

LOSCHMIDT
LABORATORIES



Analysis of protein structures



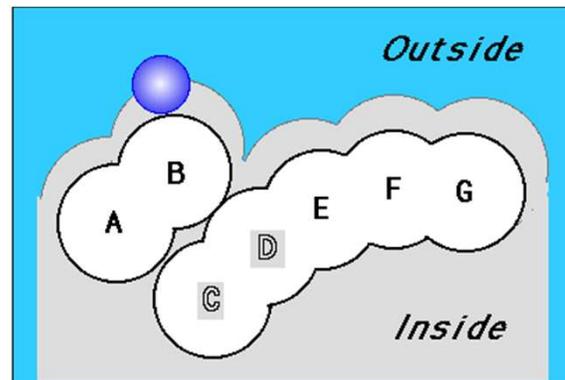
Outline

- ❑ Residue solvent accessibility
- ❑ Protein solubility
- ❑ Molecular interactions
- ❑ Functional sites
 - Binding sites
 - Transport pathways

Residue solvent accessibility



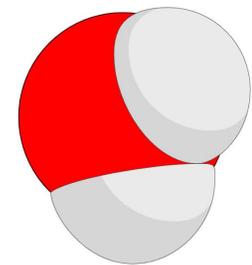
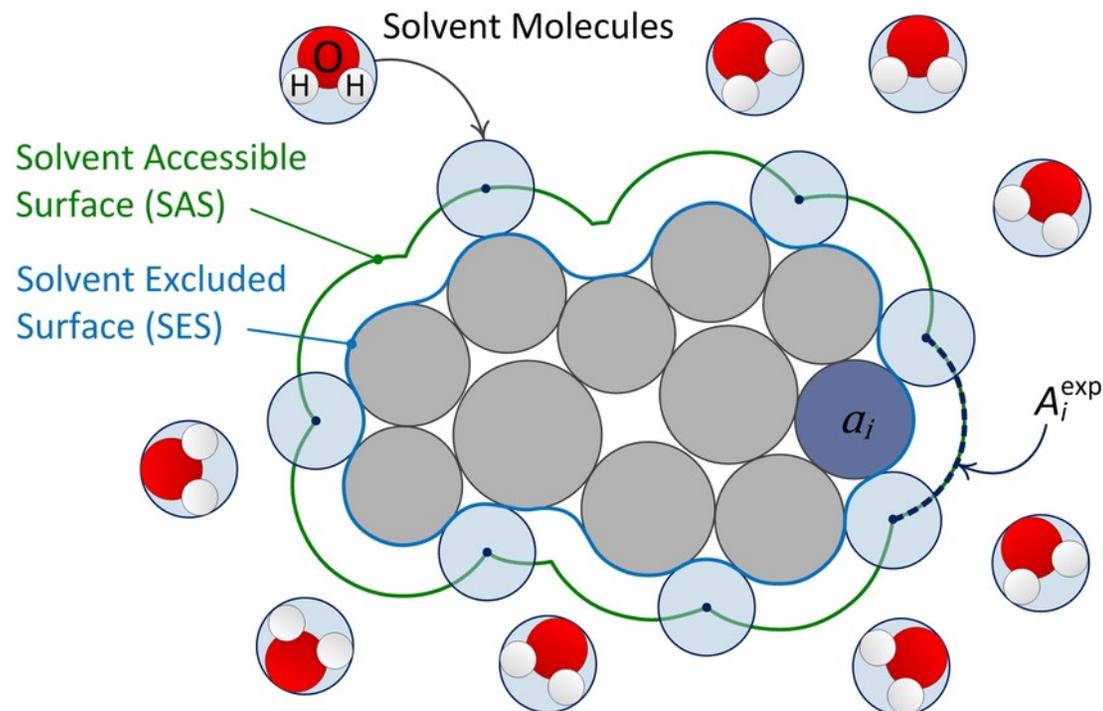
- ❑ Solvent **accessible surface area** (ASA or SASA, in \AA^2):
the extent to which a residue in a protein structure is accessible to the solvent
- ❑ Typically calculated by **rolling** a spherical **probe** of a particular radius over a protein surface and summing the area that can be accessed by this probe on each residue



Residue solvent accessibility



- ❑ Solvent **accessible surface area** (ASA, SASA or SAS, in \AA^2)
- ❑ Solvent excluded surface (SES) – also known as molecular surface, or Connolly surface area



Water radius $\approx 1.4 \text{\AA}$

Residue solvent accessibility



- Relative accessible surface area (rASA)
 - Ratio of the actual accessible area of a given residue
$$rASA = ASA / ASA_{MAX}$$
 - Enables **comparison** of accessibility of **different amino acids** (e.g., long extended vs. spherical amino acids)
- Simplified two state description
 - **Buried** vs. **exposed** residues
 - *threshold* for differentiating buried vs. surface residues is not well defined (usually 15–25 %)
 - $rASA < threshold \Rightarrow$ buried; $rASA \geq threshold \Rightarrow$ exposed

Residue solvent accessibility – programs



□ Naccess

- <http://www.bioinf.manchester.ac.uk/naccess/>
- Calculates atomic and residue ASAs and rASA for PDB files
- Adjustable settings (e.g., probe size, atomic radii,...)

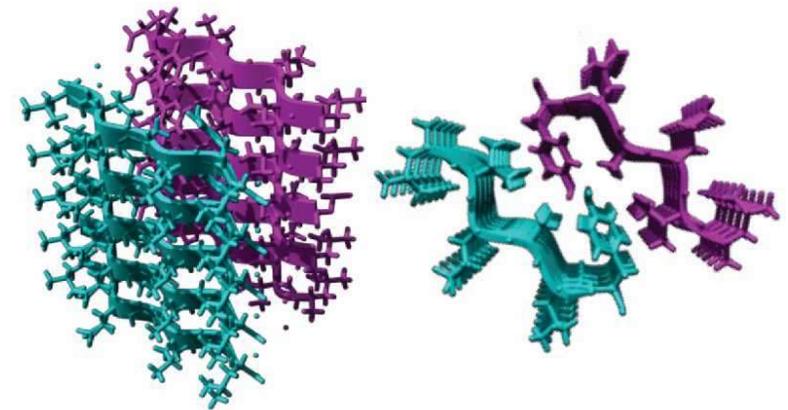
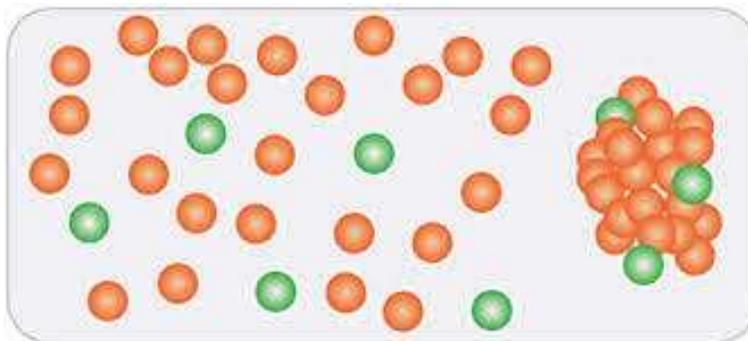
□ DSSP

- <https://swift.cmbi.umcn.nl/gv/dssp/>
- Assignment of secondary structures and calculation of ASAs of residues

Protein solubility



- ❑ Concentration of protein in saturated solution that is in equilibrium with solid phase
- ❑ For proteins expressed in the lab, it depends on
 - ❑ Hydrophilic/hydrophobic balance of the solvent-exposed residues
 - ❑ Aggregation-prone regions (APRs) – mainly hydrophobic residues prone to form beta-structures
 - ❑ Protein expressibility in the cells

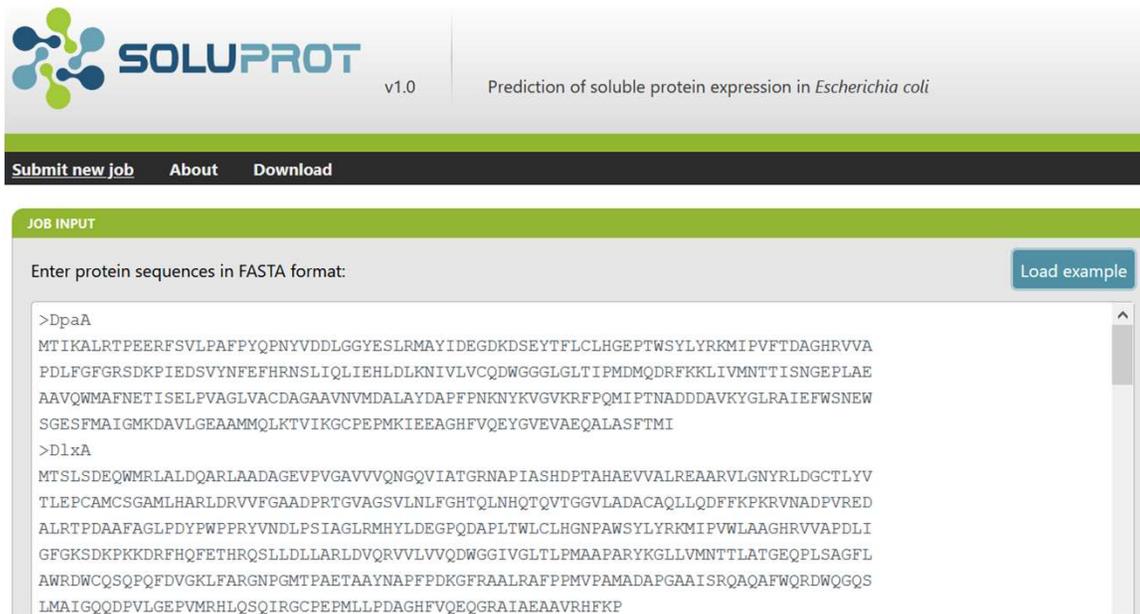


Cross-beta spines of amyloid fibrils

Protein solubility

❑ SoluProt

- <https://loschmidt.chemi.muni.cz/soluprot/>
- Soluble expression of protein sequences in *E.coli*
- Based on machine learning



SOLUPROT v1.0 Prediction of soluble protein expression in *Escherichia coli*

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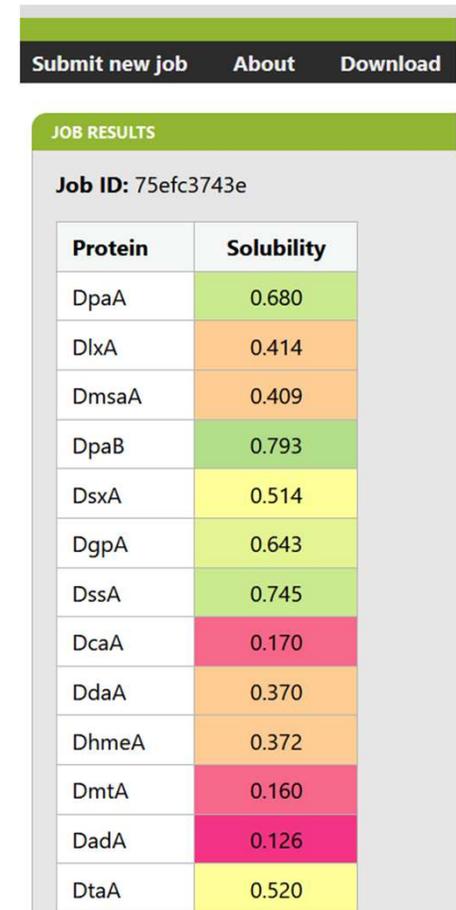
JOB INPUT

