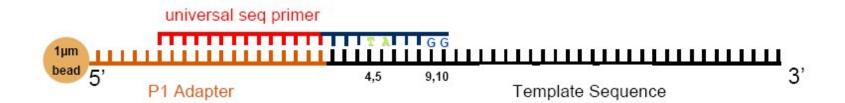
# SOLiD Chemistry System 4-color ligation Visualization (2<sup>nd</sup> cycle)



# **SOLiD Chemistry System 4-color ligation** Cleavage (2<sup>nd</sup> cycle)



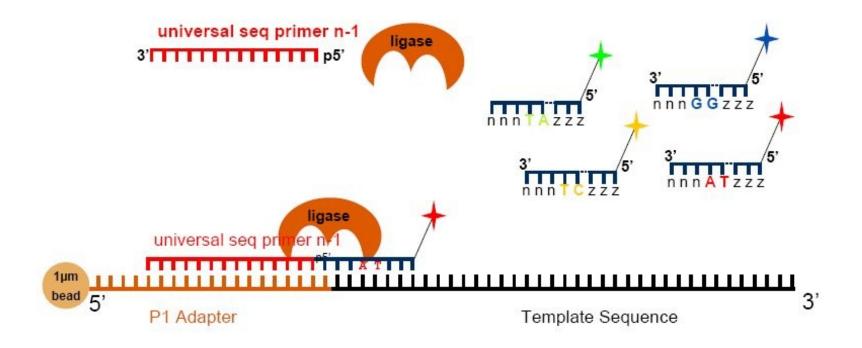
# SOLiD Chemistry System 4-color ligation interrogates every 5<sup>th</sup> base



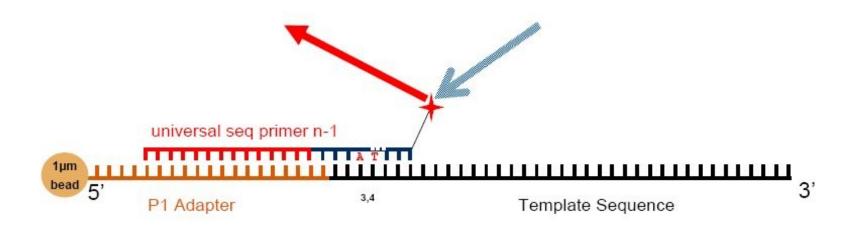
### **SOLiD Chemistry System 4-color ligation** Reset



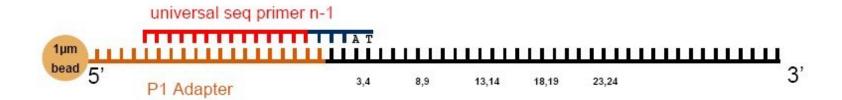
# SOLiD Chemistry System 4-color ligation (1st cycle after reset)



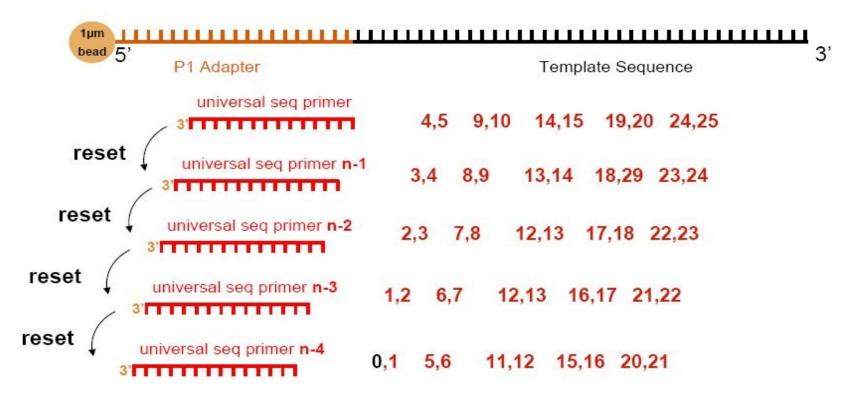
# SOLiD Chemistry System 4-color ligation (1st cycle after reset)



