

# CG920 Genomics

## Lesson 4

### Forward Genetics

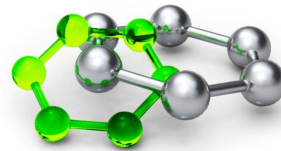
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# Outline

- Forward vs. Reverse Genetics
- Use of Libraries of Insertional Mutants in Forward Genetics
  - Searching in Libraries of Insertional Mutants According to:
    - anatomically or morphologically detectable phenotype
    - metabolic profile
    - expression of genes of interest
  - Identification of the Mutated Locus
    - plasmid rescue
    - iPCR
- Use of Libraries of Point Mutants in Forward Genetics
  - Positional Cloning

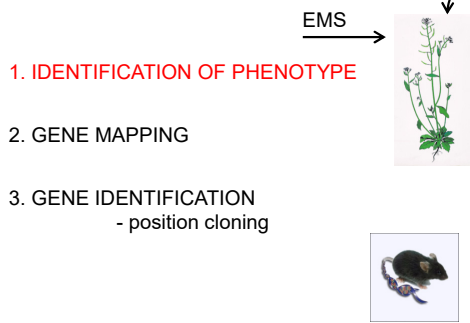
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- Forward vs. Reverse Genetics

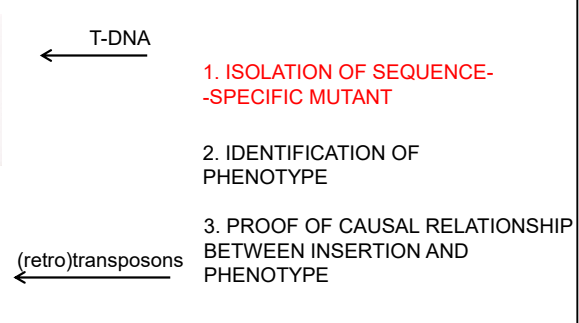
„Classical“ genetics *versus* „reverse genetics“  
approaches in functional genomics

**RANDOM MUTAGENESIS**

**„Classical genetics“ approach**



**„Reverse genetics“ approach**

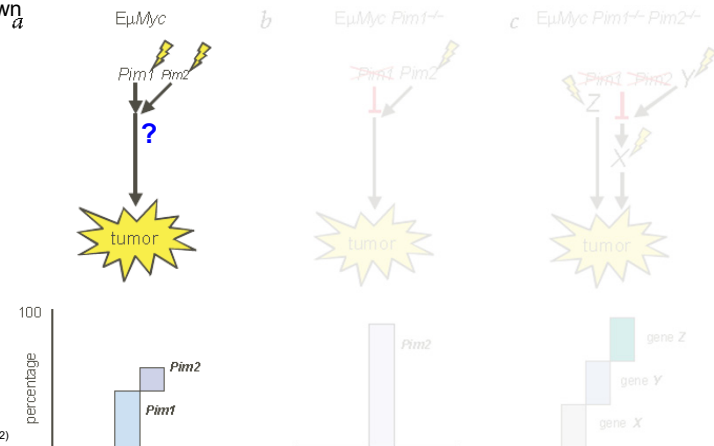


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- Forward vs. Reverse Genetics
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# Insertional mutagenesis in forward genetics approaches

- Use of insertional mutagenesis for study of carcinogenesis
  - Infection of EμMyc mice by MoMuLV retrovirus leads to lymphomas formation, which arose due to activation of Pim kinases (40 % activation of Pim1, 15 % activation of Pim2), molecular targets of these kinases were unknown

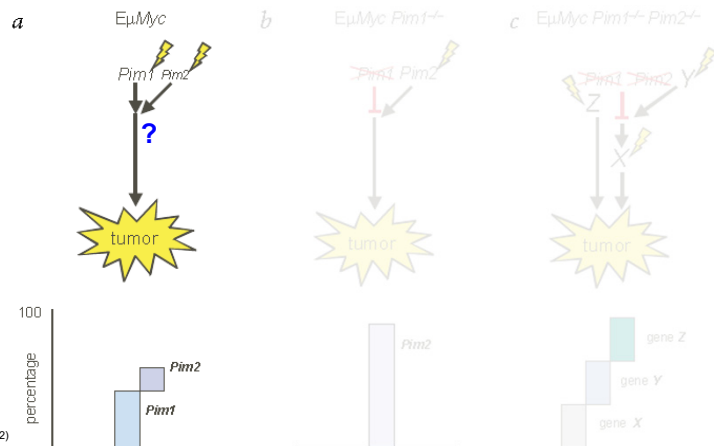


6

Mikkers et al., Nature Gen (2002)

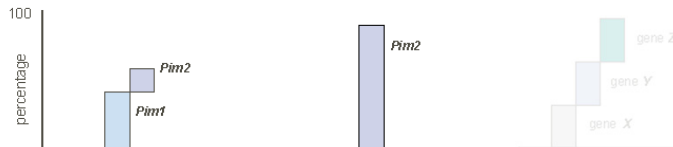
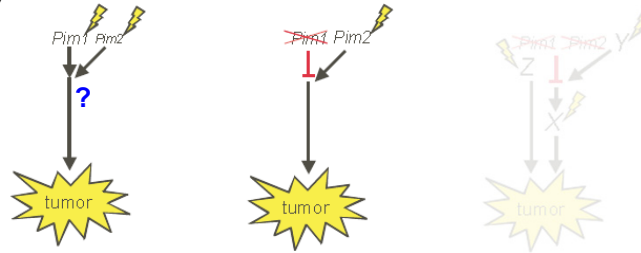
# Insertional mutagenesis in forward genetics approaches

