

LOSCHMIDT  
LABORATORIES



## Analysis of protein structures

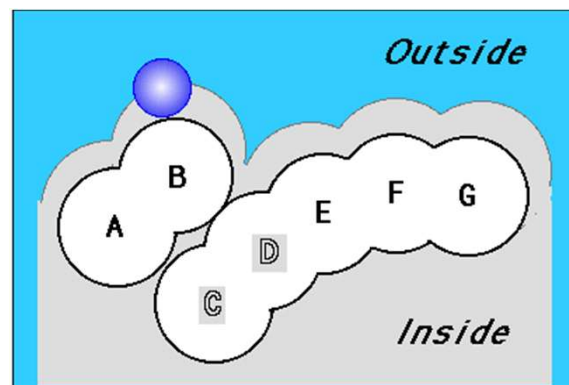
# Outline

- ❑ Residue solvent accessibility
- ❑ Protein solubility
- ❑ Molecular interactions
- ❑ Functional sites
  - Binding sites
  - Transport pathways

# Residue solvent accessibility



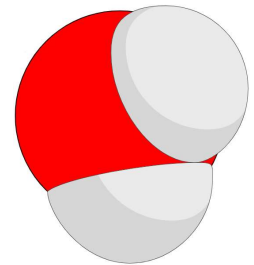
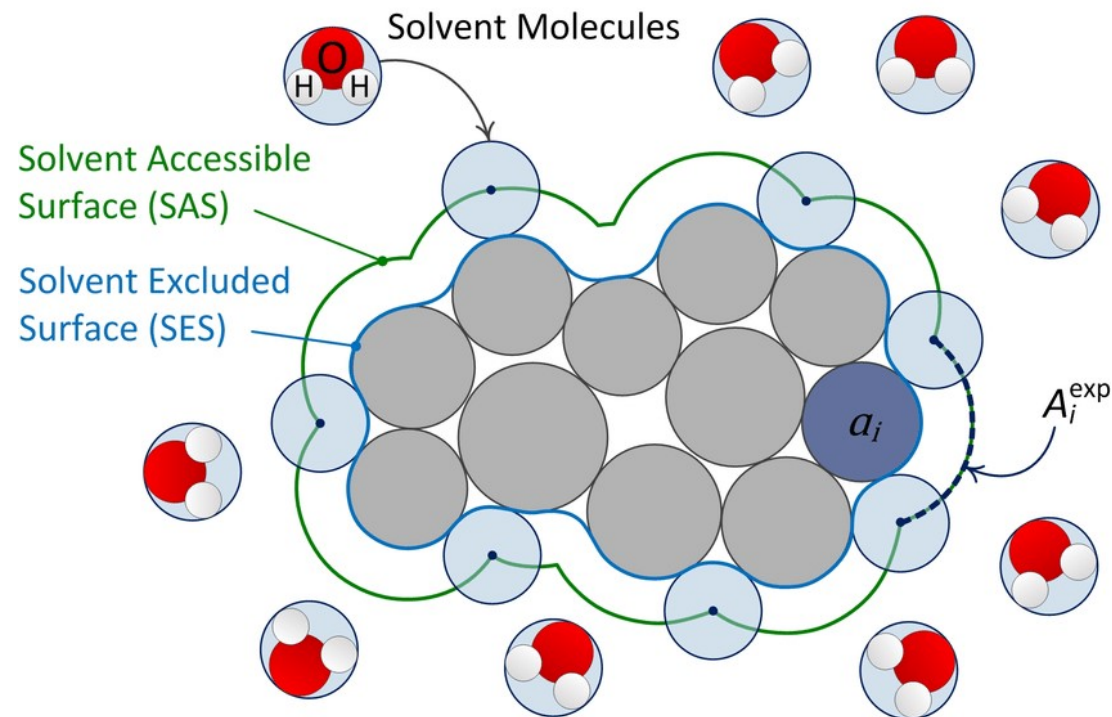
- ❑ Solvent **accessible surface area** (ASA or SASA, in  $\text{\AA}^2$ ):  
the extent to which a residue in a protein structure is accessible to the solvent
- ❑ Typically calculated by **rolling** a spherical **probe** of a particular radius over a protein surface and summing the area that can be accessed by this probe on each residue



# Residue solvent accessibility



- ❑ Solvent **accessible surface area** (ASA, SASA or SAS, in  $\text{\AA}^2$ )
- ❑ Solvent excluded surface (SES) – also known as molecular surface, or Connolly surface area



Water radius  $\approx 1.4 \text{ \AA}$

# Residue solvent accessibility



- ❑ Relative accessible surface area (rASA)
  - Ratio of the actual accessible area of a given residue
$$\text{rASA} = \text{ASA} / \text{ASA}_{\text{MAX}}$$
  - Enables **comparison** of accessibility of **different amino acids** (e.g., long extended vs. spherical amino acids)
- ❑ Simplified two state description
  - **Buried** vs. **exposed** residues
  - *threshold* for differentiating buried vs. surface residues is not well defined (usually 15–25 %)
  - $\text{rASA} < \text{threshold} \Rightarrow \text{buried}$ ;  $\text{rASA} \geq \text{threshold} \Rightarrow \text{exposed}$

# Residue solvent accessibility – programs

## ❑ Naccess

- <http://www.bioinf.manchester.ac.uk/naccess/>
- Calculates atomic and residue ASAs and rASA for PDB files
- Adjustable settings (e.g., probe size, atomic radii,...)

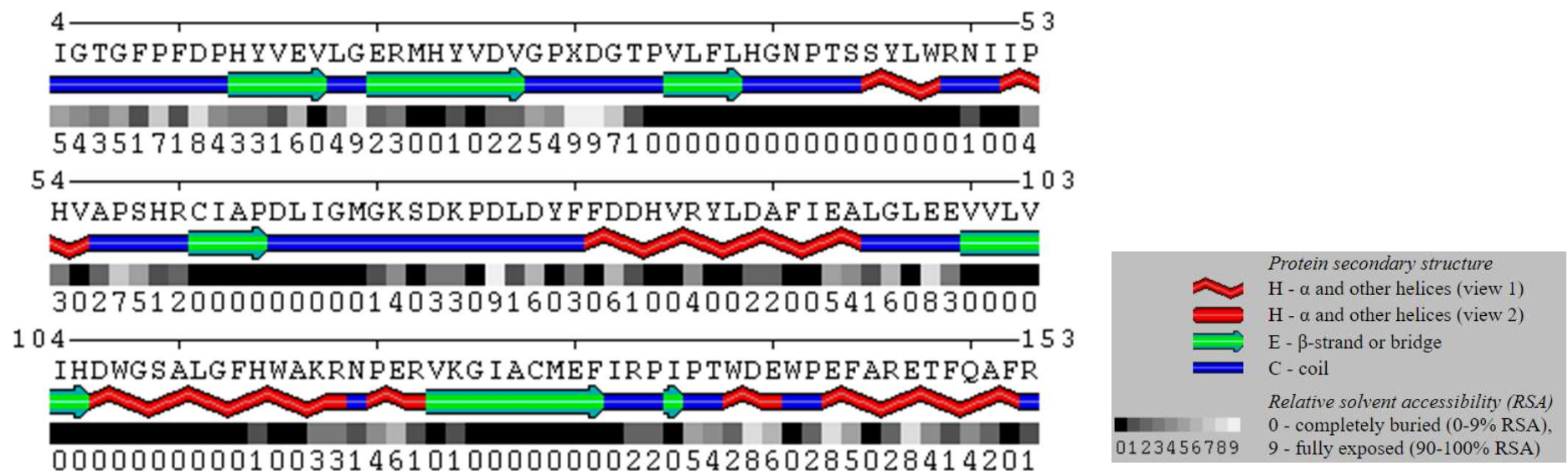
## ❑ DSSP

- <https://swift.cmbi.umcn.nl/gv/dssp/>
- Assignment of secondary structures and calculation of ASAs of residues

# Residue solvent accessibility – programs

## □ POLYVIEW-2D

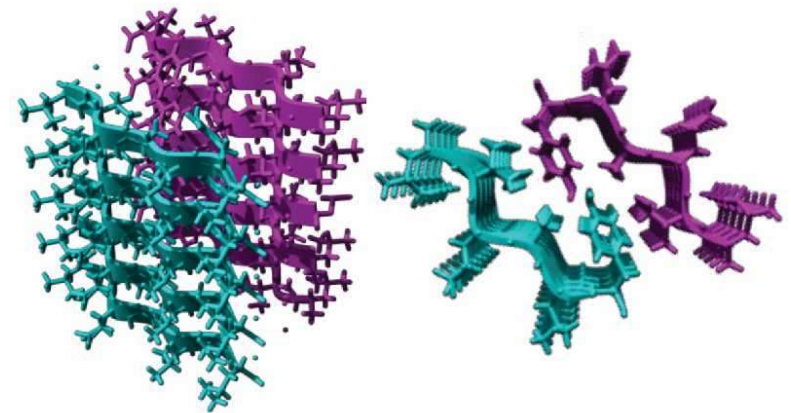
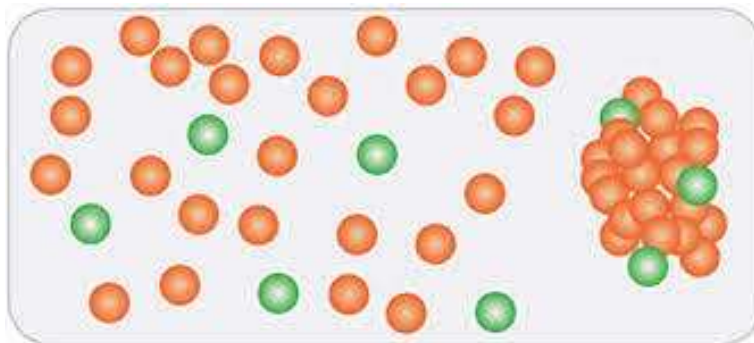
- <https://polyview.cchmc.org/>
- Visualization tool for structural and functional annotations of proteins, including solvent accessibility
- Residue SASA calculated by **DSSP** and transformed to **rASA**



# Protein solubility



- ❑ Concentration of protein in saturated solution that is in equilibrium with solid phase
- ❑ For proteins expressed in the lab, it depends on
  - ❑ Hydrophilic/hydrophobic balance of the solvent-exposed residues
  - ❑ Aggregation-prone regions (APRs) – mainly hydrophobic residues prone to form beta-structures
  - ❑ Protein expressibility in the cells



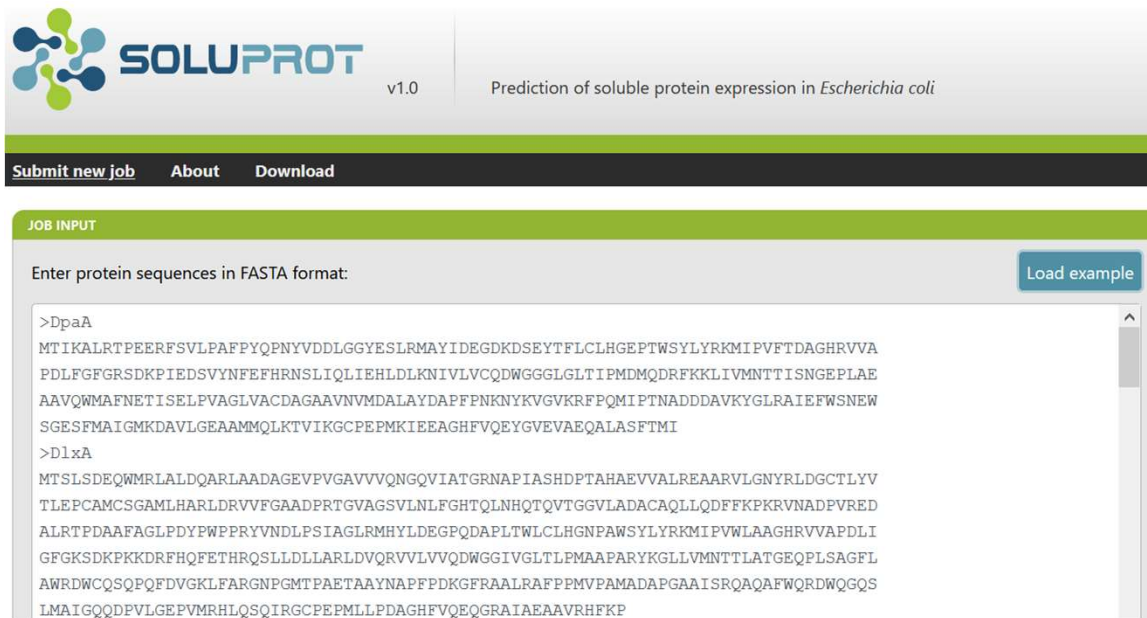
Cross-beta spines of amyloid fibrils



# Protein solubility

## ❑ SoluProt

- <https://loschmidt.chemi.muni.cz/soluprot/>
- Soluble expression of protein sequences in *E.coli*
- Based on machine learning



