

# CG920 Genomics

## Lesson 2

### Genes Identification

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# Literature

- Literature sources for Chapter 02:

- Plant Functional Genomics, ed. Erich Grotewold, 2003, Humana Press, Totowa, New Jersey
- Majoros, W.H., Pertea, M., Antonescu, C. and Salzberg, S.L. (2003) GlimmerM, Economy, and Unveil: three ab initio eukaryotic gene finders. *Nucleic Acids Research*, **31**(13).
- Singh, G. and Lykke-Andersen, J. (2003) New insights into the formation of active nonsense-mediated decay complexes. *TRENDS in Biochemical Sciences*, **28** (464).
- Wang, L. and Wessler, S.R. (1998) Inefficient reinitiation is responsible for upstream open reading frame-mediated translational repression of the maize R gene. *Plant Cell*, **10**, (1733)
- de Souza et al. (1998) Toward a resolution of the introns earlylate debate: Only phase zero introns are correlated with the structure of ancient proteins *PNAS*, **95**, (5094)
- Feuillet and Keller (2002) Comparative genomics in the grass family: molecular characterization of grass genome structure and evolution *Ann Bot*, **89** (3-10)
- Frobis, A.C., Matus, D.Q., and Seaver, E.C. (2008). Genomic organization and expression demonstrate spatial and temporal Hox gene colinearity in the lophotrochozoan Capitella sp. I. *PLoS One* **3**, e4004



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# Outline

- Forward and Reverse Genetics Approaches
  - Differences between the approaches used for identification of genes and their function
- Identification of Genes *Ab Initio*
  - Structure of genes and searching for them
  - Genomic colinearity and genomic homology
- Experimental Genes Identification
  - Constructing gene-enriched libraries using methylation filtration technology
  - EST libraries
  - Forward and reverse genetics



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# Outline

- **Forward and Reverse Genetics Approaches**
  - Differences between the approaches used for identification of genes and their function



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# Forward vs. Reverse Genetics

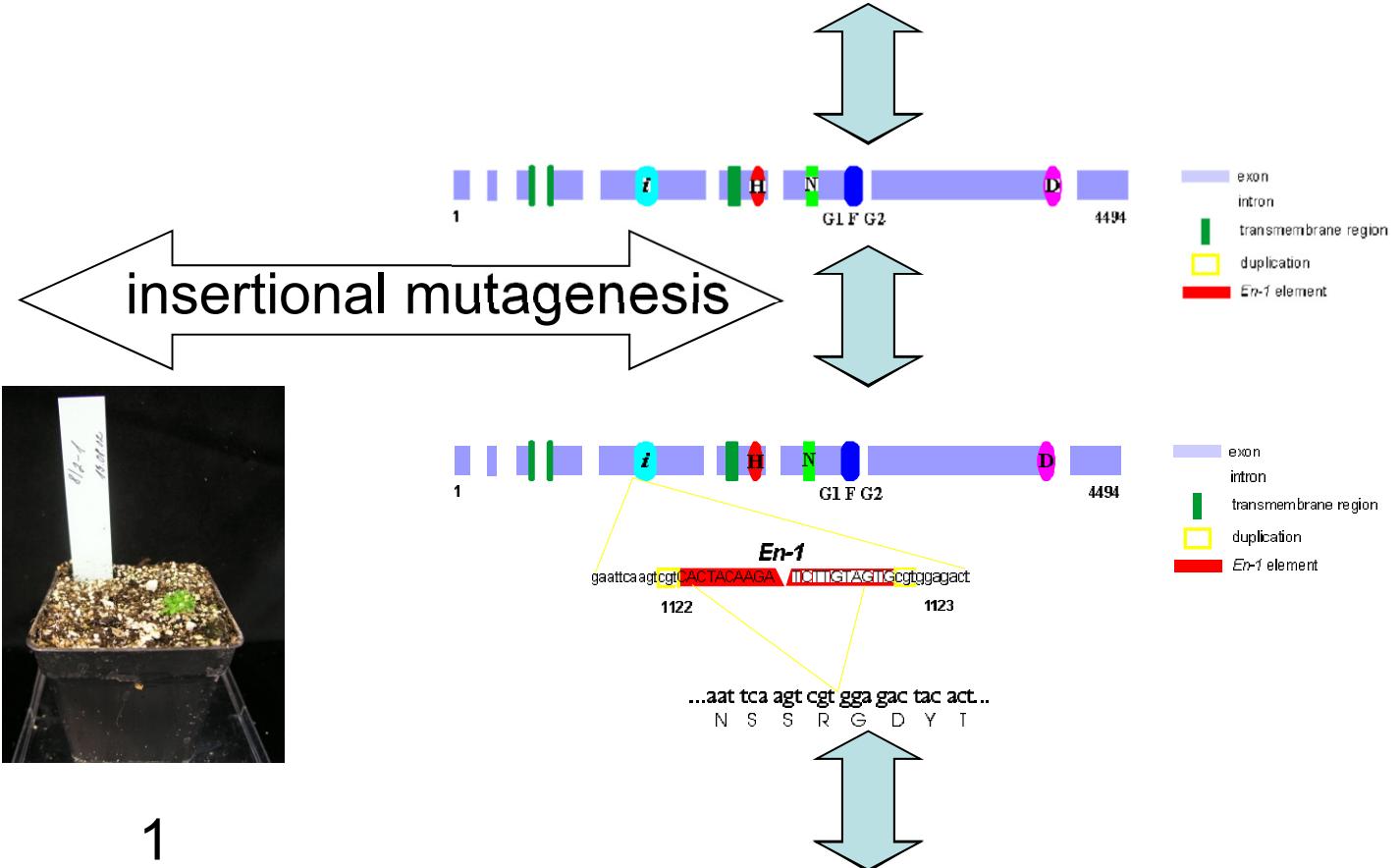
## Revolution in understanding the term „gene“

„classical“ genetics approaches



„reverse genetics“ approaches

5'TTATATATATATTAAAAAATAAAATAAAA  
GAACAAAAAAGAAAATAAAAATA....3'



3 : 1



# Identification of the role of *ARR21* gene

- Hypothetical signal transducer in two-component system of *Arabidopsis*

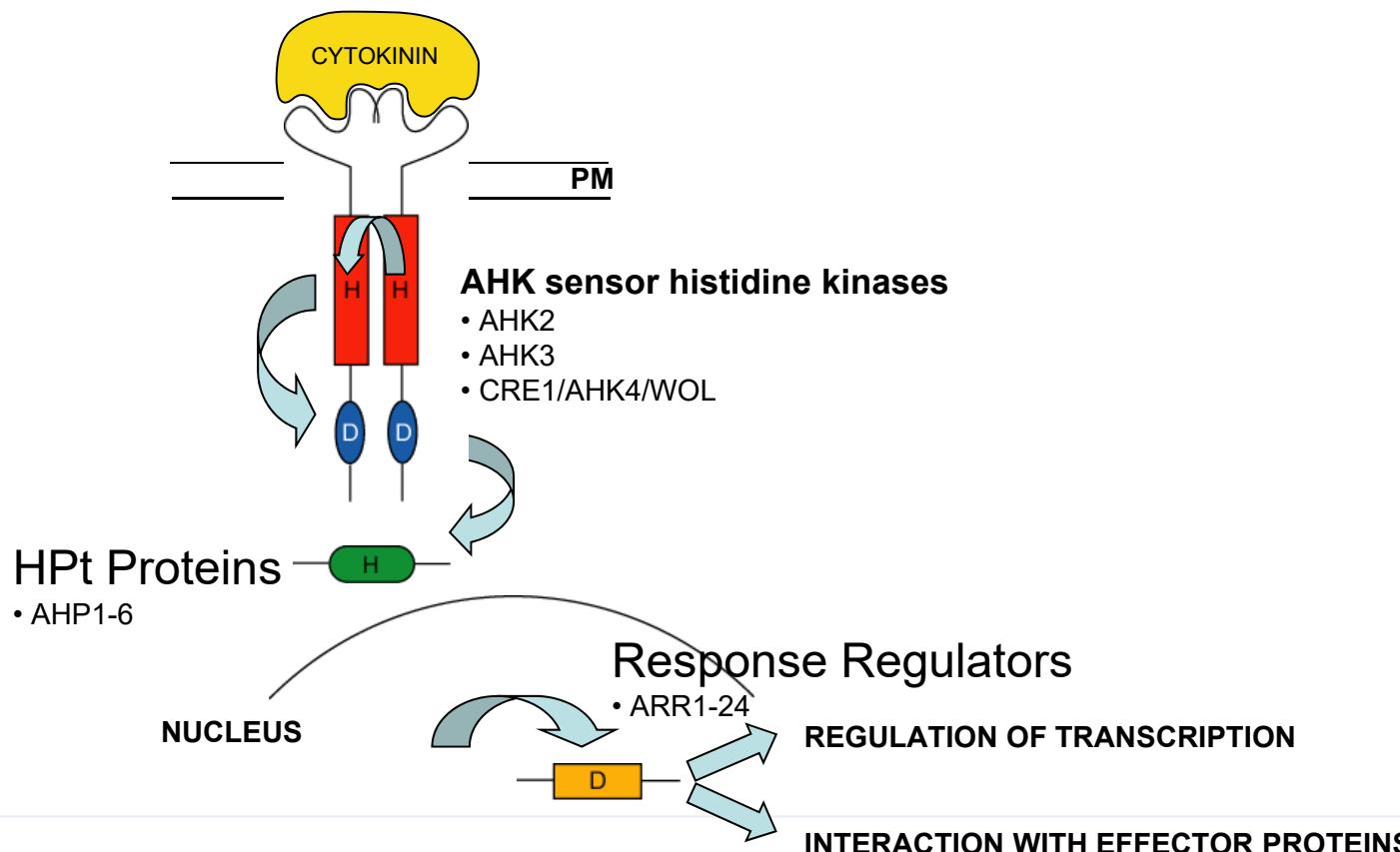


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# Identification of the role of *ARR21* gene

Recent Model of the CK Signaling via Multistep Phosphorelay (MSP) Pathway



# Identification of the role of *ARR21* gene

- Hypothetical signal transducer in two-component system of *Arabidopsis*
- Mutant identified by searching in databases of insertional mutants (SINS-sequenced insertion site) using BLAST



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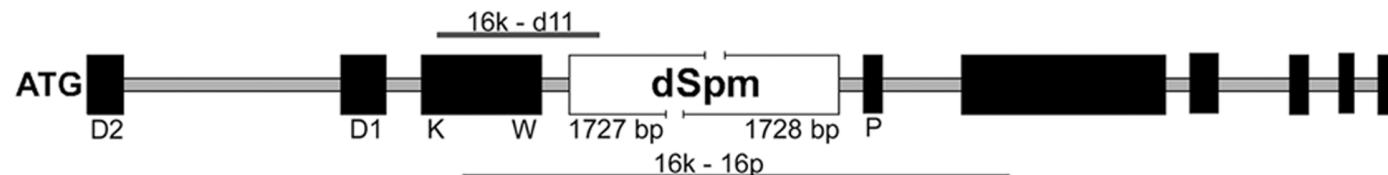
# Identification of the role of *ARR21* gene – isolation of insertional mutant

- **Searching in databases of insertional mutants (SINS)**

Insert\_SINS: 01\_09\_64  
Query: 80 tcctagcggtcatgagcgtaaccataacttgacaanaagagaacgttagccagccatgg 139  
||||||||||||||||||||||||||||||||||||||||||||||||||||  
Sbjct: 58319 tcctagcggtcatgagcgtaaccataacttgacaagagagaacgttagccagccatgg 58378  
Arr21: 1830

Insert\_SINS: 01\_09\_64  
Query: 140 tttgatatctttgtcaaaaatgttttggattttactgt 179  
||||||||||||||||||||||||||||||||||||||||  
Sbjct: 58379 tttgatatctttgtcaaaaatgttttggattttactgt 58418  
Arr21: 1890

- Localization of *dSpm* insertion in genome sequence of *ARR21* using sequenation of PCR products



# Identification of the role of *ARR21* gene

- Hypothetical signal transducer in two-component system of *Arabidopsis*
  - Mutant identified by searching in databases of insertional mutants (SINS-sequenced insertion site) using BLAST
  - Expression of *ARR21* in wild-type and inhibition of expression of *ARR21* in insertional mutant confirmed at the RNA level