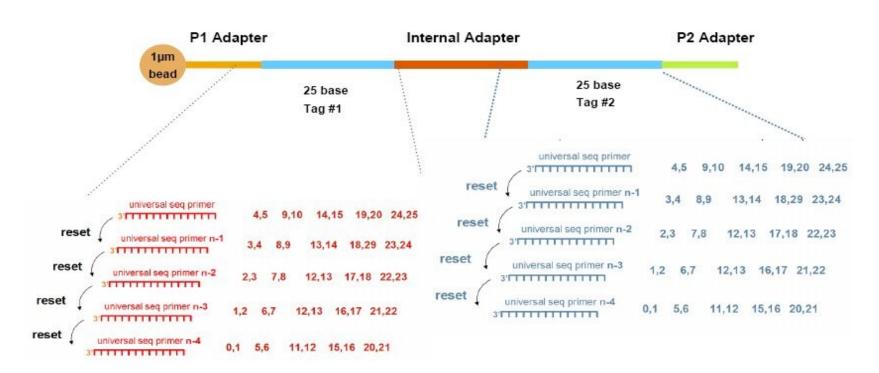
# SOLiD Chemistry System 4-color ligation (2<sup>nd</sup> Round)

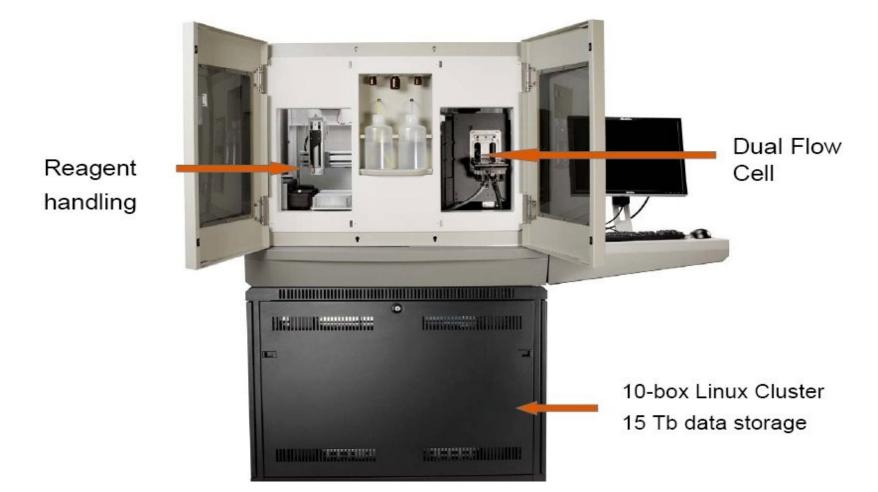


### Sequential rounds of sequencing Multiple cycles per round



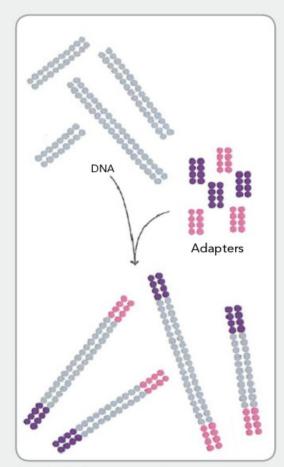
### Paired End two sequences generated Sequential rounds of sequencing Multiple cycles per round





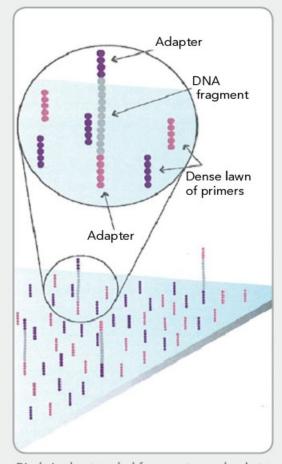
# **Solexa** (2007)

#### 1. PREPARE GENOMIC DNA SAMPLE



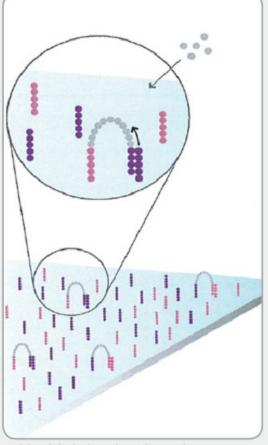
Randomly fragment genomic DNA and ligate adapters to both ends of the fragments.