**SOLUPROT** v1.0 Prediction of soluble protein expression in *Escherichia coli*

Submit new job About Download

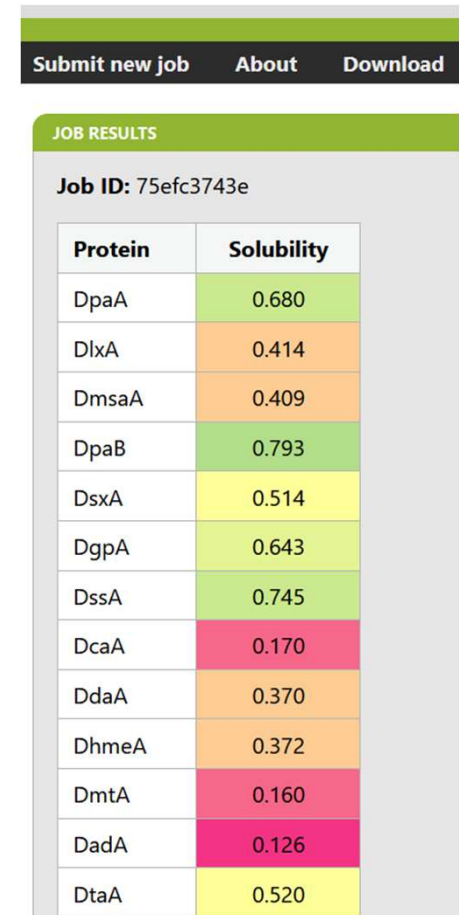
**JOB INPUT**

Enter protein sequences in FASTA format:

Load example

```
>DpaA
MTIKALRTPEERFSVLPAFFYPQPNYVDDLGGYESLRMAYIDEGDKDSEYTFCLHGEPTWSYLYRKMI PVFTDAGHRVVA
PDLFGFGRSDKPIEDSVYNFEFHRNSLIQLIEHLDLKNIVLVCQDWGGGLGLTIPMDMQDRFKKLIVMNTTISNGEPLAE
AAVQWMAFNETISELPVAGLVACDAGAAVNVM DALAYDAFFPNKNYKVGVRFPQMIPTNADDDAVKYGLRAIEFWSNEW
SGESFMAIGMKDAVLGEAAMMQLKTVIKGCPEPMKIEEAGHFVQEGVEVAEQALASFTMI
>DlxA
MTSLSDEQWMLRALDQARLAADAGEVPVGA VVVQNGQVIATGRNAPIASHDPTAHAEVVALREAA RVLGNYRLDGCTLYV
TLEPCAMCSGAMLHARLD R VVFGAADPRTGVAGSVLNLFGHTQLNHQTQVTGGVLADACAQLLQDFFKPKRVNADPVRED
ALRTPDAAFAGLPDYFPWPPRYVNDLPSIAGLRMHYLDGEPQDAPLTWLC LHGNPAWSYLYRKMI PVWLAAGHRVVAPDLI
GFGKSDKPKKDRFHQFETHRQSLDLLLARLDVQRVVLVVQDWGGIVGLTLPMAAPARYKGLLVMTT LATGEQPLSAGFL
AWRDCWCSQPQFDVVGKLFARGNPGMTPAETAAYNAPFPDKGFRAALRAFFPMVPAMADAPGAAISRQAQAFWQRDWGQGS
LMAIGQQDPVLGEFVMRHLQSQIRGCPEPMLLPDAGHFVQEQQGRAIAEAAVRHFKP
```

Input



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**JOB RESULTS**

Job ID: 75efc3743e

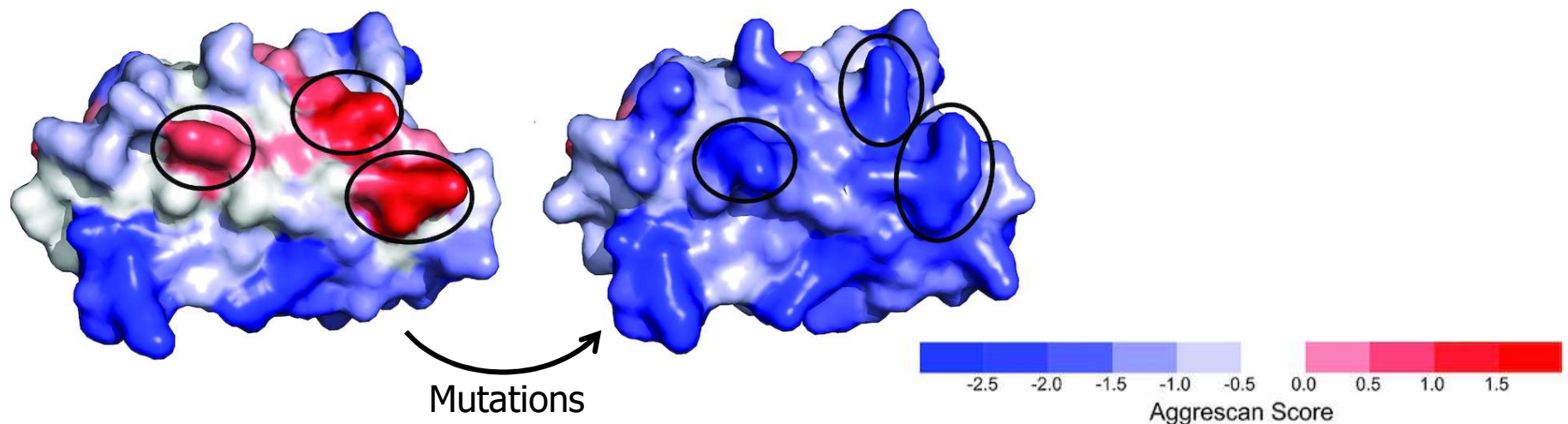
Protein	Solubility
DpaA	0.680
DlxA	0.414
DmsaA	0.409
DpaB	0.793
DsxA	0.514
DgpA	0.643
DssA	0.745
DcaA	0.170
DdaA	0.370
DhmeA	0.372
DmtA	0.160
DadA	0.126
DtaA	0.520

Output

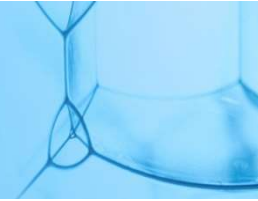
# Protein solubility

## ❑ Aggrescan3D

- <http://biocomp.chem.uw.edu.pl/A3D2/>
- Predicts the aggregation propensities by identifying APRs
- Can introduce mutations and predict impact on stability and aggregation-propensity
- Can account for protein flexibility (“dynamic mode”)



# Molecular interactions



- ❑ Assessment of the interactions within a protein structure and between proteins in an assembly is **essential** to understand the **molecular basis of stability and function** of proteins and their complexes

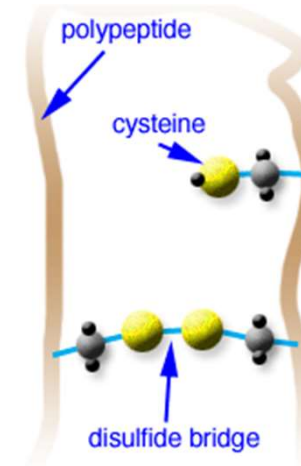
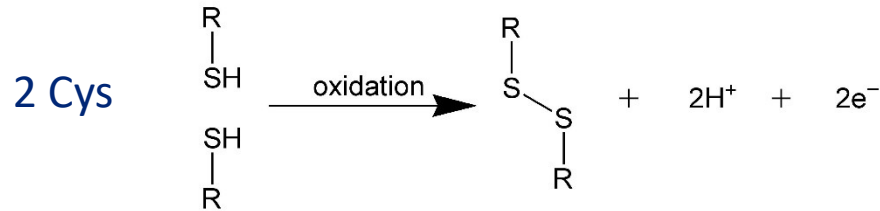
# Types of interactions



- ❑ charge-charge (ionic) interactions
  - present in charged residues; e.g. salt bridges
- ❑ hydrogen bonds (H-bonds)
  - donor and acceptor atoms sharing hydrogen
- ❑ aromatic ( $\pi$ - $\pi$ ) interactions
  - attractive interaction between aromatic rings
- ❑ van der Waals (vdW) interactions
  - between any two atoms
- ❑ hydrophobic interactions
  - entropic origin; important for hydrophobic residues

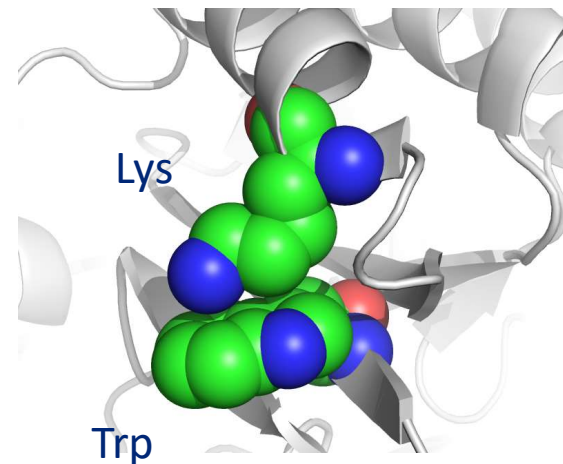
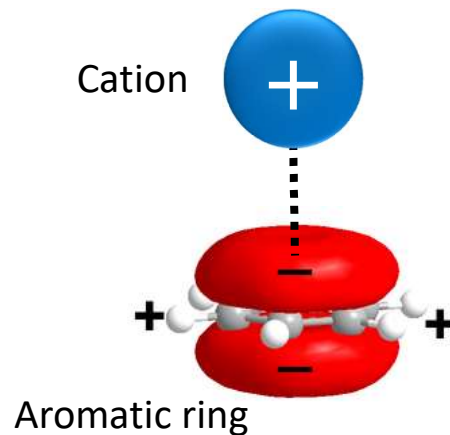
# Types of interactions

## □ disulfide bonds (cysteine bridges)



## □ cation- $\pi$ interactions

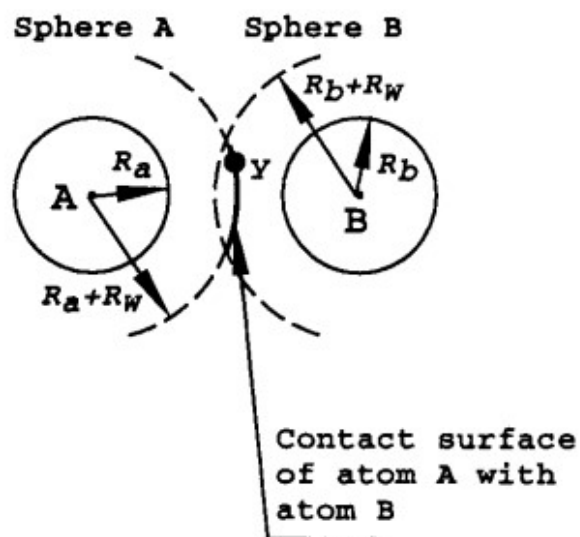
- electrostatic interaction of a positively charged residue (Lys or Arg) with an aromatic residue (Phe, Trp, or Tyr)



# Molecular interactions – how to identify?



- criteria for recognizing various types of interactions
  - geometric rules (distances, angles)
  - atom types
  - energetics (physicochemical rules)
  - contact surface area between atoms