Enter protein sequences in FASTA format: [Load example](#)

```
>DpaA
MTIKALRTPEERFSVLPAPFYQPNYVDDLGGYESLRMAYIDEGDKDSEYTFCLHGEP TWSYLYRKMIPVFTDAGHRVVA
PDLFGFGRSDKPIEDSVYNFEFHRNSLIQLIEHLDLKNIVLVCQDWGGGLGLTIPMDMQDRFKKLIVMNTTISNGEPLAE
AAVQWMAFNETISELPVAGLVACDAGAAVNVMDALAYDAPFPNKNYKVGKRFPPQMIPTNADDDAVKYGLRAIEFWSNEW
SGESFMAIGMKDAVLGEAAMMLKTVIKGCPEPMKIEEAGHFVQYEGVEVAEQALASFTMI

>DlxA
MTSLSDEQWMRLALDQARLAADAGEVPVGVAVVQNGQVIATGRNAPIASHDPTAHAEVVALREAAARVLGNYRLDGCTLYV
TLEPCAMCSGAMLHARLDRVVFVGAADPRTGVAGSVLNLFGHTQLNHQTQVTGGVLADACAQLLQDFFKPKRVNADPVRED
ALRTPDAAFAGLPDYFPWPPRYVNDLPSIAGLRMHYLDGEPQDAPLTLWCLHGNPAWSYLYRKMIPVWLAAGHRVVPDLI
GFGKSDKPKKDRFHQFETHRQSLDLLLARLDVQRVVLVVDWGGIVGLTLPMAAPARYKGLLVMTTLATGEQPLSAGFL
AWRDCWCSQPQFDVVGKLFARGNPGMTPAETAAYNAPFPDKGFRAALRAFPMPVPMADAPGAAISRQAQAFWRDQWGGQS
LMAIGQQDPVLGEPVMRHLQSQIRGCPEPMLLPDAGHFVQEQQGRAIAEAAVRHFKE
```

Input



Submit new job About Download

JOB RESULTS

Job ID: 75efc3743e

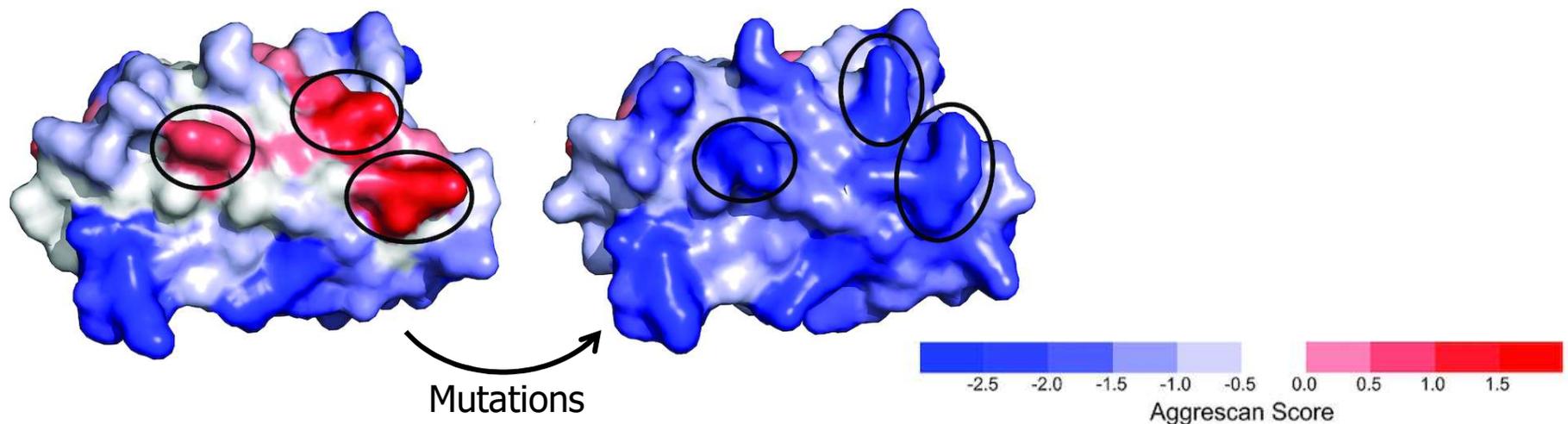
Protein	Solubility
DpaA	0.680
DlxA	0.414
DmsaA	0.409
DpaB	0.793
DsxA	0.514
DgpA	0.643
DssA	0.745
DcaA	0.170
DdaA	0.370
DhmeA	0.372
DmtA	0.160
DadA	0.126
DtaA	0.520

Output

Protein solubility

□ Aggrescan3D

- <http://biocomp.chem.uw.edu.pl/A3D2/>
- Predicts the aggregation propensities by identifying APRs
- Can introduce mutations and predict impact on stability and aggregation-propensity
- Can account for protein flexibility (“dynamic mode”)



Molecular interactions



- Assessment of the interactions within a protein structure and between proteins in an assembly is **essential** to understand the **molecular basis of stability and function** of proteins and their complexes

Types of interactions

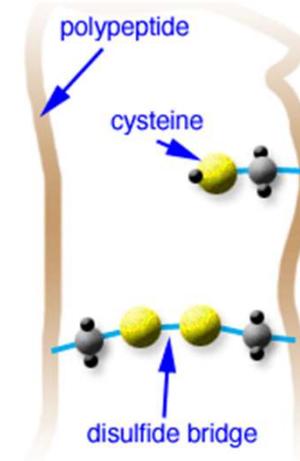
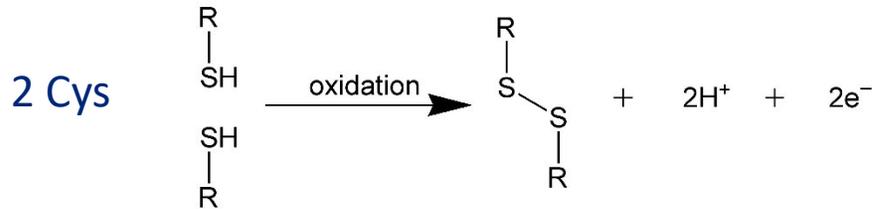


- ❑ charge-charge (ionic) interactions
 - present in charged residues; e.g. salt bridges
- ❑ hydrogen bonds (H-bonds)
 - donor and acceptor atoms sharing hydrogen
- ❑ aromatic (π - π) interactions
 - attractive interaction between aromatic rings
- ❑ van der Waals (vdW) interactions
 - between any two atoms
- ❑ hydrophobic interactions
 - entropic origin; important for hydrophobic residues

Types of interactions

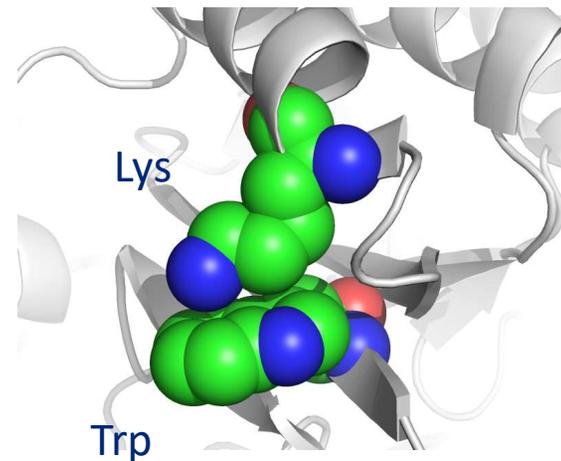
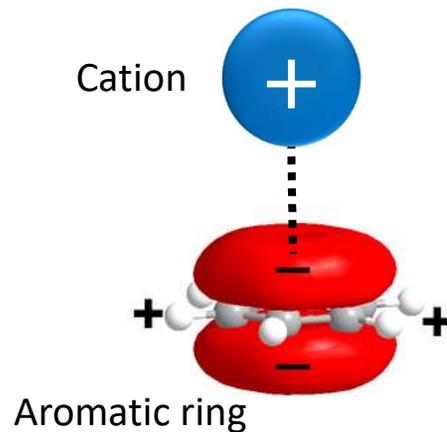


- disulfide bonds (cysteine bridges)



- cation- π interactions

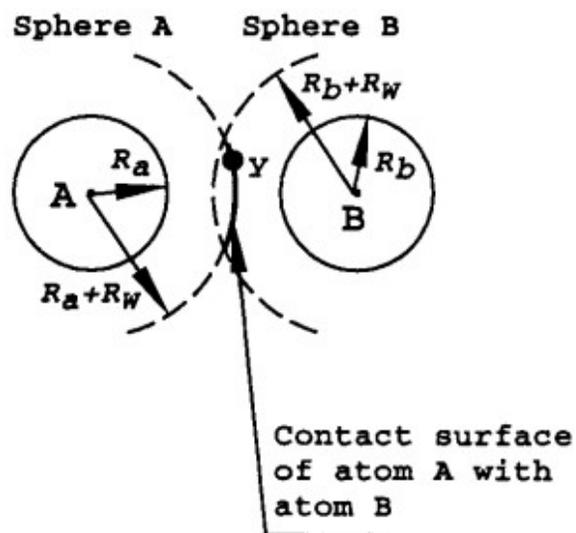
- electrostatic interaction of a positively charged residue (Lys or Arg) with an aromatic residue (Phe, Trp, or Tyr)



Molecular interactions – how to identify?



- criteria for recognizing various types of interactions
 - geometric rules (distances, angles)
 - atom types
 - energetics (physicochemical rules)
 - contact surface area between atoms

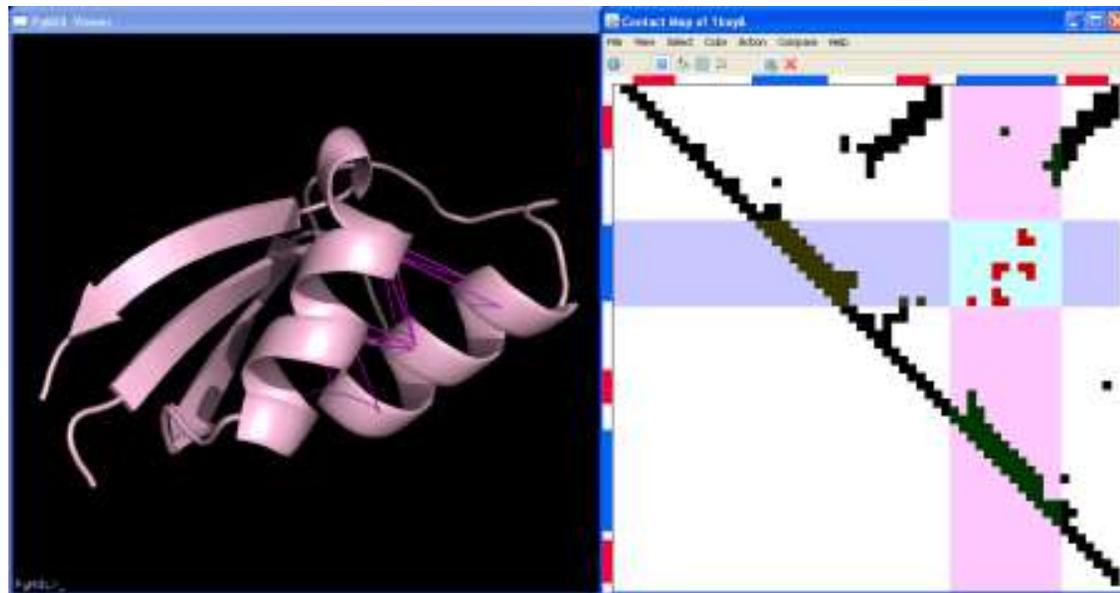


Molecular interactions – programs

- CMView

- <https://www.bioinformatics.org/cmview/>

- represents **residue-residue contacts** within a protein or between proteins in a complex in the form of a **contact map**
 - 3D visualization using PyMol



Molecular interactions – programs



- ❑ PIC (**P**rotein **I**nteraction **C**alculator)
 - <http://pic.mbu.iisc.ernet.in/>
 - identifies **various interactions** – disulfide bonds, hydrophobic interactions, ionic (charged-charged) interactions, hydrogen bonds, aromatic–aromatic, aromatic–sulfur and cation– π interactions within a protein or between proteins in a complex
 - uses standard criteria (atom types and geometry)

Molecular interactions – programs

□ PIC (Protein Interaction Calculator)

- <http://pic.mbu.iisc.ernet.in/>

INTRAPROTEIN INTERACTIONS

* This option can accept a monomeric or a multichain protein file.