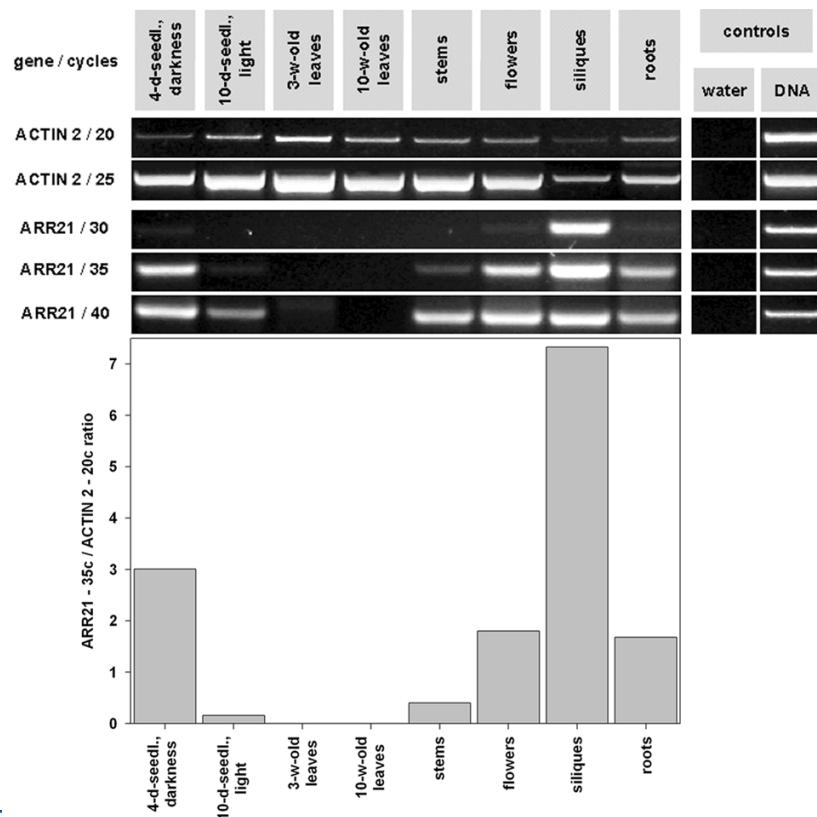


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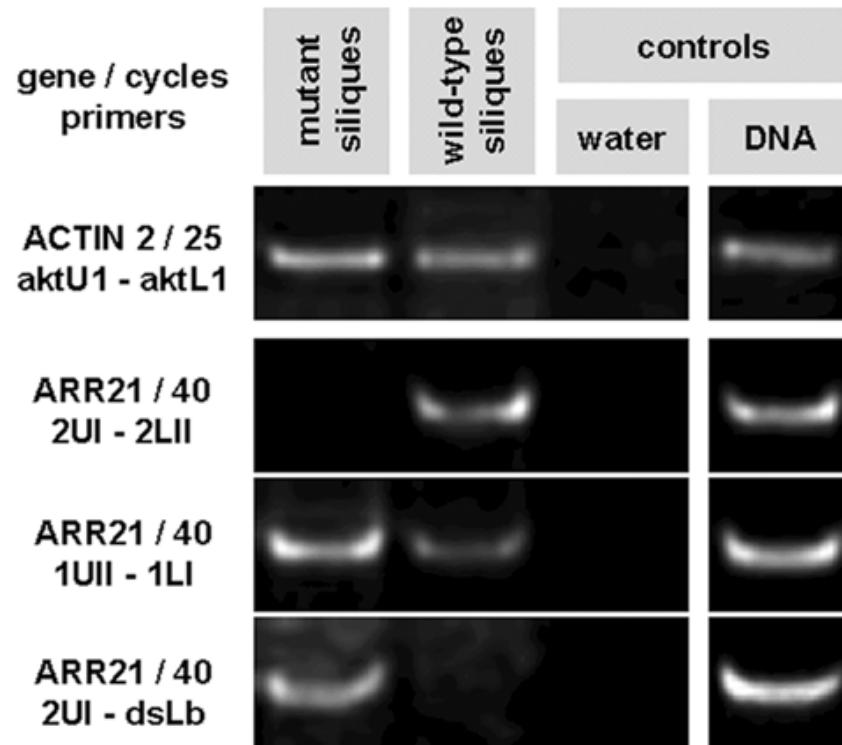
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# Identification of the role of *ARR21* gene – analysis of expression

wild type expression



insertional mutant vs wild type



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# Identification of the role of *ARR21* gene

- Hypothetical signal transducer in two-component system of *Arabidopsis*
  - Mutant identified by searching in databases of insertional mutants (SINS-sequenced insertion site) using BLAST
  - Expression of *ARR21* in wild-type and inhibition of expression of *ARR21* in insertional mutant confirmed at the RNA level
  - Phenotype analysis of insertional mutant



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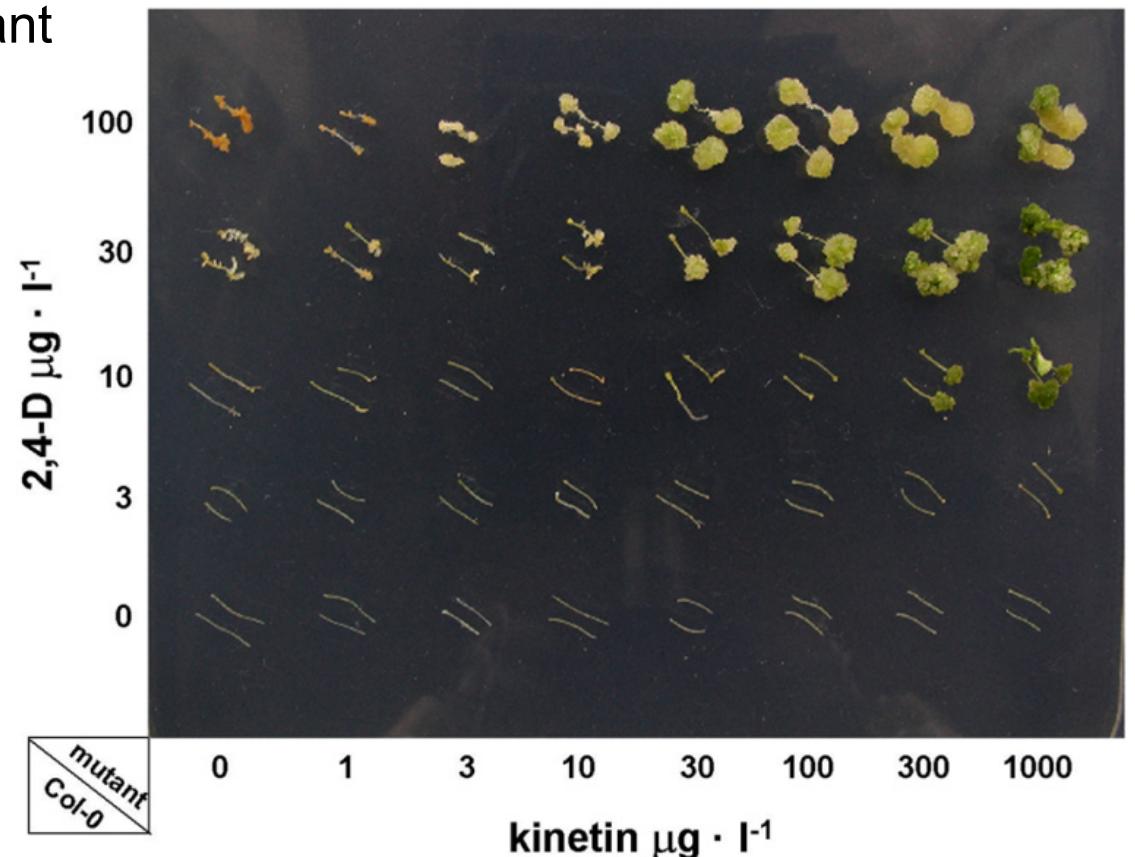


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# Identification of the role of *ARR21* gene – phenotype analysis of mutant

- Analysis of sensitivity to plant growth regulators
  - 2,4-D a kinetin
  - ethylene
  - Light of various wavelengths
- No alterations - nor in flowering, neither in the number of the seeds



# Identification of the role of *ARR21* gene – possible reasons for the absence of the phenotype

- Functional redundancy within the gene family



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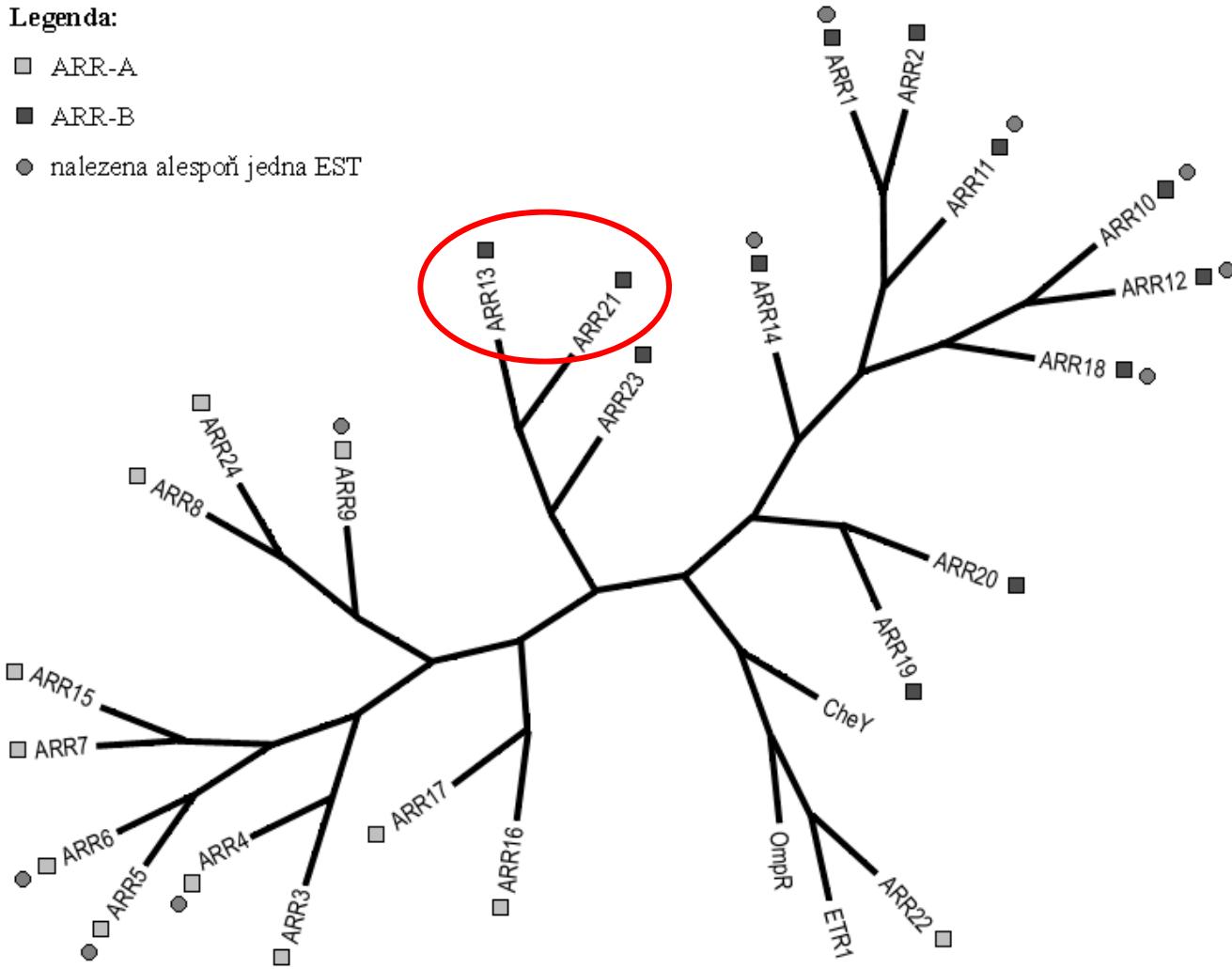
# Identification of the role of *ARR21* gene – homology of *ARR* genes

Legenda:

■ ARR-A

■ ARR-B

● nalezena alespoň jedna EST



# Identification of the role of *ARR21* gene – causes of absence of the phenotype

- Functional redundancy within the gene family?
- Phenotype only under specific conditions



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# Identification of the role of *ARR21* gene – summary