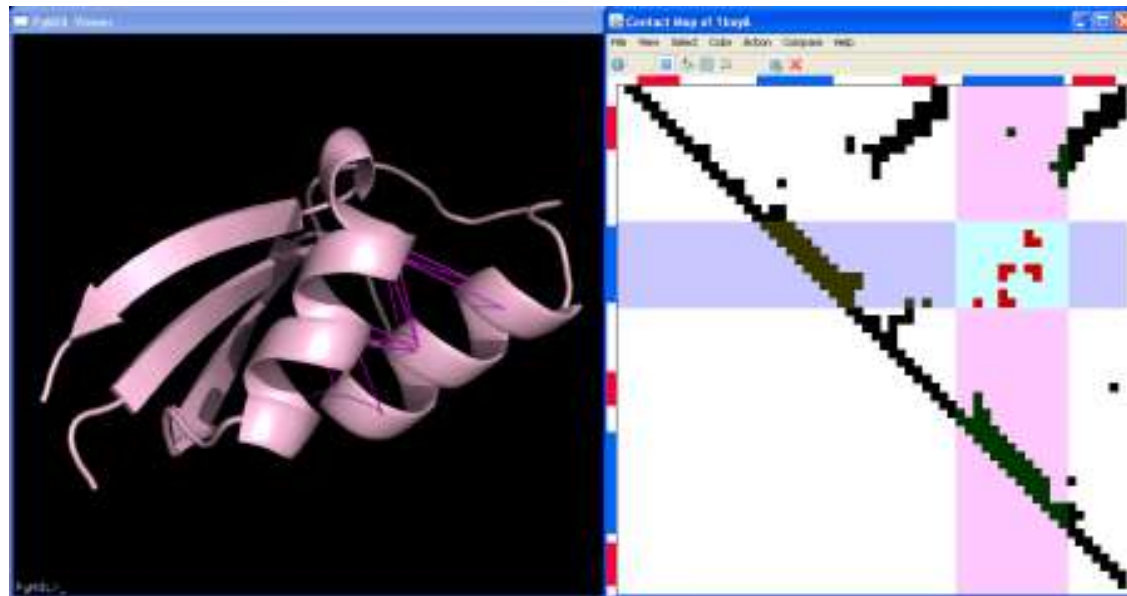


# Molecular interactions – programs

## ❑ CMView

❑ <https://www.bioinformatics.org/cmview/>

- represents **residue-residue contacts** within a protein or between proteins in a complex in the form of a **contact map**
- 3D visualization using PyMol



# Molecular interactions – programs



- ❑ PIC (**P**rotein **I**nteraction **C**alculator)
  - <http://pic.mbu.iisc.ernet.in/>
  - identifies **various interactions** – disulfide bonds, hydrophobic interactions, ionic (charged-charged) interactions, hydrogen bonds, aromatic–aromatic, aromatic–sulfur and cation– $\pi$  interactions within a protein or between proteins in a complex
  - uses standard criteria (atom types and geometry)



# Molecular interactions – programs

## ❑ PIC (Protein Interaction Calculator)

- <http://pic.mbu.iisc.ernet.in/>

**INTRAPROTEIN INTERACTIONS**

\* This option can accept a monomeric or a multichain protein file.

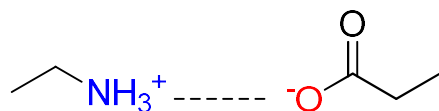
Upload a file in PDB format:  No file chosen

- ☒ **Hydrophobic Interactions** Enter the interaction cut-off value  (Default 5A)
- ☒ **Disulphide Bridges**
- ☒ **Main Chain-Main Chain Hydrogen Bonds**
- ☒ **Main Chain-Side Chain Hydrogen Bonds**
- ☒ **Side Chain-Side Chain Hydrogen Bonds**
- ☒ **Ionic Interactions** Enter the interaction cut-off value  (Default 6A)
- ☒ **Aromatic-Aromatic Interaction** : cut-off value  to  (Default 4.5A to 7A)
- ☒ **Aromatic-Sulphur Interactions** Enter the interaction cut-off value  (Default 5.3A)
- ☒ **Cation-Pi Interactions** Enter the interaction cut-off value  (Default 6A)

To E-mail the results enter a valid e-mail address:  (Optional)

# Molecular interactions – programs

- ESBRI (Evaluating the Salt BRIdges in Proteins )
  - <http://bioinformatica.isa.cnr.it/ESBRI/introduction.html>
  - analysis of salt bridges interactions (ionic interaction + H-bond)
  - checks if at least one Asp or Glu side-chain carboxyl oxygen atom and one side-chain nitrogen atom of Arg, Lys or His are within a distance  $\leq 4.0 \text{ \AA}$



Salt bridge

Residue 1	Residue 2	Distance
NZ ALYS A 11	OD1 ASP A 62	3.86
NZ ALYS A 11	OD2 ASP A 62	2.78
NZ ALYS A 11	OD2 ASP A 68	2.85
NZ BLYS A 11	OD1 ASP A 62	3.79
NZ BLYS A 11	OD2 ASP A 62	2.74
NZ BLYS A 11	OD2 ASP A 68	2.75
NH1 ARG A 46	OE1 GLU A 276	3.61

# Functional sites



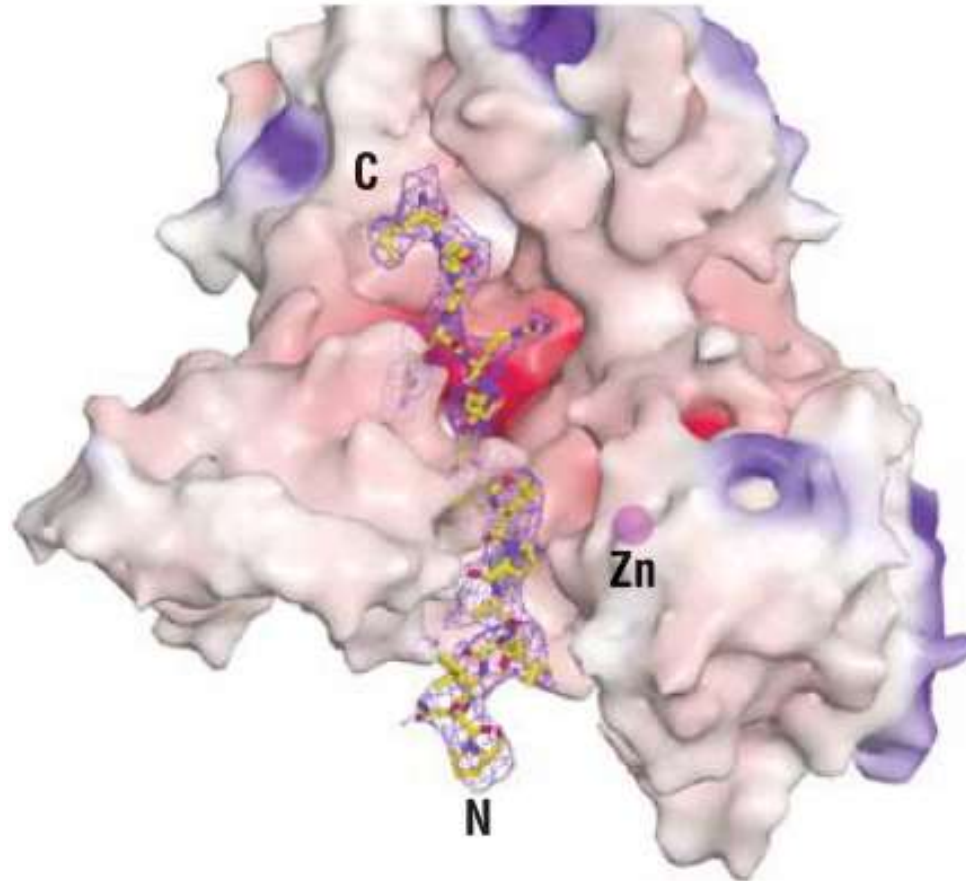
- ❑ binding sites
  - binding sites for small molecules
  - binding sites for macromolecules
- ❑ transport pathways
  - overall void
  - tunnels
  - channels

# Binding sites



- ❑ sites on the protein that provides the complementarity for the bound molecule (ligand)
  - **binding site** – its function is molecular recognition
  - **active/catalytic site** – special case of the binding site – its function is to promote chemical catalysis (break/formation of covalent bonds)
- ❑ binding involves the formation of **non-covalent interactions** between the protein and the bound molecule
- ❑ bound molecule – **small molecule** or **macromolecule**
- ❑ binding is usually **very specific** – complementarity in shape and charge distribution between the site and bound molecule

# Binding sites



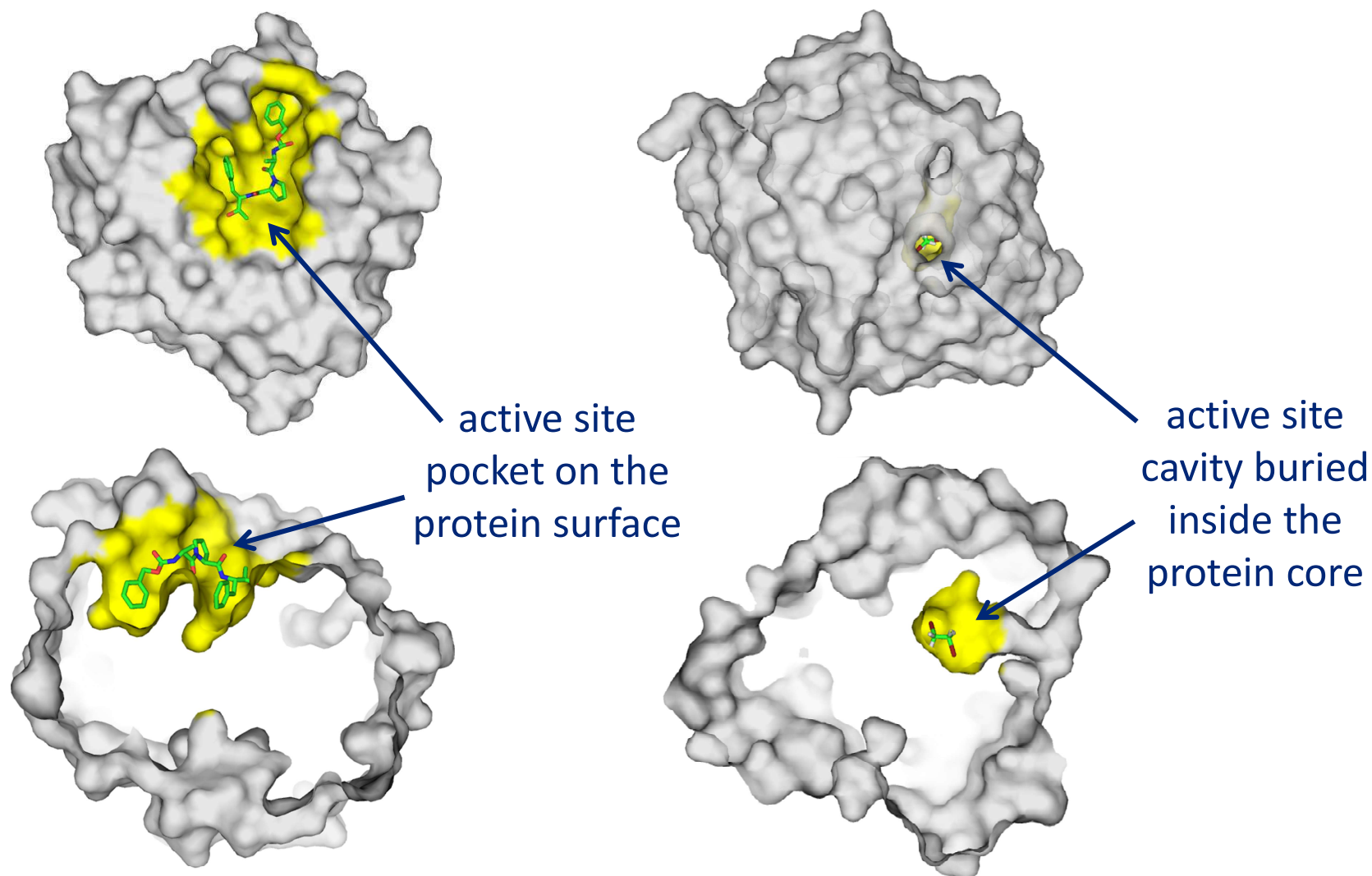
Complementarity in shape and charge distribution  
between the active site and substrate

# Binding sites for small molecules



- ❑ typically internal **cavities**, or surface **pockets** or **clefts**
  - concave regions
  - provide **microenvironment** different from that of the bulk solvent (e.g., many residues with negative charge → very strong electrostatic field enabling binding of highly charged ligands)
  - often identifiable by a simple examination of the protein structure
- ❑ highly conserved by evolution
- ❑ low desolvation energy
- ❑ characteristic physicochemical properties