- Use of insertional mutagenesis for study of carcinogenesis
  - Infection of EμMyc *pim1* mutants by MoMuLV retrovirus leads to lymphomas formation, which in 90 % contain insertion nearby (activation) *Pim2*



# Insertional mutagenesis in forward genetics approaches

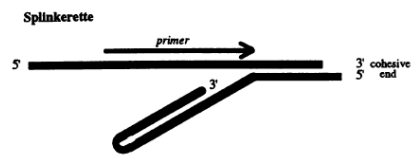
- Use of insertional mutagenesis for study of carcinogenesis
  - Infection of EμMyc double mutants *pim1*, *pim2* by MoMuLV retrovirus leads to lymphomas formation, which can be expected to activate either one of the signalling partner of Pim proteins (Y), one of the downstream proteins of Pim signalling pathway (X) or to activate some of the related pathways leading to lymphomagenesis (Z).





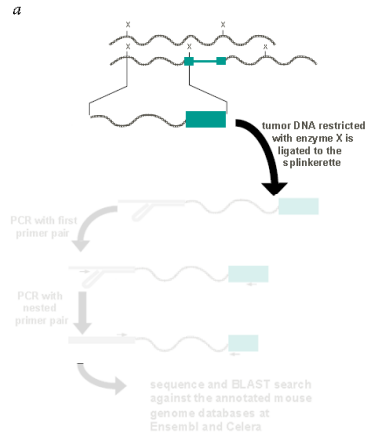
# Insertional mutagenesis in forward genetics approaches

- Isolation of genomic regions adjacent to the insertion site of the provirus
  - Cleavage of genomic DNA and ligation of special linkers, so-called *splinkerettes* (increasing the specificity of amplification)



Devon et al., Nucl Acid Res (1994)

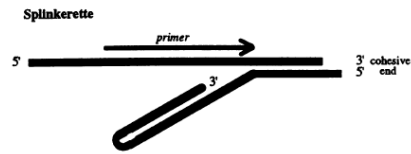
Mikkers et al., Nature Gen (2002)



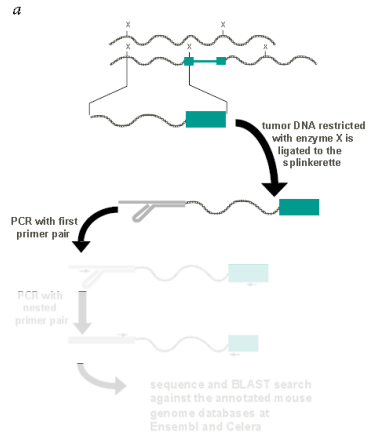
# Insertional mutagenesis in forward genetics approaches

- Isolation of genomic regions adjacent to the insertion site of the provirus

- First amplification using specific primers



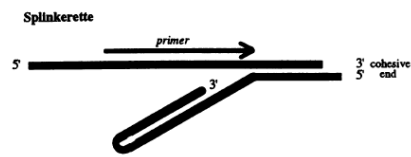
Devon et al., Nucl Acid Res (1994)



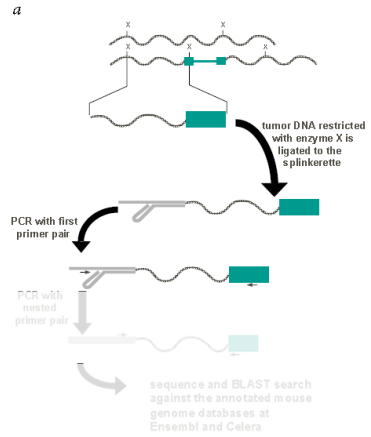
Mikkers et al., Nature Gen (2002)

# Insertional mutagenesis in forward genetics approaches

- Isolation of genomic regions adjacent to the insertion site of the provirus
  - Second amplification using **nested primers** (increasing the specificity)



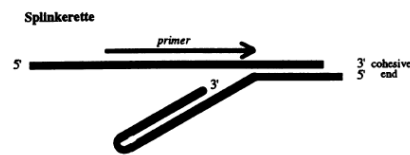
Devon et al., Nucl Acid Res (1994)



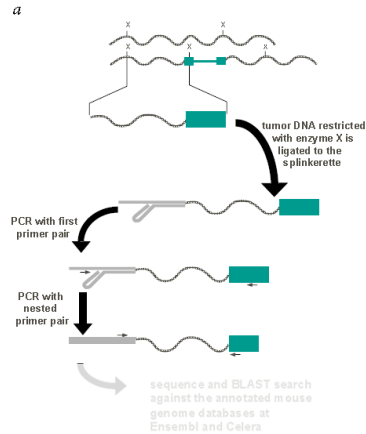
Mikkers et al., Nature Gen (2002)

# Insertional mutagenesis in forward genetics approaches

- Isolation of genomic regions adjacent to the insertion site of the provirus
  - Sequencing and localization of regions adjacent to provirus by searching in annotated databases of mouse genome



Devon et al., Nucl Acid Res (1994)



Mikkers et al., Nature Gen (2002)

In case of splinkerette, the primer is of the same sequence as the top strand and therefore it is unable to act as a primer until the complement of this strand has been synthesized (from the insert-specific primer at the right-hand side).

