



Outline
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<ul> <li>Preparation of transcriptional fusion of promoter of analysed gene with a reporter gene</li> </ul>
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## GUS Reporter in Mouse Embryos









Histone 2A-GFP in Drosophila embryo by PAM



















Microarray expression profiles of 19 fluorescently sorted GFP-marked lines were analyzed (3–9, 23, 24). The colors associated with each marker line reflect the developmental stage and cell types sampled. Thirteen transverse sections were sampled along the root's longitudinal axis (red lines) (10). CC, companion cells.



(A) The majority of enriched GO terms (hierarchically clustered) are associated with individual cell types (blue bar). A smaller number are present across multiple cell types (green bar). (fig. S2) (**B**) GO category enrichment for hair cells confirms a previous report (15). Enriched cis-elements and an enriched TF family were also identified. ( $\mathbf{C}$ ) From the top 50% of varying probe sets, 51 dominant radial patterns were identified. Pattern expression values were mean-normalized (rows) and log<sub>2</sub> transformed to yield relative expression indices for each marker line (columns). Marker line order is the same for all figures; see table S1 for marker line abbreviations. (**D**) Pattern expression peaks were found across one to five cell types. (**E** to **G**) Patterns where expression is enriched in single and multiple cell types support transcriptional regulation of auxin flux and synthesis. In all heat maps with probe sets, expression values were mean-normalized and log<sub>2</sub> transformed. Expression is false-colored in representations of a root transverse section, a cut-away of a root tip, and in a lateral root primordium. (E) Auxin biosynthetic genes (CYP79B2, CYP79B3, SUPERROOT1, and SUPERROOT2) are transcriptionally enriched in the QC, lateral root primordia, pericycle, and phloem-pole pericycle ( $P = 1.99E^{-11}$ , pattern 5). All AGI identifiers and TAIR descriptions are found in table S14. (F) Auxin amido-synthases GH3.6 and GH3.17 that play a role in auxin homeostasis show enriched expression in the columella, just below the predicted auxin biosynthetic center of the QC (P =8.82E<sup>-4</sup>, pattern 13). (G) The expression of the auxin transporter, *PIN*-FORMED2, and auxin transport regulators (PINOID, WAG1) are enriched in the columella, hair cells, and cortex ( $P = 1.03E^{-4}$ , pattern 31).



## Deconstructing and reconstructing the embryo by single-cell transcriptomics combined with spatial mapping.

(A) Single-cell sequencing of the Drosophila embryo: ~1000 handpicked stage 6 fly embryos are dissociated per Drop-seq replicate, cells are fixed and counted, single cells are combined with barcoded capture beads, and libraries are prepared and sequenced. Finally, single-cell transcriptomes are deconvolved, resulting in a digital gene expression matrix for further analysis.

**(B)** Mapping cells back to the embryo: Single-cell transcriptomes are correlated with high-resolution gene expression patterns across 84 marker genes, cells are mapped to positions within a virtual embryo, and expression patterns are computed by combining the mapping probabilities with the expression levels (virtual in situ hybridization).



Schematic flowchart of the Human Protein Atlas. For each gene, a signature sequence (PrEST) is defined from the human genome sequence, and following RT-PCR, cloning and production of recombinant protein fragments, subsequent immunization and affinity purification of antisera results inmunospecific antibodies. The produced antibodies are tested and validated in various immunoassays. Approved antibodies are used for protein profiling in cells (immunofluorescence) and tissues (immunohistochemistry) to generate the images and protein expression data that are presented in the Human Protein Atlas (Ponten et al., *J Int Med*, 2011).