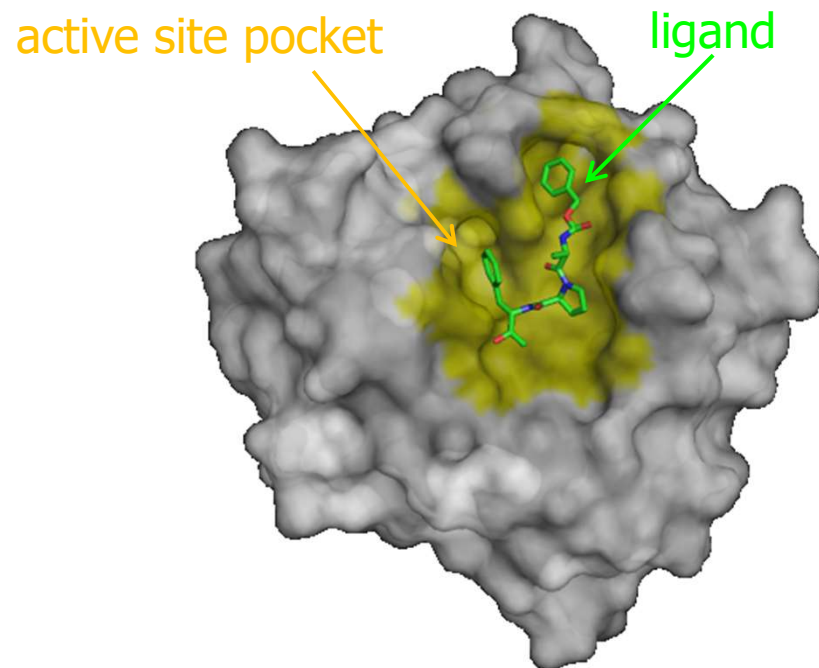
# Binding sites for small molecules





# Binding sites for small molecules

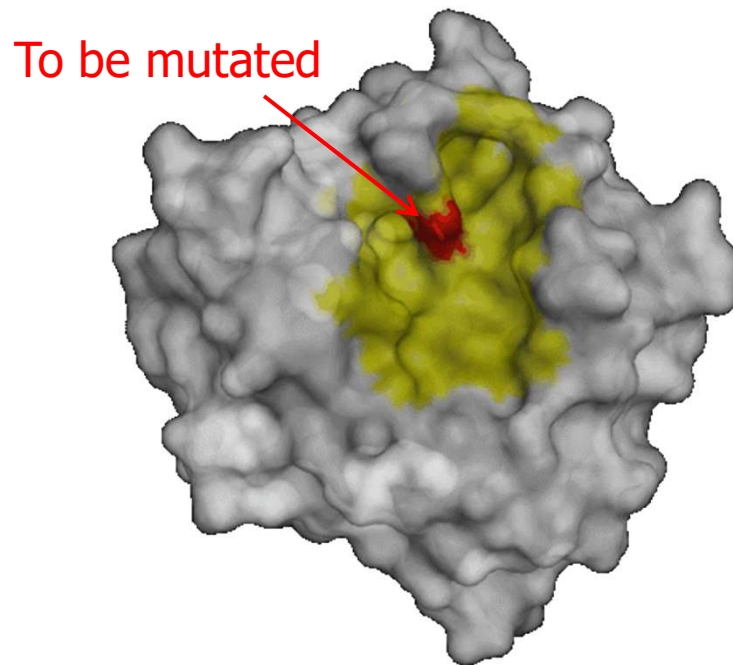
- can be very ligand-specific





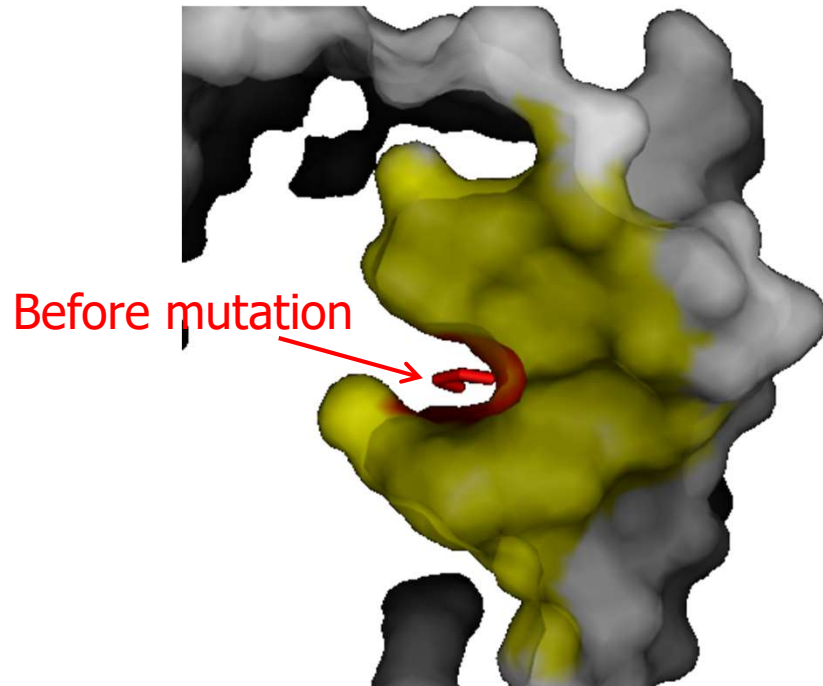
# Binding sites for small molecules

- can be very ligand-specific



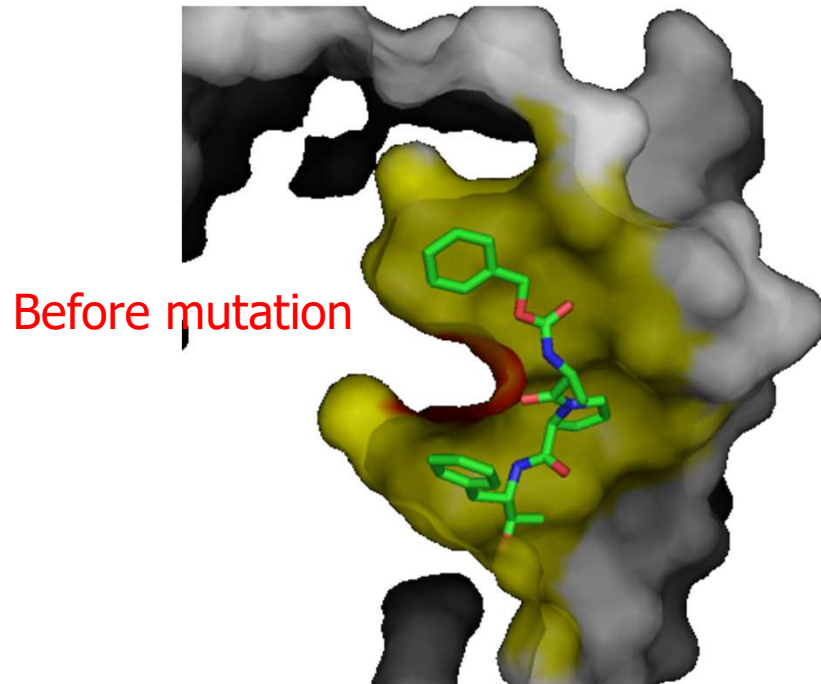
# Binding sites for small molecules

- can be very ligand-specific



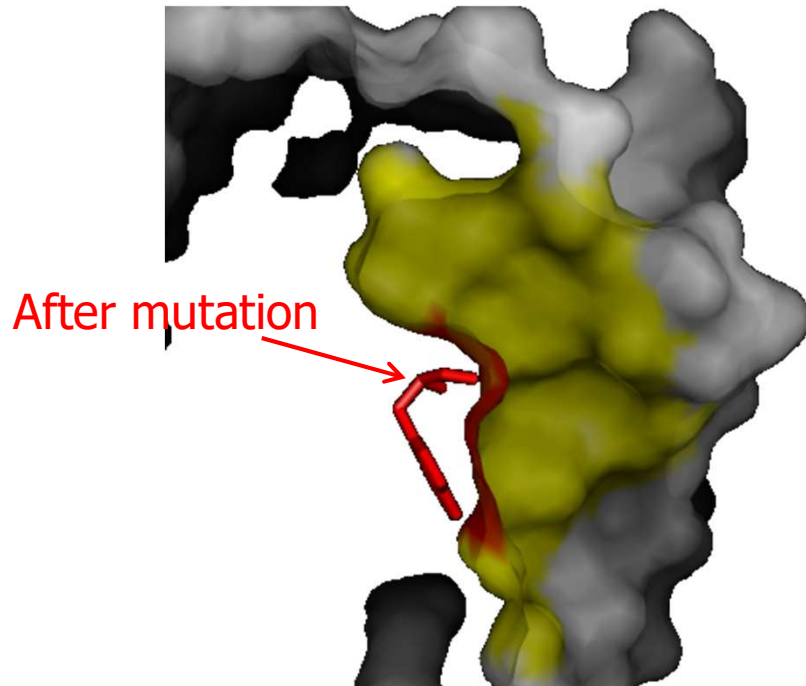
# Binding sites for small molecules

- can be very ligand-specific



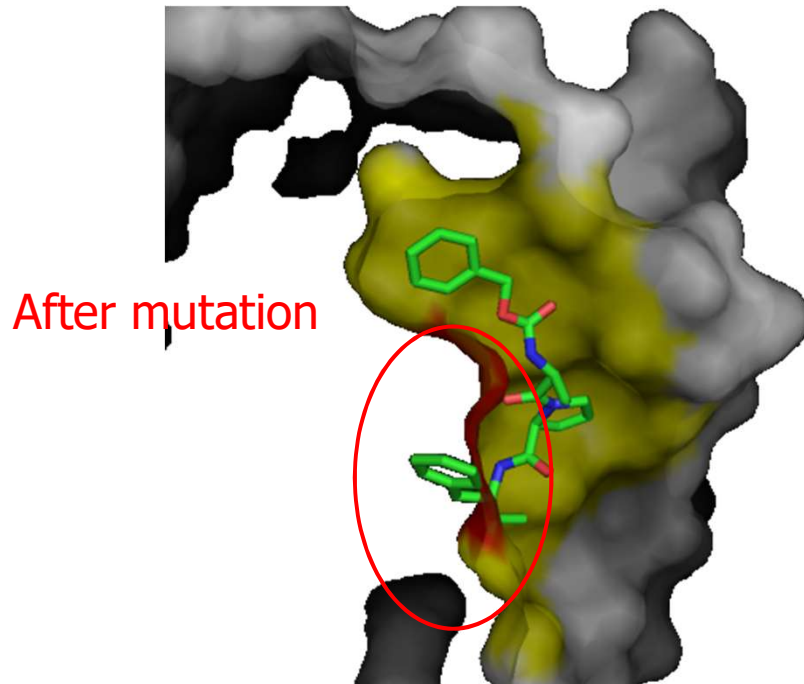
# Binding sites for small molecules

- can be very ligand-specific



# Binding sites for small molecules

- can be very ligand-specific



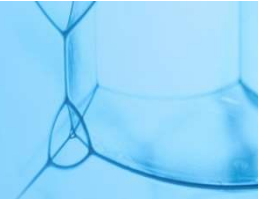
No longer a good fit!

# Binding sites for small molecules



- ❑ approaches to identify binding sites:
  - ❑ evolutionary **conservation**
  - ❑ **“pocket”** detection
    - geometry based methods
    - energy based methods
  - ❑ binding site **similarity**
    - template-based methods
    - microenvironment-based methods