Upload a file in PDB format: No file chosen

Hydrophobic Interactions Enter the interaction cut-off value (Default 5A)

Disulphide Bridges

Main Chain–Main Chain Hydrogen Bonds

Main Chain–Side Chain Hydrogen Bonds

Side Chain–Side Chain Hydrogen Bonds

Ionic Interactions Enter the interaction cut-off value (Default 6A)

Aromatic–Aromatic Interaction : cut-off value to (Default 4.5A to 7A)

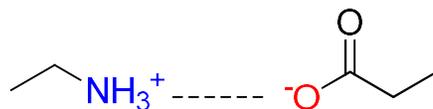
Aromatic–Sulphur Interactions Enter the interaction cut-off value (Default 5.3A)

Cation–Pi Interactions Enter the interaction cut-off value (Default 6A)

To E-mail the results enter a valid e-mail address: (Optional)

Molecular interactions – programs

- ESBRI (Evaluating the Salt BRIdges in Proteins)
 - <http://bioinformatica.isa.cnr.it/ESBRI/introduction.html>
 - analysis of salt bridges interactions (ionic interaction + H-bond)
 - checks if at least one Asp or Glu side-chain carboxyl oxygen atom and one side-chain nitrogen atom of Arg, Lys or His are within a distance $\leq 4.0 \text{ \AA}$



Salt bridge

Residue 1	Residue 2	Distance
NZ ALYS A 11	OD1 ASP A 62	3.86
NZ ALYS A 11	OD2 ASP A 62	2.78
NZ ALYS A 11	OD2 ASP A 68	2.85
NZ BLYS A 11	OD1 ASP A 62	3.79
NZ BLYS A 11	OD2 ASP A 62	2.74
NZ BLYS A 11	OD2 ASP A 68	2.75
NH1 ARG A 46	OE1 GLU A 276	3.61

Functional sites



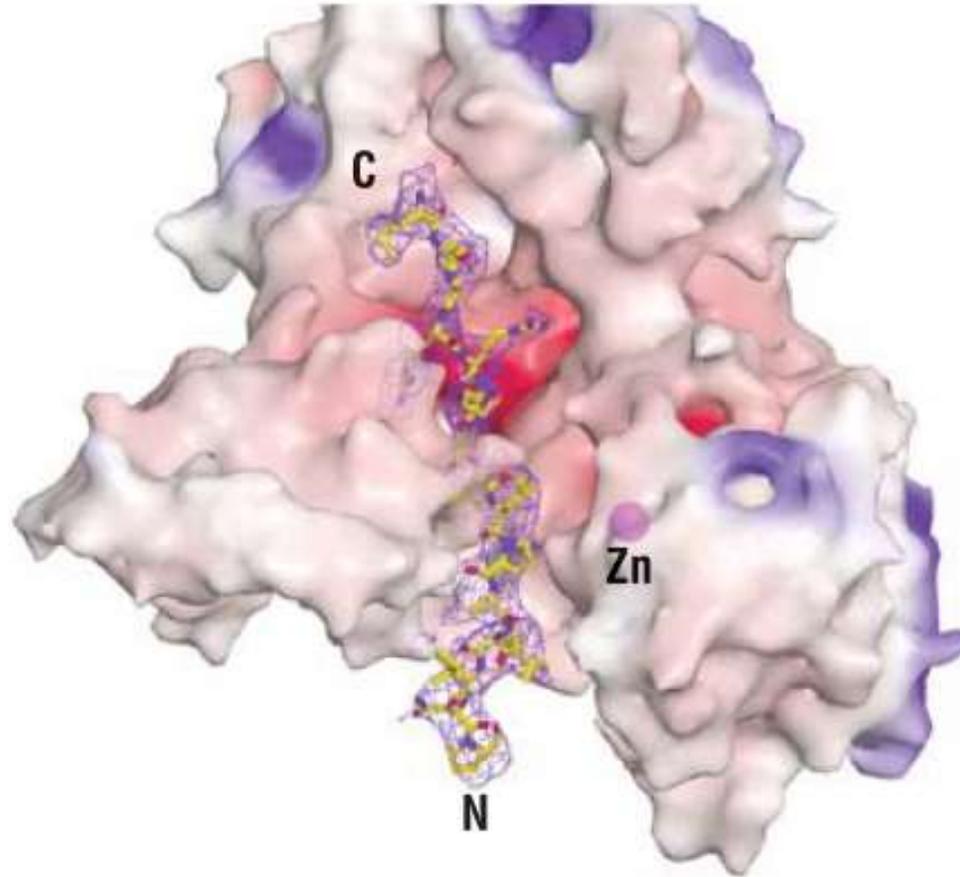
- ❑ binding sites
 - binding sites for small molecules
 - binding sites for macromolecules
- ❑ transport pathways
 - overall void
 - tunnels
 - channels

Binding sites



- ❑ sites on the protein that provides the complementarity for the bound molecule (ligand)
 - **binding site** – its function is molecular recognition
 - **active/catalytic site** – special case of the binding site – its function is to promote chemical catalysis (break/formation of covalent bonds)
- ❑ binding involves the formation of **non-covalent interactions** between the protein and the bound molecule
- ❑ bound molecule – **small molecule** or **macromolecule**
- ❑ binding is usually **very specific** – complementarity in shape and charge distribution between the site and bound molecule

Binding sites



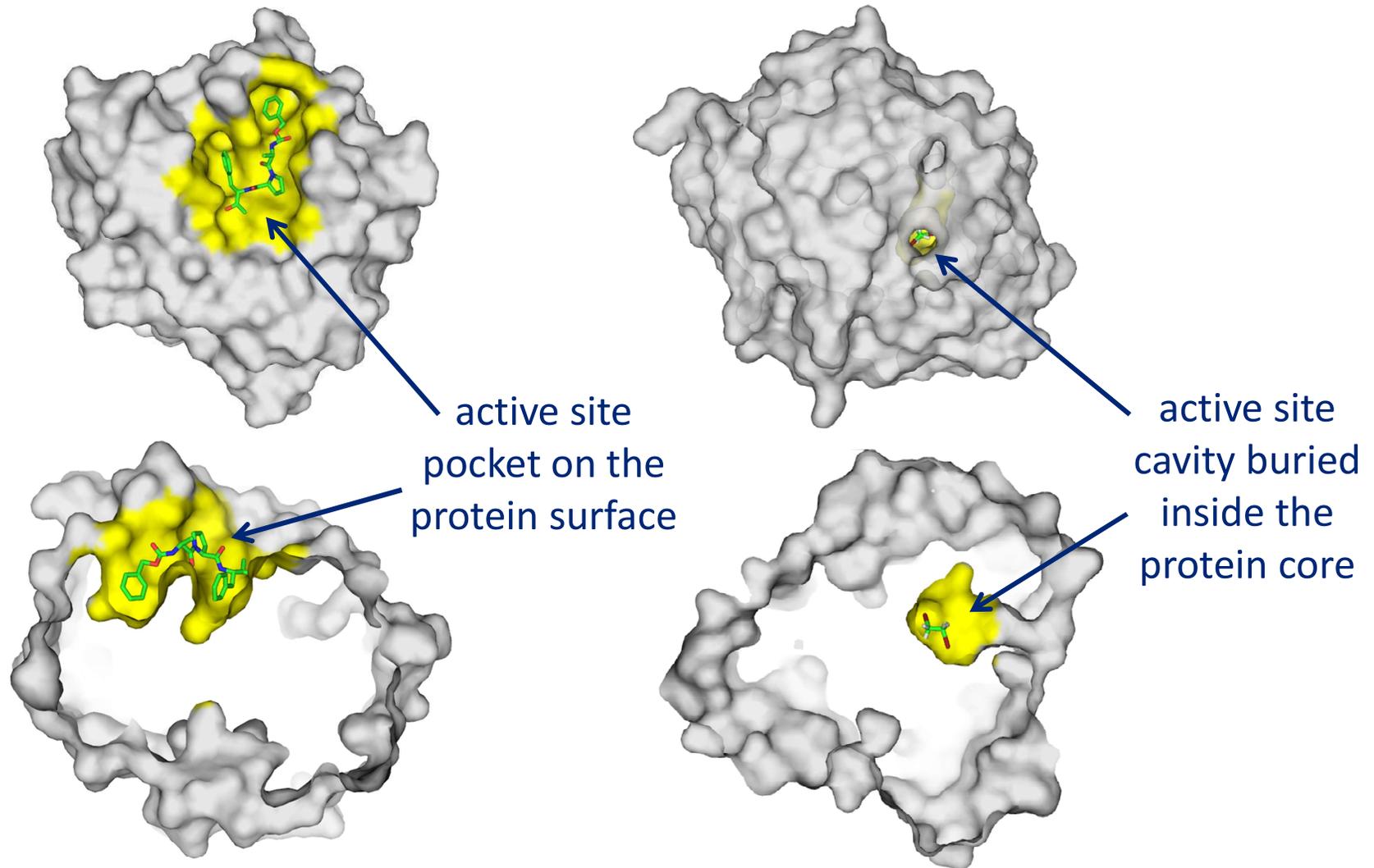
Complementarity in shape and charge distribution
between the active site and substrate

Binding sites for small molecules



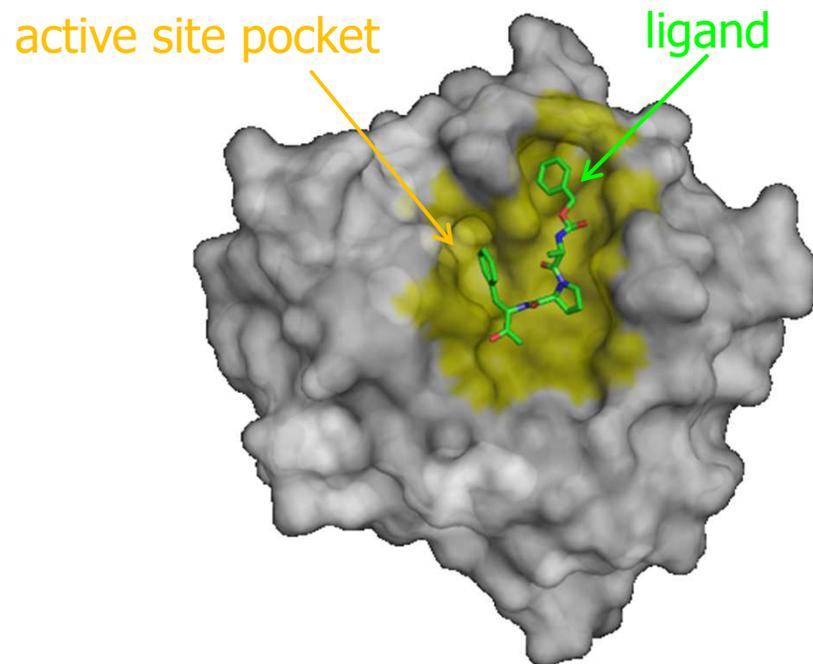
- ❑ typically internal **cavities**, or surface **pockets** or **clefts**
 - concave regions
 - provide **microenvironment** different from that of the bulk solvent (e.g., many residues with negative charge → very strong electrostatic field enabling binding of highly charged ligands)
 - often identifiable by a simple examination of the protein structure
- ❑ highly conserved by evolution
- ❑ low desolvation energy
- ❑ characteristic physicochemical properties