# Outline

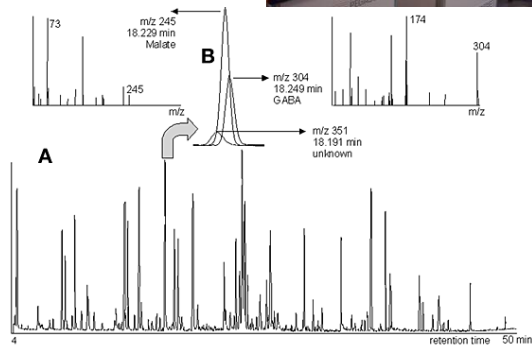
- Forward vs. Reverse Genetics
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  - Searching in Libraries of Insertional Mutants According to:
    - anatomically or morphologically detectable phenotype
    - metabolic profile

# Metabolic profiling

- Metabolic profiling of plants
  - Automated analysis of metabolites (up to 25.000) by GC-MS techniques in libraries of T-DNA mutants

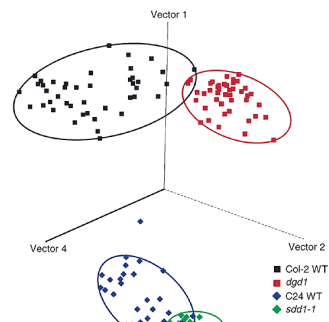


Retention Time (min)	m/z	Abundance	Identification
1.23	73	100	
1.45	245	50	
18.229	245	100	Malate
18.249	304	80	GABA
18.191	351	60	unknown
17.4	174	40	
18.04	304	30	



# Metabolic profiling

- Metabolic profiling of plants
  - Automated analysis of metabolites (up to 25.000) by GC-MS techniques in libraries of T-DNA mutants
  - Identification of interesting (even commercially interesting) mutants



# Metabolic profiling

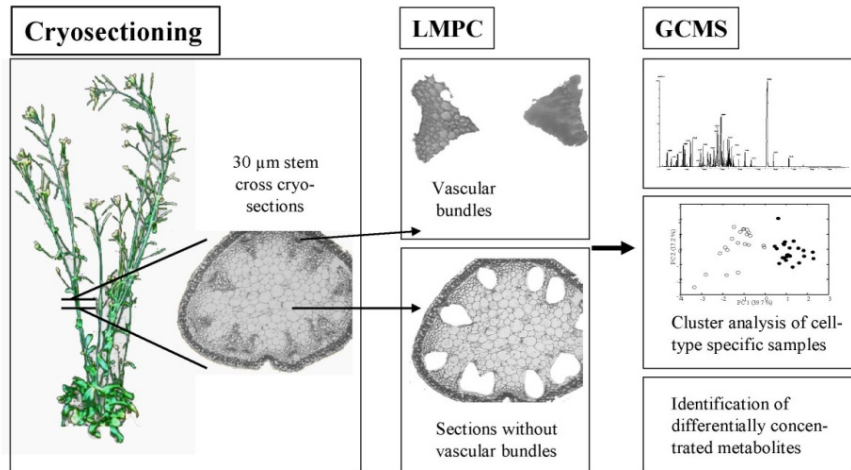
- Metabolic profiling of plants
  - Automated analysis of metabolites (up to 25.000) by GC-MS techniques in libraries of T-DNA mutants
  - Identification of interesting (even commercially interesting) mutants
  - Fast and easy isolation of genes through identification of sequences adjacent to T-DNA





# Metabolic profiling

- Metabolic profiling of plants
  - Possibility to use special techniques, e.g. [microdissection](#)

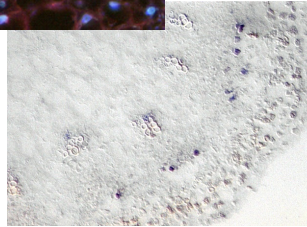
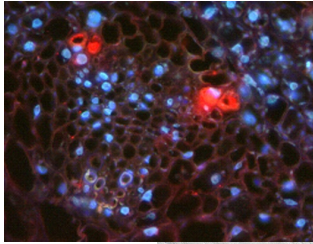


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# Expression profile

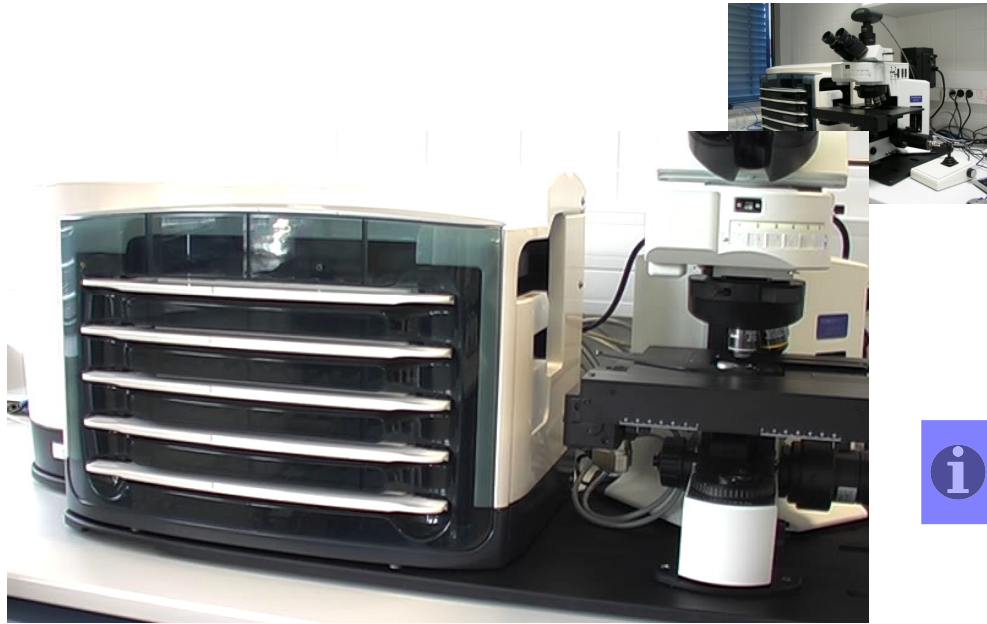
- Identification of mutants with a **change in the expression profile**
  - Analysis of **expression profile (pattern)** of the gene and identification of **mutants with altered expression pattern**