#### 2. ATTACH DNA TO SURFACE



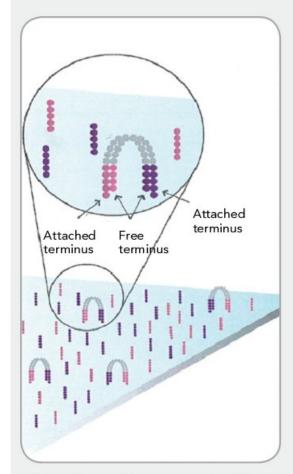
Bind single-stranded fragments randomly to the inside surface of the flow cell channels.

#### 3. BRIDGE AMPLIFICATION



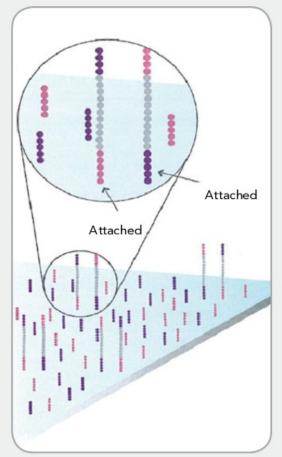
Add unlabeled nucleotides and enzyme to initiate solid-phase bridge amplification.

## 4. FRAGMENTS BECOME DOUBLE STRANDED



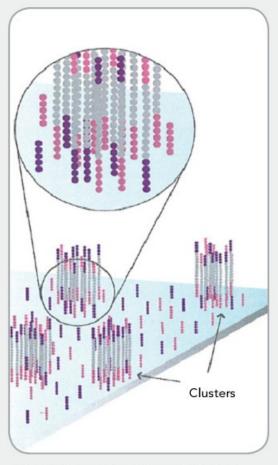
The enzyme incorporates nucleotides to build double-stranded bridges on the solid-phase substrate.

## 5. DENATURE THE DOUBLE-STRANDED MOLECULES



Denaturation leaves single-stranded templates anchored to the substrate.

## 6. COMPLETE AMPLIFICATION



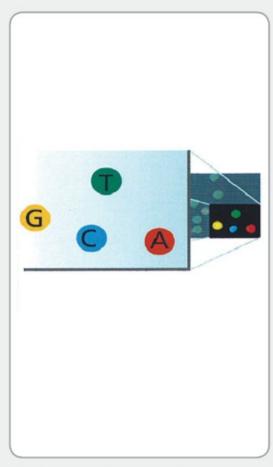
Several million dense clusters of doublestranded DNA are generated in each channel of the flow cell.

## 7. DETERMINE FIRST BASE

# Laser

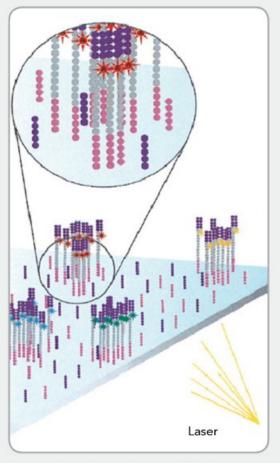
First chemistry cycle: to initiate the first sequencing cycle, add all four labeled reversible terminators, primers and DNA polymerase enzyme to the flow cell.

#### 8. IMAGE FIRST BASE



After laser excitation, capture the image of emitted fluorescence from each cluster on the flow cell. Record the identity of the first base for each cluster.

#### 9. DETERMINE SECOND BASE



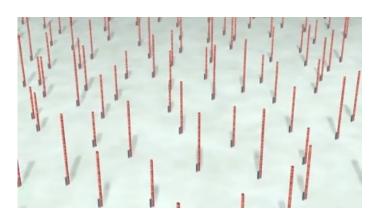
Second chemistry cycle: to initiate the next sequencing cycle, add all four labeled reversible terminators and enzyme to the flow cell.