- Gene *ARR21* identified by comparative analysis of *Arabidopsis* genome
- Based on sequence analysis, its function was predicted
- Site-specific expression of *ARR21* gene was proved at the RNA-level
- Identification of gene function by insertional mutagenesis in case of *ARR21* in development of *Arabidopsis* was not successful, probably because of functional redundancy within the gene family



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- Identification of Genes *Ab Initio*
  - Structure of genes and searching for them



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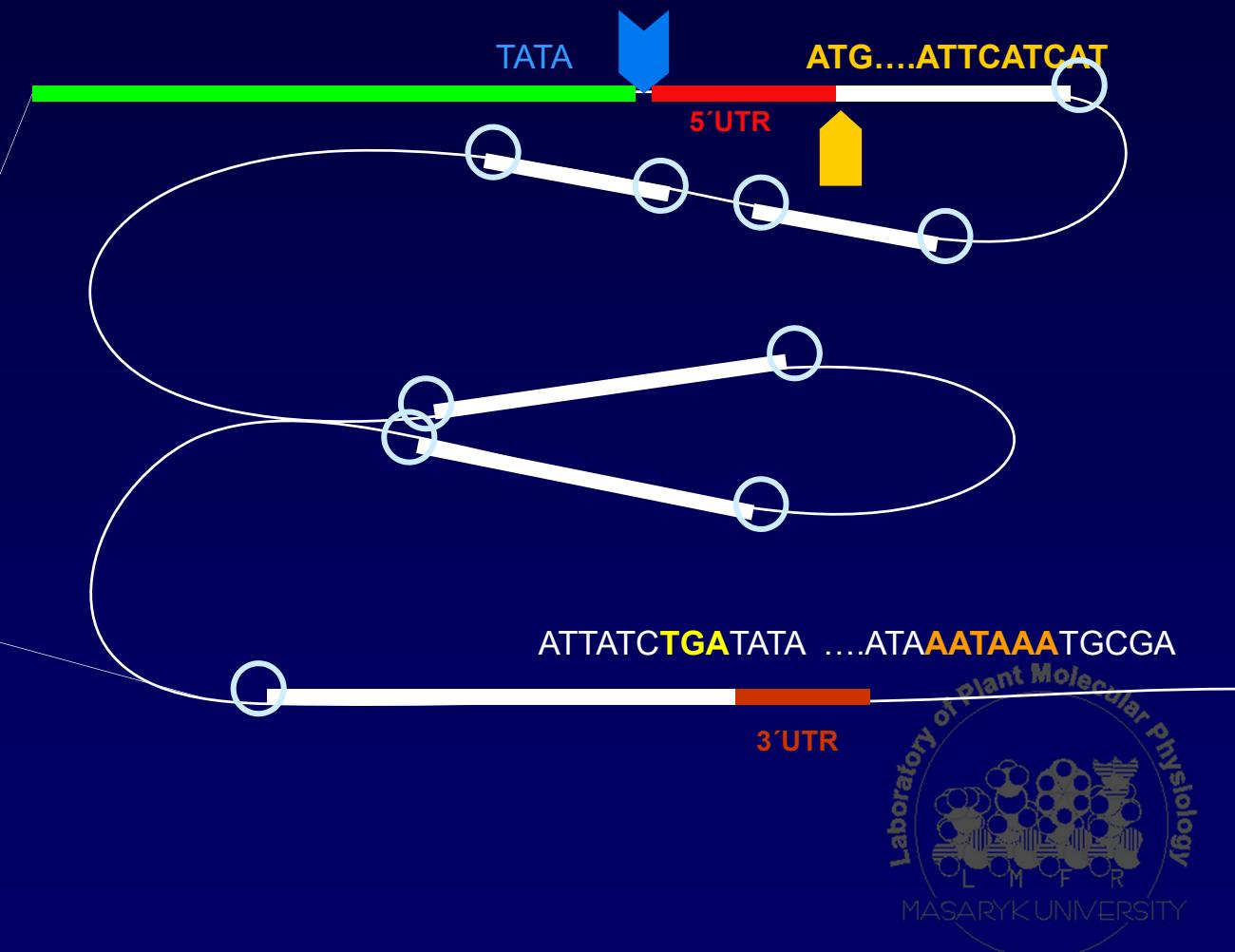


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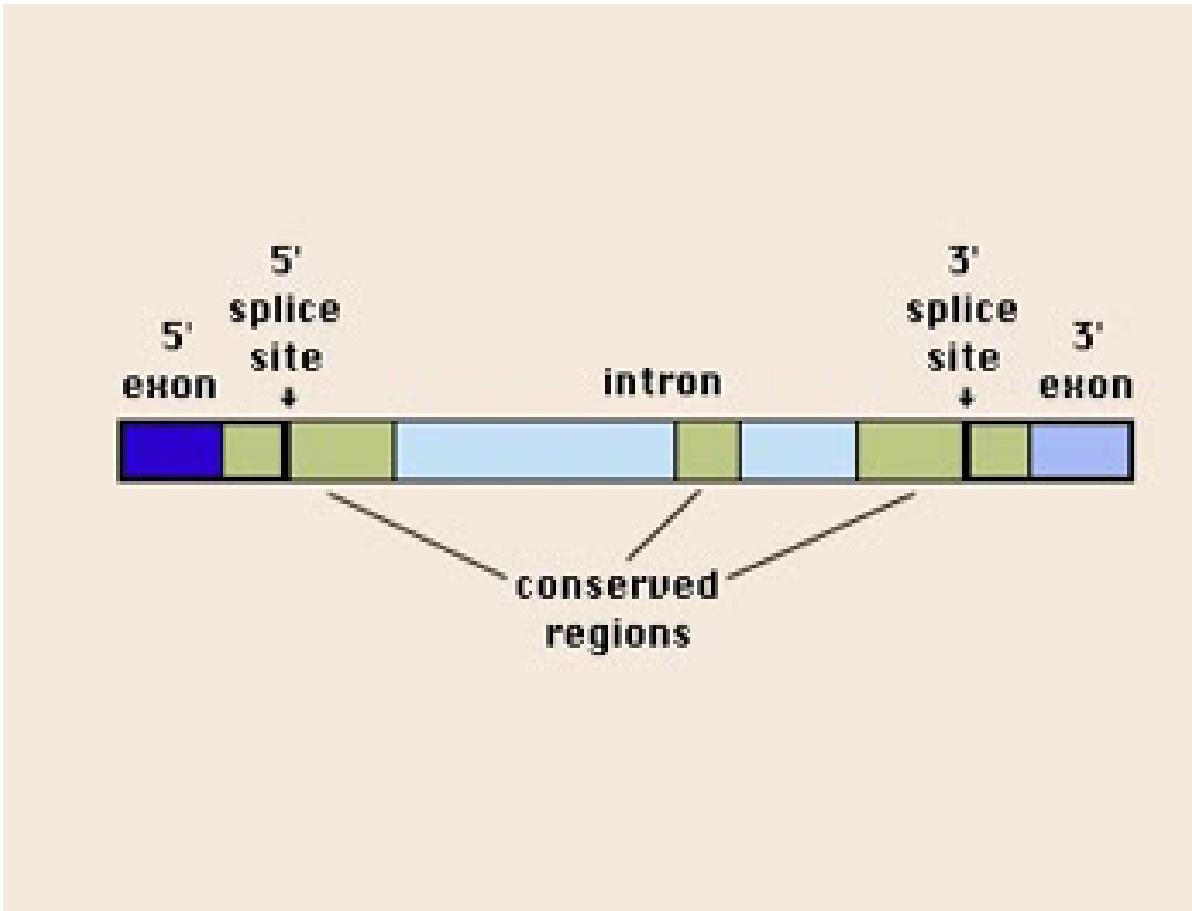
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# Genes Structure

- Promoter
- Transcriptional start
- 5'UTR
- Translational start
- Splicing sites
- Stop codon
- 3'UTR
- Polyadenylation signal



# RNA Splicing



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# Identification of Genes *Ab Initio*

- Omitting 5' and 3' UTR
- Identification of translation start (ATG) and stop codon (TAG, TAA, TGA)
- Finding donor (typically GT) and acceptor (AG) splicing sites
- Using various statistic models (e.g. Hidden Markov Model – HMM, see recommended literature, Majoros *et al.*, 2003) to evaluate and score the weight of identified donor and acceptor sites



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# Splicing Site Prediction

- Programs for splice site prediction  
(specify approximately 35 %)
  - GeneSplicer ([http://www.tigr.org/tdb/GeneSplicer/gene\\_spl.html](http://www.tigr.org/tdb/GeneSplicer/gene_spl.html))
  - SplicePredictor (<http://deepc2.psi.iastate.edu/cgi-bin/sp.cgi>)



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# SplicePredictor

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Bioinformatics 2  
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## SplicePredictor

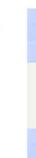
- a method to identify potential splice sites in (plant) pre-mRNA by sequence inspection using Bayesian statistical models  
(click [here](#) to access the older method using logitlinear models)

Sequences should be in the one-letter-code ( $\{a,b,c,g,h,k,m,n,r,s,t,u,w,y\}$ ), upper or lower case; all other characters are ignored during input. Multiple sequence input is accepted in **FASTA** format (sequences separated by identifier lines of the form “>SQ;name\_of\_sequence comments”) or in **GenBank** format.

Paste your genomic DNA sequence here:

```
GAGGAGGGCACAAATGACGAATATACAAAATGATCTTAAACAGCTAAACTATATTGGACATTTCGATCTCAGATATA  
AAAGATTTCAATTAAATACTTGATAAATACTCTTATTATTTCTTAGTTATTAAAAAAAACCTCTAATAAAAT  
ACGAGTTAACGTTAACAAATCGCTTAGACTAAAATACACCATATAATTCAAACGATAAAGTTACAAAGTAATATCC  
AAGTATCTCATAGTCAACATATATAGTAAATAATTAGTTGACGTATAAGAAAATAAAATAAAATTAGTATCTTAT  
TTGGGTGGTGCTGACTGGTGAUTGCTCGGAAATGGAACCATAATCCAAGACATGGTTTAGAT
```

... or upload your sequence file (specify file name):



... or type in the GenBank accession number of your sequence:



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# SplicePredictor

## What do the output columns mean?

SplicePredictor. Version of February 13, 2005.  
Date run: Wed Nov 9 11:30:14 2005

Species: Homo sapiens  
Model: 2-class Bayesian  
Prediction cutoff (2 ln[BF]): 3.00  
Local pruning: on  
Non-canonical sites: not scored

Sequence 1: your-sequence, from 1 to 9490.