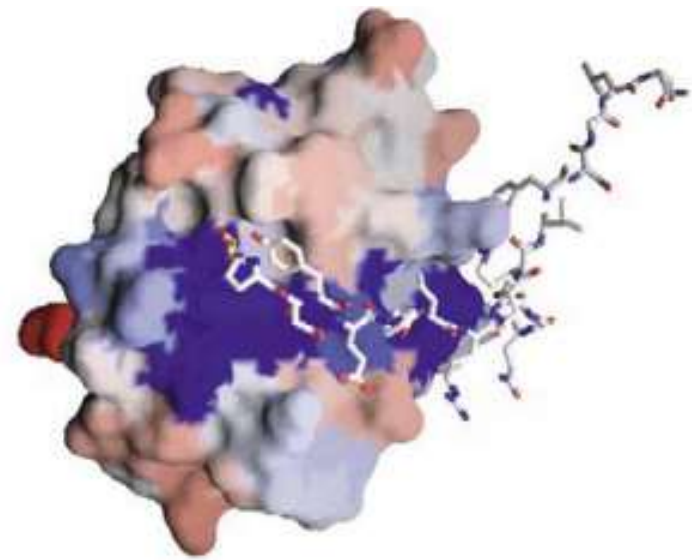
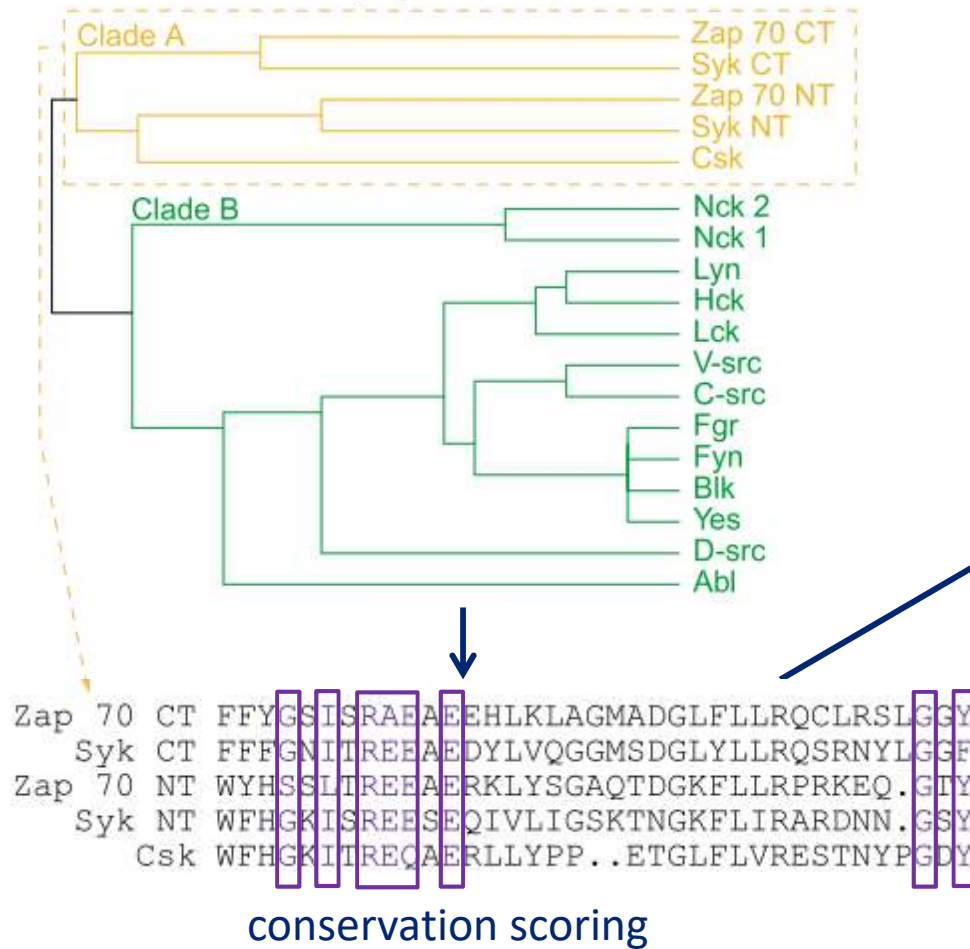
# Evolutionary conservation



- ❑ residues important for **protein function** or **stability** tend to be **highly conserved** over evolution
- ❑ residue conservation in a set of related proteins can be derived from a **multiple sequence alignment**
- ❑ **mapping of conservation** on structure can reveal patches of conserved **surface** residues – potential binding sites
- ❑ protein interior usually more conserved than surface – not suitable for prediction of buried cavities
- ❑ **not very specific** – better to combine with other features

# Evolutionary conservation

phylogenetic analysis



map evolutionary data on  
the structure



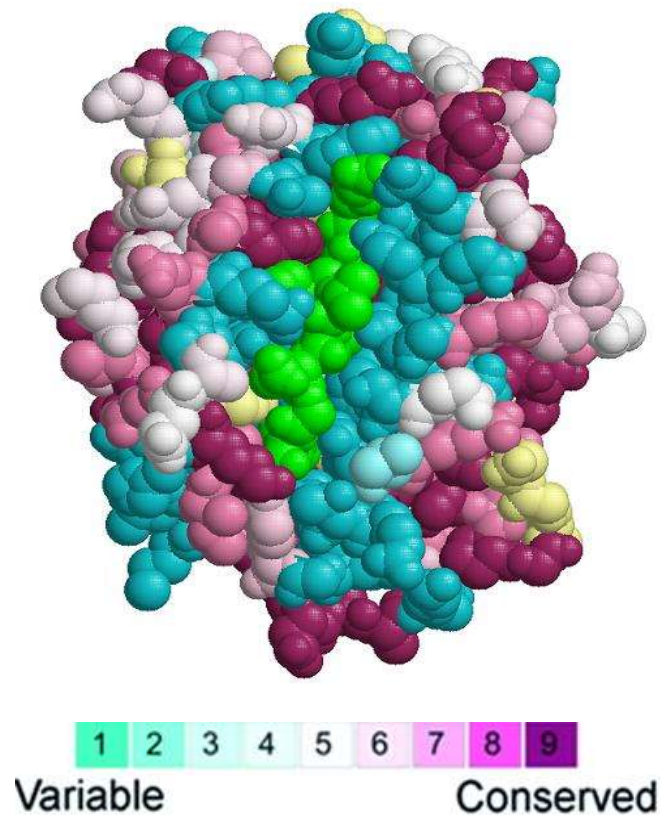
# Evolutionary conservation

## ❑ ConSurf

- <http://consurf.tau.ac.il/>
- estimates the level of **evolutionary conservation** of individual positions in protein and maps this information onto its 3D structure
- conservation score is derived based on the site-specific **evolutionary rates** calculated for each position by Rate4Site program
- **ConSurfDB** – pre-calculated conservation scores for all structures from wwPDB

# Evolutionary conservation

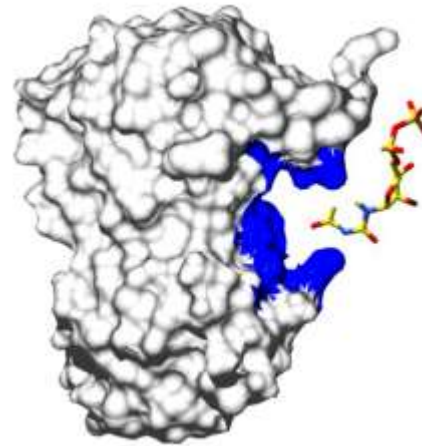
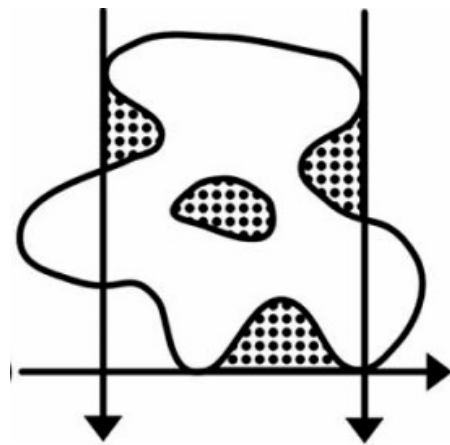
## □ ConSurf



# “Pocket” detection



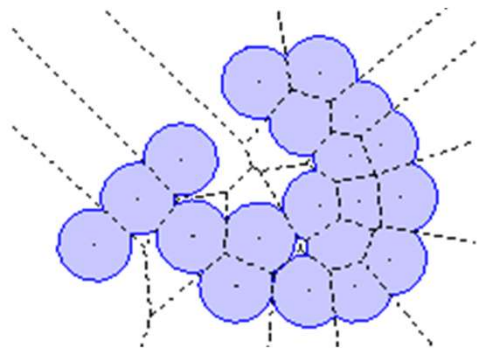
- analyze the protein surface for pockets (clefts, cavities)



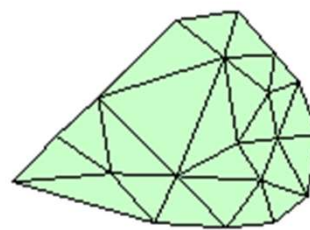
- **geometry-based** methods
  - define favorable cleft regions based on sterical assessments
- **energy-based** methods
  - define favorable cleft regions based on energetic evaluations

# Geometry-based methods

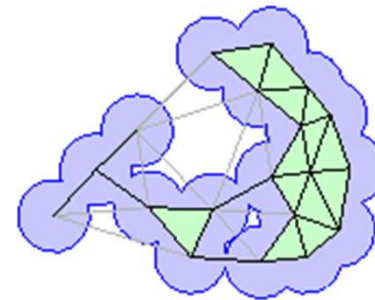
- ❑ Computed Atlas of Surface Topography of proteins (CASTp)
  - <http://sts.bioe.uic.edu/castp>
  - uses **computational geometry methods** including Delaunay triangulation, alpha shape and discrete flow theory
  - **measures the volume** and **surface area** of each pocket and cavity using the ASA model and molecular surface (Connolly) model



Voronoi  
diagram



Delaunay  
triangulation



alpha shape

# Geometry-based methods



## ❑ SURFNET

- <http://www.ebi.ac.uk/thornton-srv/software/SURFNET/>
- probe sphere filling method

## ❑ fPocket

- <http://fpocket.sourceforge.net/>
- uses alpha shape and Voronoi tessellation
- mdpocket – version for analysis of pocket dynamics

## ❑ POCASA (POcket-CAvity Search Application)

- [http://altair.sci.hokudai.ac.jp/g6/Research/POCASA\\_e.html](http://altair.sci.hokudai.ac.jp/g6/Research/POCASA_e.html)
- combination of the grid system with probe sphere filling method

## ❑ ....

# Energy-based methods



- ❑ pockets are defined by energetic criteria
- ❑ evaluate the **interaction energy** between the **protein** and a **molecular fragment – probe** (e.g., a methyl, hydroxyl, or amine groups) to locate energetically favorable binding sites
- ❑ recently combined with other methods to assess the *ligandability* (ability of a cavity to bind ligands)

**Note:** 'druggability' is usually referred to the likelihood of finding orally bioavailable small molecules that bind to a particular target in a disease-modifying way. Ligandability is a necessary but not sufficient condition for druggability.

# Energy-based methods

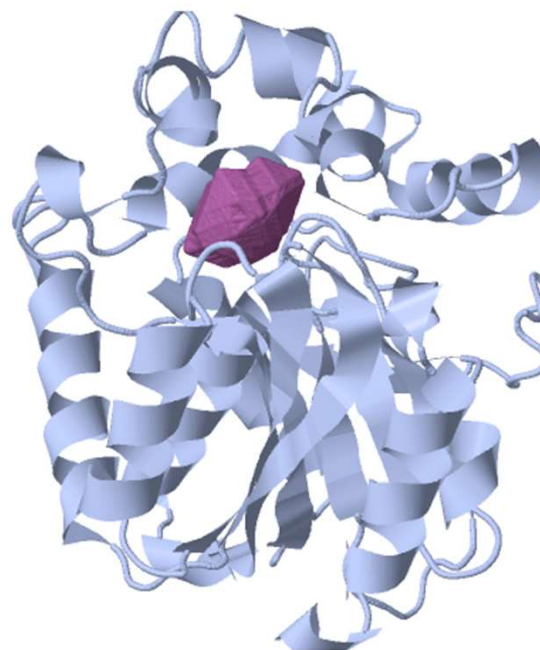


## ❑ Cavity Plus

- <http://www.pkumdl.cn/cavityplus>
- applies Cavity program to detect the potential binding sites and rank them with ligandability and druggability scores
- extracts pharmacophore features within the cavities

# Energy-based methods

## □ Cavity Plus



### ▼ Cavity Results

No. ⚡	Pred. Max pKd ? ⚡	Pred. Avg pKd ⚡	DrugScore ⚡	Druggability ? ⚡	Surface ? ⚡	Residues ? ⚡
1	10.19	6.11	493.00	less druggable	<input checked="" type="checkbox"/>	<a href="#">More</a>
2	8.87	5.66	-745.00	Undruggable	<input type="checkbox"/>	<a href="#">More</a>
3	8.16	5.42	-420.00	Undruggable	<input type="checkbox"/>	<a href="#">More</a>
4	7.87	5.32	-750.00	Undruggable	<input type="checkbox"/>	<a href="#">More</a>
5	7.11	5.06	-1105.00	Undruggable	<input type="checkbox"/>	<a href="#">More</a>
6	6.54	4.86	-992.00	Undruggable	<input type="checkbox"/>	<a href="#">More</a>
7	5.90	4.64	-1123.00	Undruggable	<input type="checkbox"/>	<a href="#">More</a>

Functional sites → binding sites → binding sites for small molecules



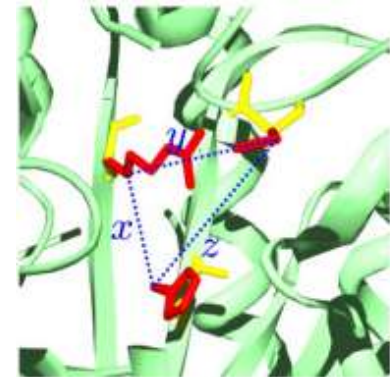
# Binding site similarity



- prediction of binding site is based on its similarity to other (known) binding site

