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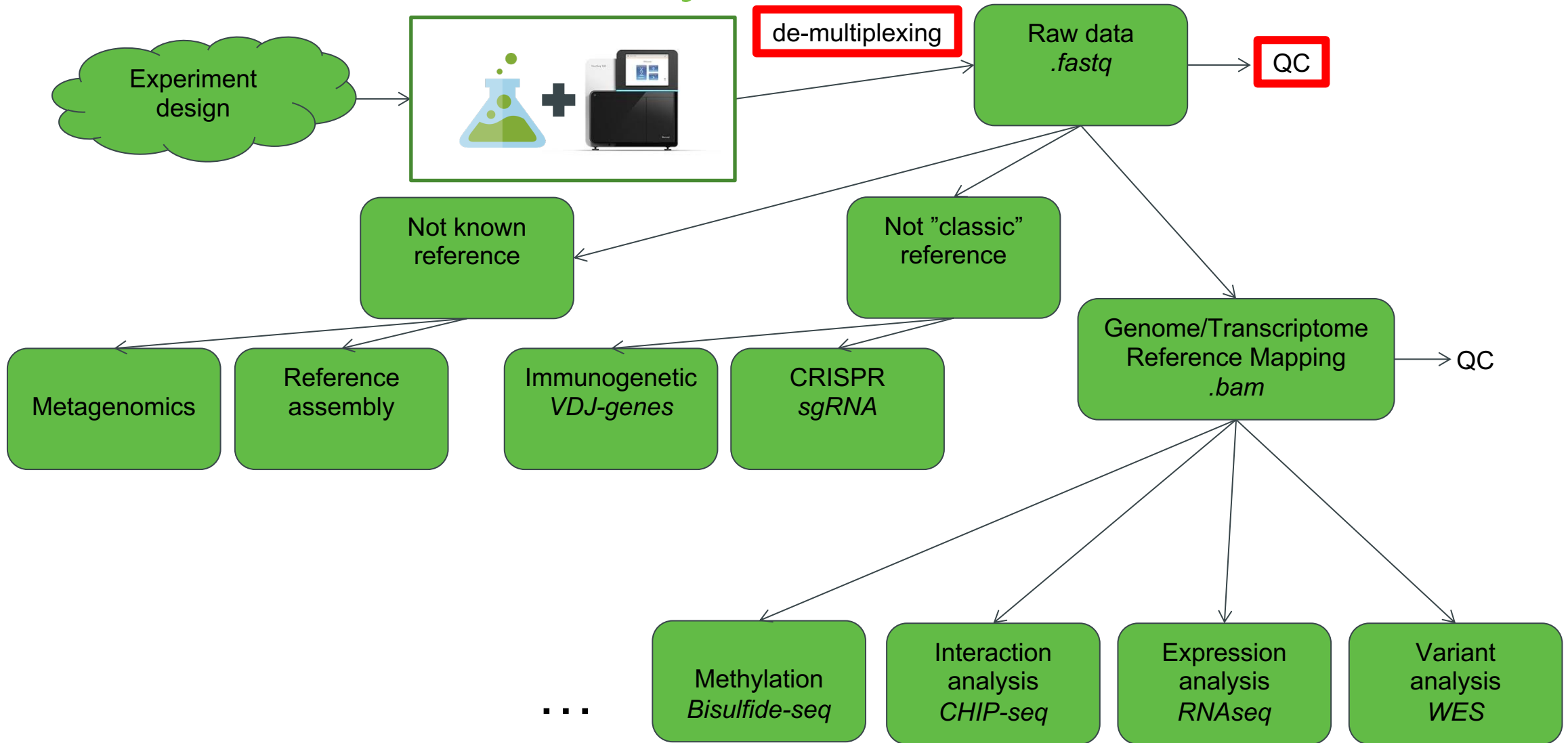
Central European Institute of Technology
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**Modern methods for genome analysis
(PřF:Bi7420)**