# Expression profile

- Identification of mutants with a change in the expression profile
  - Analysis of *expression profile (pattern)* of the gene and identification of *mutants with altered expression pattern*
  - Possibility of *partial automation* (virtual digital microscopy)

## Automated Microscopy Screening



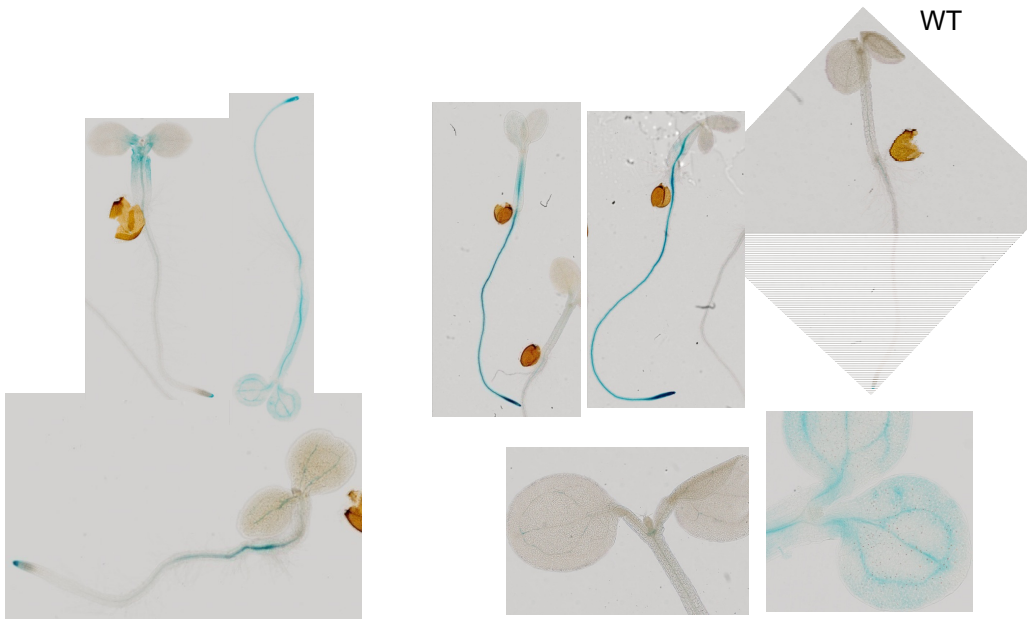
21

 CEITEC

Tady by mohla být reference zpět na CEITEC, jaké skvělé vybavení v něm je a jak dobře se vám s tím pracuje