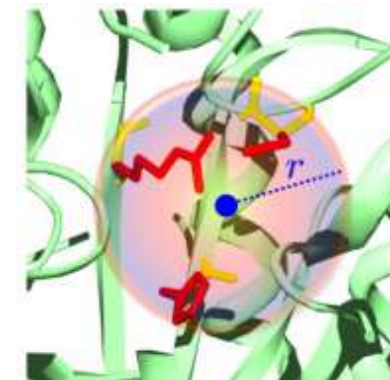
- **template-based** methods

- binding sites are represented by 3D templates



- **microenvironment-based** methods

- similar “microenvironments” associated with binding residues and their distances, physicochemical and evolutionary properties



# Template-based methods



- ❑ definition and construction of **3D templates**
  - **local structural motifs, patterns or descriptors** that characterize the binding sites (e.g., functional groups)
  - capture essence of the binding site in protein
  - usually constraints on atom type and, occasionally, sequential relationships
- ❑ search in database for structures using **template as a query**
  - identification of structures with a given binding site
- ❑ compare the **query structure** against a 3D template database
  - identification of potential binding sites in the query structure

# Template-based methods

## ❑ PINTS

- <http://www.russelllab.org/cgi-bin/tools/pints.pl>
- enables to **compare** a protein structure against a **database of 3D templates** as well as a 3D templates against a **database of protein structures**
- additionally allows comparison of two structures
- the 3D template database includes ligand-binding sites and SITE annotations from PDB files

# Template-based methods

## ❑ ProFunc

- <http://www.ebi.ac.uk/thornton-srv/databases/profunc/>
- aims to identify the most likely **function** of a protein from its 3D structure
- uses series of methods, including fold matching, residue conservation, surface cleft analysis, and functional **3D templates** (templates for enzyme active sites, ligand-binding templates, DNA-binding templates, reverse template comparison vs. structures in wwPDB)

# Template-based methods



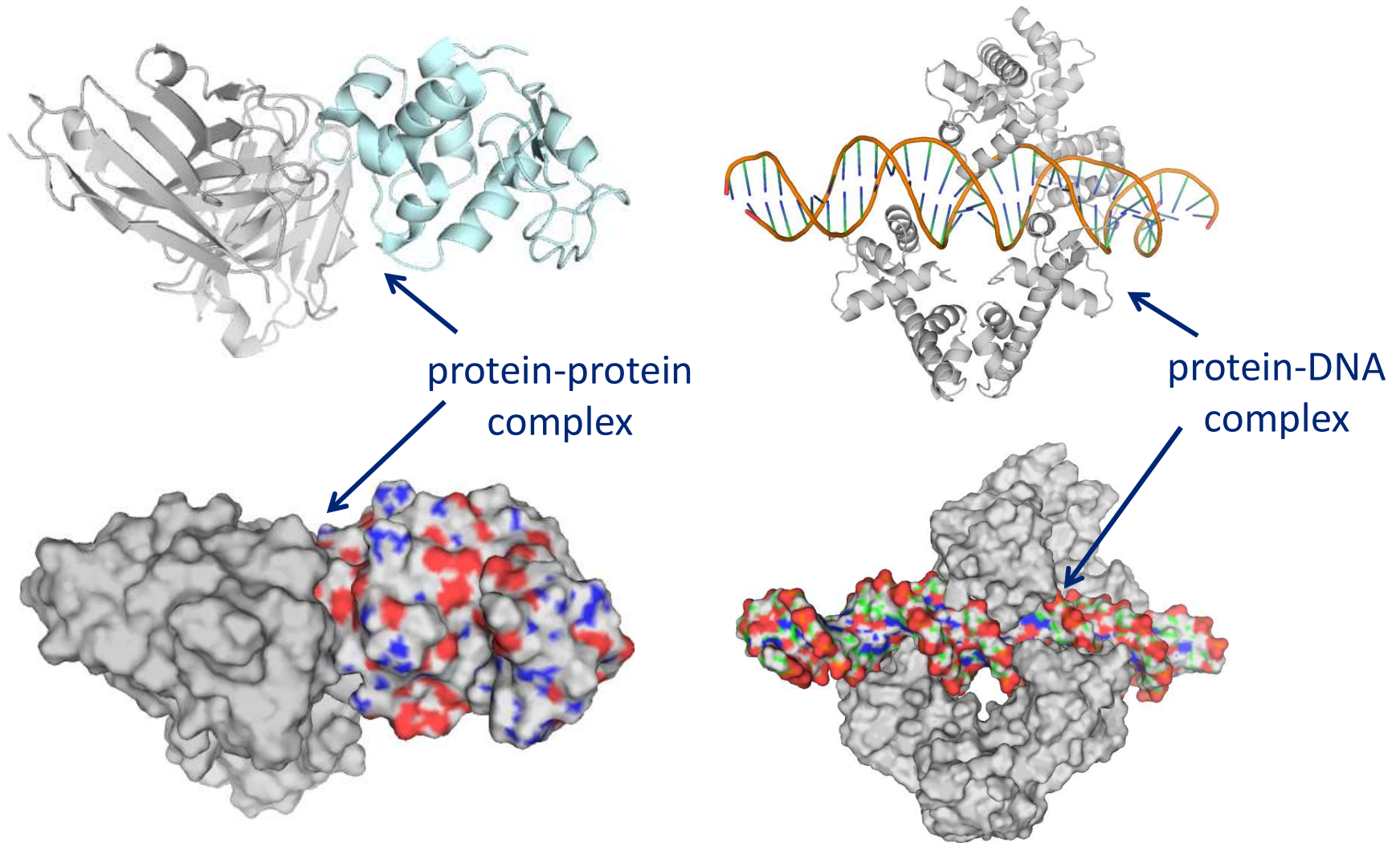
- ❑ Mechanism and Catalytic Site Atlas
  - <https://www.ebi.ac.uk/thornton-srv/m-csa/>
  - provides information about the **active sites**, **catalytic residues** and **mechanisms** in enzymes with experimentally determined 3D structure
  - defines catalytic residues as the residues directly involved in some aspect of the enzymatic reaction
  - provides **3D templates** for catalytic sites in the database

# Binding sites for macromolecules



- ❑ typically **protruding loops**, large surface **clefts** but also **flat** binding **sites** – flatter than binding sites for small molecules
  - recognition of a macromolecule involves interactions over a large continuous surface area or several discrete binding regions
  - **difficult to identify** by a simple examination of the protein structure
- ❑ high evolutionary conservation
- ❑ low desolvation energy
- ❑ characteristic physicochemical properties
- ❑ DNA binding sites have characteristic motifs and positive charged electrostatic patches

# Binding sites for macromolecules



# Binding sites for macromolecules

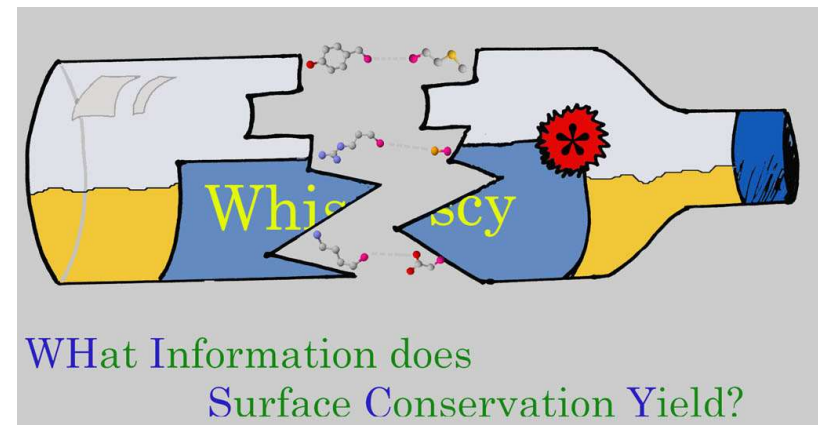


- ❑ approaches to identify binding sites
  - evolutionary **conservation**
  - **knowledge-based**
  
- ❑ meta-servers (combine several tools in a workflow)



# Evolutionary conservation methods

- ❑ same principles as for binding sites of small molecules
- ❑ WHISCY
  - <https://wenmr.science.uu.nl/whiscy/>
  - predicts protein-protein interface using conservation and structural information (interface propensities for each surface residue are used to adjust the score)



# Knowledge-based methods



- ❑ combine multiple **interface features**
  - conservation
  - residue propensity for protein-protein interface
  - physicochemical properties
  - structural properties
- ❑ use known binding sites for parameterization or training →  
**empirical scoring functions** and **machine learning** methods

# Knowledge-based methods

- ❑ cons-PPISP (consensus Protein Protein Interaction Site Predictor)
  - <http://pipe.scs.fsu.edu/ppisp.html>
  - utilizes **machine learning** to predict **protein binding** sites
  - trained on position-specific sequence profiles and solvent accessibilities of each residue and its spatial neighbors
- ❑ Patch Finder Plus
  - <http://pfp.technion.ac.il/>
  - utilizes **machine learning** primarily to find **DNA binding** regions
  - identifies the largest positive electrostatic patch on a protein surface
    - combination of residue frequency, composition and conservation, surface concavity, accessible area and H-bond potential

# Meta-servers

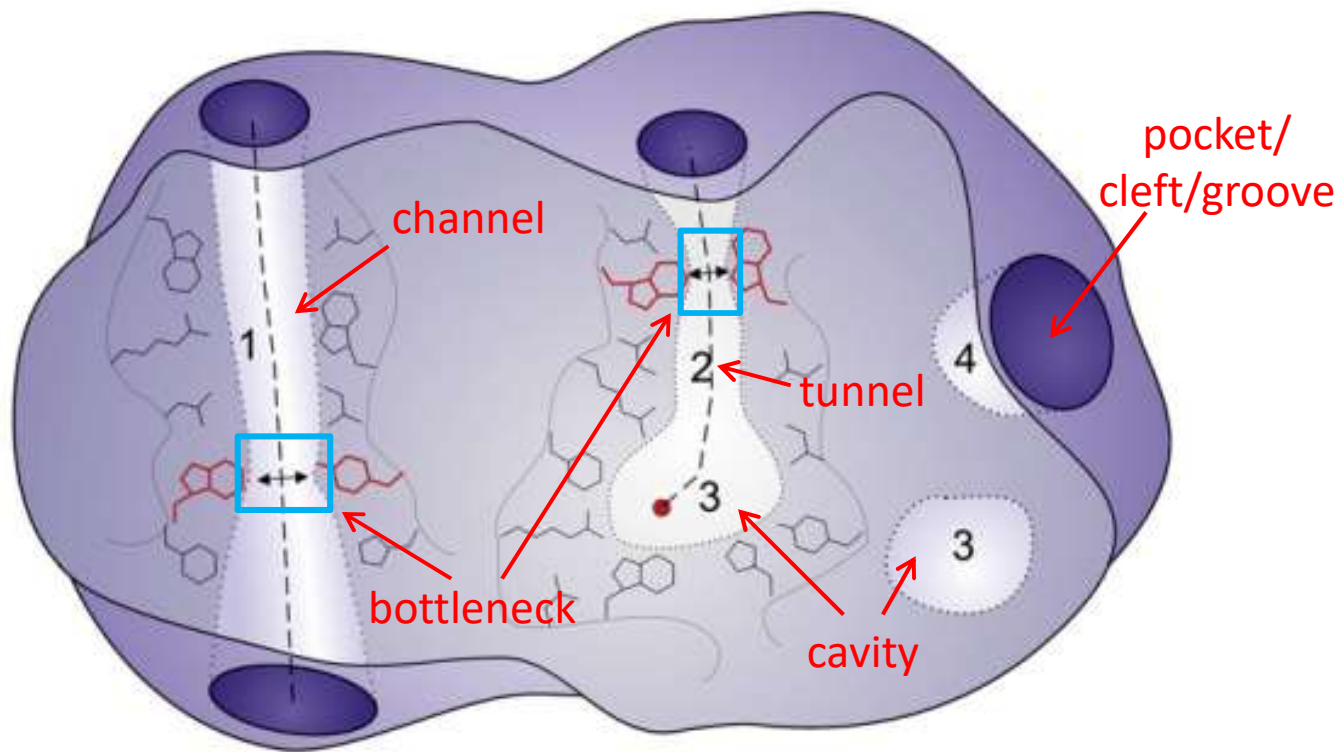
- ❑ combine **multiple methods** to improve prediction accuracy
- ❑ meta-PPISP (Protein Protein Interaction Site Predictor)
  - <http://pipe.scs.fsu.edu/meta-ppisp.html>
  - combines cons-PPISP, ProMate and PINUP
- ❑ PI<sup>2</sup>PE (Protein Interface/Interior Prediction Engine)
  - <http://pipe.scs.fsu.edu/>
  - pipeline to use five different predictors including cons-PPISP, meta-PPISP and DISPLAR