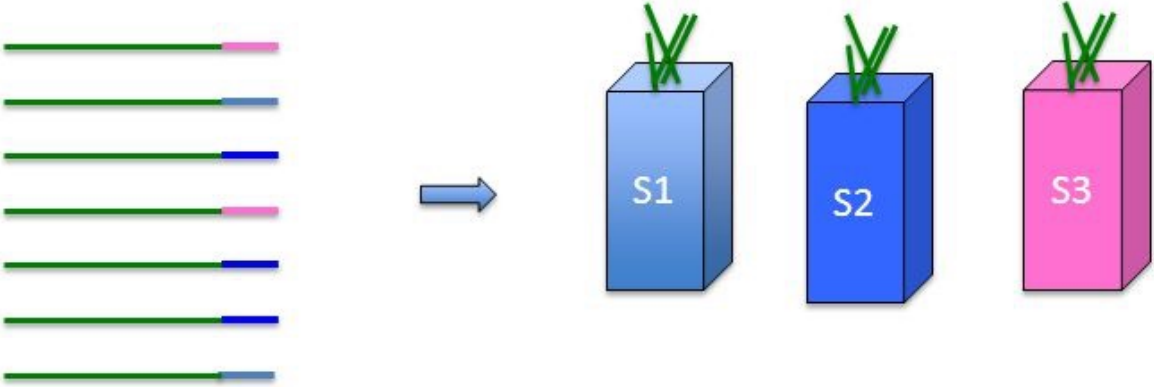
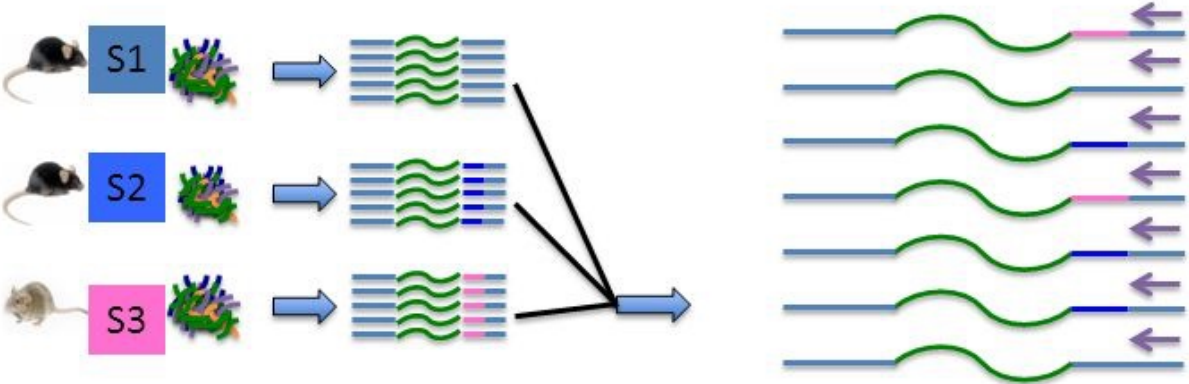
Lecture 2 : Raw NGS data quality control