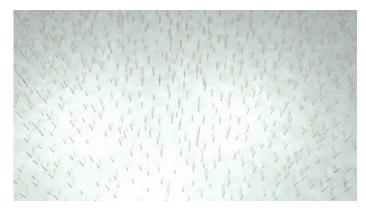
# **HELICOS (2008)**

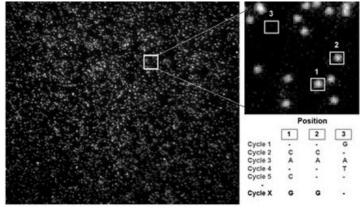




## True Single Molecule Sequencing (tSMS)



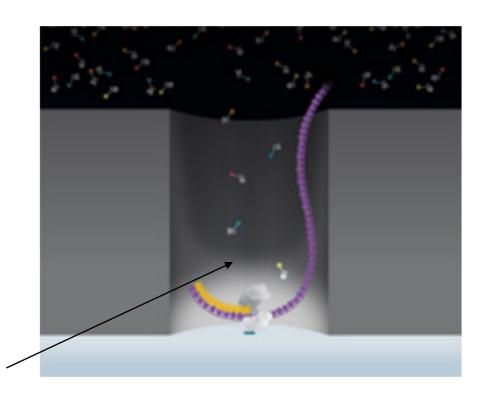








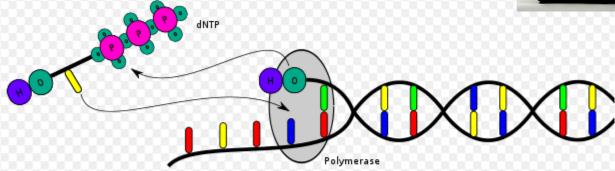
## **Pacific Biosciences**



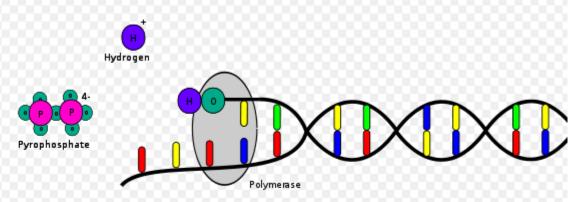
20 zeptolitrů

# **Ion Torrent**





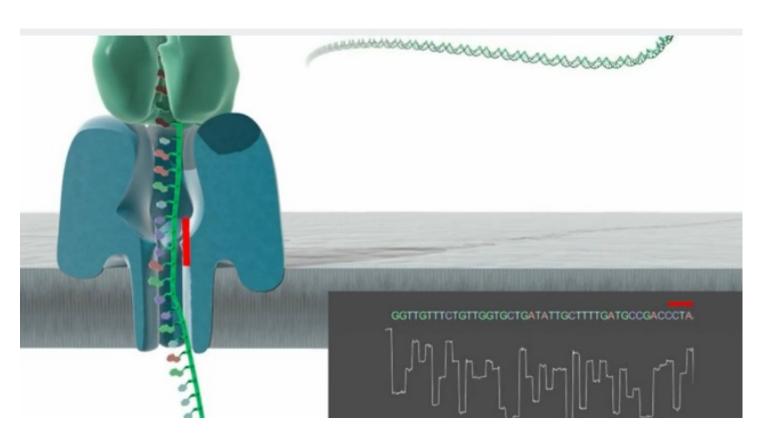
Polymerase integrates a nucleotide.



Hydrogen and pyrophosphate are released.