### Potential splice sites

t	q	loc	sequence	P	c	rho	gamma	*	P*R*G*
A	<--	75	tttttcgcatctcAGat	0.973	7.16	0.000	0.000	7	(5 1 1)
A	<--	134	atttattttttttAGtt	0.999	14.86	0.000	0.000	7	(5 1 1)
A	<--	500	gattttgttggAGtc	0.977	7.48	0.000	0.000	7	(5 1 1)
A	<--	780	tctgttatttgttAGct	0.986	8.56	0.000	0.000	7	(5 1 1)
A	<--	848	tattttttaaatAGat	0.968	6.80	0.000	0.000	7	(5 1 1)
A	<--	1051	caattttttttttaAGaa	0.930	5.19	0.000	0.000	7	(5 1 1)
A	<--	1213	tatttttttttttAGtt	0.998	12.14	0.000	0.000	7	(5 1 1)
A	<--	1373	tttccctctcacAGga	0.999	13.17	0.000	0.000	7	(5 1 1)
A	<--	1487	tttatatatattatAGtg	0.883	4.04	0.000	0.000	7	(5 1 1)
A	<--	1581	atgtgttgcgttAGga	0.982	8.03	0.000	0.000	7	(5 1 1)
A	<--	1781	ggttgtcgaaaaAGgg	0.886	4.10	0.000	0.000	7	(5 1 1)
A	<--	2440	taattaaaattttAGat	0.939	5.46	0.000	0.000	7	(5 1 1)
A	<--	2479	catctaaattttAGat	0.942	5.59	0.000	0.000	7	(5 1 1)
D	---->	2546	aagGTagta	0.909	4.61	0.885	1.903	15	(5 5 5)
A	<--	2572	tttttttttttgcAGca	0.930	5.16	0.000	0.000	7	(5 1 1)
A	<---	2763	ctcaaaatccaaAGgt	0.873	3.86	0.185	0.000	11	(5 5 1)
A	<---	2782	tttcgttttcattAGcg	0.952	5.98	0.220	0.000	11	(5 5 1)
A	<---	3022	tttgtttgtactAGct	0.956	6.16	0.221	0.000	11	(5 5 1)
A	<---	3048	cttgcataatAGga	0.973	7.15	0.229	0.000	11	(5 5 1)
A	<---	3171	cgtcgttatttAGta	0.988	8.74	0.000	0.000	7	(5 1 1)
A	<---	3284	tttttgttatcaaAGgt	0.993	10.03	0.000	0.006	8	(5 1 2)
D	---->	3372	aatGtaagg	0.933	5.28	0.855	1.849	15	(5 5 5)
A	<--	3451	aatgtttctcgAGaa	0.916	4.77	0.293	0.065	12	(5 5 2)
A	<--	3581	cgatcgccgttAGgt	0.850	3.47	0.000	0.000	7	(5 1 1)
D	---->	3649	cacGTatta	0.933	5.25	0.000	1.848	11	(5 1 5)
A	<--	3695	tttgtgttatacaaAGgt	0.907	4.56	0.000	0.000	7	(5 1 1)
A	<--	4254	attttgttcttcAGat	0.998	12.82	0.000	0.002	8	(5 1 2)
A	<--	4351	tttcttacatggAGaa	0.991	9.42	0.000	0.000	7	(5 1 1)
A	<--	4633	gttttgttttttAGgg	0.879	3.97	0.000	0.000	7	(5 1 1)
A	<--	4976	cttgtgttttcAGct	0.952	5.98	0.000	0.000	7	(5 1 1)
A	<--	5004	tttttttttttgcAGAg	0.996	11.17	0.000	0.000	7	(5 1 1)
D	---->	5356	caaGTaaat	0.821	3.04	0.387	0.000	11	(5 5 1)
D	---->	5384	ttgGtaaga	0.941	5.54	0.478	0.090	13	(5 5 3)
A	<--	5403	actctgttttttAGct	0.894	4.26	0.000	0.000	7	(5 1 1)
A	<--	5441	cttttttttttacAGaa	0.995	10.43	0.387	0.000	11	(5 5 1)
A	<--	5472	tttttttttttacAGct	0.965	6.62	0.478	0.090	13	(5 5 3)
D	---->	5745	gcgGTaaag	0.991	9.48	0.990	1.956	15	(5 5 5)
A	<--	5808	catcatatccaaAGgt	0.948	5.83	0.458	0.000	11	(5 5 1)
A	<--	6135	ggtttatttttAGgt	0.999	13.59	0.508	0.050	12	(5 5 2)
A	<--	6552	ggattttcacctcAGag	0.938	5.42	0.000	0.000	7	(5 1 1)



# Splicing Site Prediction

- Programs for splice site prediction  
(specify approximately 35 %)
  - GeneSplicer ([http://www.tigr.org/tdb/GeneSplicer/gene\\_spl.html](http://www.tigr.org/tdb/GeneSplicer/gene_spl.html))
  - SplicePredictor (<http://deepc2.psi.iastate.edu/cgi-bin/sp.cgi>)
  - NetGene2 (<http://www.cbs.dtu.dk/services/NetGene2/>)



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# NetGene2



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## NetGene2 Server

The NetGene2 server is a service producing neural network predictions of splice sites in human, *C. elegans* and *A. thaliana*.

Instructions

Output format

Abstract

Performance

### SUBMISSION

Submission of a local file with a single sequence:

File in **FASTA** format

Human

*C. elegans*

*A. thaliana*

Submission by pasting a single sequence:

Sequence name

Human

*C. elegans*

*A. thaliana*

Sequence

GAGGAGGCACAAATGACGAATATAACAAATGATCTAACAGCTAAACTATATTGGACATTTTCGATC  
TCAGATATA  
AAAGATTTCATTCAATATAACTTGGATAAAACTCTTATTATTTCTTAGTTATTAAAAAAAACCT  
CTAATAAT  
ACGAGTTAAGTCCACAAATCGCTTAGACTAAAATACACCATATAATTCAAACGATAAGTTACAAAA



NOTE: The submitted sequences are kept confidential and will be erased immediately after processing.

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# NetGene2

Prediction done

\*\*\*\*\* NetGene2 v. 2.4 \*\*\*\*\*

The sequence: Sequence has the following composition:

Length: 9490 nucleotides.  
31.8% A, 17.0% C, 19.6% G, 31.7% T, 0.0% X, 36.5% G+C

### Donor splice sites, direct strand

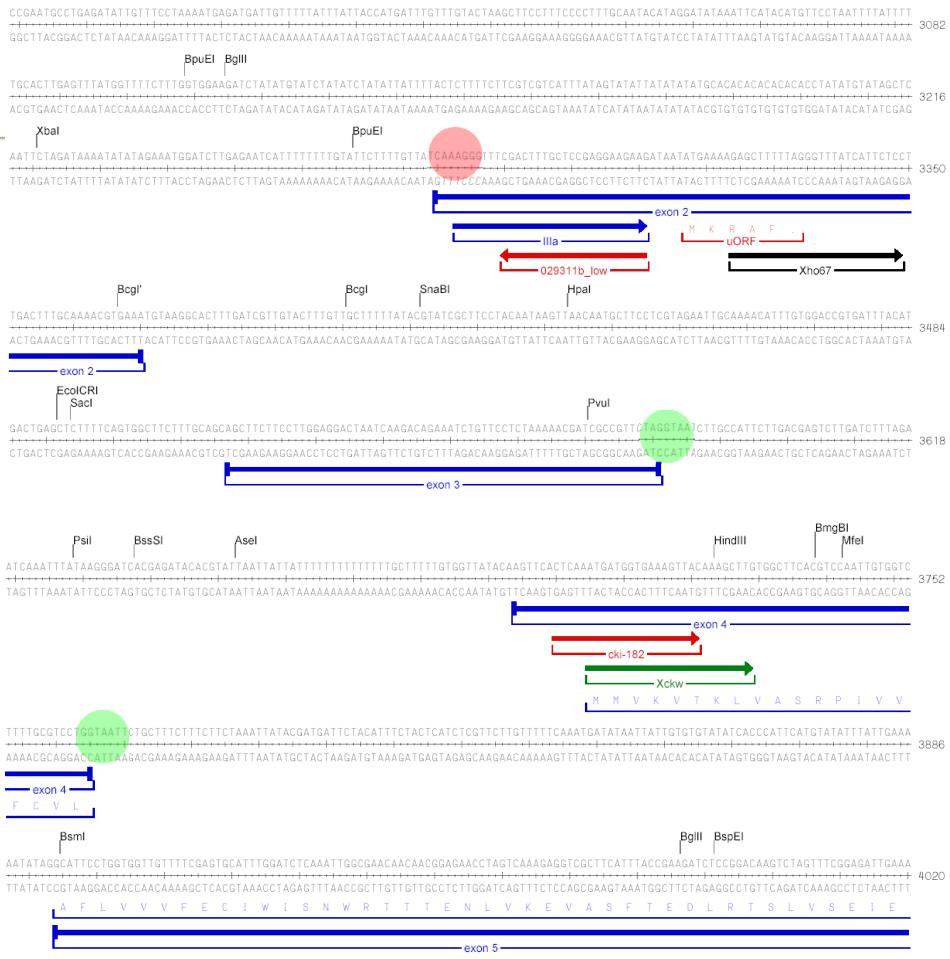
pos	5'→-3'	phase	strand	confidence	5'	exon	intron	3'
1704		0	+	0.87	TTCCAAACAC^	GTAATATT		
1906		0	+	0.99	CGGTGAACGG^	GTCAGAACAT		
3582		1	+	1.00	GCCCTTCTAG^	GTAATCTTCG	H	
3765		1	+	1.00	TTGGCTCTG^	GTAATTCTGC	H	
4134		0	+	0.74	TCAACACAG^	GTTGTTAAAA		
4619		1	+	0.74	AGCAAGAAAG^	GTCCTGTTTC		
4915		0	+	0.94	C GTT CCT CTG^	GTA AAT ACT G		
5356		0	+	0.87	TCTCAACCAA^	GTAATGTTT		
5384		1	+	1.00	GATTGGTG^	GTAAGACTCT	H	
5809		1	+	1.00	TATCTTAAG^	GTG TG TCCA A		
6057		0	+	1.00	GC ACT CTT TG^	GTA AGGACT CT	H	
6096		1	+	0.74	CTCTCACAA^	GTA AAT CTAG		
7369		0	+	1.00	GG AT GC CAA^	GTA AG GT TAA A	H	
7886		0	+	0.74	GAACAAAATG^	GT TAG AT GAA		
9323		0	+	0.74	GAAGATTAGG^	GT TTT CTCT		

#### Donor splice sites, complement strand

pos 3'->5' pos 5'->3' phase strand confidence 5' exon intron 3'

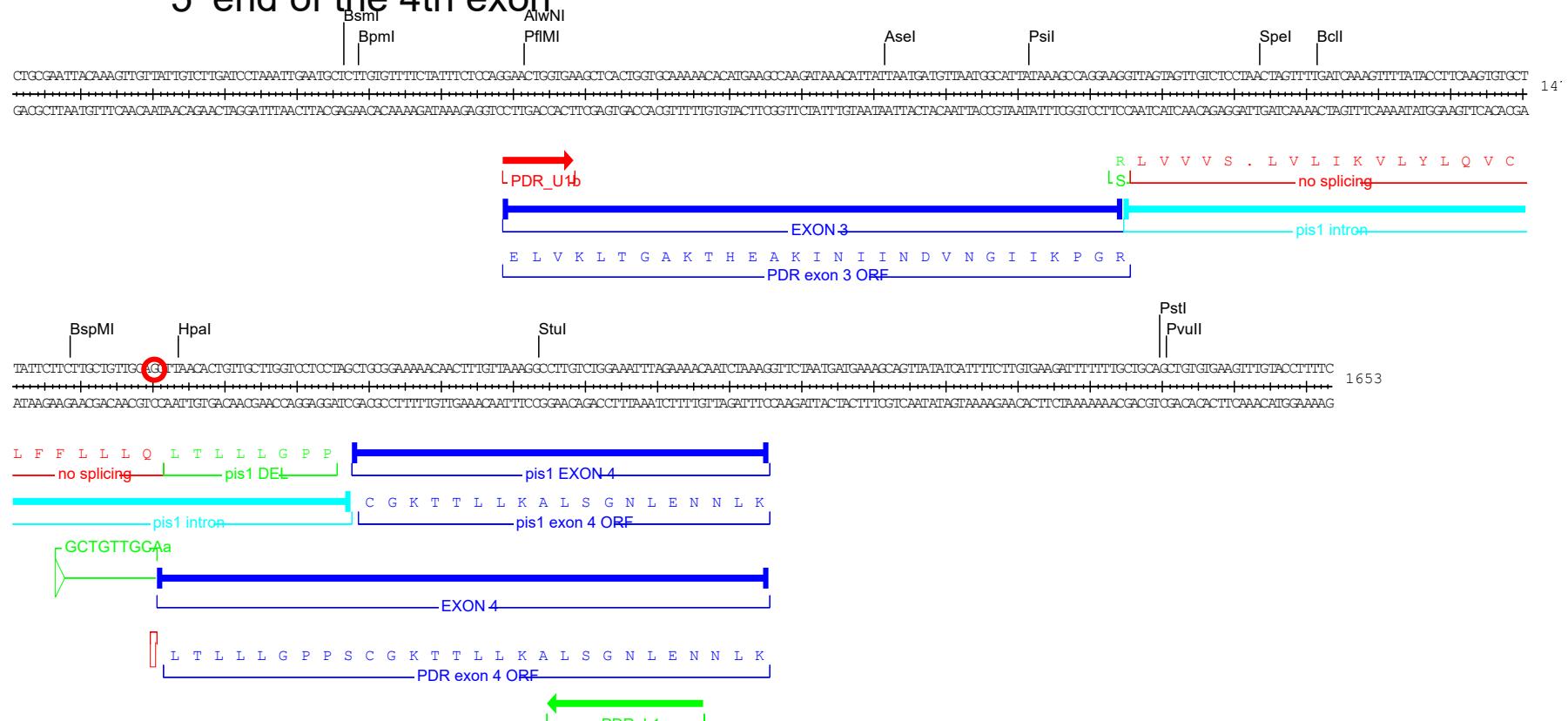
### Acceptor splice sites, direct strand

pos 5'->3'	phase	strand	confidence	5'	intron	exon	3'
1213	0	+	0.59	TATTTTTT	AG	^TTATGGAGAC	
1221	2	+	0.87	AGTTATGGAG	^ACAAGAACATCG		
1373	0	+	0.71	TCTCTCACAG	^GACACAGAAAT		
1487	1	+	0.81	ATATTGATAG	^TGGGACATTA		
3284	0	+	0.87	GTTATCAAAG	^GGTTTCGACT		
4254	0	+	1.00	TGTTCTTCAG	^ATCCGACCAT	H	
4832	2	+	0.54	AAAATTGCAG	^TTCCAGTGGC		
5004	0	+	0.94	TTTTTGCCAG	^AGATACACAC		
5472	1	+	0.96	AAAATTAACAG	^CTCTGCTCAA		
6135	0	+	1.00	ATTATTATAG	^GTAAGATTAA	H	
6490	1	+	0.90	AAAGTTACAG	^TGGTGGAGAA		
6744	0	+	0.59	TGTCAACACAG	^TTTCGTAGAG		
7447	0	+	0.96	TTCTGCACAG	^ATGCCAGAAAA		
7780	2	+	0.76	TCCATTTCAG	^ATACAGAACAA		
7786	2	+	0.92	TCAGATACAG	^AACACATGCA		