Binding sites for small molecules



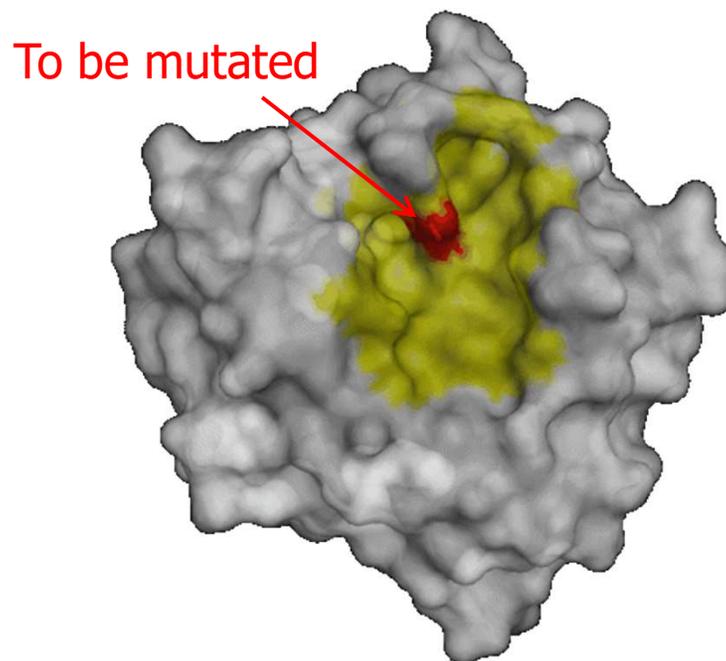
Binding sites for small molecules

- can be very ligand-specific



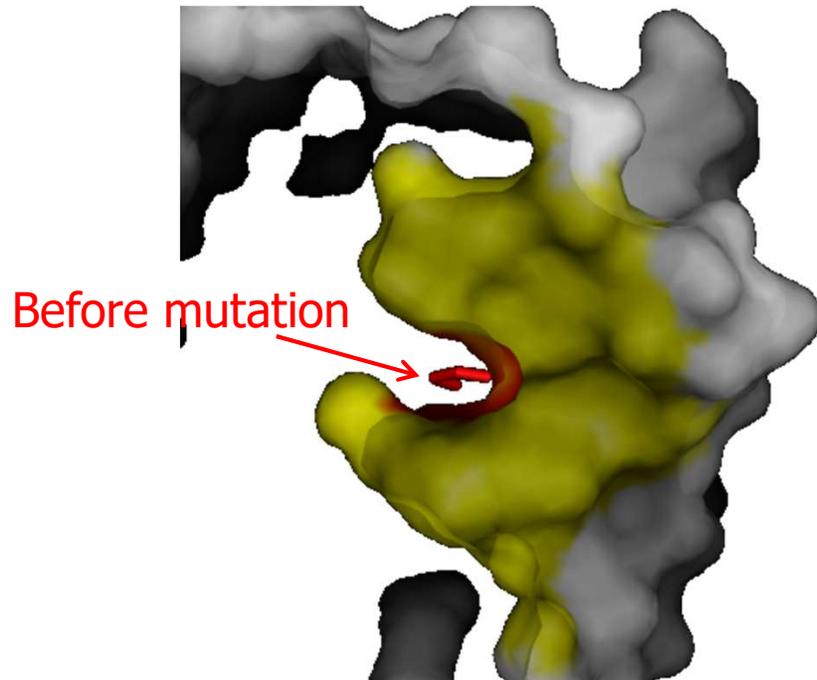
Binding sites for small molecules

- can be very ligand-specific



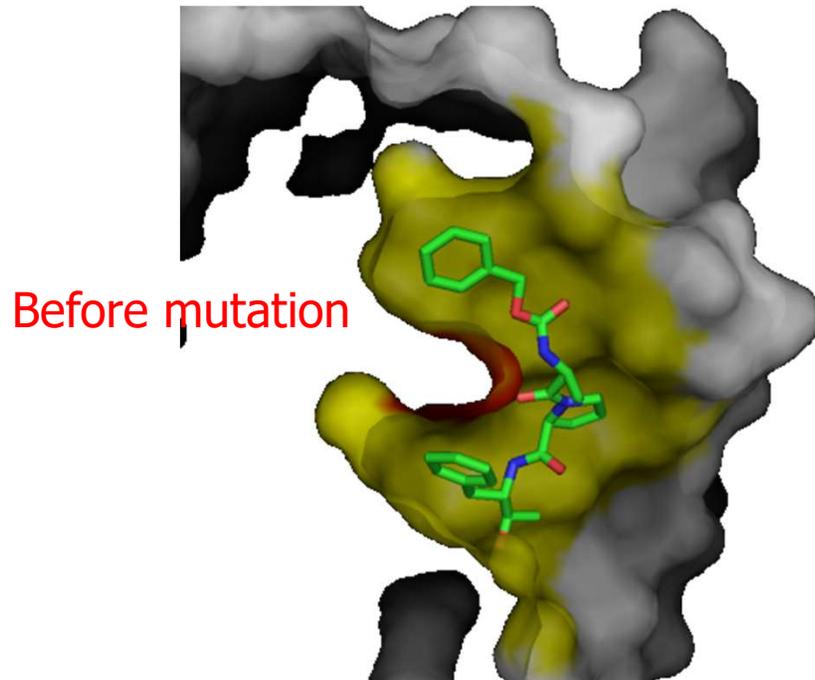
Binding sites for small molecules

- can be very ligand-specific



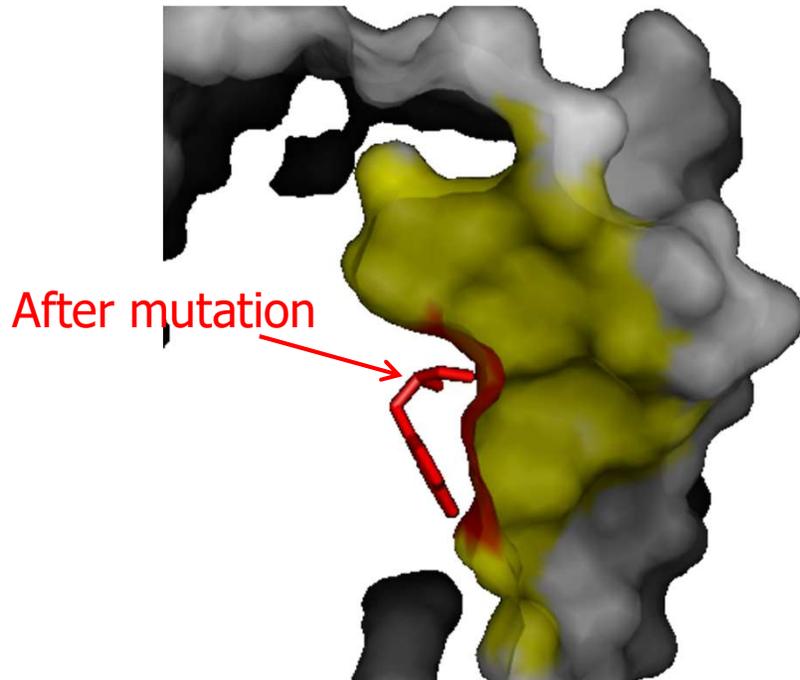
Binding sites for small molecules

- can be very ligand-specific



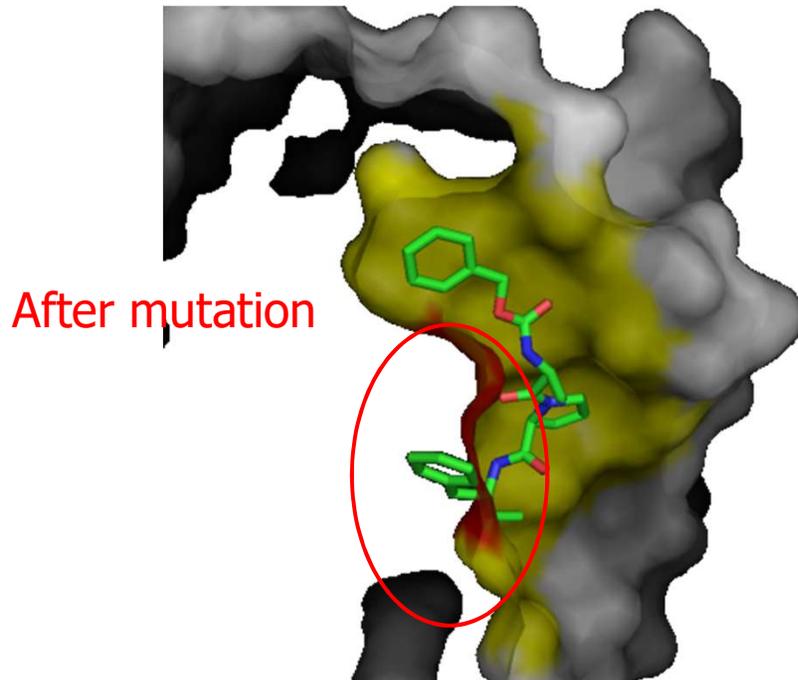
Binding sites for small molecules

- can be very ligand-specific



Binding sites for small molecules

- can be very ligand-specific



No longer a good fit!

Binding sites for small molecules



- ❑ approaches to identify binding sites:
 - ❑ evolutionary **conservation**
 - ❑ **“pocket”** detection
 - geometry based methods
 - energy based methods
 - ❑ binding site **similarity**
 - template-based methods
 - microenvironment-based methods

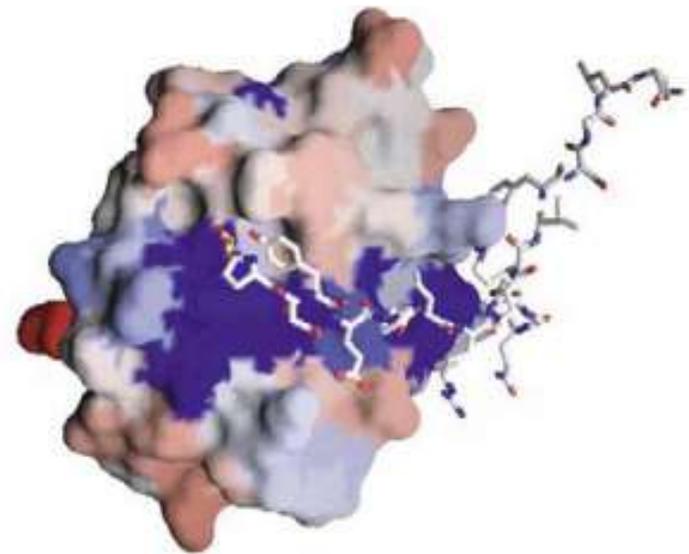
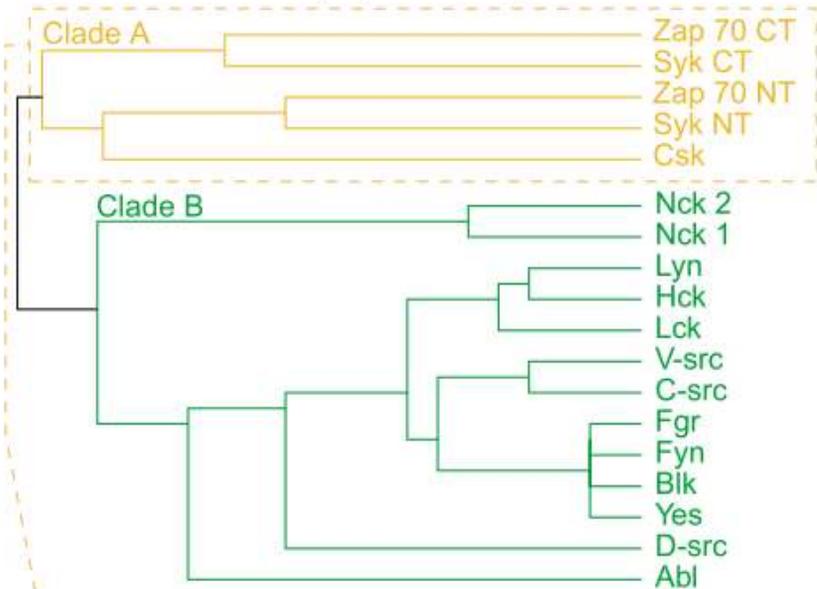
Evolutionary conservation