Pokud tam nezůstane video, ikonu bych dala pryč

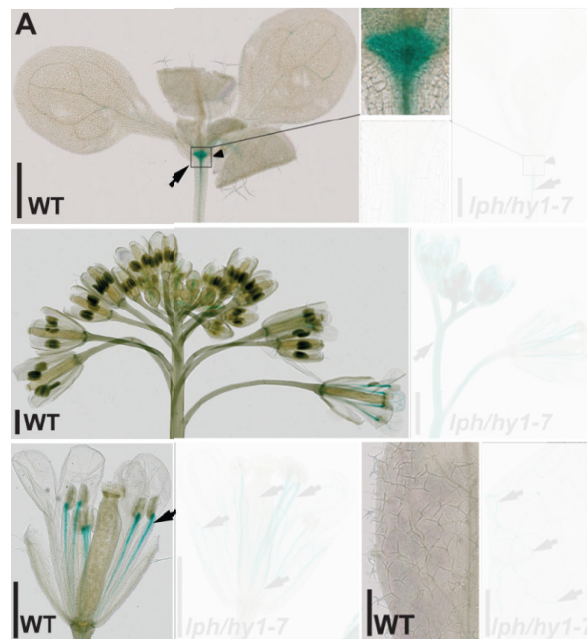
# Expression profile



22

CEITEC

## *LPH* controls expression of *CKI1*



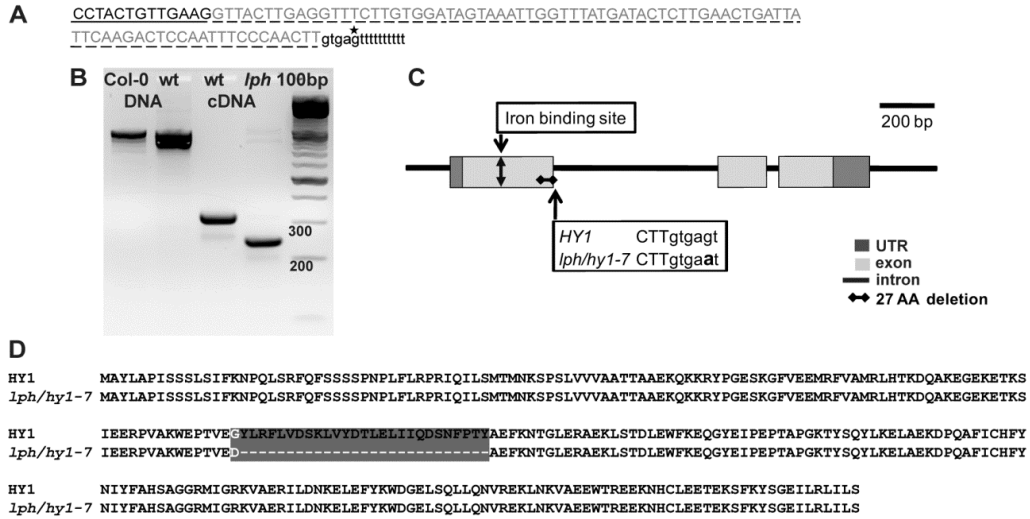
23

Dobisova et al., *Plant Physiol*, 2017

CEITEC

Dobré použití otázky „so what did we do“?

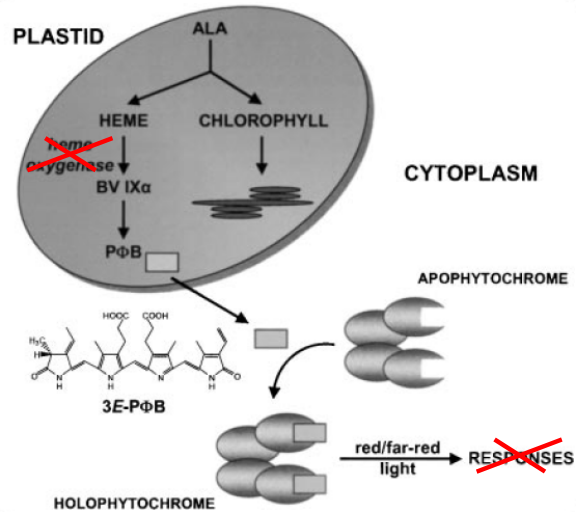
# *lph* is novel allele of HEME OXYGENASE 1



Dobisova et al., *Plant Physiol*, 2017

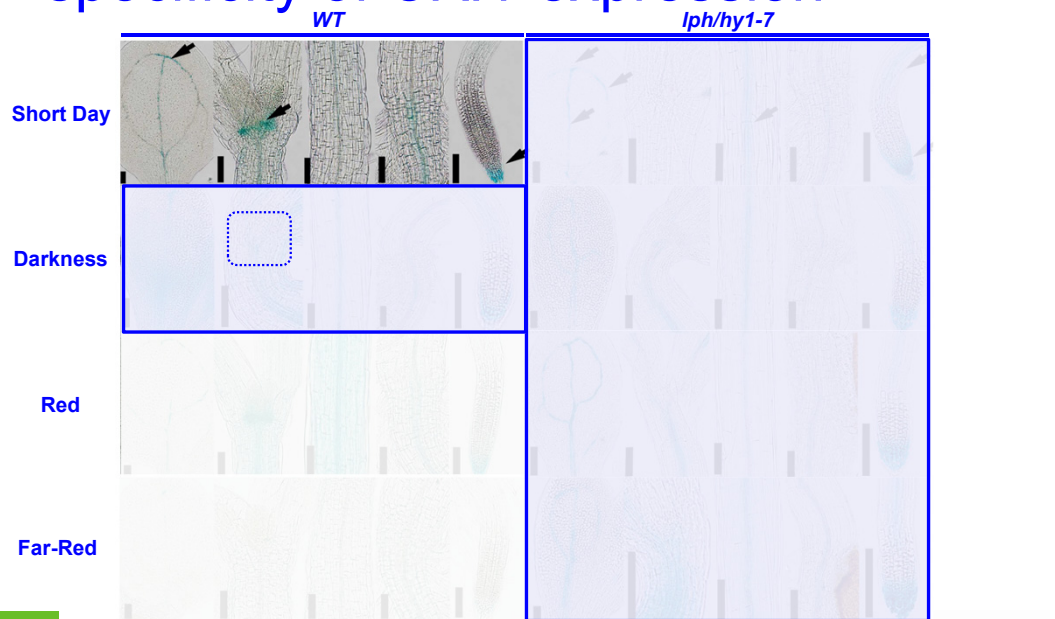


# *lph* is novel allele of HEME OXYGENASE 1



Terry et al., *Biochem Soc Trans*, 2002

# Light controls spatiotemporal specificity of *CKI1* expression



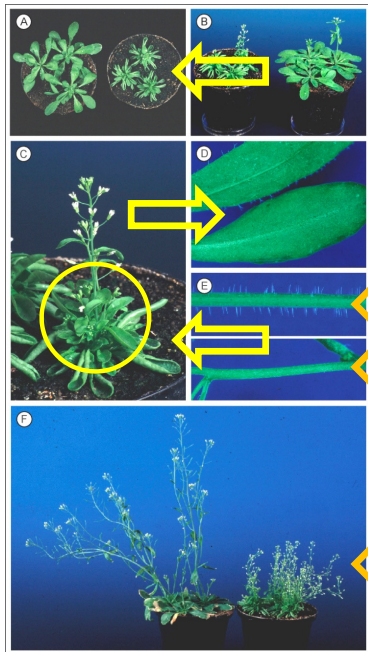
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    - plasmid rescue
    - iPCR