	R Bi	esults of ological Transcriptional regulated gene	f –o Iy R profiling	m le	ic le'	S Va	Si an mor	tuc t C	lies Conc en 7к	VS Clu diffe	S Sic erent	<b>DN</b> tially	S
gene	•		locus	sample 1	sample 2	status	value 1	value 2	log2(fold_change)	test stat	n value	a value	significant
AT1C07795			1-2414285-2414967	wT	MT	OK	value_!	1 1804	1 70760e+308	1.79769e+3	6 888850-05	0,00039180	Vee
HPS1			1:4555901 4559709	WT	мт	or		0 606592	1 707600+209	1.79769e+3	6 610040 06	4.67708e-	100
HRSI			1.4556691-4556708	vv i		UK .		0,090505	1.797098#300	1.79769e+3	0.019948-00	0,00053505	yes
ATMLO14			1:9227472-9232296	WT	мт	ок	(	0,514609	1.79769e+308	08 1.79769e+3	9.74219e-05	5 3.50131e-	yes
NRT1.6			1:9400663-9403789	WT	мт	ок	(	0,877865	1.79769e+308	08 1 79769e+3	3.2692e-08	07	yes
AT1G27570			1:9575425-9582376	WT	мт	ок	(	2,0829	1.79769e+308	08	9.76039e-06	6.647e-05	yes
AT1G60095			1:22159735-22162419	WT	мт	ок	(	0,688588	1.79769e+308	08	9.95901e-08	07	yes
AT1G03020			1:698206-698515	WT	мт	ок		1,78859	1.79769e+308	1.79769e+3 08	0,00913915	0,0277958	yes
AT1G13609			1:4662720-4663471	WT	мт	ок		3,55814	1.79769e+308	1.79769e+3 08	0,00021683	3 0,00108079	yes
AT1G21550			1:7553100-7553876	wт	мт	ок		0,562868	1.79769e+308	1.79769e+3 08	0,00115582	2 0,00471497	ves
AT1G22120			1-7806308-7809632	WT	мт	OK		0.617354	1 70760e+308	1.79769e+3	2 483020-06	1.91089e-	Vee
AT1022120			4.44000007 4400002	wr		or		4 40054	4 70700- 200	1.79769e+3	4.00500-05	0,00028514	,
ALIGNO				WT.		or		0.504001	4 70700- 200	1.79769e+3	7.07055-00	5.46603e-	,
APON10			1:13233397-13255570	VV I	MI	UK		0,581031	1.7970987308	1.79769e+3	1.010058-06	0,00037473	yes.
AT1G48700			1:18010728-18012871	WT	мт	ок	0	0,556525	1.79769e+308	08 1.79769e+3	6.53917e-05	6	yes
AT1G59077			1:21746209-21833195	WT	мт	ок	0	138,886	1.79769e+308	08 1.79769e+3	0,00122789	0,00496816	yes
AT1G60050			1:22121549-22123702	WT	мт	ок	(	0,370087	1.79769e+308	08	0,00117953	0,0048001	yes
AT4G15242			4:8705786-8706997	WT	мт	ок	0,00930712	17,9056	10,9098	-4,40523	1.05673e-05	7.13983e-05	yes
AT5G33251			5:12499071-12500433	WT	мт	ок	0,0498375	52,2837	10,0349	-9,8119	c	i	0 yes
AT4G12520			1:7421055-7421738	WT	MT	ок	0,019511	15,8516	9,66612	-3,90043	9.60217e-05	0,00052890	4 yes
AT1G60020			1:22100651-22105276	WT WT	MT	OK	0,011837	7,18823	9,24611 9 1587	-7,50382	6.19504e-14	1.4988e-12	yes 0 ves

Excample of an output of transcriptional profiling study using Illumina sequencing performed in our lab. Shown is just a tiny fragment of the complete list, copmprising about 7K genes revealing differential expression in the studied mutant.









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In the figure, there is simplified scheme of vesicle trafficking pathways, regulated by GNOM and its closest relative, GNOM-LIKE1 (GNL1).

Secretory and membrane proteins are synthesised at the ER (blue) and passed onto the Golgi apparatus (green) by anterograde trafficking in COPII-coated vesicles.

The retrograde route from the Golgi apparatus to the ER is regulated by the ARF-GEFs GNOM (GN) and GNL1, which regulate the recruitment of COPI coats to the Golgi membrane. On the secretory route, proteins are transported to the sorting station, the trans-Golgi network (TGN; lilac).

From there, proteins are either transported to the vacuole (grey) via multivesicular bodies (MVB, also called prevacuolar compartment, PVC, which corresponds to the late endosome; deep blue) or trafficked to the plasma membrane (PM).

Plasma membrane proteins like the auxin efflux carrier PIN1 (red), which accumulates at the basal PM at steady state, are continually internalised and trafficked to the TGN, which resembles the early endosome (EE) in plants.

From the TGN, PIN1 is recycled to the plasma membrane via the recycling endosome (RE; light blue). This pathway is regulated by the ARF-GEF GNOM.







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