- ❑ residues important for **protein function** or **stability** tend to be **highly conserved** over evolution
- ❑ residue conservation in a set of related proteins can be derived from a **multiple sequence alignment**
- ❑ **mapping of conservation** on structure can reveal patches of conserved **surface** residues – potential binding sites
- ❑ protein interior usually more conserved than surface – not suitable for prediction of buried cavities
- ❑ **not very specific** – better to combine with other features

Evolutionary conservation

phylogenetic analysis



Zap 70 CT FFYGSISRRAEAEHLKLAGMADGLFLLRQCLRSLGGY
 Syk CT FFFGNITREEAEDYLVQGGMSDGLYLLRQSRNYLGGF
 Zap 70 NT WYHSSLTREEAERKLYSGAQTGKFLLRPRKEQ.GTY
 Syk NT WFHGKISRRESEQIVLIGSKTNGKFLIRARDNN.GSY
 Csk WFHGKITREQAERLLYPP..ETGLFLVRESTNYPGDY

conservation scoring

map evolutionary data on the structure

Evolutionary conservation

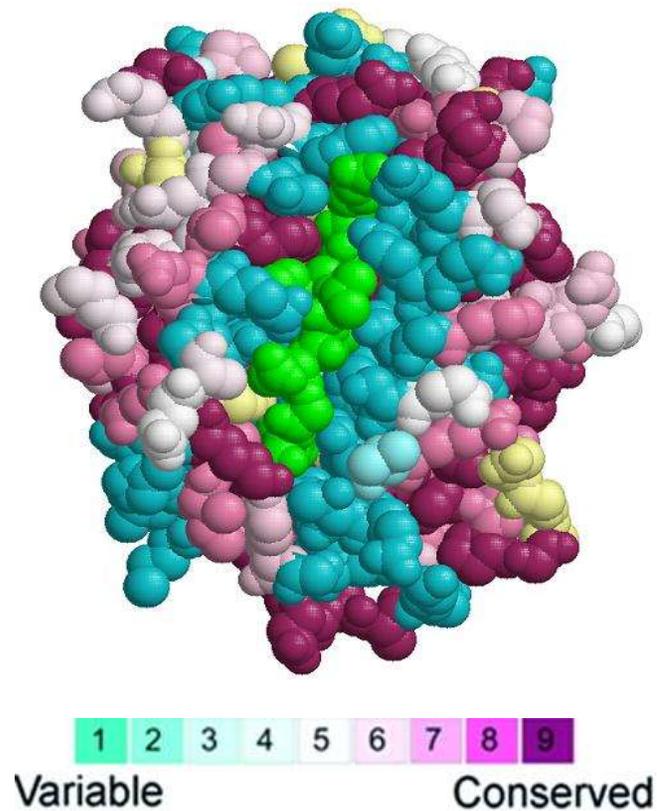


□ ConSurf

- <http://consurf.tau.ac.il/>
- estimates the level of **evolutionary conservation** of individual positions in protein and maps this information onto its 3D structure
- conservation score is derived based on the site-specific **evolutionary rates** calculated for each position by Rate4Site program
- **ConSurfDB** – pre-calculated conservation scores for all structures from wwPDB

Evolutionary conservation

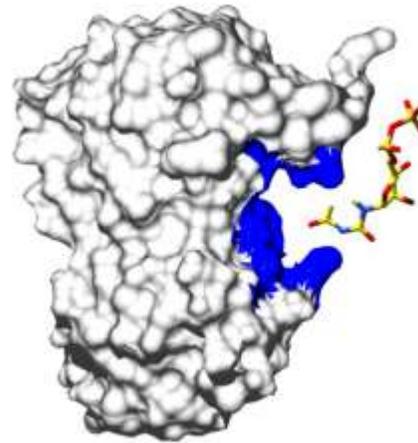
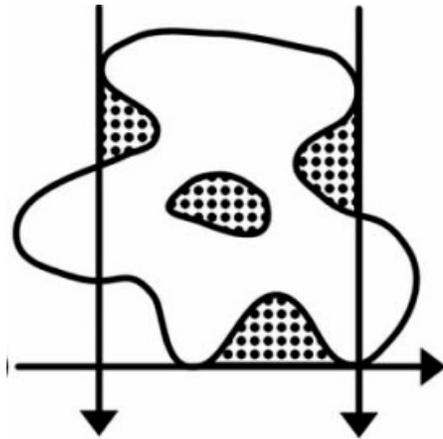
- ConSurf



“Pocket” detection



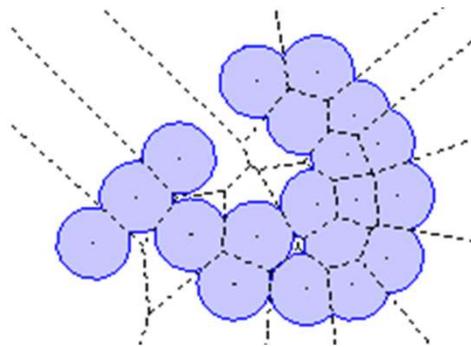
- analyze the protein surface for pockets (clefts, cavities)



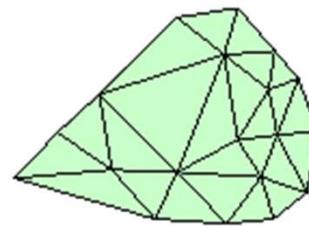
- **geometry-based** methods
 - define favorable cleft regions based on sterical assessments
- **energy-based** methods
 - define favorable cleft regions based on energetic evaluations

Geometry-based methods

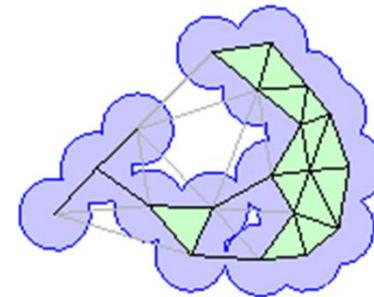
- ❑ Computed Atlas of Surface Topography of proteins (CASTp)
 - <http://sts.bioe.uic.edu/castp>
 - uses **computational geometry methods** including Delaunay triangulation, alpha shape and discrete flow theory
 - **measures the volume** and **surface area** of each pocket and cavity using the ASA model and molecular surface (Connolly) model



Voronoi
diagram



Delaunay
triangulation



alpha shape

Geometry-based methods



❑ SURFNET

- <http://www.ebi.ac.uk/thornton-srv/software/SURFNET/>
- probe sphere filling method

❑ fPocket

- <http://fpocket.sourceforge.net/>
- uses alpha shape and Voronoi tessellation
- mdpocket – version for analysis of pocket dynamics

❑ POCASA (POcket-CAvity Search Application)

- http://altair.sci.hokudai.ac.jp/g6/Research/POCASA_e.html
- combination of the grid system with probe sphere filling method

❑

Energy-based methods



- ❑ pockets are defined by energetic criteria
- ❑ evaluate the **interaction energy** between the **protein** and a **molecular fragment – probe** (e.g., a methyl, hydroxyl, or amine groups) to locate energetically favorable binding sites
- ❑ recently combined with other methods to assess the *ligandability* (ability of a cavity to bind ligands)

Note: 'druggability' is usually referred to the likelihood of finding orally bioavailable small molecules that bind to a particular target in a disease-modifying way. Ligandability is a necessary but not sufficient condition for druggability.

Energy-based methods

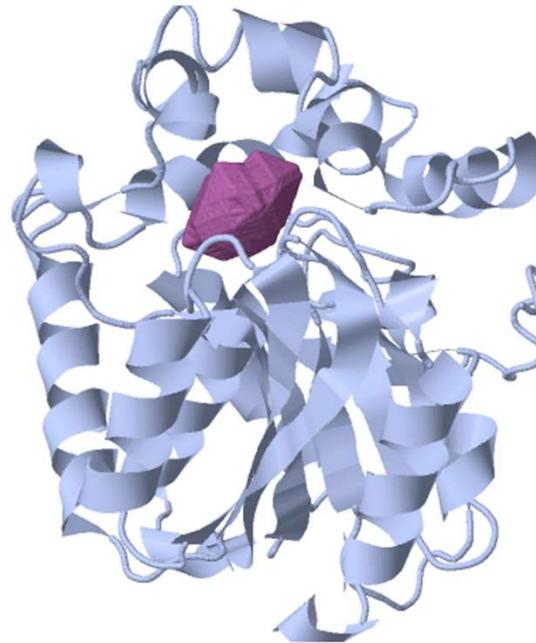


□ Cavity Plus

- <http://www.pkumdl.cn/cavityplus>
- applies Cavity program to detect the potential binding sites and rank them with ligandability and druggability scores
- extracts pharmacophore features within the cavities

Energy-based methods

□ Cavity Plus



▼ Cavity Results

No.	Pred. Max pKd ?	Pred. Avg pKd	DrugScore	Druggability ?	Surface ?	Residues ?
1	10.19	6.11	493.00	less druggable	<input checked="" type="checkbox"/>	More
2	8.87	5.66	-745.00	Undruggable	<input type="checkbox"/>	More
3	8.16	5.42	-420.00	Undruggable	<input type="checkbox"/>	More
4	7.87	5.32	-750.00	Undruggable	<input type="checkbox"/>	More
5	7.11	5.06	-1105.00	Undruggable	<input type="checkbox"/>	More
6	6.54	4.86	-992.00	Undruggable	<input type="checkbox"/>	More
7	5.90	4.64	-1123.00	Undruggable	<input type="checkbox"/>	More

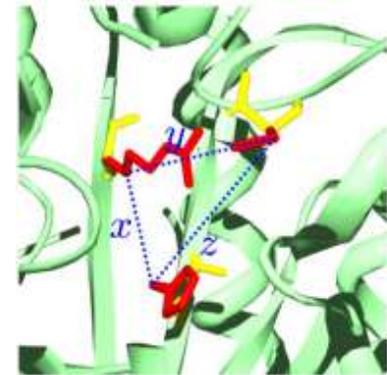
Binding site similarity



- prediction of binding site is based on its similarity to other (known) binding site

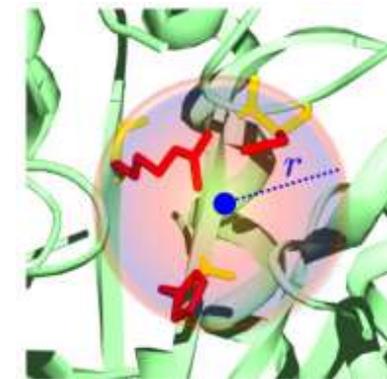
- **template-based** methods

- binding sites are represented by 3D templates



- **microenvironment-based** methods

- similar “microenvironments” associated with binding residues and their distances, physicochemical and evolutionary properties