## Identification of mutated locus

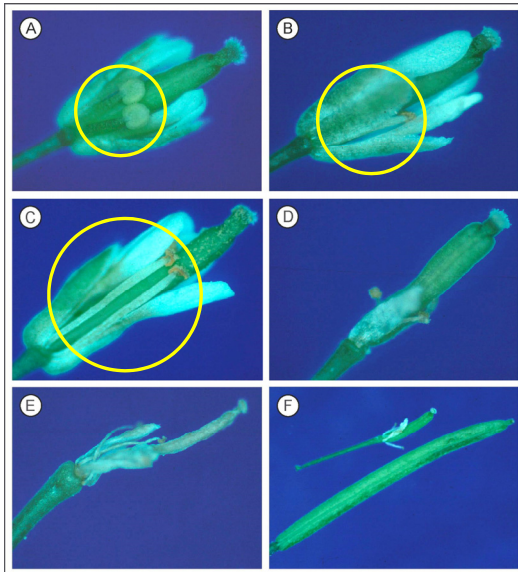
- Identification of chromosomal rearrangements responsible for bushy phenotype of *Arabidopsis*
  - Description of phenotype

## Identification of mutant



- Crinkled leaves
- Bushy phenotype (branching defective)
- No trichomes on leaves and stems
- Late senescence

## Identification of mutant



- Male sterility, defects in stamen filament elongation (A,B)

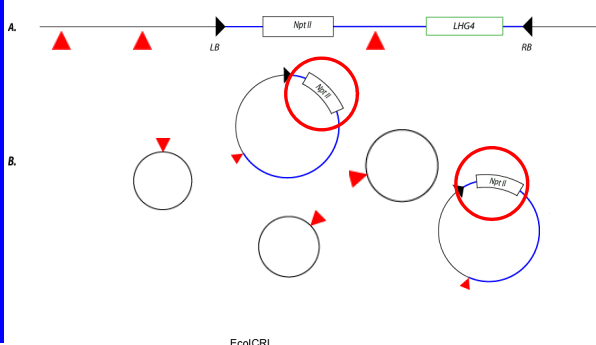
(compare with wild type C)

## Identification of mutated locus

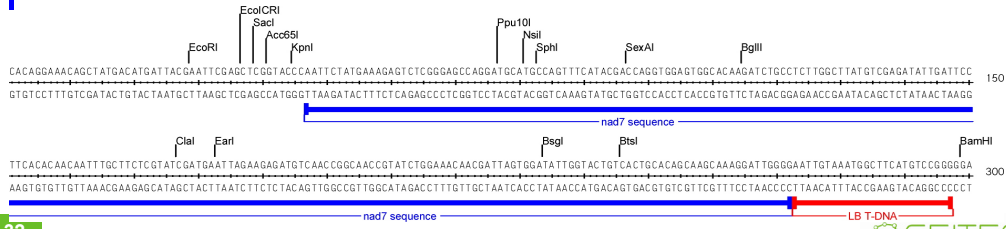
- Identification of chromosomal rearrangements responsible for bushy phenotype of *Arabidopsis*
  - Description of phenotype
  - Identification of T-DNA mutated region

# Identification of mutated locus

## 1. Identification of region of genomic DNA adjacent to the *left border* using *plasmid rescue*



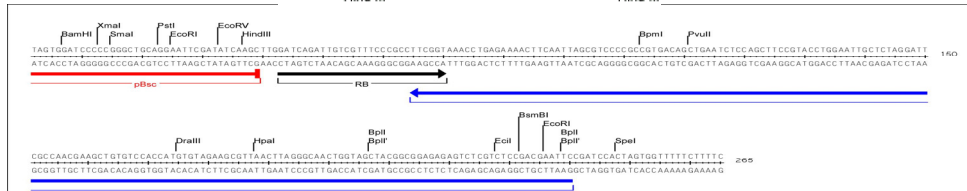
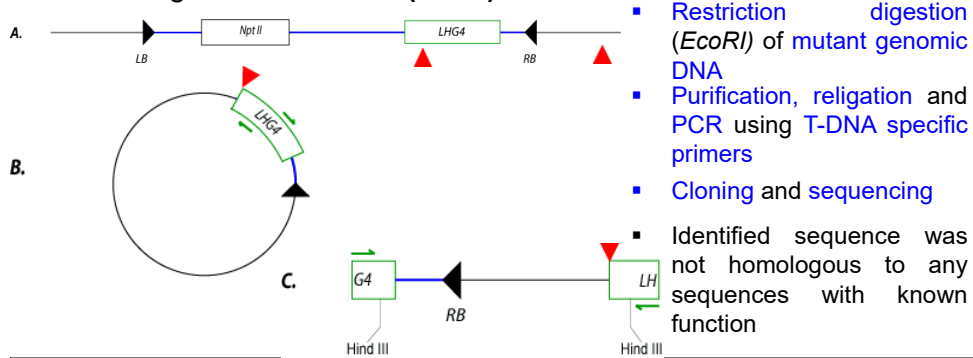
- Restriction digestion (*EcoRI*) of mutant genomic DNA
- Religation and transformation of *E. coli*
- Isolation of plasmid DNA from positively selected clones
- Identified sequence was identical to gene for *NAD7* coded by *mtDNA*