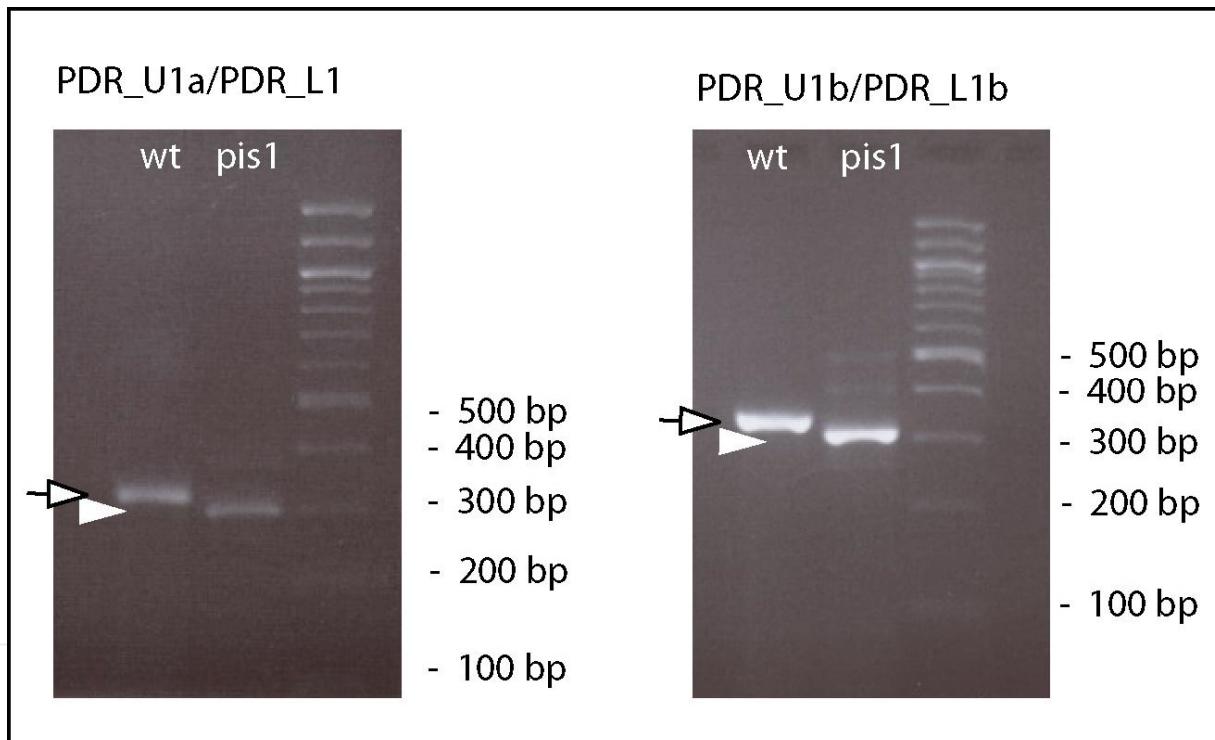
# RNA Splicing and Adaptation

- Flexibility in splicing site recognition in plants in practice – example of developmental plasticity of (not only) plants
  - Identification of mutant with point mutation (transition G→A) exactly at the splice site at the 5' end of the 4th exon



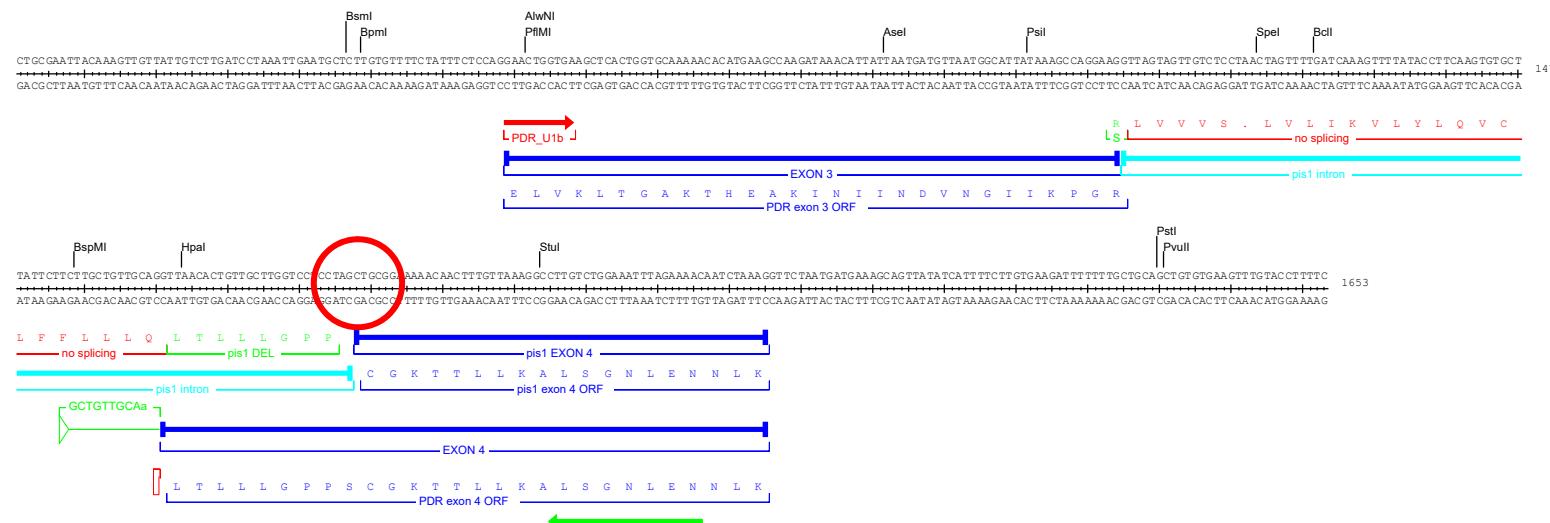
# RNA Splicing and Adaptation

- Identification of mutant with point mutation (transition G→A) exactly at the splice site at the 5' end of the 4th exon
- Analysis by RT PCR proved the presence of a fragment shorter than cDNA should be after the typical splicing event



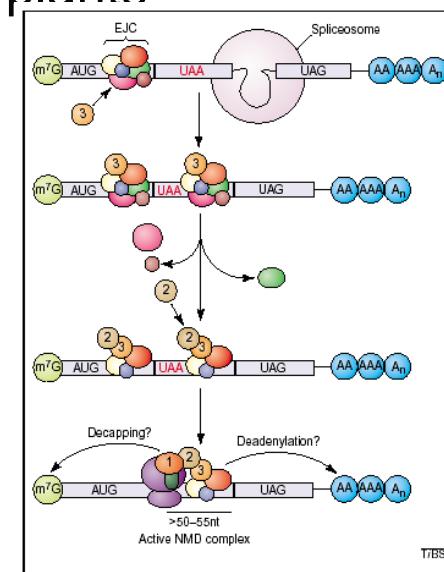
# RNA Splicing and Adaptation

- Flexibility in splicing site recognition in plants in practice – example of developmental plasticity of (not only) plants
  - Identification of mutant with point mutation (transition G→A) exactly at the splice site at the 5' end of the 4th exon
  - Analysis by RT PCR proved the presence of a fragment shorter than cDNA should be after the typical splicing event
  - Sequenation of this fragment then suggested alternative splicing with the closest possible splice site in exon 4



# RNA Splicing and Adaptation

- Divergencies at splice site recognition in plants in practice – example of developmental plasticity of (not only) plants
  - Identification of mutant with point mutation (transition G→A) exactly at the splice site at the 5' end of the 4th exon
  - Analysis by RT PCR proved the presence of a fragment shorter than cDNA should be after the typical splicing event
  - Sequenation of this fragment then suggested alternative splicing with the closest possible splice site in exon 4
  - Existence of similar defense mechanisms was proven in different organisms as well (e.g. Instability of mutant mRNA with early stop codon formation (> 50 - 55 bp before typical stop codon) in eukaryotes, see recommended literature – Singh and Lykke-Andersen, 2003)



INVESTICE DO ROZVOJE VZDĚLÁVÁNÍ

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# Identification of Genes *Ab Initio*

- Programs for exon prediction
  - 4 types of exons (according to location in the gene):
    - initial
    - internal
    - terminal
    - single
  - Programs predict splice sites and they take into account the structure of the type of exon as well
- initial:
  - Genescan (<http://hollywood.mit.edu/GENSCAN.html>)
  - GeneMark.hmm (<http://opal.biology.gatech.edu/GeneMark/>)
- internal:
  - MZEF (<http://rulai.cshl.org/tools/genefinder/>)



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# GENSCAN

**The New GENSCAN Web Server at MIT**

**Identification of complete gene structures in genomic DNA**

For information about Genscan, click here

This server provides access to the program Genscan for predicting the locations and exon-intron structures of genes in genomic sequences from a variety of organisms.

This server can accept sequences up to 1 million base pairs (1 Mbp) in length. If you have trouble with the web server or if you have a large number of sequences to process, request a local copy of the program (see instructions at the bottom of this page) or use the [GENSCAN email server](#). If your browser (e.g., Lynx) does not support file upload or multipart forms, use the [older version](#).