Template-based methods



- ❑ definition and construction of **3D templates**
 - **local structural motifs, patterns or descriptors** that characterize the binding sites (e.g., functional groups)
 - capture essence of the binding site in protein
 - usually constraints on atom type and, occasionally, sequential relationships
- ❑ search in database for structures using **template as a query**
 - identification of structures with a given binding site
- ❑ compare the **query structure** against a 3D template database
 - identification of potential binding sites in the query structure

Template-based methods



□ PINTS

- <http://www.russelllab.org/cgi-bin/tools/pints.pl>
- enables to **compare** a protein structure against a **database of 3D templates** as well as a 3D templates against a **database of protein structures**
- additionally allows comparison of two structures
- the 3D template database includes ligand-binding sites and SITE annotations from PDB files

Template-based methods



□ ProFunc

- <http://www.ebi.ac.uk/thornton-srv/databases/profunc/>
- aims to identify the most likely **function** of a protein from its 3D structure
- uses series of methods, including fold matching, residue conservation, surface cleft analysis, and functional **3D templates** (templates for enzyme active sites, ligand-binding templates, DNA-binding templates, reverse template comparison vs. structures in wwPDB)

Template-based methods



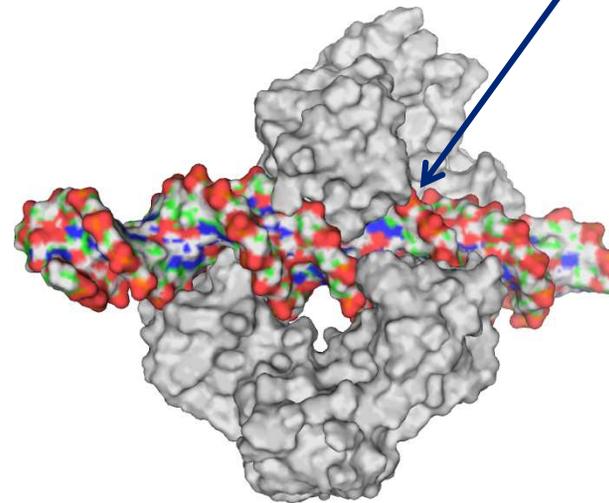
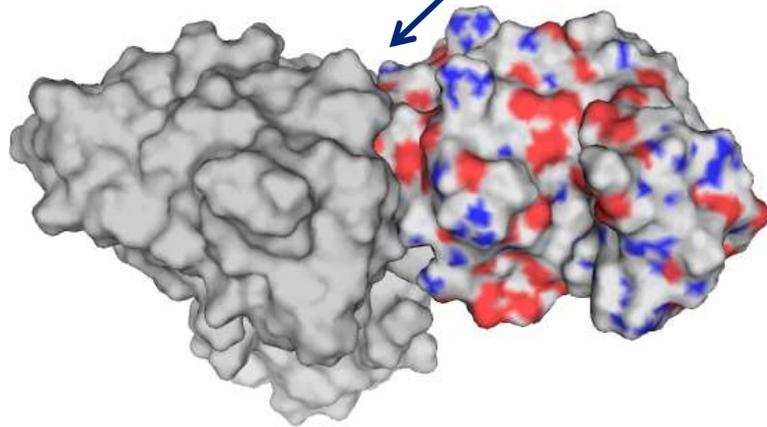
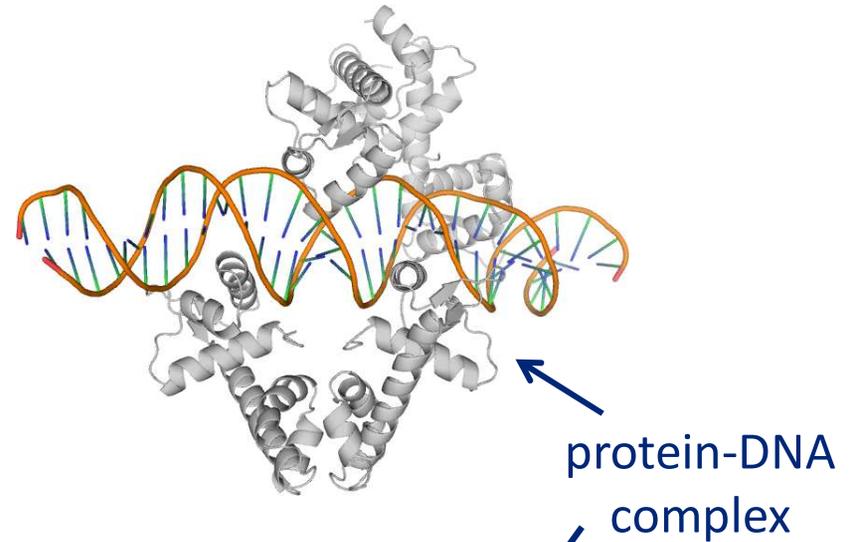
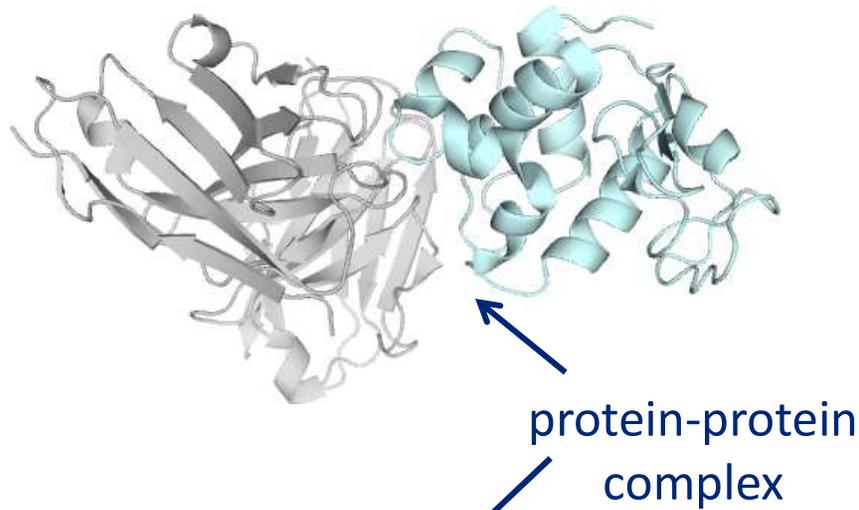
- ❑ Mechanism and Catalytic Site Atlas
 - <https://www.ebi.ac.uk/thornton-srv/m-csa/>
 - provides information about the **active sites**, **catalytic residues** and **mechanisms** in enzymes with experimentally determined 3D structure
 - defines catalytic residues as the residues directly involved in some aspect of the enzymatic reaction
 - provides **3D templates** for catalytic sites in the database

Binding sites for macromolecules



- ❑ typically **protruding loops**, large surface **clefts** but also **flat** binding **sites** – flatter than binding sites for small molecules
 - recognition of a macromolecule involves interactions over a large continuous surface area or several discrete binding regions
 - **difficult to identify** by a simple examination of the protein structure
- ❑ high evolutionary conservation
- ❑ low desolvation energy
- ❑ characteristic physicochemical properties
- ❑ DNA binding sites have characteristic motifs and positive charged electrostatic patches

Binding sites for macromolecules



Binding sites for macromolecules

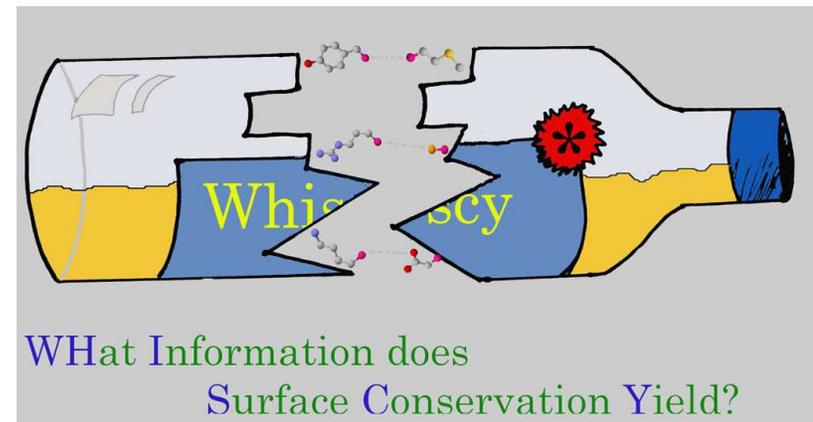


- approaches to identify binding sites
 - evolutionary **conservation**
 - **knowledge-based**

- meta-servers (combine several tools in a workflow)

Evolutionary conservation methods

- ❑ same principles as for binding sites of small molecules
- ❑ WHISCY
 - <https://wenmr.science.uu.nl/whiscy/>
 - predicts protein-protein interface using conservation and structural information (interface propensities for each surface residue are used to adjust the score)



Knowledge-based methods



- ❑ combine multiple **interface features**
 - conservation
 - residue propensity for protein-protein interface
 - physicochemical properties
 - structural properties
- ❑ use known binding sites for parameterization or training → **empirical scoring functions** and **machine learning** methods

Knowledge-based methods



- ❑ cons-PPISP (consensus Protein Protein Interaction Site Predictor)
 - <http://pipe.scs.fsu.edu/ppisp.html>
 - utilizes **machine learning** to predict **protein binding** sites
 - trained on position-specific sequence profiles and solvent accessibilities of each residue and its spatial neighbors

- ❑ Patch Finder Plus
 - <http://pfp.technion.ac.il/>
 - utilizes **machine learning** primarily to find **DNA binding** regions
 - identifies the largest positive electrostatic patch on a protein surface
 - combination of residue frequency, composition and conservation, surface concavity, accessible area and H-bond potential

Meta-servers



- ❑ combine **multiple methods** to improve prediction accuracy

- ❑ meta-PPISP (Protein Protein Interaction Site Predictor)
 - <http://pipe.scs.fsu.edu/meta-ppisp.html>
 - combines cons-PPISP, ProMate and PINUP