## Identification of mutated locus

### 2. Identification of region of genomic DNA adjacent to the *right border* using *inversion PCR* (iPCR)

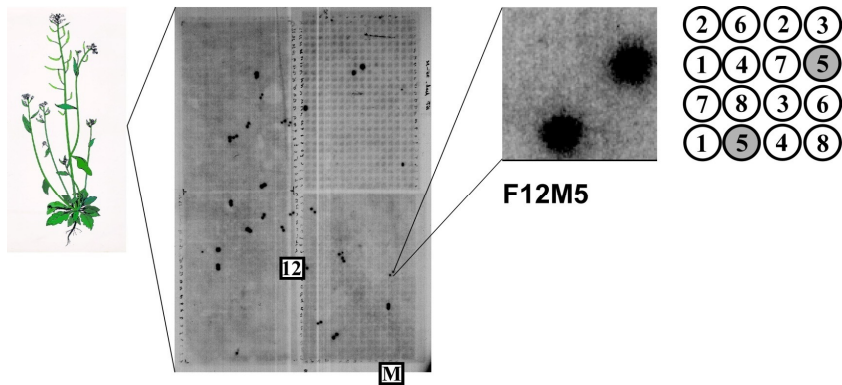


## Identification of mutated locus

- Identification of chromosomal rearrangements responsible for bushy phenotype of *Arabidopsis*
  - Description of phenotype
  - Identification of T-DNA mutated region
  - Localization of T-DNA insertion site in *Arabidopsis* genome

### Searching in library IGF-BAC

- Genome library containing 10.752 clones with an average size of an insert of 100 kb
- Bacterial clones arranged in the microtiter plates
- Library loaded onto nylon filters for hybridization with the radiolabeled probe



## Mapping with IGF-BAC database

### **I. Sequences adjacent to the left border of T-DNA**

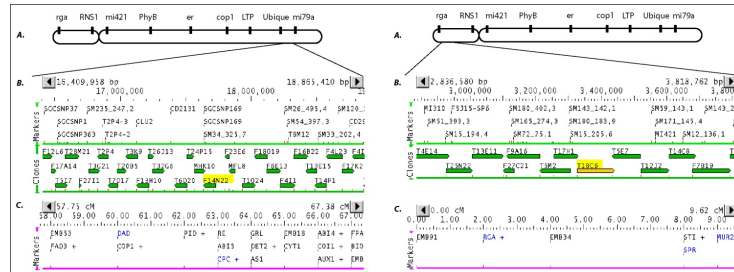
- 28 positively hybridizing clones in total
- 19 of them located on chromosome 2
- 18 of them similar with mtDNA

### **II. Sequences adjacent to the right border of T-DNA**

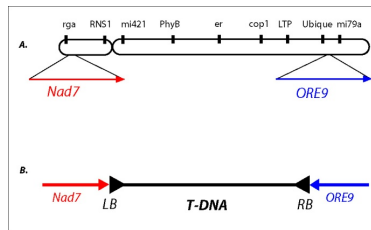
- 6 positively hybridizing clones in total
- all of them located on chromosome 2

## Localization of genomic T-DNA adjacent to both left and right T-DNA borders on chromosome 2

Sequences adjacent to *right* and *left* border of T-DNA



- There was probably an inversion of almost entire chromosome 2



# Outline

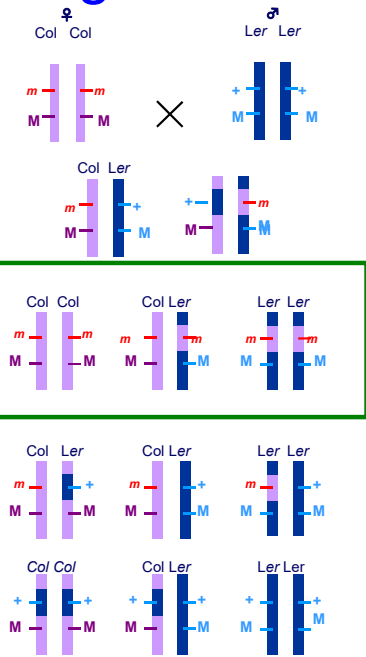
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  - Positional Cloning

# Identification of mutated locus

- Positional cloning
  - Principle: **co-segregation analysis** of **segregating population** (mostly of offspring of backcrosses) with **molecular markers**
  - **SSLP** (Simple Sequence Length Polymorphism)
    - Polymorphism of genome (PCR products) length, amplified using specific primers
  - **RFLP** (Restriction Fragment Length Polymorphism)
    - Detection by Southern blot (PCR after digestion of the genomic DNA and ligation of adapters)
  - **CAPS** (Cleaved Amplified Polymorphic Sequence)
    - Restriction fragment length polymorphism, genome segments amplified by PCR
  - **RAPD** (Randomly Amplified Polymorphic DNA)
    - Polymorphism of length of randomly amplified genome segments, using short 8-10bp primers

# Positional cloning

Preparation of mapping population

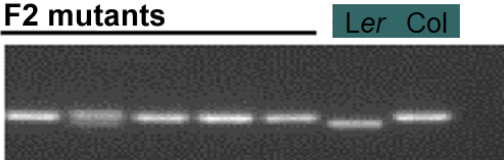




**Recombinant analysis** – determining the percentage of recombination between mutation and molecular marker

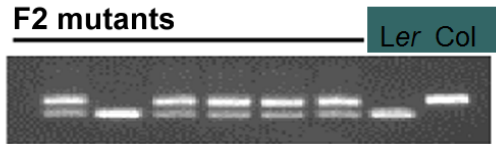
$$r [\%] = \frac{\text{number of chromosomes of Col}}{\text{number of all the chromosomes}} \times 100$$