## **Oxford nanopore**





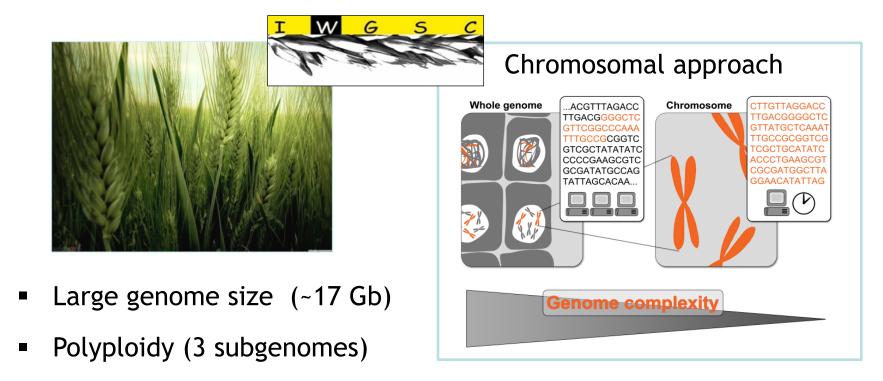
# Další technologie

- Mikroelektroforéza
- Sekvenování na bázi microarray

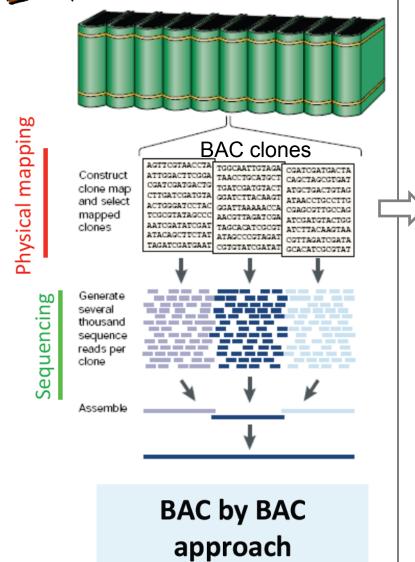
# CHALLENGES IN GENOME SEQUENCING

De novo genome assemblies using only short read data of NGS technologies are generally incomplete and highly fragmented due to

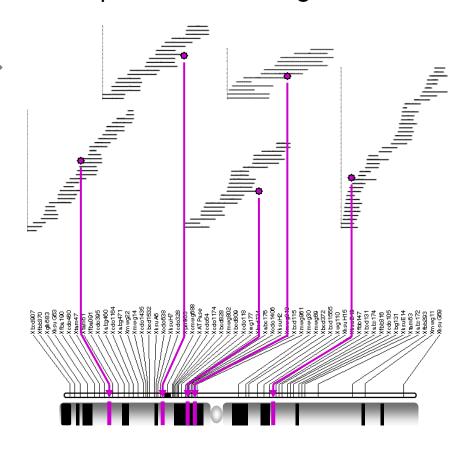
- Large duplications chromosomal approach, BAC-by-BAC sequencing
- High proportion of repetitive DNA challenge!



# **BAC-BY-BAC SEQUENCING**



- Physical map is composed of contigs of overlapping BAC clones
- BAC contigs are landed on the chromosome through markers comprised in the contigs



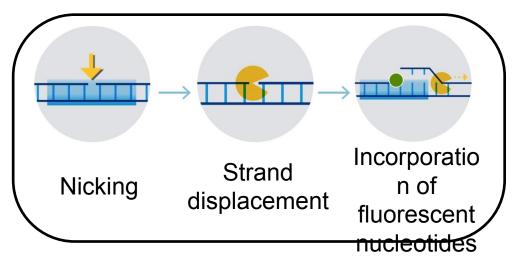
## SOLUTIONS FOR THE REPEATS

- Long mate-pair reads > 10 kb
- Long read technologies PacBio, Oxford Nanopore
- Optical mapping
  - Single-molecule mapping of genomic DNA hundreds of kilobases to several megabases in size
  - Creates sequence-motif maps, which provide long-range template for ordering genomic sequences
  - Visualisation of reality "Seeing is Believing"

## **OPTICAL MAPPING**

# Three enzymatic approaches

- restriction enzymes: sequence-specifically cleave DNA immobilized on a surface
- nicking enzymes:
   fluorescent labelling
   of the nicking site
   in solution (BioNano
   Genomics Irys)



methyltransferase enzymes:
 labelling with ultra-high density

## BIONANO GENOME MAPPING ON NANOCHANEL ARRAYS

1 Sequence-specific labeling

Nickase (Nt.BspQI)

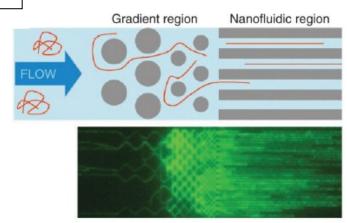
5'-ATGCGCTCTTCCATGAATGCGAGC-3'
3'-TACGCGAGAAGGTACTTACGCTCG-5'



5'-ATGC**GCTCTTC**CAUGAAUGCGAGC-3'

3'-TACGCGAGAAGGTACTTACGCTCG-5'

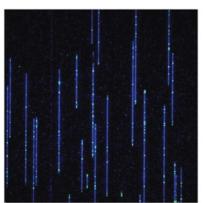
2 DNA linearization



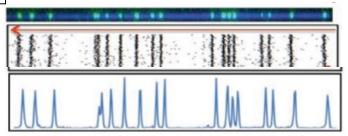
Lam et al., Nat. Biotechnol. 30(8) 2012

3 Fluorescence imaging





4 Map construction



5 Building consensus map