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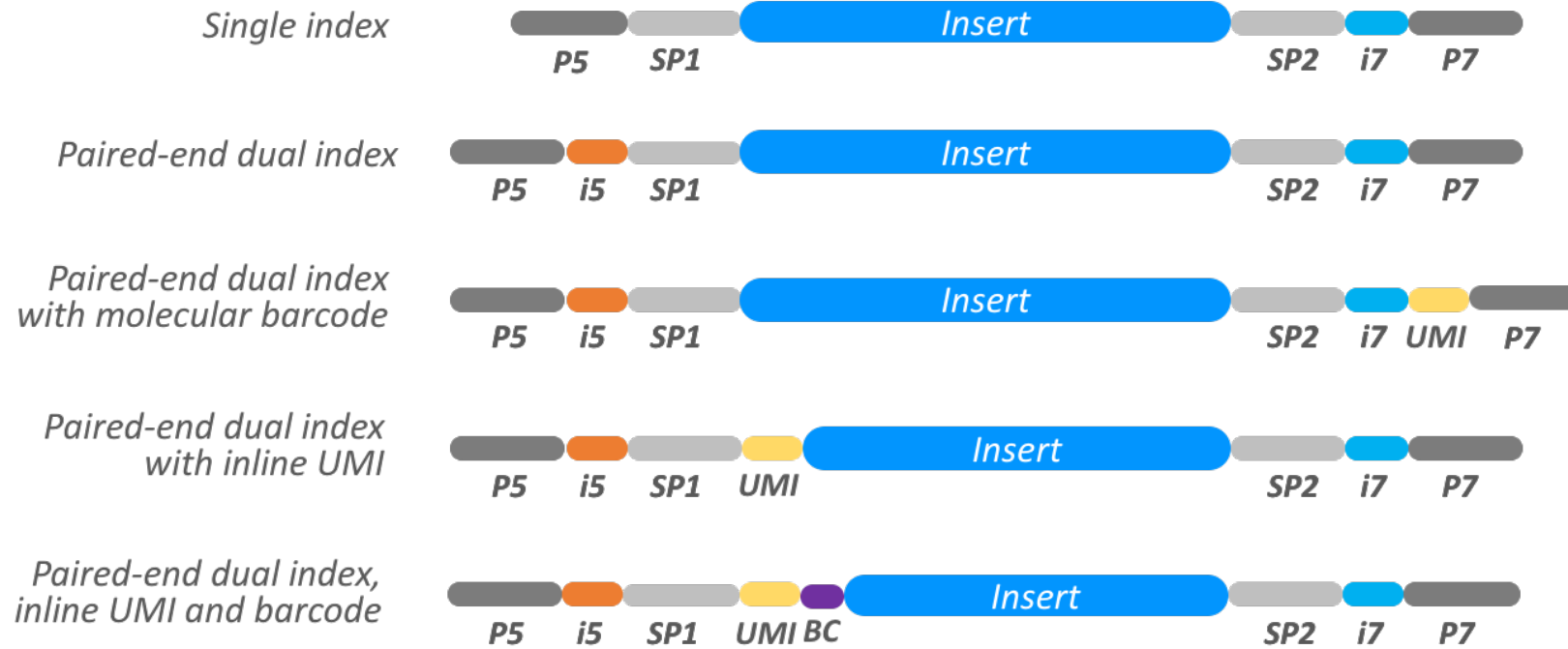
NGS data analysis