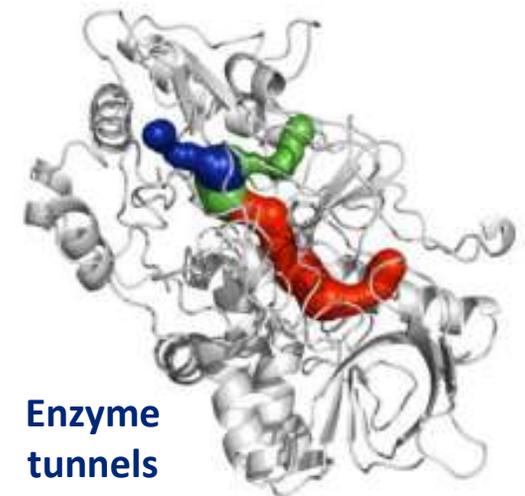
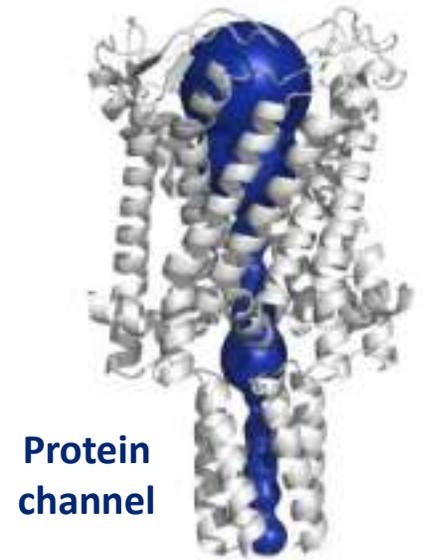
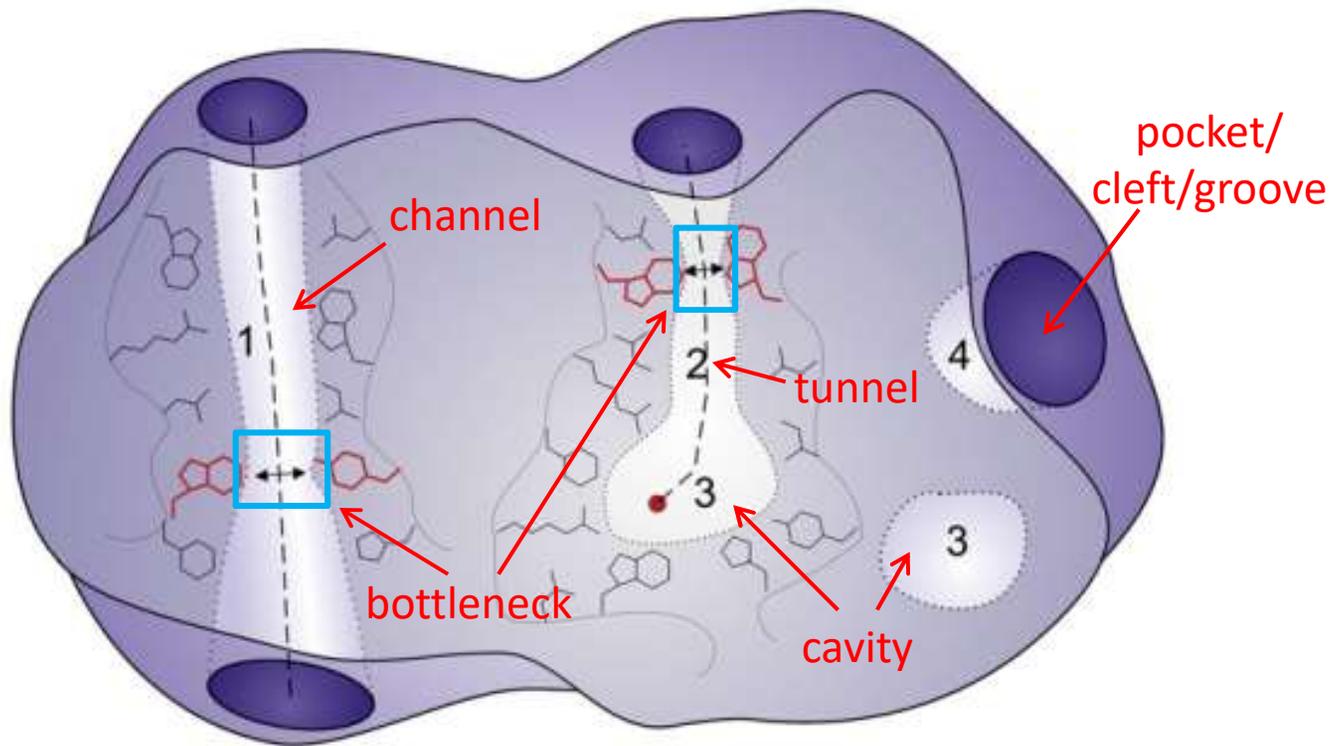
- ❑ PI²PE (Protein Interface/Interior Prediction Engine)
 - <http://pipe.scs.fsu.edu/>
 - pipeline to use five different predictors including cons-PPISP, meta-PPISP and DISPLAR

Transport pathways



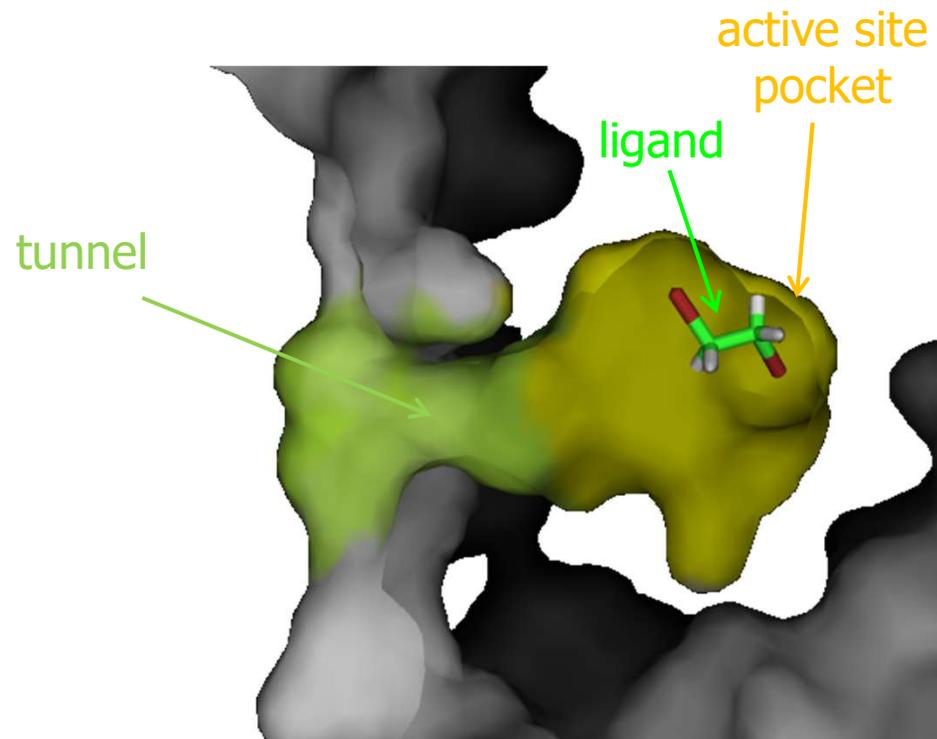
- ❑ mediate transport of ions and small molecules in proteins – an essential role in functioning of large variety of proteins
 - **channels/pores** – transport of substances across membranes
 - **tunnels** – exchange of ligands between the active/binding site and bulk solvent in proteins with buried active/binding site cavities
 - **intramolecular tunnels** – transport of reaction intermediates between two distinct active sites in bifunctional enzymes
- ❑ their **permeability** to different substances depends on the pathway size (radii), shape, length, amino acid composition (physicochemical properties) and dynamics

Transport pathways

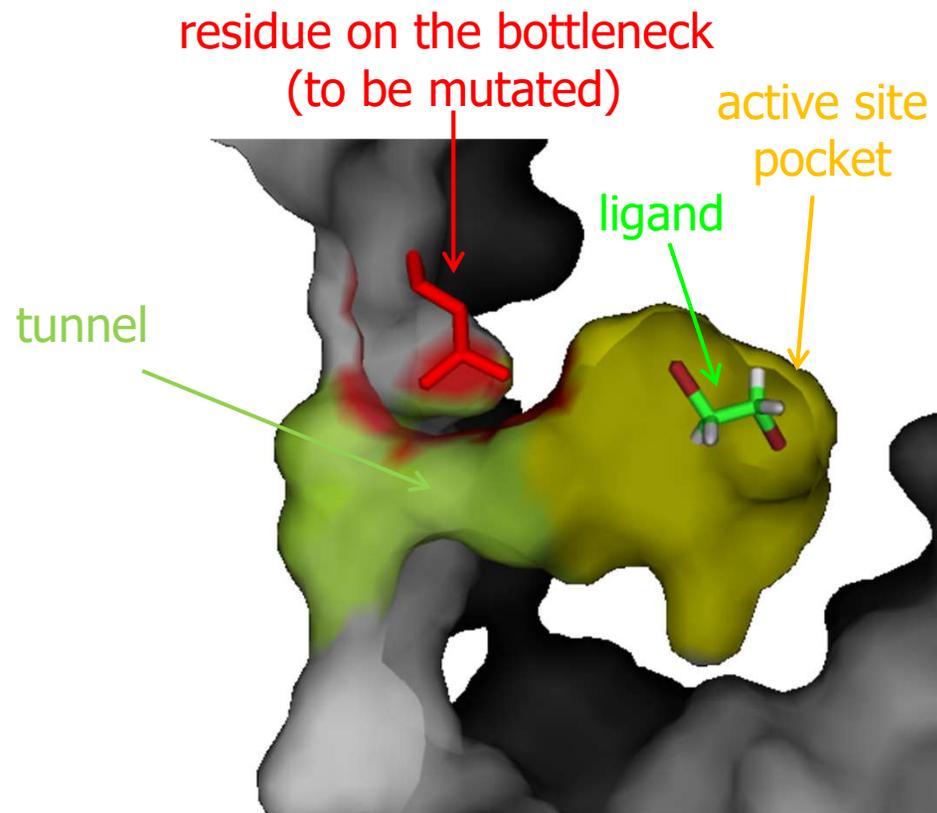


- **bottleneck** – the narrowest part of the tunnel/channel; it has critical importance to the selectivity