F2 mutants



marker I – linked  
5 mutants  
 $1/10 \times 100 = 10\%$

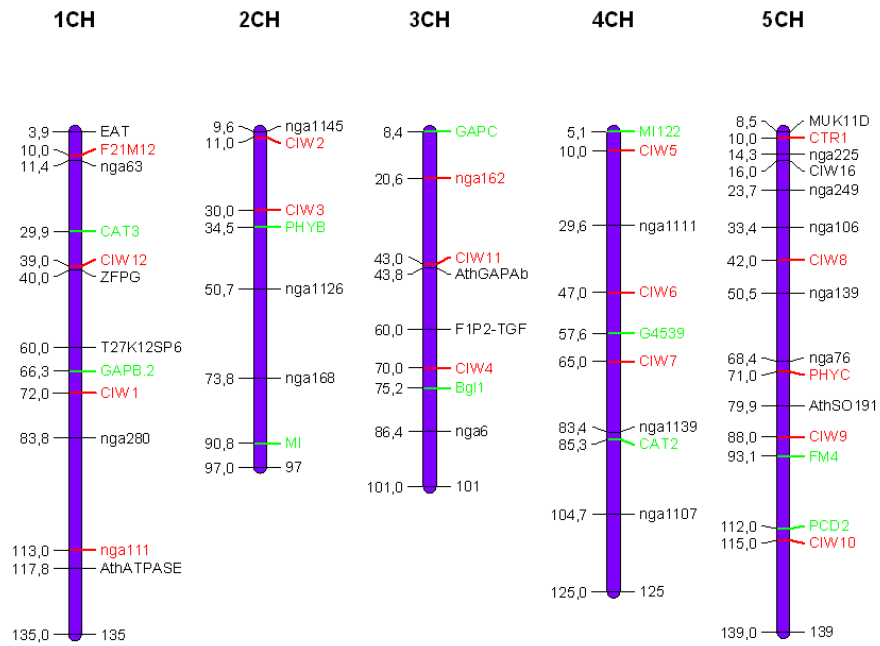
F2 mutants



marker II - no linkage  
6 mutants  
 $7/12 \times 100 = 58\%$

- Analysis of approximately 2000 mutant plants
- Determining the closest (still segregating) marker
- Identification of mutation by sequencing

## Map of DNA molecular markers



# Markers for fine mapping

- AGI Map
- Lister & Dean RI
- Classical
- mi-RFLP
- Goodman
- GoodmanBAC
- TIGR
- Finkelstein
- Altmann

## Maps for Chromosome 2

for all Maps: [Search Options](#)

Selected Maps:

Display All Rows:

[MapViewer Home](#)  
[Release Note](#)  
[View Print-Version](#)

**AGI Map**

Zoom to:

Zoom up to 200x to see genes!

Search by name (e.g. UFO)

Select range (e.g. 1500-2000)

[AGI Map color key](#)

0 bp 2,463,170 bp

0 1,000,000 2,000,000

SM130_329,3	ATPTR2_B	SM104_73,6	<b>CIW2</b>	SM115_237,0	SM12_274,4	CIC9A
SM134_72,6	SM71_250,0	RNS1	SGCSNP180	SM46_294,4	SGCSNP129	T6PS-
SM61_322,0	SM45_234,7	SM57_225,2	SM04_90,5	SM158_96,3	SGCSN	
SM122_129,8	MI320	SM206_129,8	SM121_171,6	SM14_243,3	T6PS-	
SM122_129,8	NGA1145	SM254_364,8	G4532	M497A	T6PS-	
SM253_310,7	NGA114	SM17_241,6	G4553		T6PS-	
SM121_93,6		M246	SM46_411,1		T6PS-	
	FGA		SGCSNP111		T12A:	
			SM138_120,2		SM1:	
			SM73_258,4			
			SM233_170,9			

NOR\_2 F219 T23K3 T16F16 T17M13 F19B11 F31J2 T103 F1013 F16510 T25M19 F

F23H14 T8011 F504 T8K22 T18F12 T18C20 T16B23 F2810 F15L11 T3P4 F5K7

F1QA0 F23114 T20P6 T4M0 F3C11 T23015 F5G3 T20P20 T17C22

F14H20 F7D11 T6F5

**Lister & Dean RI**

Zoom to:

Search by name (e.g. UFO)

Select range (e.g. 1500-2000)

0.00 cM 11.97 cM

0,00 5,00 10,00

TEL2N	RGA +	FK1 +	VE012	MI320	F53_2
NOR2	SGCSNP180 +		ATPTR2_B	NGA1145 +	
				AT6ST2B	
				MIT201A +	

# Key Concepts

- **Forward genetics** allows targeted screening for **interesting phenotypes**, whose **association** with **a given gene/locus** is unknown
  - Employs both **insertional mutagenes** as well as **point mutations**
    - **Insertional mutation**
      - (mostly) **loss-of-function** mutation
        - Identification via
          - iPCR
          - plasmid rescue
      - **Point mutation**
        - Both **loss-of-function** as well as
        - **gain-of-function** mutations
          - Identification via
            - **map-based cloning**
            - **GWAS**

# Discussion