Organism:  Suboptimal exon cutoff (optional):

Sequence name (optional):

Print options: Predicted peptides only

Upload your DNA sequence file (one-letter code, upper or lower case, spaces/numbers ignored):

Or paste your DNA sequence here (one-letter code, upper or lower case, spaces/numbers ignored):  

```
GAGGAGGCCAAATGACGAATACTACAAATGATCTTAAACAGCTAAACTATTTGGACATTTCGATC  
TCAGATATA  
AAAGATTTCATTCAATATAACTTGGATAAATACTCTTATTATTTCTTAGTTATTAAAAAAACCT  
CTAATAAT  
ACGAGTTTAAGTCCAAACCGCTTAGACTAAACACCCATATAATTCAAACGATAAAGTTACAAA  
GTAATATCC  
AAGTATCTCATAGTCACACATATAATAGTAATAATTAGTTGACGTATAAGAAAATAAAATAATT  
GTATCTTAT  
TTTGGGTGGTGCCTGACTGGTACTGGTGACTCGACAATGCTCGGAAATGGAACCATATCCCAAGACATGG  
GTTTTAGAT  
AGAACAAAATAAGTGTCCGAAGGAATGATATTAAAGTCAAATAGAATAATTATAAATTGTAATTAGCA  
AATAAAAAC
```

To have the results mailed to you, enter your email address here (optional):

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# GENSCAN

## GENSCANW output for sequence CKI1

GENSCAN 1.0 Date run: 10-Nov-105 Time: 02:24:26

Sequence CKI1 : 9490 bp : 36.53% C+G : Isochore 1 ( 0 - 43 C+G%)

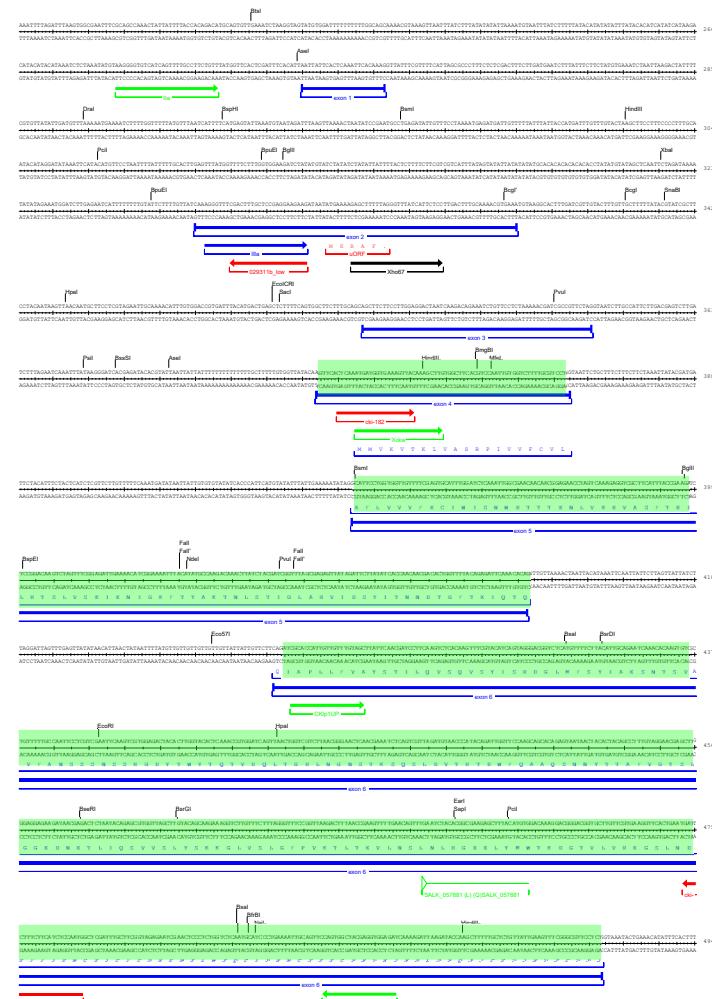
Parameter matrix: Arabidopsis.smat

Predicted genes/exons:

Gn.	Ex	Type	S	.Begin	...End	.Len	Fr	Ph	I/Ac	Do/T	CodRg	P....	Tscr..
1.00	Prom	+	1497	1536	40							-3.85	
1.01	Init	+	3708	3764	57	2	0	63	51	37	0.499	4.03	
1.02	Intr	+	3894	4133	240	2	0	-3	7	327	0.713	17.32	
1.03	Intr	+	4255	4914	660	0	0	86	59	296	0.771	22.57	
1.04	Intr	+	5005	5383	379	0	1	70	91	343	0.772	31.41	
1.05	Intr	+	5473	6056	584	2	2	38	99	582	0.722	50.76	
1.06	Intr	+	6136	7368	1233	0	0	68	108	655	0.977	56.86	
1.07	Term	+	7448	7660	213	1	0	43	35	212	0.999	12.65	
1.08	PlyA	+	7910	7915	6							-0.45	
2.03	PlyA	-	7976	7971	6							-4.83	
2.02	Term	-	8793	8050	744	0	0	107	37	542	0.997	48.46	
2.01	Init	-	9253	8936	318	1	0	105	73	386	0.999	41.18	

Suboptimal exons with probability > 0.100

Exnum	Type	S	.Begin	...End	.Len	Fr	Ph	B/Ac	Do/T	CodRg	P....	Tscr..
S.001	Init	+	1867	1905	39	0	0	64	40	57	0.298	3.74
S.002	Init	+	2374	2442	69	0	0	55	95	-11	0.132	2.40
S.003	Intr	+	3894	4110	217	2	1	-3	-34	307	0.177	11.55
S.004	Intr	+	4352	4914	563	0	2	75	59	338	0.187	26.20
S.005	Intr	+	5005	5379	375	0	0	70	8	335	0.212	22.99
S.006	Intr	+	5442	6056	615	2	0	95	99	589	0.208	57.32

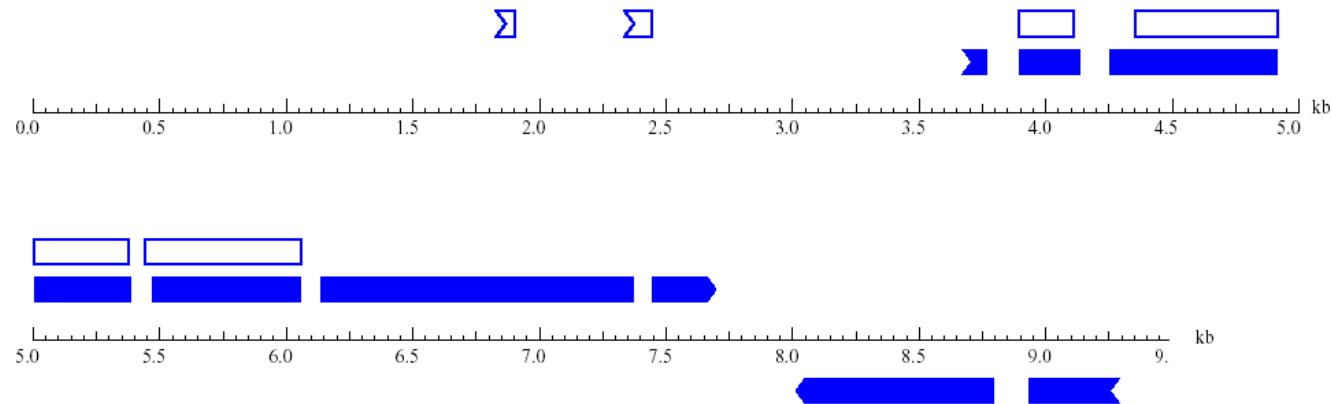


IN VLOŽIT ČIČKO RŮŽOVÝ VLÁDLE ÁVÁNÍ

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# GENSCAN

GENSCAN predicted genes in sequence 02:56:23



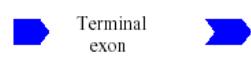
Key:



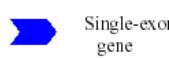
Initial  
exon



Internal  
exon



Terminal  
exon

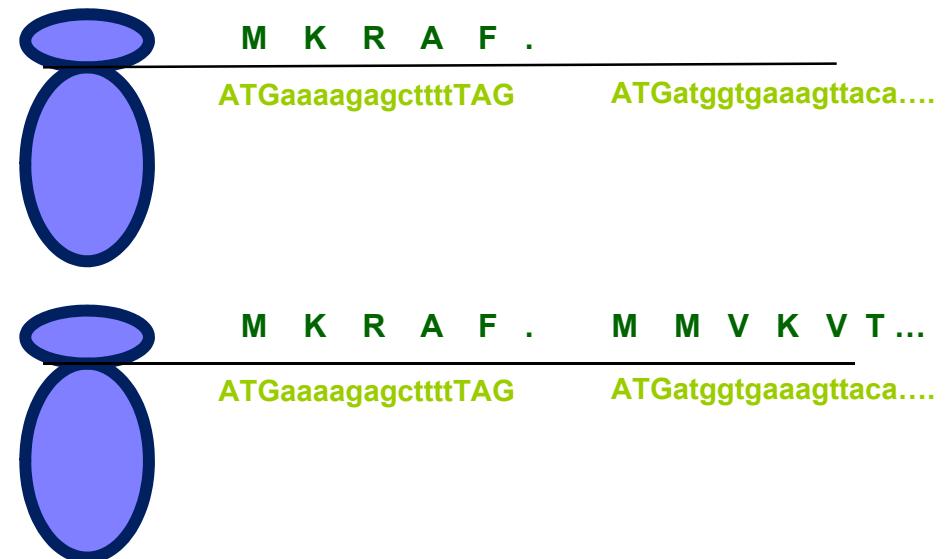


Single-exon  
gene

- Optimal exon
- Suboptimal exon

# Regulation of Translation

- Splicing in Untranslated Regions – important regulation part of genes
- Translational repression by short ORFs in 5' UTR
- Identified e.g. in maize (Wang and Wessler, 1998, see recommended literature for additional info.)
- In case of CKI1 there was an attempt to prove this mechanism of regulation using transgenic lines carrying *uidA* under control of two versions of promoter (unconfirmed so far)

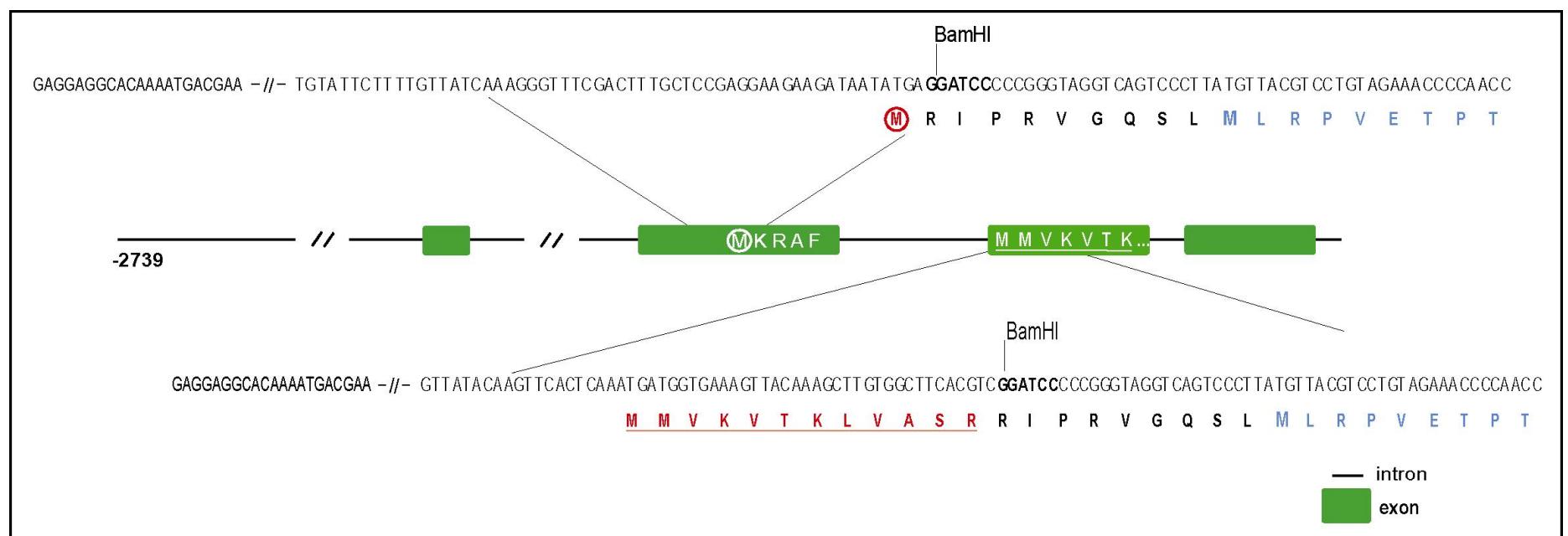


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# Regulation of translation

- Functional purpose of splicing in untranslated regions – important regulation part of genes
- In case of CKI1 there was an attempt to prove this mechanism of regulation using transgenic lines carrying *uidA* under control of two versions of promoter (unconfirmed so far)



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# Gene Modelling

- Programs for gene modelling
  - Those that take into account other parameters as well, e.g. continuity of ORFs
    - Genescan (<http://hollywood.mit.edu/GENSCAN.html>) – very good tool for prediction of exons in coding regions (tested for gene *PDR9*, Genescan identified all of the 23 (!) exons)
    - GeneMark.hmm (<http://opal.biology.gatech.edu/GeneMark/>)
    - GlimmerHMM (<https://ccb.jhu.edu/software/glimmerhmm/>)