# Transport pathways



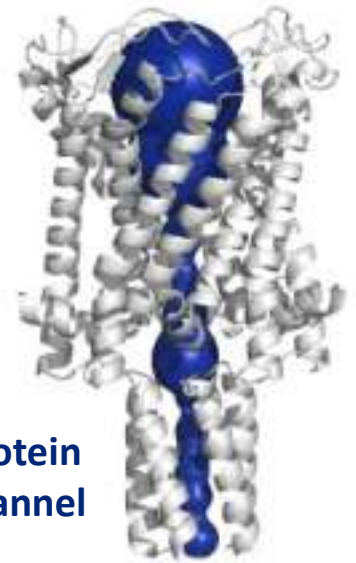
- ❑ mediate transport of ions and small molecules in proteins – an essential role in functioning of large variety of proteins
  - **channels/pores** – transport of substances across membranes
  - **tunnels** – exchange of ligands between the active/binding site and bulk solvent in proteins with buried active/binding site cavities
  - **intramolecular tunnels** – transport of reaction intermediates between two distinct active sites in bifunctional enzymes
- ❑ their **permeability** to different substances depends on the pathway size (radii), shape, length, amino acid composition (physicochemical properties) and dynamics

# Transport pathways

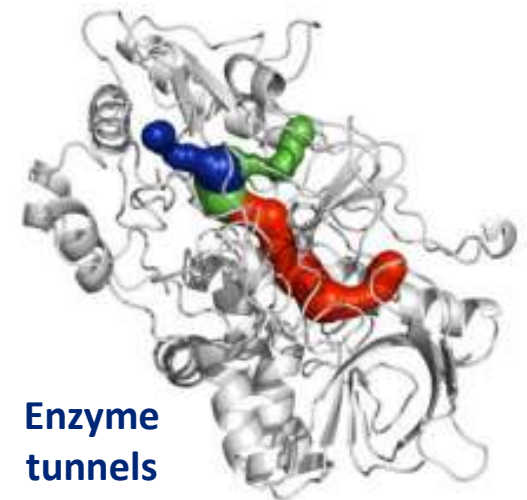


- **bottleneck** – the narrowest part of the tunnel/channel; it has critical importance to the selectivity

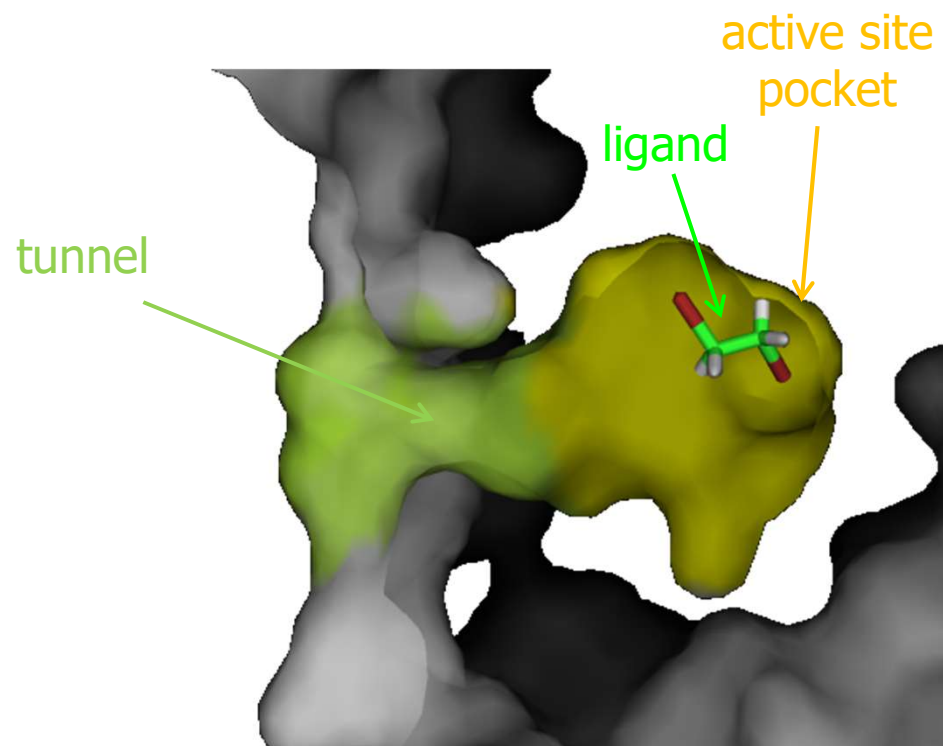
Protein channel



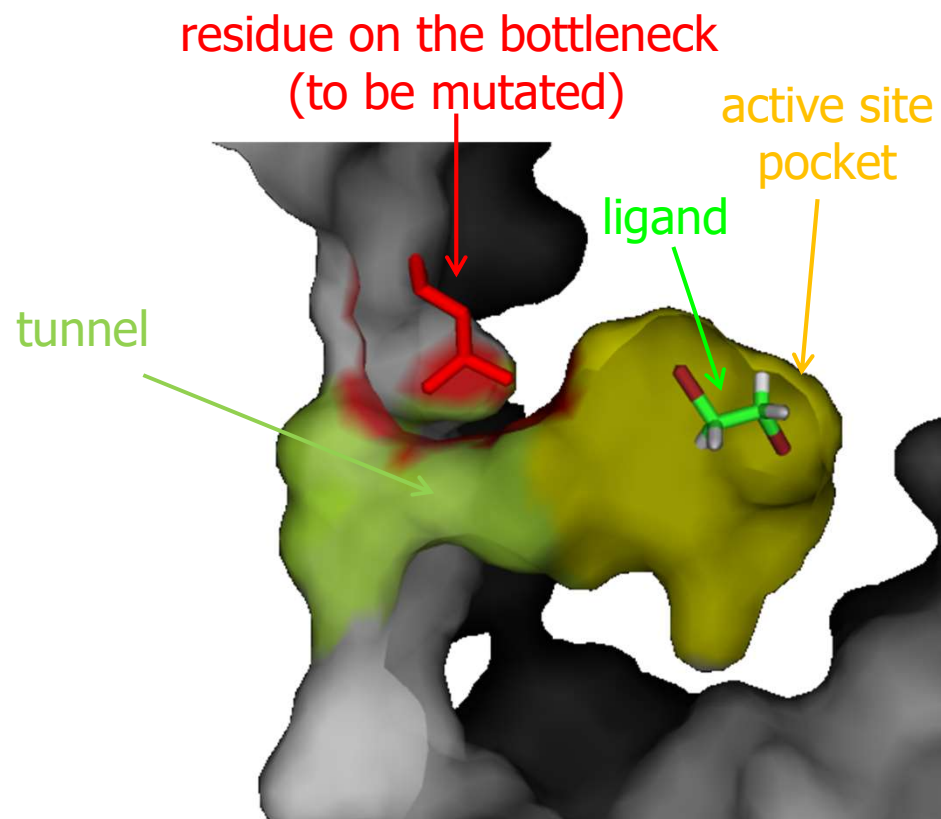
Enzyme tunnels



# Transport pathways

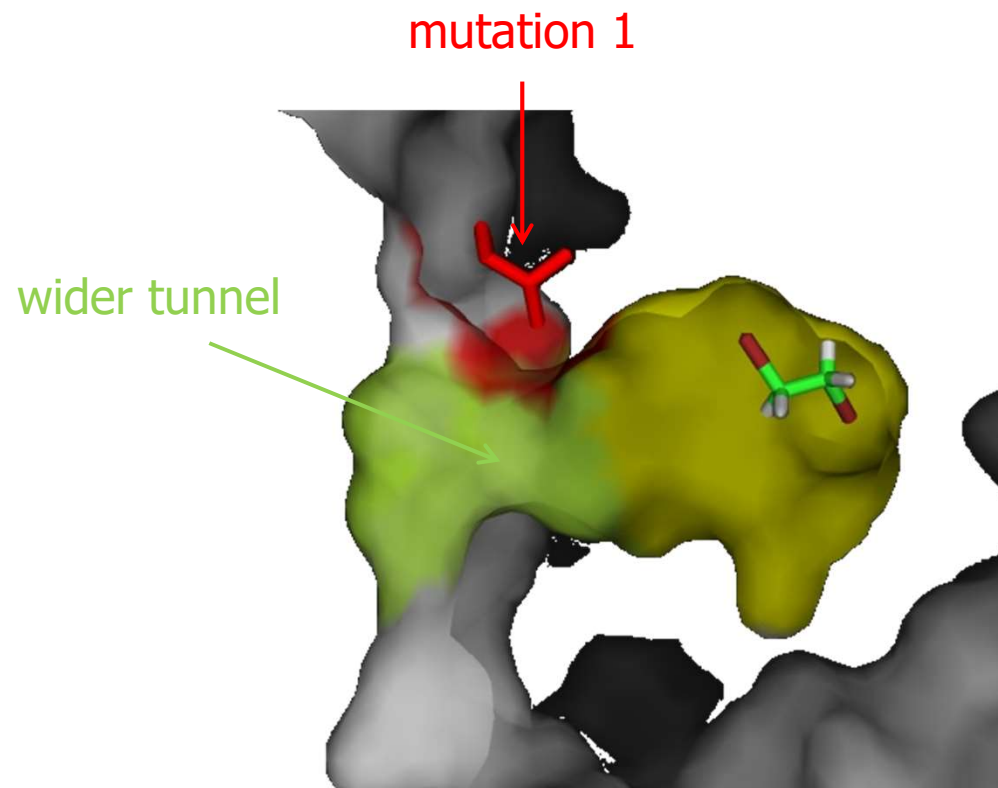


# Transport pathways

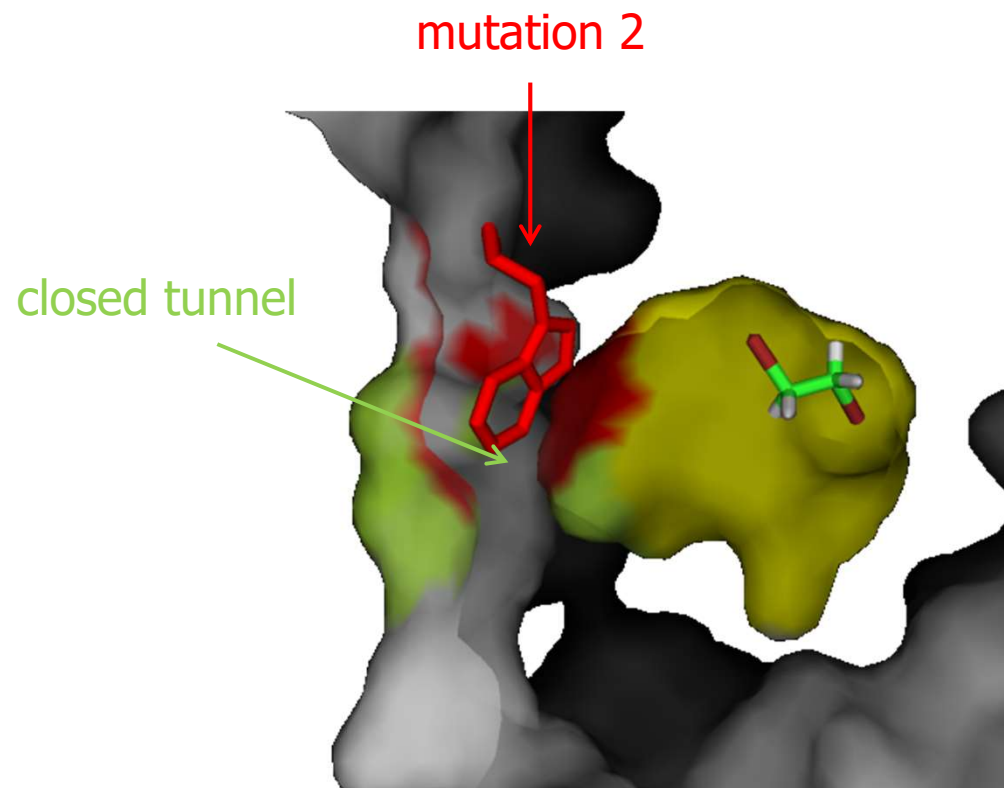




# Transport pathways



# Transport pathways



# Prediction of transport pathways



- ❑ identification of **overall voids** in proteins
- ❑ identification of **tunnels**
- ❑ identification of **channels**