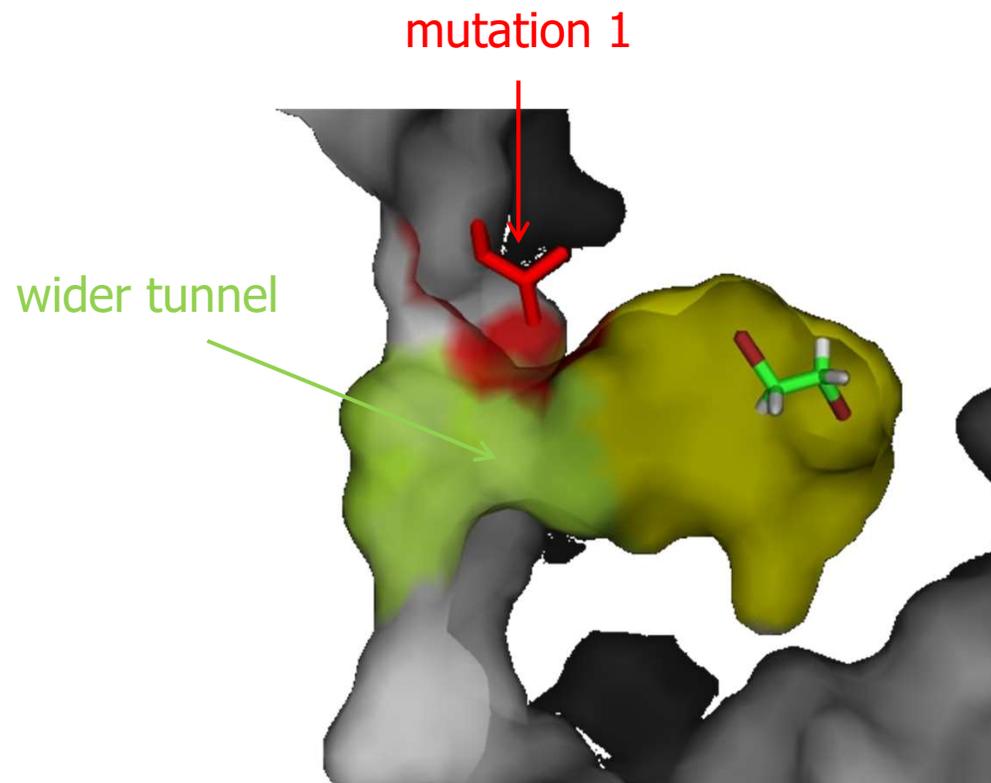
Transport pathways



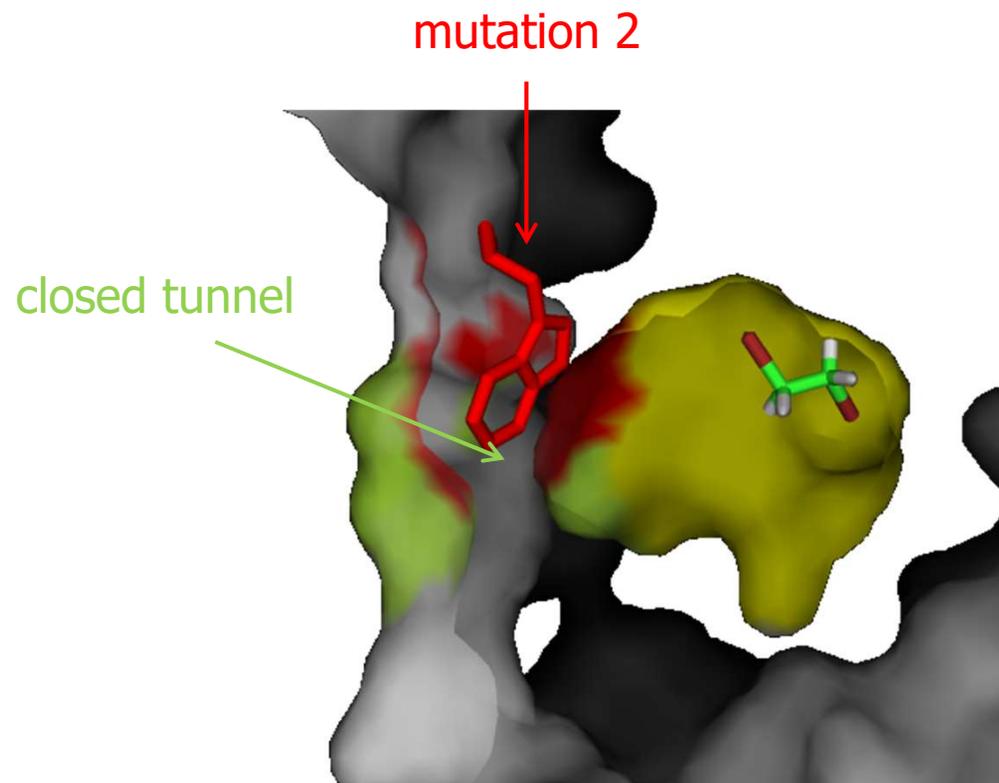
Transport pathways



Transport pathways



Transport pathways



Prediction of transport pathways



- ❑ identification of **overall voids** in proteins
- ❑ identification of **tunnels**
- ❑ identification of **channels**

Identification of overall voids



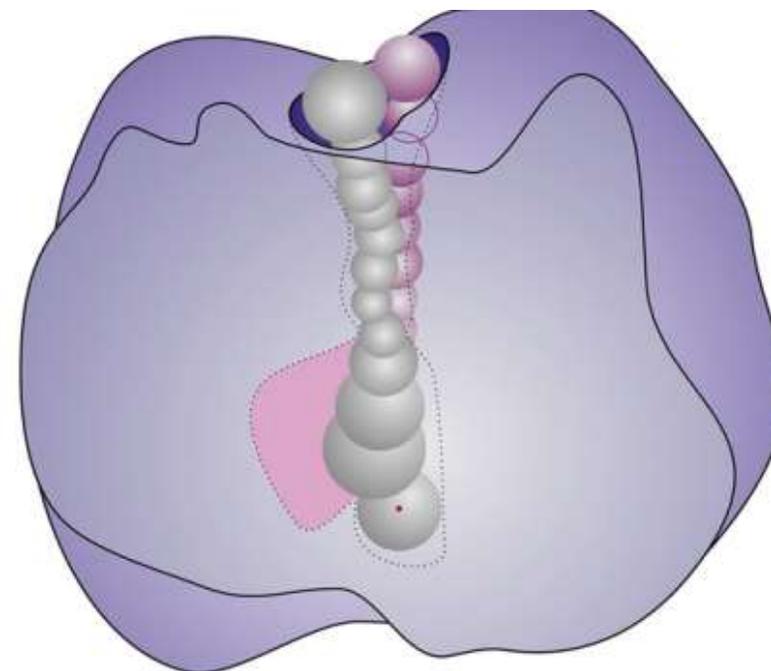
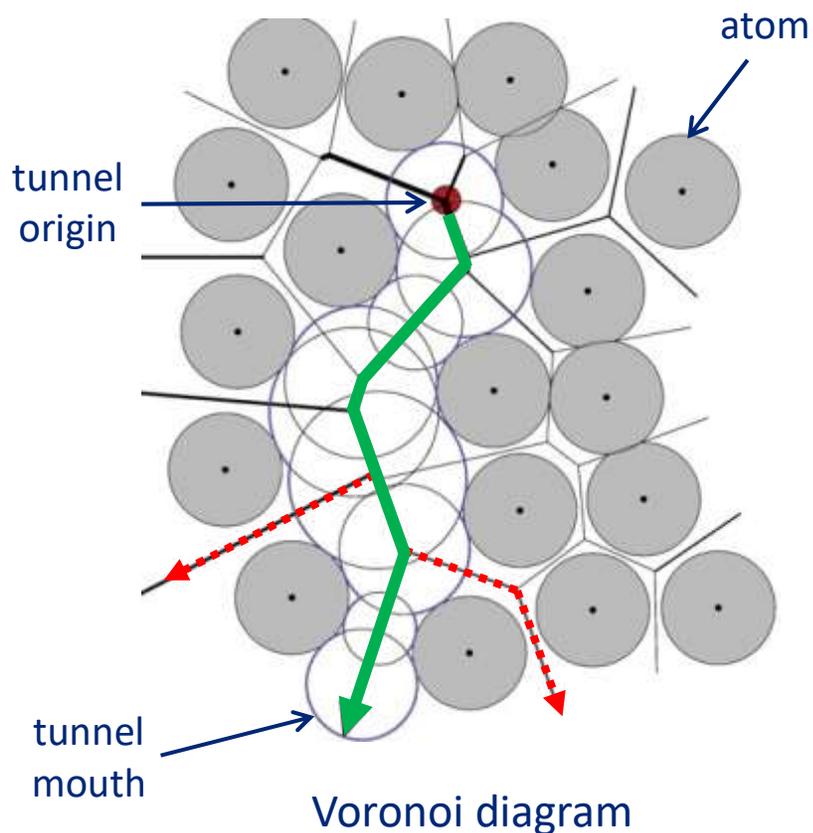
- ❑ methods that aim to accurately represent **all types of voids** in a protein structure, including channels, tunnels, surface clefts, pockets as well as internal cavities
- ❑ usually provide very **limited information** on tunnel and channel characteristics – the identified voids have to be separated from each other
- ❑ **geometry-based** methods for pocket detection
 - HOLLOW – <http://hollow.sourceforge.net/>
 - 3V – <http://3vee.molmovdb.org/>
 - LIGSITE^{csc}, PASS, CASTp, fPocket, SURFNET, POCASA ...

Identification of tunnels



- ❑ methods that calculate **tunnels** connecting occluded cavities with the surrounding bulk solvent
- ❑ identify the pathways **from a cavity** to the **protein surface**
- ❑ **Voronoi diagrams** described by the skeleton of voids between atoms to find all theoretically possible pathways connecting the starting point with the bulk solvent
- ❑ diagram is searched for optimal pathways using Dijkstra's algorithm, based on the criteria defined by a **cost function**
- ❑ the **probe size** defines the lowest radius threshold
- ❑ tunnel geometry is approximated by a sequence of spheres

Identification of tunnels



Common limitation: the tools identify two spherical tunnels instead of one asymmetric tunnel

-  **Probe size:** the minimum radius specified for the tunnel search
-  Allowed pathway according to the selected probe
-  Disallowed pathways

Identification of tunnels - programs



□ CAVER 3.0

- <http://caver.cz/>
- Command-line stand-alone
- PyMOL plugin
- GUI with CAVER Analyst 2

□ CAVER Web

- <http://loschmidt.chemi.muni.cz/caverweb/>
- interactive guide-through web server
- optimized protocol for detection of biologically relevant tunnels
- based on CAVER 3.0 program

Identification of tunnels - programs

The screenshot displays the CAVER software interface for protein tunnel analysis. The main window shows a 3D ribbon representation of a protein structure with two tunnels highlighted in yellow and purple. The interface is divided into several panels:

- Tunnels info:** A table listing identified tunnels.
- Job information:** Details about the current analysis job.
- Visualization settings:** Options for tunnel and structure visualization.
- Tunnel profile:** A graph showing the radius of a tunnel along its length.
- Details for tunnel 1:** Specific parameters for the first tunnel.

id	bottleneck radius [Å]	length [Å]	curvature	throughput
1	1.5	10.8	1.2	0.73
2	1.3	10.1	1.2	0.60

Job ID:	logovv
Title:	Untitled
Structure:	4A4E

Tunnel ID	Radius [Å]	Length [Å]
1	~2.0	0-10.8
2	~1.5	0-10.1

radius [Å]	1.5
distance from starting point [Å]	8.5
coordinates [A, A, Å]	[5.3, 2.6, 11.9]

amino acid	ID	chain
Trp	141	A
Phe	144	A
Ala	145	A
Thr	148	A

Functional sites → transport pathways

Identification of tunnels - programs

- ❑ MolAxis
 - <http://bioinfo3d.cs.tau.ac.il/MolAxis/>
- ❑ MOLE 2.0
 - <http://mole.upol.cz/>
- ❑ ChExVis
 - <http://vgl.serc.iisc.ernet.in/chexvis/>
- ❑ BetaCavityWeb
 - <http://voronoi.hanyang.ac.kr/betacavityweb/>
- ❑ ...

Identification of channels

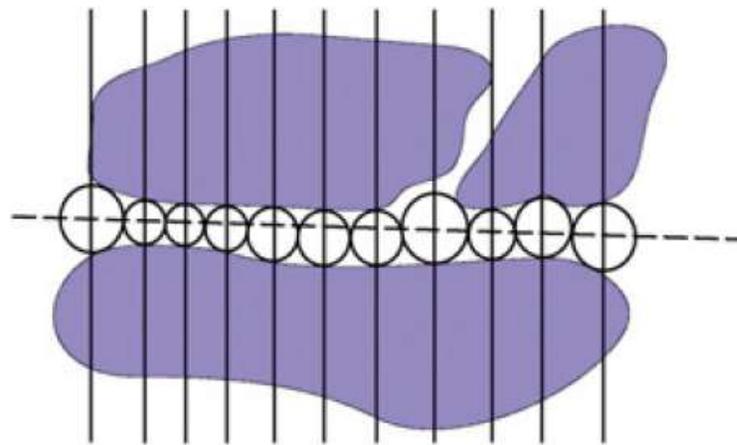


- ❑ methods calculate **channels (pores)** penetrating throughout proteins
- ❑ not convenient for identifying tunnels leading from occluded cavities
- ❑ usually analyze just one channel per structure
- ❑ usually need information about approximate position and direction of the channel (**channel axis**) – user-provided or automatically identified

Identification of channels - programs

□ POREWALKER

- <http://www.ebi.ac.uk/thornton-srv/software/PoreWalker/>
- identifies **channel axis** by heuristic iterative approach (based on the axes of transmembrane secondary structures)
- protein is divided into equally-spaced slices perpendicular to the axis; the largest spheres fitting the channel are identified

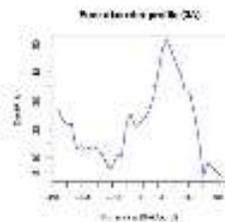


Identification of channels - programs

□ POREWALKER

Pore analysis results

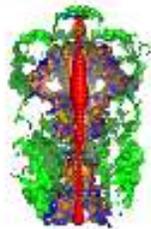
Overview of the available results:



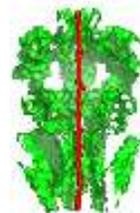
Pore shape



Slideshow of the pore cross-sections



Pore visualization



Features of the cavity

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