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# GeneMark

### **Result of last submission:**

### [View PDF Graphical Output](#)

## GeneMark hmmp Listing

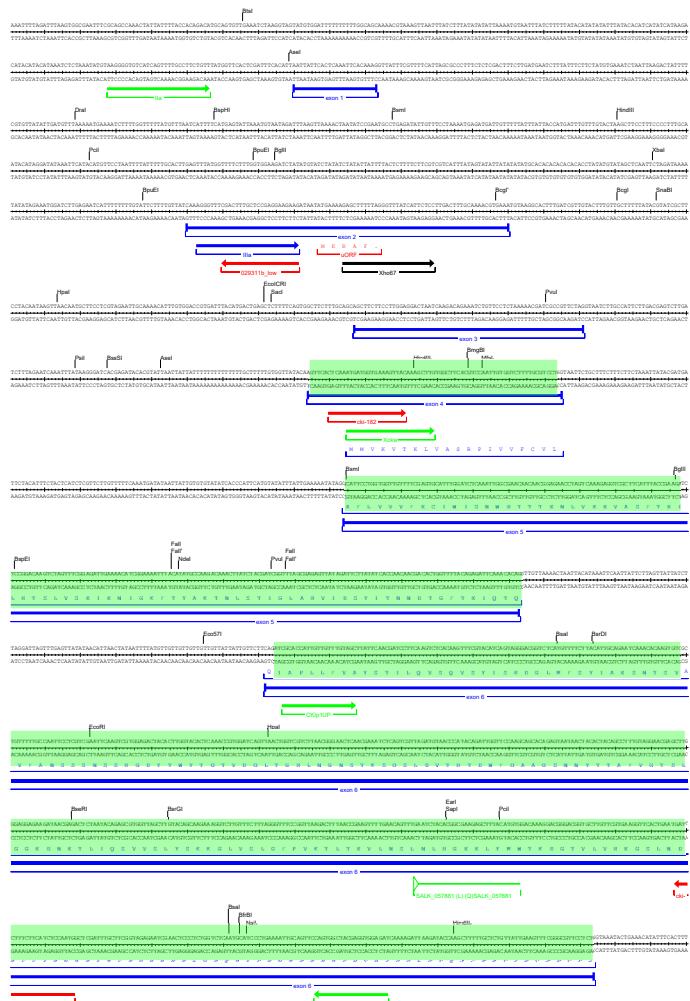
Go to: [GeneMark.hmm Protein Translations](#)

[Go to: Job Submission](#)

```
Eukaryotic GeneMark.hmm version bp 3.9 April 25, 2008
Sequence name: CKII1
Sequence length: 5043 bp
G+C content: 38.79%
Matrices file: /home/germark/euk_ghm.matrices/athaliana_hmm3.0.mod
Thu Oct 1 11:09:24 2009
```

### Predicted genes/exons

Gene	Exon	Strand	Exon	Exon Range	Exon Length	Start/End Frame
#	#		Type			
1	1	+	Initial	969 1025	57 1 3 - -	
1	2	+	Internal	1155	1394	240
1	3	+	Internal	1516	2175	660
1	4	+	Internal	2256	2544	379
1	5	+	Internal	2734	3017	584
1	6	+	Internal	3397	4529	1233
1	7	+	Terminal	4709	4921	213



/ZDĚLÁVÁNÍ

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# GeneMark

## Result of last submission:

[View PDF Graphical Output](#)

[GeneMark.hmm Listing](#)

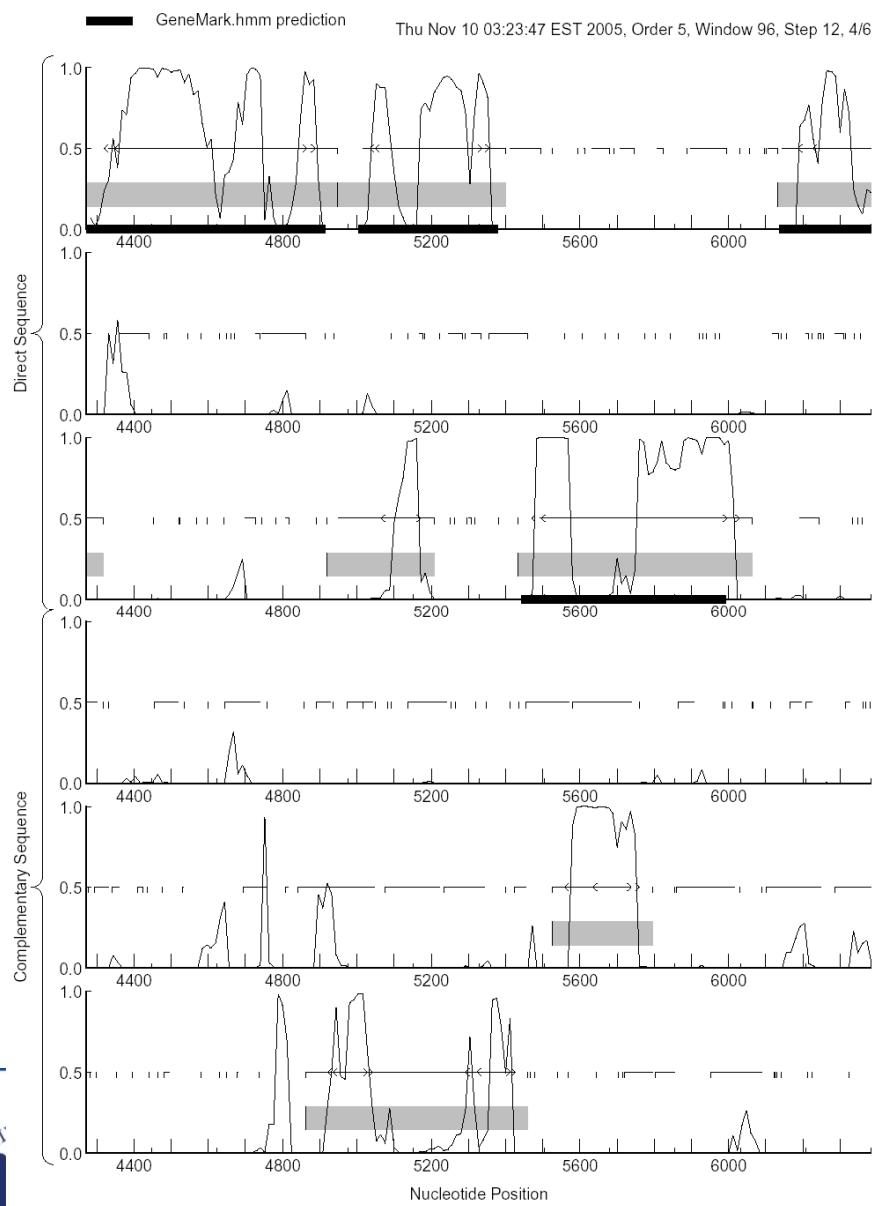
[Go to: GeneMark.hmm Protein Translations](#)

[Go to: Job Submission](#)

Bukariotyc GeneMark.hmm version bp 3.9 April 25, 2008  
Sequence name: CK11  
Sequence length: 5043 bp  
G+C content: 38.79%  
Matrices file: /home/genmark/euk\_ghm.matrices/athaliana\_hmm3.0.mod  
Thu Oct 1 11:09:24 2009

Predicted genes/exons

Gene #	Exon #	Strand	Exon Type	Exon Range	Exon Length	Start/End Frame
1	1	+	Initial	969	1025	57 1 3 - -
1	2	+	Internal	1155	1394	240 1 3 - -
1	3	+	Internal	1515	2175	660 1 3 - -
1	4	+	Internal	2266	2644	379 1 1 - -
1	5	+	Internal	2734	3317	584 2 3 - -
1	6	+	Internal	3397	4629	1233 1 3 - -
1	7	+	Terminal	4709	4921	213 1 3 - -



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# Genomic Homologies

- Searching for genes according to homologies with known sequences
  - Comparison with EST databases
    - BLASTN (<http://www.ncbi.nlm.nih.gov/BLAST/>, <http://workbench.sdsc.edu/>)
  - Comparison with protein databases
    - BLASTX (<http://www.ncbi.nlm.nih.gov/BLAST/>, <http://workbench.sdsc.edu/>)
    - Genewise (<http://www.ebi.ac.uk/Wise2/>)

They compare protein sequence with genomic DNA (after reverse transcription), therefore the aminoacid sequence is needed
  - Comparison with homologous genome sequences from related species
    - VISTA/AVID (<http://www.lbl.gov/Tech-Transfer/techs/lbnl1690.html>)



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# Outline

- Forward and Reverse Genetics Approaches
  - Differences between the approaches used for identification of genes and their function
- Identification of Genes *Ab Initio*
  - Structure of genes and searching for them
  - Genomic colinearity and genomic homology



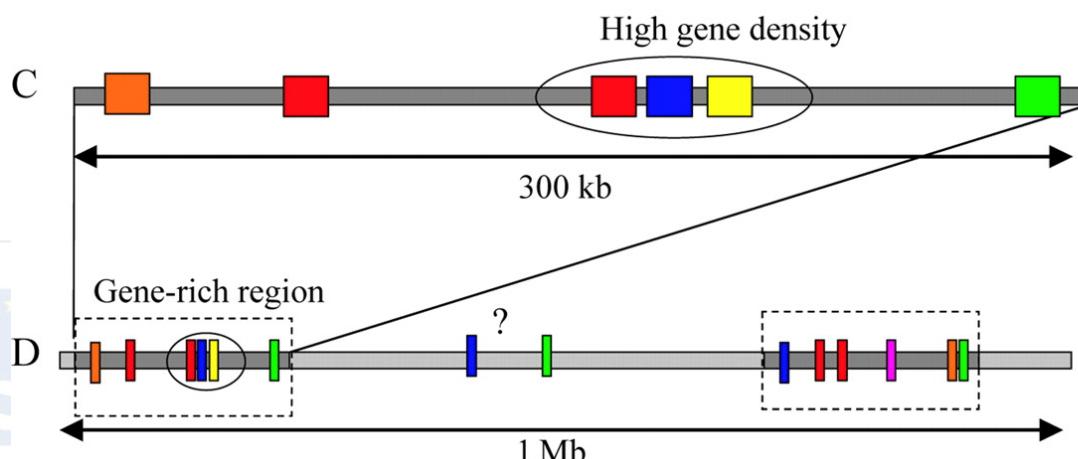
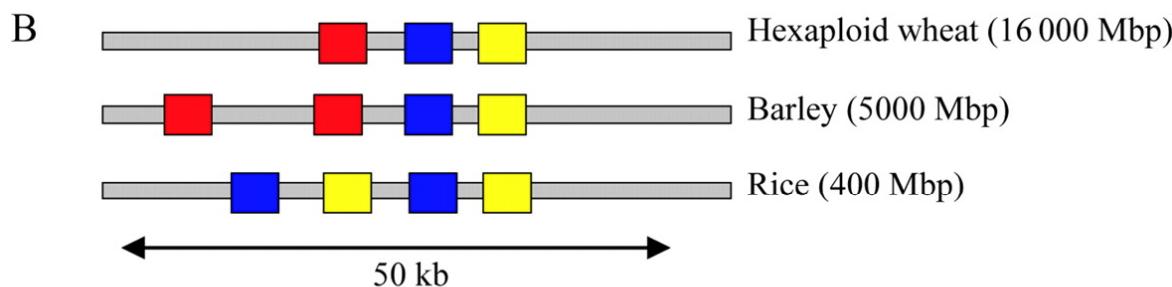
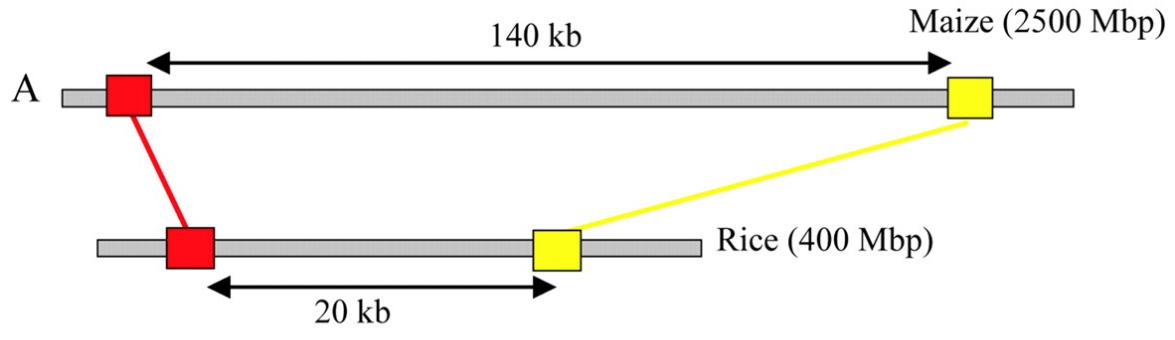
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# Genomic Colinearity