# Identification of overall voids



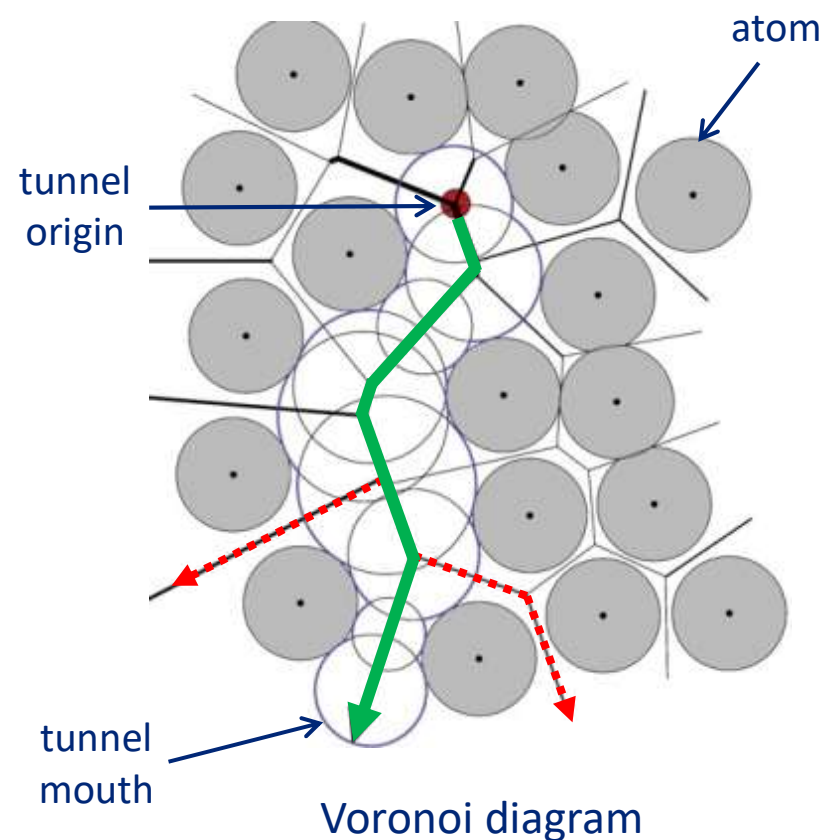
- ❑ methods that aim to accurately represent **all types of voids** in a protein structure, including channels, tunnels, surface clefts, pockets as well as internal cavities
- ❑ usually provide very **limited information** on tunnel and channel characteristics – the identified voids have to be separated from each other
- ❑ **geometry-based** methods for pocket detection
  - HOLLOW – <http://hollow.sourceforge.net/>
  - 3V – <http://3vee.molmovdb.org/>
  - LIGSITE<sup>CSC</sup>, PASS, CASTp, fPocket, SURFNET, POCASA ...




# Identification of tunnels

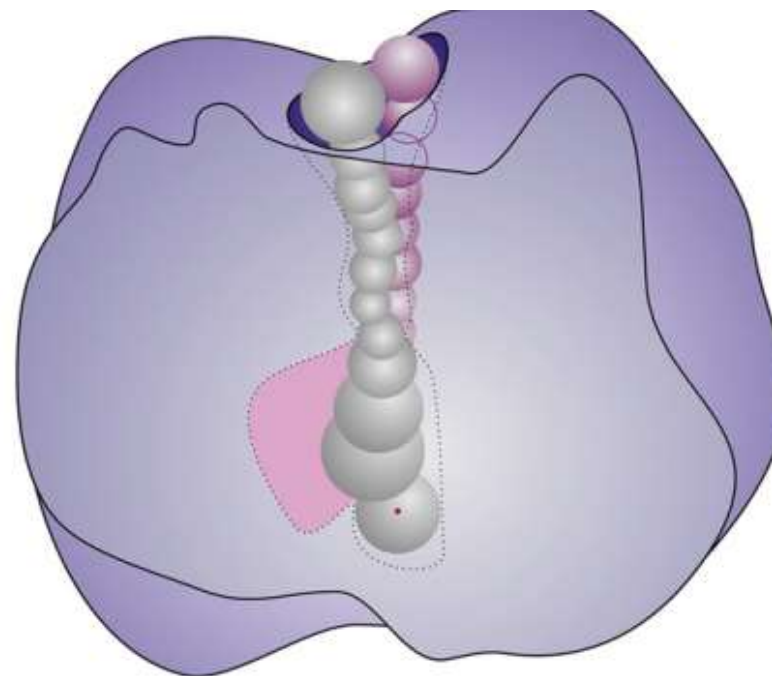


- ❑ methods that calculate **tunnels** connecting occluded cavities with the surrounding bulk solvent
- ❑ identify the pathways **from a cavity** to the **protein surface**
- ❑ **Voronoi diagrams** described by the skeleton of voids between atoms to find all theoretically possible pathways connecting the starting point with the bulk solvent
- ❑ diagram is searched for optimal pathways using Dijkstra's algorithm, based on the criteria defined by a **cost function**
- ❑ the **probe size** defines the lowest radius threshold
- ❑ tunnel geometry is approximated by a sequence of spheres

# Identification of tunnels



-  **Probe size:** the minimum radius specified for the tunnel search
-  Allowed pathway according to the selected probe
-  Disallowed pathways



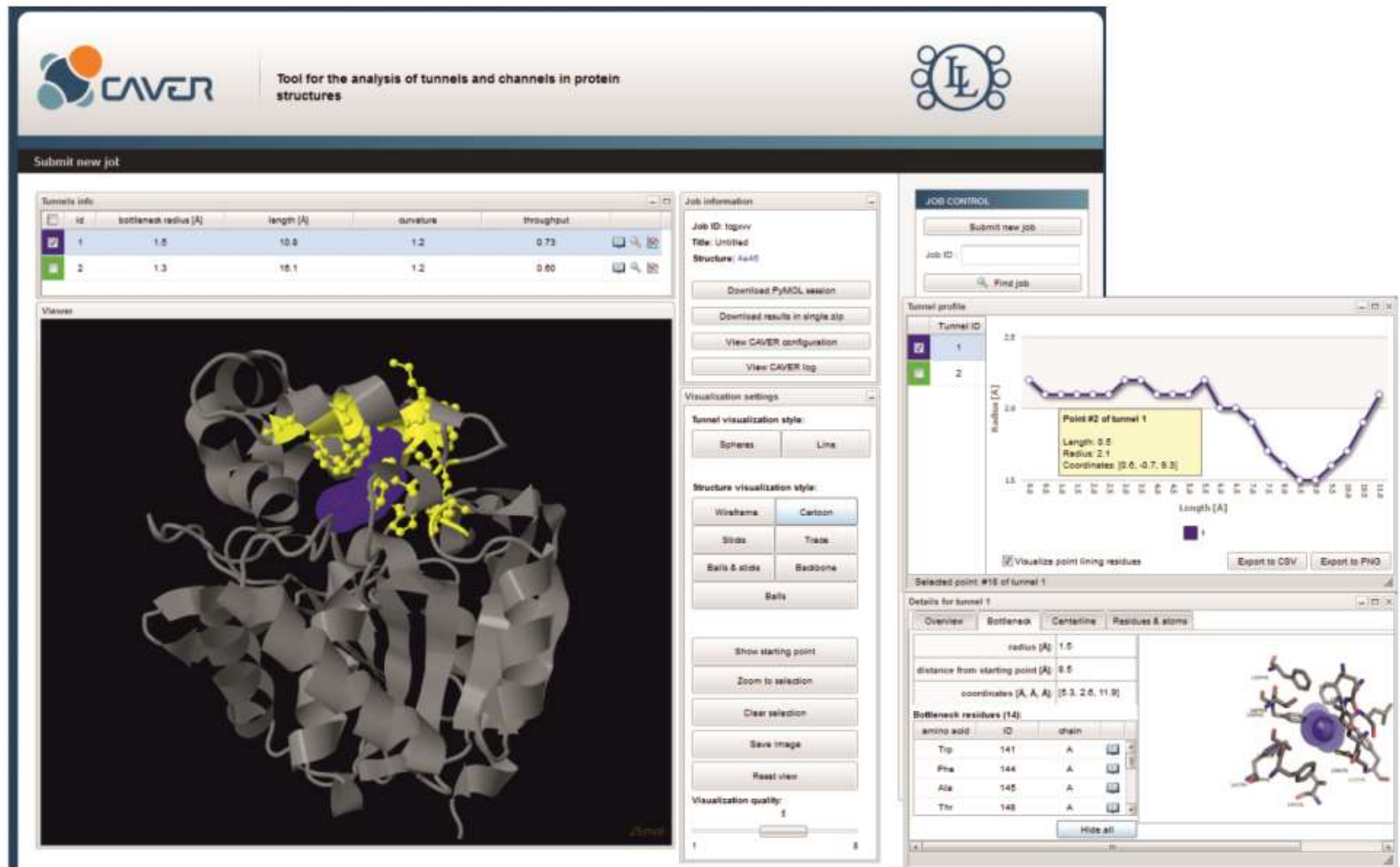
Common limitation: the tools identify two spherical tunnels instead of one asymmetric tunnel

# Identification of tunnels - programs



- ❑ CAVER 3.0
  - <http://caver.cz/>
  - Command-line stand-alone
  - PyMOL plugin
  - GUI with CAVER Analyst 2
- ❑ CAVER Web
  - <http://loschmidt.chemi.muni.cz/caverweb/>
  - interactive guide-through web server
  - optimized protocol for detection of biologically relevant tunnels
  - based on CAVER 3.0 program

# Identification of tunnels - programs



Functional sites → transport pathways

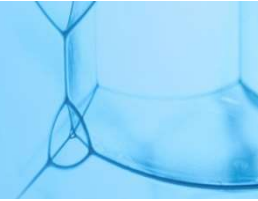


# Identification of tunnels - programs



- ❑ MolAxis
  - <http://bioinfo3d.cs.tau.ac.il/MolAxis/>
- ❑ MOLE 2.0
  - <http://mole.upol.cz/>
- ❑ ChExVis
  - <http://vgl.serc.iisc.ernet.in/chexvis/>
- ❑ BetaCavityWeb
  - <http://voronoi.hanyang.ac.kr/betacavityweb/>
- ❑ ...

# Identification of channels

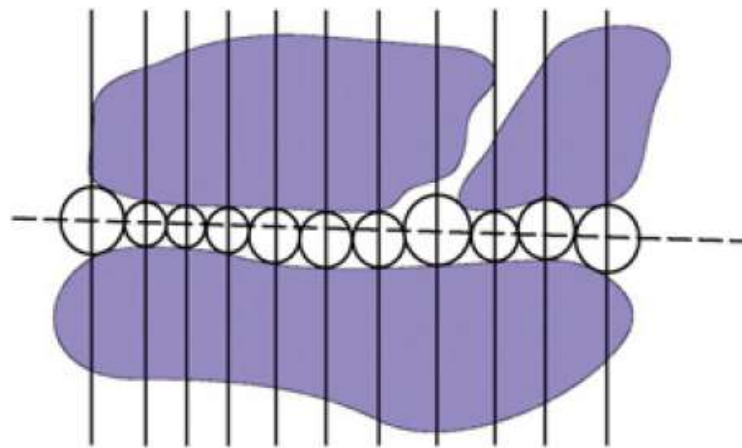


- ❑ methods calculate **channels (pores)** penetrating throughout proteins
- ❑ not convenient for identifying tunnels leading from occluded cavities
- ❑ usually analyze just one channel per structure
- ❑ usually need information about approximate position and direction of the channel (**channel axis**) – user-provided or automatically identified

# Identification of channels - programs

## ❑ POREWALKER

- <http://www.ebi.ac.uk/thornton-srv/software/PoreWalker/>
- identifies **channel axis** by heuristic iterative approach (based on the axes of transmembrane secondary structures)
- protein is divided into equally-spaced slices perpendicular to the axis; the largest spheres fitting the channel are identified

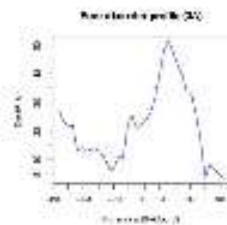


# Identification of channels - programs

## □ POREWALKER

### Pore analysis results

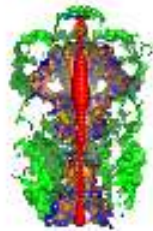
#### Overview of the available results:



Pore shape



Slideshow of the pore cross-sections



Pore visualization



Features of the cavity

# References

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