De-multiplexing

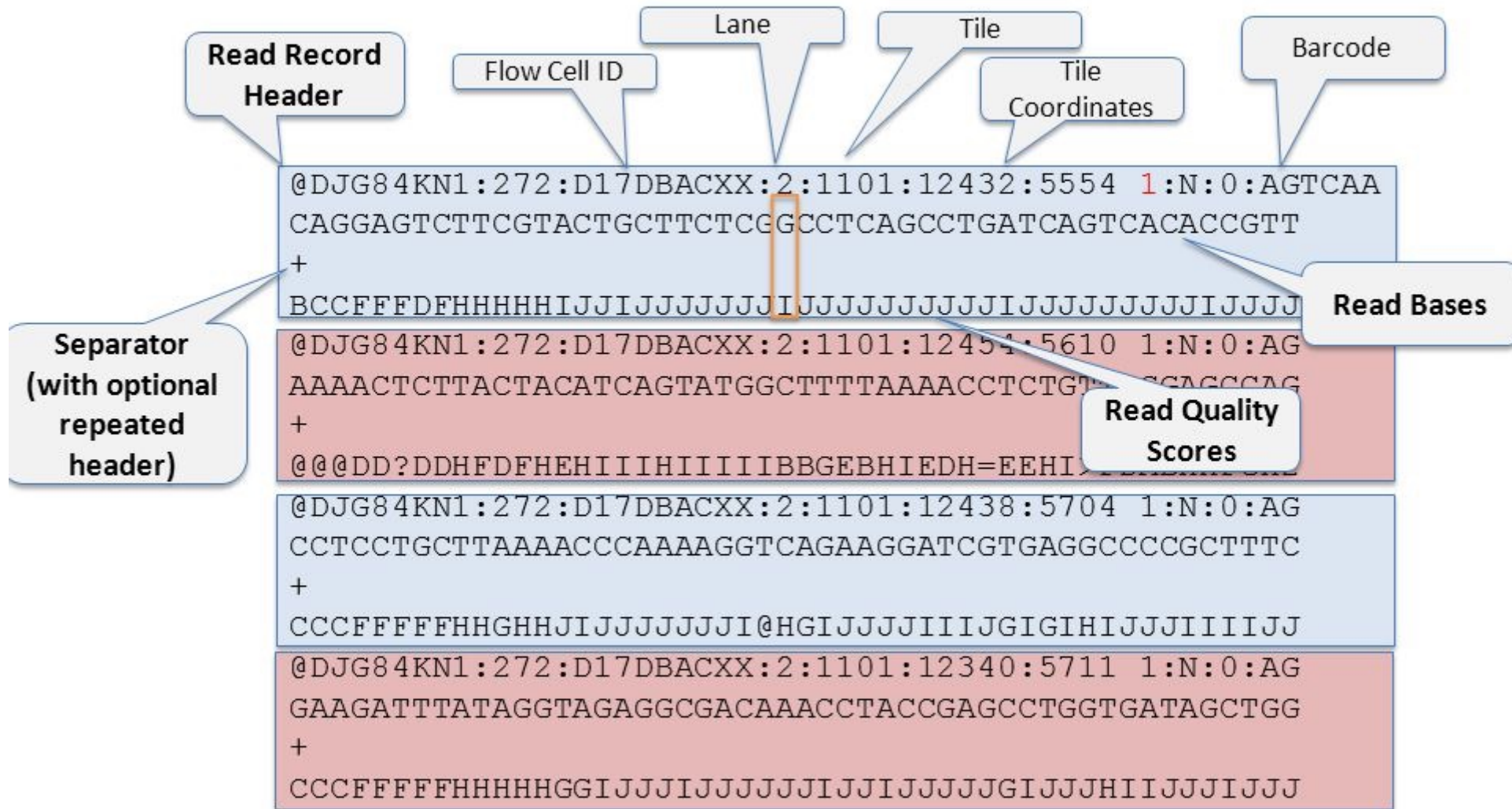


De-multiplexing



- P5/P7:** Flow cell binding sequences (platform-specific)
- SP1/2:** Sequencing primer binding sites (common for all libraries)
- i5/i7:** Sample Indexes (specific to a particular library)
- UMI:** Unique molecular index (barcode tag for individual molecules)
- BC:** User-defined barcode (unique per sample, single cell, etc.)
- Insert:** Target DNA or cDNA fragment (library-specific)

Primary data – fastq file



NOTE: for paired-end runs, there is a second file with one-to-one corresponding headers and reads.

Fastq format - quality

- Fastq - q stands for quality – coded as phred score

CGGGGEEGCGGGGGAFF87@E:++6C<++3:,8,33,,,:,,,::,,:

$$Q = -10 \cdot \log_{10} P$$

Quality	Error probability
5	31%
10	10%
20	1%
30	0.1%

- What the machine thinks is the quality
- Only account for sequencing errors
- Very good for early problem detection