- Genomes of related species (despite large differences) are characterized by similarities in sequence organization -> possibility to use this information for identification of genes in related species when searching in databases
- General scheme of work while applying genomic colinearity (also called „comparative genomics“) for experimental identification of genes in related species:
  - Mapping small genomes using low-copy DNA markers (e.g. RFLP)
  - Using these markers for identification of orthologous genes (genes with the same or similar function) of related species
  - Small genome (e.g. rice, 466 Mbp) can be used as a guide: molecular low-copy markers (e.g. RFLP) bound to gene of interest are identified and these regions are then used as a probe for searching in BAC libraries during identification of orthologous regions of large genomes (e.g. barley: 5 Gbp, or wheat: 16 Gbp)

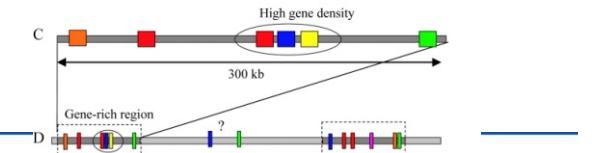
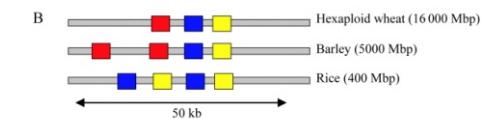
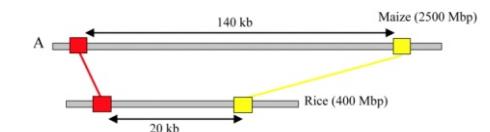
# Genomic Colinearity



Feuillet and Keller, 2002

# Genomic Colinearity

- Can be mostly used for the species of grass (e.g. using related genes of species of barely, wheat, rice, maize)
- Small genome reorganizations (deletions, duplications, inversions, translocations smaller than a few cM) are then detected by detailed sequential comparative analysis
- During evolution there's occurred some divergencies in related species, mostly in non-coding regions (invasion of retrotransposons etc.)



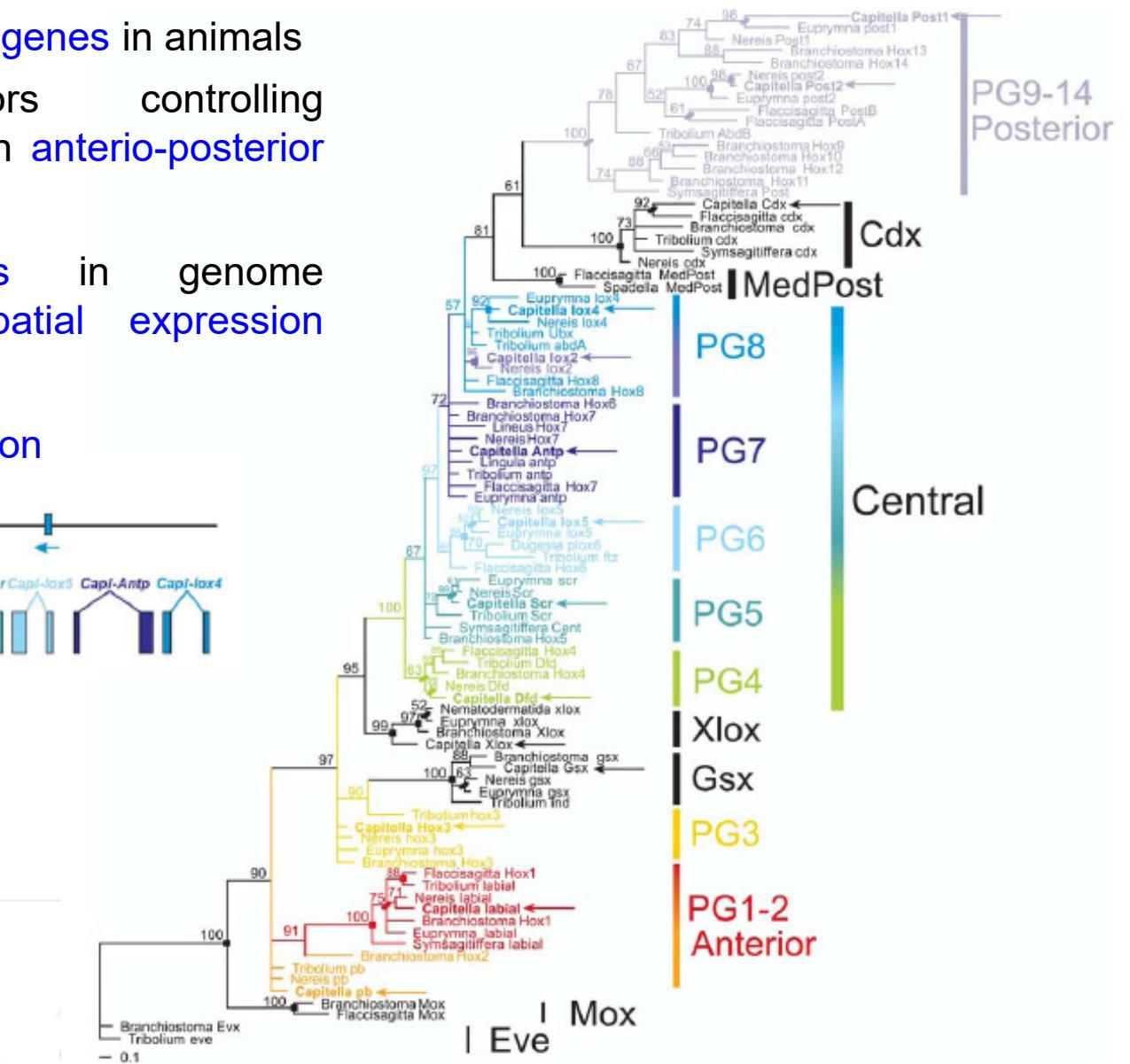
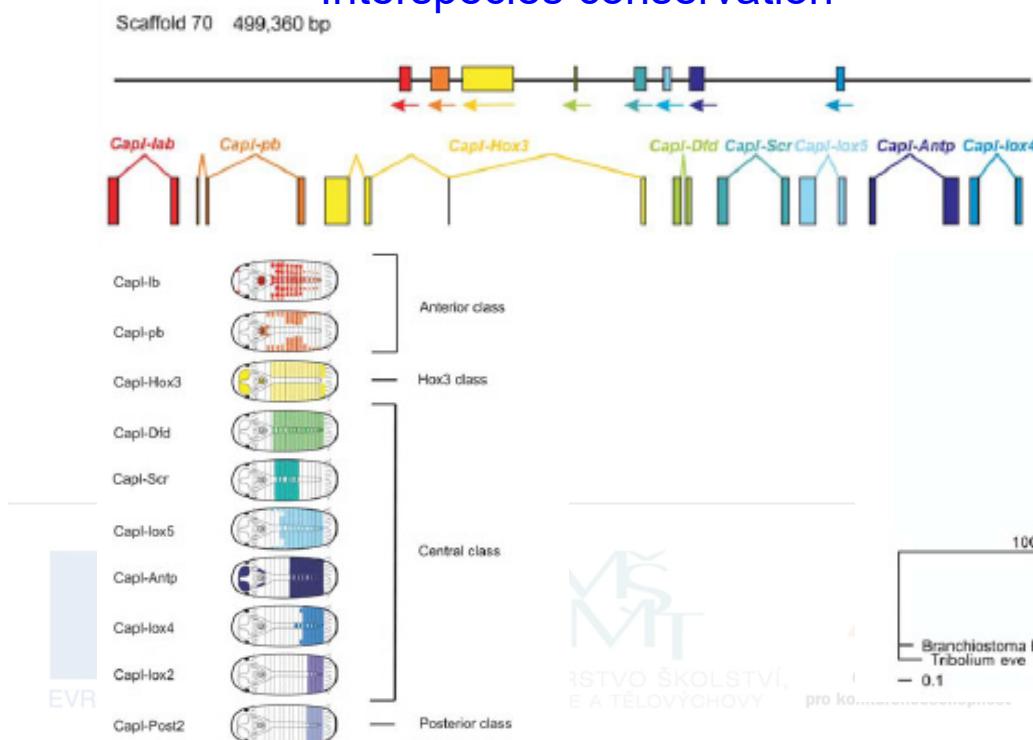
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# Genomic Colinearity

- Genomic colinearity of HOX genes in animals
    - Transcription factors controlling organisation of body in antero-posterior axis
    - Position of genes in genome corresponds with spatial expression during development
    - Interspecies conservation



# Outline

- Forward and Reverse Genetics Approaches
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  - Genomic colinearity and genomic homology
- Experimental Genes Identification
  - Constructing gene-enriched libraries using methylation filtration technology



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# Methylation Filtration

- Preparation of gene-enriched libraries by technology of methylation filtration
- genes are (mostly!) hypomethylated, noncoding regions are methylated
- using bacterial restriction-modification system, which recognizes methylated DNA with restriction enzymes McrA a McrBC
  - McrBC recognizes methylated cytosin (in DNA), which comes after purine (G or A)
  - For cleavage the distance of these sites 40-2000 bp is necessary



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# Methylation Filtration

- Preparation of gene-enriched libraries by technology of methylation filtration
- Scheme of work during preparation of BAC genome libraries using methylation filtration:
  - preparation of genomic DNA without addition of organelle DNA (chloroplasts and mitochondria)
  - fragmentation of DNA (1-4 kbp) and ligation of adaptors
  - preparation of BAC libraries in *mcrBC+* strain of *E. coli*
  - selection of positive clones
- Limited usage: enrichment of coding DNA only approx. 5 -10 %



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  - EST libraries



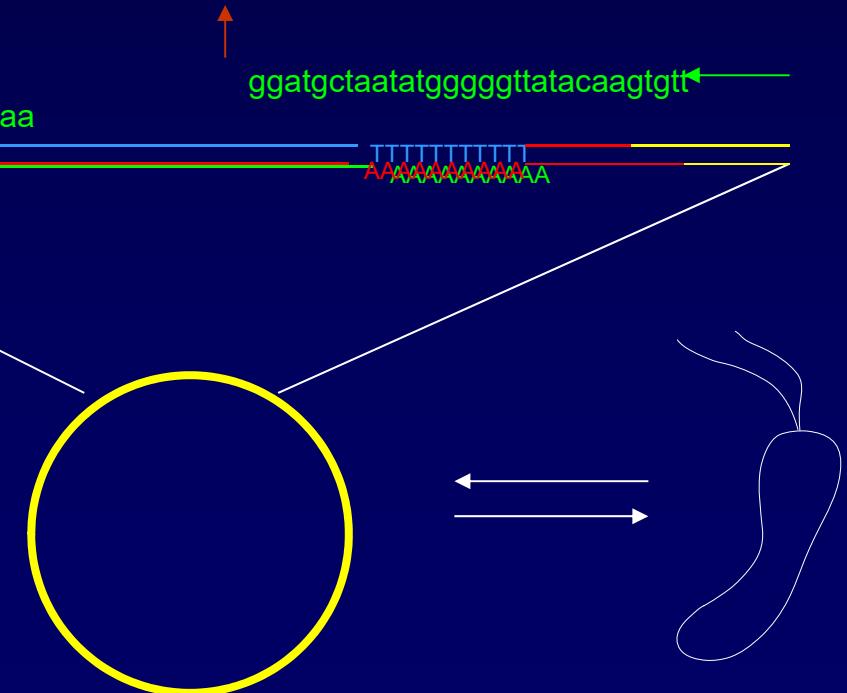
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# EST Libraries

- Preparation of EST libraries
  - Isolation of mRNA
  - Reverse transcription
  - Ligation of linkers and synthesis of second cDNA strand
  - Cloning into suitable bacterial vector
  - Transformation into bacteria and isolation of DNA →  
cctacgattataccccc当地址  
(amplification of DNA)
  - Sequencing using primers specific for used plasmid
  - Saving the results of sequencing into public database

The screenshot shows a search result for NC\_002377, which is the Agrobacterium tumefaciens Ti plasmid. The page displays the full sequence (2495 bp), its linear nature, and its complete status. It includes details like the source (Agrobacterium tumefaciens), version (NC\_002377.1), and references to scientific publications. The sequence itself is shown in a standard DNA sequence format.



# Outline

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  - Genomic colinearity and genomic homology
- Experimental Genes Identification
  - Constructing gene-enriched libraries using methylation filtration technology
  - EST libraries
  - Forward and reverse genetics



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# Discussion



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