Fastq – quality control

- How can we summarize this?
- What QC can be done?

```
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FastQC Report

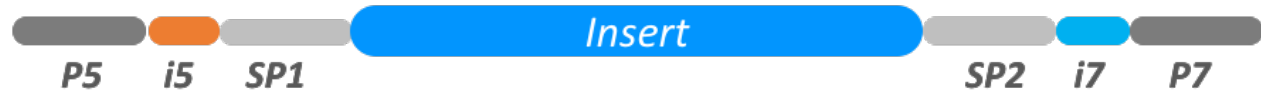
Summary

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- ✓ [Per base sequence quality](#)
- ✓ [Per tile sequence quality](#)
- ✓ [Per sequence quality scores](#)
- ! [Per base sequence content](#)
- ! [Per sequence GC content](#)
- ✓ [Per base N content](#)
- ✓ [Sequence Length Distribution](#)
- ✓ [Sequence Duplication Levels](#)
- ✓ [Overrepresented sequences](#)
- ✓ [Adapter Content](#)

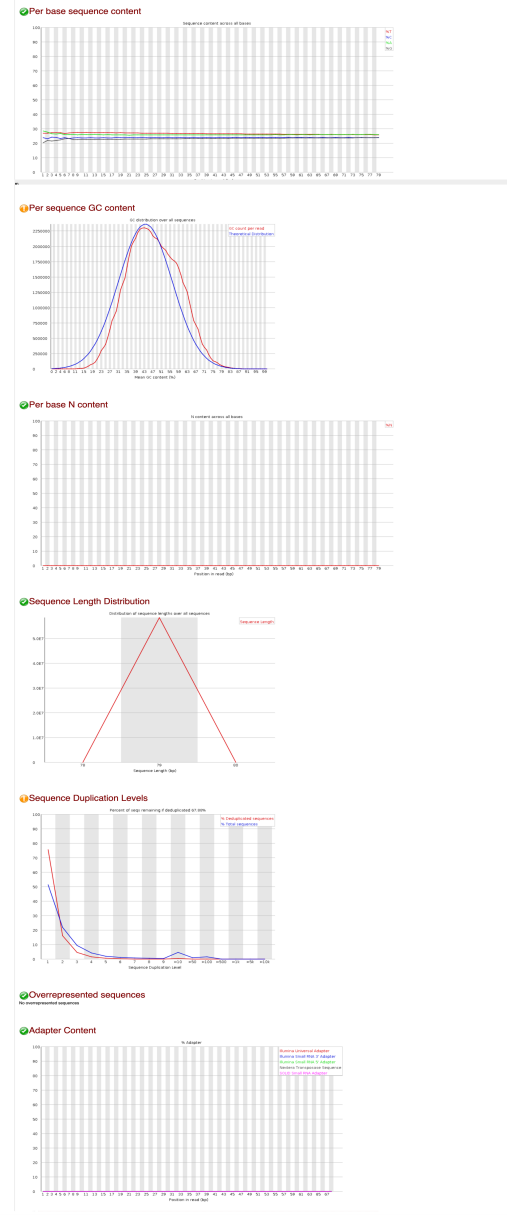
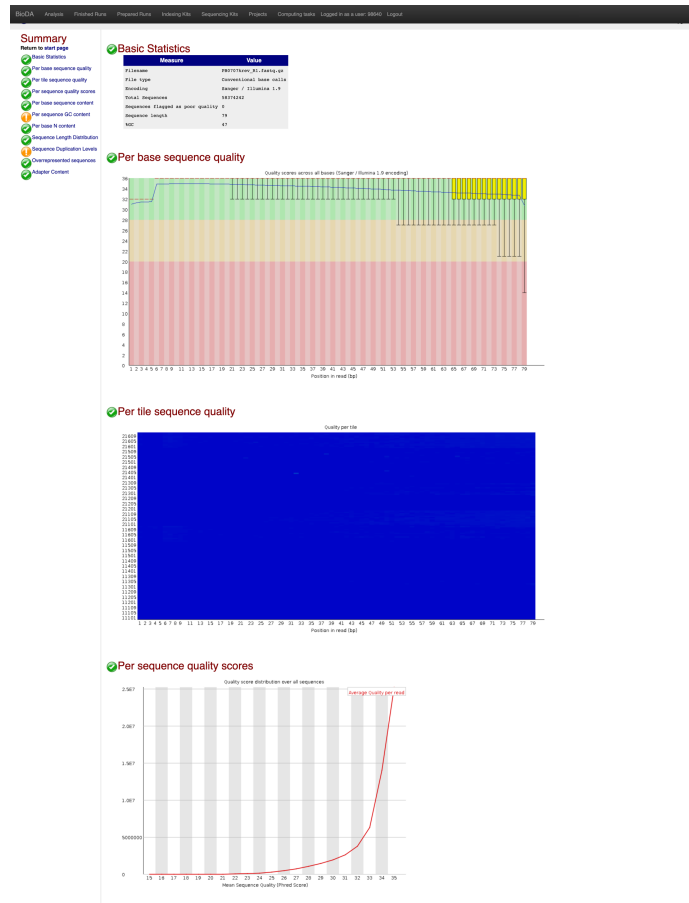
✓ Basic Statistics

Measure	Value
Filename	MU_a_ytHl_R1.fastq.gz
File type	Conventional base calls
Encoding	Sanger / Illumina 1.9
Total Sequences	252819865
Sequences flagged as poor quality	0
Sequence length	161
%GC	40



Fastq – quality control

- Fastqc - tool





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Thank you for your attention!

